

A Novel Pressure Regulating Brain Imaging Implant For Ultra-Large Field-of-View MicroscopicImaging in NHPs

Olivya Caballero¹, Jordi Chanovas¹, Manuel Ledo¹, Anirvan Nandy², Azadeh Yazdah-Shahmorad³, Edward Callaway⁴, Eyal Seidemann⁵, John Reynolds⁴, Michael Avery⁴, Peichao Li⁴, Shiming Tang⁶, Srinivastavan Kolla¹, Susana Martinez-Conde¹, Westley Hayes¹, Stephen Waite¹, John Carter¹, Yuzhi Chen⁵, Stephen Macknik¹.

¹Ophthalmology, Neurology, and Physiology/Pharmacology, State Univ. of New York, New York; ²Yale School of Medicine, New Haven, CT; ³University of Washington, Seattle, WA;

⁴The Salk Institute, La Jolla, CA; ⁵University of Texas, Austin, TX; ⁶Peking University, Beijing, China.



SUNY
DOWNSTATE
Health Sciences University

Problem Definition & Need

Problem Definition

Several problems challenge mesoscopic imaging in the brain: 1) Difficulty with positioning high-NA objectives near the brain; 2) Creating a flat imaging window against the surface of the brain; 3) Adjusting the imaging window to changes in swelling and pressure in the brain, such as those that may occur due to hydration changes and other physiological factors; 4) Preventing growth of dura and biofilms that cloud the imaging window; 5) Follow-on MRI imaging of the animal post-implantation.

Need Statement

Need a thin, strong, radiolucent, and pressure regulating implant for ultra-large field-of-view microscopic imaging in primates.

Design Inputs

Functional Requirement

Maintain a flat, clear, imaging window against the surface of the brain.

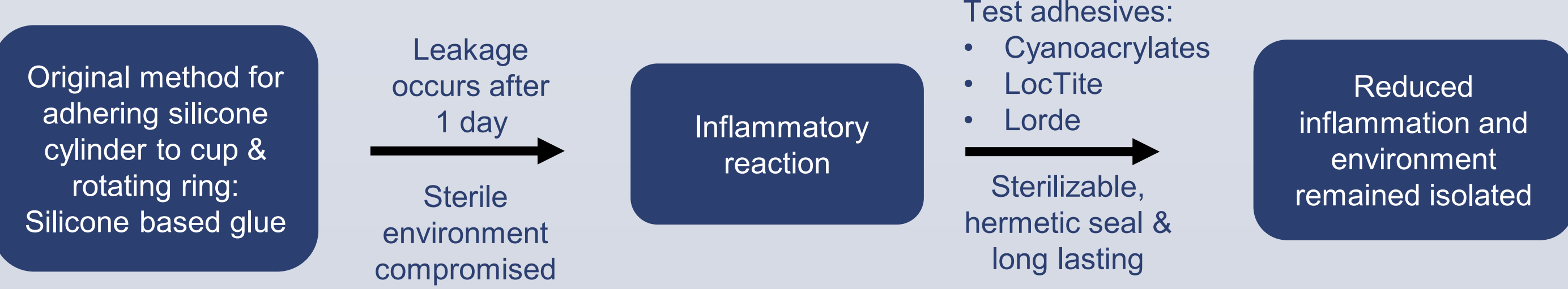
Constraints

- Radiolucent
- Thin enough to allow for an objective with a working distance of 2mm to image the brain.
- Allow for water immersion objective.
- Capable of holding attachments & bearing weight (at least 5 lbs).
- Imaging window must be at least 20 mm in diameter.
- Coverslip must move parallel to chamber to allow for manual control of the imaging window.
- Must be durable, last at least six months and be usable on awake monkeys.
- Must be hermetically sealed to prevent infection.

Failures & Modifications



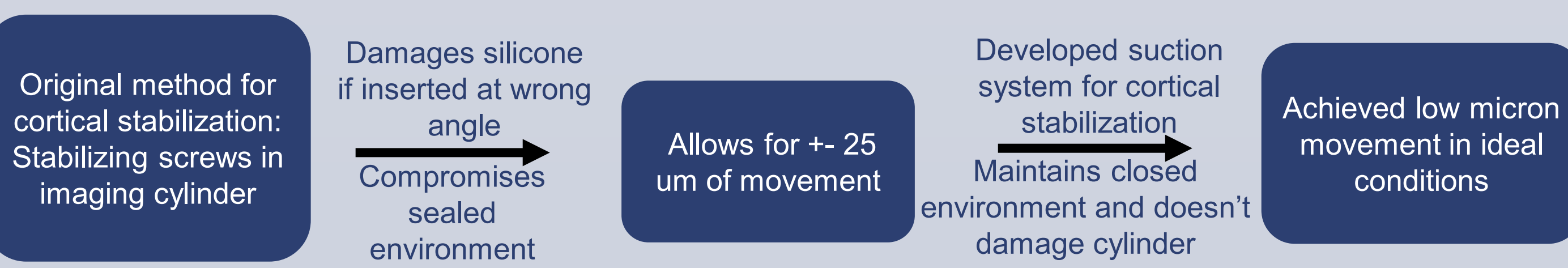
Modification: Redesigned the silicone gasket to include two O-rings. Previous design was a flat disk



Modification: Implemented use of Lorde medical grade adhesive. 3D printed structures to accommodate improved adhesion.



Modification: Switched from multi-axis titanium arm to a limited, bulkier, aluminum block.



Modification: Redesigned cylinder to have permanent, metal, 21G needle guides sealed with silicone adhesive to allow for penetration and subsequent suction. Works in conjunction with head stabilization to reduce motion.

Engineering Design Solution

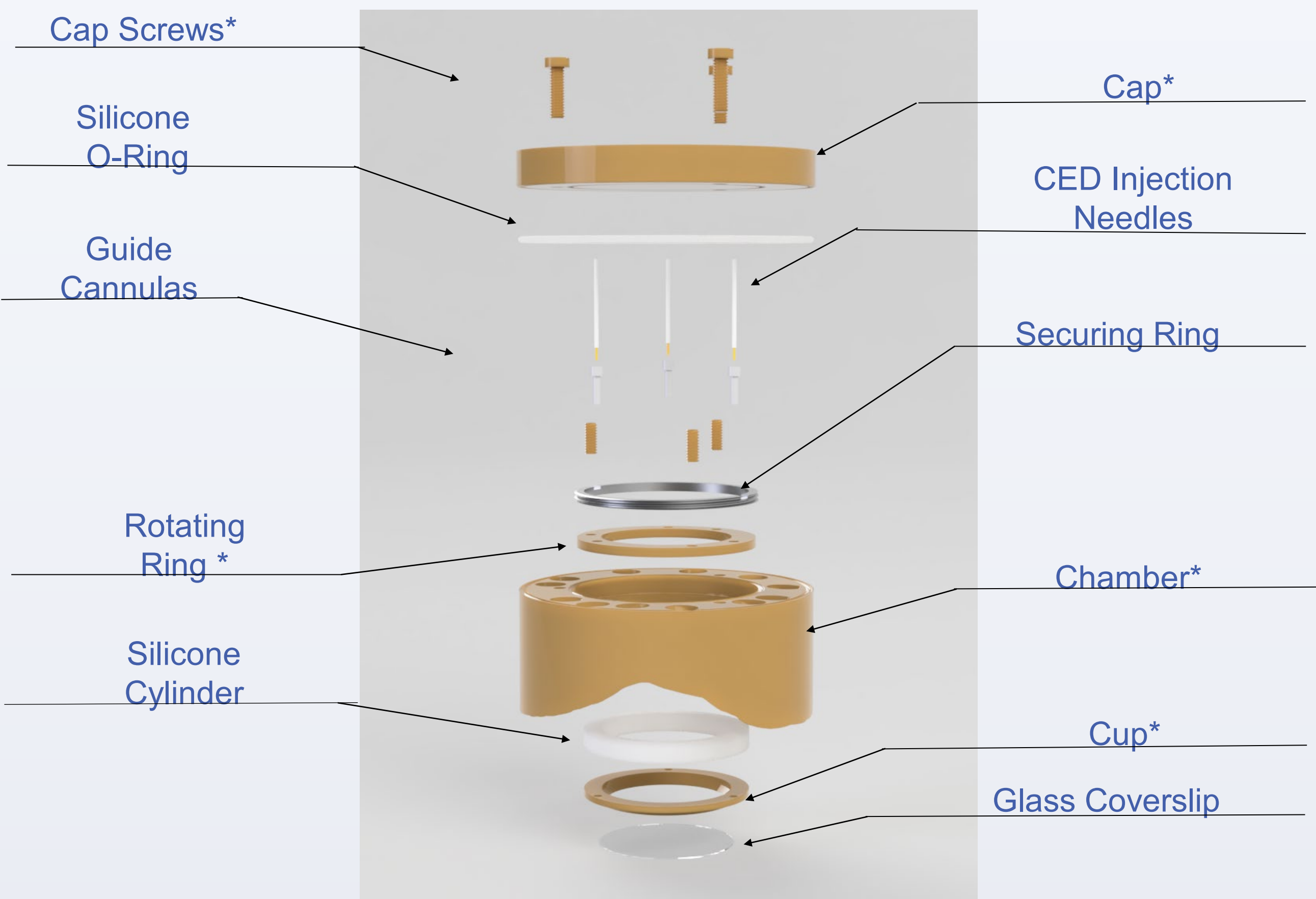


Figure 1: Exploded view of annotated imaging implant prototype. *Indicates part is made of PEEK plastic, chosen for its radiolucent properties, strength, and ability to be sterilized.

Component Listing with Description

Cap & Cap Screws- Removable cap for imaging and cleaning.

Silicone O-Ring - Approximately .4 mm thick, prevents bacterial movement between chamber and cap. There are two O-rings, one on the outer and one on the inner diameter of the chamber. The cap and chamber have indents to accommodate the rings and squeeze them when the cap is completely secured.

Thin Securing Ring - Secures rotating ring against the bottom shelf of the chamber and prevents it from moving up.

Guide Cannulas - Cannulas are threaded into the rotating ring at 3 different locations and sealed with silicone glue. These cannulas allow for 21G CED needles to penetrate the cylinder.

CED Injection Needles - Convection enhanced delivery needles. Designed for cortical injections but can also be used to deliver drugs or imaging contrast agents into the soft tissue.

Rotating ring - Multiple threaded holes for the height adjusting screws and can rotate to adjust the positions of the screws. Sits between shelf in chamber and thin securing ring.

Chamber - Has holes for bone screws that are perpendicular to the surface of the bone to increase the strength of the bond between the chamber and bone². It also includes three threaded holes on the top to allow for attachments and to secure the cap.

Silicone - Chosen for its ease in manufacturing, control of mechanical properties, and ability to be sterilized. Serves as a spring to adjust for pressure changes caused by the variations in swelling in the brain. Although not indicated in the drawing, the silicone connects to the lip of the imaging cup and rotating ring to create a sealed environment.

Imaging Cup & Glass Cover Slip - The glass is glued to the cup. Together they create a bowl that can hold liquid for a water immersion objective.

Modifications for 3D printing

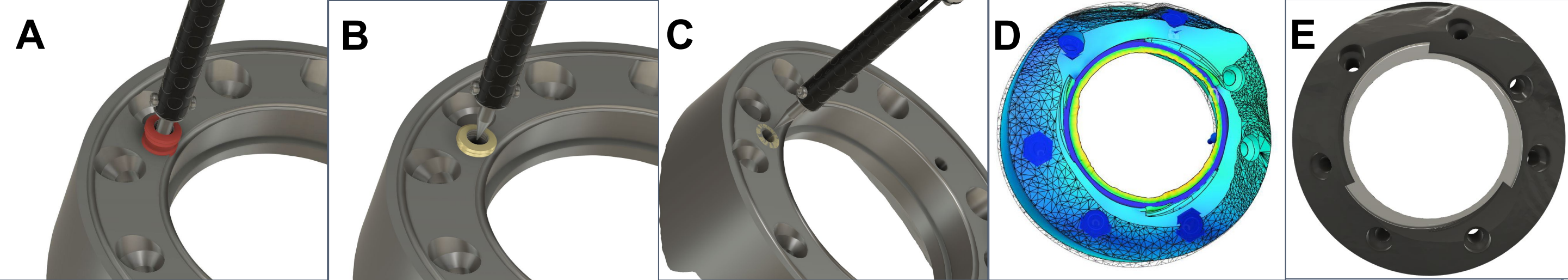


Figure 2: Panels (A), (B) & (C) show the progression of replacing damaged threads on the implant using a soldering iron to press new, metal inserts into PEEK plastic at 550 degrees Celsius. Panels (D) & (E) show a modified tab design to replace the threaded ring and it easier to print. Panel (D) is a strength simulation showing >2 e⁴ mm of displacement when 20 N of force is placed on the ring.

Viral Transduction Results

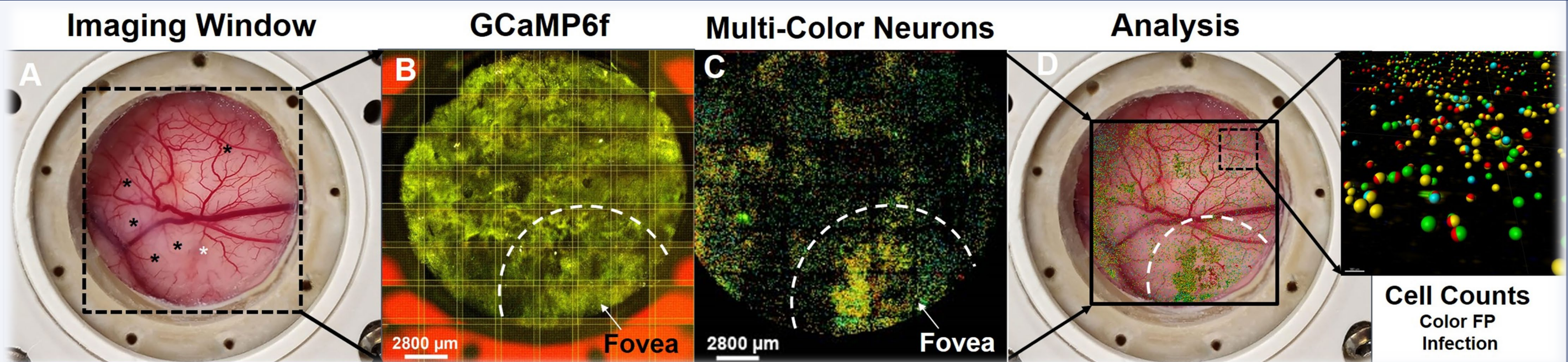


Figure 2: (A) 2cm imaging window (stars indicate previous probe sites). White star indicates the LGN CED injection site. (B) Stitched 2P image (49 tiled 2800 μm stacks) of GCaMP6f infected cells. (C) Multicolor scan of multiple fluorescent protein cells. (D) Positions of individual best of multi-color FP cells. (E) Hyperspectral brainbow analysis. We were able to successfully transfect the cortical area beneath the implant with five multi-color fluorescent proteins and GCaMP6. The implant allowed for impressive imaging using our 2-photon scope and after analysis revealed successful transfection of the virus' injected. The results indicate that the implant was successful in its ability to keep enough dural and pial regrowth at bay to allow for the imaging previously discussed.

Bioluminescence Brainbow Spectral Analysis

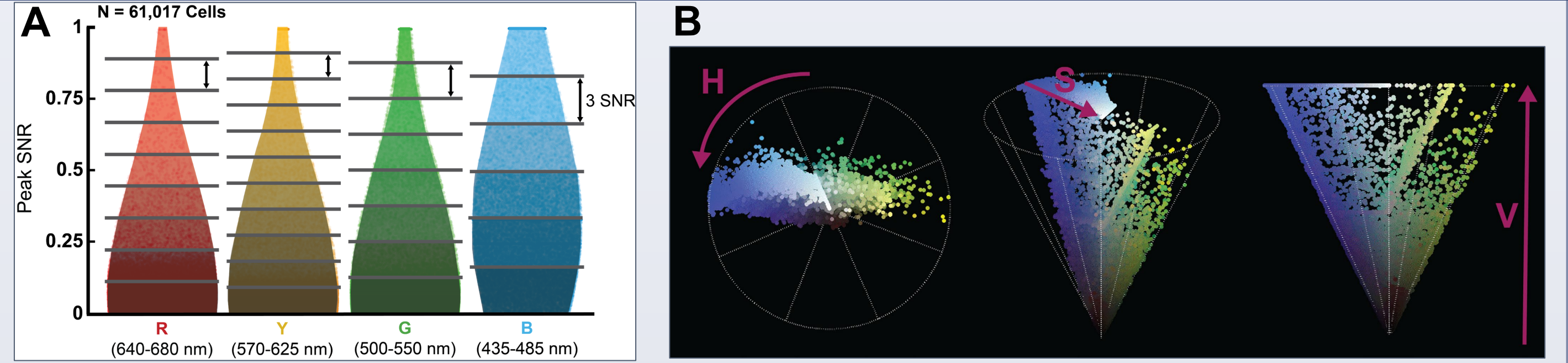


Figure 3: (A) Violin plot shows 4,752 distinguishable colors. (B) indicates the hue, saturation, and values of the cell colors.

Stability Analysis

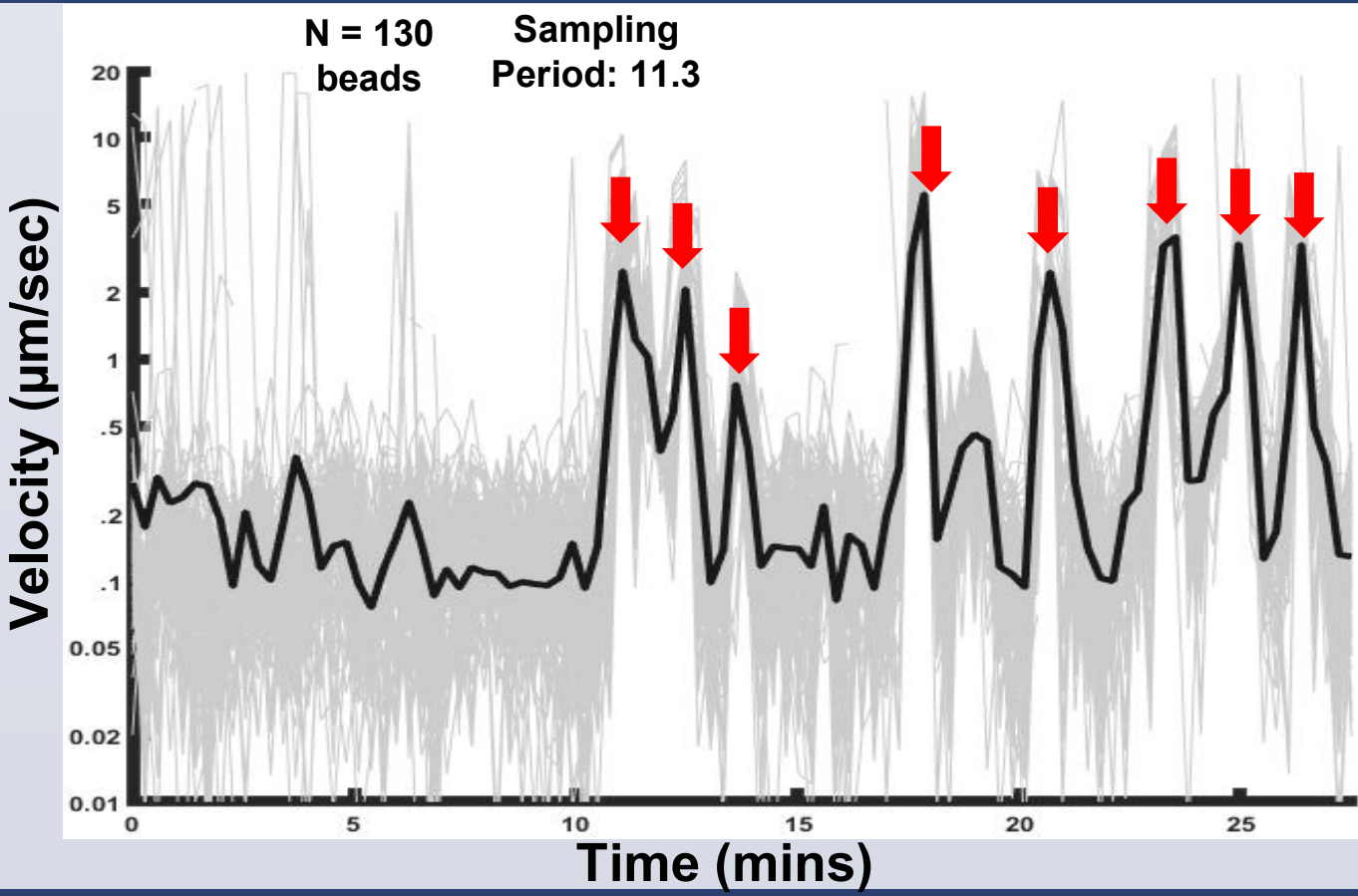
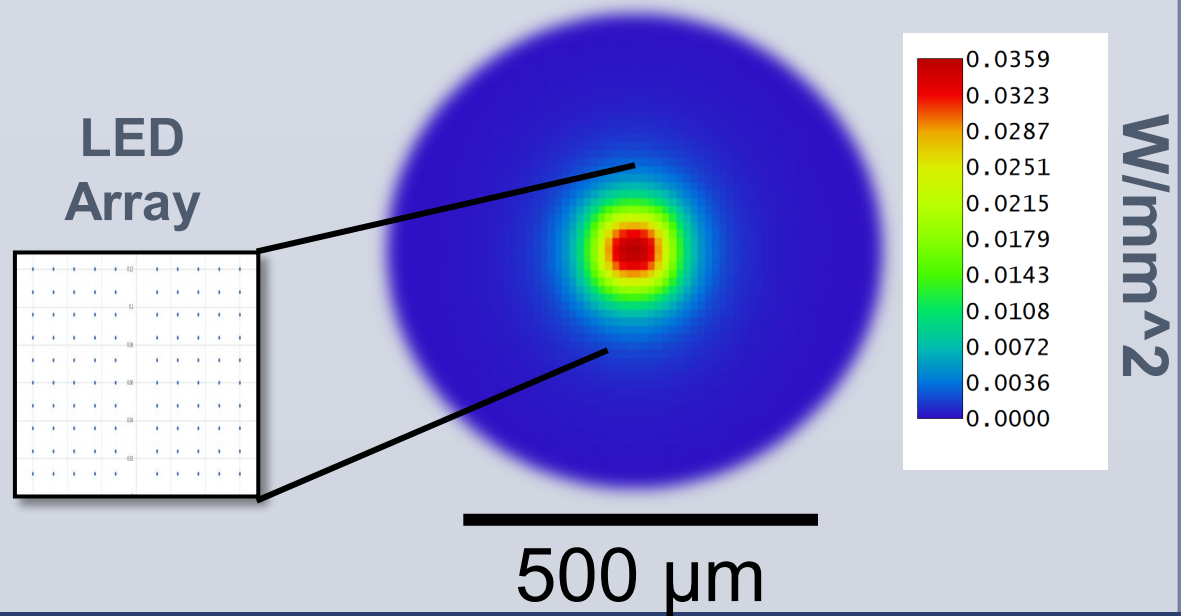


Figure 4: X-Y motion was measured by placing a clear slide of fluorescent od = microbeads on the imaging glass of the chamber. The movement of each bead (gray lines) was recorded & averaged (black line). Large bodily movements of the NHP are indicated with red arrows. As seen in the figure the NHP becomes restless after ~ 30 mins of imaging.

LED Simulation for OBServ

Figure 5: LED light scatter simulation, 1 mm deep into the cortex using Zemax Optic Studio. LED positional array of 100 16 x 16 μm LEDs separated by 8 μm, each LED operates at .012 mW/mm² of power.



References

1. <http://bionumbers.hms.harvard.edu/bionumber.aspx?id=106646>
2. Grant H. Mulliken, Robert Desimone, et al. (2015). Custom-fit radiolucent cranial implants for neurophysiological recording and stimulation.
3. Ruiz O, Roe Aw, et al. (2013). Optogenetics through windows on the brain in the nonhuman primate.
4. Yazdan-Shahmorad, A. et al. A Large-Scale Interface for Optogenetic Stimulation and Recording in Nonhuman Primates. Neuron 89, 927-939, doi:10.1016/j.neuron.2016.01.013 (2016).
5. Cai, D., Cohen, K. B., Luo, T., Lichtman, J. W. & Sanes, J. R. Improved tools for the Brainbow toolbox. Nature methods 10, 540-547, doi:10.1038/nmeth.2450 (2013).
6. Zhang, G. R. et al. Neurons can be labeled with unique hues by helper virus-free HSV-1 vectors expressing Brainbow. J Neurosci Methods 240, 77-88, doi:10.1016/j.jneumeth.2014.11.009 (2015).

Acknowledgements

We thank Molla Nawsher for technical assistance, as well as the Desimone lab for their assistance and advice. Special thanks to Rogue Research for machining our implant. This work was supported by funding from the NSF-NCS Award # 1734887.