

1 RRH: INTRATEST TRACE ELEMENT GEOCHEMISTRY

2 LRH: HUPP AND FEHRENBACHER

3

4 INTRATEST TRACE ELEMENT VARIABILITY IN POLAR AND

5 SUBPOLAR PLANKTIC FORAMINIFERA: INSIGHTS INTO VITAL

6 EFFECTS, ONTOGENY, AND BIOMINERALIZATION PROCESSES

7

8 BRITTANY N. HUPP<sup>1,2,3\*</sup> AND JENNIFER S. FEHRENBACHER<sup>2</sup>

9 <sup>1</sup>4400 University Dr. MS2B3, Department of Atmospheric, Oceanic and Earth Sciences, George  
10 Mason University, Fairfax, VA 22030 USA

11 <sup>2</sup>Ocean Administration Building, 101 SW 26th St. College of Earth, Ocean, and Atmospheric  
12 Sciences, Oregon State University, Corvallis, OR 97331 USA

13 <sup>3</sup>NOAA Climate and Global Change Postdoctoral Fellowship Program, CPAESS, UCAR,  
14 Boulder, CO, USA

15

16 \*Corresponding Author: Brittany N. Hupp ([bhupp@gmu.edu](mailto:bhupp@gmu.edu))

17

18

19

20

21

22

23

## ABSTRACT

24

25 To use planktic foraminiferal tests as paleoproxy substrates, it is necessary to delineate  
26 environmental versus biological controls on trace element incorporation. Here we utilize laser-  
27 ablation inductively coupled plasma mass spectrometry (LA-ICP-MS) to explore interspecies,  
28 chamber-to-chamber, and intratest trace element (i.e., Mg, Na, Sr, Ba, Mn, Zn) variability in  
29 thickly-calcified specimens of the polar and subpolar planktic foraminifera *Neogloboquadrina*  
30 *incompta*, *N. pachyderma*, and *Turborotalita quinqueloba* collected from plankton tows in the  
31 Northern California Current. Among the study taxa, test Mg/Ca, Na/Ca, and Sr/Ca are likely  
32 dominantly controlled by depth habitat. The neogloboquadrinids record higher Ba/Ca and Mn/Ca,  
33 and also show positive covariance between these elements, possibly due to calcifying in an  
34 oxygen-depleted marine snow microhabitat. Trace elements are found to be more enriched in the  
35 lamellar calcite than the outer chamber wall dominated by gametogenic crust. The data presented  
36 herein provide insight into potential vital effects, paleoproxy considerations, ontogeny, and  
37 biomineralization processes.

38

39

40

41

42

43

44

45

46

47 INTRODUCTION

48

49 The trace element geochemistry (TE/Ca) of foraminifera varies as a function of seawater  
50 composition, the physical conditions of the water in which they are calcifying, and the biological  
51 conditions during calcification (e.g., Lea et al., 1999; Erez, 2003; Russel et al., 2004; Katz et al.,  
52 2010; Allen et al., 2016; de Nooijer et al., 2017; Evans et al., 2018). Thus, great efforts have been  
53 made to quantitatively constrain relationships between foraminiferal test TE/Ca geochemistry and  
54 the conditions of the foraminifer's living environment, as a means for paleoenvironmental  
55 reconstruction. While some foraminiferal TE/Ca paleoproxies have been rigorously studied, such  
56 as Mg/Ca as a paleotemperature proxy (Nürnberg, 1995, Nürnberg et al., 1996; Lea et al., 1999;  
57 Anand et al., 2003; Russell et al., 2004; Martínez-Botí et al., 2011; Hönisch et al., 2013; Holland  
58 et al., 2020), incorporation of other TEs into foraminiferal calcite is less well-understood and  
59 represent new pathways of needed exploration. In addition to understanding the environmental  
60 controls on TE incorporation, it is equally important to constrain how biological processes  
61 modulate TE geochemistry. Biological processes that influence test geochemistry, broadly referred  
62 to as “vital effects” (Niebler et al., 1999; de Nooijer et al., 2014; Schiebel and Hemleben, 2017),  
63 can distort primary environmental signals, thus interfering with paleoreconstructions reliant upon  
64 test geochemistry. For example, trace element geochemistry may differ from chamber-to-chamber  
65 or when examining profiles of TEs across a chamber wall (Eggins et al., 2003; Steinhardt et al.,  
66 2014, Steinhardt et al., 2015; Davis et al. 2020; Hupp and Fehrenbacher, 2023). Furthermore,  
67 ontogenetic processes, such as test thickening observed in several species of planktic foraminifera  
68 syn- and post-gametogenesis, have been shown to impart different geochemical signatures in

69 different portions of the test wall (see Table 1 of Hupp and Fehrenbacher, 2023 and references  
70 therein). Recognizing intratest variability is informative to identifying potential vital effects, as  
71 well as providing insight into ontogeny and biomineralization processes (Hathorne et al., 2003;  
72 Eggins et al., 2003; Davis et al., 2020). Fossilized test TE/Ca variability can be further confounded  
73 by the addition of diagenetic calcite, which varies even more from primary foraminiferal calcite  
74 compositions (e.g., Boussetta et al., 2011; Kozdon et al., 2013; Edgar et al., 2015).

75 Recent advances in *in situ* technologies such as laser-ablation inductively coupled plasma mass  
76 spectrometry (LA-ICP-MS) provide a unique opportunity to investigate intratest trace element  
77 variability. LA-ICP-MS analysis allows for quantifying TEs in individual foraminifera in a  
78 population of specimens (i.e., intertest variability). By analyzing micron-scale intratest variability  
79 using LA-ICP-MS, one can also identify potential vital effects and investigate other mechanisms  
80 responsible for trace element incorporation in foraminifera (Hathorne et al., 2003; Eggins et al.,  
81 2003; Sadekov et al., 2010; Davis et al., 2020). This is necessary to identify intertest and intratest  
82 TE variability that potentially confound “averaged” TE measurements used in  
83 paleoreconstructions, as is the case in more traditional, pooled, whole-test ‘bulk’ analytical  
84 techniques, such as solution analysis.

85 LA-ICP-MS is commonly used to investigate TE geochemistry in modern planktic  
86 foraminiferal species (e.g., Hathorne et al., 2009; Steinhardt et al., 2015; Davis et al., 2020),  
87 however, these investigations predominantly focus on tropical, subtropical, and transitional  
88 species. Here, we explore the intratest variability of polar and subpolar modern planktic  
89 foraminiferal species, specifically *Neogloboquadrina incompta* (Cifelli, 1961), *Neogloboquadrina*  
90 *pachyderma* (Ehrenberg, 1861), and *Turborotalita quinqueloba* (Natland, 1938), to bolster their  
91 usage as geochemical paleoproxy substrates. Specimens investigated in this study were collected

92 from plankton tow samples throughout the Northern California Current region under a wide range  
93 of environmental conditions between 2011 and 2022. All specimens were dead upon collection as  
94 identified by their thickly-crusted test walls and, in recently collected samples, the absence of  
95 remnant cytoplasm. Only encrusted specimens were examined, as the presence of crust indicates  
96 the end or near-end of an individual's life-cycle, and thus are likely more representative of  
97 fossilized specimens found on the seafloor (Bé and Lott, 1964; Bé, 1980; Caron et al., 1990; Davis  
98 et al., 2020). By characterizing the intratest variability of crusted (i.e., dead) specimens collected  
99 across a wide range of conditions in the uppermost surface ocean, we aim to provide a foundation  
100 from which future trace element studies focused on polar and subpolar planktic foraminifera  
101 collected from sediment traps and the seafloor can be compared. Furthermore, this study illustrates  
102 the wide range of approaches that can be taken to investigating foraminiferal geochemistry when  
103 using LA-ICP-MS data. This study provides insight into vital effects and trace element  
104 incorporation into the tests of polar and subpolar planktic foraminifera taxa independent of  
105 potential trace element heterogeneity associated with travel through the water column (e.g.,  
106 scavenging; Davis et al., 2020) or early diagenesis upon the seafloor (e.g., Branson et al., 2015).  
107

108 **HYDROGRAPHIC SETTING**

109  
110 Study specimens were collected via surface plankton tow from offshore sites throughout the  
111 Northern California Current (NCC) region, ranging from northern California to the border between  
112 Washington (USA) and Canada (**Fig. 1**). This region of the Northeast Pacific Ocean is  
113 characterized by a relatively narrow, deep continental shelf (0 to 200 m) and a steep continental  
114 slope that transitions into deep (> 2500 m) water 25-50 km offshore. A well-developed seasonal

115 oxygen minimum zone (OMZ, 600-1000 m) associated with North Pacific Intermediate Water  
116 exists offshore (e.g., Paulmier and Ruiz-Pino, 2009; Pierce et al., 2012). As in many other eastern  
117 boundary ocean margins, upwelling along the margin is a highly seasonal phenomenon driven by  
118 shifts in wind strength and direction (Barth and Wheeler, 2005). During spring and summer, this  
119 region experiences strong upwelling caused by prevailing northerly winds that induce Ekman  
120 transport of deeper water from the shelf break onto the continental shelf (e.g., Kirincich et al.,  
121 2005; Perlin et al., 2005). Fall and winter display starkly different conditions, with southerly,  
122 downwelling-favorable winds, large swells, high wave energies nearshore, and the arrival of  
123 moisture-laden storms (Kniskern et al., 2011). Environmental variability due to seasonal upwelling  
124 is further overprinted by longer-term oscillations such as El-Niño Southern Oscillation and the  
125 Pacific Decadal Oscillation. Collectively, these features create a dynamic region with diverse  
126 conditions under which regional planktic foraminifera calcify (Ortiz and Mix, 1992; Taylor et al.,  
127 2018; Lane et al., 2023).

128

## 129 MATERIALS AND METHODS

130

### 131 PLANKTON TOWS AND STUDY SPECIMENS

132

133 Plankton tows were collected from several sites (**Fig. 1**) during a series of cruises conducted  
134 between 2011 and 2022. Vertical tows were conducted from 100 m depth to the surface using a  
135 150  $\mu$ m mesh net. Most tows were obtained during NOAA-led fisheries survey cruises, where  
136 samples were preserved using NaOH-buffered formalin immediately upon collection and stored in  
137 a sample archive (Feinberg and Peterson, 2003). Planktic foraminiferal tests were removed from

138 archived tows, rinsed with DI water upon removal, and stored on micropaleontological slides until  
139 further cleaning was conducted in preparation for geochemical analysis. In addition to tests from  
140 archived tows, additional tows were collected by the authors where tests were immediately  
141 removed from tows, rinsed with DI water, and stored on micropaleontological slides, without any  
142 exposure to preservative chemicals. In total, specimens examined herein originate from 39 discrete  
143 tows collected across 29 different sites over a period of 11 years. Metadata for each test examined,  
144 including tow location (site name, latitude, longitude) and archived (preserved in buffered  
145 formalin) versus recently collected (never exposed to preservatives) status is detailed in the  
146 **Supplementary Appendix 1.**

147 Specimens of *N. incompta* (N = 92), *N. pachyderma* (N = 55), and *T. quinqueloba* (N = 42)  
148 were selected for individual foraminifer geochemical analysis. All three study taxa are asymbiotic;  
149 however, *T. quinqueloba* is a spinose species whereas the neogloboquadrinids are non-spinose  
150 (Schiebel and Hemleben, 2017). Specimens selected for analysis were limited to heavily-calcified  
151 tests (i.e., clearly apparent crusts) where it is inferred or confirmed that the specimens were dead  
152 upon collection. For archive specimens, dead specimens were inferred based upon evidence of a  
153 clearly apparent gametogenic crust (see fig 1. in Hupp and Fehrenbacher, 2023 for further  
154 description of crusted versus uncrusted individuals). For the recently collected specimens, dead  
155 specimens were heavily-calcified and lacked cytoplasm.

156

#### 157 TEST PREPARATION AND CLEANING

158

159 Study specimens underwent an oxidative cleaning process to remove remnant organic matter  
160 and detritus (Mashiotta et al., 1999) which consisted of 1) rinsing the tests with methanol, 2)

161 rinsing the tests with ultrapure milliQ water three times 3) submerging tests in a 1:1 solution of  
162 30% H<sub>2</sub>O<sub>2</sub> buffered with 0.1 M NaOH and placing vials containing the tests and solution in a warm  
163 (~65°C) water bath for 10 minutes, and lastly, 4) removing the H<sub>2</sub>O<sub>2</sub>-NaOH solution and rinsing  
164 the tests with milliQ water three times. While previous studies submerge the tests for 30 minutes,  
165 the water bath time was decreased to 10 minutes to prevent test dissolution (Bonnin et al., 2019).

166

#### 167 LA-ICP-MS

168

169 Trace element geochemistry was determined via laser-ablation inductively coupled plasma  
170 mass spectrometry (LA-ICP-MS). Tests were analyzed with a Thermo Scientific iCAP RQ  
171 quadrupole ICP-MS coupled to an Applied Spectra RESOlution (193 nm) laser-ablation system  
172 housed in the Keck Collaboratory for Plasma Spectrometry at Oregon State University. While ten  
173 isotopes were measured during analysis, seven isotopes are examined herein: <sup>24</sup>Mg, <sup>23</sup>Na, <sup>88</sup>Sr,  
174 <sup>138</sup>Ba, <sup>55</sup>Mn, <sup>66</sup>Zn, and <sup>43</sup>Ca. Isotopes were measured using a rapid peak-hopping method where  
175 Mn, Zn, Ba were assigned dwell times of 0.05 s, Na a dwell time of 0.04 s, Sr and Mg dwell times  
176 of 0.02 s, and Ca a dwell time of 0.01 s (total sweep time: 0.29 s). Analysis spots were ablated  
177 from the outer chamber wall (the test surface) through the chamber wall. For the  
178 neogloboquadrinids, laser energy was set to 4.0 mJ attenuated by 87.5% whereas for *T.*  
179 *quinqueloba* the laser energy was reduced to 3.0 or 3.5 mJ, also attenuated by 87.5%. Circular spot  
180 sizes range from 24 to 50 µm and were adjusted for each specimen to maximize the ablated area.  
181 Repetition rates were adjusted between 5 and 6 Hz to optimize laser profiles. Individual laser  
182 analyses were conducted on the F0, F1, F2, F3, and F4 chambers of *T. quinqueloba* and the F0,  
183 F1, F2, and F3 chambers for *N. incompta* and *N. pachyderma*. In rare cases, due to test orientation,

184 the oldest chamber in the outer whorl (F4 and F3 for *T. quinqueloba* and the neogloboquadrinids,  
185 respectively) was not accessible to be analyzed. Repeat analyses were conducted on at least one  
186 chamber per specimen to assess reproducibility (**Supplemental Table 1**).

187 Three standard reference materials were measured bracketing every 10 to 12 specimens (i.e.,  
188 every 50 to 72 test ablation spots across 10 to 12 continuous profiles), including the glasses NIST  
189 610 and NIST 612, and the compressed-powder USGS standard MACS-3. Each standard was  
190 ablated for 60 s using a repetition rate of 6 Hz on a spot size of 50  $\mu\text{m}$  with a laser energy of 5 mJ  
191 attenuated by 50%. All TE/Ca were calibrated using the NIST 610 and 612 glasses, aside from  
192 Na/Ca which was calibrated using NIST 610 and MACS-3 due to a consistent concentration of Na  
193 within NIST glass standards. Elemental calibrations of standard reference materials measured  
194 throughout the analysis sessions yielded  $r^2 \geq 0.99$ .

195 Data was processed with the Python package *LAtools* (Branson et al., 2019) following  
196 established data reduction protocols (Longerich et al., 1996). Data reduction involves despiking  
197 and signal smoothing to remove outliers in the laser spectra, evaluation and correction for any drift  
198 observed in the standard measurements throughout the analysis session, and removal of average  
199 background counts from each data point. Data output from this process produces individual  
200 specimen files, where each spot measurement composes a distinct section of the full spectrum, as  
201 well as mean TE/Ca for each spot analysis, normalized to known TE/Ca concentrations of the  
202 standards (**Supplemental Fig. 1**; Jochum et al., 2011). Whole-test averages and individual  
203 chamber averages of all TEs for each specimen are reported in **Supplemental Appendix 2**. For  
204 chambers where multiple measurements were taken, the replicate measurements were averaged to  
205 produce the chamber values reported in **Supplemental Appendix 2**.

206

## RESULTS

## INTERSPECIES WHOLE TEST AVERAGE TE/CA VARIABILITY

The range of whole-test average TE/Ca compositions for the study specimens provides context for further investigation of the intratest geochemistry in these species (**Fig. 2, Table 1**). Whole-test averages were determined by taking the mean of the average chamber measurements for each chamber in the outer whorl (**Fig. 2A**). Note that where multiple measurements were made within a single chamber, those measurements were averaged to determine a chamber mean before being used to calculate the whole-test average. It is recognized that the whole-test average as described here is technically an interpolated average, as we are calculating the mean of all discrete measurements taken from chambers in the outer whorl as opposed to dissolving and analyzing the entire test, as done in solution analyses. However, comparing the “whole test” average as measured by solution-based ICP-MS and interpolated by laser-based ICP-MS result in negligible differences (Fehrenbacher et al., 2020). We also recognize that while much of the species variability is directly tied to environmental variability during specimen formation/period of calcification, the whole test TE/Ca geochemistry provides context for the subsequent discussion of intratest TE/Ca variability. Furthermore, examination of whole-test distributions provides insight into elements that are potentially modulated by environmental conditions. For example, TEs that do not vary significantly may suggest that these TEs are not significantly impacted by calcification conditions (e.g., Ba/Ca in *T. quinqueloba*; **Fig. 2E**).

228 Here we describe the whole-test average distributions by examining the distribution mean  
229 (**Table 1**) as well as the distribution range, as defined by the maximum whisker value minus the

230 minimum whisker value of the boxplots shown in **Figure 2**. *Neogloboquadrina incompta*  
231 generally records the lowest average Mg/Ca, Na/Ca, and Sr/Ca, followed by *N. pachyderma*, and  
232 *T. quinqueloba* (**Fig. 2B-D**). The range of the distributions is fairly consistent between all taxa  
233 for Mg/Ca (~3 mmol/mol) and Sr/Ca (~0.25 mmol/mol), but the Na/Ca range increases from: ~6  
234 mmol/mol in *N. incompta*; to 13 mmol/mol in *N. pachyderma*; and to 22 mmol/mol in *T.*  
235 *quinqueloba*, respectively. Interspecies TE/Ca differences are more irregular when comparing  
236 Ba, Mn, and Zn (**Fig. 2E-G**). While all taxa exhibit similar Ba/Ca minima, *N. incompta* exhibits  
237 the largest range of Ba/Ca values (41  $\mu$ mol/mol), followed by *N. pachyderma* (16  $\mu$ mol/mol),  
238 and lastly with the smallest range, *T. quinqueloba* (5  $\mu$ mol/mol) (**Fig. 2E**). *Neogloboquadrina*  
239 *incompta* also exhibits a larger range of Mn/Ca values (25  $\mu$ mol/mol) compared to *N.*  
240 *pachyderma* (7  $\mu$ mol/mol) and *T. quinqueloba* (11  $\mu$ mol/mol) (**Fig. 2F**). The inner quartile  
241 ranges are comparable among all three taxa for Zn/Ca, but the full range of whole test values is  
242 largest in *N. pachyderma* (977  $\mu$ mol/mol), followed by *T. quinqueloba* (728  $\mu$ mol/mol), and  
243 lastly, *N. incompta* (586  $\mu$ mol/mol). *Neogloboquadrina pachyderma* also records higher Zn/Ca  
244 values on average than *T. quinqueloba* and *N. incompta* (**Fig. 2G**).  
245

246 CHAMBER-TO-CHAMBER TE/CA VARIABILITY

247  
248 To explore how TE/Ca may change through ontogeny, chamber-to-chamber TE/Ca  
249 averages are compared to one another by mean centering the TE/Ca data (i.e., whole test average  
250 TE/Ca is subtracted from each chamber average; **Fig. 3A**). By normalizing each chamber  
251 average by the test average, the range of chamber-to-chamber variability can be examined while

252 removing variability that could be more directly attributed to calcification under different  
253 environmental conditions.

254 For all three taxa, the Mg/Ca composition decreases in each successively-added chamber  
255 in the outer whorl, with the youngest chamber (F0) exhibiting the lowest values found in the outer  
256 whorl (**Fig. 3B, E, H**). The F4 chamber of *T. quinqueloba* does not seem to follow the intratest  
257 pattern (**Fig. 3H**), however this could be an artifact, as not all F4 chambers could be analyzed in  
258 this taxon. Ratios of Na/Ca and Sr/Ca show minimal chamber-to-chamber variability in *N.*  
259 *pachyderma* (**Fig. 3F, G**) and *T. quinqueloba* (**Fig. 3I, J**). However, in *N. incompta*, the F0  
260 chamber records consistently higher Na/Ca than all other chambers in the outer whorl (**Fig. 3C**).  
261 A consistent increase in Sr/Ca is observed with each successively younger chamber in the outer  
262 whorl of *N. incompta* (**Fig. 3D**). The ratio of Ba/Ca differs little between chambers in *N. incompta*  
263 (**Fig. 4A**), however the older chambers (i.e., F2, F3) tend to record slightly higher Ba/Ca values  
264 than the ultimate and penultimate chambers of *N. pachyderma* and *T. quinqueloba* (**Fig. 4D, G**).  
265 A similar pattern of decreasing TE/Ca content in each sequentially-added chamber is observed  
266 with Mn/Ca in *N. pachyderma* (**Fig. 4E**) and *T. quinqueloba* (**Fig. 4H**), whereas *N. incompta* shows  
267 only a relative Mn/Ca depletion in the F0 chamber (**Fig. 4B**). The neogloboquadrinids exhibit an  
268 opposite pattern with Zn/Ca where Zn incorporation increases with each progressively added  
269 chamber (**Fig. 4C, F**). Conversely, *T. quinqueloba* shows a progressive decrease in Zn  
270 incorporation with each progressively-added chamber, aside from the ultimate F0 chamber (**Fig.**  
271 **4I**).

272

273 INTRATEST TE/CA VARIABILITY

274

275 In addition to characterizing the chamber-to-chamber variability, the range of values  
276 measured from individual chambers within a single test is also explored (**Fig. 5**). The intratest  
277 range is calculated by subtracting the lowest chamber average value of those measured in the outer  
278 whorl from the highest chamber average value of outer whorl chambers in a single specimen (e.g.,  
279 **Fig. 5A**). Here, the medians of the boxplot distributions (**Fig. 5**) will be discussed, as some TE/Ca  
280 exhibit several outliers influencing the distribution mean. The median intratest range of Mg/Ca  
281 values  $\sim 0.8$  mmol/mol in the neogloboquadrinids, however the distribution of ranges exhibits a  
282 greater spread for *N. pachyderma* than for *N. incompta* (**Fig. 5B**). *Turborotalita quinqueloba*  
283 exhibits a median intratest Mg/Ca range of  $\sim 1.6$  mmol/mol. The median of the intratest Na/Ca  
284 range increases from 2.0 mmol/mol in *N. incompta* to 3.4 mmol/mol in *N. pachyderma* to 8.8  
285 mmol/mol in *T. quinqueloba*, and the distribution spread of intratest ranges increase in the same  
286 taxa order (**Fig. 5C**). Distributions of intratest Sr/Ca ranges are very similar among the three study  
287 taxa, with a median intratest range of  $\sim 0.1$  mmol/mol (**Fig. 5D**). Intratest Ba/Ca ranges show an  
288 opposite pattern among taxa as observed in Na/Ca (**Fig. 5E**). The median of intratest Ba/Ca ratios  
289 was found to be 6.4  $\mu$ mol/mol in *N. incompta*, 4.7  $\mu$ mol/mol in *N. pachyderma*, and 2.4  $\mu$ mol/mol  
290 in *T. quinqueloba*. The boxplot distribution of intratest Ba/Ca ranges shows the highest maximum  
291 value in *N. incompta* (36.6  $\mu$ mol/mol), followed by *N. pachyderma* (18.1  $\mu$ mol/mol), and lastly, *T.*  
292 *quinqueloba* (9.9  $\mu$ mol/mol). For Mn/Ca, *T. quinqueloba* shows the highest median internal  
293 variability (6.6  $\mu$ mol/mol), followed by *N. incompta* (6.3  $\mu$ mol/mol), and *N. pachyderma* (3.0  
294  $\mu$ mol/mol), however, the distribution of Mn/Ca intratest ranges recorded by *N. incompta* shows  
295 the greatest spread (**Fig. 5F**). Conversely, the highest median range of intratest Zn/Ca is recorded  
296 by *N. pachyderma* (357  $\mu$ mol/mol), followed by *T. quinqueloba* (194  $\mu$ mol/mol), and *N. incompta*

297 (111  $\mu\text{mol/mol}$ ), with *N. pachyderma* exhibiting the greatest variability of intratest ranges (**Fig.**  
298 **5G**).

299 Intratest TE/Ca variability, characterized by the intratest range and standard deviation of  
300 average chamber measurements in the outer whorl, can be compared to the whole-test TE/Ca value  
301 of an individual specimen to determine if intratest variability fluctuates consistently with  
302 increasing or decreasing TE/Ca content (**Table 2**). Intratest Mg/Ca variability shows a strong  
303 (range  $r^2 = 0.67$ , and standard deviation or “SD”  $r^2 = 0.68$ ) and moderately strong (range and SD  
304  $r^2 = 0.58$ ) positive relationship with whole-test Mg/Ca values in *N. incompta* and *N. pachyderma*,  
305 respectively. No correlation is found between intratest Mg/Ca variability and the whole-test  
306 average value in *T. quinqueloba*. *Neogloboquadrina incompta* exhibits a moderate positive  
307 correlation (range  $r^2 = 0.53$ , SD  $r^2 = 0.54$ ) between intratest Na/Ca variability and the whole-test  
308 value, whereas *N. pachyderma* (range and SD  $r^2 = 0.36$ ) and *T. quinqueloba* (range  $r^2 = 0.32$ , SD  
309  $r^2 = 0.33$ ) exhibit only weak positive correlations. No relationship is found between Sr/Ca intratest  
310 variability and the whole-test Sr/Ca value in any of the three study taxa. Whole-test Ba/Ca shows  
311 a very strong positive correlation with intratest variability in *N. pachyderma* (range  $r^2 = 0.98$ , SD  
312  $r^2 = 0.99$ ), and a strong positive relationship in *N. incompta* (range and SD  $r^2 = 0.64$ ) and *T.*  
313 *quinqueloba* (range  $r^2 = 0.75$ , SD  $r^2 = 0.76$ ). A strong positive relationship is found in *N. incompta*  
314 (range  $r^2 = 0.70$ , SD  $r^2 = 0.72$ ) and *T. quinqueloba* (range  $r^2 = 0.77$ , SD  $r^2 = 0.78$ ) when comparing  
315 intratest Mn/Ca variability to whole-test values, and a moderate to weak positive correlation is  
316 found in *N. pachyderma* (range  $r^2 = 0.41$ , SD  $r^2 = 0.39$ ). Moderate positive correlations are also  
317 found between intratest Zn/Ca variability and whole-test values in *T. quinqueloba* (range  $r^2 = 0.62$ ,  
318 SD  $r^2 = 0.61$ ) and *N. pachyderma* (range  $r^2 = 0.51$ , SD  $r^2 = 0.49$ ), with a weak correlation observed  
319 in *N. incompta* (range  $r^2 = 0.37$ , SD  $r^2 = 0.38$ ).

320

321 INTRACHAMBER TE/CA VARIABILITY

322

323 Laser ablation was conducted from the outside of the test through to the inner chamber wall  
324 allowing for individual depth profiles to be used to assess how the geochemistry of the test changes  
325 through the test wall (e.g., **Fig. 6**). All specimens examined for this study were chosen because  
326 they appeared to be thickly calcified, and all three species are known to exhibit a gametogenic  
327 crust (Schiebel and Hemleben, 2017; Pearson and Kucera, 2018). The inner versus outer calcite  
328 TE/Ca data are compared to evaluate if trace elements vary between the inner chamber wall and  
329 outer gametogenic crust (**Figs. 6, 7**). For simplicity, we assume the first half of a given depth  
330 profile (the outer calcite) is dominantly composed of crust calcite; the second half of the same  
331 depth profile, represents dominantly the inner chamber calcite or “lamellar” calcite (cf., Hupp and  
332 Fehrenbacher, 2023; **Fig. 6B**). If there is little to no difference in the average of these two values  
333 (e.g., data points sit upon a 1:1 line, **Figs. 6C, 7**), we assume that TEs do not vary greatly between  
334 the outer crust and the inner chamber wall. Conversely, if a chamber exhibits higher TEs in the  
335 crust or inner chamber wall, data points will either plot above or below the line of equality,  
336 respectively. While there is some variability in the amount of gametogenic calcite precipitated at  
337 the end of ontogeny, the 50% rule is an appropriate approximation (Bé, 1980; Ariwaka, 1983;  
338 Multizta et al., 1997; Bauch et al., 2002; Simstich et al. 2003; Bolton and Marr, 2013; Hupp and  
339 Fehrenbacher, 2023). Definitively quantifying the amount of early ontogenetic versus gametogenic  
340 crust calcite in our study populations (N = 189) would require laborious analyses (e.g., thin  
341 sectioning specimens for EMPA and SEM imaging) which is outside of the scope of this study.

342 For all trace elements, aside from Ba, all three taxa show greater enrichment in TEs in the inner  
343 50% of the spectrum (i.e., the lamellar calcite) compared to the outer 50% of the same spectrum  
344 (i.e., the outer crust; **Fig. 7**). Deviation from the line of equality is greatest in Mg/Ca, Na/Ca, and  
345 Sr/Ca, whereas the inner chamber wall shows only minor enrichments in Mn/Ca and Zn/Ca  
346 compared to the crust, aside from large deviations observed in the Zn/Ca of *T. quinqueloba* (**Fig.**  
347 **7R**). Ratios of Ba/Ca are also heavily enriched (in some cases by >200%) in the inner chamber  
348 walls of the neogloboquadrinids, yet *T. quinqueloba* shows a minor enrichment of Ba in the crust  
349 relative to the lamellar calcite of the inner test wall due to a few high-Ba specimens (**Fig. 7P**).  
350

351 PATTERNS IN TE/CA COVARIANCE  
352

353 As potential controls on TE incorporation are considered, it is helpful to evaluate patterns in  
354 TE covariance. **Figure 8A-C** shows covariance matrices for the whole-test TE/Ca values of our  
355 study specimens, separated by taxon. This results section will focus upon discussing moderate and  
356 strong relationships, as several elements covary at  $r^2 < 0.40$ . For *N. incompta*, Mg/Ca moderately  
357 positively covaries with Na/Ca ( $r^2 = 0.47$ ) and Sr/Ca ( $r^2 = 0.45$ ) on the whole-test average scale  
358 (**Fig. 8A**). Moderate positive relationships are also found between Ba/Ca and Mn/Ca ( $r^2 = 0.52$ ) as  
359 well as Ba/Ca and Zn/Ca ( $r^2 = 0.43$ ). For *N. pachyderma*, a strong positive relationship is observed  
360 between Na/Ca and Sr/Ca ( $r^2 = 0.64$ ) (**Fig. 8B**). Ratios of Na/Ca also moderately positively covary  
361 with Mg/Ca ( $r^2 = 0.50$ ), Ba/Ca ( $r^2 = 0.57$ ), and Mn/Ca ( $r^2 = 0.47$ ). Moderate positive covariations  
362 are found between Mg/Ca and Sr/Ca ( $r^2 = 0.45$ ), Mg/Ca and Ba/Ca ( $r^2 = 0.47$ ), Ba/Ca and Mn/Ca  
363 ( $r^2 = 0.48$ ), and Mn/Ca and Zn/Ca ( $r^2 = 0.40$ ) in this taxon. *Turborotalita quinqueloba* exhibits the  
364 least number of correlations between trace elements, with Mg/Ca and Na/Ca exhibiting a moderate

365 positive correlation ( $r^2 = 0.49$ ), and Sr/Ca and Ba/Ca exhibiting a strong negative correlation ( $r^2 =$   
366 -0.80) (**Fig. 8C**).

367 In addition to investigating covariance in TEs on the whole-test scale, the TE covariation in  
368 individual chambers is also explored (**Fig. 8D-F**). At first glance, there are many more significant  
369 ( $p\text{-value} \leq 0.05$ ) relationships at the chamber level. For example, some level of significant positive  
370 covariance is found among all elements studied in the neogloboquadrinids (**Fig. 8D, E**). Of note,  
371 *N. incompta* exhibits moderate positive covariance between Mg/Ca, Na/Ca, Sr/Ca, and Ba/Ca with  
372  $r^2$  ranging from 0.50 and 0.66 for each of the combinations of TE comparisons (**Fig. 8D**). Moderate  
373 positive relationships also exist between Mn/Ca and Na/Ca ( $r^2 = 0.42$ ), as well as Mn/Ca and Ba/Ca  
374 ( $r^2 = 0.43$ ). *Neogloboquadrina pachyderma* also shows moderate positive correlations between  
375 Mg/Ca, Na/Ca, and Sr/Ca ( $0.44 \leq r^2 \leq 0.51$ ; **Fig. 8E**). However, the strongest positive relationships  
376 in this taxon are found between Ba and Mn ( $r^2 = 0.73$ ) and Mn and Zn ( $r^2 = 0.71$ ), with Ba and Zn  
377 exhibiting a moderate positive relationship ( $r^2 = 0.47$ ). Comparisons of chamber TEs for *T.*  
378 *quinqueloba* result in more significant covariations than found on the whole-test scale, the  
379 strongest being between Mg and Na ( $r^2 = 0.57$ ) and Na and Ba ( $r^2 = 0.52$ ; **Fig. 8F**).

380

381 DISCUSSION

382

383 WHOLE TEST TE/CA GEOCHEMISTRY OF POLAR AND SUBPOLAR PLANKTIC FORAMINIFERA

384

385 Interesting differences in whole-test trace element geochemistry are identified among the  
386 three study species (**Fig. 2, Table 1**). For example, the whole-test Mg/Ca, Na/Ca, and Sr/Ca  
387 systematically increase when comparing *N. incompta* to *N. pachyderma* to *T. quinqueloba*,

388 respectively (Fig. 2). These interspecies differences are likely reflective of differences in living  
389 depth habitat in the study region where *N. incompta* prefers the deepest depth habitat of the three  
390 study taxa, and *T. quinqueloba* prefers the shallowest. Magnesium is known to be incorporated at  
391 greater concentrations into the foraminiferal crystal lattice when the test is grown under higher  
392 temperatures (Nürnberg et al., 1996; Lea et al., 1999; Anand et al., 2003; Russell et al., 2004;  
393 Martínez-Botí et al., 2011; Hönisch et al., 2013; Holland et al., 2020). The highest whole test  
394 Mg/Ca values are found in *T. quinqueloba*, suggesting that they live closest to the relatively  
395 warmer sea surface, and the lowest Mg/Ca values are found in *N. incompta*, suggesting that they  
396 prefer a deeper, cooler depth habitat (Fig. 2B). If we are to assume that the Mg/Ca composition of  
397 the three study taxa represents their preferred depth habitat, then this general pattern in relative  
398 depth habitat among the three study taxa are consistent with those observed in much of the Nordic  
399 Seas (Simstich et al., 2003), but differ from those characterized for the subtropical eastern North  
400 Atlantic (Rebotim et al., 2017).

401 Foraminiferal Na/Ca has been identified as a potential paleosalinity proxy, where test  
402 Na/Ca compositions positively covary with salinity (Wit et al., 2013; Mezger et al., 2016; Bertlich  
403 et al., 2018; Bertlich et al., 2021; Watkins et al., 2021) or as a proxy for the major ion chemistry  
404 of seawater on long timescales (Zhou et al., 2021; Gray et al., 2023). Among the data presented  
405 herein, there is a systematic increase in Na/Ca where not only the whole-test value but also the  
406 range of whole test Na/Ca values progressively increase from *N. incompta* to *N. pachyderma* to *T.*  
407 *quinqueloba* (Fig. 2C). This observation contradicts the current understanding of foraminiferal  
408 Na/Ca and salinity. In the Northern California Current region, salinity is found to generally  
409 increase with depth (Ortiz et al., 1996; Lane et al., 2023) suggesting that one should expect to find  
410 a decrease in Na/Ca values when comparing *N. incompta* to *N. pachyderma* to *T. quinqueloba*,

411 respectively. However, existing studies on the Na/Ca composition of several planktic foraminiferal  
412 taxa have found distinct differences in species-specific Na/Ca-salinity calibrations (e.g., see figure  
413 7 in Bertlich et al., 2021). For example, tropical species *Trilobatus sacculifer* incorporates Na at  
414 lower rates than polar taxon *N. pachyderma* (Allen et al., 2016; Bertlich et al., 2018; Mezger et al.,  
415 2018; Bertlich et al., 2021). Deviation from predicted interspecies differences in Na/Ca  
416 composition with inferred depth habitat is most likely due to interspecies differences in Na  
417 partitioning. Despite the three study species not following the predicted pattern of decreased Na/Ca  
418 content with shallower depth habitats, *T. quinqueloba* does exhibit the largest range of Na/Ca  
419 values, supporting the interpretation of a shallower depth habitat where salinity conditions can be  
420 highly variable. All three taxa show moderate positive relationships between Mg and Na in the  
421 whole-test and individual chamber scale, providing support for the interpretation of temperature  
422 and salinity changes with depth habitat being the dominant controls on the modulation of these  
423 elements, or, more generally, that Mg and Na incorporation mechanisms are potentially linked  
424 (**Fig. 8**).

425 Measurements of Sr/Ca show the same systematic increase in whole test values among the  
426 three taxa, in which the lowest values are found in *N. incompta* and the highest values are found  
427 in *T. quinqueloba* (**Fig. 2D**). Distributions of whole-test values of Sr/Ca appear to have an abrupt  
428 upper limit and a skewed distribution toward lower Sr/Ca values. This distribution architecture is  
429 likely reflective of maximum Sr content in the uppermost surface ocean with greater variability of  
430 Sr/Ca values found below the surface maximum. Recent compilations of Sr/Ca ratios in seawater  
431 have found that Sr/Ca decreases with depth in the Pacific surface ocean (Lebrato et al., 2020),  
432 although at much higher values than found in foraminiferal calcite (Allen et al., 2016), supporting  
433 the linkage between depth habitat and foraminiferal Sr/Ca in the study taxa.

434 Barium and manganese incorporation may be reflective of habitat, more specifically  
435 potential growth in deeper, oxygen-poor waters or the influence of calcification in microhabitats  
436 during periods of higher productivity (Fritz-Endres et al., 2022). Non-spinose taxa have been found  
437 living and calcifying in marine snow particles, a habitat favorable for the incorporation of Ba and  
438 Mn (Fehrenbacher et al., 2018; Greco et al., 2019; Fritz-Endres et al., 2022; Richey et al., 2022).  
439 It is also noteworthy that upon picking and removing tests for this study, it was not uncommon to  
440 find neogloboquadrinid specimens embedded in marine snow. Barite forms within marine snow  
441 particles suggesting that the Ba in solution within particulate organic matter is highly elevated  
442 compared to ambient seawater (Dymond and Collier, 1996; Griffith and Paytan, 2012; Martinez-  
443 Ruiz et al., 2019; Carter et al., 2020; Martinez-Ruiz et al., 2020). During periods of high export  
444 productivity, more tests of non-spinose taxa can be found living in the abundant marine snow,  
445 leading to increased barium incorporation into the calcite crystal lattice (Fritz-Endres et al., 2022).  
446 Marine snow also acts as an oxygen-depleted microhabitat (Alldredge and Cohen, 1987), where  
447 redox-sensitive elements such as Mn could be more readily incorporated into the foraminiferal  
448 crystal lattice. However, Mn/Ca composition of some species may also be controlled by upwelling  
449 intensity (Davis et al., 2020). The Northern California Current is known for its exceptional  
450 seasonal fluctuations in upwelling intensity, in which the upwelling of nutrient-rich deep waters  
451 often lead to increased productivity in the surface ocean. Periods of higher productivity associated  
452 with upwelling or other factors are also associated with expanded oxygen minimum zones, thus  
453 potentially creating oxygen-depleted habitats on both the micro- (i.e., marine snow) and macro-  
454 scales. While the exact control on Ba and Mn incorporation in these planktic foraminiferal taxa is  
455 not well-constrained, there appears to be a similar control on the whole-test Ba/Ca and Mn/Ca of  
456 the neogloboquadrinids that helps to explain the covariation of whole-test Ba/Ca and Mn/Ca (*N.*

457 *incompta*  $r^2 = 0.52$ ; *N. pachyderma*  $r^2 = 0.48$ ; **Fig. 8A, B**). This positive relationship between Ba  
458 and Mn is either similar (i.e., *N. incompta*,  $r^2 = 0.43$ ; **Fig. 8D**) or stronger (i.e., *N. pachyderma*,  $r^2 =$   
459 0.73; **Fig. 8E**) when examining TE covariance within individual chambers. Ba/Ca and Mn/Ca are  
460 not correlated on either the whole test or individual-chamber scale for *T. quinqueloba* (**Fig. 8C,**  
461 **F**). Whole-test Mn/Ca ratios of *T. quinqueloba* are in a similar range to the neogloboquadrinids,  
462 but the Ba/Ca ratios are considerably lower (**Fig. 2**). However, the Ba/Ca ratios recorded by *T.*  
463 *quinqueloba* are elevated compared to most other spinose taxa (cf. Hönisch et al., 2011). As a  
464 spinose foraminifera, *T. quinqueloba* is unlikely to live in a marine snow microhabitat, however  
465 the elevated Ba/Ca and Mn/Ca ratios of this species may also be influenced by upwelling (Davis  
466 et al., 2020).

467 Zinc incorporation has been found to be potentially reflective of productivity, oxygen  
468 concentration, and/or carbonate ion concentration in benthic foraminifera (Marchitto et al., 2000;  
469 Bryan and Marchitto, 2011; van Dijk et al., 2017), but is relatively understudied in planktic  
470 foraminifera (Marr et al., 2013; Davis et al., 2023; Hupp and Fehrenbacher, 2023) and thus  
471 incorporation mechanisms are poorly understood. Interspecies comparison of whole-test Zn/Ca  
472 values in this study furthers this conundrum, as whole-test Zn/Ca does not appear to follow any  
473 interspecies patterns similar to those described above for the other trace elements. However,  
474 *Neogloboquadrina pachyderma* exhibits distinctly higher Zn/Ca values than the two other study  
475 taxa and comparisons of average TEs in individual chambers yields a strong positive covariance  
476 between Mn/Ca and Zn/Ca incorporation ( $r^2 = 0.71$ ; **Fig. 8E**). Zinc is also a redox-sensitive  
477 element that can become soluble under low oxygen conditions during microbial sulfate reduction,  
478 and thus could also be reflective of calcification in a marine snow microhabitat (Davis et al., 2023).  
479 Interestingly, we do not find a similar strength of relationship in our *N. incompta* population (**Fig.**

480 8). It is possible that *N. pachyderma* has a greater tolerance for calcifying under low-oxygen  
481 conditions in which more time spent in such an environment could allow for growth under different  
482 stages of the redox ladder. Dissolved manganese can be produced in hypoxic environments under  
483 manganese reduction, whereas anoxic conditions are needed for sulfate reduction to occur in which  
484 dissolved zinc could become available for incorporation in the calcite crystal lattice.

485 Interspecies differences in trace element incorporation provide useful insight into the  
486 general ecology of the study taxa in this region. Although *N. incompta* and *N. pachyderma* are of  
487 the same genus, differences in preferred depth habitat and potentially differences in marine  
488 microhabitat growth regimes create distinct geochemical signatures among these two species.  
489 *Turborotalita quinqueloba* also has a clearly different geochemistry than the neogloboquadrinids.  
490 All three study taxa are asymbiotic; however, *T. quinqueloba* is a spinose species, whereas the  
491 neogloboquadrinids are non-spinose. Despite lacking algal photosymbionts, *T. quinqueloba*  
492 appears to have a shallower depth habitat (inferred from Mg/Ca ratios) compared to *N. incompta*  
493 or *N. pachyderma* (Simstich et al., 2003). *Turborotalita quinqueloba* also exhibits very low Ba/Ca  
494 whole test ratios (**Fig. 2**) and lacks significant relationships between Ba, Mn, and Zn (**Fig. 8**),  
495 suggesting that even though it shares the asymbiotic ecology of the neogloboquadrinids, its TE/Ca  
496 incorporation mechanisms may be more similar to symbiont-bearing spinose species.

497

498 ONTOGENETIC TRENDS IN TRACE ELEMENT INCORPORATION

499

500 Investigation of intratest variability can provide insight into ontogeny. For example,  
501 chamber-to-chamber variability may be reflective of changing depth habitat or oceanographic  
502 conditions during an individual's life cycle. While some chamber-to-chamber TE comparisons

503 show little to no difference between adjacent chambers, other chamber comparisons yield  
504 interesting results (**Fig. 3, 4, Table 3**). Here, chamber-to-chamber trends demonstrate that Mg  
505 content progressively decreases in sequentially younger chambers in all three study taxa (**Fig. 3B,**  
506 **E, H**). A similar chamber-to-chamber trend has been observed in *N. dutertrei* (Steinhardt et al.,  
507 2014; Fehrenbacher et al., 2018), but has not been previously recorded in other studies of *N.*  
508 *incompta* (Bolton, 2011; Davis et al., 2017) or *N. pachyderma* (Davis et al., 2017; Hupp and  
509 Fehrenbacher, 2023). Decreasing Mg could be reflective of an increasing depth of living habitat  
510 over the course of an individual's life (e.g., Meiland et al., 2021).

511 An enrichment in Na in the F0 chamber of *N. incompta* is evident (**Fig. 3C**). Recent studies  
512 have shown that addition of a gametogenic crust at the end of a planktic foraminifers' life can lead  
513 to an overall lower concentration in the whole-test trace element geochemistry (Eggins et al., 2003;  
514 Sadekov et al., 2005; Bolton and Marr, 2013; Davis et al., 2017; Hupp and Fehrenbacher, 2023).  
515 Examination of intrachamber TE composition supports this observation, where the inner lamellar  
516 calcite records higher TEs than the outer crust calcite (**Fig. 7**). The gametogenic crust tends to be  
517 relatively thin in the F0 chamber compared to other chambers in the outer whorl. Therefore, the  
518 Na geochemistry of the F0 chamber of crusted specimens can seem to be relatively enriched.  
519 Similar observations have been made for Na and Sr in populations of *N. pachyderma* collected  
520 from plankton tows in the Northern California Current (Hupp and Fehrenbacher, 2023).

521 Chamber-to-chamber trends in Sr/Ca vary markedly between species, with *N. incompta*  
522 exhibiting a decreasing trend with each additional chamber, *N. pachyderma* exhibiting an  
523 increasing trend, and *T. quinqueloba* showing no systematic trend (**Fig. 3, Table 3**), furthering the  
524 conundrum of what environmental parameter controls Sr incorporation (Lea et al., 1999; Martin  
525 et al., 1999; Stoll et al., 1999; Billups et al., 2004; Allen et al., 2016). Compilations of systematic

526 changes in Sr among adjacent chambers have revealed a wide range of intratest patterns (see Table  
527 1 in Davis et al., 2020). Since the study of Davis et al. (2020), Hupp and Fehrenbacher (2023)  
528 found little change in chamber-to-chamber Sr/Ca variability in a population of crusted *N.*  
529 *pachyderma*, aside from an enrichment in Sr in the F0 chamber. Conversely, prior study of the  
530 non-spinose taxa *Globorotaloides hexagonus*, known to inhabit oxygen minimum zones, has  
531 shown a distinct increase in Sr incorporation with the addition of each new chamber (Davis et al.,  
532 2023). Ratios of Sr/Ca have been shown to increase with increased temperature, salinity, and  
533 carbonate ion concentration in spinose tropical to subtropical species (Allen et al., 2016). However,  
534 systematic chamber-to-chamber fluctuations in Sr/Ca could directly mimic changes in water Sr/Ca,  
535 reflective of dynamic changes in depth habitat through ontogeny.

536 Chamber Ba/Ca and Mn/Ca comparisons in *N. pachyderma* and *T. quinqueloba* show  
537 distinct decreasing trends in TE incorporation with ontogeny, whereas *N. incompta* exhibits  
538 relatively consistent Ba and Mn composition among chambers in the final whorl (**Fig. 4**).  
539 Differences in Ba and Mn content among chambers in *N. pachyderma* may reflect movement into  
540 and out of a marine snow calcifying environment during the ontogeny of an individual, in which  
541 chambers recording higher Ba/Ca and Mn/Ca values signal calcification in a marine snow  
542 microenvironment, where conditions are conducive for elevated TEs (Alldredge and Cohen, 1987;  
543 Fritz-Endres et al., 2022; Richey et al., 2022); however, this explanation would not extend to  
544 explain similar trends found in *T. quinqueloba*. While we do not find any chamber-to-chamber  
545 gradient in the Ba/Ca and Mn/Ca composition of *N. incompta*, the larger range of Ba and Mn  
546 whole-test values suggests that they too prefer a marine snow microhabitat. However, the lack of  
547 chamber variability may imply that individuals either spend nearly all of their adult life calcifying  
548 in marine snow when it is present or all of their life outside of marine snow.

549 Interspecies differences in Zn/Ca chamber-to-chamber trends inspire more questions;  
550 Zn/Ca shows a clear increasing trend in progressively younger chambers of the  
551 neogloboquadrinids, but exhibits a decreasing trend, aside from the F0 chamber, in *T. quinqueloba*.  
552 These chamber-to-chamber observations could reflect changes in the carbonate ion concentration  
553 of the calcifying microhabitat (Marchitto et al., 2000; van Dijk et al., 2017), with potential  
554 differences between spinose and non-spinose taxa. However, the mechanism for producing such  
555 differences in  $[CO_3^{2-}]$  among spinose and non-spinose taxa is unclear, particularly because all taxa  
556 examined in this study are asymbiotic and thus changing pH conditions in the local  
557 microenvironment due to the photosynthesis of algal symbionts is not an explanation. Ratios of  
558 Zn/Ca have been suggested to be sensitive to oxygen concentrations (Davis et al., 2023), whereby  
559 incorporation of redox-sensitive Zn would increase when calcifying under low-oxygen conditions.  
560 This relationship has been observed in *Globorotaloides hexagonus*, a species commonly found in  
561 living in oxygen minimum zones (Davis et al., 2023). However, oxygen conditions are unlikely to  
562 change *systematically* throughout the ontogeny, and thus is unlikely to be the cause of chamber-  
563 to-chamber Zn variability in our study taxa. Zinc incorporation into planktic foraminiferal tests  
564 continues to be an enigma.

565 While the causes of ontogenetic trends in TE incorporation are difficult to constrain, it is  
566 interesting to consider the interspecies differences in such trends (**Table 3**). For example, one  
567 would expect to potentially find similarities in ontogenetic TE incorporation among taxa of the  
568 same genus, same spinose versus non-spinose affinity, or same symbiotic versus asymbiotic status.  
569 However, we find little consistencies or natural groupings in chamber-to-chamber TE  
570 incorporation among our study taxa aside from a consistent decrease in Mg incorporation through

571 ontogeny in all study taxa and similarities in systematic Zn incorporation among the  
572 neogloboquadrinids.

573 The range of intratest chamber measurements (e.g., **Fig. 5A**) can be used to identify more  
574 sporadic changes in environmental conditions throughout an individual's lifetime. For example,  
575 the intratest Ba/Ca range is highest in the neogloboquadrinids (**Fig. 5D**), where the intensity of  
576 export production may change during an individual's life or they may choose to only live a portion  
577 of their life calcifying in a marine snow microenvironment. Generally speaking, our data show that  
578 greater intratest Mg/Ca and Na/Ca variability is likely reflective of existence in a shallower depth  
579 habitat where temperature and salinity feature stronger fluctuations during a foraminifers' lifetime.  
580 This expected relationship tied to depth habitat is well-supported by the moderate positive  
581 correlations ( $0.47 \leq r^2 \leq 0.53$ ) found between whole-test and individual chamber Mg/Ca and Na/Ca  
582 in our study taxa (**Fig. 8**). However, while *T. quinqueloba* records overall higher whole-test Sr/Ca  
583 values than the other two study taxa, we find that their intratest ranges are comparable to the  
584 neogloboquadrinids likely due to 1) the environmental condition(s) that regulate Sr incorporation  
585 not changing much in their depth habitat over the short timescale of an individual foraminifers  
586 lifetime or 2) a general insensitivity to changing Sr incorporation. Greater intratest Ba/Ca and  
587 Mn/Ca in the neogloboquadrinids and, in the case of *N. pachyderma*, Zn/Ca variability is reflective  
588 of ontogeny lived in a deeper depth habitat, likely intersecting with the oxygen minimum zone,  
589 where fluctuations in export production and oxygen saturation modulate trace element  
590 incorporation into the foraminiferal crystal lattice over an individual's lifetime. Further  
591 characterization of the preferred depth habitat of these taxa in the Northern California Current will  
592 strengthen our understanding of these geochemical-environmental relationships.

593 In addition to examining chamber-to-chamber variability to delineate ontogenetic controls  
594 on trace element incorporation, we can investigate how trace elements vary with depth through the  
595 chamber wall. Previous studies have investigated either a few individuals or a single population of  
596 tests to examine how test trace element geochemistry may change with the addition of gametogenic  
597 crust (e.g., Table 1 in Hupp and Fehrenbacher, 2023). However, the dataset presented herein  
598 allows for investigation into the potential for consistent patterns in the trace element differences  
599 between the gametogenic crust and early ontogenetic calcite in individuals grown under a wide  
600 range of environmental conditions. Here we find that for all three study taxa and nearly all trace  
601 elements examined herein, the inner chamber wall (i.e., dominantly lamellar calcite) has  
602 consistently higher trace element compositions than the outer chamber wall, dominated by the  
603 gametogenic crust (**Fig. 7**). This observation indicates that the biomineralization pathway for  
604 precipitating the crust post- or syn-reproduction differs from the “normal” biomineralization  
605 mechanism used for growing a test throughout the vast majority of a foraminifer's life. While the  
606 gametogenic crusting process has been found to occur quickly (on the scale of hours to days; Bé,  
607 1980), it is possible that rapid calcification rates could be linked to lower TE incorporation.  
608 However, studies of symbiont-bearing planktic foraminifer have found that calcification rate does  
609 not typically exert a dominant control on trace element partitioning (Allen et al., 2016).  
610 Furthermore, if calcification rate were to exert a control on TE incorporation, one might expect to  
611 find higher TE compositions in calcite precipitated at higher rates, as commonly found with trace  
612 elements incorporated during inorganic calcite precipitation (e.g., Lorens, 1981; Mucci, 1988;  
613 Tesoriero and Pankow, 1996; Nehrke et al., 2007). Yet, the rapidly-precipitated gametogenic  
614 calcite exhibits consistently lower TE concentrations than the lamellar calcite among the study  
615 taxa, supporting the idea that calcification rate is unlikely to be the cause of such TE differences

616 throughout the chamber wall. More research is needed to better understand differences between  
617 and the motivation for these two modes of biomineralization.

618

## POTENTIAL VITAL EFFECTS AND PALEOPROXY CONSIDERATIONS

620

621 Constraining controls on interspecies, interspecimen, and intratest trace element variability  
622 is critical to refining our understanding and utilization of foraminifera as paleoproxy substrates.  
623 The data explored herein provide insight into several considerations for foraminiferal-based  
624 paleoreconstructions. Firstly, interspecies Ba/Ca differences and variability in the range of intratest  
625 Ba/Ca chamber values lend support for the recently proposed Ba/Ca-productivity proxy in non-  
626 spinose foraminifera (Fehrenbacher et al., 2018; Fritz-Endres et al., 2022; Richey et al., 2022). In  
627 both neogloboquadrinid species we find elevated whole-test Ba/Ca and a wide range of Ba/Ca  
628 intratest values, suggesting that some of our study specimens grew under higher productivity  
629 regimes. Thus far, positive relationships between Ba/Ca and export productivity have been  
630 documented in nonspinose taxa *Neogloboquadrina dutertrei*, *Pulleniatina obliquiloculata*, and  
631 *Globorotalia truncatulinoides* (Fehrenbacher et al., 2018; Fritz-Endres et al., 2022; Richey et al.,  
632 2022). This study suggests that a similar relationship may exist in *N. incompta* and *N. pachyderma*,  
633 as well. To identify periods of high productivity in the fossil record, Ba/Ca ratios should be  
634 measured in individual specimens from each time slice of interest to examine both the  
635 interspecimen and intraspecimen (e.g., in *N. pachyderma*) variability in a population. A similar  
636 approach has been taken with examining changes in populations of individual  $\delta^{18}\text{O}$  foraminiferal  
637 analyses to characterize changes El Niño intensity over time (Thirumalai et al., 2013).

638 Correlations between Ba/Ca and Mn/Ca (**Fig. 8**) are supportive of not only Ba as a  
639 productivity proxy, but also Mn as a potential dissolved oxygen proxy. Although research has

640 explored dissolved oxygen concentrations as the primary driver of Mn incorporation in benthic  
641 foraminifera, much less research has been done to identify such a relationship in planktic  
642 foraminifera (e.g., Marr et al., 2013; Davis et al., 2023). Here we show that Mn, and possibly Zn  
643 in the case of *N. pachyderma*, incorporation may be a direct result of redox conditions during  
644 calcification, and after further development could serve as critical proxies for reconstructing  
645 changes in oxygen minimum zone behavior and, when paired with Ba/Ca, particulate organic  
646 carbon flux to the seafloor.

647 The Na/Ca-paleosalinity proxy was recently proposed, and while field-based and culturing  
648 studies have produced promising results (Mezger et al., 2016; Bertlich et al., 2018; Dämmer et al.,  
649 2020; Bertlich et al., 2021), others are skeptical of the applicability of this ratio as a salinity proxy  
650 in deep time (Zhou et al., 2021; Gray et al., 2023). Although the data provided herein cannot settle  
651 this debate, these data contribute to fill the large data-gap in quantifying the Na/Ca composition of  
652 different planktic foraminiferal species collected from their natural living environment. Whole-  
653 test Na/Ca averages of our three study taxa ranged from approximately 10.7 to 41.7 mmol/mol  
654 (mean = 15.0 mmol/mol) for *N. incompta*, 11.3 to 25.1 mmol/mol (mean = 17.3 mmol/mol) for *N.*  
655 *pachyderma*, and 14.5 to 93.1 mmol/mol (mean = 24.3 mmol/mol) for *T. quinqueloba* (**Table 1**).  
656 These values are consistent with those that have been previously published on *N. pachyderma* from  
657 the Northern California Current region (Hupp and Fehrenbacher, 2023), but are higher than values  
658 reported from *N. pachyderma* collected from the Weddell Sea that were subsequently grown in  
659 culture (Bertlich et al., 2021). To our knowledge, these are the first Na/Ca ratios acquired from *N.*  
660 *incompta* and *T. quinqueloba* collected from plankton tows. In general, all published values of *N.*  
661 *pachyderma* are higher than those found in tropical spinose species (e.g., *T. sacculifer*, *G. ruber*;  
662 Mezger et al., 2016; Bertlich et al., 2018), suggesting that interspecies Na/Ca differences and Na

incorporation mechanisms are far from being understood. It is possible that elevated Na/Ca is associated with storage in NaOH-buffered preservatives (i.e., formalin) or exposure to NaOH-buffered solutions during the pre-analysis chemical cleaning procedure. However, Na/Ca data shown herein is consistent with that previously published by Hupp and Fehrenbacher (2023), in which all specimens were collected from “fresh” tows and were not exposed to preservatives. A constrained cleaning study is needed to determine the potential impact of using NaOH-buffered cleaning solutions and preservatives on individual specimens analyzed by laser ablation or solution-based ICP-MS. Regardless of this potential source of bias, we are confident in the relative differences found between species, between specimens, and within individual tests.

Contrasts in interspecies TE composition described herein appear to reflect differences in preferred habitat and therefore variation in calcification conditions. However, it is worth noting that interspecies differences are also in part controlled by the relative number of specimens collected from variable environmental regimes. Furthermore, it is recognized that individual species of foraminifera often require species-specific TE-environmental calibrations when developing paleoproxies; this is because each taxon may have different sensitivities to environmental conditions or variable approaches to calcification that produce taxon-specific partition coefficients. We can already identify some evidence of different sensitivities to trace element incorporation from the data presented herein. For example, *T. quinqueloba* does not exhibit covariance between many of the trace elements examined on the whole-test or individual chamber scale (**Fig. 8C, F**), suggesting that either 1) their trace element incorporation is less sensitive to calcifying under high productivity, low oxygen, or variable carbonate chemistry conditions and/or 2) their preferred habitat is relatively homogenous in these environmental conditions (aside from changes in temperature and salinity). We also find that *N. pachyderma*

686 shows a distinct Zn enrichment and strong positive correlation between Mn and Zn that is not  
687 found in either of the other study taxa (**Fig. 2, 8**), showing further taxon-specific patterns in trace  
688 element incorporation.

689 Herein we characterize the amount of chamber-to-chamber and general intratest variability  
690 with the aim of identifying vital effects that could influence the appropriateness of these polar to  
691 subpolar taxa for paleoreconstruction purposes. Chamber-to-chamber measurements show that on  
692 average individual chamber Mg/Ca only varies from the test average by <0.5 mmol/mol across the  
693 three study taxa (**Fig. 3**) with intratest ranges being < 1 mmol/mol for the neogloboquadrinids and  
694 < 2 mmol/mol for *T. quinqueloba* (**Fig. 5B**). Published calibrations for *N. pachyderma* and *N.*  
695 *incompta* (Davis et al., 2017) suggest that intratest variability of 1 mmol/mol would equate to a  
696 few degrees C depending on the location along the logarithmic Mg/Ca-temperature calibration  
697 curve. A Mg/Ca-temperature relationship has not been determined for *T. quinqueloba*.  
698 Measurements of Na/Ca indicate chamber measurements vary by < 1 mmol/mol across the three  
699 study taxa (**Fig. 3**) when compared to the test average and intratest ranges are found to be < 4  
700 mmol/mol for the neogloboquadrinids and < 10 mmol/mol for *T. quinqueloba* (**Fig. 5C**). Published  
701 Na/Ca-salinity calibrations are relatively limited and thus a difference of 1 to 10 mmol/mol could  
702 equate to a change of 2 to >10 psu. Strong species-specific (e.g., Bertlich et al., 2021) and  
703 instrument-specific (e.g., Gray et al., 2023) differences in Na/Ca make it difficult to constrain the  
704 impact of intratest differences in Na/Ca on paleoreconstructions at this time; however, further work  
705 illuminating the caveats of the proposed Na/Ca-salinity proxy will help to place these intratest  
706 differences into context in regards to their potential effect on salinity reconstructions. In general,  
707 intratest variability in Mg/Ca and Na/Ca is found to be greatest in *T. quinqueloba* of our three  
708 study taxa. Minimal intratest variability is found in Sr/Ca (<0.05 mmol/mol comparing chambers

709 to test average; ~1 mmol/mol intratest range) among the three study taxa (**Fig. 3, 5D**), and with  
710 the dominant control on Sr/Ca remaining elusive, we do not pinpoint a need for concern of  
711 ontogenetic influences on paleoreconstruction at this time.

712 When comparing chamber values to the test average for Ba/Ca, we find variability of < 2  
713  $\mu\text{mol/mol}$  (**Fig. 4**), and intratest ranges among all chambers in the outer whorl of < 5  $\mu\text{mol/mol}$   
714 (**Fig. 5E**). While Ba/Ca-export productivity calibrations have not been published for the study  
715 taxa, published calibrations of *Neogloboquadrina dutertrei* and *Pulleniatina obliquiloculata*  
716 suggest that differences of 2  $\mu\text{mol/mol}$  would result in a change of interpreted export productivity  
717 by  $\sim 0.5 \text{ mol C/m}^2/\text{yr}$  and a difference of 5  $\mu\text{mol/mol}$  could further offset export productivity  
718 estimates by up to approximately 2  $\text{mol C/m}^2/\text{yr}$  (Fritz-Endres et al., 2022). Individual chamber  
719 comparisons to test averages for Mn/Ca show offsets of < 3  $\mu\text{mol/mol}$  (**Fig. 4**) and intratest  
720 chamber ranges of < 7  $\mu\text{mol/mol}$  (**Fig. 5F**). Similar to Ba/Ca, calibrations between Mn/Ca and  
721 dissolved oxygen have not been published for these taxa. However, published calibrations between  
722 Mn/Ca and dissolved oxygen measured from the oxygen minimum zone-dwelling foraminifer  
723 *Globorotaloides hexagonus* indicate that intratest differences of this magnitude would likely have  
724 very little impact on dissolved oxygen reconstructions (Davis et al., 2023). Furthermore, studies  
725 linking Mn/Ca to upwelling variability found differences in Mn/Ca for specimens of *Globigerina*  
726 *ruber* (pink), *Orbulina universa*, and *Globorotalia menardii* grown during non-upwelling and  
727 upwelling times to be > 10  $\mu\text{mol/mol}$  on average (Davis et al., 2020). This further suggests that  
728 intratest Mn/Ca found in the three study taxa herein are unlikely to obscure primary environmental  
729 signals interpreted from whole-test values. As discussed, the primary control on Zn/Ca is  
730 ambiguous. Individual chamber values of Zn/Ca vary by <100  $\mu\text{mol/mol}$  when compared to the  
731 test average (**Fig. 4**). However, the intratest range of Zn/Ca values can reach concentrations up to

732 nearly 400  $\mu\text{mol/mol}$ . As our understanding of the primary controls of Zn/Ca evolves, so will the  
733 implications of these intratest differences in the three study taxa.

734 The magnitude of intratest variability described above could have impacts on Mg/Ca-  
735 temperature reconstructions, particularly for *T. quinqueloba*. Vital effects in the form of intratest  
736 differences could also impact Na/Ca-salinity reconstructions and Ba/Ca-export productivity  
737 reconstructions, though much more work is needed to refine our understanding of these potential  
738 proxies, particularly in the three taxa examined in this study. While the degree of intratest  
739 variability may be alarming, it is important to recognize that the depth habitat of an individual is  
740 not stagnant through ontogeny. Observations of population dynamics as a means to investigate  
741 reproductive strategies of planktic foraminifera living in the Red Sea have shown that individuals  
742 often deviate from a consistent depth habitat and that the depth habitat of a foraminifer changes  
743 throughout its lifetime (Meiland et al., 2021). However, statistically robust investigations of test  
744 populations show that the population, and thus the chemistry of their tests, will largely represent a  
745 consistent change in depth habitat during ontogeny. Therefore, intratest variability, such as that  
746 characterized in this study, is likely largely reflective of changes in habitat conditions or depth  
747 during an individuals' lifetime.

748 The data provided herein also lead to suggestions on how to approach LA-ICP-MS trace  
749 element analyses for the study taxa. Here we find that intratest trace element variability increases  
750 in tests with higher whole-test trace element values (**Table 2**). This pattern is observed as a  
751 moderate to strong positive relationship in all three taxa for most elements, aside from Sr/Ca.  
752 Therefore, when analyzing tests via LA-ICP-MS, it is imperative to conduct individual spot  
753 analyses in *all* chambers in the final whorl to determine your test average signal and not restrict

754 your analyses to the ultimate or penultimate chambers. Measuring all chambers in the final whorl  
755 will also allow for more direct comparison to solution-based data (Fehrenbacher et al., 2020).

756 Lastly, we have shown that for our three study taxa, there are predictable intrachamber  
757 differences where early ontogenetic calcite records higher trace element compositions than the  
758 gametogenic crust for nearly all trace elements (**Fig. 7**). Differences in trace element geochemistry  
759 between lamellar calcite and crust calcite are most apparent in the Mg/Ca and Na/Ca of all three  
760 taxa, as well as in the Ba/Ca composition of the neogloboquadrinids and the Zn/Ca composition  
761 of *T. quinqueloba* (**Fig. 7**). This observation has implications for how TE-environmental  
762 calibrations are constructed and how we analyze fossil specimens. For example, TE-environmental  
763 calibrations are often established from controlled culture experiments. However, tests grown in  
764 culture do not often develop a thick gametogenic crust (Davis et al., 2017; Fehrenbacher et al.,  
765 2017). We therefore echo the importance of following the recommendations and considerations  
766 brought forth by Davis et al. (2017) and Hupp and Fehrenbacher (2023), where either 1)  
767 calibrations linking TE composition to environmental variables must be established from only  
768 crusted specimens, to allow for a more direct comparison to the fossil record which is dominated  
769 by crusted specimens (Caron et al., 1990; Johnstone et al., 2010); or 2) test measurements for  
770 paleoreconstructions should use LA-ICP-MS to measure and isolate the trace element signature of  
771 the early ontogenetic calcite which more accurately records signals from the foraminifers living  
772 environment.

773

## 774 CONCLUSIONS

775

776 Here we explore interspecies, chamber-to-chamber, and intrachamber trace element  
777 variability in three subpolar to polar planktic foraminiferal taxa: *Neogloboquadrina incompta*, *N.*  
778 *pachyderma*, and *Turborotalita quinqueloba*. By examining large populations of thickly-calcified  
779 tests (i.e., dead upon collection) collected under a wide range of environmental conditions  
780 throughout the dynamic Northern California Current region, we investigate environmental versus  
781 biological controls on trace element incorporation. The three study taxa show distinctly different  
782 whole-test trace element values that are largely reflective of preferred living habitat. The whole-  
783 test Mg/Ca and Sr/Ca ratios are highest in *T. quinqueloba* and lowest in *N. incompta*, representing  
784 the shallowest and deepest depth habitats among the study taxa, respectively (Fig. 2).  
785 *Turborotalita quinqueloba* also exhibits wide distributions of whole-test averages and intratest  
786 ranges for Mg/Ca and Na/Ca, reflective of a shallow living depth with higher temperature and  
787 salinity variability over an individual's ontogeny (Figs. 2, 5). Whole-test and individual chamber  
788 Mg/Ca and Na/Ca compositions positively covary in all three study taxa further supporting their  
789 incorporation mechanisms being tied to living depth habitat. The neogloboquadrinids exhibit  
790 elevated Ba/Ca and Mn/Ca, and strong positive covariance between these TEs compared to *T.*  
791 *quinqueloba*, indicating that some individuals calcified in an oxygen-depleted marine snow  
792 microhabitat, likely during periods of elevated export production (Figs. 2, 5, 8). These  
793 observations provide support for using the Ba/Ca of these non-spinose foraminifers as a  
794 paleoproduction proxy and potentially Mn as a paleoredox proxy. Furthermore, strong  
795 correlations are found between Mn/Ca and Zn/Ca in *N. pachyderma*, suggesting that this species  
796 may be able to calcify more readily under variable redox conditions than the other study taxa in  
797 which hypoxic versus anoxic conditions allow for greater incorporation of Mn and Zn, respectively  
798 (Fig. 8E). Tests with higher whole-test trace element values tend to have greater intratest trace

799 element variability (**Table 2**) and the outer chamber walls composed of dominantly gametogenic  
800 crust, consistently record lower trace element compositions than the lamellar calcite of the inner  
801 chamber wall formed throughout most of a foraminifera's ontogeny; this pattern was found for  
802 nearly all trace elements in all three study taxa (**Fig. 7**). Intratest chamber-to-chamber variability  
803 can be quite high (**Figs. 3 – 5**), and thus laser-ablation ICP-MS studies of planktic foraminifera  
804 should always measure spot analyses of every chamber in the outer whorl. Lastly, results provide  
805 insight into the intratest trace element variability in subpolar and polar planktic foraminifera, and  
806 vast amounts of data to better understand controls on trace element incorporation in these  
807 understudied high-latitude taxa.

808

809

#### 810 ACKNOWLEDGEMENTS

811

812 This work was supported by an NSF OCE grant 2222365 to B.H. and J.F. and NSF OCE 2049143  
813 to J.F. This research was further supported by the NOAA Climate and Global Change Postdoctoral  
814 Fellowship Program to B.H., administered by UCAR's Cooperative Programs for the  
815 Advancement of Earth System Science (CPAESS) under award #NA21OAR4310383. Special  
816 thanks to J. Fisher for assistance in accessing archived tow samples and the staff at the Oregon  
817 State University Keck Collaboratory for Plasma Spectrometry.

818

#### 819 REFERENCES

820

821 Alldredge, A. L., and Cohen, Y., 1987, Can microscale chemical patches persist in the sea?

822 Microelectrode Study of Marine Snow, Fecal Pellets: Science, v. 235, p. 689–691, doi:  
823 [10.1126/science.235.4789.689](https://doi.org/10.1126/science.235.4789.689).

824 Allen, K. A., Hönisch, B., Eggins, S. M., Haynes, L. L., Rosenthal, Y., and Yu, J., 2016, Trace  
825 element proxies for surface ocean conditions: A synthesis of culture calibrations with  
826 planktic foraminifera: *Geochimica et Cosmochimica Acta*, v. 193, p. 197–221, doi:  
827 [10.1016/j.gca.2016.08.015](https://doi.org/10.1016/j.gca.2016.08.015).

828 Anand, P., Elderfield, H., and Conte, M. H., 2003, Calibration of Mg/Ca thermometry in  
829 planktonic foraminifera from a sediment trap time series: *Paleoceanography*, v. 18, p.  
830 2002PA000846, doi: [10.1029/2002PA000846](https://doi.org/10.1029/2002PA000846).

831 Ariwaka, R., 1983, Distribution and taxonomy of *Globigerina pachyderma* (Ehrenber) off the  
832 Sanriku Coast, Northeast Honshu, Japan: *Tohoku University Scientific Reports*, v. 53, p.  
833 103-157.

834 Barth, J. A., and Wheeler, P. A., 2005, Introduction to special section: Coastal Advances in Shelf  
835 Transport: *Journal of Geophysical Research: Oceans*, v. 110, p. 2005JC003124, doi:  
836 [10.1029/2005JC003124](https://doi.org/10.1029/2005JC003124).

837 Bauch, D., Erlenkeuser, H., Winckler, G., Pavlova, G., and Thiede, J., 2002, Carbon isotopes and  
838 habitat of polar planktic foraminifera in the Okhotsk Sea: the “carbonate ion effect” under  
839 natural conditions: *Marine Micropaleontology*, v. 45, p. 83-99.

840 Bé A. W. H., and Lott, L., 1964, Shell growth and structure of planktonic foraminifera: *Science*,  
841 v. 145, p. 823–824, doi:

842 Bé, A. W. H., 1980, Gametogenic calcification in a spinose planktonic foraminifer,  
843 *Globigerinoides sacculifer* (Brady): Marine Micropaleontology, v. 5, p. 283–310, doi:  
844 [10.1016/0377-8398\(80\)90014-6](https://doi.org/10.1016/0377-8398(80)90014-6).

845 Bertlich, J., Nürnberg, D., Hathorne, E. C., de Nooijer, L. J., Mezger, E. M., Kienast, M.,  
846 Nordhausen, S., Reichart, G.-J., Schönfeld, J., and Bijma, J., 2018, Salinity control on Na  
847 incorporation into calcite tests of the planktonic foraminifera *Trilobatus sacculifer* –  
848 evidence from culture experiments and surface sediments: Biogeosciences, v. 15, p. 5991–  
849 6018, doi: [10.5194/bg-15-5991-2018](https://doi.org/10.5194/bg-15-5991-2018).

850 Bertlich, J., Gussone, N., Berndt, J., Arlinghaus, H. F., and Dieckmann, G. S., 2021, Salinity  
851 effects on cultured *Neogloboquadrina pachyderma* (sinistral) from high latitudes: new  
852 paleoenvironmental insights: Geo-Marine Letters, v. 41, p. 2, doi: [10.1007/s00367-020-00677-1](https://doi.org/10.1007/s00367-020-00677-1).

853

854 Billups, K., Rickaby, R.E.M., and Schrag, D.P., 2004, Cenozoic pelagic Sr/Ca records:  
855 Exploring a link to paleoproductivity: Paleoceanography, v. 19, p. PA3005,  
856 doi:10.1029/2004PA001011.

857 Bolton, A., 2011, LA-ICP-MS Trace Element Analysis of Planktonic Foraminifera and  
858 Application to Marine Isotope Stage 31 in the Southwest Pacific Ocean. Thesis.

859 Bolton, A., and Marr, J. P., 2013, Trace element variability in crust-bearing and non crust-  
860 bearing *Neogloboquadrina incompta*, P–D intergrade and *Globoconella inflata* from the  
861 Southwest Pacific Ocean: Potential paleoceanographic implications: Marine  
862 Micropaleontology, v. 100, p. 21–33, doi: [10.1016/j.marmicro.2013.03.008](https://doi.org/10.1016/j.marmicro.2013.03.008).

863 Bonnin, E. A., Zhu, Z., Fehrenbacher, J. S., Russell, A. D., Hönisch, B., Spero, H. J., and  
864 Gagnon, A. C., 2019, Submicron sodium banding in cultured planktic foraminifera shells:  
865 *Geochimica et Cosmochimica Acta*, v. 253, p. 127–141, doi: [10.1016/j.gca.2019.03.024](https://doi.org/10.1016/j.gca.2019.03.024).

866 Boussetta, S., Bassinot, F., Sabbatini, A., Caillon, N., Nouet, J., Kallel, N., Rebaubier, H.,  
867 Klinkhammer, G., and Labeyrie, L., 2011, Diagenetic Mg-rich calcite in Mediterranean  
868 sediments: Quantification and impact on foraminiferal Mg/Ca thermometry: *Marine  
869 Geology*, v. 280, p. 195–204.

870 Branson, O., Read, E., Redfern, S. A. T., Rau, C., and Elderfield, H., 2015, Revisiting diagenesis  
871 on the Ontong Java Plateau: Evidence for authigenic crust precipitation in *Globorotalia  
872 tumida*: *Paleoceanography*, v. 30, p. 1490–1502, doi: [10.1002/2014PA002759](https://doi.org/10.1002/2014PA002759).

873 Branson, O., Fehrenbacher, J. S., Vetter, L., Sadekov, A. Y., Eggins, S. M., and Spero, H. J.,  
874 2019, LAtools: A data analysis package for the reproducible reduction of LA-ICPMS data:  
875 *Chemical Geology*, v. 504, p. 83–95, doi: [10.1016/j.chemgeo.2018.10.029](https://doi.org/10.1016/j.chemgeo.2018.10.029).

876 Bryan, S. P., and Marchitto, T. M., 2010, Testing the utility of paleonutrient proxies Cd/Ca and  
877 Zn/Ca in benthic foraminifera from thermocline waters: *Geochemistry, Geophysics,  
878 Geosystems*, v. 11, p. 2009GC002780, doi: [10.1029/2009GC002780](https://doi.org/10.1029/2009GC002780).

879 Caron, D. A., Roger Anderson, O., Lindsey, J. L., Faber, W. W., and Lin Lim, E. E., 1990,  
880 Effects of gametogenesis on test structure and dissolution of some spinose planktonic  
881 foraminifera and implications for test preservation: *Marine Micropaleontology*, v. 16, p. 93–  
882 116, doi: [10.1016/0377-8398\(90\)90031-G](https://doi.org/10.1016/0377-8398(90)90031-G).

883 Carter, S. C., Paytan, A., and Griffith, E. M., 2020, Toward an improved understanding of the  
884 marine barium cycle and the application of marine barite as a paleoproductivity proxy:  
885 *Minerals*, v. 10, p. 421, doi: [10.3390/min10050421](https://doi.org/10.3390/min10050421).

886 Dämmer, L. K., de Nooijer, L., Van Sebille, E., Haak, J. G., and Reichart, G.-J., 2020,  
887 Evaluation of oxygen isotopes and trace elements in planktonic foraminifera from the  
888 Mediterranean Sea as recorders of seawater oxygen isotopes and salinity: Climate of the  
889 Past, v. 16, p. 2401–2414, doi: [10.5194/cp-16-2401-2020](https://doi.org/10.5194/cp-16-2401-2020).

890 Davis, C. V., Fehrenbacher, J. S., Hill, T. M., Russell, A. D., and Spero, H. J., 2017,  
891 Relationships between temperature, pH, and crusting on Mg/Ca ratios in laboratory-grown  
892 *Neogloboquadrina* foraminifera: Paleoceanography, v. 32, p. 1137–1152, doi:  
893 [10.1002/2017PA003111](https://doi.org/10.1002/2017PA003111).

894 Davis, C. V., Fehrenbacher, J. S., Benitez-Nelson, C., and Thunell, R. C., 2020, Trace element  
895 heterogeneity across individual planktic foraminifera from the modern Cariaco Basin:  
896 Journal of Foraminiferal Research, v. 50, p. 204–218, doi: [10.2113/gsjfr.50.2.204](https://doi.org/10.2113/gsjfr.50.2.204).

897 Davis, C. V., Doherty, S., Fehrenbacher, J., and Wishner, K., 2023, Trace element composition  
898 of modern planktic foraminifera from an oxygen minimum zone: Potential proxies for an  
899 enigmatic environment: Frontiers in Marine Science, v. 10, p. 1145756, doi:  
900 [10.3389/fmars.2023.1145756](https://doi.org/10.3389/fmars.2023.1145756).

901 de Nooijer, L. J., Spero, H. J., Erez, J., Bijma, J., and Reichart, G. J., 2014, Biomineralization in  
902 perforate foraminifera: Earth-Science Reviews, v. 135, p. 48–58, doi:  
903 [10.1016/j.earscirev.2014.03.013](https://doi.org/10.1016/j.earscirev.2014.03.013).

904 de Nooijer, L. J., Brombacher, A., Mewes, A., Langer, G., Nehrke, G., Bijma, J., and Reichart,  
905 G.-J., 2017, Ba incorporation in benthic foraminifera: Biogeochemistry: Biomineralization,  
906 accessed February 27, 2024, at <https://bg.copernicus.org/preprints/bg-2017-45/bg-2017-45.pdf>.

908 Dymond, J., and Collier, R., 1996, Particulate barium fluxes and their relationships to biological  
909 productivity: Deep Sea Research Part II: Topical Studies in Oceanography, v. 43, p. 1283–  
910 1308, doi: [10.1016/0967-0645\(96\)00011-2](https://doi.org/10.1016/0967-0645(96)00011-2)

911 Edgar, K.M., Anagnostou, E., Pearson, P.N., and Foster, G.L., 2015, Assessing the impact of  
912 diagenesis on  $\delta^{11}\text{B}$ ,  $\delta^{13}\text{C}$ ,  $\delta^{18}\text{O}$ , Sr/Ca and B/Ca values in fossil planktic foraminiferal  
913 calcite: *Geochimica et Cosmochimica Acta*, v. 166, p. 189–209.

914 Eggins, S., De Deckker, P., and Marshall, J., 2003, Mg/Ca variation in planktonic foraminifera  
915 tests: implications for reconstructing palaeo-seawater temperature and habitat migration:  
916 *Earth and Planetary Science Letters*, v. 212, p. 291–306, doi: [10.1016/S0012-821X\(03\)00283-8](https://doi.org/10.1016/S0012-821X(03)00283-8).

918 Erez, J., 2003, The source of ions for biomineralization in foraminifera and their implications for  
919 paleoceanographic proxies: *Reviews in mineralogy and geochemistry*, v. 54, p. 115–149,  
920 doi: [10.2113/0540115](https://doi.org/10.2113/0540115)

921 [Evans, D., Müller, W., and Erez, J., 2018, Assessing foraminifera biomineralization models](#)  
922 [through trace element data of cultures under variable seawater chemistry: \*Geochimica et\*](#)  
923 [Cosmochimica Acta, v. 236, p. 198–217.](#)

924 Fehrenbacher, J. S., Russell, A. D., Davis, C. V., Gagnon, A. C., Spero, H. J., Cliff, J. B., Zhu,  
925 Z., and Martin, P., 2017, Link between light-triggered Mg-banding and chamber formation  
926 in the planktic foraminifera *Neogloboquadrina dutertrei*: *Nature Communications*, v. 8, p.  
927 15441, doi: [10.1038/ncomms15441](https://doi.org/10.1038/ncomms15441).

928 Fehrenbacher, J. S., Russell, A. D., Davis, C. V., Spero, H. J., Chu, E., and Hönisch, B., 2018,  
929 Ba/Ca ratios in the non-spinose planktic foraminifer *Neogloboquadrina dutertrei*: Evidence

930 for an organic aggregate microhabitat: *Geochimica et Cosmochimica Acta*, v. 236, p. 361–  
931 372, doi: [10.1016/j.gca.2018.03.008](https://doi.org/10.1016/j.gca.2018.03.008).

932 Fehrenbacher, J., Marchitto, T., and Spero, H. J., 2020, Comparison of laser ablation and  
933 solution-based ICP-MS results for individual foraminifer Mg/Ca and Sr/Ca analyses:  
934 *Geochemistry, Geophysics, Geosystems*, v. 21, p. e2020GC009254, doi:  
935 [10.1029/2020GC009254](https://doi.org/10.1029/2020GC009254).

936 Feinberg, L. R., and Peterson, W. T., 2003, Variability in duration and intensity of euphausiid  
937 spawning off central Oregon, 1996–2001: *Progress in Oceanography*, v. 57, p. 363–379,  
938 doi: [10.1016/S0079-6611\(03\)00106-X](https://doi.org/10.1016/S0079-6611(03)00106-X).

939 Fritz-Endres, T., Fehrenbacher, J. S., Russell, A. D., and Cynar, H., 2022, Increased productivity  
940 in the equatorial Pacific during the deglaciation inferred from the Ba/Ca ratios of non-  
941 spinose planktic foraminifera: *Paleoceanography and Paleoclimatology*, v. 37, p.  
942 e2022PA004506, doi: [10.1029/2022PA004506](https://doi.org/10.1029/2022PA004506).

943 Gray, W. R., Evans, D., Henehan, M., Weldeab, S., Lea, D. W., Müller, W., and Rosenthal, Y.,  
944 2023, Sodium incorporation in foraminiferal calcite: An evaluation of the Na/Ca salinity  
945 proxy and evidence for multiple Na-bearing phases: *Geochimica et Cosmochimica Acta*, v.  
946 348, p. 152–164, doi: [10.1016/j.gca.2023.03.011](https://doi.org/10.1016/j.gca.2023.03.011).

947 Greco, M., Jonkers, L., Kretschmer, K., Bijma, J., and Kucera, M., 2019, Depth habitat of the  
948 planktonic foraminifera *Neogloboquadrina pachyderma* in the northern high latitudes  
949 explained by sea-ice and chlorophyll concentrations: *Biogeosciences*, v. 16, p. 3425–3437,  
950 doi: [10.5194/bg-16-3425-2019](https://doi.org/10.5194/bg-16-3425-2019).

951 Griffith, E. M., and Paytan, A., 2012, Barite in the ocean – occurrence, geochemistry and  
952 palaeoceanographic applications: *Sedimentology*, v. 59, p. 1817–1835, doi: [10.1111/j.1365-3091.2012.01327.x](https://doi.org/10.1111/j.1365-3091.2012.01327.x).

953

954 Hathorne, E. C., Alard, O., James, R. H., and Rogers, N. W., 2003, Determination of intratest  
955 variability of trace elements in foraminifera by laser ablation inductively coupled plasma-  
956 mass spectrometry: *Geochemistry, Geophysics, Geosystems*, v. 4, p. 2003GC000539, doi:  
957 [10.1029/2003GC000539](https://doi.org/10.1029/2003GC000539).

958 Hathorne, E. C., James, R. H., and Lampitt, R. S., 2009, Environmental versus biomineralization  
959 controls on the intratest variation in the trace element composition of the planktonic  
960 foraminifera *G. inflata* and *G. scitula*: *Paleoceanography*, v. 24, p. 2009PA001742, doi:  
961 [10.1029/2009PA001742](https://doi.org/10.1029/2009PA001742).

962 Holland, K., Branson, O., Haynes, L. L., Hönisch, B., Allen, K. A., Russell, A. D., Fehrenbacher,  
963 J. S., Spero, H. J., and Eggins, S. M., 2020, Constraining multiple controls on planktic  
964 foraminifera Mg/Ca: *Geochimica et Cosmochimica Acta*, v. 273, p. 116–136, doi:  
965 [10.1016/j.gca.2020.01.015](https://doi.org/10.1016/j.gca.2020.01.015).

966 Hönisch, B., Allen, K. A., Russell, A. D., Eggins, S. M., Bijma, J., Spero, H. J., Lea, D. W., and  
967 Yu, J., 2011, Planktic foraminifers as recorders of seawater Ba/Ca: *Marine  
968 Micropaleontology*, v. 79, p. 52–57, doi: [10.1016/j.marmicro.2011.01.003](https://doi.org/10.1016/j.marmicro.2011.01.003).

969 Hönisch, B., Allen, K. A., Lea, D. W., Spero, H. J., Eggins, S. M., Arbuszewski, J., deMenocal,  
970 P., Rosenthal, Y., Russell, A. D., and Elderfield, H., 2013, The influence of salinity on  
971 Mg/Ca in planktic foraminifers – Evidence from cultures, core-top sediments and  
972 complementary  $\delta^{18}\text{O}$ : *Geochimica et Cosmochimica Acta*, v. 121, p. 196–213, doi:  
973 [10.1016/j.gca.2013.07.028](https://doi.org/10.1016/j.gca.2013.07.028).

974 Hupp, B. N., and Fehrenbacher, J. S., 2023, Geochemical differences between alive, uncrusted  
975 and dead, crusted Shells of *Neogloboquadrina pachyderma* : Implications for  
976 paleoreconstruction: Paleoceanography and Paleoclimatology, v. 38, p. e2023PA004638,  
977 doi: [10.1029/2023PA004638](https://doi.org/10.1029/2023PA004638).

978 Jochum, K. P., Weis, U., Stoll, B., Kuzmin, D., Yang, Q., Raczek, I., Jacob, D. E., Stracke, A.,  
979 Birbaum, K., Frick, D. A., Günther, D., and Enzweiler, J., 2011, Determination of  
980 Reference Values for NIST SRM 610–617 Glasses Following ISO Guidelines:  
981 Geostandards and Geoanalytical Research, v. 35, p. 397–429, doi: [10.1111/j.1751-908X.2011.00120.x](https://doi.org/10.1111/j.1751-982 908X.2011.00120.x).

983 Johnstone, H. J. H., Schulz, M., Barker, S., and Elderfield, H., 2010, Inside story: An X-ray  
984 computed tomography method for assessing dissolution in the tests of planktonic  
985 foraminifera: Marine Micropaleontology, v. 77, p. 58–70, doi:  
986 [10.1016/j.marmicro.2010.07.004](https://doi.org/10.1016/j.marmicro.2010.07.004).

987 Katz, M. E., Cramer, B. S., Franzese, A., Honisch, B., Miller, K. G., Rosenthal, Y., and Wright,  
988 J. D., 2010, Traditional and emerging geochemical proxies in foraminifera: The Journal of  
989 Foraminiferal Research, v. 40, p. 165–192, doi: [10.2113/gsjfr.40.2.165](https://doi.org/10.2113/gsjfr.40.2.165).

990 Kirincich, A. R., Barth, J. A., Grantham, B. A., Menge, B. A., and Lubchenco, J., 2005, Wind-  
991 driven inner-shelf circulation off central Oregon during summer: Journal of Geophysical  
992 Research: Oceans, v. 110, p. 2004JC002611, doi: [10.1029/2004JC002611](https://doi.org/10.1029/2004JC002611).

993 Kniskern, T. A., Warrick, J. A., Farnsworth, K. L., Wheatcroft, R. A., and Goñi, M. A., 2011,  
994 Coherence of river and ocean conditions along the US West Coast during storms:  
995 Continental Shelf Research, v. 31, p. 789–805, doi: [10.1016/j.csr.2011.01.012](https://doi.org/10.1016/j.csr.2011.01.012).

996 Kozdon, R., Kelly, D.C., Kitajima, K., Strickland, A., Fournelle, J.H., and Valley, J.W., 2013, *In*  
997 *situ*  $\delta^{18}\text{O}$  and Mg/Ca analyses of diagenetic and planktic foraminiferal calcite preserved in a  
998 deep-sea record of the Paleocene-Eocene thermal maximum: *Paleoceanography*, v. 28, p. 1-  
999 12, doi:10.1002/palo.20048

1000 Lane, M. K., Fehrenbacher, J. S., Fisher, J. L., Fewings, M. R., Crump, B. C., Risien, C. M.,  
1001 Meyer, G. M. L., and Schell, F., 2023, Planktonic foraminiferal assemblages reflect  
1002 warming during two recent mid-latitude marine heatwaves: *Frontiers in Marine Science*, v.  
1003 10, p. 1155761, doi: [10.3389/fmars.2023.1155761](https://doi.org/10.3389/fmars.2023.1155761).

1004 Lea, D. W., Mashiotta, T. A., and Spero, H. J., 1999, Controls on magnesium and strontium  
1005 uptake in planktonic foraminifera determined by live culturing: *Geochimica et*  
1006 *Cosmochimica Acta*, v. 63, p. 2369–2379, doi: [10.1016/S0016-7037\(99\)00197-0](https://doi.org/10.1016/S0016-7037(99)00197-0).

1007 Lebrato M., et al., 2020, Global variability in seawater Mg:Ca and Sr:Ca ratios in the modern  
1008 ocean: *Proceedings of the National Academy of Sciences*, v. 117, 22281-22292, doi:  
1009 10.1073/pnas.1918943117.

1010 Longerich, H. P., Jackson, S. E., and Günther, D., 1996. Inter-laboratory note. Laser ablation  
1011 inductively coupled plasma mass spectrometric transient signal data acquisition and analyte  
1012 concentration calculation: *Journal of Analytical Atomic Spectrometry*, v. 1, p. 899–904, doi:  
1013 [10.1039/ja9961100899](https://doi.org/10.1039/ja9961100899)

1014 Lorens, 1981, Sr, Cd, Mn and Co distribution coefficients in calcite as a function of calcite  
1015 precipitation rate: *Geochimica et Cosmochimica Acta*, v. 45, p. 553-561, doi:  
1016 10.1016/0016-7037(81)90188-5.

1017 Marchitto, T. M., Curry, W. B., and Oppo, D. W., 2000, Zinc concentrations in benthic  
1018 foraminifera reflect seawater chemistry: *Paleoceanography*, v. 15, p. 299–306, doi:  
1019 [10.1029/1999PA000420](https://doi.org/10.1029/1999PA000420).

1020 Marr, J. P., Carter, L., Bostock, H. C., Bolton, A., and Smith, E., 2013, Southwest Pacific Ocean  
1021 response to a warming world: Using Mg/Ca, Zn/Ca, and Mn/Ca in foraminifera to track  
1022 surface ocean water masses during the last deglaciation: *Paleoceanography*, v. 28, p. 347–  
1023 362, doi: [10.1002/palo.20032](https://doi.org/10.1002/palo.20032).

1024 Martínez-Botí, M. A., Mortyn, P. G., Schmidt, D. N., Vance, D., and Field, D. B., 2011, Mg/Ca  
1025 in foraminifera from plankton tows: Evaluation of proxy controls and comparison with core  
1026 tops: *Earth and Planetary Science Letters*, v. 307, p. 113–125, doi:  
1027 [10.1016/j.epsl.2011.04.019](https://doi.org/10.1016/j.epsl.2011.04.019).

1028 Martinez-Ruiz, F., Paytan, A., Gonzalez-Muñoz, M. T., Jroundi, F., Abad, M. M., Lam, P. J.,  
1029 Bishop, J. K. B., Horner, T. J., Morton, P. L., and Kastner, M., 2019, Barite formation in the  
1030 ocean: Origin of amorphous and crystalline precipitates: *Chemical Geology*, v. 511, p. 441–  
1031 451, doi: [10.1016/j.chemgeo.2018.09.011](https://doi.org/10.1016/j.chemgeo.2018.09.011).

1032 Martinez-Ruiz, F., Paytan, A., Gonzalez-Muñoz, M. T., Jroundi, F., Abad, M. M., Lam, P. J.,  
1033 Horner, T. J., and Kastner, M., 2020, Barite precipitation on suspended organic matter in the  
1034 mesopelagic zone: *Frontiers in Earth Science*, v. 8, p. 567714, doi:  
1035 [10.3389/feart.2020.567714](https://doi.org/10.3389/feart.2020.567714).

1036 Mashiotta, T. A., Lea, D. W., and Spero, H. J., 1999, Glacial–interglacial changes in  
1037 Subantarctic sea surface temperature and  $\delta^{18}\text{O}$ -water using foraminiferal Mg: *Earth and*  
1038 *Planetary Science Letters*, v. 170, p. 417–432, doi: [10.1016/S0012-821X\(99\)00116-8](https://doi.org/10.1016/S0012-821X(99)00116-8).

1039 Meilland, J., Siccha, M., Kaffenberger, M., Bijma, J., and Kucera, M., 2021, Population  
1040 dynamics and reproduction strategies of planktonic foraminifera in the open ocean:  
1041 Biogeosciences, v. 18, p. 5789–5809, doi: [10.5194/bg-18-5789-2021](https://doi.org/10.5194/bg-18-5789-2021).

1042 Mezger, E. M., de Nooijer, L. J., Boer, W., Brummer, G. J. A., and Reichart, G. J., 2016, Salinity  
1043 controls on Na incorporation in Red Sea planktonic foraminifera: Paleoceanography, v. 31,  
1044 p. 1562–1582, doi: [10.1002/2016PA003052](https://doi.org/10.1002/2016PA003052).

1045 Mezger, E.M., de Nooijer, L.J., Siccha, M., Brummer, G.J.A., Kucera, M., and Reichart, G.J.,  
1046 2018, Taphonomic and ontogenetic effects on Na/Ca and Mg/Ca in spinose planktonic  
1047 foraminifera from the Red Sea: Geochemistry, Geophysics, Geosystems, v. 19, p. 4174-  
1048 4194, doi: 10.1029/2018GC007852.

1049 Mucci, A., 1988, Manganese uptake during calcite precipitation from seawater: conditions  
1050 leading to the formation of a pseudokutnahorite: Geochimica et Cosmochimica Acta, v. 52,  
1051 p. 1859-1868.

1052 Multizta, S., Dürkoop, A., Hale, W., Wefer, G., and Stefan Niebler, H., 1997, Planktonic  
1053 foraminifera as recorders of past surface-water stratification: Geology, v. 25, p. 335, doi:  
1054 [10.1130/0091-7613\(1997\)025<0335:PFAROP>2.3.CO;2](https://doi.org/10.1130/0091-7613(1997)025<0335:PFAROP>2.3.CO;2).

1055 Nehrke, G., Reichart, G.J., Van Cappellen, P., Meile, C., and Bijma, J., 2007, Dependence of  
1056 calcite growth rate and Sr partitioning on solution stoichiometry: Geochimica et  
1057 Cosmochimica Acta, v. 71, p. 2240-2249.

1058 Niebler, H. S., Hubberten, H. W., and Gersonde, R. (1999). Oxygen isotope values of planktic  
1059 foraminifera: A tool for the reconstruction of surface water stratification. In G. Fischer, and  
1060 G. Wefer (eds.), Use of proxies in Paleoceanography: Examples from the South Atlantic  
1061 (pp. 165–189). Springer.

1062 Nürnberg, D., 1995, Magnesium in tests of *Neogloboquadrina pachyderma* sinistral from high  
1063 northern and southern latitudes: *Journal of Foraminiferal Research*, v. 25, p. 350-368.

1064 Nürnberg, D., Bijma, J., and Hemleben, C., 1996, Assessing the reliability of magnesium in  
1065 foraminiferal calcite as a proxy for water mass temperatures: *Geochimica et Cosmochimica  
1066 Acta*, v. 60, p. 803-814.

1067 Ortiz, J. D., and Mix, A. C., 1992, The spatial distribution and seasonal succession of planktonic  
1068 foraminifera in the California Current off Oregon, September 1987 – September 1988:  
1069 Geological Society, London, Special Publications, v. 64, p. 197–213, doi:  
1070 [10.1144/GSL.SP.1992.064.01.13](https://doi.org/10.1144/GSL.SP.1992.064.01.13).

1071 Ortiz, J.D., Mix, A.C., Rugh, W., Watkins, J.M., and Collier, R.W., 1996, Deep-dwelling  
1072 planktonic foraminifera of the northeastern Pacific Ocean reveal environmental control of  
1073 oxygen and carbon isotopic disequilibria: *Geochimica et Cosmochimica Acta*, v. 60, p.  
1074 4509-4523.

1075 Paulmier, A., and Ruiz-Pino, D., 2009, Oxygen minimum zones (OMZs) in the modern ocean:  
1076 *Progress in Oceanography*, v. 80, p. 113–128, doi: [10.1016/j.pocean.2008.08.001](https://doi.org/10.1016/j.pocean.2008.08.001).

1077 Pearson, P.N., and Kucera, M., 2018, Chapter 12: Taxonomy, biostratigraphy, and phylogeny of  
1078 Oligocene *Turborotalita* in Wade, B.S., Olsson, R.K., Pearson, P.N., Huber, B.T., and  
1079 Berggren, W.A. (eds.), 2018, *Atlas of Oligocene Planktonic Foraminifera*: Cushman  
1080 Foundation for Foraminiferal Research Volume 46

1081 Perlin, A., Moum, J. N., Klymak, J. M., Levine, M. D., Boyd, T., and Kosro, P. M., 2005, A  
1082 modified law-of-the-wall applied to oceanic bottom boundary layers: *Journal of  
1083 Geophysical Research: Oceans*, v. 110, p. 2004JC002310, doi: [10.1029/2004JC002310](https://doi.org/10.1029/2004JC002310).

1084 Pierce, S. D., Barth, J. A., Shearman, R. K., and Erofeev, A. Y., 2012, Declining Oxygen in the  
1085 Northeast Pacific: *Journal of Physical Oceanography*, v. 42, p. 495–501, doi: [10.1175/JPO-D-11-0170.1](https://doi.org/10.1175/JPO-D-11-0170.1).

1087 Rebotim, A., Voelker, A.H.L., Jonkers, L., Waniek, J.J., Meggers, H., Schiebel, R., Fraile, I.,  
1088 Schulz, M., and Kucera, M., 2017, Factors controlling the depth habitat of planktonic  
1089 foraminifera in the subtropical eastern North Atlantic: *Biogeosciences*, v. 14, p. 827-859,  
1090 doi:10.5194/bg-14-827-2017.

1091 Richey, J. N., Fehrenbacher, J. S., Reynolds, C. E., Davis, C. V., and Spero, H. J., 2022, Barium  
1092 enrichment in the non-spinose planktic foraminifer, *Globorotalia truncatulinoides*:  
1093 *Geochimica et Cosmochimica Acta*, v. 333, p. 184–199, doi: [10.1016/j.gca.2022.07.006](https://doi.org/10.1016/j.gca.2022.07.006).

1094 Russell, A. D., Hönisch, B., Spero, H. J., and Lea, D. W., 2004, Effects of seawater carbonate  
1095 ion concentration and temperature on shell U, Mg, and Sr in cultured planktonic  
1096 foraminifera: *Geochimica et Cosmochimica Acta*, v. 68, p. 4347–4361, doi:  
1097 [10.1016/j.gca.2004.03.013](https://doi.org/10.1016/j.gca.2004.03.013).

1098 Sadekov, A.Y., Eggins, S.M., and De Dekker, P., 2005, Characterization of Mg/Ca distributions  
1099 in planktonic foraminifera species by electron microprobe mapping: *Geochemistry,*  
1100 *Geophysics, Geosystems*, v. 6, p. Q12P06, doi:10.1029/2005GC000973.

1101 Sadekov, A. Yu., Eggins, S. M., Klinkhammer, G. P., and Rosenthal, Y., 2010, Effects of  
1102 seafloor and laboratory dissolution on the Mg/Ca composition of *Globigerinoides sacculifer*  
1103 and *Orbulina universa* tests — A laser ablation ICPMS microanalysis perspective: *Earth*  
1104 and *Planetary Science Letters*, v. 292, p. 312–324, doi: [10.1016/j.epsl.2010.01.039](https://doi.org/10.1016/j.epsl.2010.01.039).

1105 Schiebel, R., and Hemleben, C., 2017, *Planktic Foraminifers in the Modern Ocean*: Springer  
1106 Berlin Heidelberg, Berlin, Heidelberg.

1107 Simstich, J., Sarnthein, M., and Erlenkeuser, H., 2003, Paired  $\delta^{18}\text{O}$  signals of *Neogloboquadrina*  
1108 *pachyderma* (s) and *Turborotalita quinqueloba* show thermal stratification structure in  
1109 Nordic Seas: *Marine Micropaleontology*, v. 48, p. 107-125.

1110 Steinhardt, J., Cléroux, C., Ullgren, J., de Nooijer, L., Durgadoo, J. V., Brummer, G.-J., and  
1111 Reichart, G.-J., 2014, Anti-cyclonic eddy imprint on calcite geochemistry of several  
1112 planktonic foraminiferal species in the Mozambique Channel: *Marine Micropaleontology*,  
1113 v. 113, p. 20–33, doi: [10.1016/j.marmicro.2014.09.001](https://doi.org/10.1016/j.marmicro.2014.09.001).

1114 Steinhardt, J., de Nooijer, L. L. J., Brummer, G., and Reichart, G., 2015, Profiling planktonic  
1115 foraminiferal crust formation: *Geochemistry, Geophysics, Geosystems*, v. 16, p. 2409–  
1116 2430, doi: [10.1002/2015GC005752](https://doi.org/10.1002/2015GC005752).

1117 Stoll H.M., Schrag, D.P., and Clemens, S.C., 1999, Are seawater Sr/Ca variations preserved in  
1118 quaternary foraminifera?: *Geochimica et Cosmochimica Acta*, v. 63, p. 3535-3547.

1119 Taylor, B. J., Rae, J. W. B., Gray, W. R., Darling, K. F., Burke, A., Gersonde, R., Abelmann, A.,  
1120 Maier, E., Esper, O., and Ziveri, P., 2018, Distribution and ecology of planktic foraminifera  
1121 in the North Pacific: Implications for paleo-reconstructions: *Quaternary Science Reviews*, v.  
1122 191, p. 256–274, doi: [10.1016/j.quascirev.2018.05.006](https://doi.org/10.1016/j.quascirev.2018.05.006).

1123 Tesoriero, A.J., and Pankow, J.F., 1996, Solid solution partitioning of  $\text{Sr}^{2+}$ ,  $\text{Ba}^{2+}$ , and  $\text{Cd}^{2+}$  to  
1124 calcite: *Geochimica et Cosmochimica Acta*, v. 60, p. 1053-1063.

1125 Thirumalai, K., Partin, J. W., Jackson, C. S., and Quinn, T. M., 2013, Statistical constraints on El  
1126 Niño Southern Oscillation reconstructions using individual foraminifera: A sensitivity  
1127 analysis: *Paleoceanography*, v. 28, p. 401–412, doi: [10.1002/palo.20037](https://doi.org/10.1002/palo.20037).

1128 van Dijk, I., de Nooijer, L. J., and Reichart, G.-J., 2017, Trends in element incorporation in  
1129 hyaline and porcelaneous foraminifera as a function of  $p\text{CO}_2$ : Biogeosciences, v. 14, p.  
1130 497–510, doi: [10.5194/bg-14-497-2017](https://doi.org/10.5194/bg-14-497-2017).

1131 Watkins, C. S., Schmidt, M. W., and Hertzberg, J. E., 2021, Calibrating *Trilobatus sacculifer*  
1132 Na/Ca ratios from Atlantic core-tops as a proxy for sea surface salinity: Paleoceanography  
1133 and Paleoclimatology, v. 36, p. e2021PA004277, doi: [10.1029/2021PA004277](https://doi.org/10.1029/2021PA004277).

1134 Wit, J.C., de Nooijer, L.J., Wolthers, M., and Reichart, G.J., 2013, A novel salinity proxy based  
1135 on Na incorporation into foraminiferal calcite: Biogeosciences, v. 10, p. 6375-6387,  
1136 doi:10.5194/bg-10-6375-2013.

1137 Zhou, X., Rosenthal, Y., Haynes, L., Si, W., Evans, D., Huang, K.-F., Hönisch, B., and Erez, J.,  
1138 2021, Planktic foraminiferal Na/Ca: A potential proxy for seawater calcium concentration:  
1139 *Geochimica et Cosmochimica Acta*, v. 305, p. 306–322, doi: [10.1016/j.gca.2021.04.012](https://doi.org/10.1016/j.gca.2021.04.012).

1140

1141

1142

1143

1144 CAPTIONS

1145

1146 **Table 1.** Summary statistics of the populations of whole-shell TE/Ca values including number of  
1147 individuals measured, the mean, standard deviation, maximum and minimum values, the range,  
1148 and inter-quartile range. Note that these values were calculated to include outliers that are not  
1149 incorporated into the boxplots shown in Figure 2.

1151 **Table 2.** Covariance values ( $r^2$  and p-value) comparing the intratest range and intratest standard  
1152 deviation (SD) of a given TE/Ca to the whole-test average of that same TE/Ca, evaluated for  
1153 each of the study taxa. Asterisks indicate regressions where outliers were removed because they  
1154 had a strong bias on the covariance results.

1155

1156 **Table 3.** Summary of systematic chamber-to-chamber differences in TE/Ca composition with the  
1157 addition of each new chamber, as determined in this study and shown in Figures 3 and 4.

1158

1159 **Figure 1.** Bathymetric map of plankton tow sites (red circles) in the Northern California Current  
1160 off the coast of Washington (WA), Oregon (OR), and California (CA) in the United States. The  
1161 scale bar is in kilometers.

1162

1163 **Figure 2.** A) Schematic approach used to calculate the whole-test average for each individual  
1164 test. B-G) Violin plots with inset boxplots showing the distribution of whole-test TE/Ca values  
1165 for the study specimens belonging to the three study species (*N. incompta*, N = 92; *N.*  
1166 *pachyderma*, N = 55; and *T. quinqueloba*, N = 42) for B) Mg/Ca, C) Na/Ca, D) Sr/Ca, E) Ba/Ca,  
1167 F) Mn/Ca, and G) Zn/Ca. Foraminifer cartoon in panel A adapted from Hupp and Fehrenbacher  
1168 (2023).

1169

1170 **Figure 3.** A) Schematic approach used to normalize each chamber value (mean centered) to  
1171 account for growth under variable environmental conditions. The distribution normalized  
1172 chamber values (e.g.,  $F3_{\text{normalized}}$ ) for each chamber and taxon is shown in the remaining panels.  
1173 B-J) Violin plots with inset boxplots showing the distribution of TE/Ca averages for each

1174 individual chamber normalized to the test average TE/Ca composition for B-D) *N. incompta*, E-  
1175 G) *N. pachyderma*, and H-J) *T. quinqueloba*. Trace elements examined include Mg/Ca (panels B,  
1176 E, H), Na/Ca (panels C, F, I), and Sr/Ca (panels D, G, J). F0 refers to the youngest chamber in  
1177 the outer whorl, with progressively older chambers referred to as F1, F2, F3, and F4,  
1178 respectively. Numbers listed below each chamber along the x-axis indicate the number of  
1179 measurements contributing to the TE/Ca distributions associated with that chamber for each  
1180 taxon. Note the variable y-axes between panels.

1181

1182 **Figure 4.** Violin plots with inset boxplots showing the distribution of TE/Ca averages for each  
1183 individual chamber normalized to the test average TE/Ca composition for A-C) *N. incompta*, D-  
1184 F) *N. pachyderma*, and G-I) *T. quinqueloba*. Trace elements examined include Ba/Ca (panels A,  
1185 D, G), Mn/Ca (panels B, E, H), and Zn/Ca (panels C, F, I). F0 refers to the youngest chamber in  
1186 the outer whorl, with progressively older chambers referred to as F1, F2, F3, and F4,  
1187 respectively. Note that the same approach illustrated in Figure 3A was used to determine the  
1188 normalized chamber values plotted in these violin plots. Numbers listed below each chamber  
1189 along the x-axis indicate the number of measurements contributing to the TE/Ca distributions  
1190 associated with that chamber for each taxon. Note the variable y-axes between panels.

1191

1192 **Figure 5.** A) Schematic showing a hypothetical example of how the intratest range was  
1193 calculated for each individual test. The range was determined by subtracting the chamber with  
1194 the lowest average from the chamber with the highest average from the same test. B-G) Violin  
1195 plots with inset boxplots showing the intratest range of TE/Ca values measured in individual  
1196 tests of the three study taxa (*N. incompta*, N = 92; *N. pachyderma*, N = 55; and *T. quinqueloba*,

1197 N = 42). Trace elements examined include B) Mg/Ca, C) Na/Ca, D) Sr/Ca, E) Ba/Ca, F) Mn/Ca,  
1198 and G) Zn/Ca.

1199

1200 **Figure 6.** Schematic workflow illustrating how the spectrum from a single spot measurement can  
1201 be evaluated to assess intrachamber variability.

1202

1203 **Figure 7.** Comparison of averages of the inner 50% of each chamber wall spectrum to the outer  
1204 50% of the same test spectrum. The dashed line denotes the line of equality and the solid blue  
1205 line represents the trendline of the data. Data is shown for A-F) *N. incompta*, G-L) *N.*  
1206 *pachyderma*, and M-R) *T. quinqueloba*. Note the variable axes between panels.

1207

1208 **Figure 8.** Covariance matrices of whole-test average TE/Ca values (A-C) and covariance  
1209 matrices comparing the average TE composition of individual chambers(D-F). Panels shown are  
1210 separated by taxa for A and D) *N. incompta*, B and E) *N. pachyderma*, and C and F) *T.*  
1211 *quinqueloba*. Boxes are colored according to the strength and sign (blue = positive correlation,  
1212 red = negative correlation) of the  $R^2$  value, consistent with the scale to the right of each matrix.  
1213 Only correlations where the p-value is  $\leq 0.05$  are shown (i.e., blanks represent regressions where  
1214 p-values were  $> 0.05$ ).

1215

1216

1217

1218

1219

1220

**Table 1.**

Taxon	Statistic	Mg/Ca (mmol/mol)	Na/Ca (mmol/mol)	Sr/Ca (mmol/mol)	Ba/Ca ( $\mu$ mol/mol)	Mn/Ca ( $\mu$ mol/mol)	Zn/Ca ( $\mu$ mol/mol)
<i>N. incompta</i> (N = 92)	Mean	2.11	15.01	1.34	14.03	9.91	256.02
	Std Dev. ( $2\sigma$ )	2.10	6.85	0.15	38.79	17.49	445.53
	Max.	7.62	41.66	1.79	136.95	47.53	1353.77
	Min.	1.04	10.73	1.04	1.12	1.16	30.13
	Range	6.58	30.92	0.75	135.83	46.36	1323.64
	IQR	0.82	1.76	0.06	16.69	8.97	240.83
<i>N. pachyderma</i> (N = 55)	Mean	2.37	17.25	1.37	26.77	6.52	501.18
	Std Dev. ( $2\sigma$ )	1.66	5.74	0.16	152.07	15.67	641.16
	Max.	5.22	25.07	1.53	386.34	40.23	1587.37
	Min.	1.01	11.34	1.06	0.98	1.40	81.44
	Range	4.21	13.73	0.47	385.36	38.82	1505.93
	IQR	1.04	3.84	0.10	7.14	3.55	368.94
<i>T. quinqueloba</i> (N = 42)	Mean	2.88	24.28	1.43	10.87	8.83	370.85
	Std Dev. ( $2\sigma$ )	1.83	25.25	0.30	65.85	14.87	821.12
	Max.	5.32	93.07	1.55	191.91	40.77	2090.17
	Min.	1.64	14.54	0.59	1.32	2.86	93.40
	Range	3.68	78.53	0.95	190.59	37.90	1996.77
	IQR	1.17	8.45	0.07	2.54	5.15	309.79

1230

1231

**Table 2.**

TE/Ca	Intratest:	<i>N. incompta</i>		<i>N. pachyderma</i>		<i>T. quinqueloba</i>	
		r <sup>2</sup>	p-value	r <sup>2</sup>	p-value	r <sup>2</sup>	p-value
Mg/Ca	Range	0.67	2.2•10 <sup>-16</sup>	0.58	9.1•10 <sup>-12</sup>	0.18	0.0029
	SD	0.68	2.2•10 <sup>-16</sup>	0.58	9.1•10 <sup>-12</sup>	0.16	0.0057
Na/Ca	Range	0.53	2.2•10 <sup>-16</sup>	0.36	6.7•10 <sup>-7</sup>	0.32	5.8•10 <sup>-5</sup>
	SD	0.534	2.2•10 <sup>-16</sup>	0.36	8.3•10 <sup>-7</sup>	0.33	4.1•10 <sup>-5</sup>
Sr/Ca	Range	4.3•10 <sup>-4</sup>	0.31	0.0047*	0.27	-0.022*	0.66
	SD	7.7•10 <sup>-4</sup>	0.31	0.0099*	0.23	-0.025*	0.77
Ba/Ca	Range	0.64	2.2•10 <sup>-16</sup>	0.98	2.2•10 <sup>-16</sup>	0.75	6.9•10 <sup>-14</sup>
	SD	0.64	2.2•10 <sup>-16</sup>	0.99	2.2•10 <sup>-16</sup>	0.76	5.4•10 <sup>-14</sup>
Mn/Ca	Range	0.70	2.2•10 <sup>-16</sup>	0.41	7.2•10 <sup>-8</sup>	0.77	2.4•10 <sup>-14</sup>
	SD	0.72	2.2•10 <sup>-16</sup>	0.39	2.6•10 <sup>-7</sup>	0.78	4.1•10 <sup>-15</sup>
Zn/Ca	Range	0.37	6.6•10 <sup>-11</sup>	0.51	5.3•10 <sup>-10</sup>	0.62	3.3•10 <sup>-10</sup>
	SD	0.38	3.4•10 <sup>-11</sup>	0.49	1.6•10 <sup>-9</sup>	0.61	7.5•10 <sup>-10</sup>

1232

1233

1234

1235

1236

1237

1238

1239

1240

1241

1242

1243

1244

**Table 3.**

TE/Ca	<i>N. incompta</i>	<i>N. pachyderma</i>	<i>T. quinqueloba</i>
Mg/Ca	Decrease	Decrease	Decrease (except F4)
Na/Ca	Increase (F0 only)	--	--
Sr/Ca	Increase	Decrease	--
Ba/Ca	--	Decrease	Decrease
Mn/Ca	--	Decrease	Decrease
Zn/Ca	Increase	Increase	Decrease (except F0)

1245

1246

1247

1248

1249

1250

1251

1252

1253

1254

1255

1256

1257

1258

1259

1260

1261

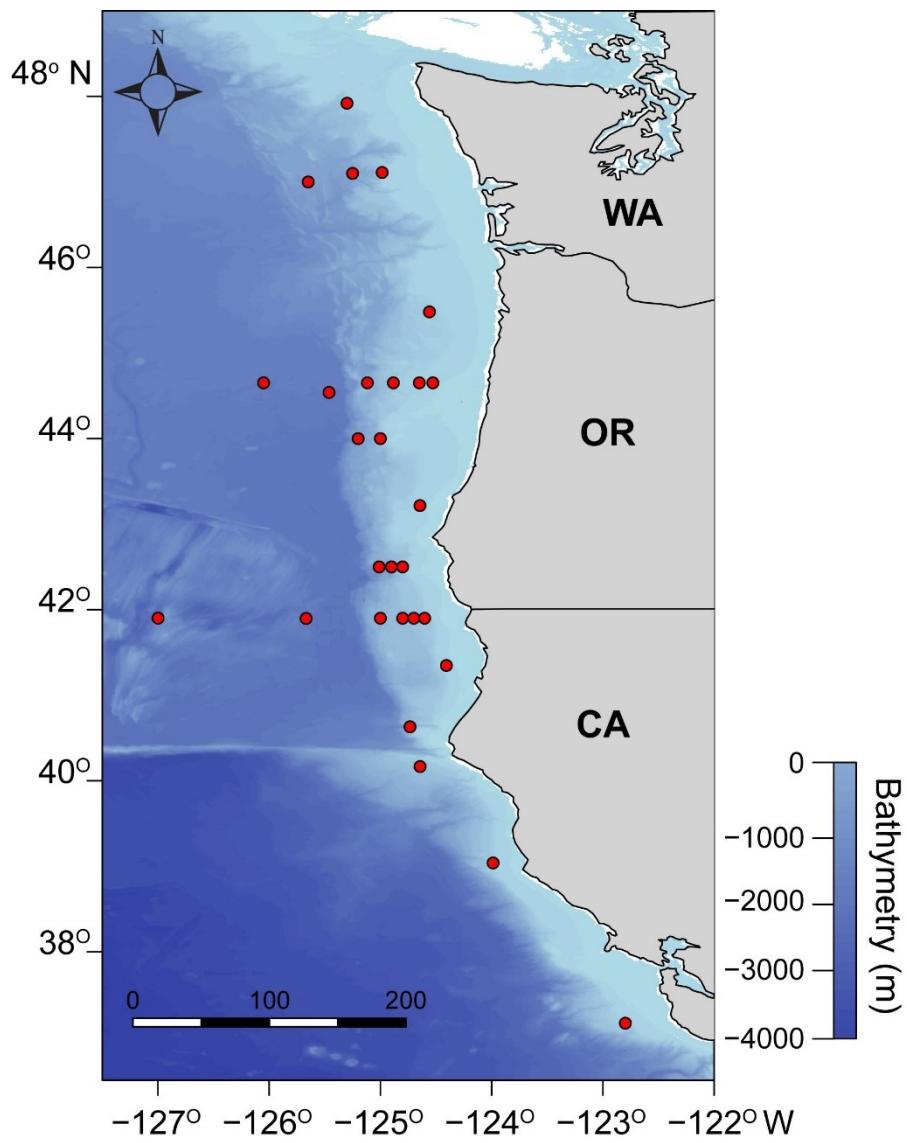
1262

1263

1264

1265

1266 **Figure 1**



1267

1268

1269

1270

1271

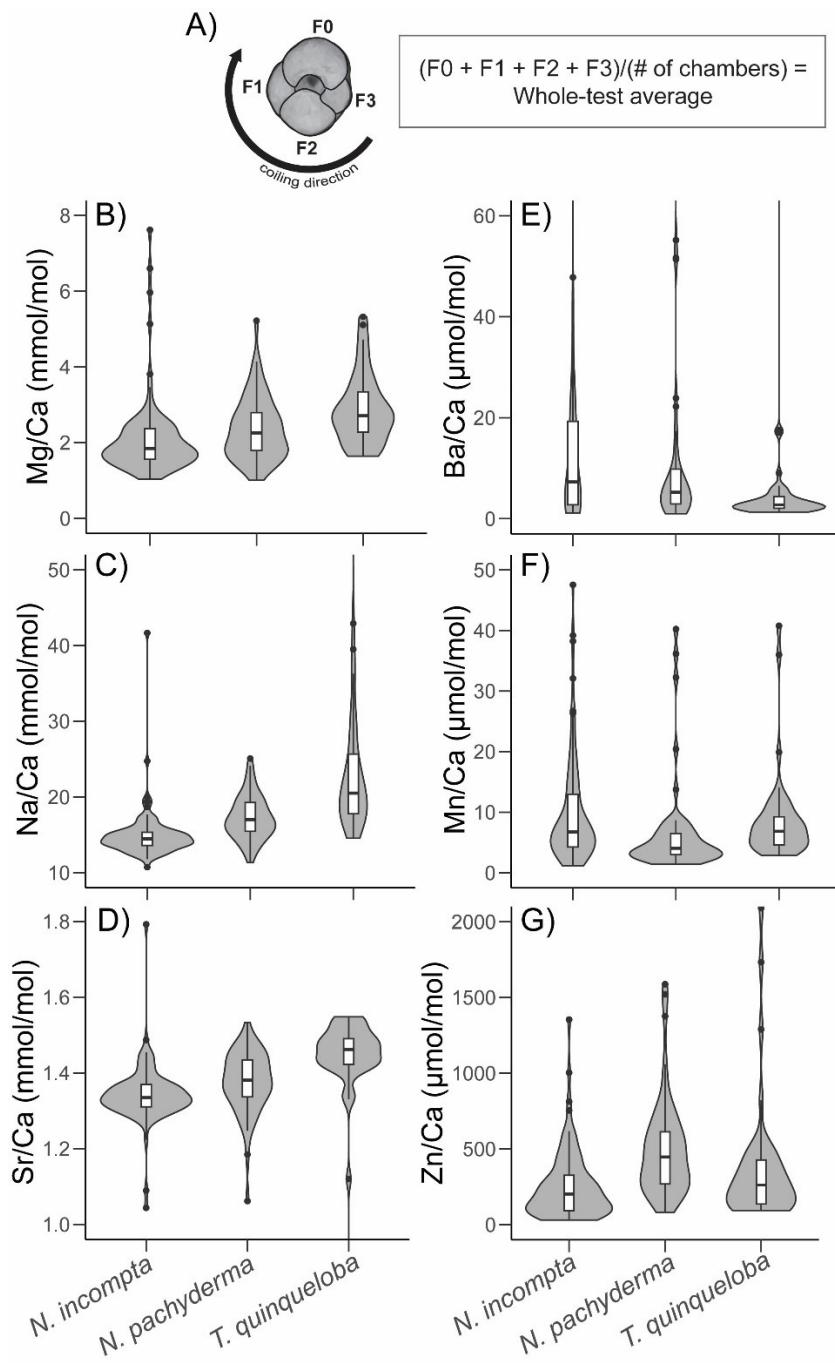
1272

1273

1274

1275

1276 **Figure 2**



1277

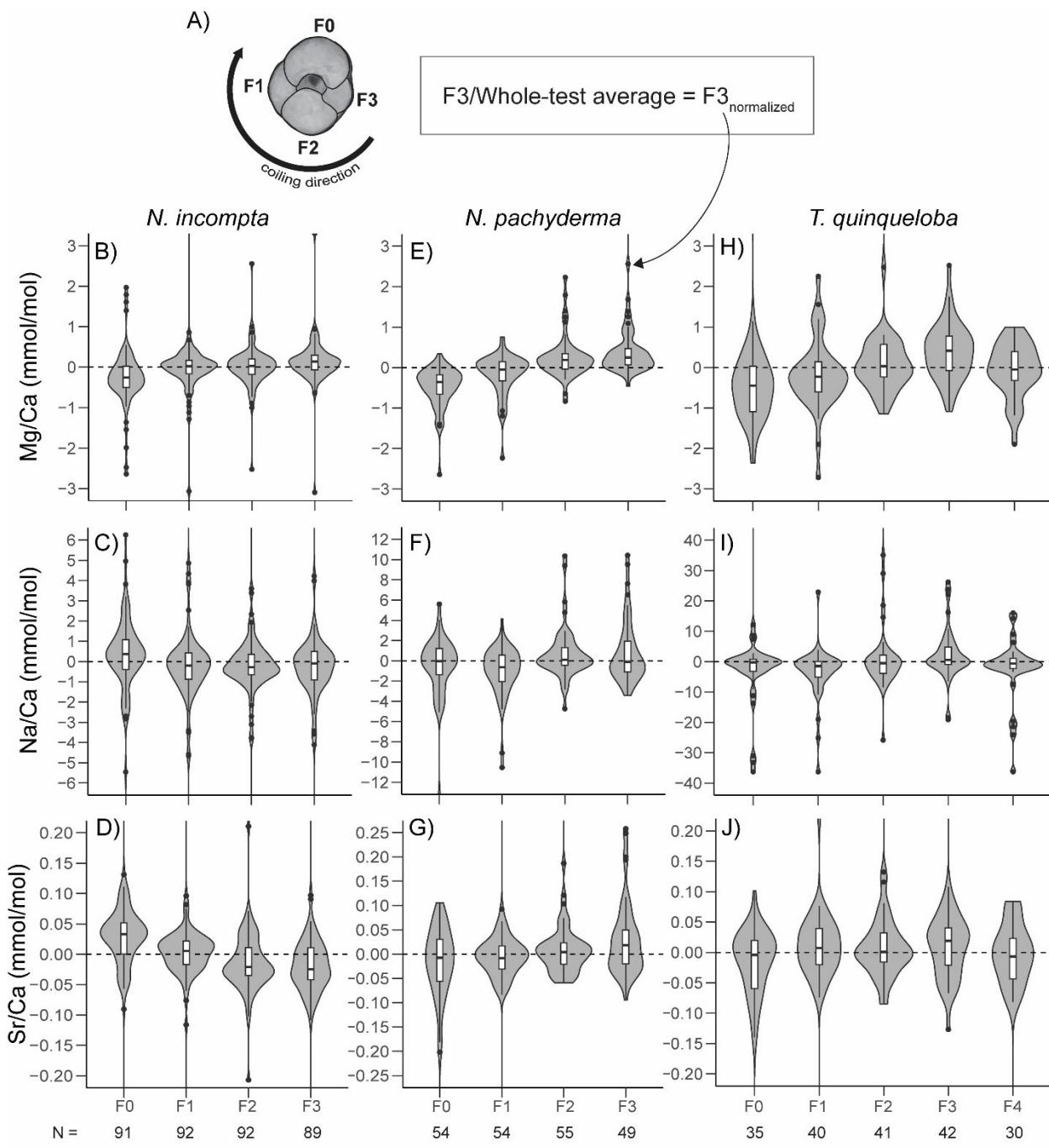
1278

1279

1280

1281

1282 **Figure 3.**



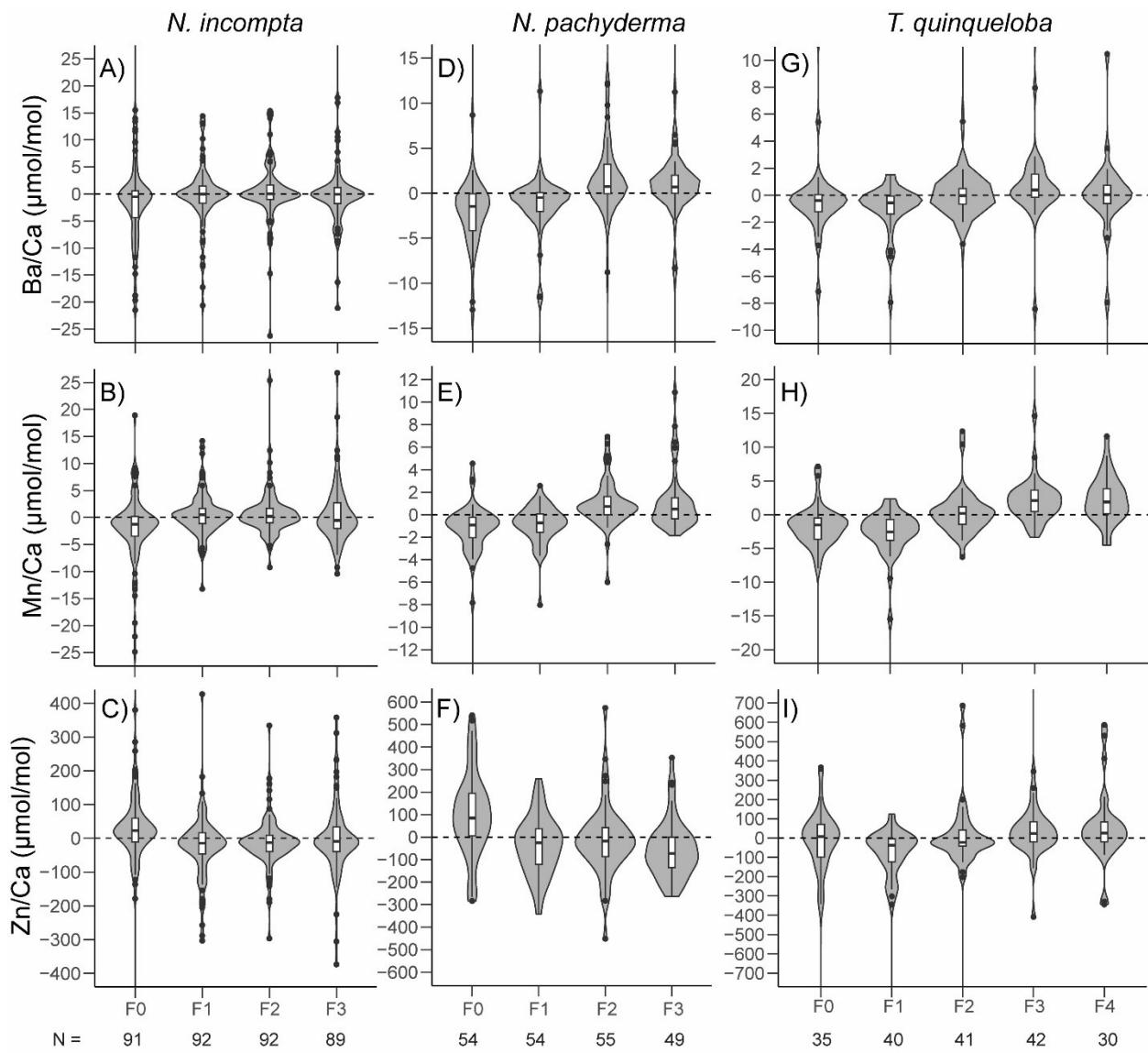
1284

1285

1286

1287

1288 **Figure 4.**



1289

1290

1291

1292

1293

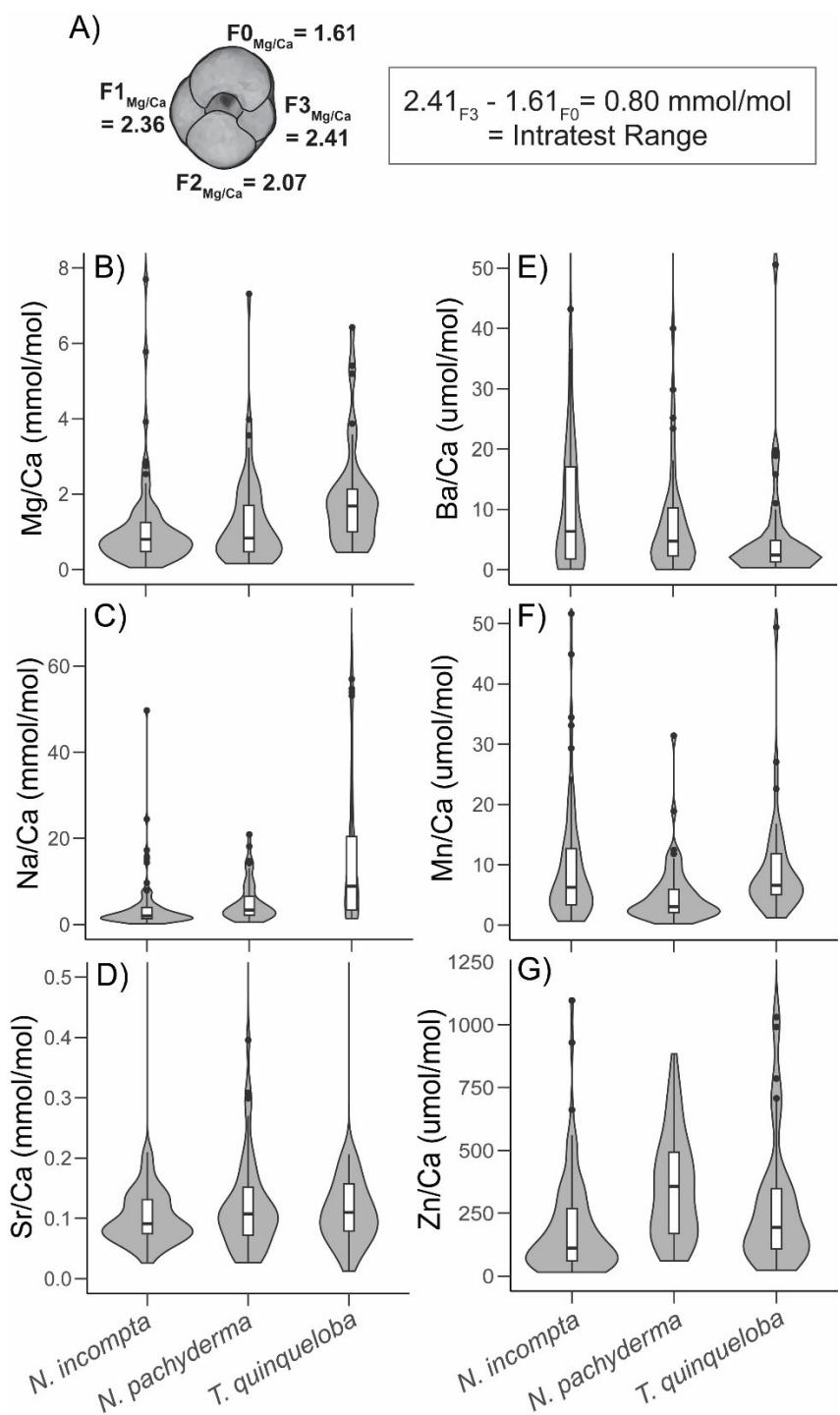
1294

1295

1296

1297

1298

1299 **Figure 5.**

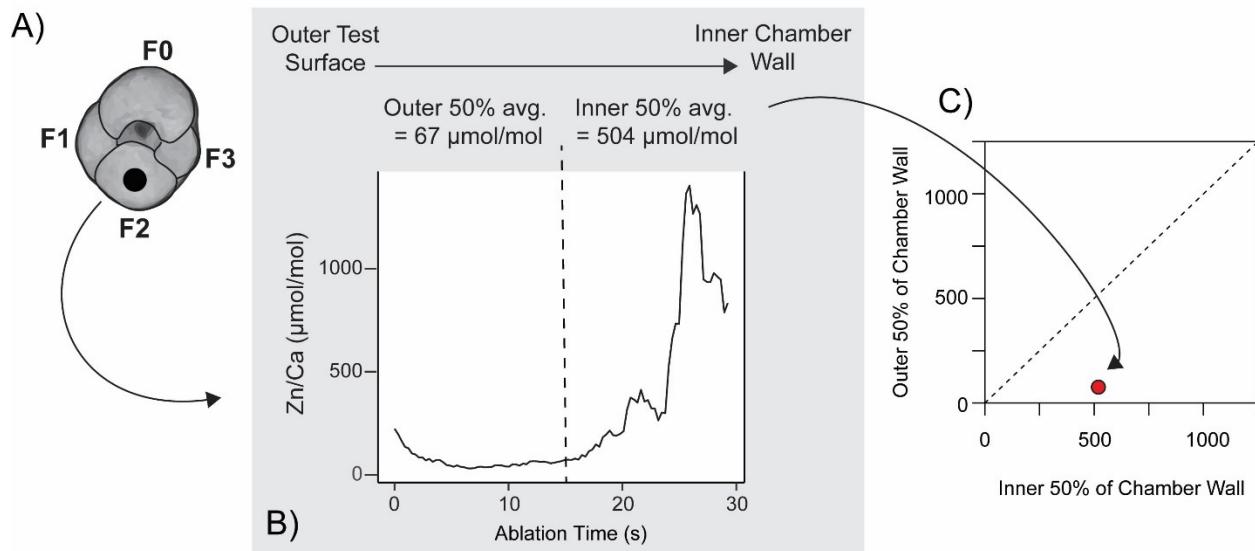
1300

1301

1302

1303 **Figure 6.**

Approach to Assessing Intrachamber Variability



1304

1305

1306

1307

1308

1309

1310

1311

1312

1313

1314

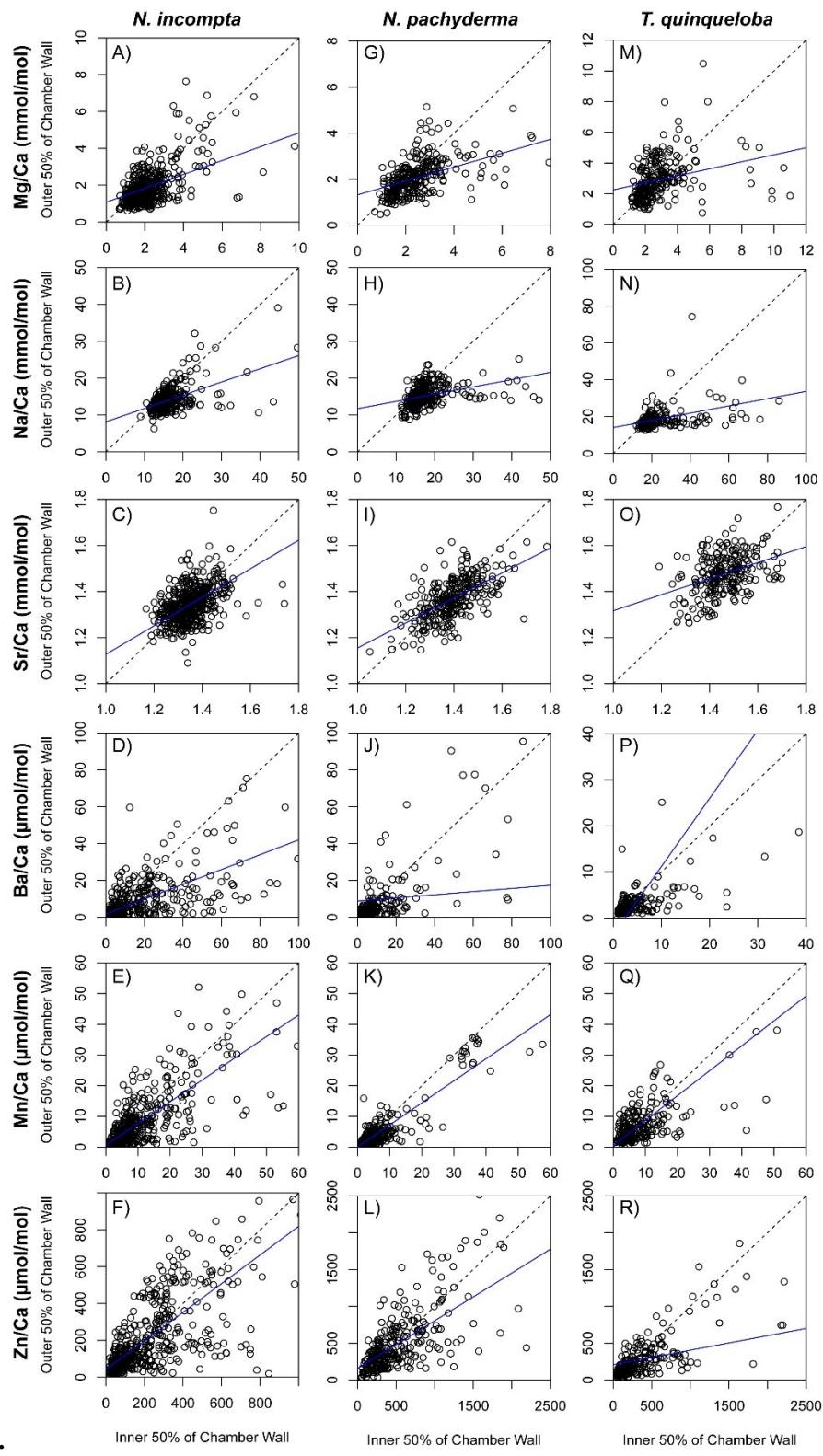
1315

1316

1317

1318

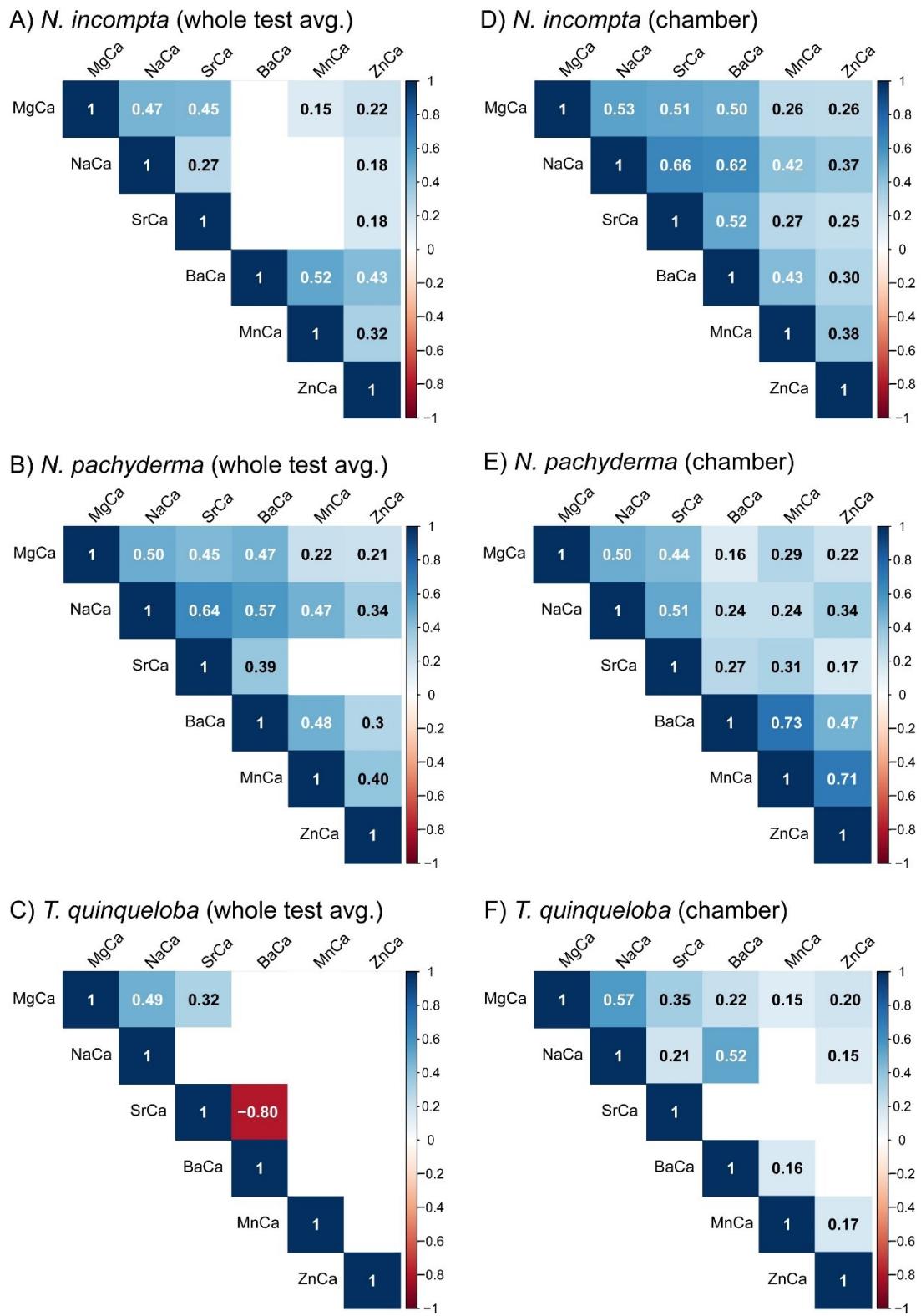
1319 Figure 7



1320

1321

1322 **Figure 8.**



1323

1324