Trapping of the Enoyl-Acyl Carrier Protein Reductase • Acyl Carrier Protein Interaction

Lorillee Tallorin,[†] Kara Finzel,[†] Quynh G. Nguyen, Joris Beld,[‡] James J. La Clair, and Michael D. Burkart*

Department of Chemistry and Biochemistry, University of California, San Diego, 9500 Gilman Drive, La Jolla, California 92093-0358,United States

Supporting Information Placeholder

ABSTRACT: An ideal target for metabolic engineering, fatty acid biosynthesis remains poorly understood on a molecular level. These carrier protein dependent pathways require fundamental protein • protein interactions to guide reactivity and processivity, and their control has become one of the major hurdles in successfully adapting these biological machines. Our laboratory has developed methods to prepare acyl carrier proteins (ACPs) loaded with substrate mimetics and crosslinkers to visualize and trap interactions with partner enzymes, and we continue to expand the tools for studying these pathways. We now describe application of the slow-onset, tight-binding inhibitor triclosan to explore the interactions between the type II fatty acid ACP from *Escherichia coli*, AcpP, and its corresponding enoyl-ACP reductase, FabI. We show that the AcpP • triclosan complex demonstrates nM binding, inhibits *in vitro* activity and can be used to isolate FabI in complex proteomes.

Machines involved in primary metabolism, particularly the production of fatty acids, have garnered increased attention over the last decade due to their potential for biofuel production and as antibiotic targets. These machines share a common choreography, whereby acetyl-coenzyme A (CoA) and malonyl-CoA are assembled sequentially in an iterative fashion to form elongated fatty acids. All intermediates are tethered to an acyl carrier protein (ACP),[1] which carries its cargo along the assembly line of modifying partner enzymes until release by a thioesterase or transfer *via* an acyltransferase. While this modular machinery appears ideal for metabolic engineering, many of the leading efforts, such as heterologous pathway assembly,[2] have been met with limited success. We and others have shown that this arises from our lack of understanding the protein • protein interactions that guide the processivity between the ACP and its associated partner enzymes (Fig. S1). Unfortunately, structural studies on these systems continue to pose challenges due to the transient nature of these interactions.

Our laboratory has developed a suite of tools to study

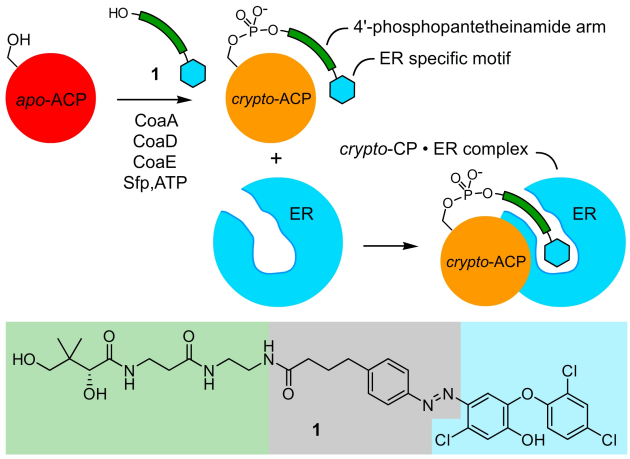


Figure 1. Developing an enoyl reductase (ER) probe **1** from triclosan (light blue), a linker (grey) and a pantetheine arm (green). An *apo*-ACP is chemoenzymatically modified with probe **1** yielding *crypto*-**1**-ACP, which contains the ER specific motif (blue hexagon). The resulting *crypto*-**1**-ACP can be used to bind to ER and trap the *crypto*-**1**-ACP • ER complex. A full depiction of the role of the ER in fatty acid biosynthesis is provided in Fig. S1.

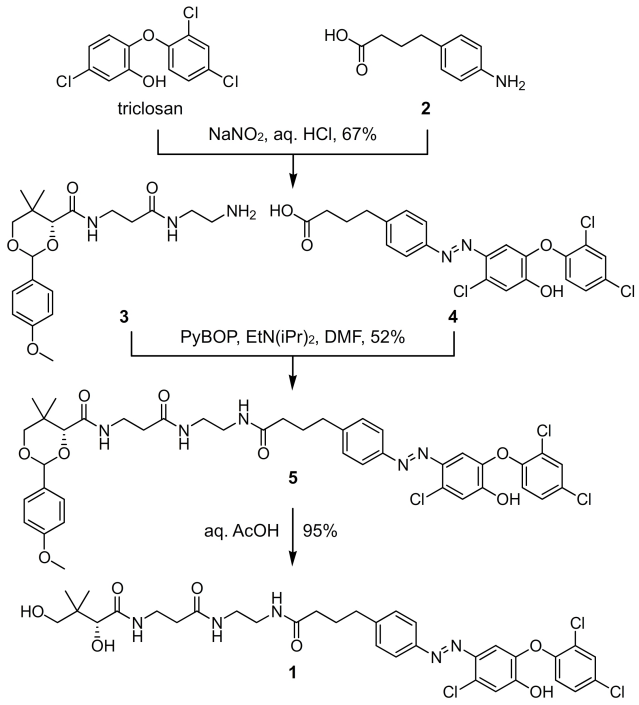
the interactivity between ACP and associated enzymes through the chemoenzymatic preparation of ACPs that bear a diversity of tethered functionality on their pantetheine terminus.[3] These synthetic probes can be converted with CoaA, CoaD and CoaE to the corresponding CoA analogs and *in situ* loaded onto the *apo*-ACP by the promiscuous 4'-phosphopantetheinyl transferase (PPTase) Sfp, resulting in a *crypto*-ACP bearing a terminal domain specific motif (Fig. S2).[4] We now describe expansion of this approach to study enoyl reductase (ER) domains (Fig. 1) using the enoyl-ACP reductase (FabI) from the *E. coli* fatty acid synthase as a model. Understanding these protein • protein interactions is key to engineering and drug discovery efforts.

FabI, a member of the short chain alcohol dehydrogenase/reductase (SDR) family, is responsible for the

reduction of *trans*-2-*enoyl*-AcpP to *acyl*-AcpP *via* its NADH cofactor.[5] It is also characterized as playing a determinant role in completing cycles during fatty acid biosynthesis in *E. coli*.[6] As one of the eight ACP • partner protein structures, a 2.7 Å structure of the AcpP • FabI complex has been solved.[7] This structure contains a tetrameric FabI bound with two *trans*-2-dodecenoyl thioester loaded AcpPs. However, due to the transient nature of this interaction, the interface between AcpP and FabI was not well resolved.

We first sought to leverage our previous work with ketosynthase (KS), thioesterase (TE) and dehydratase (DH) domains (Fig. S2),[2,8] and apply this approach to deliver ER domain probes. However, the design is complicated by the fact that ER enzymes typically do not involve covalent active site intermediates, but rather act *via* a NADH cofactor. We hypothesized that appending a tight non-covalent ER inhibitor[9] to the terminus of a pantetheinamide probe would, after chemoenzymatic loading, provide a *crypto*-ACP with sufficient binding to study ACP • ER interactions.[10] We began by exploring triclosan (Fig. 1), a broad-spectrum antibiotic and prototypical inhibitor for FabI,[11] which is characterized by slow-onset, tight-binding inhibition. Previous studies have accounted this strong inhibition to the stable ternary complex formed when triclosan noncovalently interacts with both FabI and NAD^+.[12,13] Additionally, it is suggested that AcpP interacts with basic residues adjacent to the FabI substrate binding loop. This loop is disordered in the FabI • cofactor binary complex and becomes ordered upon binding of NAD^+ and triclosan.[7,14]

Scheme 1. Synthesis of the triclosan probe 1.



We began synthesizing probe **1** (Scheme 1), which is comprised of a pantetheine portion, a linker and triclosan (Fig. 1). The linker was developed from literature precedent,[15] based on it being long enough to span the distance between the AcpP and deep pocket of FabI. As depicted in Scheme 1, probe **1** was prepared in three steps from triclosan, 4-aminophenylbutyric acid (**2**) and amine **3**. The synthesis began by forming a diazonium salt from **2** and $NaNO_2$ in the presence of HCl, which then was coupled *in situ* to triclosan *via* an electrophilic aromatic substitution. The resulting ~5:1 mixture of *trans*- to *cis*-azoacids **4**[15] was coupled with **3** to yield **5** in 52% yield. Samples of probe **1** were achieved at 35% overall yield from triclosan after deprotection of **5** in aq. AcOH.

Our biochemical studies began by evaluating the inhibition of FabI. We found that **1** had an IC_{50} value of 49.3 ± 0.2 µM (Fig. S3), which was 1,000-fold greater (reduced affinity) than triclosan (IC_{50} value of ~0.04 µM).[13] While this activity was less than desired, the slow off-rate associated with triclosan may still allow it to sufficiently trap ACP • ER complexes. Hence, we turned our attention to preparation of the corresponding *crypto*-**1**-AcpP.

Recombinant CoaA, CoaD, and CoaE were utilized to convert probe **1** into the corresponding CoA analog, which was used to post-translationally modify AcpP *in situ* using Sfp (Fig. 1). We confirmed the loading of probe **1** onto *apo*-AcpP using conformationally-sensitive urea-PAGE[16] (Fig. S4) and LC-MS (ESI) analyses (Fig. S5).

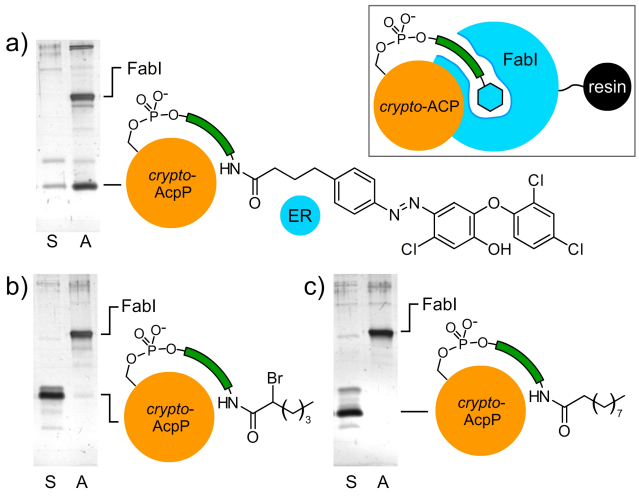


Figure 2. FabI resin was used to selectively isolate *crypto*-AcpP. SDS-PAGE gels shown from application of 20 µL of resin bearing 80 µM FabI to 20 µL of a solution containing 100 µM of **a)** the ER specific *crypto*-**1**-AcpP. Negative controls including: **b)** an α-bromoamide *crypto*-AcpP and **c)** a fatty acid *crypto*-AcpP (see control probe structures in Fig. S2). Lanes depict supernant (S) from step 3 (Fig. S6) and affinity (A) purified fractions from step 4 (Fig. S6).

We used an affinity assay to explore the specificity of the *crypto*-**1**-AcpP to FabI (see schematic representation

in Fig. S6). FabI was covalently immobilized on Affi-Gel 10 resin and mixed with a panel of *crypto*-AcpPs to explore the selectivity of the domain specific motif. As seen in Fig. 2a, only minimal levels of *crypto*-**1**-AcpP were observed in the supernatant while both FabI (released from the resin) and *crypto*-**1**-AcpP were largely observed in the affinity-isolated fraction, indicating that *crypto*-**1**-AcpP interacts with FabI. Alternatively, AcpP was not obtained when repeating the same procedure using two control probes bearing α-bromohexanoate (Fig. 2b) and decanoate (Fig. 2c) tethered to AcpP. This data indicated the binding of AcpP to FabI was only engaged when the ER domain-specific unit was present on the pantetheinamide terminus.

To further test selectivity for FabI, we reversed the affinity system. *Crypto*-**1**-AcpP was appended to Affi-Gel 10 and screened for its ability to isolate FabI from a series of lysates. As shown in Fig. 3, *crypto*-**1**-AcpP resin was able to selectively isolate FabI from *E. coli* K12 lysate spiked with pure FabI (lysate 1, Fig. 3), lysate from *E. coli* engineered to overexpress FabI (lysate 2, Fig. 3), and *E. coli* K12 lysate (lysate 3, Fig. 3).

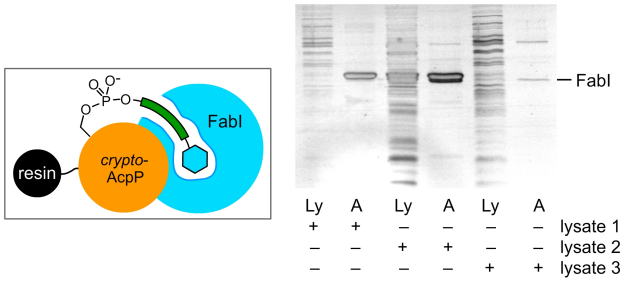


Figure 3. *Crypto*-**1**-AcpP resin was used to isolate FabI from different lysates (Ly). Lysate 1 contained 70 µL of K12 lysate (1.0 mg/mL in total protein) spiked with 10 µL of 80 µM FabI. Lysate 2 contained 70 µL of *E. coli* overexpressing FabI lysate (1.0 mg/mL in total protein). Lysate 3 contained 70 µL of K12 lysate (1.0 mg/mL in total protein). Affinity isolated fractions (A) were generated by using 15 µL of resin containing 75 µM of the *crypto*-**1**-AcpP.

Using purified recombinant proteins, we evaluated the ability of *crypto*-**1**-AcpP to inhibit FabI (Fig. 4a). The inhibition of *crypto*-**1**-AcpP (IC_{50} value of 1.1±0.1 µM) (Fig. 4a and Fig. S7) was 50-fold greater than probe **1** (IC_{50} value of 49.3±0.2 µM) (Fig. S3), therein highlighting the importance of AcpP interactions.

Next, we tested the ability of FabI to discriminate cognate and non-cognate carrier proteins. ActACP, the ACP from the type II actinorhodin polyketide synthase from *S. coelicolor*,[17] was loaded with **1** using the chemoenzymatic labeling protocol (Fig. 1). *Crypto*-**1**-ActACP did not show comparable inhibition of FabI to the cognate *crypto*-**1**-AcpP in the same range (Fig. 4a). ActACP (PDB ID: 2K0X) docking studies with FabI structures (PDB IDs: 2FHS, 1DFI, and 1QSG) indicated no catalytically active orientation (Fig. S8a). As opposed to AcpP, where 20% of structures show Ser36 pointing towards the FabI binding pocket (Fig. S8b), the site of phosphopanteheine attachment on ActACP, Ser42, does not point towards the FabI active site (Fig. S8a). No FabI inhibition was observed for *crypto*-**1**-ActACP, *crypto*-acyl-AcpP, *apo*-AcpP, or *holo*-AcpP (Fig. S7) highlighting the importance of the loaded AcpP cargo.

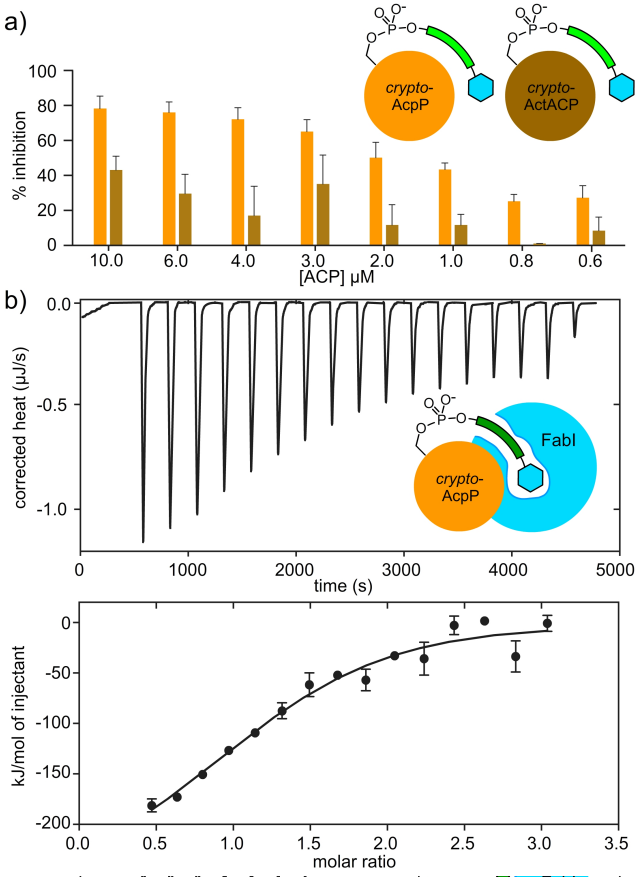


Figure 4. Inhibition and binding data. **a)** Comparison of inhibition of 0.02 µM FabI with either *crypto*-**1**-ActACP (brown) or *crypto*-**1**-AcpP (orange) at concentrations ranging from 0.15 µM to 10.00 µM. **b)** ITC analysis of the binding of *crypto*-**1**-AcpP to FabI. The initial concentration of the FabI monomer was 4.6 µM in the cell and 18 injections of 3 µL of 40 µM *crypto*-**1**-AcpP were delivered sequentially. Data was collected in duplicate with a standard error less than 5%.

Since probe **1** does not allow for covalent attachment of FabI and AcpP, we wondered whether the *crypto*-**1**-AcpP and FabI interaction was strong enough to stabilize the complex for future studies. We therefore utilized isothermal titration calorimetry (ITC) to measure the binding affinity of the two proteins (Fig. 4b). Reports indicate that the 110.9 kDa FabI tetramer consists of monomers made up of seven β-strands packed by eight helices.[7] The resulting *crypto*-**1**-AcpP stoichiometry of binding was calculated to be 1:1 to each FabI monomer (n=1.2+0.1), suggesting that each site is independent and

identical. Interestingly, the binding stoichiometry of *holo*-AcpP to the ketosynthase domain, KSII (FabF) from *E. coli*,[18] was also calculated to be 1:1. The FabI • AcpP crystal structure showed a stoichiometry of 2:1, albeit with poor resolution of the AcpP interface.[7]

We then explored the biophysical parameters guiding the interaction of *crypto*-**1**-AcpP to FabI. ITC analysis (Fig. 4b) returned a K_d value of 711.9±1.3 nM, which is 3-10 fold lower than other AcpP • partner protein K_d values.[19] This interaction was exothermic, with ΔG=-34.0±0.4 kJ/mol (ΔH=-234.9±26.7 kJ/mol), and had an entropic loss (ΔS=-677.4±94.9 J•mol/K) characteristic of an enthalpy-driven binding event, presumably due to disordered of the protein • protein interaction.[20]

The present study extends our collection of chemoenzymatic AcpP tools with the first inhibitor-based non-covalent triclosan probe **1**. This probe was appended to AcpP and was able to recognize and isolate FabI from complex lysates. The low micromolar inhibition of FabI with *crypto*-**1**-AcpP reveals a strong interaction of our proposed probe with FabI. This was further supported by the enhanced binding of *crypto*-**1**-AcpP to FabI.

Bacterial and apicomplexan ER domains from fatty acid synthases are currently targeted by several antibiotics, but resistance is increasing. Small molecules that disrupt the interface between AcpP and ER domains may offer a viable route for antibiotic design to combat resistance, but more structural information about this interaction is necessary. We envision the use of probe **1** or related probes to aid further structural characterization of the AcpP • ER interaction. A structural understanding of how ACPs interact with their cognate enzymes will also pave the way for metabolic engineering of biosynthetic pathways for the synthesis of pharmaceutically relevant metabolites, while also identifying essential ACP interactions for pathogenic organisms.

ASSOCIATED CONTENT

Additional figures, synthetic methods, protocols, and spectroscopic data has been provided. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

mburkart@ucsd.edu

Author Contributions

†These authors contributed equally

‡Current address: Department of Microbiology and Immunology, Drexel University College of Medicine.

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REFERENCES

(1) (a) Prescott, D. J.; Vagelos, P. R., *Adv. Enzymol. Relat. Areas Mol. Biol.* **1972**, *36*, 269; (b) Rock, C. O.; Cronan, J. E., *Biochim. Biophys. Acta* **1996**, *1302*, 1.

(2) Blatti, J. L.; Beld, J.; Behnke, C. A.; Mendez, M.; Mayfield, S. P.; Burkart, M. D., *PLoS One* **2012**, *7*, e42949.

(3) Worthington, A. S.; Burkart, M. D. *Org. Biomol. Chem.* **2006**, *4*, 44.

(4) (a) Nazi, I.; Koteva, K. P.; Wright, G. D., *Anal. Biochem.* **2004**, *324*, 100; (b) La Clair, J. J.; Foley, T. L.; Schegg, T. R.; Regan, C. M.; Burkart, M. D., *Chem. Biol.* **2004**, *11*, 195.

(5) Bergler, H.; Fuchsbichler, S.; Hçgenauer, F.; Turnowsky, F. *Eur. J. Biochem.* **1996**, *242*, 689.

(6) Heath, R. J.; Rock, C. O., *J. Biol. Chem.* **1995**, *270*, 26538.

(7) Rafi, S.; Novichenok, P.; Kolappan, S.; Zhang, X.; Stratton, C. F.; Rawat, R.; Kisker, C.; Simmerling, C.; Tonge, P. J., *J. Biol. Chem.* **2006**, *281*, 39285; (b) Finzel, K.; Lee, D. J.; Burkart, M. D. *Chembiochem*. **2015**, *16*, 528.

(8) Ishikawa, F.; Haushalter, R. W.; Lee, D. J.; Finzel, K.; Burkart, M. D., *J. Am. Chem. Soc.* **2013**, *135*, 8846.

(9) (a) Pan, P.; Tonge, P. J. *Curr. Top. Med. Chem.* **2012**, *12*, 672; (b) Lu, H.; Tonge, P. J. *Acc. Chem. Res*. **2008**, *41*, 11.

(10) (a) Saito, J.; Yamada, M.; Watanabe, T.; Iida, M.; Kitagawa, H.; Takahata, S.; Ozawa, T., Takeuchi, Y.; Ohsawa F. Protein Sci. 2008, 17, 691; (b) Rafi, S., Novichenok, P., Kolappan, S.; Zhang, X.; Stratton, C. F.; Rawat, R.; Kisker, C.; Simmerling, C.; Tonge, P. J. *J. Biol. Chem.* **2006**, *281*, 39285.

(11) (a) Heath, R. J.; Yu, Y. T.; Shapiro, M. A.; Olson, E.; Rock, C. O., *J. Biol. Chem.* **1998**, *273*, 30316; (b) Swinney, D. C., *Nat. Rev. Drug Discov.* **2004**, *3*, 801; (c) Sivaraman, S.; Sullivan, T. J.; Johnson, F.; Novichenok, P.; Cui, G.; Simmerline, C.; Tonge, P. J., *J. Med. Chem.* **2004**, *47*, 509.

(12) (a) Heath, R. J.; Rubin, J. R.; Holland, D. R.; Zhang, E.; Snow, M. E.; Rock, C. O., *J. Biol. Chem.* **1999**, *274*, 11110; (b) Qiu, X.; Abdel-Meguid, S. S.; Janson, C. A.; Court, R. I.; Smyth, M. G.; Payne, D. J., *Protein Sci.* **1999**, *8*, 2529; (c) Heath, R. J.; Su, N.; Murphy, C. K.; Rock, C. O., *J. Biol. Chem.* **2000**, *275*, 40128; (d) Kapoor, M.; Reddy, C. C.; Krishnasastry, M. V.; Surolia, N.; Surolia, A., *Biochem. J.* **2004**, *381*, 719.

(13) (a) Ward, W. H.; Holdgate, G. A.; Roswell, S.; McLean, E. G.; Pauptit, R. A.; Clayton, E.; Nichols, W. W.; Colls, J. G.; Minshull, C. A.; Jude, D. A.; Mistry, A.; Timms, D.; Camble, R.; Hales, N. J.; Britton, C. J.; Taylor, I. W., *Biochem.* **1999**, *38*, 12514; (b) Sivaraman, S.; Zwahlen, J.; Bell, A. F.; Hedstrom, L.; Tonge, P. J., *Biochem.* **2003**, *42*, 4406; (c) Tallorin, L.; Durrant, J. D.; Nguyen, Q. G.; McCammon, J. A.; Burkart, M. D., *Bioorg. Med. Chem.* **2014,** *22*, 6053.

(14) (a) Nguyen, C.; Haushalter, R. W.; Lee, D. J.; Markwick, P. R.; Bruegger, J.; Caldara-Festin, G.; Finzel, K.; Jackson. D. R.; Ishikawa, F.; O'Dowd, B.; McGammon, A.; Opella, S. J.; Tsai, S. C.; Burkart, M. D., *Nature* **2013**, *505*, 427; (b) Zhang, Y. M.; Rao, M. S.; Heath, R. J.; Price, A. C.; Olson, A. J.; Rock, C. O.; White, S. W., *J. Biol. Chem.* **2001**, *276*, 8231; (c) Zhang, Y. M.; Wu, B.; Zheng, J.; Rock, C. O.; *J. Biol. Chem.* **2003**, *278*, 52935.

(15) Brun, E. M.; Bonet, E.; Puchades, R.; Maquieira, A. *Environ. Sci. Technol.* **2008**, *42*, 1665.

(16) Rock, C. O.; Jackowski, S., *J. Biol. Chem.* **1982**, *257*, 10759.

(17) Haushalter, R. W.; Filipp, F. V., Ko, K.; Yu, R.; Opella, S. J.; Burkart, M. D., *ACS Chem. Biol.* **2011**, *6*, 4.

(18) Worthington, A. S.; Porter, D. F.; Burkart M. D. *Org. Biomol. Chem.* **2010**, *8*,1769.

19) (a) Arthur, C.J.;Williams, C.; et.al. *ACS Chem. Bio.* **2009**, *4*, 625.

(20) Keskin, O.; Gursoy, A.; Ma, B.; Nussinov, R. *Chem. Rev.* **2008**, *108*, 1225-1244.

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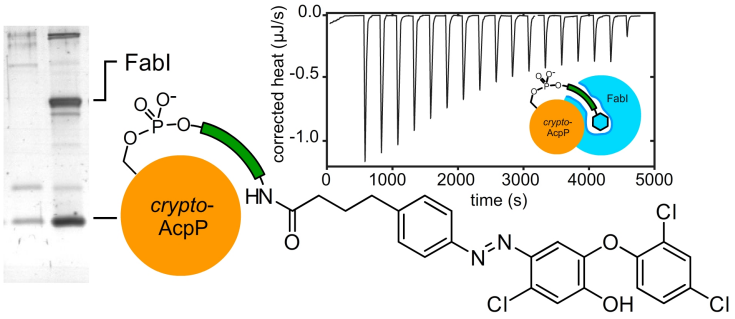
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