

Intermittent bioirrigation and oxygen dynamics in permeable sediments: An experimental and modeling study of three tellinid bivalves

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1 Abstract

2 To explore the dynamic nature of geochemical conditions in bioirrigated marine permeable
3 sediments, we studied the hydraulic activity of 3 tellinacean bivalve molluscs (the Pacific species
4 *Macoma nasuta* and *Macomona liliana*, and the northern Atlantic and Pacific species *Macoma*
5 *balthica*). We combined porewater pressure sensing, time-lapse photography and oxygen
6 imaging to quantify the durations and frequencies of tellinid irrigation activity and the associated
7 oxygen dynamics in the sediment. Porewater pressure records of all tellinids were dominated by
8 intermittent porewater pressurization, induced by periodic water injection into the sediment
9 through their excurrent siphons, which resulted in intermittent oxygen supply to subsurface
10 sediments. The durations of irrigation (2–12 min long) and intervals between subsequent
11 irrigation bouts (1.5–13 min) varied among tellinid species and individual sizes. For large *M.*
12 *liliana* and *M. nasuta*, the average durations of intervals between irrigation bouts were
13 sufficiently long (10 min and 4 min, respectively) to allow complete oxygen consumption in
14 between irrigation bouts in all tested sediment types. Irrigation patterns of smaller conspecifics
15 and the smaller species *M. balthica* were characterized by significantly shorter separation of
16 irrigation bouts, which resulted in more continuous oxygenation of the sediment. Transport-
17 reaction modeling confirmed these species- and size-specific geochemical signatures and
18 indicated that the geochemical character of the sediment is largely conditioned by the interplay
19 between temporal irrigation patterns and sedimentary oxygen consumption rates. For large
20 tellinids, model simulations indicated that oscillatory rather than stationary geochemical
21 conditions are prevalent in a wide range of sediment types, with oxic pockets collapsing
22 completely between periods of active irrigation. Based on the model results we developed
23 analytical approximations that allow estimation of spatio-temporal characteristics of sediment

oxygenation for a wide range of sediment types and infaunal activity patterns. Our results emphasize the need to consider the intermittent nature of bioirrigation when studying the geochemical impact of infauna in permeable sediments.

27

28 1. Introduction

Bioturbation by infaunal organisms has diverse impacts on the functioning and ecology of benthic systems. It alters the sediment characteristics and the distribution of porewater constituents (Aller, 1980; Boudreau and Marinelli, 1994; Berg *et al.*, 2001; Volkenborn *et al.*, 2007), affects microbial diversity and rates of microbially mediated processes (Aller and Yingst, 1985; Kristensen, 1988; Na *et al.*, 2008), and influences the structure of micro-, meio- and macrofaunal benthic communities (Reise, 1981; Meyers *et al.*, 1987; Woodin *et al.*, 1998; Marinelli *et al.*, 2002; Volkenborn and Reise, 2006; Engel *et al.*, 2012). The significant impact of single or multiple species on benthic–pelagic exchange fluxes is well documented (Vopel *et al.*, 2003; Lohrer *et al.*, 2004; Karlson *et al.*, 2007; D’Andrea and DeWitt, 2009, Michaud *et al.*, 2009), but time–integrated approaches such as benthic chamber incubations are likely to average over the dynamic nature of sediment reworking (Solan *et al.*, 2004) and porewater bio–advection (Wetthey *et al.*, 2008; Volkenborn *et al.*, 2010). Bioturbation is the product of infaunal activities, e.g. burrowing, pumping, feeding, or defecating, and consequently the rates and direction of particle movement and fluid flow depend on the frequencies, durations, and the sequence of behaviors. A number of experimental studies have shown that infaunal organisms irrigate their burrows in a discontinuous fashion on a timescale of minutes (e.g. Kristensen *et al.*, 1991) to hours (e.g., in intertidal areas where the ventilation by infaunal organisms ceases during ebb tide; Aller and Yingst, 1978; Aller *et al.*, 1983), implying that bioturbated sediments are subject to

47 dynamic geochemical conditions on this time scale (Timmermann *et al.*, 2006; Volkenborn *et al.*,
48 2010, 2012). Typically, irrigation activity patterns are governed by endogenous pacemakers, but
49 for many organisms they are also altered in response to changes in environmental conditions
50 (e.g., O₂ concentration in the irrigated water; Dales *et al.*, 1970).

51

52 For a better mechanistic understanding of how infaunal organisms affect biogeochemical
53 processes, it is essential to perform measurements and modeling on temporal and spatial scales
54 that reflect the activities of individual organisms. The need to implement discontinuous irrigation
55 into bioirrigation models was first addressed by Boudreau and Marinelli (1994), who developed
56 a derivative of the cylinder microenvironment model introduced by Aller (1980) to estimate the
57 time dependence of solute fluxes. In diffusion-dominated systems intermittent flushing of
58 burrows causes strong temporal variations in the instantaneous solute fluxes (Boudreau and
59 Marinelli, 1994). In advection-dominated systems the role of intermittent irrigation is less well
60 understood, though larger sediment volumes are supplied with reactive chemical species. The
61 pocket injection model developed by Meysman *et al.* (2005) captures the main spatial features of
62 porewater advection when water is forced into and through permeable sediments continuously
63 and at a constant rate, but its implementation under intermittent irrigation conditions is lacking.
64 As recently shown for some important bioirrigating organisms such as arenicolid polychaetes
65 and thalassinid crustaceans, bioadvective porewater transport is driven by species- and behavior-
66 specific porewater pressure dynamics (Wetthey *et al.*, 2008; Woodin *et al.*, 2010; Volkenborn *et*
67 *al.*, 2010, 2012). These pressure gradients can be detected tens of centimeters away from the
68 organisms and result in unsteady and bidirectional porewater transport (Wetthey and Woodin,
69 2005; Wetthey *et al.*, 2008; Volkenborn *et al.*, 2010). As a consequence, substantial sediment

70 volumes surrounding infaunal organisms and their burrows experience intermittent oxygen
71 supply and frequent oscillations between oxic and anoxic conditions (Volkenborn *et al.*, 2012).

72

73 This study provides experimental evidence that links hydraulic activities of bioirrigating infauna
74 with oscillation of geochemical conditions in sediments, using three tellinid bivalve mollusc
75 species as model organisms. Tellinids are cosmopolitan nearshore bivalves with separate inhalant
76 and excurrent siphons. They are facultative deposit- and suspension feeders (Levinton, 1991).
77 While the inhalant siphon is used for the uptake of surface particles and overlying water, the
78 excurrent siphon remains several cm deep in the sediment. During feeding, water passes through
79 the bivalves and is injected into the sediment at a depth of 2–10 cm, resulting in intermittent
80 sediment pressurization (Woodin *et al.*, 2010, 2012). Here we have investigated temporal
81 patterns of bioirrigation associated with deposit-feeding by tellinid bivalves and their impact on
82 oxygen dynamics in different sediment types.

83

84 Our focus is on the dynamics of O₂ as the energetically most favorable electron acceptor in
85 biogeochemical reactions and as a widely used proxy for organic matter mineralization (see
86 review by Glud, 2008). Specifically, we explore the volume of sediment that is characterized by
87 either permanently oxic or intermittently oxic conditions, which can be regarded as
88 geochemically distinct environments (Aller, 1994). Sediments experimentally exposed to oxic–
89 anoxic oscillatory conditions – at least on the scale of days to weeks (we are not aware of studies
90 at minute to hour timescales) – have been shown to exhibit higher mineralization rates of organic
91 matter, including pollutants, than those with stable redox conditions (Sun *et al.*, 1993; Aller,
92 1994; Sun *et al.*, 2002; Caradec *et al.*, 2004; Cravo-Laureau *et al.*, 2011; Bennett *et al.*, 2012).

93 Given these changes in mineralization rates, quantification of the frequencies of redox oscillation
94 and time scales of the behaviors driving these rates is critical. Focusing on redox oscillations and
95 using empirical data, we parameterized a transport-reaction model to estimate the relationship
96 between temporal irrigation patterns and (intermittently) oxic sediment volumes in a wide range
97 of sediment types inhabited by these bivalves and by other important bioturbating organisms.

98

99 2. Methods

100 Hydraulic activities of tellinid species were investigated in laboratory containers and narrow
101 aquaria using isolated individuals. Porewater pressure sensing was combined with time-lapse
102 photography to identify porewater pressure waveforms associated with specific tellinid behaviors
103 (as previously described in Volkenborn *et al.*, 2010; for pressure sensor specifications see
104 Wethey and Woodin, 2005; Volkenborn *et al.*, 2012). Time synchronized planar optode imaging
105 of oxygen was used to investigate the related oxygen dynamics in the porewater (for details see
106 Matsui *et al.*, 2011; Volkenborn *et al.*, 2012). The experimental set-up differed among species,
107 and the detailed information on aquarium sizes, overlying water exchange rates, sediment
108 characteristics, temperature, light, salinity and experimental replication and duration are given
109 below. In all experiments, sediments were sieved through a coarse mesh (5–10 mm) to remove
110 larger organisms. In parallel to the aquaria setup, sediment cores (10 cm² area) were filled with
111 10–15 cm of sediment for the measurement of sediment permeability by the constant head
112 method (Klute and Dirksen, 1986). Sub-samples were taken with 5 mL cut-off syringes and used
113 for the determination of porosity (weight loss by drying). Sediment reactivity, approximated by
114 the rate of oxygen consumption, was determined by injecting oxygenated water into the sediment
115 at locations close to the optode but far from the organisms and subsequently measuring

116 porewater oxygen concentrations in 10 s intervals until all oxygen was consumed (which took 1-
117 5 min, depending on the sediment type). The volumetric rate of sedimentary oxygen
118 consumption, K_{OX} , expressed in micromoles of O_2 per volume of sediment (L_s) and time (μmol
119 $O_2 L_s^{-1} \text{ min}^{-1}$), was then estimated as the initial rate of decrease in the measured porewater O_2
120 concentrations multiplied by sediment porosity (Polerecky *et al.*, 2005; Volkenborn *et al.*, 2010).

121

122 *a. Study species and experimental setup*

123 *Macomona liliana* is a common bivalve in New Zealand intertidal sandflats (Roper *et al.*, 1992),
124 where it lives up to 10 cm below the sediment surface (Powell, 1979). Twenty large individuals
125 (shell length of 4.1–5.0 cm) were collected in February 2010 from an intertidal sandflat near the
126 northern entrance of Tauranga Harbour, North Island, New Zealand (37°27.77'S 175°57.90'E)
127 and four small individuals (shell length of 2.3–2.4 cm) were collected nearby, west of Tuapiro
128 Point (37°29.37'S 175°57.0'E). For porewater pressure measurements, circular containers (11.1
129 cm diameter and 10.5 cm deep for large individuals; 17.5 cm diameter and 11 cm deep for small
130 individuals) were filled with sediment and immersed in running artificial seawater (Red Sea Salt,
131 Red Sea Aquatics Ltd, Eilat, Israel) (31–34 ‰ salinity). For measurements with large individuals
132 eight containers were filled with either muddy sand or sandy sediment. For measurements with
133 small individuals we only used muddy sand sediment. Each container was equipped with a
134 pressure sensor at approx. 8 cm depth. Time-lapse photographs (every 15 s) were taken from
135 above with SLR cameras with flash (Nikon D200 and D300). Experiments were done at a
136 temperature of 17–19°C under ambient light and lasted for 6 days. Oxygen imaging was done on
137 four *M. liliana* individuals, incubated in narrow aquaria (20 cm × 4.5 cm surface, 20 cm deep).
138 Two of these aquaria were filled to a depth of approximately 15 cm with muddy sand and sandy

139 sediment, respectively. Overlying water was exchanged at a rate of 100 mL min^{-1} , resulting in
140 complete exchange every 5 min. Each aquarium was equipped with a pressure sensor that was
141 inserted from above to about 8 cm depth, and with oxygen sensitive foils ($20 \text{ cm} \times 20 \text{ cm}$) that
142 were glued to the inside of both large walls. Incubations were done in the dark for 7 days at 20–
143 23°C . Oxygen images were taken simultaneously from 2 aquaria every 15 s and time-lapse
144 photographs were taken from above 5 s after each optode image. The aquaria were exchanged
145 and rotated opportunistically in order to capture periods with tellinids irrigating close to the
146 optode.

147

148 *Macoma nasuta* is abundant in intertidal flats in the eastern North Pacific Ocean, where it
149 burrows 10–20 cm deep into the sediment (Levinton, 1991; Meyerhöfer, 1985). Six specimens
150 with a shell length of 4.7–5.2 cm were collected in April 2010 from the mid to high intertidal of
151 False Bay, Washington, USA ($48^{\circ}29.35'\text{N}$, $123^{\circ}4.03'\text{W}$). Individuals were incubated in six
152 square aquaria ($10 \times 10 \text{ cm}$ surface, 20 cm deep), filled to approximately 15 cm with either
153 muddy sand or sandy sediment. One wall of each aquarium was equipped with an oxygen
154 sensitive foil ($10 \text{ cm} \times 20 \text{ cm}$) and pressure sensors were deployed from above at approx. 8 cm
155 depth. Overlying water (from Friday Harbor Laboratories seawater system) was exchanged at
156 100 mL min^{-1} , resulting in a complete exchange approx. every 5 min. Oxygen images from the
157 side were taken simultaneously from the six aquaria and top-down time-lapse flash-illuminated
158 photographs were taken with a 5 s delay. Measurements were done over a period of 4 days in the
159 dark at a temperature of $12\text{--}14^{\circ}\text{C}$ and a salinity of 34 ‰.

160

161 *Macoma balthica* is a cosmopolitan nearshore bivalve abundant in intertidal areas of the northern
162 parts of the Atlantic and Pacific Oceans. Porewater pressure measurements and time-lapse
163 photography were done in Germany in May 2007 and August 2011. Four individuals with shell
164 lengths between 1.8–2.0 cm were collected from a mid-intertidal sandflat in Königshafen, Sylt,
165 Germany (55°02.21'N, 8°24.53'E) and individuals were held in sediment-filled 1 L beakers (in
166 2007) or 5 L buckets (in 2011) immersed in running seawater from the Wadden Sea Station
167 seawater system. Measurements were done over 6 days at a temperature of 15–18°C and a
168 salinity of 33–35 ‰. Oxygen imaging was done in Finland in August 2011. Six individuals of *M.*
169 *balthica* (~1.6 cm shell length) were collected from a subtidal location near the Tvärminne
170 Zoological Station (59°50.60'N, 23°15.71'E). The set-up was the same as in the *M. nasuta*
171 experiments (see above). Measurements were done in the dark at 12–15°C and a salinity of 6 ‰.

172

173 ***b. Porewater pressure analysis***

174 Porewater pressure data were analyzed with respect to time allocation to feeding activity,
175 burrowing, and inactivity for each individual. Intervals of inactivity were defined as periods
176 without porewater pressure dynamics for >30 min. Burrowing episodes were defined as time
177 intervals with high magnitude pressure oscillations with intermittent sediment pressurization,
178 where there were < 20 min between subsequent intervals of high magnitude pressure oscillations.
179 The first and last high magnitude pressure pulses were used as start and end points to define a
180 burrowing episode. For periods of deposit-feeding the average durations of pressurized (T_{PUMP})
181 and non-pressurized (T_{REST}) intervals and the average frequencies of feeding bouts were
182 determined for each individual. Feeding bouts were defined as distinct if separated by a > 20 s
183 gap. Effects of the sediment type on these measures of hydraulic activity were tested for each

184 species by one-way factorial ANOVA. For *Macomona liliana* the effect of tellinid size on the
185 temporal patterns in hydraulic activity was additionally tested by a one-way factorial ANOVA.

186

187 *c. Transport-reaction modeling*

188 To study the spatial and temporal dynamics of O₂ in an intermittently bioirrigated sediment, a
189 transport-reaction model was set up in COMSOL 4.3, a finite-element modeling environment.

190 The 2D axisymmetric model domain consisted of a cylinder (radius 20 cm, sufficiently large to

191 minimize boundary effects) with the top 2 cm filled with water and the bottom 20 cm with

192 sediment. The mesh consisted of approximately 5500 finite elements with side lengths between

193 <1 mm near the injection pocket and the sediment water interface and 6 mm in regions with low

194 flow and small concentration gradients. A spherical injection pocket with a radius of 0.2 cm was

195 positioned on the central symmetry axis 5.75 cm below the sediment surface. The transport of

196 the overlying water and porewater was modeled using COMSOL's implementation of Navier–

197 Stokes and Brinkman equations describing free flow in an isotropic porous media (Le Bars and

198 Grae Worster, 2006), accounting for pressure gradients and viscous forces but neglecting inertial

199 terms. As a solver we used the sparse direct solver MUMPS (<http://graal.ens-lyon.fr/MUMPS>).

200 The top of the domain was set as open (zero normal stress), and the normal flow at the bottom

201 and vertical domain boundaries was set to zero. Intermittent irrigation was modeled by imposing

202 the fluid velocity at the injection boundary to values that varied between zero (during the resting

203 interval T_{REST}) and the ratio of the instantaneous volumetric pumping rate and the surface area of

204 the injection pocket (during the pumping interval T_{PUMP}). Permeability of 5×10^{-12} m² and

205 porosity 0.45 were used in all simulations. Because irrigation was implemented by imposing

206 flow velocity rather than pressure at the inlet boundary, the flow velocity field and the results
207 derived from it were not affected by this choice of sediment permeability.

208

209 Pumping rates of tellinid bivalves for model parameterization were derived from the literature.
210 For *Macoma nasuta* of comparable sizes to those used in our experiments (5.0 cm shell length),
211 Meyerhöfer (1985) measured instantaneous pumping rates of 1.8 mL min⁻¹ with thermistor
212 probes. Based on this instantaneous pumping rate, and given the proportional time of active
213 irrigation by *M. nasuta* measured in this study (7 irrigation bouts h⁻¹, 5 min duration; see
214 Results), time-integrated pumping rates would be around 1.1 mL min⁻¹, which is in the range of
215 pumping rates estimated with the “clambox” when averaged over 12 min intervals (Specht and
216 Lee, 1989). Assuming the *Macoma nasuta* dry weight versus instantaneous pumping rate
217 relationship estimated by Meyerhöfer (1985), and the size versus dry weight relationship found
218 by Gallucci and Hylleberg (1976), we estimated the instantaneous pumping rates for the size
219 classes used in our experiments to be 1.8, 1.6, 0.43, and 0.35 mL min⁻¹ for *M. nasuta*, large and
220 small *M. liliana*, and *M. balthica*, respectively. In additional simulations exploring the
221 relationship between pumping rates and oxic sediment volumes, instantaneous pumping rates
222 were varied between 0.1 and 3 mL min⁻¹.

223

224 O₂ concentrations in the porewater, denoted as C , were computed using

$$225 \quad \phi \frac{\partial C}{\partial t} = \nabla \cdot (D \nabla C) - \vec{u} \cdot \nabla C + R \quad (1)$$

226 where $\vec{u}$ is the advection velocity field (m s⁻¹), ϕ is porosity, and D (in m² s⁻¹) parameterizes
227 diffusion when in the sediment ($D = \phi / (1 - 2 \times \ln(\phi)) \times D_{\text{mol}}$, with $D_{\text{mol}} = 1.9 \cdot 10^{-9}$ m² s⁻¹; Schulz,
228 2000) and dispersion when in the overlying water ($D = D_{\text{mol}} + 0.4 \cdot \nu / 363 \cdot (z \cdot u^* / \nu)^3$, where ν is the

229 kinematic viscosity ($10^{-6} \text{ m}^2 \text{ s}^{-1}$), u^* is shear velocity and z is the distance from the sediment–
 230 water interface; Boudreau, 2001). The shear velocity was set to 10^{-5} m s^{-1} , which was sufficient
 231 to establish a homogeneous O_2 concentration distribution in the overlying water; its value did not
 232 significantly affect the burrow-associated oxic sediment volume discussed below. Hydrodynamic
 233 dispersion in the sediment was neglected because it plays only a minor role at low flow velocities
 234 (Meysman *et al.*, 2006). The net O_2 consumption rate R was set to 0 in the overlying water and
 235 was modeled as $-K_{\text{OX}} \times C / (K_{\text{M}} + C)$ in the sediment, where K_{OX} is the maximum sedimentary
 236 oxygen consumption rate (see above) and K_{M} is the half-saturation constant. In muddy sands,
 237 K_{OX} values are typically in the range of $5\text{--}30 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$ (Glud *et al.* 2003; Volkenborn *et*
 238 *al.*, 2010). In more muddy sediments, they may exceed $50 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$ (e.g., Volkenborn *et*
 239 *al.*, 2012), and are typically $<5 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$ in highly permeable sands (Billerbeck *et al.*,
 240 2006; Werner *et al.*, 2006; Volkenborn *et al.*, 2012). Model simulations used K_{OX} values in the
 241 range from 2.25 to $45 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$, thus covering the range found in natural sediments. The
 242 half-saturation constant K_{M} was set to $1 \mu\text{M}$, with comparable results achieved when setting it to
 243 $10 \mu\text{M}$ (not shown) which is in the range of K_{M} values used in previous studies (e.g., Van
 244 Cappellen and Wang, 1995). In all simulations, a no flux condition was imposed at the bottom
 245 and side of the domain, and O_2 concentration of $220 \mu\text{M}$ was imposed at the top of the domain,
 246 whereas in the injected water it was set to $88 \mu\text{M}$, consistent with the maximal porewater O_2
 247 concentrations detected at the aquarium wall in our experiments (typically around 40% air
 248 saturation). This reduction in O_2 can be mostly attributed to the respiratory O_2 uptake by the
 249 pumping organisms.

250

251 3. Laboratory results

252 *a. Tellinid behaviors and related porewater pressure waveforms.* All three tellinid species
253 engaged in a range of hydraulic activities that caused behavior-specific porewater pressure
254 waveforms. An exemplary pressure record from *Macomona liliana* is shown in Figure 1. Most of
255 the time, all 3 species were surface-deposit-feeding (videos 1–3 in the appendix). Episodes of
256 deposit-feeding lasted from several hours to a few days and were characterized by intermittent
257 irrigation as indicated by frequent alternations of pressurized and non-pressurized intervals (Fig.
258 1). From time to time tellinids relocated their excurrent siphons, which could be seen in time-
259 lapse images as a change in the location of particle movement and accumulation of fecal pellets
260 at the sediment surface. Movements of the excurrent siphon occurred while the sediment was
261 pressurized and caused short positive pressure pulses at typical frequencies of 2 pulses min⁻¹
262 (Fig. 1C). Movements of the incurrent siphon were associated with short positive or negative
263 pressure pulses around the hydrostatic pressure baseline. Deposit-feeding was accompanied by
264 frequent sediment blow-outs through the incurrent and excurrent siphons, i.e. pseudofeces
265 expulsions and defecations, respectively, both causing very similar short negative or positive
266 pressure pulses (<4 s decreasing pressure, followed by an active recovery or sediment-dependent
267 pressure recovery over 2–7 s). Burrowing caused substantial sediment movement, and was
268 characterized by large positive and negative pressure pulses (Fig. 1B). The frequencies and
269 durations of hydraulic activities varied among species (Figs. 2 and 3; Table 1). However, for all
270 of them, the time-integrated pressures (pressure amplitude × duration) during the pulses
271 associated with feces or pseudofeces expulsions were typically <5 % of the pressures integrated
272 over one irrigation bout. Thus, though large in amplitude, the short duration of the pressure

pulses associated with feces and pseudofeces expulsions suggests little contribution of these behaviors to the transport of porewater by the bivalves, which is consistent with findings for arenicolids (Wetthey *et al.*, 2008). Thus, we have focused the analysis on the intermittent porewater pressurization during deposit-feeding, since this behavior was the most important in terms of time allocation (Table 1) and porewater advection.

b. Species-specific irrigation patterns. Analysis of the pressure records revealed that the average durations of pumping (T_{PUMP}) and resting (T_{REST}) differed among tellinid species and size classes (Fig. 2, Table 1) The temporal irrigation patterns of individuals were characterized by a substantial variation around the individual means. On average, the standard deviation of T_{PUMP} and T_{REST} of individuals was 40% of the mean duration (Fig. 3).

The large *Macomona liliana* specimens were hydraulically active 94% of the time ($n=16$ individuals, 8 in mud, 8 in sand over 5 days). $88 (\pm 4) \%$ of the time these individuals were deposit-feeding. Feeding episodes were characterized by intermittent porewater pressurization, with $8.3 (\pm 2.5)$ min of pressurization separated by $10.0 (\pm 1.8)$ min intervals of no hydraulic activity (mean and SD of 16 individuals with 50–317 bouts analyzed per individual). This corresponded to $3.3 (\pm 0.5)$ feeding bouts h^{-1} and pressurization of the sediment during $45 (\pm 10) \%$ of the time while feeding. Sediment type had no significant effect on the durations of pressurized ($F_{1,14}=0.94$; $p=0.35$) and non-pressurized ($F_{1,14}=1.14$; $p=0.30$) intervals or feeding bout frequencies ($F_{1,14}=0.11$; $p=0.74$). Feeding activity by the small *M. liliana* specimens ($n=4$, all in muddy sand, 82 to 122 bouts analyzed per individual) was characterized by barely significantly shorter pressurized (4.4 ± 1.0 min, $F_{1,10}=4.8$; $p=0.051$), significantly shorter non-pressurized (6.7 ± 1.8 min; $F_{1,10}=12.9$; $p=0.005$) intervals, and significantly higher feeding bout

297 frequencies (5.6 ± 1.2 min; $F_{1,10}=18.6$; $p=0.002$) when compared to the larger conspecifics
298 feeding in the same muddy sediment type.

299

300 *Macoma nasuta* was hydraulically active 93% of the time ($n=5$ individuals over 3–4 days). After
301 the initial burrowing, they were actively deposit-feeding $83 (\pm 20)$ % of the time (mean and SD
302 of 5 individuals), with 2 individuals allocating a significant portion of the time to suspension
303 feeding (28% and 19%). While deposit-feeding, the incurrent siphon was continuously moving
304 around, ingesting surface sediment. Deposit-feeding was characterized by $5.0 (\pm 1.0)$ min of
305 pressurization separated by intervals of $3.7 (\pm 1.1)$ min ($n=5$ individuals, with 94–167 feeding
306 bouts analyzed per individual). The average frequency of feeding bouts was $7.1 (\pm 1.4) \text{ h}^{-1}$.
307 Burrowing accounted for $2.4 (\pm 3)$ % of the time budget and burrowing episodes lasted $68 (\pm 20)$
308 min. 3 of 6 individuals did not burrow at all after the episode of initial burrowing.

309

310 *Macoma balthica* individuals were small compared to the other tellinid species and the
311 magnitude of porewater pressurization was very small most of the time. Time allocation of
312 deposit-feeding was therefore determined from time-lapse photography. Over the 6–7 days of
313 observation the two individuals were actively feeding during 57% and 45% of the time. Feeding
314 occurred in bouts of $2.3 (\pm 0.2)$ min duration separated by intervals of $1.7 (\pm 0.2)$ min (78–197
315 bouts of 3 individuals analyzed). The feeding bout frequency was $15.1 (\pm 0.2) \text{ h}^{-1}$. Burrowing
316 occurred $0.62 (\pm 0.25)$ times per 24 h and lasted $12.9 (\pm 7.6)$ min.

317

318 ***c. Linking tellinid behaviors and oxygen dynamics.*** Over the course of the measurements, seven
319 of the 28 tellinid individuals positioned their excurrent siphons for some hours close to the

320 oxygen sensitive foil. While feeding, the sediment was intermittently supplied with oxygen.
321 Oxygen dynamics were directly related to porewater pressurization, with oxygen concentrations
322 increasing during pressurized and decreasing during non-pressurized intervals (Fig. 4). Oxic and
323 anoxic intervals typically alternated on the scale of minutes in synchrony with the pressure
324 records, and maximal oxygen concentrations were typically around 40% air saturation.
325 Sometimes, as a result of water injection occurring at some distance from the optode, the
326 detection of oxygen by the optode was delayed relative to the onset of porewater pressurization
327 (Fig. 4A).

328

329 The differences in feeding bout frequencies and interval durations (Table 1; Fig. 2) caused
330 species-specific oxygen dynamics in the sediment (Figs. 4, 5; Appendix video 4). For *M. liliana*,
331 the oxygen delivered during feeding bouts was typically entirely consumed during the resting
332 periods, and oscillations from oxic to anoxic and back to oxic conditions occurred at frequencies
333 of < 3 times per hour, with oxic and anoxic periods typically lasting around 10 min (Figs. 4, 5).
334 With *M. nasuta* oscillations between oxic and anoxic conditions occurred more frequently (up to
335 6 h⁻¹), congruent with the higher feeding bout frequencies documented by porewater pressure
336 sensing for this species (Table 1). In the one occasion when a *M. balthica* specimen irrigated
337 close to the optode, oxygen was present almost continuously, reflecting incomplete oxygen
338 depletion during the short resting period between irrigation bouts.

339

340 4. Modeling intermittent bioadvection and O₂ dynamics

341 Based on the irrigation patterns determined with porewater pressure sensing, the size-specific
342 instantaneous pumping rates from the literature and the oxygen concentrations of the injected

343 water determined with planar optode imaging, we modeled the dynamics of the oxic sediment
344 volume (V_{O_2} , defined as the volume of sediment with porewater O_2 concentrations $>2 \mu M$ and
345 depths >1 cm, i.e., below the depth to which O_2 penetrates by diffusion from the overlying water
346 in our conditions) for the three species at 3 different sediment reactivities (Fig. 6). These
347 reactivities correspond approximately to the range of sediments types that tellinids inhabit - from
348 permeable clean sands to low permeability muddy sands.

349

350 **a. Intermittent irrigation by tellinids.** There was an increase in V_{O_2} during each irrigation bout
351 and a decrease after the irrigation stopped. Because of the long time intervals between irrigation
352 bouts, the injection pocket of *Macomona liliana* experienced oxic to fully anoxic oscillations
353 regardless of their oxygen consumption rates (Fig. 6A). For *Macoma nasuta*, model simulations
354 predicted oxic-anoxic oscillatory conditions only for sediment with high and intermediate
355 reactivity (Fig. 6B). For *Macoma balthica* oxygen was entirely consumed between subsequent
356 irrigation bouts only in highly reactive sediments, while in sediments of intermediate and low
357 reactivity portions of the irrigated pocket remained oxic due to the short intervals between
358 subsequent irrigation intervals (Fig. 6C). Thus, the spatio-temporal patterns of oxygen
359 distributions largely depend on the species- and size-specific irrigation patterns in relation to the
360 sedimentary oxygen consumption rates.

361

362 **b. Spatial extent of oxygenation.** Model simulations showed that the maximal volume of the
363 oxic sphere, $V_{O_{2max}}$, depends on both the instantaneous pumping rate, Q , and the sediment
364 reactivity, K_{OX} (Fig. 7). A relationship between these quantities derived from the model
365 simulations was $V_{O_{2max}} = m \times Q / K_{OX}$, where the proportionality constant $m = 81.5 \mu M$ (Fig. 8).

366 This relationship is consistent with the idea that, at steady state, the amount of oxygen injected
 367 into the sediment per unit time ($Q \times O_2^{\text{injected}}$, where O_2^{injected} is the oxygen concentration in the
 368 injected water) is equal to the rate at which O_2 is consumed in the oxic sediment volume. For a
 369 low K_M value (1 μM), the rate of O_2 consumption in the sediment is approximately constant and
 370 equal to K_{OX} for a wide range of relevant O_2 concentrations. Therefore, the consumption rate can
 371 be approximated by $V_{O2\text{max}} \times K_{OX}$, resulting in

$$372 \quad V_{O2\text{max}} \approx (Q \times O_2^{\text{injected}}) / K_{OX}. \quad (2)$$

373 Indeed, $O_2^{\text{injected}} = 88 \mu\text{M}$ in this expression closely approximates the proportionality constant m
 374 $= 81.5 \mu\text{M}$ derived from the model results. The small difference, attributed to the combined
 375 effects of neglecting diffusive transport, neglecting reaction kinetics at low O_2 concentrations,
 376 and the use of a 2 μM threshold for defining oxic sediment, indicates that the role of these effects
 377 is minor.

378

379 Our experimental data support this estimate for the oxic sediment volume. For $O_2^{\text{injected}} = 88 \mu\text{M}$,
 380 $K_{OX} = 16 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$, and $Q = 1.6 \text{ mL min}^{-1}$, Eq. 2 predicts a $V_{O2\text{max}}$ of 8.8 cm^3 . Under these
 381 conditions we detected an intermittently oxic area of 4 cm^2 when *M. liliانا* was injecting water
 382 into the sediment close to the O_2 sensitive foil (Fig. 5). The ratio of the predicted (intermittently)
 383 oxic volume and the measured oxic area suggests that the detected area extended roughly 2.2 cm
 384 into the sediment away from the oxygen sensitive foil, which seems reasonable, given the
 385 uncertainties with respect to the shape of the oxic pocket (e.g. we do not know if we are seeing a
 386 cross-section through the center or through the outer part of the oxic pocket).

387

388 *c. Temporal dynamics of oxygenation.* Given the intermittent irrigation activity of tellinids, the
 389 questions arise as to whether V_{O2max} is reached over a single irrigation bout and whether the oxic
 390 pocket becomes fully anoxic between subsequent irrigation bouts.

391

392 The metric of interest for redox oscillation is the fraction of V_{O2max} that is reached after a given
 393 duration of pumping and not the size of V_{O2max} . The size of V_{O2max} depends on the instantaneous
 394 pumping rate (Fig. 8; Eq. 2); however, the fraction of V_{O2max} reached after a given duration of
 395 pumping does not (Fig. 9 A and B). The fraction of V_{O2max} that is reached after a given duration
 396 of pumping does depend on K_{OX} (Fig. 9A). Furthermore, as shown by the solid lines in Fig. 9A,
 397 the increase of V_{O2} towards V_{O2max} is well approximated by

$$398 \quad V_{O2} / V_{O2max} \approx A \times \{1 - \exp[(-K_{OX} \times T_{PUMP}) / (O_2^{injected} \times \phi)]\} + B, \quad (3a)$$

399 where T_{PUMP} is the time since the start of the pumping activity. The fitting parameters A and B
 400 depend on K_{OX} approximately as

$$401 \quad A \approx a_0 + a_1 K_{OX} + a_2 K_{OX}^2, \quad (3b)$$

$$402 \quad B \approx b_0 + b_1 K_{OX}, \quad (3c)$$

403 where $a_0 = 1.444$, $a_1 = -3.985 \times 10^{-3} \text{ L}_s \text{ min } \mu\text{mol}^{-1}$, $a_2 = 5.16 \times 10^{-5} \text{ L}_s^2 \text{ min}^2 \mu\text{mol}^{-2}$, $b_0 = -0.01$,
 404 $b_1 = 1.06 \times 10^{-3} \text{ L}_s \text{ min } \mu\text{mol}^{-1}$. The regression coefficient R^2 for equation 3b is 0.98 and 0.99 for
 405 equation 3c. Model simulations also show that the time required for the volume of oxic sediment
 406 to increase from zero to 95% of V_{O2max} , denoted as T_{O2max} , is also independent of the
 407 instantaneous pumping rate but increases with decreasing values of K_{OX} (Fig. 9C). This
 408 relationship is well approximated by

$$409 \quad T_{O2max} \approx O_2^{injected} \times \phi / K_{OX}. \quad (4)$$

410

Based on the average durations of T_{PUMP} for the tellinids used in our experiments (Table 1), equation 4 allows prediction of the K_{OX} value above which a specific tellinid species and size class will establish 95% of V_{O2max} over the course of a single irrigation bout (see dotted horizontal lines in Fig. 9C). For example, at $K_{\text{OX}} > 20 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ even the short pumping intervals of *Macoma balthica* will establish 95% of V_{O2max} , while at $K_{\text{OX}} < 5 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ none of the tellinids in our experiments would be, on average, pumping sufficiently long to reach 95% of V_{O2max} . Similarly, equations 3a-c allow prediction of the fraction of V_{O2max} reached by pumping water with oxygen concentration $\text{O}_2^{\text{injected}}$ into permeable sediment of a given reactivity K_{OX} for a given interval T_{PUMP} , provided that the injection pocket at the start of pumping is fully anoxic. For example, for $\text{O}_2^{\text{injected}} = 88 \mu\text{M}$, the volume of the oxic sediment induced by pumping for 2 min would be 50% of V_{O2max} for $K_{\text{OX}} = 9 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ but 80% of V_{O2max} for $K_{\text{OX}} = 18 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ (Fig. 9A). It should be noted that these approximations are valid only for such combinations of values of K_{OX} , T_{PUMP} and $\text{O}_2^{\text{injected}}$ that the predicted ratio $V_{\text{O2}} / V_{\text{O2max}}$ falls within the range 1% to 95%.

425

Model simulations show that the fraction of V_{O2max} that is still oxic after a given duration of no irrigation (T_{REST}) does not depend on the instantaneous pumping rate but only on K_{OX} (Fig. 9B), and that it is well approximated by

$$V_{\text{O2}} / V_{\text{O2max}} \approx A \times \{1 - (T_{\text{REST}} \times K_{\text{OX}}) / (\text{O}_2^{\text{injected}} \times \phi)\} + B \quad (5)$$

The fitting parameters A and B depend on K_{OX} approximately as Eq 3b and Eq 3c, where $a_0 = 1.005$, $a_1 = 1.573 \times 10^{-4} \text{L}_s \text{min} \mu\text{mol}^{-1}$, $a_2 = 2.62 \times 10^{-5} \text{L}_s^2 \text{min}^2 \mu\text{mol}^{-2}$, $b_0 = 0.012$, $b_1 = -2.93 \times 10^{-3} \text{L}_s \text{min} \mu\text{mol}^{-1}$. The regression coefficient R^2 is 0.99 for both.

433

Likewise, the time it takes for the oxic sediment to decrease from V_{O2max} to a volume $< 1\%$ of V_{O2max} , denoted as T_{O2zero} , is independent of V_{O2max} , depends only on K_{OX} (Fig. 9D), and is well approximated by (Fig. 9D)

$$T_{O2zero} \approx O_2^{injected} \times \phi / K_{OX}. \quad (6)$$

Thus, based on the average durations of T_{REST} for the tellinids used in the experiments (Table1), equation 6 allows prediction of the K_{OX} value above which oxygen is completely consumed in the course of an average resting interval for any of the tellinid species and size classes (see dotted lines in Fig. 9D). For example, at $K_{OX} > 23 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ even the short resting interval of *Macoma balthica* is sufficiently long to allow complete oxygen depletion, while at $K_{OX} < 3.7 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ none of the tellinids in our experiments would be, on average, resting sufficiently long to allow complete oxygen depletion in the injection pocket. Similarly, equation 5 allows prediction of the fraction of V_{O2max} reached in a sediment of a given reactivity K_{OX} after resting for a given interval T_{REST} , provided that $V_{O2} = V_{O2max}$ at the start of resting. For example, after 2 min of resting the fraction would be about 55% for $K_{OX} = 9 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ but only about 10 % for $K_{OX} = 18 \mu\text{mol L}_s^{-1} \text{min}^{-1}$ (Fig. 9B).

5. Discussion

a. Intermittent irrigation and redox oscillations in permeable sediments. Bioturbation creates heterogeneous and dynamic geochemical conditions in sediments (Aller, 1994), but our ability to quantify and predict the spatio-temporal dynamics is still limited, especially in advection-dominated systems. In these environments infaunal activities can be a predominant driver of porewater transport (Meysman *et al.*, 2005; Volkenborn *et al.*, 2007; Wetthey *et al.*, 2008) that

457 brings oxygenated fluids into contact with suboxic or anoxic sediment. Models of continuous
458 bioirrigation adequately predict gradients in porewater solutes (Aller, 1980; Na *et al.*, 2008), but
459 they do not allow characterization of the temporal dynamics of the geochemical conditions
460 associated with infaunal activity. Intermittent irrigation is not unique to tellinids, but is
461 characteristic for many different types of infauna, such as arenicolid polychaetes (Kristensen,
462 2001; Timmermann *et al.*, 2006; Volkenborn *et al.*, 2010), nereid polychaetes (Foster-Smith,
463 1978; Kristensen, 2001; Pischedda *et al.*, 2012), terebellid polychaetes (Aller and Yingst 1987),
464 thalassinid crustaceans (Forster and Graf, 1995; Volkenborn *et al.*, 2012) or chironomid insect
465 larvae (Polerecky *et al.*, 2006; see also review by Riisgård and Larsen, 2005). Thus, periodic
466 oxygen supply is the rule rather than the exception in bioturbated sediments. The resulting
467 spatial-temporal patterns in redox conditions are largely driven by (i) durations and frequencies
468 of active irrigation intervals, (ii) instantaneous pumping rates, and (iii) sediment reactivity, e.g.,
469 sedimentary oxygen consumptions rates. Given the potential effects of redox oscillations on
470 organic matter mineralization as well as the probable interruption of anoxic or oxic pathways as
471 the oscillation occurs, it is critical to explore the factors constraining stable or oscillatory redox
472 conditions.

473

474 The instantaneous pumping rates of individuals constrain the maximal sediment volumes
475 (V_{O2max}) that are supplied with oxygen (Figs. 7B and 8A), but they do not affect the temporal
476 dynamics of the intermittently oxic pockets. Model simulations indicate that the wax and wane
477 of the oxic pockets are solely governed by the temporal irrigation patterns (T_{PUMP} and T_{REST}), the
478 sedimentary oxygen consumption rates K_{OX} , and the oxygen concentrations in the irrigation
479 water (Fig. 7A). The model results (Figs. 8 and 9) and the analytical approximations (equations

2-6) allow predictions of the volumes of sediment that are supplied with oxygen as well as of the relative proportion of sediment that experiences oscillatory or continuously oxic conditions for a wide range of sediment types and temporal irrigation patterns.

Overall, model simulations suggest that over a wide range of sediment types (i.e., with $K_{OX} > 23 \mu\text{mol L}_s^{-1} \text{min}^{-1}$) the volumes of subsurface sediment pockets that become oxic as a result of intermittent irrigation activity by the studied tellinids are similar to those derived from continuous irrigation models, and that the entire pockets switch between oxic and anoxic conditions during an average T_{PUMP} - T_{REST} cycle. In a typical muddy sand sediment ($\phi = 0.45$; $K_{OX} = 25 \mu\text{mol L}_s^{-1} \text{min}^{-1}$; Volkenborn *et al.*, 2010) with 50 tellinids m^{-2} , the intermittent injection of water with an instantaneous pumping rate of $1.6 \text{ mL min}^{-1} \text{ind.}^{-1}$ (Meyerhöfer, 1985) with an oxygen concentration of $88 \mu\text{M}$ will result in a sediment volume of approximately $280 \text{ cm}^3 \text{ m}^{-2}$ that oscillates between oxic and anoxic conditions several times per hour.

However, short durations of T_{PUMP} and T_{REST} in combination with low reactivities of the sediment make it more likely that at the end of T_{PUMP} the oxic sediment pocket is not in steady-state and that oxygen is not fully consumed at the end of T_{REST} (Figs. 9 A and B). Under these conditions, different irrigation patterns with similar time-integrated pumping rates may have very different geochemical consequences. For example, in a sediment with $K_{OX} = 18 \mu\text{mol L}_s^{-1} \text{min}^{-1}$, irrigation patterns of $T_{PUMP} = 3 \text{ min}$ and $T_{REST} = 3 \text{ min}$ will result in an oscillating sediment volume of $V_{O2\text{max}}$, whereas for $T_{PUMP} = 1 \text{ min}$ and $T_{REST} = 1 \text{ min}$ the maximal volume of oxygenated sediment will be about 50% smaller and $< 50\%$ of this volume will be oscillating (Fig. 9A and B).

503

504 Given the variability of T_{PUMP} for the studied tellinid individuals (Fig. 3), the maximum volume
505 of oxygenated sediment (V_{O2max} ; see Eq. 2) will not be established during some of the shorter
506 irrigation bouts (compare Figs. 3 and 9), which is congruent with our observation that oxic-
507 anoxic oscillations are less frequent in the outer region of the oxygenated pockets (see images of
508 F_{REDOX} for *M. nasuta* and *M. liliana* in Fig. 5). Similarly, given the temporal variability of T_{REST}
509 (Fig. 3), some intervals of resting will be too short for the sediment to become fully anoxic
510 before the onset of the next pressurization interval. This is apparent in the optode imagery for *M.*
511 *nasuta* during minute 20 to 35, when T_{REST} intervals were short, in contrast to minute 45 to 60
512 min, when T_{REST} intervals were longer and there was an anoxic interval between the subsequent
513 feeding bouts (Fig. 4).

514

515 The tellinids were all engaged in the same suite of hydraulic behaviors, but the temporal patterns
516 of these activities and, consequently, their geochemical impact varied among species and size
517 classes. Besides species-specific aspects it was the size of the tellinids that constrained the
518 duration of T_{PUMP} and T_{REST} . Very likely, the smaller gills provide less space to accumulate
519 particles during deposit-feeding, which forces smaller individuals to expel particles more
520 frequently. Given that the geochemical dynamics are directly linked to the durations of pumping
521 and resting intervals, the geochemical signature of an individual is likely to change also as a
522 result of behavioral response to the environmental conditions. In our experiments, tellinids were
523 deposit-feeding most of the time (Table 1), but tellinid behavior is known to vary depending on
524 the environmental conditions. Deposit-feeding dominates in fine sediments (Olafson, 1986) and
525 at low water flow velocities (Levinton, 1991), while the time spent suspension feeding increases

526 at high population densities (Marinelli and Williams, 2003) and high food availability in the
527 water column (Hummel, 1985; Lin and Hines, 1994). *Macoma balthica* shows reversible
528 morphological changes in response to changes in food conditions, with smaller gill (particle
529 collection) to palp (particle sorting) ratios in finer sediments (Drent *et al.*, 2004). The location of
530 subsurface water injection can also change with environmental conditions. The burrowing depth
531 of tellinids is affected by animal size, length of the incurrent siphon, sediment type, season, and
532 the presence of predators (Zwarts and Wanink, 1989; Tallqvist, 2001). In our narrow aquaria the
533 excurrent siphons were often located in the upper 4 cm of sediment, and it may well be that the
534 constrained conditions in ant-farm and container settings as well as the relatively short duration
535 of experiments may have limited burrowing depth.

536

537 We did not find a significant difference in hydraulic activity in response to sediment type, but for
538 some individuals we occasionally observed periods of irrigation without obvious uptake of
539 sediment particles. The duration of sediment pressurization increased during such periods. For
540 example, for one *M. nasuta* individual that was pumping without particle uptake, the durations of
541 irrigation and resting intervals increased from 5.2 min and 4.9 min while deposit-feeding (Table
542 1) to 20 min and 10 min, respectively (mean of 26 bouts over a 5 hour period). Model
543 simulations predict that at, e.g., $K_{OX} = 4.5 \mu\text{mol L}_s^{-1} \text{ min}^{-1}$ such a change in the irrigation pattern
544 will result in a 1.7 fold larger oxic pocket (Fig. 9A), and instead of continuously oxic conditions
545 within 40 % of the pocket, the entire volume will switch between oxic and anoxic conditions
546 (Fig. 9B). Thus, environmental factors that are known to affect temporal infaunal activity
547 patterns, such as temperature (Stief *et al.*, 2010), oxygen concentrations (Dales *et al.*, 1970;
548 Mangum and Burnett, 1975), organism densities (Marinelli and Williams, 2003), flow velocities

549 (Levinton, 1991), food availability in the water column (Hummel, 1985; Lin and Hines, 1994),
550 the presence of predators and injury (Kamermans and Huitema, 1994), or a combination of these
551 factors (e.g., Kolar and Rahel, 1993), are also likely to influence the magnitude and dynamics of
552 sediment oxygenation. The relatively low variability in the means of T_{PUMP} and T_{REST} in tellinids
553 of the same species and size class (Fig. 3, Table 1) suggests that the observed irrigation patterns
554 were robust representatives of species- and size class- specific activity patterns in our
555 experimental settings. However, given the responsiveness of tellinids to a wide range of
556 environmental variables, such external factors may, via their impact on infaunal behavior,
557 ultimately also impact the geochemical dynamics in the sediment.

558

559 ***b. Simplifying model assumptions.*** Model simulations enabled us to show that the interplay
560 between biological parameters (instantaneous pumping rates, temporal patterns of irrigation,
561 efficiency of respiratory O_2 uptake by the animal) and the sediment characteristics (sedimentary
562 oxygen consumption rates, sediment porosity) result in specific oxic-anoxic oscillatory
563 conditions in the sediment. It should be noted that O_2 concentrations in the irrigated water as
564 well as porosity of the sediment lie within a relatively narrow range compared to instantaneous
565 pumping rates and sedimentary oxygen consumption rates, which both may vary over 2 orders of
566 magnitude and more. Thus, it is the volumetric instantaneous pumping rate (related to organism
567 size and species), the O_2 consumption rate of the sediment and the duration of irrigation and
568 resting intervals that constrain the oxygenated sediment volume (Figs. 8 and 9). In model
569 simulations we did not account for the spatial – and maybe even more importantly the temporal –
570 variability of K_{OX} . From repeated injections of oxygenated water close to the planar optode and
571 measurements of subsequent decays of oxygen in time, we know that over the first few injections

572 K_{OX} typically decreases, which can be attributed to the initially rapid oxidation of a highly
573 reactive pool of reduced compounds (Polerecky *et al.*, 2005). With repeated injections of oxic
574 water over the course of hours to days, K_{OX} is expected to change as a result of changes in
575 microbial activity and abundance, as well as changes in sediment porosity and organic matter
576 content. Organic rich fecal pellets, for example, accumulate around the injection pocket and are
577 characterized by high nitrifying activity (Henriksen *et al.*, 1983), which will alter oxygen
578 consumption rates and may relate to the repositioning of the bivalve on timescales of days.
579 Dynamic changes of K_{OX} around the injection pockets may have been responsible for some of
580 the mismatches between planar optode measurements and model output data. For example, for
581 very comparable values of K_{OX} the steady-state oxic volume (V_{O2max}) was established for shorter
582 time intervals in one of our measurements (Fig. 4a: *M. liliانا* at $K_{OX} = 16 \mu\text{mol L}_s^{-1} \text{min}^{-1}$) than
583 predicted from model simulations (Fig. 6A: *M. liliانا* at $K_{OX} = 18 \mu\text{mol L}_s^{-1} \text{min}^{-1}$). Estimates of
584 K_{OX} from the oxygen decrease after this tellinid had stopped pumping during the time interval
585 shown in Fig. 4a varied between 8-10 $\mu\text{mol L}_s^{-1} \text{min}^{-1}$. Thus, K_{OX} was lower than our estimate
586 based on water injections in distant anoxic sediment and the model indeed predicted shorter
587 intervals of steady-state oxic sediment volumes at lower values of K_{OX} (Fig. 6).

588

589 K_{OX} varies also seasonally due to changes in temperature, organic content and microbial activity
590 (Glud *et al.*, 2003; Billerbeck *et al.*, 2006; Werner *et al.*, 2006). For example, Werner *et al.*
591 (2006) documented a 20-fold change in sedimentary oxygen consumption rates between summer
592 ($K_{OX} \approx 14 \mu\text{mol L}_s^{-1} \text{min}^{-1}$) and winter ($K_{OX} \approx 0.7 \mu\text{mol L}_s^{-1} \text{min}^{-1}$). As a consequence of such
593 variation in sediment reactivity, the geochemical signature of a species with a specific irrigation
594 pattern may change, e.g., from a mostly continuously oxic pocket at lower K_{OX} (i.e., in winter) to

595 a mostly oscillatory pocket at higher K_{OX} (i.e., in summer). Thus, due to the dynamic and
596 heterogeneous nature of oxygen consumption rates predictions of (intermittently) oxic sediment
597 volumes based on the (average) reactivity of the bulk sediment will only be as accurate as
598 estimates of K_{OX} within this sediment volume.

599

600 Another crucial assumption in our model was homogenous sediment permeability. This resulted
601 in close to spherical oxic pockets in the model. However, the optode data suggest that the
602 tellinids frequently created rather vertical cylinders (Figs. 4, 5), suggesting locally enhanced
603 upward movement of injected water. Visual observations as well as the porewater pressure data
604 (Fig. 1) suggest that tellinids are capable of injecting water at much higher flow rates, e.g.,
605 during defecations. These “jets” can induce sedimentary cracks to the surface (Matsui *et al.*,
606 2011). Furthermore, fecal pellets accumulate over time around the injection pockets, which
607 further alter not only the reactivity but also the physical characteristics of the sediment.
608 Occasionally, we observed suboxic water exiting the sediment, especially when injection
609 occurred at a relatively shallow depth. In concert, these phenomena are likely to impact the shape
610 and size of the oxygenated pockets beyond the factors assessed in the numerical model and
611 analytical expressions presented here.

612

613 6. Conclusions and perspectives

614 Intermittent irrigation by infaunal organisms has been shown to create geochemically dynamic
615 zones in the sediment where oxic and anoxic conditions alternate on the scale of minutes
616 (Volkenborn *et al.*, 2012). This study is a first attempt to integrate the intermittent nature of
617 bioirrigation into a numerical transport-reaction model that allows an exploration of the related

618 redox dynamics. Model simulations as well as the simple analytical approximations derived from
619 these simulations indicate that temporal irrigation patterns in concert with sedimentary oxygen
620 consumption rates play an important role in constraining the intermittently or continuously oxic
621 sediment volumes. Furthermore, the simulations predict that oscillatory rather than stationary
622 redox conditions are prevalent in a wide range of sediments inhabited by tellinid bivalves.

623

624 We expect that the transient availability of oxygen in the sediment has significant implications
625 for biogeochemical processes. Oxic-anoxic oscillations on the scale of days to weeks have been
626 found to increase rates of organic degradation when compared to continuous oxic or continuous
627 anoxic conditions (Aller, 1994; Sun *et al.*, 1993). Though insights into how microbial
628 assemblages respond to fluctuating conditions are limited (Cravo-Laureau *et al.*, 2011), the
629 higher efficiency in organic mineralization is thought to result from the activation of different
630 microbial assemblages during oxic and anoxic conditions (Sun *et al.*, 1993). Here, we have
631 shown that oxic-anoxic oscillations not only occur on the scale of hours to days, e.g., due to
632 repositioning of excurrent siphons, but also on much shorter time scales, with oxic and anoxic
633 conditions alternating up to six times per hour within these ephemeral oxic pockets (Figs. 4, 5).
634 How microbial assemblages deal with these rapid changes is largely unexplored. For example,
635 oxic-anoxic fluctuations on comparable short time-scales occur when oxic surface sediment is
636 ingested by deposit-feeding infauna and is exposed to anoxic conditions within the digestive
637 tract (Stief *et al.*, 2009). The residence time of the sediment in the gut is typically in the range of
638 30 min. It was found that this duration is not sufficient to allow complete denitrification, which
639 resulted in significant release of nitrous oxides. Thus, rapid oscillations between oxic and anoxic
640 conditions not only reduce the time during which aerobic mineralization can take place, they

641 may also affect rates and benthic fluxes. Quantifying the spatial-temporal dynamics in redox
642 conditions is critical for understanding early diagenesis and argues for the use of 2D optical
643 sensors that allow high resolution measurements of chemical species, such as pH (Hulth *et al.*,
644 2002; Zhu *et al.*, 2006, Larsen *et al.*, 2012), ammonia (Strömberg and Hulth, 2005) carbon
645 dioxide (Zhu *et al.*, 2006; Schröder *et al.*, 2007), ferrous iron (Zhu and Aller, 2012) and other
646 porewater trace metals (Stahl *et al.*, 2012). The combination of measurements and transport-
647 reaction models on appropriate spatial-temporal scales that account for the intermittency of
648 infaunal activity are needed to better understand biogeochemical processes in biologically active
649 sediments.

650

651 Acknowledgments and author contributions

652 This publication was supported by the National Science Foundation (OCE 0928002 to SAW and
653 DSW, and OCE 0751882 to CM), the Office of Naval Research (N00014-0310352 to SAW and
654 DSW), National Oceanic and Atmospheric Administration (NA04NOS4780264 to DSW and
655 SAW), National Aeronautics and Space Administration (NNX11AP77 to DSW and SAW) and
656 the Max Planck Society (LP). The Friday Harbor Laboratories of the University of Washington
657 and Wadden Sea Station Sylt of the Alfred Wegener Institute for Polar and Marine Research in
658 the Helmholtz Association provided laboratory and living space as well as access to field sites.
659 Writing of the manuscript was led by NV, CM and LP, and all authors contributed intellectually
660 to this project, which originated with NV, DSW and SAW. Data collection was done by NV,
661 SAW and DSW; data analysis was led by NV; transport-reaction modeling was led by CM, NV
662 and LP; optode data capture and optode and pressure data analysis software were written by LP
663 and DSW; CP, JEH and SFT in New Zealand and AN, JN and CP in Finland (supported by the

664 EU BONUS+ project HYPER to AN) hosted and supported the laboratory work and specimen
665 collection in the field. We thank Robert C. Aller and two anonymous reviewers for valuable
666 comments on this manuscript.

667

668 Appendix

669 Videos 1-3: Time-lapse movies of the tellinid bivalves *Macomona liliana*, *Macoma nasuta* and
670 *Macoma balthica* (72 hour real-time in 27 s; 9600-fold speed). Particle reworking is related to
671 deposit-feeding activity. Occasionally the approximate position of the subsurface excurrent
672 siphon can be deduced from the locations of ongoing particle displacement.

673

674 Video 4: Oxygen dynamics (450-fold speed) induced by intermittent irrigation by the tellinid
675 bivalves *Macomona liliana*, *Macoma nasuta* and *Macoma balthica*, as revealed by planar optode
676 imaging of oxygen.

677 References

- 678 Aller, R. C. 1980. Quantifying solute distributions in the bioturbated zone of marine sediments
679 by defining an average microenvironment. *Geochim. Cosmochim. Acta*, *44*, 1955–1965.
- 680
- 681 Aller, R. C. 1994. Bioturbation and remineralization of sedimentary organic matter: effects of
682 redox oscillation. *Chem. Geol.*, *114*, 331–345.
- 683
- 684 Aller, R. C. and J. Y. Yingst. 1985. Effects of the marine deposit-feeders *Heteromastus filiformis*
685 (Polychaeta), *Macoma balthica* (Bivalvia) and *Tellina texana* (Bivalvia) on averaged
686 sedimentary solute transport, reaction rates, and microbial distributions. *J. Mar. Res.*, *43*, 615–
687 645.
- 688
- 689 Aller, R. C. and J. Y. Yingst. 1978. Biogeochemistry of tube-dwellings: A study of the sedentary
690 polychaete *Amphitrite ornata* (Leidy). *J. Mar. Res.*, *36*, 201–254.
- 691
- 692 Aller, R. C., J. Y. Yingst and W. J. Ullman. 1983. Comparative biogeochemistry of water in
693 intertidal *Onuphis* (Polychaeta) and *Upogebia* (Crustacea) burrows: temporal pattern and causes.
694 *J. Mar. Res.*, *41*, 571–704.
- 695
- 696 Berg, P., S. Rysgaard, P. Funch and M. K. Sejr. 2001. Effects of bioturbation on solutes and
697 solids in marine sediments. *Aquat. Microb. Ecol.*, *26*, 81–94.
- 698

699 Bennett, W. W., P. R. Teasdale, J. G. Panther, D. T. Welsh, H. Zhao, D. F. Jolley. 2012.
700 Investigating arsenic speciation and mobilization in sediments with DGT and DET: a mesocosm
701 evaluation of oxic-anoxic transitions. *Environ. Sci. Technol.*, *46*, 3981–3989.
702
703 Billerbeck, M., U. Werner, L. Polerecky, E. Walpersdorf, D. de Beer and M. Huettel. 2006.
704 Surficial and deep pore water circulation governs spatial and temporal scales of nutrient
705 recycling in intertidal sand flat sediment. *Mar. Ecol. Prog. Ser.*, *326*, 61–76.
706
707 Boudreau, B. P. 2001. Solute transport above the sediment–water interface, *in* The Benthic
708 Boundary Layer, Boudreau, B .P. and B. B. Jørgensen, eds., Oxford University Press, New York,
709 104–126.
710
711 Boudreau, B. P. and R. L. Marinelli. 1994. A modeling study of discontinuous biological
712 irrigation. *J. Mar. Res.*, *52*, 947–968.
713
714 Caradec, S., V. Grossi, F. Gilbert, C. Guigue and M. Goutx. 2004. Influence of various redox
715 conditions on the degradation of microalgal triacylglycerols and fatty acids in marine sediments.
716 *Org. Geochem.*, *35*, 277–287.
717
718 Cravo-Laureau, C., G. Hernadez-Raquet , I. Vitte, R. Jézéquel, V. Bellet, J. J. Godon, P.
719 Caumette, P. Balaguer and R. Duran. 2011. Role of environmental fluctuations and microbial
720 diversity in degradation of hydrocarbons in contaminated sludge. *Res. Microbiol.*, *162*, 888–895.
721

722 Dales, R. P., C. P. Mangum and J. C. Tichy. 1970. Effects of changes in oxygen and carbon
 723 dioxide concentrations on ventilation rhythms in onuphid polychaetes. J. Mar. Biol. Ass. U.K.,
 724 50, 365–380.
 725
 726 D’Andrea, A. F. and T. H. DeWitt. 2009. Geochemical ecosystem engineering by the mud
 727 shrimp *Upogebia pugettensis* (Crustacea: Thalassinidae) in Yaquina Bay, Oregon: density–
 728 dependent effects on organic matter remineralization and nutrient cycling. Limnol. Oceanogr.,
 729 54, 1911–1932.
 730
 731 Drent, J., P. C. Luttikhuisen and T. Piersma. 2004. Morphological dynamics in the foraging
 732 apparatus of a deposit feeding marine bivalve: phenotypic plasticity. Funct. Ecol., 18, 349–356.
 733
 734 Engel, M., A. Behnke, J. Klier, C. Buschbaum, N. Volkenborn and T. Stoeck. 2012. Effects of
 735 the bioturbating lugworm *Arenicola marina* on the structure of benthic protistan communities.
 736 Mar. Ecol. Prog. Ser., 471, 87–99.
 737
 738 Forster, S. and G. Graf. 1995. Impact of irrigation on oxygen flux into the sediment: intermittent
 739 pumping by *Callianassa subterranea* and “piston-pumping” by *Lanice conchilega*. Mar. Biol.,
 740 123, 335–346.
 741
 742 Foster-Smith, R. L. 1978. An analysis of water flow in tube-living animals. J. Exp. Mar. Biol.
 743 Ecol., 34, 73–95.
 744

745 Gallucci, V. F. and J. Hylleberg. 1976. A quantification of some aspects of growth in the bottom-
 746 feeding bivalve *Macoma nasuta*. *Veliger*, 19, 59–67.

747

748 Glud, R. N. 2008. Oxygen dynamics in marine sediments. *Mar. Biol. Res.*, 4, 243–289.

749

750 Glud, R. N., J. K. Gundersen, H. Røy and B. B. Jørgensen. 2003. Seasonal dynamics of benthic
 751 O₂ uptake in a semi-enclosed bay: importance of diffusion and fauna activity. *Limnol.*
 752 *Oceanogr.*, 48, 1265–1276.

753

754 Henriksen, K., M. B. Rasmussen and A. Jensen. 1983. Effect of bioturbation on microbial
 755 nitrogen transformations in the sediment and fluxes of ammonium and nitrate to the overlying
 756 water. *Ecol. Bull.*, 35, 193–205.

757

758 Hulth, S., R. C. Aller, P. Engstrom, and E. Selander. 2002. A pH plate fluorosensor (optode) for
 759 early diagenetic studies of marine sediments. *Limnol. Oceanogr.*, 47, 212–220.

760

761 Hummel, H. 1985. Food intake of *Macoma balthica* (Mollusca) in relation to seasonal changes in
 762 its potential food on a tidal flat in the Dutch Wadden Sea. *Neth. J. Sea Res.*, 19, 52–76

763

764 Kamermans, P. and H. J. Huitema. 1994. Shrimp (*Crangon crangon* L.) browsing upon siphon
 765 tips inhibits feeding and growth in the bivalve *Macoma balthica* (L.). *J. Exp. Mar. Biol. Ecol.*,
 766 175, 59–75.

767 Karlson, K., E. Bonsdorff and R. Rosenberg. 2007. The impact of benthic macrofauna for
 768 nutrient fluxes from Baltic Sea sediments. *Ambio*, 36, 161–167.
 769
 770 Klute, A. and C. Dirksen. 1986. Hydraulic conductivity and diffusivity: laboratory methods, *in*
 771 Methods of Soil Analysis, Part 1, Physical and Mineralogical Methods, A. Klute, ed., American
 772 Society of Agronomy, Madison, WI, 687–700.
 773
 774 Kolar, C. S. and F. J. Rahel. 1993. Interaction of a biotic factor (predator presence) and an
 775 abiotic factor (low oxygen) as an influence on benthic invertebrate communities. *Oecologia*, 95,
 776 210–219.
 777
 778 Kristensen, E. 1988. Benthic fauna and biogeochemical processes in marine sediments:
 779 microbial activities and flux, *in* Nitrogen Cycling in Coastal Marine Environments, Blackburn,
 780 T.H. and J. Sørensen, eds., Wiley, New York, 275–299.
 781
 782 Kristensen, E. 2001. Impact of polychaetes (*Nereis* spp. and *Arenicola marina*) on carbon
 783 biogeochemistry in coastal marine sediments. *Geochem. Trans.*, 2, 92–103.
 784
 785 Kristensen E., M. H. Jensen and R. C. Aller. 1991. Direct measurement of dissolved inorganic
 786 nitrogen exchange and denitrification in individual polychaete (*Nereis virens*) burrows. *J. Mar.*
 787 *Res.*, 49, 355–377.
 788

789 Larsen, M., S. M. Borisov, B. Grunwald, I. Klimant and R. N. Glud. 2012. A simple and
 790 inexpensive high resolution color ratiometric planar optode imaging approach: application to
 791 oxygen and pH sensing. *Limnol. Oceanogr.: Methods*, 9, 348–360.
 792
 793 Le Bars, M. and M. Grae Worster. 2006. Interfacial conditions between a pure fluid and a porous
 794 medium: implications for binary alloy solidification. *J. Fluid Mech.*, 550, 149–173.
 795
 796 Levinton, J. S. 1991. Variable feeding behavior in three species of *Macoma* (Bivalvia:
 797 Tellinacea) as a response to water flow and sediment transport. *Mar. Biol.*, 110, 375–383.
 798
 799 Lin, J. and A. H. Hines. 1994. Effects of suspended food availability on the feeding mode and
 800 burial depth of the Baltic clam, *Macoma balthica*. *Oikos*, 69, 28–36.
 801
 802 Lohrer, A. M., S. F. Thrush, M. M. Gibbs. 2004. Bioturbators enhance ecosystem function
 803 through complex biogeochemical interactions. *Nature*, 431, 1092–1095.
 804
 805 Mangum, C. P. and L. E. Burnett. 1975. The extraction of oxygen by estuarine invertebrates, *in*
 806 *Physiological Ecology of Estuarine Organisms*, F. J. Vernberg, ed., University of South Carolina
 807 Press, Columbia, SC, 147–163.
 808
 809 Marinelli, R. L., C. R. Lovell, S. G. Wakeham, D. B. Ringelberg and D. C. White. 2002.
 810 Experimental investigation of the control of bacterial community composition in macrofaunal
 811 burrows. *Mar. Ecol. Prog. Ser.*, 235, 1–13.

812

813 Marinelli, R.L. and T. J. Williams. 2003. Evidence for density dependent effects of infauna on
814 sediment biogeochemistry and benthic–pelagic coupling in nearshore systems. *Estuar. Coast.*
815 *Shelf Sci.*, 57, 179–192.

816

817 Matsui, G.Y., N. Volkenborn, L. Polerecky, U. Henne, D. S. Wetthey, C. R. Lovell and S. A.
818 Woodin. 2011. Mechanical imitation of bidirectional bioadvection in aquatic sediments. *Limnol.*
819 *Oceanogr.: Methods*, 9, 84–96.

820

821 Meyerhöfer, E. 1985. Comparative pumping rates in suspension–feeding bivalves. *Mar. Biol.*, 85,
822 137–142.

823

824 Meyers, M. B., H. Fossing and E. N. Powell. 1987. Microdistribution of interstitial meiofauna,
825 oxygen and sulfide gradients, and the tubes of macro–infauna. *Mar. Ecol. Prog. Ser.*, 35, 223–
826 241.

827

828 Meysman F. J. R., O. S. Galaktionov, B. Gribsholt and J. J. Middelburg. 2006. Bio-irrigation in
829 permeable sediments: An assessment of model complexity. *J. Mar. Res.*, 64, 589–627.

830

831 Meysman, F. J. R., E. S. Galaktionov and J. J. Middelburg. 2005. Irrigation patterns in permeable
832 sediments induced by burrow ventilation: a case study of *Arenicola marina*. *Mar. Ecol. Prog.*
833 *Ser.*, 303, 195–212.

834

835 Michaud, E., G. Desrosiers, R. C. Aller., F. Mermillod-Blondin, B. Sundby and G. Stora. 2009.
836 Spatial interactions in the *Macoma balthica* community control biogeochemical fluxes at the
837 sediment–water interface and microbial abundances. J. Mar. Res., 67, 43–70.
838
839 Na, T., B. Gribsholt, O. S. Galaktionov, T. Lee and F. J. R. Meysman. 2008. Influence of
840 advective bio-irrigation on carbon and nitrogen cycling in sandy sediments. J. Mar. Res., 66,
841 691–722.
842
843 Olafsson, E. B. 1986. Density dependence in suspension–feeding and deposit–feeding
844 populations of the bivalve *Macoma balthica*: A field experiment. J. Anim. Ecol., 55, 517–526.
845
846 Pischedda, L., P. Cuny, J. L. Esteves, J. –C. Poggiale and F. Gilbert. 2012. Spatial oxygen
847 heterogeneity in a *Hediste diversicolor* irrigated burrow. Hydrobiologia, 680, 109–124.
848
849 Polerecky, L., U. Franke, U. Werner, B. Grunwald and D. de Beer. 2005. High spatial resolution
850 measurement of oxygen consumption rates in permeable sediments. Limnol. Oceanogr.:
851 Methods, 3, 75–85.
852
853 Polerecky, L., N. Volkenborn and P. Stief. 2006. High temporal resolution oxygen imaging in
854 bioirrigated sediments. Environ. Sci. Technol., 40, 5763–5769.
855
856 Powell, A. W. B. 1979. New Zealand Mollusca: marine, land and freshwater shells. Auckland,
857 Collins, 500 pp.

858

859 Reise, K. 1981. High abundance of small zoobenthos around biogenic structures in tidal
860 sediments of the Wadden Sea. *Helgol. Meeresunters.*, *34*, 413–425.

861

862 Riisgård, H. U. and P. S. Larsen. 2005. Water pumping and analysis of flow in burrowing
863 zoobenthos—an overview. *Aquat. Ecol.*, *39*, 237–258.

864

865 Roper, D. S., R. D. Pridmore and S. F. Thrush. 1992. Recruitment to the macrobenthos of
866 *Macomona liliana* (Bivalvia: Tellinidae) in Manukau Harbour, New Zealand. *N. Z. J. Mar.*
867 *Freshwater Res.*, *26*, 385–392.

868

869 Schröder, C. R., L. Polerecky and I. Klimant. 2007. Time-resolved pH/pO₂ mapping with
870 luminescent hybrid sensors. *Anal. Chem.*, *79*, 60–70.

871

872 Schulz, H. D. 2000. Quantification of early diagenesis: Dissolved constituents in marine pore
873 water, *in* Marine geochemistry, Schulz, H. D. and M. Zabel, eds., Springer, New York, 87–128.

874

875 Solan, M., B. D. Wigham, I. R. Hudson, R. Kennedy, C.H. Coulon, K. Norling, H.C. Nilsson and
876 R. Rosenberg. 2004. In situ quantification of bioturbation using time-lapse fluorescent sediment
877 profile imaging (f-SPI), luminophore tracers and model simulation. *Mar. Ecol. Prog. Ser.*, *271*,
878 1–12.

879

880 Specht, D.T. and H. Lee II. 1989. Direct measurement technique for determining ventilation rate
881 in the deposit feeding clam *Macoma nasuta* (Bivalvia, Tellinaceae). Mar. Biol., 101, 211–218.
882

883 Stahl, H., K. W. Warnken, L. Sochaczewski, R. N. Glud, W. Davison and H. Zhang. 2012. A
884 combined sensor for simultaneous high resolution 2-D imaging of oxygen and trace metals
885 fluxes. Limnol. Oceanogr.: Methods, 10, 389–401.
886

887 Stief, P., L. Polerecky, M. Poulsen and A. Schramm. 2010. Control of nitrous oxide emission
888 from *Chironomus plumosus* larvae by nitrate and temperature. Limnol. Oceanogr., 55, 872–884.
889

890 Stief, P., M. Poulsen, L. P. Nielsen, H. Brix and A. Schramm. 2009. Nitrous oxide emission by
891 aquatic macrofauna. PNAS, 106, 4296–4300.
892

893 Strömberg, N. and S. Hulth. 2005. Assessing an imaging ammonium sensor using time correlated
894 pixel-by-pixel calibration. Anal. Chim. Acta., 550, 61–68.
895

896 Sun, M. Y., R. C. Aller, C. Lee and S. G. Wakeham. 2002. Effects of oxygen and redox
897 oscillation on degradation of cell associated lipids in surficial marine sediments. Geochim.
898 Cosmochim. Acta, 66, 2003–2012.
899

900 Sun, M. Y., C. Lee and R. C. Aller. 1993. Laboratory studies of oxic and anoxic degradation of
901 chlorophyll-a in Long Island Sound sediments. Geochim. Cosmochim. Acta, 57, 147–157.
902

903 Tallqvist, M. 2001. Burrowing behaviour of the baltic clam *Macoma balthica*: effects of
 904 sediment type, hypoxia and predator presence. Mar. Ecol. Prog. Ser., 212, 183–191.
 905

906 Timmermann, K., G. T. Banta and R. N. Glud. 2006. Linking *Arenicola marina* irrigation
 907 behavior to oxygen transport and dynamics in sandy sediment. J. Mar. Res., 64, 915–938.
 908

909 Volkenborn, N. and K. Reise. 2006. Lugworm exclusion experiment: responses by deposit
 910 feeding worms to biogenic habitat transformations. J. Exp. Mar. Biol. Ecol., 330, 169–179.
 911

912 Volkenborn, N., S. I. C. Hedtkamp, J. E. E. van Beusekom and K. Reise. 2007. Effects of
 913 bioturbation and bioirrigation by lugworms (*Arenicola marina*) on physical and chemical
 914 sediment properties and implications for intertidal habitat succession. Estuar. Coast. Shelf Sci.,
 915 74, 331–343.
 916

917 Volkenborn, N., L. Polerecky, D. S. Wetthey and S. A. Woodin. 2010. Oscillatory porewater
 918 bioadvection in marine sediments induced by hydraulic activities of *Arenicola marina*. Limnol.
 919 Oceanogr., 55, 1231–1247.
 920

921 Volkenborn, N., L. Polerecky, D. S. Wetthey, T. H. DeWitt and S. A. Woodin. 2012. Hydraulic
 922 activities by ghost shrimp *Neotrypaea californiensis* induce oxic-anoxic oscillations in
 923 sediments. Mar. Ecol. Prog. Ser., 455, 141–156.
 924

925 Vopel, K., D. Thistle and R. Rosenberg. 2003. Effect of the brittle star *Amphiura filiformis*
 926 (Amphiuridae, Echinodermata) on oxygen flux into the sediment. *Limnol. Oceanogr.*, *48*, 2034–
 927 2045.

928

929 Wang, Y. and P. Van Cappellen. 1996. A multicomponent reactive transport model of early
 930 diagenesis: Application to redox cycling in coastal marine sediments. *Geochim. Cosmochim.*
 931 *Acta*, *60*, 2993-3014.

932

933 Werner, U., M. Billerbeck, L. Polerecky, U. Franke, M. Huettel, J. E. E. van Beusekom and D.
 934 de Beer. 2006. Spatial and temporal patterns of mineralization rates and oxygen distribution in a
 935 permeable intertidal sand flat (Sylt, Germany). *Limnol. Oceanogr.*, *51*, 2549–2563.

936

937 Wetthey, D. and S. A. Woodin. 2005. Infaunal hydraulics generate porewater pressure signals.
 938 *Biol. Bull.*, *209*, 139–145.

939

940 Wetthey, D. S., S. A. Woodin, N. Volkenborn and K. Reise. 2008. Porewater advection by
 941 hydraulic activities of lugworms, *Arenicola marina*: a field, laboratory and modeling study. *J.*
 942 *Mar. Res.*, *66*, 255–273.

943

944 Woodin, S.A., R.L. Marinelli and S.M. Lindsay. 1998. Process-specific cues for recruitment in
 945 sedimentary environments: Geochemical signals? *J. Mar. Res.*, *56*, 535–558.

946

947 Woodin, S. A., D. S. Wethey, J. E. Hewitt and S. F. Thrush. 2012. Small scale terrestrial clay
 948 deposits on intertidal sandflats: behavioral changes and productivity reduction. *J. Exp. Mar. Biol.*
 949 *Ecol.*, *413*, 184–191.
 950
 951 Woodin, S. A., D. S. Wethey and N. Volkenborn. 2010. Infaunal hydraulic ecosystem engineers:
 952 cast of characters and impacts. *Integr. Comp. Biol.*, *50*, 176–187.
 953
 954 Zhu, Q. and R. C. Aller. 2012. Two-dimensional dissolved ferrous iron distributions in marine
 955 sediments as revealed by a novel planar optical sensor. *Mar. Chem.*, *136–137*, 14–23.
 956
 957 Zhu, Q., R. C. Aller and Y. Fan. 2006.. Two-dimensional pH distributions and dynamics in
 958 bioturbated marine sediments. *Geochim. Cosmochim. Acta*, *70*, 4933–4949.
 959
 960 Zwarts, L. and J. Wanink. 1989. Siphon size and burying depth in deposit- and suspension-
 961 feeding benthic bivalves. *Mar. Biol.*, *100*, 227–24.

962 Figure legends

963 **Figure 1:** Porewater pressure dynamics associated with hydraulic activities of the tellinid bivalve
964 *Macomona liliana* (shell length 5.0 cm, sediment permeability $4.3 \times 10^{-12} \text{ m}^2$). The 22 hour
965 sequence starts and ends with burrowing episodes. In between, this individual was deposit-
966 feeding, characterized by intermittent pressurization and positive and negative hydraulic pulses
967 associated with feces and pseudofeces expulsions and siphon movements. B and C are
968 enlargements of the pressure data as indicated by the dashed boxes in A.

969

970 **Figure 2:** Intermittent porewater pressurization associated with deposit-feeding by *Macomona*
971 *liliana* (shell length 4.9 cm, permeability $4.3 \times 10^{-12} \text{ m}^2$), *Macoma nasuta* (shell length 4.9 cm,
972 permeability $2.3 \times 10^{-12} \text{ m}^2$) and *Macoma balthica* (shell length 2.3 cm, unknown permeability).
973 Dotted lines indicate baseline hydrostatic pressure ($p=0$).

974

975 **Figure 3:** Temporal characteristics of intermittent porewater pressurization during deposit-
976 feeding by tellinid bivalves. Shown are irrigation characteristics of individuals that belong to
977 different species and size classes in sediment types with different sedimentary oxygen
978 consumption rates (K_{OX} in $\mu\text{mol L}_s^{-1} \text{ min}^{-1}$; 'na' not available). Durations of pressurized (T_{PUMP})
979 and non-pressurized intervals (T_{REST}) are displayed as box-and-whisker plots. Boxes correspond
980 to the 25th and 75th percentiles, vertical lines span the 5th and 95th percentiles and the horizontal
981 lines within the boxes correspond to the median durations. Maximal and minimal durations are
982 indicated by small horizontal bars. Numbers in the middle correspond to the average frequencies
983 of irrigation bouts per hour calculated as $60/(T_{\text{PUMP}} + T_{\text{REST}})$.

984 **Figure 4:** Dynamics of porewater oxygen concentrations and pressure recorded simultaneously
985 during deposit-feeding by two tellinid species in sediments with volumetric oxygen consumption
986 rates of $16 \mu\text{mol L}_s^{-1} \text{min}^{-1}$. The left images are O_2 snapshots while the right images are the
987 vertically orientated time-series of the horizontal profile indicated by dotted lines in the O_2
988 snapshots (a and b). The white lines in the O_2 profile time-series are the time-synchronous
989 porewater pressure records. Pressure plateaus above the hydrostatic baseline pressure (horizontal
990 dotted line) correspond to feeding bouts and result in oxygenation of subsurface sediment.

991

992 **Figure 5:** Representative spatial-temporal patterns of sediment oxygenation during 2–4 hours of
993 deposit-feeding by 3 tellinid species. The images show (from top to bottom) the probability of
994 oxygen being present over the analyzed period (P_{O_2}), the average oxygen concentration over the
995 oxic intervals (c_{O_2}), the frequency of alterations from oxic to anoxic and back to oxic conditions
996 per hour (F_{REDOX}), and the average durations of the oxic (T_{OX}) and anoxic (T_{ANOX}) periods.
997 Sediment reactivity for *M. liliana* and *M. nasuta* was $16 \mu\text{mol L}_s^{-1} \text{min}^{-1}$.

998

999 **Figure 6:** Modeled variations of the oxic sediment volumes (V_{O_2} , defined as the volume of
1000 sediment with porewater O_2 concentrations $>2 \mu\text{M}$ and below a sediment depth of 1 cm) with
1001 intermittent irrigation by tellinids. Simulations were run for 3 different sediment reactivities
1002 (K_{OX}) that approximately correspond to the range of sediment types that these bivalves inhabit –
1003 from permeable clean sand ($K_{\text{OX}} = 4.5 \mu\text{mol L}_s^{-1} \text{min}^{-1}$) to low permeability fine sand ($K_{\text{OX}} = 40$
1004 $\mu\text{mol L}_s^{-1} \text{min}^{-1}$). Simulations are based on the means of the species-specific irrigation patterns
1005 derived from porewater pressure analysis and instantaneous pumping rates from the literature
1006 (*M. liliana*: $Q = 1.6 \text{ mL min}^{-1}$; $T_{\text{PUMP}} = 8.1 \text{ min}$; $T_{\text{REST}} = 10 \text{ min}$; *M. nasuta*: $Q = 1.8 \text{ mL min}^{-1}$;

1007 $T_{\text{PUMP}} = 4.8 \text{ min}$; $T_{\text{REST}} = 3.7 \text{ min}$; *M. balthica*: $Q = 0.35 \text{ mL min}^{-1}$; $T_{\text{PUMP}} = 2.3 \text{ min}$; $T_{\text{REST}} = 1.7$
1008 min). Bars below species names depict the type of activity (white = rest; black = pumping).

1009

1010 **Figure 7:** Modeled variations of the oxic sediment volume, V_{O_2} , around the injection pocket
1011 over a 40 min long period of constant irrigation followed by a resting period sufficiently long to
1012 reach anoxic conditions. A: Variation of V_{O_2} for a specific instantaneous pumping rate (1.5 mL
1013 min^{-1}) at various sediment reactivities. B: Variation of V_{O_2} for a specific sediment reactivity (18
1014 $\mu\text{mol L}_s^{-1} \text{ min}^{-1}$) and various instantaneous pumping rates. The time required to reach 95% of a
1015 steady state oxic volume ($T_{\text{O}_2\text{max}}$) as well as the time required for a complete consumption of
1016 oxygen within the volume $V_{\text{O}_2\text{max}}$ ($T_{\text{O}_2\text{zero}}$) are independent of the pumping rate and are governed
1017 by the reactivity of the sediment only.

1018

1019 **Figure 8:** Modeled relationships between the instantaneous pumping rates (Q) and the maximal
1020 oxic volumes ($V_{\text{O}_2\text{max}}$) established with continuous water injection into sediments of different
1021 reactivities (K_{OX}). K_{OX} values are indicated at the distal end of the lines (A). The slope of the
1022 linear regression $V_{\text{O}_2\text{max}}$ versus Q , denoted as m , depends on K_{OX} and can be described by a
1023 hyperbolic function (B). Symbols correspond to values derived from model simulations, while
1024 solid lines correspond to the analytical approximations shown.

1025

1026 **Figure 9:** Spatio-temporal dynamics of oxygenated pockets induced by intermittent water
1027 injection into sediments of different reactivity. A and B show the fraction of $V_{\text{O}_2\text{max}}$ that is
1028 reached after a given duration of pumping and resting, assuming the initial conditions of $V_{\text{O}_2}=0$
1029 and $V_{\text{O}_2}=V_{\text{O}_2\text{max}}$, respectively. Symbols correspond to means obtained by modeling of the

1030 fraction of V_{O2max} established after certain T_{PUMP} and T_{REST} for 6 different pumping rates ranging
1031 from 0.1 mL min^{-1} to 3 mL min^{-1} ; the error bars, if shown, would be the size of the symbol and
1032 values were not different as a function of pumping rate. Solid lines correspond to analytical
1033 approximations of V_{O2}/V_{O2max} (Eqs. 3 and 5). Panels C and D show the durations of pumping and
1034 resting that are necessary to reach 95 % and 1 % of V_{O2max} (T_{O2max} and T_{O2zero}), respectively.
1035 Filled symbols depict the means of T_{O2max} and T_{O2zero} derived from simulations with the same 6
1036 pumping rates (0.1 mL min^{-1} to 3 mL min^{-1}), while the solid lines correspond to the analytical
1037 approximations of T_{O2max} and T_{O2zero} (Eqs. 4 and 6). Open squares with error bars on the right
1038 side of C and D depict mean durations and standard deviations of tellinid irrigation bouts (T_{PUMP})
1039 and resting intervals (T_{REST}) measured with porewater pressure sensors for different tellinid
1040 species and sizes. Shown are the means of n individuals of large *M. liliana* (n=16), small *M.*
1041 *liliana* (n=7), large *M. nasuta* (n=5), and *M. balthica* (n=3). The horizontal dotted lines through
1042 the means of T_{PUMP} and T_{REST} and the vertical dotted lines from their intersections with the
1043 hyperbolic curves were added for orientation. All simulation results shown used $O_2^{injected} = 88$
1044 μM .

1045 **Table 1:** Tellinids and sediment characteristics during porewater pressure measurements and
1046 derived behavioral time allocation to behaviors (deposit-feeding, burrowing, inactive) and
1047 temporal characteristics of intermittent irrigation during deposit-feeding. 'N' is number of
1048 individuals; 'k' is sediment permeability; 'K_{OX}' is sedimentary volumetric oxygen consumption
1049 rate; 'na' not available, *derived from time-lapse photography of 2 individuals.

Table 1 Volkenborn *et al.*

specimen			sediment characteristics				behavioral time allocation			irrigation patterns during deposit-feeding					
species	N	shell length	k	porosity	K _{ox}		deposit feeding	burrowing	inactive	bout frequency	T _{PUMP}		T _{REST}	bouts analyzed per ind.	
		cm	10 ⁻¹² m ²	vol %	μmol L ⁻¹ min ⁻¹			% of time		h ⁻¹	min		min	n	
		mean			mean	sd					mean	sd	mean	sd	
<i>M. liliana</i>	8	4.7	4.3	0.49	16	2	86.2	6.3	7.5	3.4	7.7	2.3	10.5	1.9	
<i>M. liliana</i>	8	4.7	0.24	0.56	43	11	89.5	6.7	3.7	3.3	8.9	2.7	9.5	1.8	
<i>M. liliana</i>	4	2.4	0.10	0.56	43	11	95.8	0.9	3.2	5.6	4.4	1.0	6.7	1.8	
<i>M. nasuta</i>	2	5.0	7.3	0.43	16	4	93.2	2.5	4.4	6.1	5.2	1.2	4.9	0.7	
<i>M. nasuta</i>	3	5.0	2.3	0.51	31	6	85.8	3.2	11.0	8.0	4.6	0.8	2.9	0.2	
<i>M. balthica</i>	3	2.1	na	na	na	na	*50.4	*0.5	*49.1	15.1	2.3	0.2	1.7	0.2	

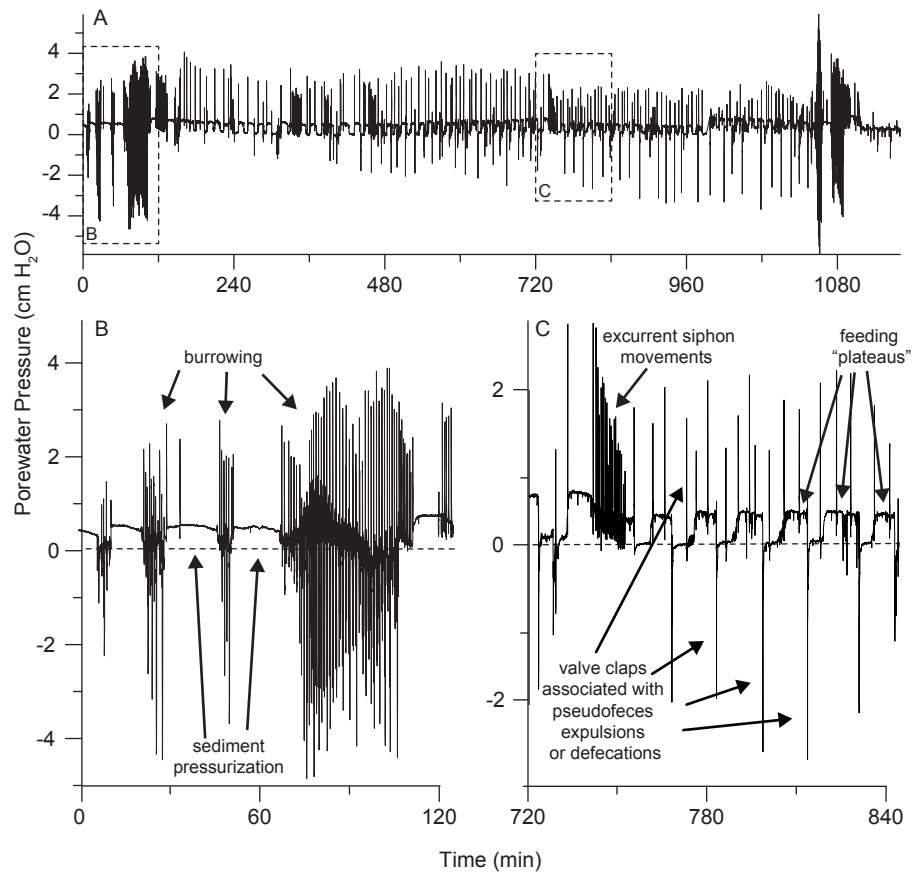


Figure 1 Volkenborn *et al.*

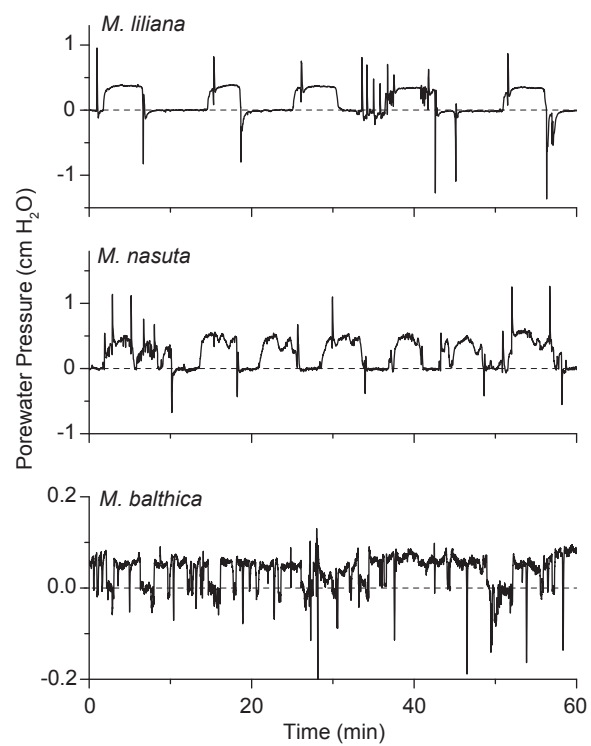


Figure 2 Volkenborn *et al.*

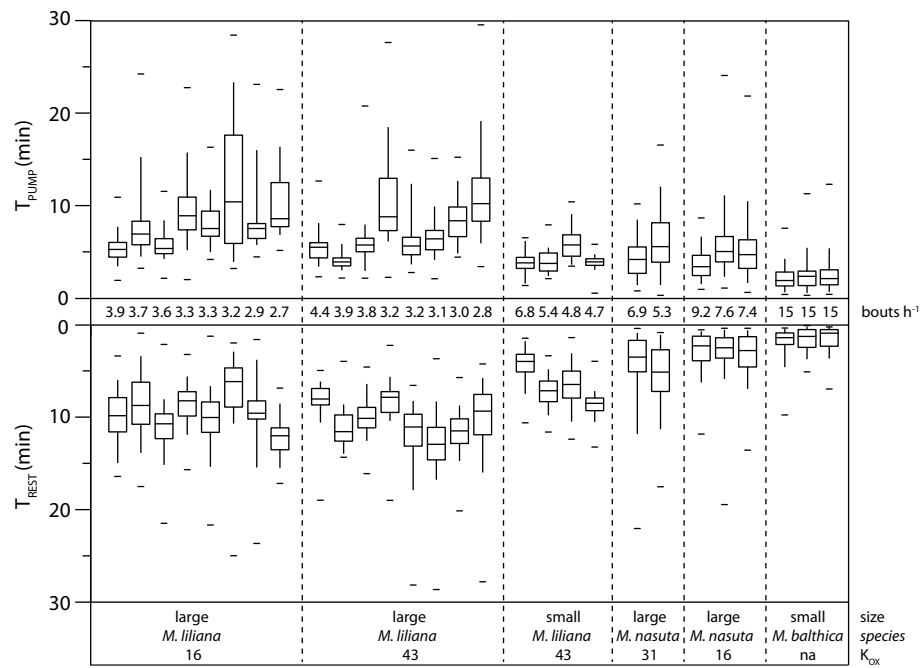


Figure 3 Volkenborn *et al.*

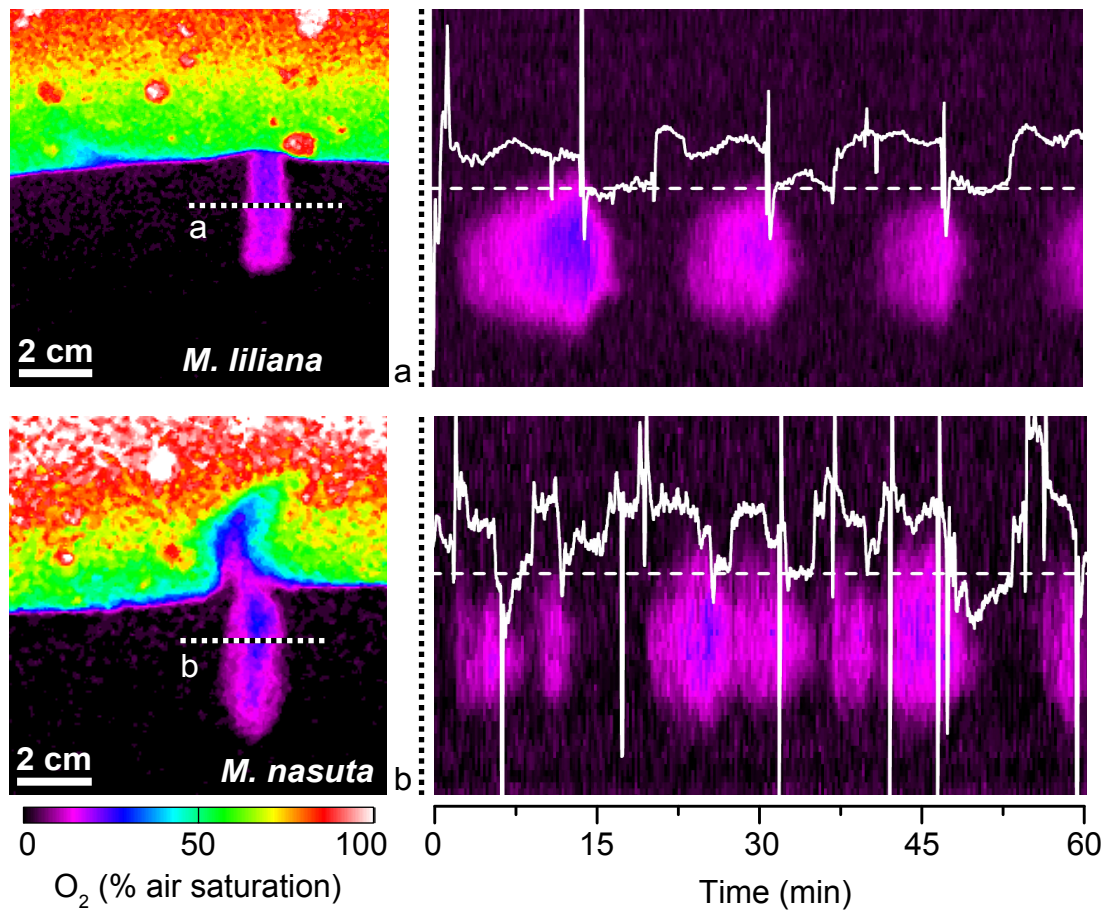


Figure 4 Volkenborn *et al.*

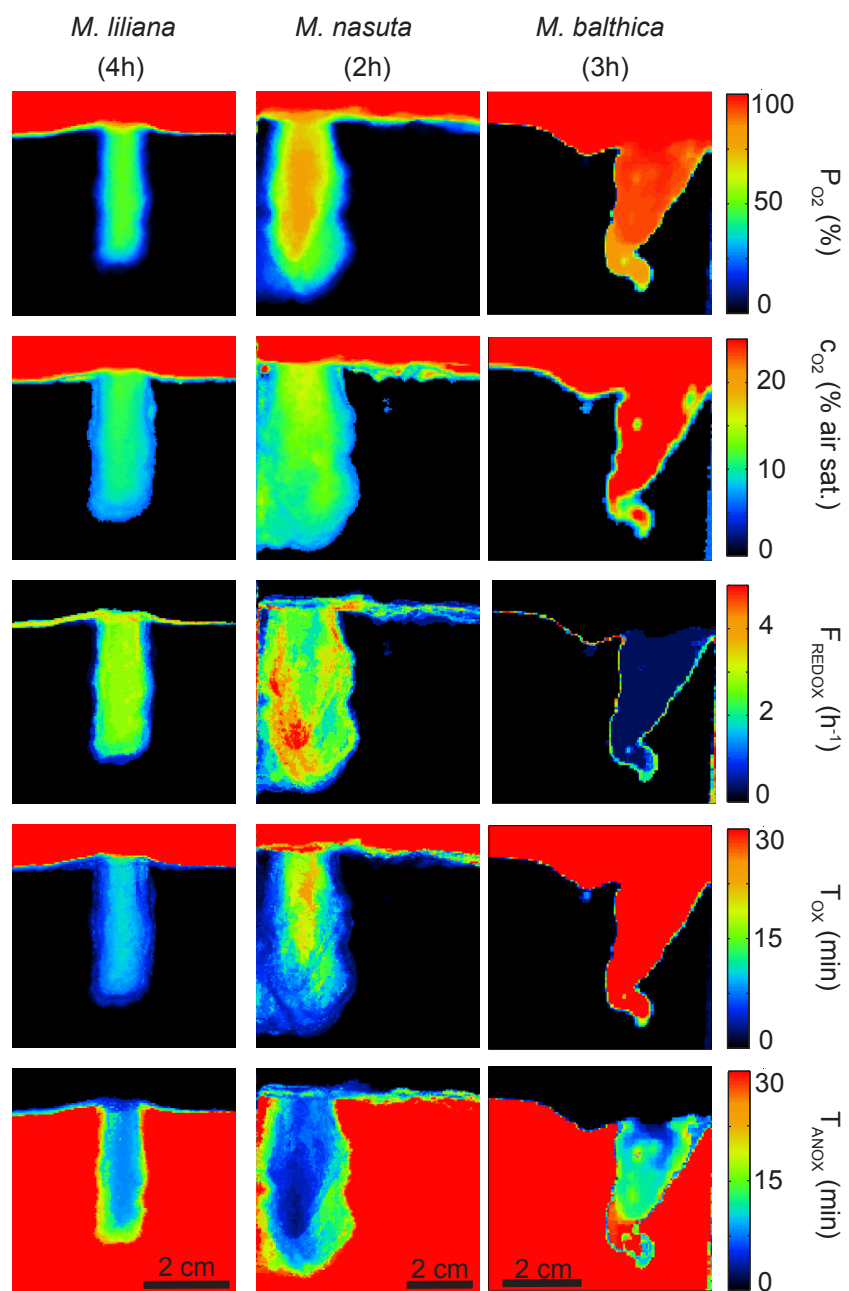


Figure 5 Volkenborn *et al.*

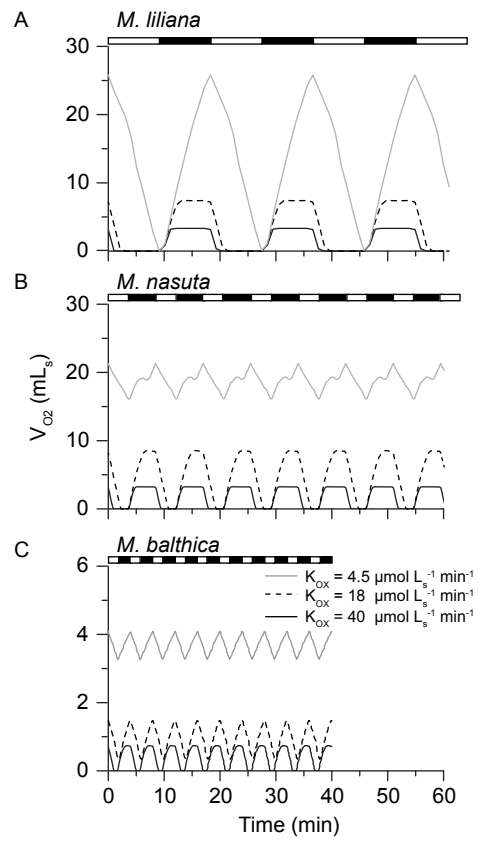


Figure 6 Volkenborn *et al.*

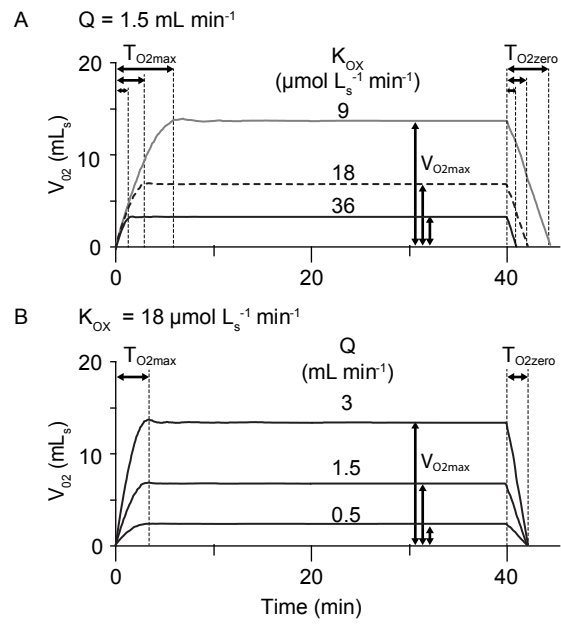


Figure 7 Volkenborn *et al.*

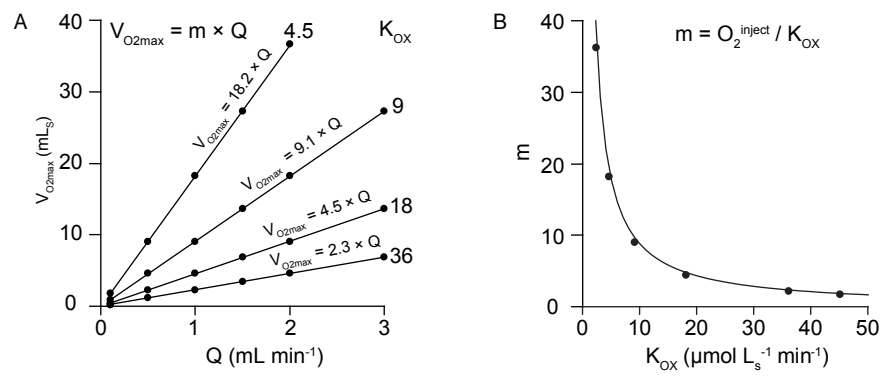


Figure 8 Volkenborn *et al.*

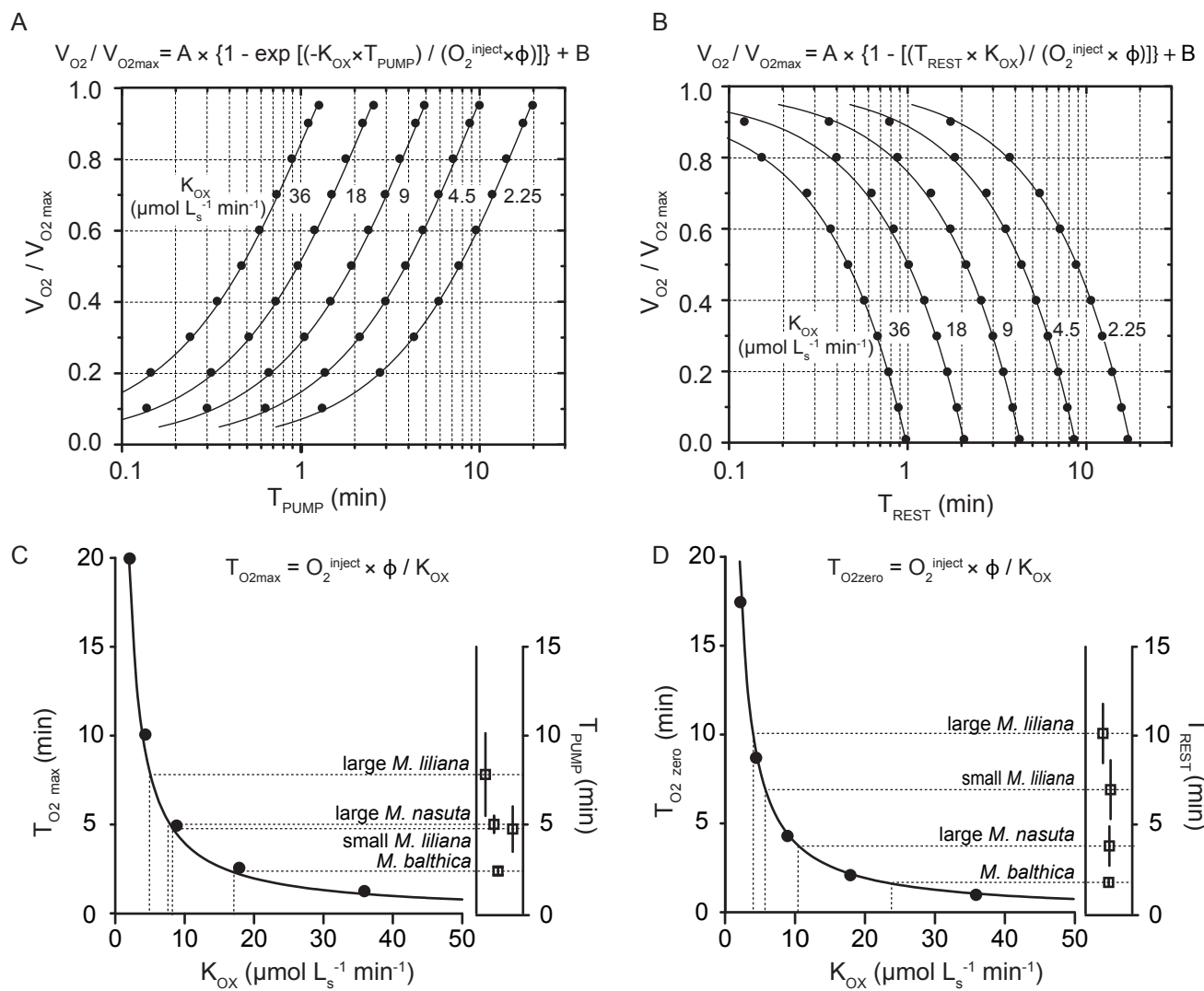


Figure 9 Volkenborn *et al.*