

Assessment of Novel Curcumin Derivatives as Potent Inhibitors of Inflammation and Amyloid- β Aggregation in Alzheimer's Disease

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Abstract

Alzheimer's disease (AD) is the most common neurodegenerative disorder affecting the elderly population worldwide. Brain inflammation plays a key role in the progression of AD. Deposition of senile plaques in the brain stimulates an inflammatory response with the overexpression of pro-inflammatory mediators, such as the neuroinflammatory cytokine. interleukin-6. Curcumin has been revealed to be a potential agent for treating AD following different neuroprotective mechanisms, such as inhibition of aggregation and decrease in brain inflammation. We

synthesized new curcumin derivatives with the aim of providing good anti-aggregation capacity but also improved anti-inflammatory activity. Nine curcumin derivatives were synthesized by etherification and esterification of the aromatic region. From these derivatives, compound **8** exhibited an anti-inflammatory effect similar to curcumin, while compounds **3**, **4**, and **10** were more potent. Moreover, when the anti-aggregation activity is considered, compounds **3**, **4**, **5**, **6**, and **10** showed biological activity *in vitro*. Compound **4** exhibited a strong anti-aggregation effect higher than curcumin. Monofunctionalized curcumin derivatives showed better bioactivity than difunctionalized compounds. Moreover, the presence of bulky groups in the chemical structure of curcumin derivatives decreased bioactivity.

INTRODUCTION

Alzheimer's disease (AD) is the most common neurodegenerative condition, affecting more than 29 million people around the world, a number that is expected to triple by 2050. AD neuropathology is characterized by intraneuronal neurofibrillary tangles and extracellular senile plaques in the brain. Neurofibrillary tangles are composed of hyperphosphorylated tau proteins, while senile plaques originate from amyloid- β (A β) aggregation [1].

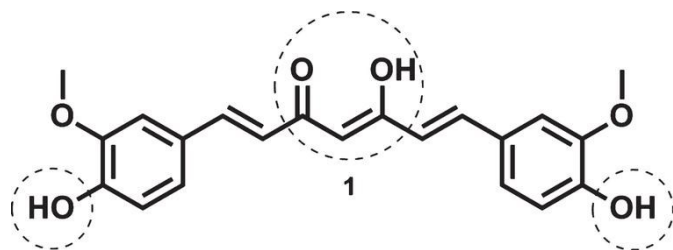
Inflammation is a complex biological response to a harmful stimuli or cell/tissue damage [2, 3]. Chronic brain inflammation is a distinctive feature of AD in which the microglia, astrocytes, and, to a certain extent, neurons are thought to be strongly involved in the inflammatory process. Furthermore, the overexpression of pro-inflammatory mediators, such as tumor necrosis factor - α and interleukin (IL)-6, and acute proteins are evident in different regions of an AD brain [4]. A synergistic pattern between AD senile plaques and pro-inflammatory cytokines increases the neurological damage to the brain [5, 6]. Thus, an increased deposition of A β proteins potentiate the production of pro-inflammatory cytokines, while these cytokines promote the formation of other constitutive proteins of the senile plaques.

The strong association between brain inflammation and AD pathology has stimulated research toward the discovery of new therapeutic agents that are likely to provide benefits to patients with AD. In this sense, the anti-inflammatory activity of natural products, which therefore decrease the impact of AD in patients, has been studied.

Curcumin (Fig. 1), a major polyphenol of the rhizome of *Curcuma longa*, is a potent anti-inflammatory and neuroprotective natural product [1]. Studies *in vitro* have revealed that curcumin inhibits amyloid β -aggregation, the activities of the enzymes β -secretase and acetylcholinesterase, and A β -induced inflammation [7, 8]. *In vivo*, this polyphenol inhibits A β oligomerization, A β deposition, and tau phosphorylation in AD animal models [7, 8].

Fig.1

Curcumin and its major reactive sites.



The anti-inflammatory activity of curcumin is mediated by modulation of several molecules involved in the inflammatory process.

In vitro, curcumin inhibits the production of pro-inflammatory cytokines, regulates the activity of inflammatory enzymes (COX-2, and the inducible nitric oxide synthase), and downregulates the expression of chemokines (MCP-1 and interferon-inducible protein) [9]. Meanwhile, *in vivo* experiments show it regulates the activation of transcription factors such as activating protein-1 and nuclear factor- κ B [9]. The lack of toxicity of curcumin at high concentrations makes it a potential nonsteroidal anti-inflammatory drug. Its low bioavailability, due to susceptibility to degradation in biological systems and poor solubility in water and plasma has, however, prevented the medical use of curcumin [11]. Although it is plausible that the anti-inflammatory activity of curcumin can be improved through chemical modification, there have been only few studies on the synthesis of curcumin analogs with this aim [10–13]. Thus, we sought to design and synthesize new anti-inflammatory curcumin derivatives with a higher anti-inflammatory effect than curcumin and good capacity to inhibit A β aggregation.

MATERIALS AND METHODS

Synthesis

Chemical reagents used were commercially available (Tedia, Applichem, Chem-Impex International, Sigma Aldrich, Oakwood Products, Lancaster Avocado, Alfa-Aesar, Fisher). All reactions were conducted with magnetic stirring under an argon atmosphere in oven-dried flasks. Reactions were monitored until deemed complete by TLC using silica-gel-coated glass plates (Merck Kieselgel 60 F254). Plates were visualized under UV light (254nm). Plates were dyed with 10% phosphomolybdic acid (PMA) in ethanol. ^1H , and ^{13}C NMR spectra were recorded at 500 (^1H), and 125MHz (^{13}C) on an Agilent Inova 500 spectrometer; and at 400 (^1H), 100MHz (^{13}C) on Eclipse 400MHz spectrometer (JEOL, Peabody, MA, USA). Chemical shifts (δ) are reported in parts per million (ppm) using the residual solvent peak and coupling constants (J) are given in Hz. Proton multiplicity is reported as singlet (s), doublet (d), triplet (t), quartet (quart.), quintet (quint.), septet (sept), multiplet (m), and broad (br). Infrared spectrophotometry was carried out on a Platinum ATR Alpha instrument (Brucker, Billerica, MA, USA). The molar masses were determined with a micrOTOF-QIII spectrometer Bruker Daltonics, Billerica, MA, USA), with electrospray ionization (ESI) and positive ion detection mode. The detailed synthetic procedures and spectral characterization are described below.

General procedure (GP1) for the synthesis of alkyl succinates *S1-S3*

An oven-dried round bottom flask was charged with the given alcohol (14.7mmol), dichloromethane (5 mL), and *N,N*-diisopropylethylamine (1.3 mL, 7.35mmol, 0.5 equiv.) at room temperature (RT). After 2h, succinic anhydride (735mg, 7.35mmol, 0.5 equiv.), and 4-

dimethylaminopyridine (448mg, 3.67mmol, 0.25 equiv.) were added, and the reaction stirred at RT. After 48h, the reaction mixture was diluted with brine/1 M HCl (3:1, 10mL). The aqueous layer was extracted with dichloromethane (3×10mL). The combined organic phases were dried over anhydrous sodium sulfate (Na₂SO₄), filtered, and concentrated under reduced pressure. The crude material was washed with hexanes (3×20mL) to obtain the desired product.

Synthesis of 4-(allyloxy)-4-oxobutanoic acid (S1): According to GP1, allyl alcohol (854mg, 14.7mmol) yielded **S1** (790mg, 40%).

Synthesis of 4-(benzyloxy)-4-oxobutanoic acid(S2): According to GP1, benzyl alcohol (3g, 27.7mmol, 2.88mL), pyridine (20mL), 4-dimethylaminopyridine (508mg, 4.15mmol, 0.15 equiv.), and succinic anhydride (1.3g, 13.8mmol, 0.5 equiv.) were combined, and the reaction was stirred for 6h at 100C to yield **S2** (3.74g, 65%).

Synthesis of 4-(cyclopentyloxy)-4-oxobutanoic acid (S3): According to GP1, cyclopentanol (949mg, 11mmol, 1mL) and N,N-diisopropylethylamine (958μL, 5.5mmol, 0.5 equiv.) were stirred in dichloromethane (5mL) at RT. After 2h, succinic anhydride (550mg, 5.5mmol, 0.5 equiv.) and 4-dimethylaminopyridine (672mg, 5.5mmol, 0.5 equiv.) were added and the reaction stirred for 48h at RT to yield **S3** (1g, 50%).

General procedure (GP2) for the synthesis of dialkylcurcumin and monoalkylcurcumin (5–10)

An oven-dried round bottom flask, equipped with magnetic stirrer and 3 Å molecular sieves, was flushed with argon and charged with alkyl succinate (207mg, 1.3mmol, 10 equiv.), pyridine (3mL), 4-dimethylaminopyridine (366mg, 0.39mmol, 3 equiv.), and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (75mg, 0.39mmol, 3 equiv.). The reaction was stirred for 4h at RT. Concurrently, a solution of curcumin (**1**) (50mg, 0.13mmol) in pyridine (3mL) was stirred for 4h at RT. The curcumin solution was then added to the alkyl succinate reaction allowed to stir for 48h at RT. The reaction mixture was diluted with a 0.5 M aqueous solution of Na₂CO₃/brine (1:1, 10mL), and the aqueous layer extracted with ethyl acetate (EtOAc) (3×10mL). The combined organic phases were dried over Na₂SO₄, filtered, concentrated under reduced pressure, and purified by preparative HPLC using normal phase silica gel column (Phenomenex, Sphereclone, 250×10mm, 5μm) with an *n*-hexane to ethyl acetate gradient system in 20min at 2mL/min to obtain the desired products.

Synthesis of diallyl O,O'-(((1E,3Z,6E)-3-hydroxy-5-oxohepta-1,3,6-triene-1,7-diyl)bis(2-methoxy-4,1-phenylene)) disuccinate (5), and allyl (4-((1E,4Z,6E)-5-hydroxy-7-(4-hydroxy-3-methoxyphenyl)-3-oxohepta-1,4,6-trien-1-yl)-2-methoxyphenyl) succinate (6): According to GP2, **S1** (207mg, 0.67mmol, 10 equiv), pyridine (3mL), 4-dimethylaminopyridine (366mg, 0.39mmol, 3 equiv.), and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (75mg, 0.39mmol, 3 equiv.) were combined and stirred at RT. Concurrently, a solution of curcumin (50mg, 0.13mmol) in pyridine (3mL) was stirred at RT. After 4h, the reactions were combined and allowed to stir for 48h at RT. The crude product was purified by HPLC with diode array UV detection (DAD) to obtain **5** (19mg, 22%) and **6** (27mg, 39%).

Synthesis of dibenzyl O,O'-(((1E,3Z,6E)-3-hydroxy-5-oxohepta-1,3,6triene-1,7-diyl)bis(2-methoxy-4,1-phenylene)) disuccinate (7), and benzyl (4-((1E,4Z,6E)-5-hydroxy-7-(4-hydroxy-3-

methoxyphenyl)-3-oxohepta-1,4,6-trien-1-yl)-2-methoxyphenyl) succinate (8) : According to GP2, **S2** (64mg, 1.3mmol, 10 equiv.), pyridine (3mL), 4-dimethylaminopyridine (366mg, 0.39mmol, 3 equiv.), and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (75mg, 0.39mmol, 3 equiv.) were stirred at RT. Concurrently, a solution of curcumin (50mg, 0.13mmol) in pyridine (3mL) was stirred at RT. After 4h, the solutions were combined and allowed to stir for 48h at RT. The crude product was purified by HPLC-DAD to obtain **7** (21mg, 21%) and **8** (28mg, 37%).

Synthesis of dicyclopentyl O,O'-(((1E,3Z,6E)-3-hydroxy-5-oxohepta-1,3,6-triene-1,7-diyl)bis(2-methoxy-4,1-phenylene)) disuccinate (9), and cyclopentyl 4-((1E,4Z,6E)-5-hydroxy-7-(4-hydroxy-3-methoxyphenyl)-3-oxohepta-1,4,6-trien-1-yl)-2-methoxyphenyl) succinate (10) : According to GP2, **S3** (245mg, 1.3mmol, 10 equiv.), pyridine (3mL), 4-dimethylaminopyridine (366mg, 0.39mmol, 3equiv.), and 1-ethyl-3-(3-dimethylamino-propyl)carbodiimide hydrochloride (75mg, 0.39mmol, 3 equiv.) was stirred at RT. Concurrently, a solution of curcumin (50mg, 0.13mmol) in pyridine (3mL) was stirred at RT. After 4h, the solutions were combined and allowed to stir for 48h at RT. The crude product was purified by HPLC-DAD to obtain **9** (17mg, 18%) and **10** (25mg, 34%).

General procedure (GP3) for the synthesis of etherification and esterification of curcumin (2–4)

An oven dried round bottom flask equipped with magnetic stirrer and 3 Å molecular sieves was flushed with argon and charged with curcumin (200mg, 0.54mmol), solvent (6mL), base (150mg, 1.08mmol, 2 equiv.), and alkyl halide (300μL, 2.7mmol, 5 equiv.). The reaction was stirred for 48h at RT. The reaction mixture was diluted with water and extracted with EtOAc (3×10mL). The combined organic phases were dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was washed with *n*-hexane (3×20mL) to obtain the desired product.

Synthesis of (1E,6E)-1,7-bis(3-methoxy-4-(prop-2-yn-1-yloxy)phenyl)hepta-1,6-diene-3,5-dione (2) : According to GP3, curcumin (200mg, 0.54mmol), *N,N*-dimethylformamide (6mL), K₂CO₃ (150mg, 1.08mmol, 2 equiv.), and propargyl bromide (300μL, 2.7mmol, 5 equiv.) were stirred for 48h at RT. The reaction mixture was diluted with water (10mL) and extracted with EtOAc (3×10mL). The combined organic phases were dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was washed with *n*-hexane (3×20mL) to yield **2** (168mg, 70%).

Synthesis of (1E, 4Z, 6E)-1-(4-(benzyloxy)-3-methoxyphenyl)-5-hydroxy-7-(4-hydroxy-3-methoxyphenyl)hepta-1,4,6-trien-3-one (3) : According to GP3, curcumin (100mg, 0.27mmol), acetone (5mL), Cs₂CO₃ (88mg, 0.27mmol, 1 equiv.), and benzyl bromide (1mL, 8.1mmol, 30 equiv.) were stirred at 56°C for 24h. The reaction mixture was diluted with water (20mL) and extracted with EtOAc (3×10mL). The combined organic phases were dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was purified by column chromatography [*n*-hexane/EtOAc, silica gel] to obtain **3** (98mg, 80%).

Synthesis of 4-((1E,4Z,6E)-5-hydroxy-7-(4-hydroxy-3-methoxyphenyl)-3-oxohepta-1,4,6-trien-1-yl)-2-methoxyphenyl acetate (4) : According to GP3, curcumin (100mg, 0.27mmol) and K₂CO₃ (38mg, 0.27mmol, 1 equiv.) in dichloromethane (10mL) were stirred at RT. After 8h, a solution

of acetyl chloride (96 μ L, 1.35 mmol, 5 equiv.) in dichloromethane (5 mL) was sonicated for 5 min and added to the curcumin solution. The reaction was stirred for 24 h at RT followed by column chromatography [*n*-hexane/EtOAc, silica gel] to obtain **4** (79 mg, 71%).

A complete detail of the methods used for synthesis and for the characterization of the synthesized compounds can be found in the [Supplementary material](#).

Mice

Female C57Bl/6 mice, 8 weeks of age, were provided by INDICASAT's animal facility. Animals were maintained in 12 h light/dark cycle at a constant temperature of 24°C with free access to food and water. Experimental procedures were performed following the ethical guidelines related to the handling of lab animals in accordance with international and institutional regulations. The Institutional Animal Care and Use Committee of INDICASAT approved the protocol (IACUC-15-004).

Cell culture and cytokine determination

To determine the anti-inflammatory capacity of compounds (**1–6**, **8**, **10**), elicited peritoneal macrophages from C57Bl/6 mice were obtained by peritoneal washing with chilled RPMI after 5 days of i.p. instillation of 2 mL of 3% thioglycollate. Cells were seeded in RPMI with 10% FCS at 2×10^5 cells/well in 96-well plates and incubated at 37°C in a 5% CO₂ atmosphere. Cells were stimulated with LPS (10 ng/mL) in the presence or absence of different concentrations of compounds (1, 3, 10, and 30 μ M). All the treatments and controls were performed in the presence of 0.5% DMSO, the vehicle for test compounds. Supernatants were collected 18 h after stimulation with LPS, and the concentrations of IL-6 were determined by ELISA. Concentrations of cytokine were measured following the manufacturer's protocol (DuoSet kit, R&D System).

Cytotoxicity assay

To determine the cytotoxicity of the compounds tested (**1–6**, **8**, **10**), 100 μ L of MTT at a concentration of 0.5 mg/mL in RPMI was added to each well after removing the supernatant and were incubated for 2 h at 37°C in 5% CO₂. MTT (soluble tetrazolium salt) is reduced to formazan (insoluble) by the activity of succinate dehydrogenase of living cells' mitochondria. The supernatants were removed, and the formazan crystals were dissolved in 100 μ L of 0.04 M HCl in isopropanol. The color was analyzed at 570 nm using a plate reader. The percentage of viable cells was calculated as % viability = (OD sample/OD control) \times 100%. Non-stimulated cells cultured in media represent 100% viability.

Thioflavin T assay

The aggregation of A β ₄₂ was evaluated by the thioflavin T assay. The rA β ₄₂ (rPeptide) was resuspended according to the manufacturer's instruction in 1% NH₄OH, at a concentration of 1 mg/mL after 1 min of hydration. rA β ₄₂ (10 μ M) was combined with or without different concentrations (1, 3, 10, and 30 μ M) of compounds, and 200 μ L of the mix was plated in a 96-well black plate. For this assay, curcumin (5 μ M) was used as inhibition control. All of the reactions were performed in the presence of 0.1% of DMSO of the vehicle for the compounds. The plate was incubated at 37°C for 48 h. After incubation, 20 μ M of thioflavin T was added and the fluorescence was measured in a Synergy HT multi-reader from Biotek (Winooski, VT),

excitation 450nm, emission 485nm. Fluorescence values were determined by subtracting the baseline fluorescence of thioflavin T.

Statistical analysis

Data were analyzed by using the statistical software package GraphPad Prism5. Statistical analysis was performed with the unpaired *t* test. A significant difference between groups was considered to be when $p < 0.05$. The half maximal inhibitory concentration (IC_{50}) was calculated by adjusting a sigmoidal dose-response curve following the procedure in GraphPad Prism5.

RESULTS

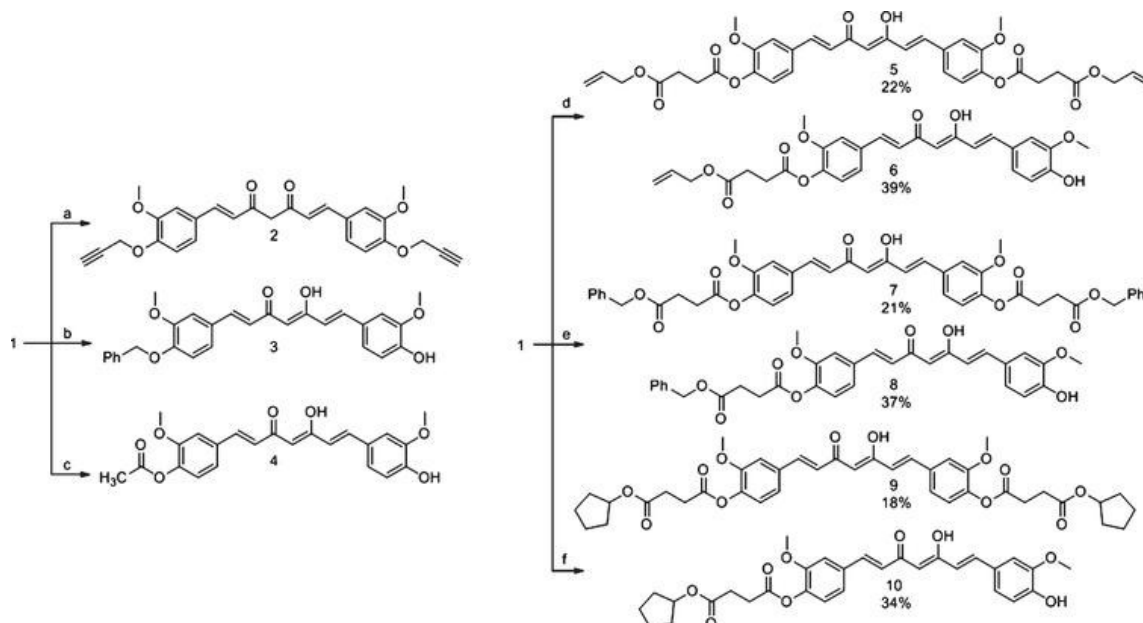
Curcumin derivatives

In this study, seven novel curcumin derivatives (**3**, **5**, **6**, **7**, **8**, **10**) were synthesized. Moreover, two known compounds (**2**, **4**) were also synthesized with the purpose of establishing a structure-activity relationship (SAR).

Compounds **2-3** were synthesized by etherification reactions, while compounds **4-10** were formed by esterification (Fig. 2). The reaction between curcumin and propargyl bromide in the presence of K_2CO_3 and DMF afforded **2** in 70% yield. Curcumin and benzyl bromide reacted in the presence of Cs_2CO_3 and DMF to afford **3** in 80% yield. The reaction between curcumin and acetyl chloride in the presence of K_2CO_3 and acetone produced **4** in 71% yield (Fig. 2).

Fig.2

Synthesis of curcumin derivatives with ether, ester, and diester groups. Reagents and conditions: a) $CHCCH_2Br$, K_2CO_3 , DMF, RT, 48 h, 70%. b) $C_6H_5CH_2Br$, Cs_2CO_3 , CH_3COCH_3 , 56°C, 24 h, 80%. c) CH_3COCl , K_2CO_3 , CH_2Cl_2 , RT, 24 h, 71%. d) S1, DMAP, EDC, pyridine, RT, 48 h, 22% (**5**), and 39% (**6**). e) S2, DMAP, EDC, pyridine, RT, 48 h, 21% (**7**), and 37% (**8**). f) S3, DMAP, EDC, pyridine, RT, 48 h, 18% (**9**), and 34% (**10**).



The reaction between curcumin and 4-(allyloxy)-4-oxobutanoic acid in the presence of EDC, DMAP, and pyridine produced difunctionalized (**5**, 22%), and monofunctionalized (**6**, 39%) succinate analogs ([Fig. 2](#)). On the other hand, when curcumin reacted with 4-(benzyloxy)-4-oxobutanoic acid in the presence of EDC, DMAP, and pyridine, difunctionalized ester **7** and monofunctionalized ester **8** derivatives were produced in 21% and 37% yield, respectively. Finally, when curcumin reacted with 4-(cyclopentyloxy)-4-oxobutanoic acid in the presence of EDC, DMAP, and pyridine, esterified analogs **9** and **10** were produced in 18% and 34% yield, respectively.

Anti-inflammatory activity of curcumin derivatives

To evaluate the anti-inflammatory activity of the curcumin analogs and curcumin, we measured the secretion of IL-6 by murine macrophages stimulated with LPS in the presence or absence of the compounds. Compounds **2–6**, **8**, and **10** reduced macrophage response, while compounds **7** and **9** did not show any anti-inflammatory activity. Compounds **2–4** completely suppressed the production of IL-6 at 10 μ M ([Fig. 3a](#)). This inhibition was not due to a cytotoxic effect of the compounds, since cell viability was not affected at the evaluated concentrations ([Fig. 3b](#)). Compounds **5**, **6**, **8**, and **10** showed mid-to-strong inhibition of IL-6 production ([Fig. 3c](#)), with a complete inhibition of IL-6 at 30 μ M. Compounds **6**, **8**, and **10** did not affect the cell viability at the concentrations tested ([Fig. 3d](#)). Compound **5** exhibited cytotoxicity at a concentration of 30 μ M. The IC₅₀ values for curcumin and its analogs are shown in [Table 1](#).

Fig.3

Anti-inflammatory activity of curcumin derivatives with ether, ester, and diester groups. Peritoneal macrophages from C57BI/6 mice were treated with different concentrations (1, 3, 10, or 30 μ M) of the evaluated compounds 1 h before the stimulus with 10 ng/mL of LPS. After 24 h, concentration of IL-6 was determined by the ELISA method in the supernatant of cells treated with curcumin derivatives with ether and ester groups (a) or with diester groups (c). Cells viability was assessed by the MTT assay after supernatant collection (b and d). All results are represented as Mean \pm S.E.M from three independent experiments performed in duplicate. * p <0.05 relative to the LPS stimulus alone. C, negative control.

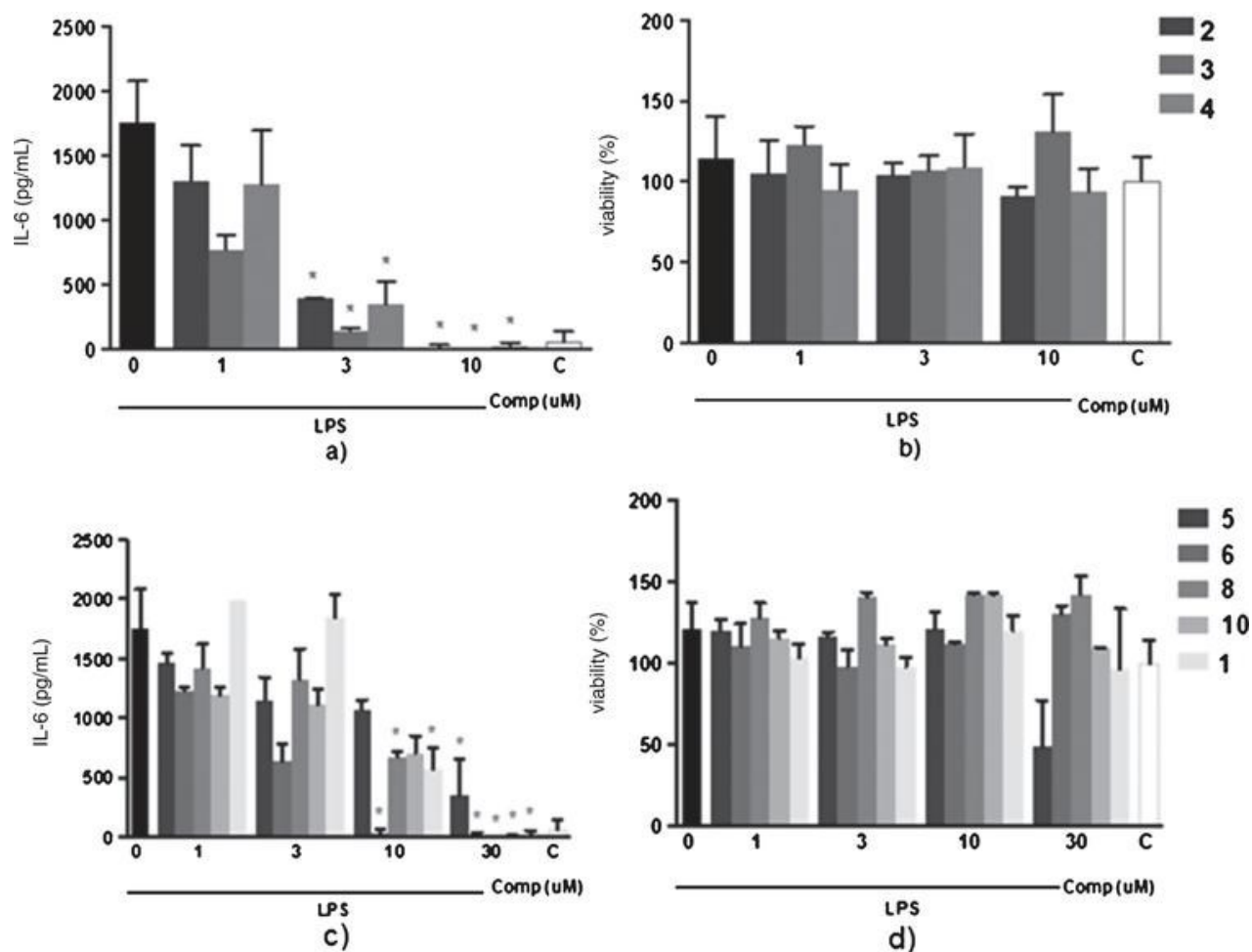


Table 1

Chemical structures and anti-inflammatory and anti-amyloid aggregation activity of synthetic curcumin derivatives

Compound Structure

^aIC₅₀±S.D (μM) ^bThT assay

IC₅₀ (μM)

2		2.23±0.84	NA
3		1.81±1.31	2.05±0.1

Compound	Structure	^a IC ₅₀ ±S.D (μM) ^b ThT assay	
		IC ₅₀ (μM)	
4		2.21±0.93	1.32±0.7
5		14.2±12.8	1.96±0.7
6		2.50±0.92	1.77±0.9
8		8.28±3.08	NA
10		3.22±1.34	2.04±0.7
1		8.25±1.25	1.49±0.9

^{a,b}Values represent average of IC₅₀ from three independent experiments performed in duplicate±S.D.

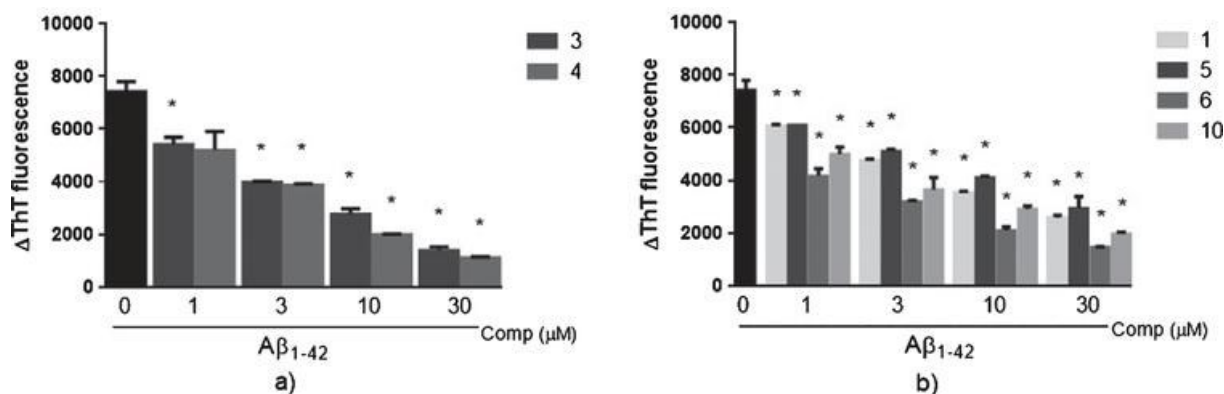
Curcumin derivatives decrease Aβ aggregation

To evaluate the effect of curcumin derivatives on Aβ aggregation, Aβ₁₋₄₂ was incubated for 48h in the presence or absence of compounds. Fibrils of Aβ were detected by a thioflavin T assay. Compounds **3**, **4**, **5**, **6**, and **10** inhibited the aggregation of Aβ in a concentration-dependent manner ([Fig. 4](#)) with IC₅₀ values ranging from 1.32±0.7μM to 2.05±0.1μM ([Table 1](#)). These compounds showed a similar effect on amyloid aggregation as compared to curcumin (**1**) (IC₅₀=1.4±0.9). Compounds **2**, **8**, and **9** did not present anti-aggregation activity.

Fig.4

Effect of curcumin derivatives with ether, ester, and diester groups on Aβ aggregation. Aβ₄₂ recombinant peptide was incubated during 48 h with or without different concentrations (1, 3, 10, or 30μM) of compounds. After incubation, ThT (20μM) was added and was determined

changes in its fluorescence intensity (Δ ThT fluorescence) in the presence of curcumin derivatives with ether and ester groups (a) or with diester groups (b). Results are represented as Mean \pm S.D. from three independent experiments performed in duplicate. * p <0.05 relative to A β ₄₂ alone.



DISCUSSION

New curcumin derivatives were prepared by etherification and esterification in order to provide structural changes that could potentially increase anti-inflammatory activity. Inhibition of the secretion of pro-inflammatory cytokines, such as IL-6, is frequently used as readout of an anti-inflammatory activity. We evaluated the effect of curcumin derivatives on the production of IL-6 by macrophages stimulated with LPS. Assessment of the bioactivity *in vitro* showed that compounds **2–6**, **8**, and **10** decreased the secretion of IL-6, depending on the chemical modification. Compounds **2–4** downregulated the production of IL-6 in a concentration-dependent manner, with a negligible release at 10 μM. A structure-activity relationship of curcumin derivatives was investigated by introducing changes on the hydroxyl groups located on the aromatic rings and evaluating the anti-inflammatory activity. Curcumin modified with small groups by etherification of the hydroxyl groups on both aromatic rings (**2**) showed a much higher anti-inflammatory activity than did unmodified curcumin (2.23 \pm 0.84 versus 8.25 \pm 1.25) (Table 1). Furthermore, introduction of a benzene ring etherified at one of the curcumin rings led to a curcumin derivative with the most potent anti-inflammatory activity (**3**). Acetylation at only one side of the molecule resulted in strong biological activity. However, when the complexity and length of the groups attached to both rings increased, a reduced (**5**), or null biological activity (**7**, **9**), occurred as compared to curcumin. Indeed, a strong improvement on the activity was achieved when modifications were done at only one of the aromatic rings (**6**, **8**, **10**). When monofunctionalized diester curcumin derivatives are considered (**6**, **8**, **10**), it was observed that the improvement of the anti-inflammatory activity was reduced by the presence of bulky groups in the molecule. These bulky groups may disturb the molecular mechanisms by which these derivatives inhibit the production of IL-6 in a fashion that needs further investigation.

Based on the results obtained from the anti-inflammatory activity of curcumin and its analogues, it can be concluded that hydroxyl groups on the aromatic rings of the curcumin are pharmacophores, required for reducing the production of IL-6. Moreover, modifications on curcumin to produce new analogs with potential anti-inflammatory activity by inhibition of IL-6 should take into account the following salient points (1) that at least one hydroxyl group of the aromatic rings should not be modified, and (2) etherification and esterification of only one of the

hydroxyl groups present in the benzene rings will strongly enhance the activity depending on the complexity of the substituent added.

At the molecular level, AD is characterized by the presence of extracellular A β senile plaques in the brain [1, 5]. Senile plaques are produced by the cleavage of the amyloid- β protein precursor (A β PP) by the enzyme β -secretase at the A β PP beta site, leading to a small soluble A β PP β fragment and a C-99 fragment of A β PP. The C-99 fragment is then broken by the enzyme γ -secretase into two fragments of A β PP intracellular domain protein and the pathological A β_{42} peptide, which polymerizes forming amyloid fibrils, leading to cell death in the brain [1].

The effect of curcumin on A β aggregation has been extensively studied [14, 15]. This effect appears to be favoring the generation of non-toxic A β intermediates during the formation of fibrils [16]. It has also been proposed that curcumin disaggregates A β fibrils [17]. Compounds **3**, **4**, **5**, **6** and **10** presented anti-aggregation activity *in vitro* (Table 1). SAR evaluation indicates that curcumin derivatives etherified with small groups at both sides of the molecule lose the anti-aggregation activity (**2**). Nevertheless, etherification at only one side of curcumin maintains the bioactivity, yet is lower than curcumin (**3**). Acetylation at only one side of the molecule led to an analog with a larger biological activity (**4**). Monofunctionalized diester analogs (**6**, **8**, **10**) showed a decreased or null bioactivity compared to curcumin. This decrease in the anti-aggregation activity is associated with the complexity and length of the analog. The presence of bulky groups in monofunctionalized diester derivatives impacted negatively the activity. When bifunctional diester is taken into account (**5**), a reduced biological activity was also observed. It has been suggested that phenolic compounds, such as curcumin, are able to produce anti-aggregation activity due to π - π interactions and the formation of hydrogen bonds between the peptide and the phenolic rings [18]. These anti-aggregation mechanisms might be also followed by compounds **3**, **4**, **5**, **6**, and **10**, yet the exact mechanism needs to be studied. Since we measured the anti-aggregation activity of the compounds in a time period of 48h, we cannot rule out the possibility that these curcumin derivatives might have disaggregation potential. Kinetics studies are necessary to evaluate a potential disaggregate effect of these new curcumin derivatives.

Inhibition of A β aggregation is a promising approach for the identification of new agents for AD treatment. Several synthetic and natural compounds have been tested as inhibitors for A β aggregation [1]. Some molecules have reached different phases of clinical trials; however, until now, no molecule has been approved as new therapeutic. Hence, the search of new molecules with anti-aggregation effect continues to be of great interest.

Our findings suggest that the novel curcumin derivatives **3**, **4**, **6**, and **10** have potential as therapeutic compounds for the treatment of AD.

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Appendices

The supplementary material is available in the electronic version of this article: <http://dx.doi.org/10.3233/JAD-170071>.

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