

Article

On-Surface Locomotion of Particle Based Microrobots Using Magnetically Induced Oscillation

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Abstract: The low Reynolds number condition presents a fundamental constraint on designing locomotive mechanisms for microscale robots. We report on the use of an oscillating magnetic field to induce on-surface translational motion of particle based microrobots. The particle based microrobots consist of microparticles, connected in a chain-like manner using magnetic self-assembly, where the non-rigid connections between the particles provide structural flexibility for the microrobots. Following the scallop theorem, the oscillation of flexible bodies can lead to locomotion at low Reynolds numbers, similar to the beating motion of sperm flagella. We characterized the velocity profiles of the microrobots by measuring their velocities at various oscillating frequencies. We also demonstrated the directional steering capabilities of the microrobots. This work will provide insights into the use of oscillation as a viable mode of locomotion for particle based microrobots near a surface.

Keywords: microrobotics; magnetic control; low Reynolds number

1. Introduction

The concept of using microrobotics for both in vitro and in vivo applications had been well explored; such as micromanipulation and microfabrication [1,2], drug delivery [3,4], tissue manipulation [2,5], and in situ sensing [6], such as in vivo diagnostics [7–9]. At the center of the development of microrobotics has been the study of locomotive mechanisms at low Reynolds numbers. The understanding of locomotion in the context of low Reynolds numbers is critical, as the negligible inertia at low Reynolds numbers requires the use of nonreciprocal motion for movement. Because of this, designing locomotive mechanisms has been one of the difficulties in low Reynolds number motion since many conventional swimming strategies, such as a rigid flapping fin, are ineffective at the micro- and nanoscale due to their reciprocity. According to Purcell, the flexible oar is one of the two major classifications of biological mechanisms for low Reynolds number locomotion; the other being the rotating corkscrew [10]. The bending of an oscillating flexible oar allows for nonreciprocal motion; the oscillating flagella of sperm and the beating cilia of protozoa are biological examples of the flexible oar.

To overcome low Reynolds number constraints, various mechanisms and strategies have been employed in a number of different microrobots such as the helical chiral swimmers [11–19], the bacteria powered microrobots (BPMs) [20–22], the magnetically steered swimming cells [23], the magnetically controlled Mag-μBot [24], the optically-deformed three-bead systems [25], the self-assembled nanoparticle swimmers [26], the biflagellate micro-objects [27], and the chemically-driven robots [28–35]. In this work, we focus on a class of microrobots that utilize their flexible bodies for motion. Notable examples of existing microrobots in this category include

microscopic artificial swimmers with flexible DNA linkages [36], flexible nanowire motors [37], MagnetoSperm [38], sperm-shaped microrobots [39,40], and soft micromachines [41].

The chain-like conjugation of beads that makes up a particle based microrobot is flexible since the connections between the beads are not completely rigid. Note that although the previously reported three-bead achiral microswimmers were claimed to be rigid while rotating [42], this does not apply to particle based microrobots undergoing oscillation. A flexible object rotating at a constant rate will eventually reach a steady state shape where the structure of this object can be modeled as near rigid; this was reported for bacterial flagella where a flexible flagellar filament can be modeled as a rigid helix when rotating [43–46]. This is caused by the constant viscous force experienced by the object under a constant rate rotation at steady state. Viscous forces on an object under oscillation, however, are time varying; thus, the object will undergo deformation in a cyclic manner as seen in the flexible oar example [10].

In this study, we investigate the motion of particle based microrobots induced by an oscillating magnetic field as an alternative strategy of locomotion for near surface movements. The particle based microrobot converts rotational motion into translation motion and swims in the bulk fluid [42,47,48]. When near a boundary, the microrobot's flow field interacts with the boundary, leading to rolling motion [42]. The rolling motion leads to movement in the direction perpendicular to the swimming direction. Thus, rotation based microrobots, including particle based microrobots and helical swimmers, move diagonally when near a boundary. Oscillation, on the other hand, does not lead to rolling motion. In a situation where a rotating microrobot navigates through tight boundaries, rolling will cause the microrobots to deviate from the expected path. While oscillation based locomotion does not present an advantage in terms of speed, the elimination of rolling can provide reliability in predicting the trajectory for near boundary navigation. Therefore, oscillation can be considered as an alternative locomotive strategy for the particle based microrobots when moving near a boundary.

2. Materials and Methods

2.1. Fabrication

The particle based microrobots used in this study consisted of six or more magnetic microparticles (4.35 μm diameter) connected in a chain-like manner. The process of connecting beads using streptavidin-biotin linkages and magnetic attraction is illustrated in Figure 1. First, two batches of coated magnetic microparticles are prepared; one batch with streptavidin coating and the other with biotin coating. The coated particles are commercially available (SpheroBeads). Alternatively, particles can be coated using streptavidin and biotin solutions with the same results [42]. Next, the two batches are mixed together and then placed inside a magnetic field. Finally, the magnetized particles will actively move towards each other and then bond together by magnetic forces and the streptavidin-biotin linkages. One of the advantage of using a magnetic field in the fabrication process is the alignment of dipoles, which ensures that the particles bond in a chain-like manner. Note that the chains are not perfectly linearly due to other factors in the mixing process, such as electrostatic forces, surface roughness, sphericity of particles, etc. The connections between the particles in the chain provide flexibility which allows for swimming or fluid transport at low Reynolds number. Previous work on tethered flexible magnetic bead cilia, fabricated in a similar manner as the particle based microrobots, had demonstrated that particle chains can be used to transport fluids at low Reynolds number by using nonreciprocal beating [49–52]. Similarly, the particle based microrobots are untethered swimmers that can generate a propulsive force when their body oscillate.

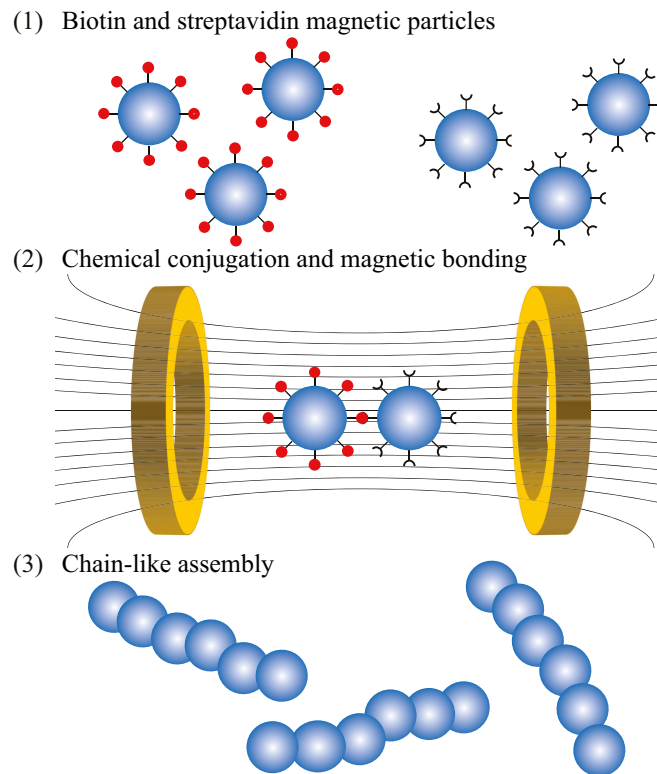


Figure 1. Fabrication process of the particle-based microrobots. The process follows the main steps: (1) preparation of the biotin and streptavidin coating of the magnetic particles; (2) chemical conjugation and magnetic bonding of the particles; (3) self-assembly of the particles into chain-like structures.

2.2. Motion Control Using Magnetic Fields

There are several advantages in using magnetic fields as the actuation method for microrobots. Magnetic fields can permeate and transmit power over long ranges for propulsion, motion control, and magnetic resonance targeting (MRT) [53,54]. The control system used here was designed to generate an oscillating magnetic field at any desired oscillating frequency and field strength. The system consists of an electromagnetic coil system, three bipolar power supplies (Kepco), a National Instrument (NI) data acquisition (DAQ) device, a computer, an inverted microscope (Leica DM IRB), and a camera (Point Grey). In experiments, we can control the magnetic field using a LabVIEW interface. The inputs from LabVIEW are executed by the NI DAQ device which generates sinusoidal signals to the power supplies for the electromagnetic coils. The camera provides real time visual feedback and records videos at 90 frames per second. The schematic for the control system is shown in Figure 2.

The electromagnetic coils are arranged in a configuration similar to Helmholtz coils. The distance between each pair of coils is the same as the outer diameter of the coils plus the thickness of the coils; whereas a true Helmholtz configuration has a distance equal to the radius of the coils. This configuration allows for the generation of a near-uniform magnetic field at the center of the coils. This type of magnetic field is necessary in order to apply a magnetic torque on the microrobots without introducing translational magnetic force. In other words, the magnetic field will cause the microrobots to align with the field without pulling the microrobots to move along the field gradient. This is crucial for this study because the uniform field ensures that the movements of the microrobots were caused by the oscillation of their bodies. The intensity of the magnetic field generated from a pair of coils can be calculated using a modified version of the Biot-Savart law [55]

$$B_{\text{coils}} = \frac{\mu_0 n I R^2}{2(R^2 + x^2 - 2dx + d^2)^{\frac{3}{2}}} + \frac{\mu_0 n I R^2}{2(R^2 + x^2 + 2dx + d^2)^{\frac{3}{2}}} \quad (1)$$

$$B_{\text{coils}} = \frac{\mu_0 n I R^2}{2(R^2 + x^2 - 2dx + d^2)^{\frac{3}{2}}} + \frac{\mu_0 n I R^2}{2(R^2 + x^2 + 2dx + d^2)^{\frac{3}{2}}} \quad (1)$$

where μ_0 is the permeability, n is the number of turns of wires per coil, I is the electrical current passing through the wires, R is the effective radius of the coil, d is the distance between the center of a coil pair, and x is the coil distance to a point. The modification from the original Biot-Savart law accounts for the distance between the pairs of coils for this particular configuration.

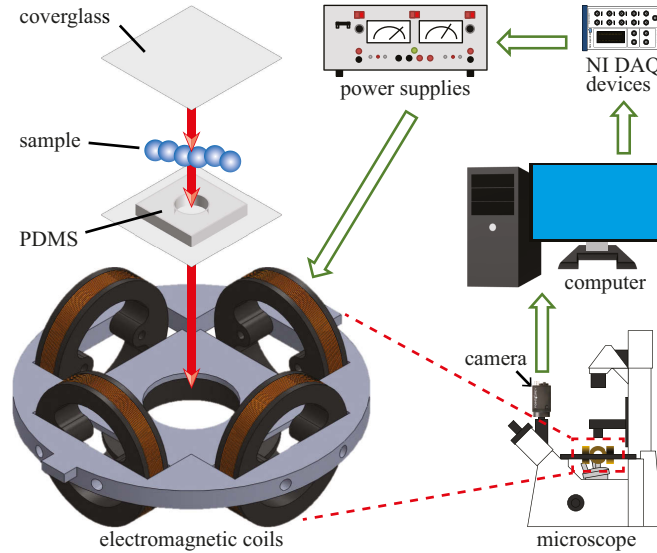


Figure 2. Experiment setup and control system for motion control using oscillating magnetic fields.

The magnetic field strength (mT), direction of the oscillating field, and oscillation frequency (Hz) of the field generated by the coils can be controlled through the control system. To create an oscillating magnetic field, two pairs of electromagnetic coils are needed. The resultant field B can be expressed as

$$B = B_0 \begin{bmatrix} -\left(1 - \sin(c\pi)\right) \cos(\theta) \left(\frac{|\cos(\omega t)|}{(1 - \sin(c\pi))} + \frac{\sin(c\pi)}{(1 - \sin(c\pi))} \right) + \cos(c\pi) \sin(\theta) \sin(\omega t) \\ \left(1 - \sin(c\pi)\right) \sin(\theta) \left(\frac{|\cos(\omega t)|}{(1 - \sin(c\pi))} + \frac{\sin(c\pi)}{(1 - \sin(c\pi))} \right) + \cos(c\pi) \cos(\theta) \sin(\omega t) \\ \left(1 - \sin(c\pi)\right) \sin(\theta) \left(\frac{|\cos(\omega t)|}{(1 - \sin(c\pi))} + \frac{\sin(c\pi)}{(1 - \sin(c\pi))} \right) + \cos(c\pi) \cos(\theta) \sin(\omega t) \end{bmatrix} \quad (2)$$

where B_0 is the amplitude of the magnetic field, ω is the oscillating frequency of the field, θ is the direction of oscillation, c is a coefficient that determines the angle of oscillation α , and t is time. The coefficient c where B_0 is the amplitude of the magnetic field, ω is the oscillating frequency of the field, θ is the direction of oscillation, c is a coefficient that determines the angle of oscillation α , and t is time. In motion control experiments, the angle θ controls the movement direction and ω controls the movement speed. The coefficient c and the angle of oscillation α will be discussed further in the proceeding section. The oscillating magnetic fields generated using Equation (2) offer a few advantages over methods used for previous swimmers [36,39,40]: (1) they allow for swimming at any directions based on the angle θ , (2) they generate near uniform fields similar to Helmholtz coils, and (3) they maintain a near constant field strength for the full cycle of each oscillation.

2.3. Oscillatory Swimming Motion

The microrobots can undergo oscillating motion at any desired frequency using the parameter ω from Equation (2). A frame by frame representation of the oscillating motion of a microrobot is shown in Figure 3a. The microrobot oscillated at 4 Hz for one full cycle from $t = 0$ s to $t = 0.27$ s. The merged image from Equation (2). A frame by frame representation of the oscillating motion of a microrobot is shown in Figure 3a. The microrobot oscillated at 4 Hz for one full cycle from $t = 0$ s to $t = 0.27$ s. The merged image shows the full range of the oscillation with an angle of oscillation (α) of 54° . Using the control parameter c in Equation (2), we can manipulate the angle of oscillation of the microrobots. In Figure 3b, six arcs are drawn using Equation (2) to represent full cycles of oscillation of the magnetic field at six angles of oscillation. The value of B_0 does not matter here since B_0 only serves as a scaling parameter. The frequency of oscillation ω also does not matter for the static representation of the oscillation. From the arcs, six different angles of oscillation are presented along with the corresponding c values.

oscillation. The value of B_0 does not matter here since B_0 only serves as a scaling parameter. The frequency of oscillation ω also does not matter for the static representation of the oscillation. From the arcs, six different angles of oscillation are presented along with the corresponding c values. For example, the angle of oscillation of 54° from Figure 3a was obtained using $c = 0.2$ in Equation (2). It should be noted that c is a dimensionless coefficient used to yield a specific angle of oscillation α ; the value of c does not have a physical meaning.

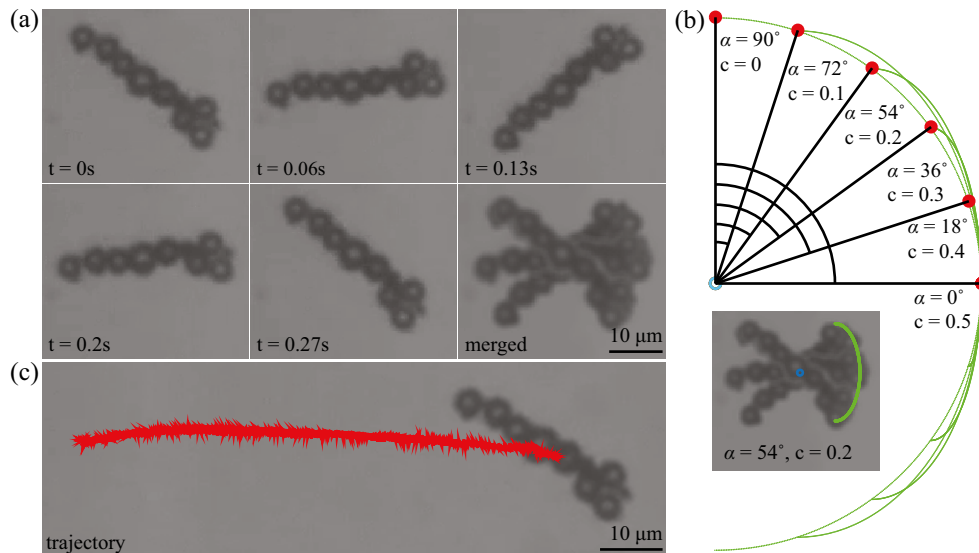


Figure 3. Oscillatory motion of the microbots and magnetic field. (a) Frame-by-frame representation of a full oscillation cycle from $t = 0$ s to $t = 27$ s. (b) An illustration of six angles of oscillation α and the corresponding c values. The inset images show the microbot from (a) with an overlay at an angle of oscillation of 54° and a corresponding value of 0.2 . (c) Translational motion of the oscillating microbot in (a) also see Video S1. The scale bars are $10\text{ }\mu\text{m}$.

3. Results

3.1. Velocity Profile

To demonstrate the motion control capability, we characterized the velocities of the microbots at various oscillating frequencies (0 Hz, 10 Hz, 20 Hz, 20 Hz, and 30 Hz) (30 Hz is the value of ω kept at $c = 0.2$, which corresponded to an angle of oscillation of 54°). The magnetic field strength B_0 was also kept at a constant value applied 0.81 mT . The applied field was strong enough to oscillate the microbots at the desired frequency and weak enough to keep the electromagnetic coils from overheating. For experiments, we placed the microbots into a simple Polydimethylsiloxane (PDMS) chamber filled with water. The chamber was sealed as shown in Figure 2. The chamber was then placed inside the electromagnetic coil system.

A MATLAB tracking algorithm was used to analyze the motion of the microbots. The algorithm performed image processing on videos that were recorded at 30 frames per second at 640×512 resolution. The image processing extracted the centroid position (x, y) of the microbots at each consecutive frame. The process followed four steps: image binarization using a grayscale thresholding, structure definition based on pixel continuity, size thresholding to get rid of objects with outlier sizes, and calculation of the centroid. The centroids from each consecutive frame were used to calculate their velocities.

The velocity profiles and images of the five oscillating microbots are shown in Figure 4. The five different microbots were tracked and exhibited linear relationships between the swimming velocity and the oscillation frequency. The approximate length of the microbots were $29.11\text{ }\mu\text{m}$, $23.63\text{ }\mu\text{m}$, $28.06\text{ }\mu\text{m}$, $27.30\text{ }\mu\text{m}$, and $33.21\text{ }\mu\text{m}$, respectively. The velocities were offset by subtracting the background flow. The background flow velocities were determined by observing the movement of the nonmotile microbots at 0 Hz. After the offset, velocities at 0 Hz became $0\text{ }\mu\text{m/s}$, and the

μm , 27.30 μm , and 33.21 μm , respectively. The velocities were offset by subtracting the background flow. The background flow velocities were determined by observing the movement of the nonmotile microrobots at 0 Hz. After the offset, velocities at 0 Hz became 0 $\mu\text{m/s}$, and the velocities at other frequencies were offset accordingly by the background flow. It should be noted that the background velocities were very small due to how close the microrobots were to the surface. From these data, it can be seen that the velocities of the microrobots vary from one another due to their geometry. Due to the fabrication process, the geometry of the microrobots is not the same, therefore, we chose microrobots with lengths within 10 μm from each other. Although we measured the length of the microrobots, the images in Figure 4 show that geometrical variation between the microrobots goes beyond length, since some microrobots are more linear than others, thus, we did not attempt make a conclusion based on length and speed. To draw a relationship between the geometry and swimming speed, precise geometry and deformation were needed, in order to examine the friction parallel and perpendicular to the bodies of the microrobots. The deformation of microrobots were 0.4824, 0.3880 and 0.2384, respectively. The information for the fitted line in Figure 4 is listed in Table 1.

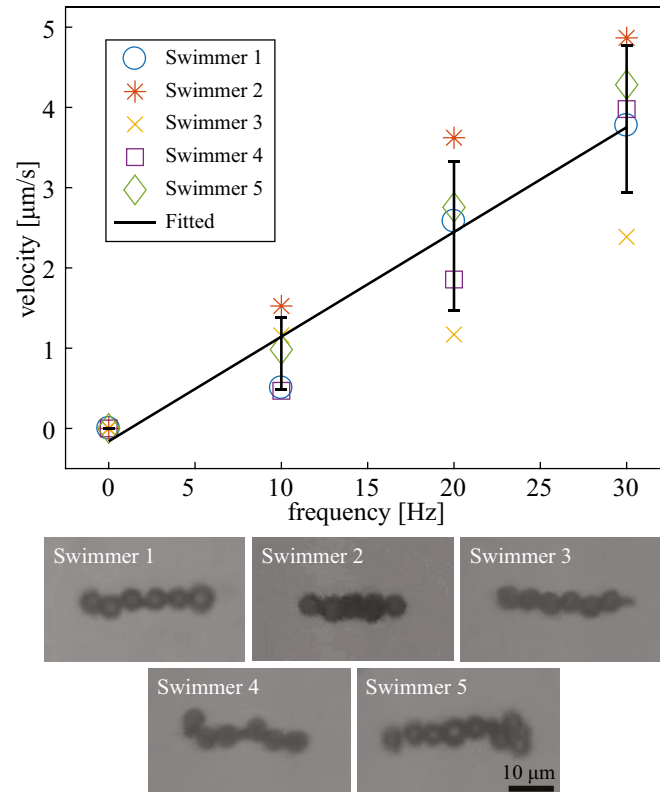


Figure 4. Velocity profiles for oscillating microrobots plotted against oscillation frequency. The plot includes five different microrobots. The fitted line is a linear fit of the mean velocities of the five microrobots. The error bars represent standard errors. The images below the plot show the geometry for each microrobot represented in the plot. The sharpness and contrast of the images were enhanced for visibility.

We applied a linear fit to the average velocities of the five robots with a slope of 0.13 and a R^2 value of 0.99. The standard deviation for the error bars at 10, 20, and 30 Hz were 0.4478 $\mu\text{m/s}$, 0.9301 $\mu\text{m/s}$, and 0.9198 $\mu\text{m/s}$, respectively. The coefficient of variations were 0.4824, 0.3880, and 0.2384, respectively. The information for the fitted line in Figure 4 is listed in Table 1.

Table 1. Values for the fitted line in Figure 4.

Frequency	Mean Velocity ($\mu\text{m/s}$)	Standard Deviation ($\mu\text{m/s}$)	Coefficient of Variations
10	0.9284	0.4478	0.4824
20	2.3973	0.9301	0.3880
30	3.8580	0.9198	0.2384

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<i>Micromachines</i> 2017 , <i>8</i> , 46	20	2.3973	0.9301	0.3880	7 of 10
	30	3.8580	0.9198	0.2384	

3.2. Motion Control

Using Equation (2), the movement direction of the microrobots can be manipulated by changing the angle θ . To demonstrate the turning capability of the oscillating microrobots, a representative example of a microrobot that steered to travel in an N pattern is shown in Figure 5a. The oscillating frequency was kept at 15 Hz for the entire duration of the experiment. This illustrates the ability of making sharp turns while using the oscillating mode. The velocity of the microrobot was measured at one second intervals and is plotted in Figure 5b. The drops in velocity at around $t = 15$ s to $t = 20$ s, $t = 40$ s, and $t = 43$ s were attributed to pauses that were used to facilitate turning.

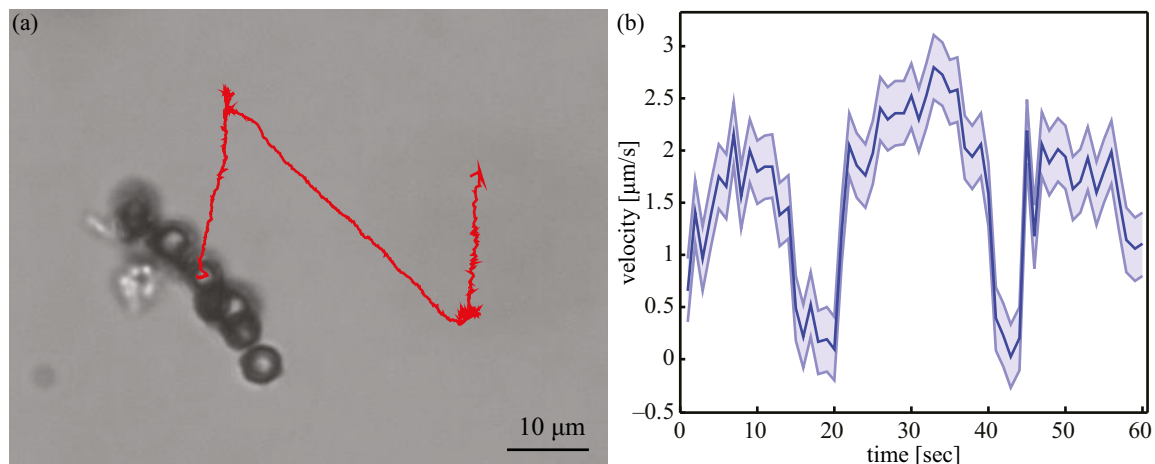


Figure 5. Steering of the microrobots. (a) An oscillating microrobot traveled in an N pattern. (b) The velocity profile plotted against time for the microrobot in (a).

4. Discussion

The results of this work demonstrate movement speed control and directional control of magnetically actuated oscillating microrobots. The velocity profile in Figure 4 illustrated a linear relationship between movement speed and oscillation frequency. The R^2 value of 0.99 of the linear curve fit suggests a high linear correlation between the velocity of the microrobots and the applied oscillation frequency. Furthermore, the relatively small coefficient of variance illustrates the consistencies between the velocity profiles of the microrobots. This linear relationship will be useful in controlling and predicting the movement speed by using the oscillation frequency as a control parameter. This will be particularly useful for developing a qualitative kinematic model that can be used for autonomous feedback control. Furthermore, the demonstration of sharp turns suggests that the microrobots can be controlled as omnidirectional robots, which is a common property among microrobots controlled using externally applied magnetic fields.

5. Conclusions

In summary, the demonstration of using oscillation as a mode of motion for particle based microrobots presents the possibility to move on or near surfaces where rotational motion might not be preferred. As experiments have shown, the microrobots can be assembled en masse using chemical and magnetic bonding. To show the viability of using oscillation as a strategy for locomotion, experiments were performed to characterize the velocity profiles of five different microrobots at incremental frequencies. The linearity of the velocity profiles demonstrated the feasibility of manipulating the movement speed by controlling the oscillation frequency. The steerability of the microrobot was also investigated, to demonstrate that both speed and direction control can be achieved using magnetically induced oscillation.

Supplementary Materials: The following are available online at www.mdpi.com/2072-666X/8/2/46/s1. Video S1: On surface translational motion of an oscillating microrobot.

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Author Contributions: U Kei Cheang built the experimental apparatus, designed the experiments, carried out the fabrication and experiments, and performed the image and data analysis. Jamel Ali and Louis Rogowski helped carry out the experiments. Hoyeon Kim helped build the experimental apparatus. Min Jun Kim helped design the experiments. U Kei Cheang wrote the paper after discussing the results with Jamel Ali, Hoyeon Kim, Louis Rogowski, and Min Jun Kim, who also commented on and revised the manuscript.

Conflicts of Interest: The authors declare no conflict of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

References

1. Ferreira, A.; Agnus, J.; Chaillet, N.; Breguet, J.-M. A smart microrobot on chip: Design, identification, and control. *Trans. Mechatron.* **2004**, *9*, 508–519. [[CrossRef](#)]
2. Zhang, H.; Huttmacher, D.W.; Chollet, F.; Poo, A.N.; Burdet, E. Microrobotics and MEMS-based fabrication techniques for scaffold-based tissue engineering. *Macromol. Biosci.* **2005**, *5*, 477–489. [[CrossRef](#)] [[PubMed](#)]
3. Dogangil, G.; Ergeneman, O.; Abbott, J.J.; Pané, S.; Hall, H.; Muntwyler, S.; Nelson, B.J. Toward targeted retinal drug delivery with wireless magnetic microrobots. In Proceedings of the IEEE/RSJ International Conference on Intelligent Robots and Systems, Nice, France, 22–26 September 2008; pp. 1921–1926.
4. Fusco, S.; Chatzipirpiridis, G.; Sivaraman, K.M.; Ergeneman, O.; Nelson, B.J.; Pané, S. Chitosan electrodeposition for microrobotic drug delivery. *Adv. Healthc. Mater.* **2013**, *2*, 1037–1044. [[CrossRef](#)] [[PubMed](#)]
5. Kim, S.; Qiu, F.; Kim, S.; Ghanbari, A.; Moon, C.; Zhang, L.; Nelson, B.J.; Choi, H. Fabrication and characterization of magnetic microrobots for three-dimensional cell culture and targeted transportation. *Adv. Mater.* **2013**, *25*, 5863–5868. [[CrossRef](#)] [[PubMed](#)]
6. Liu, X.; Kim, K.; Zhang, Y.; Sun, Y. Nanonewton force sensing and control in microrobotic cell manipulation. *Int. J. Robot. Res.* **2009**, *28*, 1065–1076.
7. Martel, S.; Mathieu, J.-B.; Felfoul, O.; Chanu, A.; Aboussouan, E.; Tamaz, S.; Poupponeau, P.; Yahia, L.; Beaudoin, G.; Soulez, G. Automatic navigation of an untethered device in the artery of a living animal using a conventional clinical magnetic resonance imaging system. *Appl. Phys. Lett.* **2007**, *90*, 114105. [[CrossRef](#)]
8. Grady, M.; Howard, M., III; Molloy, J.; Ritter, R.; Quate, E.; Gillies, G. Nonlinear magnetic stereotaxis: Three-dimensional, in vivo remote magnetic manipulation of a small object in canine brain. *Med. Phys.* **1990**, *17*, 405–415. [[CrossRef](#)] [[PubMed](#)]
9. Mathieu, J.-B.; Beaudoin, G.; Martel, S. Method of propulsion of a ferromagnetic core in the cardiovascular system through magnetic gradients generated by an mri system. *Trans. Biomed. Eng.* **2006**, *53*, 292–299. [[CrossRef](#)] [[PubMed](#)]
10. Purcell, E.M. Life at low reynolds number. *Am. J. Phys.* **1977**, *45*, 3–11. [[CrossRef](#)]
11. Tottori, S.; Zhang, L.; Qiu, F.; Krawczyk, K.K.; Franco-Obregón, A.; Nelson, B.J. Magnetic helical micromachines: Fabrication, controlled swimming, and cargo transport. *Adv. Mater.* **2012**, *24*, 811–816. [[CrossRef](#)] [[PubMed](#)]
12. Ghosh, A.; Fischer, P. Controlled propulsion of artificial magnetic nanostructured propellers. *Nano Lett.* **2009**, *9*, 2243–2245. [[CrossRef](#)] [[PubMed](#)]
13. Cheang, U.K.; Roy, D.; Lee, J.H.; Kim, M.J. Fabrication and magnetic control of bacteria-inspired robotic microswimmers. *Appl. Phys. Lett.* **2010**, *97*, 213704. [[CrossRef](#)]
14. Temel, F.Z.; Yesilyurt, S. Magnetically actuated micro swimming of bio-inspired robots in mini channels. In Proceedings of the IEEE International Conference on Mechatronics (ICM), Istanbul, Turkey, 13–15 April 2011; pp. 342–347.
15. Peyer, K.E.; Zhang, L.; Nelson, B.J. Localized non-contact manipulation using artificial bacterial flagella. *Appl. Phys. Lett.* **2011**, *99*, 174101. [[CrossRef](#)]

16. Gao, W.; Feng, X.; Pei, A.; Kane, C.R.; Tam, R.; Hennessy, C.; Wang, J. Bioinspired helical microswimmers based on vascular plants. *Nano Lett.* **2013**, *14*, 305–310. [[CrossRef](#)] [[PubMed](#)]
17. Yan, X.; Zhou, Q.; Yu, J.; Xu, T.; Deng, Y.; Tang, T.; Feng, Q.; Bian, L.; Zhang, Y.; Ferreira, A. Magnetite nanostructured porous hollow helical microswimmers for targeted delivery. *Adv. Funct. Mater.* **2015**, *25*, 5333–5342. [[CrossRef](#)]
18. Li, J.; Sattayasamitsathit, S.; Dong, R.; Gao, W.; Tam, R.; Feng, X.; Ai, S.; Wang, J. Template electrosynthesis of tailored-made helical nanoswimmers. *Nanoscale* **2014**, *6*, 9415–9420. [[CrossRef](#)] [[PubMed](#)]
19. Barbot, A.; Decanini, D.; Hwang, G. On-chip microfluidic multimodal swimmer toward 3D navigation. *Sci. Rep.* **2016**, *6*, 19041. [[CrossRef](#)] [[PubMed](#)]
20. Steager, E.B.; Sakar, M.S.; Kumar, V.; Pappas, G.J.; Kim, M.J. Electrokinetic and optical control of bacterial microrobots. *J. Micromech. Microeng.* **2011**, *21*, 035001. [[CrossRef](#)]
21. Kim, H.; Cheang, U.K.; Julius, A.A.; Kim, M.J. Dynamic obstacle avoidance for bacteria-powered microrobots. In Proceedings of the IEEE/RSJ International Conference on Intelligent Robots and Systems, Hamburg, Germany, 28 September–2 October 2015; pp. 2000–2005.
22. Kim, H.; Kim, M.J. Electric field control of bacteria-powered microrobots using a static obstacle avoidance algorithm. *Trans. Robot.* **2016**, *32*, 125–137. [[CrossRef](#)]
23. Kim, D.H.; Cheang, U.K.; Kohidai, L.; Byun, D.; Kim, M.J. Artificial magnetotactic motion control of tetrahymena pyriformis using ferromagnetic nanoparticles: A tool for fabrication of microbiorobots. *Appl. Phys. Lett.* **2010**, *97*, 173702. [[CrossRef](#)]
24. Diller, E.; Floyd, S.; Pawashe, C.; Sitti, M. Control of multiple heterogeneous magnetic microrobots in two dimensions on nonspecialized surfaces. *IEEE Trans. Robot.* **2012**, *28*, 172–182. [[CrossRef](#)]
25. Leoni, M.; Kotar, J.; Bassetti, B.; Cicuta, P.; Lagomarsino, M.C. A basic swimmer at low reynolds number. *Soft Matter* **2009**, *5*, 472–476. [[CrossRef](#)]
26. Cheang, U.K.; Kim, M. Self-assembly of robotic micro- and nanoswimmers using magnetic nanoparticles. *J. Nanopart. Res.* **2015**, *17*, 1–11. [[CrossRef](#)]
27. Mori, N.; Kuribayashi, K.; Takeuchi, S. Artificial flagellates: Analysis of advancing motions of biflagellate micro-objects. *Appl. Phys. Lett.* **2010**, *96*, 083701. [[CrossRef](#)]
28. Ismagilov, R.F.; Schwartz, A.; Bowden, N.; Whitesides, G.M. Autonomous movement and self-assembly. *Angew. Chem.* **2002**, *114*, 674–676. [[CrossRef](#)]
29. Paxton, W.F.; Kistler, K.C.; Olmeda, C.C.; Sen, A.; St. Angelo, S.K.; Cao, Y.; Mallouk, T.E.; Lammert, P.E.; Crespi, V.H. Catalytic nanomotors: Autonomous movement of striped nanorods. *J. Am. Chem. Soc.* **2004**, *126*, 13424–13431. [[CrossRef](#)] [[PubMed](#)]
30. Wang, Y.; Hernandez, R.M.; Bartlett, D.J.; Bingham, J.M.; Kline, T.R.; Sen, A.; Mallouk, T.E. Bipolar electrochemical mechanism for the propulsion of catalytic nanomotors in hydrogen peroxide solutions. *Langmuir* **2006**, *22*, 10451–10456. [[CrossRef](#)] [[PubMed](#)]
31. Orozco, J.; García-Gradilla, V.; D’Agostino, M.; Gao, W.; Cortés, A.; Wang, J. Artificial enzyme-powered microfish for water-quality testing. *ACS Nano* **2012**, *7*, 818–824. [[CrossRef](#)] [[PubMed](#)]
32. Manesh, K.M.; Cardona, M.; Yuan, R.; Clark, M.; Kagan, D.; Balasubramanian, S.; Wang, J. Template-assisted fabrication of salt-independent catalytic tubular microengines. *ACS Nano* **2010**, *4*, 1799–1804. [[CrossRef](#)] [[PubMed](#)]
33. Solovev, A.A.; Mei, Y.; Bermúdez Ureña, E.; Huang, G.; Schmidt, O.G. Catalytic microtubular jet engines self-propelled by accumulated gas bubbles. *Small* **2009**, *5*, 1688–1692. [[CrossRef](#)] [[PubMed](#)]
34. Gibbs, J.; Zhao, Y.-P. Autonomously motile catalytic nanomotors by bubble propulsion. *Appl. Phys. Lett.* **2009**, *94*, 163104. [[CrossRef](#)]
35. Kline, T.R.; Paxton, W.F.; Mallouk, T.E.; Sen, A. Catalytic nanomotors: Remote-controlled autonomous movement of striped metallic nanorods. *Angew. Chem.* **2005**, *117*, 754–756. [[CrossRef](#)]
36. Dreyfus, R.; Baudry, J.; Roper, M.L.; Fermigier, M.; Stone, H.A.; Bibette, J. Microscopic artificial swimmers. *Nature* **2005**, *437*, 862–865. [[CrossRef](#)] [[PubMed](#)]
37. Gao, W.; Kagan, D.; Pak, O.S.; Clawson, C.; Campuzano, S.; Chuluun-Erdene, E.; Shipton, E.; Fullerton, E.E.; Zhang, L.; Lauga, E.; et al. Cargo-towing fuel-free magnetic nanoswimmers for targeted drug delivery. *Small* **2012**, *8*, 460–467. [[CrossRef](#)] [[PubMed](#)]
38. Khalil, I.S.; Dijkslag, H.C.; Abelman, L.; Misra, S. Magnetosperm: A microrobot that navigates using weak magnetic fields. *Appl. Phys. Lett.* **2014**, *104*, 223701. [[CrossRef](#)]

39. Khalil, I.S.; Tabak, A.F.; Hosney, A.; Klingner, A.; Shalaby, M.; Abdel-Kader, R.M.; Serry, M.; Sitti, M. Targeting of cell mockups using sperm-shaped microrobots in vitro. In Proceedings of the 6th IEEE International Conference on Biomedical Robotics and Biomechatronics (BioRob), UTown, Singapore, 26–29 June 2016; pp. 495–501.
40. Khalil, I.S.; Tabak, A.F.; Hosney, A.; Mohamed, A.; Klingner, A.; Ghoneima, M.; Sitti, M. Sperm-shaped magnetic microrobots: Fabrication using electrospinning, modeling, and characterization. In Proceedings of the IEEE International Conference on Robotics and Automation (ICRA), Stockholm, Sweden, 16–21 May 2016.
41. Huang, H.-W.; Sakar, M.S.; Petruska, A.J.; Pané, S.; Nelson, B.J. Soft micromachines with programmable motility and morphology. *Nat. Commun.* **2016**, *7*, 12263. [[CrossRef](#)] [[PubMed](#)]
42. Cheang, U.K.; Meshkati, F.; Fu, H.; Kim, M. Magnetic control of rigid achiral microswimmers. *Bull. Am. Phys. Soc.* **2013**, *58*, 18.
43. Rodenborn, B.; Chen, C.-H.; Swinney, H.L.; Liu, B.; Zhang, H. Propulsion of microorganisms by a helical flagellum. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, E338–E347. [[CrossRef](#)] [[PubMed](#)]
44. Liu, B.; Breuer, K.S.; Powers, T.R. Propulsion by a helical flagellum in a capillary tube. *Phys. Fluids* **2014**, *26*, 011701. [[CrossRef](#)]
45. Duprat, C.; Shore, H.A. *Fluid-Structure Interactions In Low-Reynolds-Number Flows*; Royal Society of Chemistry: London, UK, 2015; p. 483.
46. Bray, D. *Cell Movements: From Molecules to Motility*; Garland Science: New York, NY, USA, 2001; p. 372.
47. Cheang, U.K.; Lee, K.; Julius, A.A.; Kim, M.J. Multiple-robot drug delivery strategy through coordinated teams of microswimmers. *Appl. Phys. Lett.* **2014**, *105*, 083705. [[CrossRef](#)]
48. Cheang, U.K.; Meshkati, F.; Kim, H.; Lee, K.; Fu, H.C.; Kim, M.J. Versatile microrobotics using simple modular subunits. *Sci. Rep.* **2016**, 30472. [[CrossRef](#)] [[PubMed](#)]
49. Wang, Y.; Gao, Y.; Wyss, H.; Anderson, P.; den Toonder, J. Out of the cleanroom, self-assembled magnetic artificial cilia. *Lab Chip* **2013**, *13*, 3360–3366. [[CrossRef](#)] [[PubMed](#)]
50. Vilfan, M.; Potočnik, A.; Kavčič, B.; Osterman, N.; Poberaj, I.; Vilfan, A.; Babič, D. Self-assembled artificial cilia. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 1844–1847. [[CrossRef](#)] [[PubMed](#)]
51. Babataheri, A.; Roper, M.; Fermigier, M.; Du Roure, O. Tethered fleximags as artificial cilia. *J. Fluid Mech.* **2011**, *678*, 5–13. [[CrossRef](#)]
52. Coq, N.; Bricard, A.; Delapierre, F.-D.; Malaquin, L.; Du Roure, O.; Fermigier, M.; Bartolo, D. Collective beating of artificial microcilia. *Phys. Rev. Lett.* **2011**, *107*, 014501. [[CrossRef](#)] [[PubMed](#)]
53. Abbott, J.J.; Peyer, K.E.; Lagomarsino, M.C.; Zhang, L.; Dong, L.; Kaliakatsos, I.K.; Nelson, B.J. How should microrobots swim? *Int. J. Robot. Res.* **2009**, *28*, 1434–1447. [[CrossRef](#)]
54. Nelson, B.J.; Kaliakatsos, I.K.; Abbott, J.J. Microrobots for minimally invasive medicine. *Annu. Rev. Biomed. Eng.* **2010**, *12*, 55–85. [[CrossRef](#)] [[PubMed](#)]
55. Griffiths, D.J.; College, R. *Introduction to Electrodynamics*; Prentice Hall: Upper Saddle River, NJ, USA, 1999; Volume 3.

