

Opinion

Symbiotic Dinoflagellate Functional Diversity Mediates Coral Survival under Ecological Crisis

David J. Suggett,¹ Mark E. Warner,² and William Leggat^{3,*}

Coral reefs have entered an era of 'ecological crisis' as climate change drives catastrophic reef loss worldwide. Coral growth and stress susceptibility are regulated by their endosymbiotic dinoflagellates (genus *Symbiodinium*). The phylogenetic diversity of *Symbiodinium* frequently corresponds to patterns of coral health and survival, but knowledge of functional diversity is ultimately necessary to reconcile broader ecological success over space and time. We explore here functional traits underpinning the complex biology of *Symbiodinium* that spans free-living algae to coral endosymbionts. In doing so we propose a mechanistic framework integrating the primary traits of resource acquisition and utilisation as a means to explain *Symbiodinium* functional diversity and to resolve the role of *Symbiodinium* in driving the stability of coral reefs under an uncertain future.

Global Deterioration of Reefs through Coral Bleaching

Coral reefs have become a global ecological casualty of the Anthropocene. Ecosystem services provided by coral reefs sustain nearly 10% of all people on Earth and support billion dollar industries in tourism and fisheries [1]. However, coral reefs worldwide have moved into an era of 'ecological crisis' from accelerating overexploitation and persistent anthropogenic threats. Elevated seawater temperature from climate change poses the greatest threat, driving mass coral bleaching (see [Glossary](#)) and associated mortality across entire regions with increasing frequency and intensity [2].

The ecological foundation of coral reefs rests on the symbiosis between reef-building corals and dinoflagellate microalgae ('zooxanthellae') of the genus *Symbiodinium* (Dinophyceae, Suessiales). This symbiosis disassociates when surface seawater temperature (SST) exceeds the long-term maximum monthly mean for extended durations [3], leading to rapid loss of *Symbiodinium* cells and/or their pigmentation. Bleaching is defined by the conspicuous whitening of the coral tissue [4] and, although there is a clear hierarchy of bleaching susceptibility among coral species [5] (e.g., [Figure 1](#)), the broad-scale impacts of elevated SST on individual coral colonies are modified by a variety of physical and biological processes, including the inherent properties of the coral host and *Symbiodinium* [6,7]. Together these processes result in complex bleaching mosaics across reefs with distinct inter- and intraspecific responses ([Figure 1](#)) that modify bleaching thresholds and hence the impact of heating. Consequently, mass bleaching manifests when ecologically dominant coral–*Symbiodinium* associations are exposed to temperatures above their thermal thresholds [2,5] ([Figure 1](#)).

Trends

Coral ecosystem health is strongly influenced by *Symbiodinium* diversity.

The ecological success of *Symbiodinium* cannot be resolved from phylogenetic diversity alone.

Traits describing resource acquisition and incorporation capture the functional diversity of *Symbiodinium*.

Symbiodinium species shifts reflect the changing metabolic requirements of the host.

Functional diversity will determine the resilience of coral reefs to environmental change.

¹ Climate Change Cluster, University of Technology Sydney, Broadway, NSW 2007, Australia

² College of Earth, Ocean, and Environment, University of Delaware, Lewes, DE 19958, USA

³ Australian Research Council (ARC) Centre of Excellence for Coral Reef Studies, College of Public Health, Medical, and Veterinary Sciences, and the Comparative Genomics Centre, James Cook University, Townsville, QLD 4811, Australia

*Correspondence: bill.leggat@jcu.edu.au (W. Leggat).

Heat stress-induced coral bleaching characterises continual progression of the symbiosis disassociation that is governed by a variety of host and/or *Symbiodinium* responses (Figure 1), but distilling this process into a generalised response for *Symbiodinium* is still unresolved (Box 1). Unlike in higher plants where thermal stress primarily affects ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) activase [8], the cellular target broadly underpinning *Symbiodinium* thermal stress susceptibility remains unknown. Various putative sites, including light-harvesting complexes, the reaction centre complex of photosystem II, thylakoid membranes, RuBisCO, and carbon-concentrating mechanisms (CCMs), have all been described as targets of heat stress [9-12], implying broad functional diversity with which cellular networks have evolved to govern stress susceptibility across the genus [13]. However, potentially unifying traits that govern stress tolerance have still not been identified (Box 1), thereby highlighting that second-order traits governing stress tolerance may simply be poor metrics to represent the complex physiologies ultimately regulating broad ecological success. Instead we propose that functional diversity underpinning stress susceptibility should be based on the key first-order traits that govern *Symbiodinium* metabolic functioning, and thus the growth and performance of their coral hosts, under both optimum and suboptimum environmental conditions.

Ecosystem Stability from *Symbiodinium* Diversity

Molecular-level markers have established exceptional phylogenetic diversity inherent within the genus *Symbiodinium* (Box 2), and this plays a major role in whether and how coral reef ecosystems respond to environmental perturbations [14]. Molecular ecology-based studies have now repeatedly demonstrated that the viability of the entire coral symbiosis over space and time [15,16-18] often corresponds to the species (or genetic variant) of *Symbiodinium* that is present. Similarly, corals populating relatively unfavourable environments, such as hot-acidic lagoons [19] or hot-saline catchments [20], typically associate with specific *Symbiodinium* taxa (Box 2). Continued improvements to molecular tools have therefore unquestionably established a central role for *Symbiodinium* diversity in shaping environmental thresholds for coral productivity and ultimately reef growth.

Basic evolutionary theory requires that the maintenance of phylogenetic diversity must be driven by functional differences in *Symbiodinium*. However, we now lag far behind molecular ecology in any comprehensive understanding of the *Symbiodinium* primary physiological attributes that determine coral ecological success given the immense phylogenetic diversity of the alga. This is clearly problematic where a change (or maintenance) in function does not reflect a parallel change to phylogenetic diversity. Notably, convergent evolution across bioregions such as the Caribbean (dominated by clades A and B) versus the Indo-Pacific (dominated by clades C and D) is likely to reflect common environmental histories selecting for the same functional responses (e.g., high versus low light 'ecotypes' [21]) that are genetically distinct. Conversely, divergent evolution due to local-scale environmental differences select among closely related genotypes and/or populations [22,23]. The net outcome is that clades, species, and even genotypes differ in their functional responses to changes in key resources such as light [21] and CO₂ [24], as well as temperature stress [25]. Phylogeny thus cannot provide an exclusive currency with which to resolve *Symbiodinium* diversity with ecological function.

Disciplines ranging from oceanography [26,27] to plant ecology [28] have overcome such difficulties linking diversity to ecological functioning by turning to the inherent traits ('emergent properties' of individual organisms) that ultimately govern the processes defining ecosystem health. Functional traits provide a standardised measure of the biogeochemical role organisms play, for example photosynthetic rate or nutrient turnover, but also capture fundamental trade-offs with fitness such as investing energy into cellular maintenance versus growth or nutritional mode [26-29]. Functional traits thus define the ecological success of species, and hence

Glossary

Coral bleaching: process in which corals pale (whiten) from loss of *Symbiodinium* cells and/or pigmentation from host coral tissues. First- versus second-order traits: traits that form the foundations for functioning are considered 'central' (first-order) whereas other traits that arise from the operation of first-order traits (or only under some environmental conditions) are considered to be second-order. Fitness: the capacity of an organism to pass its genes to successive generations, as determined by the ability to survive and reproduce by inherent competitive traits.

Photosystem II (PSII): the protein complex that generates electrons for photosynthesis by oxidizing water in algae. Dysfunction of PSII activity is a common assay of heat stress sensitivity in *Symbiodinium*.

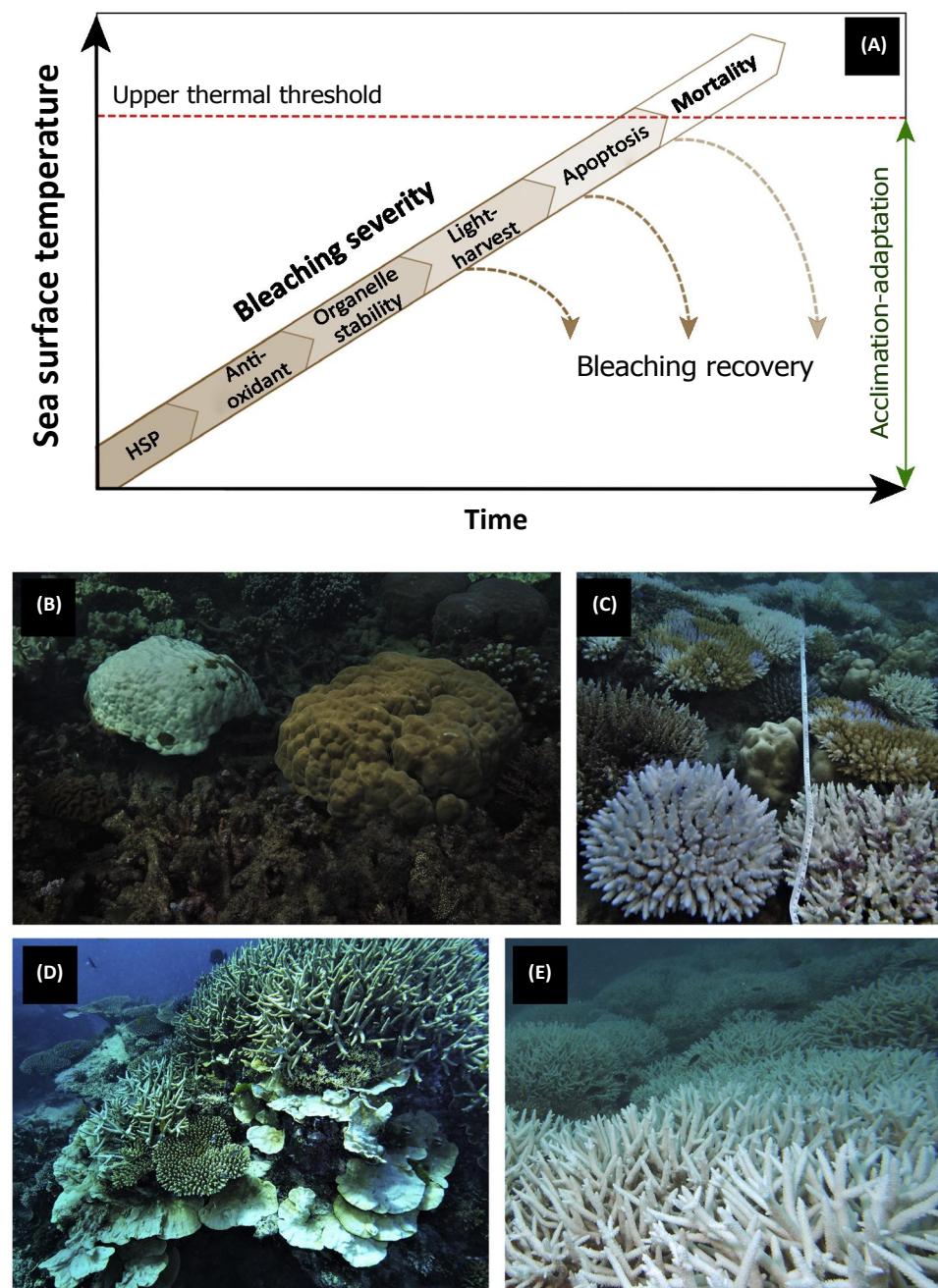
Reactive oxygen species (ROS): chemically active molecules containing oxygen ('free radicals') produced via mitochondria and/or chloroplast metabolic pathways; notably, singlet oxygen, superoxide, and hydrogen peroxide.

Ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCo): the enzyme involved in the first step of CO₂ fixation, considered to be the most abundant enzyme on Earth. It is modulated by the catalytic chaperone RuBisCo activase.

Secondary metabolites: organic compounds that are not directly required for growth and reproduction; for example, toxins and volatiles that negatively affect the fitness of competitors.

Traits: measurable characteristics of an organism that are inherited or environmentally controlled. Can encompass cellular, physiological, morphological, and life-history characteristics.

Vertical versus horizontal transmission: in corals the process where larvae retain *Symbiodinium* cells from parent colonies (vertical transmission or 'closed symbiosis') versus uptake from the surrounding environment (horizontal transmission or 'open symbiosis').



Trends in Ecology & Evolution

Figure 1. Generalised Scheme of Coral–Symbiodinium Functional Trait Responses to Thermal Stress-Induced Bleaching. (A) Increased temperatures over time induce a cascade of responses (Box 1) as the coral–Symbiodinium association is pushed towards the upper thermal thresholds for symbiosis viability ('point of no return'). These responses are either common to both coral and symbiont [upregulation of ROS detoxification networks, reduction to light harvesting through increased host fluorescence protein expression or decreased symbiont light-harvesting complex (LHC) pigments, and induction of apoptosis], or are specific to coral or symbiont (preferential alteration of heat shock proteins, HSPs; mitochondrial versus thylakoid stabilisation for host and symbiont, respectively). Sustained heating at or beyond the threshold causes mortality or requires acclimation or adaptation by selecting for upregulation of the various traits driving tolerance to heat stress (in which case the upper threshold temperature alters by changing how traits conferring thermal tolerance are expressed). Cooling can initiate recovery. How these processes operate across host–Symbiodinium associations in reef systems is highly dynamic and can manifest as different severities of bleaching at (B) intraspecies (e.g., *Porites lutea* colonies) [23] and (C) interspecies levels (e.g., alternative species within the same genus of

(See figure legend on the bottom of the next page.)

Box 1. *Symbiodinium* As Cellular Sources of Coral Stress

Coral host-specific responses that operate to effectively increase thermal tolerance [7] have become increasingly well described; notably reduced oxidative stress via mitochondrial excitation pressure that produces ROS (e.g., fluorescent proteins, tentacular retraction [68]), increased production of ROS detoxifying proteins and organelle stability (e.g., mitochondria [69]), or silencing of ROS-triggered caspases that in turn induce apoptosis [70]. ROS are produced by the coral microbial community, in particular by *Symbiodinium* photosynthetic dysfunction and associated bacterial metabolism, as well as by host mitochondria [71]. However, how physiological dysfunction initiates and progresses to drive ROS emissions remains unresolved (but see [72]); in the case of *Symbiodinium* this has reflected challenges in utilising genomics to unlock the inherent cellular networks and how they are regulated.

Dinoflagellates as a group have a variety of unique characteristics [47], including permanently condensed chromosomes, extremely large genomes, and a significantly higher reliance on post-translational regulation, in contrast to transcriptional regulation, compared to other organisms. Therefore, while the magnitude of gene expression changes is generally less than twofold, up to 30% of the transcriptome can alter [73,74], making it difficult to identify specific responses at this scale. Even so, such tools have begun to highlight parallels with how the host responds to heat stress; notably, a major reorganisation of the ROS antioxidant network in heat-tolerant *Symbiodinium* [73-75], which clearly reflects simultaneous physiological observations of reduced ROS emissions for more heat-tolerant *Symbiodinium* [12,13,73]. Enhancing ROS detoxification capability in fact appears to be a key mechanism by which heat tolerance can be acquired transgenerationally in *Symbiodinium* populations [76]. Such responses may thus be core and hence impose cellular trade-offs to processes that are secondary in affording thermal tolerance; for example, under stress clear upregulation in heat shock proteins (HSPs) is noted for the coral host (>32-fold) whereas *Symbiodinium* may [77] or may not [75] downregulate HSPs. Therefore, although the 'source' of stress is becoming well documented, understanding how this is driven by (or feeds back to) a unifying target regulating *Symbiodinium* cellular dysfunction remains unknown [9-13].

overcome the uncertainties associated with phylogenetic resolution and how it is applied to reconcile ecological success. The central concept of functional diversity is not new to coral ecology, and has in fact been recently considered to be a likely key operational unit driving ecological success of corals [30] and coral-*Symbiodinium* associations [31]. Coral reef management is increasingly turning to knowledge of key traits that regulate (or are indicative of) coral health for innovative management practices [32], while state-of-the-art ecological models that can evaluate winners and losers under complex environmental conditions rely on knowledge of quantifiable traits governing competitive ability [29,33]. However, fundamentally, the 'choice' of trait(s) that best defines *Symbiodinium* functional diversity remains largely unexplored.

***Symbiodinium* spp. Fitness Traits and Trade-Offs**

Understanding the functional roles that underpin the ecological success of *Symbiodinium* spp. within the holobiont landscape ('what makes a good endosymbiont?') demands knowledge of resource acquisition and utilisation. Algal [27] and plant [28] trait-based models commonly rest on end-to-end tracking of resources that govern growth and cellular maintenance, and thus provide a logical conceptual framework. *Symbiodinium* spp. genetic variants have clearly adapted to thrive across a broad range of habitats and host associations where resource availability will differ (Figure 2). Such diversity of ecological niche exploitation and optimisation would suggest major selection pressure for trade-offs among key traits [29,33]; for example, the broad ecological success of phytoplankton can generally be explained via an evolved continuum of 'r' versus 'k' strategies [34,35], whereby cell size operates as a 'master trait' governing allometric scaling rules for light harvesting [33] and inorganic nutrient assimilation [29,36]. *Symbiodinium* spp. genetic variants in fact exist across a cell size continuum, albeit in a relatively narrow range (ca. 7-14 mm), that appears to explain variation in light harvesting but

Acropora), or at the level of (D) alternative genera (e.g., bleaching-sensitive plating *Montipora* sp. versus species of *Acropora*) [2,5] within any given reef area. Monospecific host-*Symbiodinium* sp. associations can manifest as (E) mass bleaching (e.g., *Acropora muricata* beds hosting *Symbiodinium* ITS2 type C3 in the Seychelles). Photographs (B-E) courtesy of Emma Camp, University of Technology Sydney.

Box 2. Species Diversity among the Genus *Symbiodinium*

Molecular tools continue to unlock the immense phylogenetic diversity inherent to the genus *Symbiodinium* [14,16-18]. At the broadest scale *Symbiodinium* spp. are divided into nine distinct evolutionary lineages (i.e., clades, A-I) via divergence of the small ribosomal subunit RNA (SSU). More variable DNA regions, including the internal transcribed spacer regions (ITS), the chloroplast large subunit (cp23S), and cytochrome oxidase b (cob) have subsequently revealed immense subcladal diversity [16], typically classified alphanumerically (e.g., C1, C3z). Of these, ITS2 has been most widely adopted, but requires consideration alongside additional rapidly evolving regions (e.g., psbA_{ref}) to resolve evolutionarily distinct species (multilocus barcoding [78,20]). Integration of barcoding-based phylogeny with fundamental biological (e.g., morphology, physiology) and ecological (e.g., host specificity) patterns has provided the core framework for novel *Symbiodinium* species descriptions [20,37,22]. Molecular platforms have recently transitioned to high-throughput pyrosequencing for barcode retrieval, and added further depth to phylogenetic differentiation through more accurate detection of low-abundance background *Symbiodinium* [17,18,23]. In this case phylogeny is considered within an operational taxonomic unit (OTU) framework to identify ecologically discrete entities [18,23], including the role of intragenomic variability, to resolve taxonomic subgroups [79].

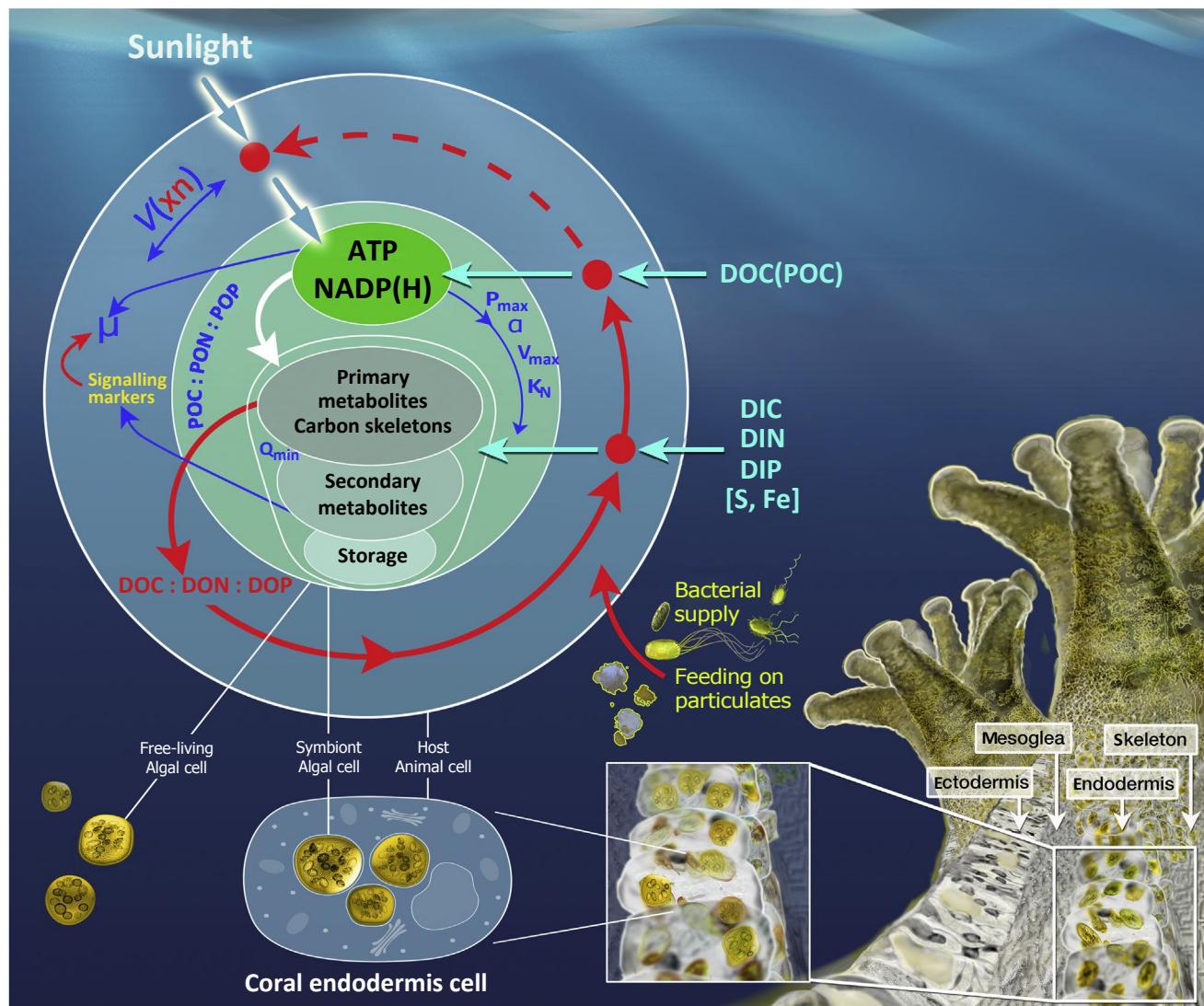
Analysis of genetic recombination has become an important complimentary tool to examine *Symbiodinium* species-level diversity [19]. Using the biological species concept, populations that exchange alleles through sexual recombination are the same species, and hence population genetics based on allele frequency similarity across multiple loci can delimit species over space and time. Such delimitation of species based on this 'incompatible breeding' may be complicated where populations frequently reproduce asexually, but only very rarely sexually, and/or where conspecific populations that previously diverged in isolation of one another become mixed [23]. Even so, the approach has proved powerful for establishing novel *Symbiodinium* species boundaries that persist over broad geographic regions [14].

Molecular-based technical advances thus continue to highlight immense phylogenetic variation and speciation. ITS2 variation alone suggests the existence of 10s to 100s of *Symbiodinium* species, but this is probably an underestimate. Multilocus and high-throughput techniques suggest 100s to 1000s of putative species, and these are changing our ecological view of *Symbiodinium* ITS2 types that were previously considered to be widespread generalist species -for example ITS2 type C3 that is harboured by many highly stress-sensitive coral species of *Acropora* [20,79].

not utilisation capability [37,21]; however, whether this central principle similarly applies to inorganic nutrient acquisition is as yet unexplored.

Dinoflagellates in particular have acquired a broad spectrum of physiological and life-history traits that have enabled ecological diversification beyond the boundaries set by allometric scaling rules ('dirty tricks', *sensu* [38]). An array of strategies associated with light harvesting and photoprotection [39,40] have been relatively well described for *Symbiodinium* spp. However, partitioning *Symbiodinium* spp. genetic variants according to differences in light harvesting and utilisation actually results in few functional groups [13,21], suggesting that trade-offs associated with nutrient acquisition and allocation strategies may in fact be pivotal in explaining their diverse niche exploitation [41] (Figure 2). Such strategies in other microalgae include plasticity of (i) the number of inorganic nutrient uptake ('porter') sites [36,42], which in the case of inorganic carbon is further complicated by the nature of CCMs and RuBisCO affinity (including *Symbiodinium* spp. [24]); (ii) minimum cellular requirements for different inorganic macro- and micronutrients [43], reflecting both pool size (active and stored) and turnover of key constituents that support cellular growth versus maintenance; and (iii) supplementing cellular energy (ATP) production through heterotrophy. *Symbiodinium* spp. are notably active mixotrophs that can supplement their phototrophic metabolism by feeding on bacteria [44] and simple sugars [45].

Accounting for these various factors that are associated with resource acquisition and utilisation introduces immense functional complexity, but it is possible to initially distil this complexity to several first-order measurable traits, as commonly employed for dynamic energy budget (DEB) modelling [46]; specifically, nutrient uptake kinetics and cellular nutrient content relative to growth rate and cell size (Figure 2), which together describe nutrient competitive ability [29]. Algae can preferentially trade-off these resources into opportunistic growth versus persistent maintenance (a classical view of *r* versus *k* selection; e.g., [28]); however,



Trends in Ecology & Evolution

Figure 2. Generalised Scheme of Resource Acquisition and Utilisation Underpinning *Symbiodinium* Competitive Fitness. External light and dissolved inorganic (DIC, DIN, DIP, S, Fe, etc.) and/or organic (POC, DOC) nutrient uptake drive cellular functioning as well as determining the competitive outcome of phytoplankton in general [34]. The generation of energy (ATP) and reductant [NADP(H)] in *Symbiodinium* cell chloroplasts and mitochondria sustains active uptake and assimilation of nutrients into organic compounds. In turn these compounds are stored, drive the formation of primary metabolites (e.g., the key carbon 'skeletons': lipids, carbohydrates, and proteins) and secondary metabolites that may also act in signalling. According to theory developed for phytoplankton [26,29], trade-offs in how these resources are acquired and utilised can explain ecological competitiveness, and generally employ several key terms, highlighted in blue: the extent of light absorption (a), maximum photosynthesis rate (P_{max}), maximum uptake rate (V_{max}), half saturation constant (K_N), and the minimum quota (Q_{min}) for any one nutrient; the quantity and hence stoichiometry of cellular particulate (POC:PON:POP:etc.) and excreted dissolved (DOC:DON:DOP:etc.) nutrients; as well as cell volume (V). Together these terms govern maintenance versus division, and hence the net achievable growth rate (m), to reflect first-order traits of competitive fitness. Such terms are governed by the growth environment and thus will be regulated when cells are *in hospite* (indicated by red lines and arrows) as well as free-living: host corals also acquire external nutrients from both feeding and their broader microbial associations. Hosts modify the inherent light field [68] for *Symbiodinium* and have been suggested to excrete specific dissolved compounds [48] (signalling markers) that control delivery of inorganic nutrients back to the symbiont, in other words distinct metabolites ('host release factors' [56]). Again, the net outcome is regulation of the net achievable growth (m), but *in hospite* will be relative to the overall population size necessary to meet host metabolic demands (*Symbiodinium* cell number, n). All the aforementioned processes play a central role in the trade-off between cellular maintenance in the algae versus the direct release of translocated material to the host coral. Abbreviations: DI(C/N/P), dissolved inorganic carbon/nitrogen/phosphorus; DO(C/N/P), dissolved organic C/N/P; PO(C/N/P), particulate organic C/N/P.

Symbiodinium, as with many other dinoflagellates [47], can potentially short-circuit this trade-off through additional secondary traits that likely disproportionately alter their competitive fitness. Examples of such secondary traits include the extent to which fixed inorganic nutrients are either excreted as dissolved organics [48], including by pathways such as photorespiration that effectively aid photoprotection, or are stored as particulate organics for mobilisation during transient resource limitation [42,43]. Although differences in the biochemical foundation for cellular fitness can be established from knowledge of cellular nutrient (elemental) stoichiometry [43], and hence first-order traits, allocation to specific constituents that enhance fitness are arguably secondary traits of interest. For example, *Symbiodinium* spp. tolerance to stressors that promote bleaching is enhanced not only by increasing protein pools that dissipate reactive oxygen species (ROS; Box 1) but also through the production of biogenic volatile signalling molecules [47].

Metabolic Coupling of Coral–*Symbiodinium* Associations

Functional traits of interest need to span the complex life-history dynamics of *Symbiodinium* where environmental constraints on fitness posed by life in symbiosis are very different from those for free-living algal cells. A defining characteristic to consider initially is the degree of specificity between certain *Symbiodinium* species and their coral hosts, as well as the mode of symbiont acquisition (vertical vs horizontal transmission). Most coral species as adults associate with a single *Symbiodinium* type (or share a few closely related types) [14], although some exceptional coral species may host as many as 5–7 distinct types as codominant [49] or rare [50] populations. Consequently, genetically unique *Symbiodinium* populations may fluctuate in some coral species or persist across others [14]. Even so, types that contribute minimally to the total population pool may ultimately yield a low net metabolic contribution to their host [51]. Coral species that do harbour multiple *Symbiodinium* types in abundance within a single colony appear to reflect complex alga-derived niche partitioning (e.g., photoacclimation to different light levels) [50]. Similarly, for coral species with shifts in dominant *Symbiodinium* type, 'shuffling' is best described in the context of environmental history, for example the thermal trends driving bleaching and subsequent recovery [41,52,53], or complex multivariate interactions of several physical/chemical (temperature, light, nutrient availability) and biological factors acting in tandem [54] that are rarely fully characterised. Unique *Symbiodinium* populations, especially within horizontally transmitted systems, may further represent true localised adaptive radiations to specific *in hospite* environmental conditions [14,20,23]. Thus trait-based characterisation of *Symbiodinium* functional performance is equally appropriate to best describe their realised niche space when *in hospite* as for cells that are free-living.

Metabolic coupling within the coral–*Symbiodinium* relationship is exceptionally complex and likely extends further to the milieu of constituents representing the true holobiont (i.e., bacteria, archaea, fungi, and viruses) [55]. Historically, efforts to understand this coupling have focussed on photosynthetically derived carbon translocation from *Symbiodinium* to coral, in the context of host 'control' over algal populations via nitrogen (N) and metabolite ('host release factor') exchange ([56] for extensive review; [57]) (Figure 2). Much of this work originated from other symbioses (e.g., *Hydra* and the green alga *Chlorella* sp., or in anemone–*Symbiodinium* systems) but provides important evidence for host-controlled N limitation as a source of slower algal mitotic division [56]. First principles would suggest N-limitation to cause an imbalance in the C:N ratio (and hence C:N:P [43]), and in turn reduced symbiont growth, but continued translocation of photosynthetically fixed carbon [57]. However, exposure to inorganic nutrient supplements intriguingly leads to a rebalance of symbiont C:N ratios toward nutrient sufficiency but sustained algal growth arrest [58,59]. External eutrophication events can drive elevated *Symbiodinium* N:P ratios as a result of direct inorganic N stimulation of the alga [59] or fuelling the coral N-fixing bacterial community via indirect DOC enrichment [60]. Such N enrichment drives P-starvation (higher N:P) to result in significant *Symbiodinium* photoinhibition that

exacerbates thermally induced coral bleaching [59]. Conversely, host feeding post 'starvation' re-establishes *Symbiodinium* nutrient quotas and algal growth [58], and substantially ameliorates photoinhibition and coral bleaching during thermal stress [61].

Existing evidence of changes to *Symbiodinium* physiological performance from altered nutrient availability would suggest that cellular nutrient content relative to cell size could provide a first-order measurable trait to evaluate *Symbiodinium* functional diversity and competitive ability *in hospite*, and hence importantly provide a direct comparison currency with cells that are free-living (Figure 2). Bulk elemental stoichiometries could not only encapsulate how the growth environment regulates resource availability relative to inherent requirements across different *Symbiodinium* but also overcome the challenge in balancing symbiont type versus population size (cell size and number of a single genetic variant) to fulfil overall translocation demands [62,63]. Corals that maintain flexibility in association with more than one *Symbiodinium* type would require different population sizes to offset any differences in nutrient uptake and/or release across types to ensure that translocation output is sustained.

Host corals regulate both light [4,64] and CO₂ (dissolved inorganic carbon, DIC) [65] availability for *Symbiodinium* photosynthesis. In fact, the host may regulate DIC delivery more heavily than N or P. When *Symbiodinium* are present, numerous symbiotic anthozoans show substantial transcriptional upregulation for carbonic anhydrase (CA), the enzyme responsible for inter-converting CO₂ and HCO₃⁻, providing DIC for photosynthesis as well as calcification. Recent work has confirmed both external as well as internal CA activity in several corals [66,67], and corals harness a sharp proton gradient to significantly lower the pH (down to ~4.0) surrounding the symbiont sitting within the host-derived membrane, or 'symbiosome', via a vacuolar H - ATPase [65]. Hence, substantial energetic investment by the host supports the DIC demands of photosynthesis [67]. While the dynamics describing the light dependency of *Symbiodinium* photosynthesis *in hospite* are generally well described [64], those describing DIC (indeed other dissolved inorganic nutrients, e.g., N, P) dependency are not; clearly these therefore also represent promising first-order traits with which to define *Symbiodinium* functional performance and how it alters over space and time (Figure 2).

Fundamentally, we have sparse knowledge regarding the cellular nutrient quotas and uptake kinetics for *Symbiodinium*; however, as has been repeatedly demonstrated across other microalgae [26,29], these traits inherently modulate the physiological and competitive response of cells. Clearly this represents an area ripe for exploring the ever-widening gap in knowledge between diversity and ecological success for *Symbiodinium*, which *in hospite* likely drive metabolic trade-offs for the host coral. Inherent nutrient supply, together with *in hospite* light and thermal conditions, may be key attributes in determining the interspecific competitive outcome among different but compatible symbionts. Thus, 'shuffling' of compatible symbionts may have less to do with specific host 'control' but instead reflect an outcome of shifting host metabolic processes [55], and hence a function of trade-offs among first-order algal traits (e.g., cell size, macro- and micronutrient/elemental quotas, and strategies for light/temperature acclimation) [41]. Variation in the first-order traits that determine the cellular energy budget drives broad niche exploitation [46] and hence the scope for functional diversity across *Symbiodinium* genetic variants. However, the exact first-order trait profile will determine the ecological success of any given symbiont; specifically: how, when, and to what extent *Symbiodinium*-coral associations are sustained, and the capacity for *Symbiodinium* to thrive *ex hospite*. In focussing on second-order traits of interest (e.g., ROS production, Box 1) for some time as key factors influencing *Symbiodinium*-coral fitness, we have in fact overlooked the first-order traits that functionally connect *Symbiodinium* to their surrounding environments (Figure 2).

Concluding Remarks

Understanding how *Symbiodinium* spp. are optimised to function across different host corals and reef environments is more crucial than ever as reefs face global 'ecological crisis'. Functional diversity theoretically mediates the response of *Symbiodinium* to changing environmental conditions, and provides a means to reconcile (and complement) the growing wealth of knowledge aligning *Symbiodinium* phylogenetic diversity with coral ecological success. In evaluating key traits that govern cellular growth and physiology, we have proposed a mechanistic physiological framework (Figure 2) that directly complements the rapid uptake of molecular-based descriptors of both *Symbiodinium* phylogeny and function. Trait-based models provide a means to evaluate this physiological framework against ecological success, but only through measuring the key first-order traits. We therefore call for renewed focus into resource acquisition and utilisation as a fundamental regulator of competitive ability (see Outstanding Questions) as a first step to resolve *Symbiodinium* spp. niche boundaries across habitats and specific host–symbiont associations, and hence to resolve the role of *Symbiodinium* in driving productive and diverse coral reefs as they enter an uncertain future.

Acknowledgments

We wish to extend special thanks to John Parkinson and three additional anonymous reviewers whose detailed and insightful comments improved an earlier draft. Funding was provided to D.J.S. and W.L. from an ARC Discovery Project (DP160100271), W.L. from an ARC Centre of Excellence (CE0561435) and to M.E.W. from the US National Science Foundation (IOS-1258065, OCE-1635695).

References

1. Costanza, R. *et al.* (2014) Changes in the global value of ecosystem services. *Glob. Environ. Change* 26, 152–158
2. Hughes, T.P. *et al.* (2017) Global warming and recurrent mass bleaching of corals. *Nature* 543, 373–377
3. Heron, S.F. *et al.* (2016) Warming trends and bleaching stress of the world's coral reefs 1985–2012. *Sci. Rep.* 6, 38402
4. Suggett, D.J. and Smith, D.J. (2011) Interpreting the sign of coral bleaching as friend vs. foe. *Glob. Change Biol.* 17, 45–55
5. Swain, T.D. *et al.* (2016) Coral bleaching response index: a new tool to standardize and compare susceptibility to thermal bleaching. *Glob. Change Biol.* 22, 2475–2488
6. Glynn, P.W. (1993) Coral reef bleaching: ecological perspectives. *Coral Reefs* 12, 1–17
7. Baird, A.H. *et al.* (2009) Coral bleaching: the role of the host. *Trends Ecol. Evol.* 24, 16–20
8. Salvucci, M.E. *et al.* (2001) Exceptional sensitivity of Rubisco activase to thermal denaturation *in vitro* and *in vivo*. *Plant Physiol.* 127, 1053–1064
9. Warner, M.E. *et al.* (1999) Damage to photosystem II in symbiotic dinoflagellates: a determinant of coral bleaching. *Proc. Natl. Acad. U. S. A.* 96, 8007–8012
10. Jones, R.J. *et al.* (1998) Temperature-induced bleaching of corals begins with impairment of the CO₂ fixation mechanism in zooxanthellae. *Plant Cell Environ.* 21, 1219–1230
11. Takahashi, S. *et al.* (2008) Heat stress causes inhibition of the *de novo* synthesis of antenna proteins and photobleaching in cultured *Symbiodinium*. *Proc. Natl. Acad. Sci. U. S. A.* 105, 4203–4208
12. Tchernov, D. *et al.* (2004) Membrane lipids of symbiotic algae are diagnostic of sensitivity to thermal bleaching in corals. *Proc. Natl. Acad. Sci. U. S. A.* 101, 13531–13535
13. Goyen, S. *et al.* (2017) A molecular physiology basis for functional diversity of hydrogen peroxide production amongst *Symbiodinium* spp. (Dinophyceae). *Mar. Biol.* 164, 46
14. Thornhill, D.J. *et al.* (2017) Population genetics of reef coral endosymbionts (*Symbiodinium*, Dinophyceae). *Mol. Ecol.* 26, 2640–2659
15. Silverstein, R.N. *et al.* (2017) Tenacious *D. Symbiodinium* in clade D remain in reef corals at both high and low temperature extremes despite impairment. *J. Exp. Biol.* 220, 1192–1196
16. Sampayo, E.M. *et al.* (2009) Cohesive molecular genetic data delineate species diversity in the dinoflagellate genus *Symbiodinium*. *Mol. Ecol.* 18, 500–519
17. Arif, C. *et al.* (2014) Assessing *Symbiodinium* diversity in scleractinian corals via next-generation sequencing-based genotyping of the ITS2 rDNA region. *Mol. Ecol.* 23, 4418–4433
18. Ziegler, M. *et al.* (2017) Biogeography and molecular diversity of coral symbionts in the genus *Symbiodinium* around the Arabian Peninsula. *J. Biogeogr.* 44, 674–686
19. Pettay, D.T. *et al.* (2015) Microbial invasion of the Caribbean by an Indo-Pacific coral zooxanthella. *Proc. Natl. Acad. Sci. U. S. A.* 112, 7513–7518
20. Hume, B.C.C. *et al.* (2016) Ancestral genetic diversity associated with the rapid spread of stress-tolerant coral symbionts in response to Holocene climate change. *Proc. Natl. Acad. Sci. U. S. A.* 113, 4416–4421
21. Suggett, D.J. *et al.* (2015) Functional diversity of photobiological traits within the genus *Symbiodinium* appears to be governed by the interaction of cell size with cladal designation. *New Phytol.* 208, 370–381
22. Parkinson, J.E. *et al.* (2015) New species of clade B *Symbiodinium* (Dinophyceae) from the greater Caribbean belong to different functional guilds: *S. aenigmatum* sp. nov., *S. antillogorgium* sp. nov., *S. endomadracis* sp. nov., and *S. pseudominutum* sp. nov. *J. Phycol.* 51, 850–858
23. Howells, E.J. *et al.* (2016) Microsatellite allele sizes alone are insufficient to delineate species boundaries in *Symbiodinium*. *Mol. Ecol.* 25, 2719–2723
24. Bradling, P. *et al.* (2013) Contrasting modes of inorganic carbon acquisition amongst *Symbiodinium* (Dinophyceae) phyotypes. *New Phytol.* 200, 432–442
25. Swain, T.D. *et al.* (2017) Consensus thermotolerance ranking for 110 *Symbiodinium* phyotypes: an exemplar utilization of a novel iterative partial-rank aggregation tool with broad application potential. *Funct. Ecol.* 31, 172–183
26. Mutshinda, C.M. *et al.* (2016) Ecological equivalence of species within phytoplankton functional groups. *Funct. Ecol.* 30, 1714–1722
27. Mitra, A. *et al.* (2016) Defining planktonic protist functional groups on mechanisms for energy and nutrient acquisition: incorporation of diverse mixotrophic strategies. *Protist* 167, 106–120

Outstanding Questions

Can first-order traits (e.g., cellular uptake and allocation) alone explain niche breadth, including anomalous stress tolerance, or must they be considered alongside second order-trait (e.g., capacity to upregulate ROS detoxification pathways)?

How diverse are micro- (trace) relative to macronutrient (C, N, P, S) uptake and utilization properties in describing *Symbiodinium* 'functional types'?

Does ecological resilience through 'symbiont shuffling' reflect match/mismatch between changing host metabolic requirements and *Symbiodinium* consortia with alternative metabolic (nutrient uptake and allocation) profiles?

What extent of metabolic and resource trade-off is necessary to persist across alternative life-history stages? Does *Symbiodinium* require 'host resource surrogates' as a free-living alga (e.g., obligate associations with other microbes)?

How important are heterotrophic strategies for supporting *Symbiodinium* nutritional and/or metabolic requirements? Can *Symbiodinium* feed on host as well as microbial metabolites?

To what extent does functional diversity of nutrient strategy reflect evolutionary radiation (and/or potentially support phylogenetic reconstructions) of the genus *Symbiodinium*?

28. Reisch, P.B. (2014) The world-wide 'fast–slow' plant economics spectrum: a traits manifesto. *J. Ecol.* 102, 275–301

29. Litchman, E. *et al.* (2007) The role of functional traits and trade-offs in structuring phytoplankton communities: scaling from cellular to ecosystem level. *Ecol. Lett.* 10, 1170–1181

30. Madin, J.S. *et al.* (2016) A trait-based approach to advance coral reef science. *Trends Ecol. Evol.* 31, 419–426

31. Parkinson, J.P. *et al.* (2015) Intraspecific diversity among partners drives functional variation in coral symbioses. *Sci. Rep.* 5, 15667

32. van Oppen, M.J.H. *et al.* (2017) Shifting paradigms in restoration of the world's coral reefs. *Glob. Change Biol.* Published online March 1, 2017. <http://dx.doi.org/10.1111/gcb.13647>

33. Edwards, K.F. *et al.* (2015) Light and growth in marine phytoplankton: allometric, taxonomic, and environmental variation. *Limnol. Oceanogr.* 60, 540–552

34. Kilham, P. and Hecky, R.E. (1988) Comparative ecology of marine and freshwater phytoplankton. *Limnol. Oceanogr.* 33, 776–795

35. Smayda, T.J. and Reynolds, C.S. (2001) Community assembly in marine phytoplankton: application of recent models to harmful dinoflagellate blooms. *J. Plankton Res.* 23, 447–461

36. Aksnes, D.L. and Cao, F.J. (2011) Inherent and apparent traits in microbial nutrient uptake. *Mar. Ecol. Prog. Ser.* 440, 41–51

37. LaJeunesse, T.C. *et al.* (2012) A genetics-based description of *Symbiodinium minutum* sp. nov. and *S. psymophilum* sp. nov. (Dinophyceae), two dinoflagellates symbiotic with cnidarian. *J. Phycol.* 48, 1380–1391

38. Thingstad, T.F. (1998) Theoretical approach to structuring mechanisms in the pelagic food web. *Hydrobiologia* 363, 59–72

39. McCabe-Reynolds, J. *et al.* (2008) Enhanced photoprotection pathways in symbiotic dinoflagellates of shallow-water corals and other cnidarians. *Proc. Natl. Acad. Sci. U. S. A.* 105, 13674–13678

40. Slavov, C. *et al.* (2016) 'Super-quenching' state protects *Symbiodinium* from thermal stress – implications for coral bleaching. *Biochim. Biophys. Acta* 1857, 840–847

41. Baker, D.M. *et al.* (2013) Nitrate competition in a coral symbiosis varies with temperature among *Symbiodinium* clades. *ISME J.* 7, 1248–1251

42. Lindemann, C. *et al.* (2016) Scaling laws in phytoplankton nutrient uptake affinity. *Front. Mar. Sci.* 3, 26

43. Moore, C.M. *et al.* (2013) Processes and patterns of oceanic nutrient limitation. *Nat. Geosci.* 6, 701–710

44. Jeong, H.J. *et al.* (2008) Heterotrophic feeding as a newly identified survival strategy of the dinoflagellate *Symbiodinium*. *Proc. Natl. Acad. Sci. U. S. A.* 109, 12604–12609

45. Xiang, T. *et al.* (2013) Isolation of clonal axenic strains of the symbiotic dinoflagellate *Symbiodinium* and their growth and host specificity. *J. Phycol.* 49, 447–458

46. Geček, S. (2017) Autotrophs' challenge to dynamic energy budget theory: comment on 'Physics of metabolic organization' by Marko Jusup *et al.* *Phys. Life Rev.* 20, 46–48

47. Murray, S.A. *et al.* (2016) Unravelling the functional genetics of dinoflagellates: a review of approaches and opportunities. *Perspect. Phycol.* 3, 37–52

48. Hillyer, K.E. *et al.* (2017) Mapping carbon fate during bleaching in a model cnidarian symbiosis: the application of ^{14}C metabolism. *New Phytol.* 214, 1551–1562

49. Kemp, D.W. *et al.* (2015) Spatially distinct and regionally endemic *Symbiodinium* assemblages in the threatened Caribbean reef-building coral *Orbicella faveolata*. *Coral Reefs* 34, 535–547

50. Boulotte, N.M. *et al.* (2016) Exploring the *Symbiodinium* rare biosphere provides evidence for symbiont switching in reef-building corals. *ISME J.* 10, 2693–2701

51. Lee, M.J. *et al.* (2016) Most low-abundance background *Symbiodinium* spp. are transitory and have minimal functional significance for symbiotic corals. *Microb. Ecol.* 71, 771–783

52. Grottoli, A.G. *et al.* (2014) The cumulative impact of annual coral bleaching can turn some coral species winners into losers. *Glob. Change Biol.* 20, 3823–3833

53. Cunning, R. *et al.* (2015) Investigating the causes and consequences of symbiont shuffling in a multi-partner reef coral symbiosis under environmental change. *Proc. Biol. Sci.* 282, 20141725

54. Kennedy, E.V. *et al.* (2016) *Symbiodinium* biogeography tracks environmental patterns rather than host genetics in a key Caribbean reef-builder, *Orbicella annularis*. *Proc. R. Soc. B* 283, 20161938

55. Aranda, M. *et al.* (2016) Genomes of coral dinoflagellate symbionts highlight evolutionary adaptations conducive to a symbiotic lifestyle. *Sci. Rep.* 6, 39734

56. Davy, S.K. *et al.* (2012) Cell biology of cnidarian–dinoflagellate symbiosis. *Microbiol. Mol. Biol. Rev.* 76, 229–261

57. Dubinsky, Z. and Berman-Frank, I. (2001) Uncoupling primary production from population growth in photosynthesizing organisms in aquatic ecosystems. *Aquat. Sci.* 63, 4–17

58. Fitt, W.K. and Cook, C.B. (2001) The effects of feeding or addition of dissolved inorganic nutrients in maintaining the symbiosis between dinoflagellates and a tropical marine cnidarian. *Mar. Biol.* 13, 507–517

59. Wiedermann, J. *et al.* (2012) Nutrient enrichment can increase the susceptibility of reef corals to bleaching. *Nat. Clim. Change* 3, 160–164

60. Pogoreutz, C. *et al.* (2017) Sugar enrichment provides evidence for a role of nitrogen fixation in coral bleaching. *Glob. Change Biol.* Published online April 21, 2017. <http://dx.doi.org/10.1111/gcb.13695>

61. Tolosa, I. *et al.* (2011) Impact of feeding and short-term temperature stress on the content and isotopic signature of fatty acids, sterols, and alcohols in the scleractinian coral *Turbinaria reniformis*. *Coral Reefs* 30, 763–774

62. Leal, M.C. *et al.* (2015) Symbiont type influences trophic plasticity of a model cnidarian–dinoflagellate symbiosis. *J. Exp. Biol.* 218, 858–863

63. Cunning, R. *et al.* (2015) Dynamic regulation of partner abundance mediates response of reef coral symbioses to environmental change. *Ecology* 96, 1411–1420

64. Anthony, K.R.N. and Hoegh-Guldberg, O. (2003) Variation in coral photosynthesis, respiration and growth characteristics in contrasting light microhabitats: an analogue to plants in forest gaps and understoreys? *Funct. Ecol.* 17, 246–259

65. Barott, K.L. *et al.* (2014) Coral host cells acidify symbiotic algal microenvironment to promote photosynthesis. *Proc. Natl. Acad. Sci. U. S. A.* 112, 607–612

66. Tansik, A.L. *et al.* (2015) External carbonic anhydrase in three Caribbean corals: quantification of activity and role in CO_2 uptake. *Coral Reefs* 34, 703–713

67. Tansik, A.L. *et al.* (2017) Inorganic carbon is scarce for symbionts in scleractinian corals. *Limnol. Oceanogr.* Published online April 3, 2017. <http://dx.doi.org/10.1002/limo.10550>

68. Palmer, C.V. *et al.* (2009) Coral fluorescent proteins as antioxidants. *PLoS One* 4, e7298

69. Dixon, G.B. *et al.* (2015) Genomic determinants of coral heat tolerance across latitudes. *Science* 348, 1460

70. Tchernov, D. *et al.* (2011) Apoptosis and the selective survival of host animals following thermal bleaching in zooxanthellate corals. *Proc. Natl. Acad. Sci. U. S. A.* 108, 9905–9909

71. Diaz, J.M. *et al.* (2016) Species-specific control of external superoxide levels by the coral holobiont during a natural bleaching event. *Nat. Commun.* 7, 13801

72. Hawkins, T.D. and Warner, M.E. (2017) Warm preconditioning protects against acute heat-induced respiratory dysfunction and delays bleaching in a symbiotic sea anemone. *J. Exp. Biol.* 220, 969–983

73. Levin, R.A. *et al.* (2016) Sex, scavengers, and chaperones: transcriptome secrets of divergent *Symbiodinium* thermal tolerances. *Mol. Biol. Evol.* 33, 2201–2215

74. Gierz, S.L. *et al.* (2017) Transcriptomic analysis of thermally stressed *Symbiodinium* reveals differential expression of stress and metabolism genes. *Front. Plant Sci.* 8, 271

75. Baumgarten, S. *et al.* (2013) Integrating microRNA and mRNA expression profiling in *Symbiodinium microadriaticum*, a dinoflagellate symbiont of reef-building corals. *BMC Genomics* 14, 704

76. Chakravarti, L. *et al.* (2017) Rapid thermal adaptation in photosymbionts of reef-building corals. *Glob. Change Biol.* Published online April 27, 2017. <http://dx.doi.org/10.1111/gcb.13702>
77. Leggat, W. *et al.* (2011) Differential responses of the coral host and their algal symbiont to thermal stress. *PLoS One* 6, e26687
78. LaJeunesse, T.C. and Thornhill, D.J. (2011) Improved resolution of reef-coral endosymbiont (*Symbiodinium*) species diversity, ecology, and evolution through *psbA* non-coding region genotyping. *PLoS One* 6, e29013
79. Smith, E.G. *et al.* (2017) Host specificity of *Symbiodinium* variants revealed by an ITS2 metahaplotype approach. *ISME J.* 11, 1500–1503