

**Merging “Morphology/Performance/Fitness” and Life-History Theories: An Empirical Assessment in the Eagle Lake Garter Snake Research Project**

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Complete List of Authors:	Addis, Elizabeth; Gonzaga University, Biology Gangloff, Eric; Iowa State University, Ecology, Evolution & Organismal Biology Palacios, Maria; CONICET CENPAT Carr, Katherine; Gonzaga University, Biology Bronikowski, Anne; Iowa State University, Ecology, Evolution & Organismal Biology
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Manuscripts

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5 Assessment in the Eagle Lake Garter Snake Research Project.  
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13 Elizabeth A. Addis <sup>\*,†,‡</sup> Eric Gangloff <sup>\*,‡</sup>, Maria G. Palacios <sup>\*,§</sup>, Katherine E. Carr <sup>†</sup>, Anne M.  
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15 Bronikowski <sup>\*,1</sup>  
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20 <sup>‡</sup>The first two authors contributed equally to this work.  
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25 <sup>\*</sup>Department of Ecology, Evolution, and Organismal Biology, Iowa State University, Ames, IA  
26  
27 50011, USA; <sup>†</sup>Department of Biology, Gonzaga University, Spokane, WA 99258 USA; <sup>§</sup>Centro  
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<sup>1</sup>E-mail: abroniko@iastate.edu

RRH: Ecomorphology and life history of garter snakes

## Synopsis

The morphology-performance-fitness method for estimating selection on morphological traits has seen decades of successful application. At the same time, life-history approaches using matrix methods and perturbation studies have also allowed the direct estimate of selection acting on vital rates and the traits that comprise them. Both methodologies have been successfully applied to the garter snakes of the long-term Eagle Lake research project to reveal selection on morphology, such as color pattern, number of vertebrae, gape size; and life-history traits such, such as birth size, growth rates, and juvenile survival. Here we conduct a reciprocal transplant study in a common laboratory environment to partition genetic and environmental sources of variation in morphology and life-history. To place our results in the ecomorphology paradigm, we measure performance outcomes (feeding, growth, insulin growth factor 1 titres) of morphological variation (body size, condition) and their fitness consequences for juvenile survival – a trait that has large fitness sensitivities in these garter snake populations and therefore to be subject to strong selection. To better merge these two complementary theories, we end by discussing our findings in a continuum of morphology – performance – fitness – life-history to highlight what these approaches, when combined, can reveal about selection in the wild.

## Introduction

The “morphology-performance-fitness” paradigm was put forth by Stevan Arnold in these pages (Arnold 1983) from a SICB (formerly American Society of Zoologists) 1981 symposium on snake feeding mechanisms. In that paper, Arnold presented a statistical method to test for the fitness consequences, and thereby adaptive significance, of changes to morphology that impact fitness through organismal performance traits, such as behavior or physiology. This ecomorphology theory included regression methods for tracing the effects of morphological variation on organismal performance, and in turn, on an individual’s fitness. When framed in microevolutionary selection theory, the strength of natural selection on these phenotypes could be quantitatively assessed (Arnold and Wade 1984b; Lande and Arnold 1983) and the adaptive significance of this variation tested. Ensuing studies of morphology/performance/fitness – i.e., the fitness consequences of morphological variation as mediated through behavior and physiology – helped to change the field of micro-evolution from adaptive story-telling (Gould and Lewontin 1979) to a comprehensive and robust assessment of adaptation in diverse taxa (Dudley 1996; Losos 1990; Schluter 1995).

At the same time, the field of life-history theory (Stearns 1992), and the advent of matrix methods to test for the adaptive significance of variation in life-history traits (Caswell 2001) – complex traits that would not yield easily to the ecomorphology paradigm - rapidly enhanced our ability to ask how and when variation in life-history traits (themselves mediated by morphology and performance) could reveal adaptation. Specifically, matrix models of population demography allowed for the quantification of the strength of natural selection on individual vital rates and life-history traits, as well as comprehensive tests for adaptive significance of such variation – i.e., parallel questions could be addressed both in the realms of morphology/behavior

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3 and life-history/demography. Because variation in vital rates (such as birth rates, juvenile  
4 survival, etc.) are underlain by variation in morphology and performance, these two theoretical  
5 constructs have a straight-forward complementarity in their use to assess selection gradients  
6  
7 (Lande 1982b).

8  
9 Arnold's long-term Eagle Lake garter snake study system (Lassen County, California)  
10 has figured prominently in empirical tests of ecomorphology theory and, more recently, in  
11 empirical tests of life-history theory. On the ecomorphology side, Arnold's pioneering work on  
12 the adaptive significance of morphological variation – feeding mechanisms, scale counts, and  
13 vertebral variation - produced textbook examples of the utility of using this framework to draw  
14 strong inferences on the adaptive significance of morphological variation e.g., (Arnold 1992;  
15 Arnold and Wade 1984a; Ayres and Arnold 1983; Kelley and others 1997; Manier and others  
16 2007). see also (Arnold and Bennett 1984) for *T. radix* example. Furthermore, this paradigm was  
17 extended to link behavioral variation to subsequent variation in performance and fitness (cf.  
18 (Arnold 1988). At the same time, discovered through our decades of mark/recapture efforts,  
19 these same populations of Eagle Lake garter snakes harbor two distinct life-history phenotypes  
20 whose study have provided examples of evolution of life histories within closely located  
21 populations (Bronikowski 2000; Bronikowski and Arnold 1999; Miller and others 2011; Miller  
22 and others 2014).

23  
24 Conventionally, this ecomorphological theory has been concerned with measuring  
25 traditional morphology (e.g., coloration, shape, number of vertebrae) and mapping these traits in  
26 a statistical model with traditional performance traits (e.g., prey capture, slither speed) and  
27 fitness. We contend that the possibilities for considering performance are much broader and  
28 include traits that represent rates or endpoints of rates. For example, in our previous work on the  
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3 Eagle Lake garter snake system, we have measured non-traditional estimates of performance  
4 such as innate and acquired immune function (measures of natural antibodies production,  
5 bactericidal capability, and mitogen-stimulated lymphocyte proliferation (Palacios and others  
6 2013; Palacios and others 2011)); hormonal titers of glucocorticoids ((Palacios and others 2012)  
7 and IGF-1 (Reding and others 2016; Sparkman and others 2009) because they are the main  
8 mediators of stress and growth hormone signaling, respectively; and metabolic function  
9 (Gangloff and others 2015). Measurement of these traits are in addition to more standard  
10 measures of performance such as growth efficiency and feeding behavior (Gangloff and others  
11 2015). Of these, we focus on the endocrine system in this study, proposing that it mediates  
12 relationships among morphology, fitness, and life-history traits similar to its role in mediating  
13 trade-offs among life-history traits (Ketterson and Nolan 1999; Lailvaux and Husak 2014;  
14 Ricklefs and Wikelski 2002; Zera and Bottsford 2001). Specifically, insulin-like growth factor-1  
15 (IGF1) facilitates variation in life-histories (e.g., (Dantzer and Swanson 2012; Lewin and others  
16 2017; Reding and others 2016; Schwartz and Bronikowski 2016). IGF1, the primary hormone of  
17 the insulin/insulin-like signaling (IIS) system, is a peptide hormone that is a paralog of insulin  
18 and has been highly conserved across amniotes (Annunziata and others 2011; Denley and others  
19 2005; Duan 1998; McGaugh and others 2015; Sparkman and others 2012; Zhu and others 2017).  
20 On the cellular level, IGF1 stimulates cell proliferation, differentiation, and migration, while on  
21 the organismal scale it triggers overall growth and maturation.  
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Lailvaux and Husak (Lailvaux and Husak 2014) argued convincingly to make the  
linkages between ecomorphology and life-history direct and explicit. Following their lead, to  
better merge ecomorphology and life-history theory, and to make clear the connections between  
them, we undertake here an explicit test of the relationships among morphology, performance,

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3 fitness, life-history, and demography in the Eagle Lake populations of garter snakes. Because of  
4 the clear function that IGF1 has for organismal growth, and because of the association of IGF1  
5 with life-history phenotypes in the Eagle Lake garter snake system, we focus our performance  
6 measures on IGF1, feeding, and growth. Specifically, we address: 1) How does morphological  
7 variation at birth translate into variation in performance - measured as feeding, growth, and  
8 circulating IGF1? 2) How do variation in these morphological and performance traits impact  
9 fitness – measured as survival until sexual maturation? And 3) how do phenotypes defined along  
10 specific morphology, performance, fitness trajectories map to variation in population  
11 demographic estimates of fitness – measured as population growth rates?

## 24 Methods

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### 26 Study system

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28 Populations of western terrestrial garter snakes (*Thamnophis elegans*) in the vicinity of Eagle  
29 Lake (Lassen County), California are characterized by either slow or fast-paced life history  
30 ecotypes. Lower elevation lakeshore snakes exhibit fast growth, large adult body size, early  
31 reproductive maturation with large reproductive effort, and low annual survival. Higher elevation  
32 meadow snakes exhibit slow growth, smaller adult body size, later maturation, low annual  
33 reproduction, high annual survival, and measurable demographic senescence with advancing  
34 adult age (“L-fast” versus “M-slow” ecotypes; (Bronikowski and Arnold 1999; Schwartz and  
35 others 2015). Selection gradients vary between these fast and slow ecotypes for survival and  
36 growth in the pre-adult stages (Miller and others 2011). As well, genetic differentiation persists  
37 among populations of contrasting ecotypes with significant  $F_{ST}$  for both nuclear markers (Manier  
38 and others 2007), and mitochondrial genomes (Schwartz and others 2015). Many ecological  
39 factors differ between the two ecotypes: predation rates are higher in the lakeshore habitat  
40

(Sparkman and others 2013) as is prey abundance and average temperature (Bronikowski and Arnold, 1999). In wild-caught animals, variation in total plasma IGF-1 is driven by interactions among body size (snout-to-vent length), ecotype, and life-history stage. IGF-1 is also higher in wild-caught pregnant snakes Sparkman and others (2009). At the same time, IGF-1 is sensitive to ecological variation: levels vary across year and covary with precipitation – likely due to the relationship between precipitation and prey availability (Miller and others 2011; Sparkman and others 2009).

## Study animals and treatment groups

### Wild IGF1 and body size

To assess the long-term trends in IGF1 plasma levels versus body size in wild animals, we assayed IGF1 plasma concentrations in an additional set (N=31) of wild-caught animals (males and non-pregnant females) from 2009 and 2010 to complement our published study of IGF1 concentration in the wild from 2006 – 2008 (Sparkman and others 2009).

### Cohort morphology, growth, feeding, and survival

In June 2010, we collected 40 pregnant females from six populations of *Thamnophis elegans* around Eagle Lake, California: three replicate populations of each of the lakeshore (L-fast) and meadow (M-slow) ecotypes. We transported these pregnant females to Iowa State University and housed them individually through gestation in 10-gallon glass aquaria with ground corncob substrate and a plastic bowl that served as both water dish and retreat site. We maintained them in a 20°C room with their tanks placed on a thermal gradient for 24 hours per day (range: 25-

34°C across each tank), kept on a 12:12 light:dark schedule, and offered 1-2 mice once a week until parturition.

Offspring ( $n=257$ ) from these 40 pregnant females were born between 12 August and 19 September 2010 (Table 1). Within 24 hours of birth, we measured offspring sex, weight, and snout-vent length (SVL, mm), and placed individual neonates in plastic home cages with paper substrate and a water bowl. We divided litters randomly, with sex split nearly evenly, into two temperature-treatment groups designed to mimic the differing thermal regimes of the warmer L-fast and cooler M-slow habitats (Bronikowski 2000). Ambient room temperature was 20°C, with the warmer rearing treatment (“warm”) receiving 16 hours of supplemental heating per day and the cooler rearing treatment (“cool”) receiving supplemental heating for only 8 hours per day. This supplemental heating, supplied by under-tank heating elements, provided a gradient of 22°C to 32°C in each home box during heating (20°C during hours of no heat), allowing the animals to behaviorally thermoregulate. All juveniles were kept on a 12:12 light:dark schedule and offered thawed pinky mice once a week. Individuals that consumed all food were offered a greater amount in subsequent feedings. At every feeding, food consumption was recorded in grams of mouse eaten. L-fast and M-slow juveniles residing in the warm and cool treatments are referred to throughout as Lwarm, Lcool, Mwarm, and Mcool. Animals were housed accordingly for life.

#### Repeated measures of IGF1 and growth

A subset of the full colony, sampled from each sex  $\times$  ecotype  $\times$  rearing treatment group, were sampled for short-term repeated measures analysis of total plasma IGF1 concentration and growth to assess the relationship between IGF1 titer and short-term growth rate. These 130 snakes from the 2010 cohort (Table 1) were weighed and measured on 6 May 2011; 14 Sept.

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3 2011; 1 November 2011; and 12 January 2012 – approximately 10, 14, 16, and 18 months of age  
4 – and blood-sampled on the latter three dates. Blood was collected from the caudal vein in  
5 heparin-rinsed and dried syringes. Plasma was separated from red blood cells by centrifugation  
6 and was frozen and stored at -80°C. Treatment of all experimental animals was in accordance  
7 with Iowa State University Institutional Animal Care and Use Committee protocol #3-2-5125-J.  
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### IGF-1 Radioimmunoassay

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22 Plasma IGF-1 concentrations were determined by radioimmunoassay (RIA) using GroPep  
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24 Protocol #3002 (GroPep Ltd., Adelaide, Australia) via competitive binding with  $^{125}\text{I}$  labeled IGF-  
25 1 (PerkinElmer #NEZ033). This assay uses an anti-human IGF-1, but was validated for the  
26 Western terrestrial garter snake by Sparkman and others (2009). IGF binding proteins were  
27 removed from IGF-1 by acid-ethanol extraction. All samples were assayed in duplicate within a  
28 single assay. Intra-assay variation was 7.5%.  
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### Statistical analysis

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40 All data were analyzed using SAS v9.4 (SAS Institute, Cary, NC). Significance was set at  $\alpha =$   
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42 0.05. All data were first inspected for normality and homogeneity of variances and transformed  
43 to meet these assumptions as necessary (noted below). In all linear mixed model analyses,  
44 denominator degrees of freedom for  $F$ -tests were estimated using the Kenward-Roger Degrees of  
45 Freedom Approximation, which weights the denominator degrees of freedom according to the  
46 variance of the effect (Kenward and Roger 1997). Figures were made with the ‘ggplot’ package  
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48 Freedom Approximation, which weights the denominator degrees of freedom according to the  
49 variance of the effect (Kenward and Roger 1997). Figures were made with the ‘ggplot’ package  
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51 (Wickham 2009) for R (R Core Team 2014).  
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5 Wild IGF1 and body size  
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8 We analyzed blood plasma concentrations of IGF-1 in field-caught animals, excluding  
9 reproductive females, from both ecotypes across five years (2006-2010). Data from 2006 – 2008  
10 were from Sparkman and others (2009). Data for 2009 and 2010 were from our standard blood  
11 field collecting. We utilized generalized linear models and concentrations were  $\log_{10}$ -  
12 transformed to meet assumptions of normality. The initial model included the main effects of  
13 size (SVL in mm), ecotype (L-fast, M-slow), year (2006 – 2010), and life-history stage (adult or  
14 juvenile, with juvenile defined as snakes with SVL < 425 mm for L-fast and < 400 mm for M-  
15 slow; Sparkman and others 2009), the four-way interaction of these effects, and all lower-order  
16 interactions. We retained statistically significant interactions and those of biological interest,  
17 resulting in a final model of:  
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$$34 \log_{10}IGF1 = \mu + SVL + Ecotype + Life\ stage + Year + SVL \times Ecotype + SVL \times Life\ stage + \\ 35 \\ 36 \qquad \qquad \qquad SVL \times Year + SVL \times Ecotype \times Life\ stage + \varepsilon \\ 37 \\ 38 \\ 39 \\ 40$$

41 where  $\mu$  represents the grand mean and  $\varepsilon$  is the error term.  
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46 Cohort morphology, growth, feeding, and survival  
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49 Morphology

50 We analyzed body size across lifetime as the snout-vent length (SVL in mm) of individual  
51 snakes from birth until death. We used mixed linear models to test for the effects of ecotype,  
52 rearing treatment (warm/cool), age (in days, assuming an average birthday of 1 Sept. 2010), and  
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3 interactions thereof, on body size. We also included sex as a fixed effect in this model, though no  
4 interactions with this term because models did not converge when a large number of interactions  
5 were included and these interactions are not of primary biological concern. Population nested  
6 within ecotype was included in the model as a fixed effect to account for among-population  
7 habitat heterogeneity within ecotypes (Palacios and others 2013). We also included the random  
8 effect of litter nested within population and ecotype, which accounts for among-litter variation  
9 within populations (Robert and Bronikowski 2010). The final mixed linear models used for  
10 analysis of body size was:

$$22 \quad SVL = \mu + Ecotype + Rearing Treatment + Age + Sex + Ecotype \times Rearing Treatment + \\ 23 \\ 24 \quad Ecotype \times Age + Rearing Treatment \times Age + Ecotype \times Rearing Treatment \times Age + \\ 25 \\ 26 \quad Population(Ecotype) + Litter(Population Ecotype) + \varepsilon$$

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32 Growth  
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34 We analyzed growth as change in snout-vent length (SVL in mm) from the first uniform  
35 measurement date (29 Nov. 2010) to 10 Dec. 2014. We used mixed linear models to test for the  
36 effects of ecotype (L-fast, M-slow), rearing treatment (warm, cool), sex, and interactions thereof,  
37 on growth. To account for variation in growth due to size, we used initial size (i.e., at 29 Nov.  
38 2010, approximately 2 months post-birth) as a covariate in the analysis. To account for elapsed  
39 time, we used number of growth days (days between 29 Nov. 2010 and an individual's final  
40 length measurement). As above, we included population nested within ecotype as a fixed effect  
41 and litter nested within population and ecotype as a random effect. The interaction of ecotype  $\times$   
42 rearing treatment was left in the model as this interaction is of biological interest; the remaining  
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3 non-significant interaction terms were removed from the model. The final mixed linear model  
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5 used for our analysis of lifetime growth was:  
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10  $Change\ in\ SVL = \mu + Growth\ Days + Initial\ Size + Ecotype + Rearing\ Treatment + Sex +$   
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12  $Ecotype \times Rearing\ Treatment + Population(Ecotype) + Litter(Population\ Ecotype) + \varepsilon$   
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16 Feeding  
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19 We analyzed the amount of food consumed (the number of fractional pieces of frozen/thawed  
20 pinkies consumed, converted to grams) from 29 Nov. 2010 until death. Covariates included  
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22 Feeding Days, which is the number of days between 29 Nov. 2010 and death, and Initial Size,  
23 which is the 29 Nov. 2010 SVL. Using a model-reduction process identical to the above, we  
24 removed non-significant interaction terms and utilized this final model of lifetime food  
25 consumption:  
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29  $Amount\ Consumed = \mu + Feeding\ Days + Initial\ Size + Ecotype + Rearing\ Treatment + Sex +$   
30  
31  $Ecotype * Rearing\ Treatment + Population(Ecotype) + Litter(Population\ Ecotype) + \varepsilon$   
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34 Survival  
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37 We analyzed survivorship of captive-born snakes from ages 1 to 4 years (Sept. 2011 to Dec.  
38 2014) with semi-parametric Cox proportional hazards using PROC PHREG in SAS. We  
39 conditioned on surviving to age 1 to correspond to the timing of the repeated measures of growth  
40 and IGF1 study, and because we were interested in the effects of morphology and performance  
41 on survival after the duration of high neonatal mortality. We included the categorical factors of  
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3 rearing treatment, ecotype, and sex, as well as all possible two- and three-way interaction terms.  
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6 Because the three-way interaction was significant, we retained this term and all lower-order  
7 terms in our final model. To test for the effect of morphology on survival, we included the  
8 continuous covariates of body size at one year (SVL in mm measured in Sept. 2011), and body  
9 condition at one year (residual of the log-mass on log-SVL regression). To test for the effect of  
10 performance on survival, we included lifetime feeding rate (calculated as the amount of food (in  
11 g) consumed before death divided by the number of days alive), and lifetime growth rate  
12 (calculated as the specific growth rate, a measure of growth scaled to body size:  $SGR = 100 \times$   
13  $[\ln(SVL_2) - \ln(SVL_1)]/days$ ; (Killen 2014; Reid and others 2011). No direct test of IGF1 titer on  
14 survival was performed because the IGF1 titers are plastic and can vary greatly over the lifetime;  
15 instead our analyses incorporates growth, for which we separately tested for an effect of IGF1  
16 concentration (below). Additionally, we included population nested within ecotype as a fixed  
17 effect and litter as a random effect to account for potential covariance among siblings.  
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#### Repeated measures of IGF1 and growth

To test for the main effects of ecotype and temperature on growth rates during the short term  
IGF-1 sampling intervals, and then to test for the effects of ecotype, temperature, and growth on  
IGF-1 levels, we used repeated measures mixed linear models with litter (nested within  
population) as a random effect.

#### Repeated Growth

We modeled short-term growth (change in SVL) during the three intervals of IGF-1 sampling  
with the main fixed effects of Sex; Interval (3 intervals: 10-14 months, 14-16 months, and 16-18

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3 months of age); Ecotype (L-fast and M-slow); Rearing Temperature Treatment (warm, cool); and  
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5 Population nested within Ecotype to reflect heterogeneity among similar ecotype populations.  
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8 We also included the time-varying covariates of size at the start of each interval and food  
9 consumed in the preceding interval. Food consumption was not significant, likely due to  
10 confounding with body size and rearing treatment over this short time-course, and was therefore  
11 removed from the model. For this subsample of the full cohort, offspring from two neighboring  
12 M-slow populations were combined because only one litter was included from one M-slow  
13 population. For the main effects of Time, Treatment and Ecotype, all interactions were included  
14 in the model. Our final model for growth (repeated increases in body length) was:  
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$$\begin{aligned} 27 \text{Change in SVL} = & \mu + \text{Interval start-size} + \text{Sex} + \text{Interval} + \text{Ecotype} + \text{Rearing Treatment} + \\ 28 & \text{Ecotype} \times \text{Interval} + \text{Rearing Treatment} \times \text{Interval} + \text{Ecotype} \times \text{Rearing Treatment} + \\ 29 & \text{Ecotype} \times \text{Rearing Treatment} \times \text{Interval} + \text{Population}(\text{Ecotype}) + \text{Litter}(\text{Population}) + \varepsilon. \\ 30 \\ 31 \\ 32 \\ 33 \\ 34 \\ 35 \\ 36 \end{aligned}$$

### 37 Repeated Plasma IGF1 Concentration

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39 For the analysis of repeated measures of plasma IGF-1 concentrations, all IGF-1 values (ng/mL)  
40 were  $\log_{10}$  transformed to meet normality assumptions. Because IGF-1 is a main mediator of  
41 growth hormone, we considered three different proxies for growth in our analyses: body size  
42 (mm), growth in the interval preceding the IGF-1 measure (“prior interval growth”), and growth  
43 in the interval following the IGF-1 measure (“subsequent interval growth”). This latter variable  
44 of subsequent interval growth had far fewer observations available for the last IGF-1 measure  
45 because most animals were placed immediately into hibernation after the last blood draw. Our  
46 models using each of these explanatory growth variables were in general agreement with each  
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3 other (data not shown), but because we were interested in the effect of growth *per se* (and not  
4 body size) on IGF1 we included growth in the preceding interval as the covariate in our final  
5 model. “Prior Interval Growth” was calculated by taking the change in SVL divided by the  
6 duration of the interval. Because interactions with sex were never significant, we considered only  
7 the two and three way interactions of Time, Treatment, and Ecotype. Our initial model included  
8 food consumption as a covariate over the same period as growth, but as it was not significant, we  
9 removed it from the model:  
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$$\begin{aligned} \log_{10} \text{IGF1} = & \mu + \text{Prior Interval Growth} + \text{Sex} + \text{Time} + \text{Ecotype} + \text{Rearing Treatment} + \\ & \text{Ecotype} \times \text{Time} + \text{Rearing Treatment} \times \text{Time} + \text{Ecotype} \times \text{Rearing Treatment} + \text{Ecotype} \times \text{Rearing} \\ & \text{Treatment} \times \text{Time} + \text{Population}(\text{Ecotype}) + \text{Litter}(\text{Population}) + \varepsilon. \end{aligned}$$

## 35 Results

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### 37 Wild IGF1 plasma concentration.

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40 IGF-1 concentrations in field-caught snakes were dependent on the interaction of SVL, ecotype,  
41 and life-history stage such that larger adult L-fast snakes had higher concentrations of plasma  
42 IGF-1, whereas size did not affect IGF-1 concentrations in juvenile L-fast snakes or M-slow  
43 snakes (Table 2, Fig. 1). Additionally, plasma IGF-1 concentrations were heterogeneous across  
44 years.  
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### 1 2 3 Cohort morphology, growth, feeding, and survival 4 5

6 Cohort morphology, growth, feeding  
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9 For the repeated measures of body size from birth to age 4 yr, all two-way interactions of  
10 ecotype, rearing treatment, and age were significant in determining absolute body size across  
11 lifetime (Table 3, Fig. 2). The main effect of ecotype was significant as well, with L-fast snakes  
12 maintaining larger body size across lifetime than M-slow snakes. Additionally, sex was a  
13 significant factor, with males slightly larger in body size than females after correcting for age.  
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16 For the single measure of growth over the first 4 years, after accounting for the number of  
17 growth days and initial size, the effects of ecotype, rearing treatment, and sex were significant in  
18 determining lifetime growth. L-fast snakes grew more than M-slow snakes while snakes in the  
19 warm rearing treatment grew more than snakes in the cool rearing treatment regardless of  
20 ecotype. Additionally, female snakes grew more than male snakes, despite males maintaining  
21 slightly greater absolute size (Table 4, Fig. 3A). For amount of food consumed (g) from birth  
22 through 4 years, after accounting for the number of feeding days and initial size, the main effects  
23 of both ecotype and rearing treatment, as well as their interaction, were significant factors in  
24 determining feeding rate (Table 5, Fig. 3B). L-warm snakes ate significantly more food than  
25 snakes in any other ecotype/treatment group combination. Additionally, female snakes ate more  
26 than male snakes.  
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### 45 46 Cohort survival 47 48

49 Survivorship to age 4 was dependent on the interaction of ecotype, rearing treatment, and sex  
50 (Table 6, Figure 4). Generally, L-fast snakes lived longer than M-slow snakes while snakes in the  
51 warm temperature treatment outlived snakes in the cool treatment. The effect of sex was  
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3 dependent on ecotype, with females experiencing higher survivorship in the L-fast ecotype and  
4 males in the M-slow. Additionally, snakes larger at one year of age and snakes with slower  
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6 growth rates lived longer than small snakes and fast-growers.  
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12 **Repeated growth and IGF1**  
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15 Repeated measures of growth and IGF1 plasma concentrations were undertaken over three  
16 intervals spanning 6 May, 2011 – 12 Jan. 2012, with blood draws at each interval end date.  
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18 Snakes with larger body length at the start of each interval grew faster than smaller snakes during  
19 the interval (Table 7). As well, females grew faster than males. L-fast animals grew faster than  
20 M-slow animals in the first interval, but not in the second or third intervals (Table 7, Fig. 5). All  
21 animals grew faster in the warm treatment during the first and second intervals, and in the cool  
22 treatment during third interval (Fig. 5). These short-term interval results contrast with overall  
23 growth over the entire experiment in which animals in the warm treatment both consumed more  
24 food and grew more than animals in the cool treatment (see Figs. 3 and 4) for both ecotypes.  
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27 In the analysis of IGF-1 concentration, growth in the interval preceding IGF-1  
28 concentrations negatively affected IGF-1 concentration (Table 8, Figure 6); higher rates of  
29 growth in the interval preceding IGF1 measurements corresponded to lower levels of circulating  
30 IGF-1. This negative correlation between previous growth and current IGF1 was consistent  
31 across all three intervals, particularly intervals 1 and 3. Consistent with this negative association  
32 between IGF1 plasma concentration and growth, males grew slower than females across these  
33 three sampling intervals and males had overall higher levels of plasma IGF1. Other significant  
34 effects in the model of IGF1 variation included a marginally significant 3-way interaction among  
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3 ecotype, treatment, and time (Fig. 6). Across sampling periods, snakes in the warm treatment  
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5 had higher IGF-1 levels than those in the cool treatment (Table 8).  
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## 11 Discussion

  
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13 Life-history theory postulates trade-offs among suites of life-history traits – such as along a  
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15 “pace-of-life” continuum – where species are arrayed along an axis from “slow” to “fast” living  
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17 (Promislow and Harvey 1990; Wikelski and others 2003). At one end are species with slow  
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19 growth and delayed sexual maturation, but with extended lifespan and iteroparity. On the “fast”  
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21 end of the continuum, species exhibit fast growth, rapid reproduction, semelparity or high  
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23 reproductive effort in relatively fewer bouts and, a short lifespan e.g., (Lemaitre and others  
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25 2015). Extensive work has documented the reality of this continuum and the presence of  
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27 underlying trade-offs among growth, reproduction, maintenance, and survival (e.g., (Ghalambor  
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29 and Martin 2001; Lawson and others 2012; Lee and others 2012; Smith and others 1989);  
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31 reviewed in (Nylin and Gotthard 1998; Schluter and others 1991). However, less research has  
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33 focused on the genetic and evolutionary mechanisms that underlie these trade-offs (e.g., (Hendry  
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35 and others 2004; Johnston and others 2013; Nussey 2009; Roff 2000) and even less research has  
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37 sought to understand the physiological processes by which these trade-offs occur (Cohen and  
38  
39 others 2012). Few studies have looked at whole organism physiology, focusing on how  
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41 allocation of resources is differentially controlled (but see (Cox and others 2010; Gangloff and  
42  
43 others 2015). Many of these animal studies of the mechanisms underlying a species’ life-history  
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45 placement on this continuum have been conducted in birds and mammals (Gaillard and others  
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47 1989), but work in snakes and lizards has also found that they exhibit life history characteristics  
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49 congruent with a slow-to-fast pace of life continuum (Shine and Charnov 1992; Sparkman and  
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3 others 2007). This pattern is frequently documented across species; however, some studies have  
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5 documented the existence of a slow-fast continuum even within a single species among  
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7 populations that have experienced different selective pressures over generations (e.g. *Anolis*  
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9 *heterodermis*, (Moreno-Arias and Urbina-Cardona 2013); *Thamnophis elegans*, (this system);  
10  
11 *Sceloporus grammicus*, (Pérez-Mendoza and Zúñiga-Vega 2014). It is here, where intra-specific  
12  
13 variation in life-history strategies exist, that it is most fruitful to understand these polymorphisms  
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15 in the ecomorphology framework. Specifically, when morphological and performance variation  
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17 give rise to fitness differences among individuals, these fitness differences provide natural  
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19 selection the basis to shape variation in population vital rates, including survivorship and  
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21 reproduction.

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24 On the morphological axis, we found that variation in body size can be accounted for by  
25  
26 fixed differences between the ecotypes and interactions with thermal rearing treatment and age.  
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28 Generally, warmer older snakes are larger than cooler younger snakes, particularly in the fast-  
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30 pace-of-life ecotype. In turn, variation in body size affects both short-term growth – a  
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32 performance measure – (with larger snakes growing more than smaller snakes) as well as  
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34 survival through the juvenile stage (1 to 4 yr) – a fitness measure – (with larger snakes  
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36 experiencing higher juvenile survival). Moreover, growth itself was predictive of survival with  
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38 slow growth (after age 1yr) predicting higher juvenile survival. In addition to growth rate, we  
39  
40 considered two other performance measures: feeding and circulating IGF1. Variation among  
41  
42 individuals in feeding was not related to morphological variation directly. Generally, feeding  
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44 was highly impacted by thermal rearing treatment, with snakes in the warmer treatment eating  
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46 more, resulting in these animals growing more and obtaining larger body sizes. Such  
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48 temperature-based plasticity of feeding behavior was further impacted by ecotype, with warm L-  
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3 fast snakes eating disproportionately more than M-slow warm animals. In the short-term  
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5 repeated measures growth and IGF1 experiment, feeding (grams food consumed) and growth  
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7 were essentially interchangeable – food consumption and rearing treatment were confounded;  
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9 animals in the warm treatment ate more and therefore grew more. Our final performance  
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11 measure, IGF1 plasma concentration (ng/mL), was considered over two time scales. First, we  
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13 analyzed 5 years of field-collected blood plasma. Here we found a significant interaction among  
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15 body size, ecotype, and life state (juvenile versus adult; Figure 1). These results show that for  
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17 adult snakes, the manner in which body size associates with IGF1 titers is ecotype-dependent: L-  
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19 fast adults have increasing IGF1 with increasing body size; M-slow adults show no dependence  
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21 of circulating IGF1 on body size. Our previous work has shown that whereas M-slow adults  
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23 essentially stop growing once they reach adulthood, L-fast adults continue growing over their  
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25 adult lifespans with ever increasing size-associated fecundity (Bronikowski and Arnold 1999;  
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27 Sparkman and others 2007).  
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34 Without additional data on individual growth rates, these field data cannot distinguish  
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36 between competing functions of IGF1 among growth, reproduction, and survival in juveniles and  
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38 adults. Thus we undertook here a short-term common garden growth and IGF1 study in which  
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40 we document, among other effects, a short term negative association with IGF1 and growth rates  
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42 (see also (Reding and others 2016). This is in contrast (discussed in more detail below) to studies  
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44 in other systems (e.g., (Lodjak and others 2017).  
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## 50 51 **IGF1 as a mediator of ecomorphology to life-history transition**

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53 We originally envisioned IGF1 as a measure of performance that may impact survival and which  
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55 is itself potentially influenced by body size or body condition. Our results suggest it may be  
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more appropriate think of IGF1 as a mediator of the morphology-to-performance-to-fitness-to-life histories causative continuum. We found that variation in IGF1 is not explained by food consumption per se, and negatively relates to growth preceding IGF1 measurement. Others have found that IGF1 responds to food and/or nutrition availability (Beckman 2011; Duncan and others 2015; Hetz and others 2015; Pierce and others 2005). And indeed we have hypothesized elsewhere that lower levels of IGF1 in M-slow snakes in the wild may be related to their unpredictable food availability (Robert and Bronikowski 2010). For example, in *Sceloporus undulatus* both hepatic IGF1 mRNA levels and circulating levels of IGF1 were reduced in response to food restriction but returned to normal levels at the cessation of food restriction (Duncan and others., 2015).

From work primarily conducted on mammals and fish, IGF1 has been shown to play an integral role in coordinating growth with internal and external cues (e.g. (Dantzer and Swanson 2012; Duan 1998; Fox and others 2006). However, as the insulin/insulin-like signaling (IIS) system is highly conserved across vertebrate taxa, it likely is involved in integrating environmental cues with growth responses in additional vertebrate species as well (e.g., Sparkman and others, 2010, Duncan and others 2015, Dantzer and Swanson 2012, McGaugh and others 2015). Most investigations of IGF1 as a mediator of life-history trade-offs have focused on mammals (e.g., above citations and see (Lewin and others 2017)). However, unlike in mammals, in reptiles environmental temperature per se can affect IGF1 levels (Avila-Mendoza and others 2016) and has a much greater impact on growth because of their ectothermic physiology. Further, most studies to date that have examined the relationship between growth and IGF1 have been correlative (but see (Duncan and others 2015; Ladjak and others 2017; Sparkman and others 2010)).

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3 Food consumption is positively correlated with growth in garter snakes – both in the  
4 present study and in the wild. However, in the short-term, prior food consumption was not  
5 correlated with IGF1 levels beyond its positive association with warm rearing treatments. Yet,  
6 prior growth was negatively correlated with titers of IGF1. These results suggest that the effects  
7 of food on growth are not directly mediated through IGF1, similar to findings in the brown house  
8 snake (Sparkman et al. 2010) and Japanese quail (Ronning et al., 2009), but contrary to the  
9 findings in other species (Pierce et al., 2001, Duncan et al. 2015). It is possible that IGF1 levels  
10 would only respond to extreme changes in food availability; since our experimental snakes were  
11 offered food *ad lib*, there was no variation in available food to affect IGF1 levels. Another  
12 possible explanation could be that food affects transcription or translation of the IGF1 gene, but  
13 we do not see those effects manifested into circulating IGF1 protein.  
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16 We found a negative correlation between prior growth and titers of circulating IGF1. This  
17 result is opposite to the pattern found in brown house snakes (Sparkman and others 2010), great  
18 tits (Lodjak and others, 2014), *Sceloporus* lizards (Duncan and others 2015), pied flycatchers  
19 (Lodjak and others 2017), and spotted hyenas (Lewin and others, 2017, but note this study  
20 examined mass not growth), but similar to that found in meadow *T. elegans* in the field  
21 (Sparkman and others 2009, but this study was looking at size not growth). Because it is unclear  
22 if IGF1 levels are indicative of previous growth, we had also looked for a correlation between  
23 IGF1 levels and growth after the measurement time point, but no relationship existed. More  
24 frequent measurements of size and IGF1 could have revealed a different relationship than what  
25 we found. For example, in salmon, the strongest relationship between IGF1 levels and growth  
26 occurred within a month prior to the hormone sampling (Beckman and others 2004).  
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3 One key component of the insulin/insulin-like signaling system axis this study did not  
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5 examine was localized expression of IGF1. As we only looked at circulating levels of IGF1, we  
6  
7 do not know how temperature and food may have affected localized production of the hormone.  
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10 A companion study by Reding et al. (2016) found that mRNA of IGF1 in liver and muscle was  
11  
12 higher in cool animals compared to those same tissues in warm animals. These results support  
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14 our findings of higher levels of circulating IGF1 in cool animals at our final time point, although  
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16 Reading and others (2016) found that circulating IGF1 levels were higher in warm animals. The  
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18 importance of paracrine and autocrine production vs. endocrine release of IGF1 is equivocal  
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20 (e.g., (Chauvigne and others 2003; Eppler and others 2007).  
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### Growth variation as a mediator of survival

As has been seen in snakes and other ectothermic vertebrates (Folkvord and Ottera, 1993; Gangloff et al. 2015; Sinervo and Adolph, 1989), initial size affects how much an individual grows. However, when considering growth over 4 years, the initial importance of birth size (or an interval's start size) disappears as animals reach sexual maturation. In the present study, this was true in both ecotypes and in both temperature treatments. Food consumption also positively predicted growth rate with more food consumed in the warm treatment and as animals increased in size (see also (Lodjak and others 2014; Sparkman and others 2010). Previous work in this system, including a different subset of individuals from this cohort, demonstrates that growth efficiency is higher in larger animals and animals in the warm rearing treatment (Gangloff and others 2015). Whole organism resting metabolic rate – an additional performance measure – may shed light on long-term versus short-term trends in our study. In both ecotypes, snakes with higher mass-independent resting metabolic rates had lower growth efficiency (a measure of

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3 conversion of ingested food into body substance(Gangloff and others 2015). This trend of higher  
4 resting metabolism negatively associating with growth occurs in other reptiles as well (e.g.,  
5 (Steyermark 2002), and is postulated to occur because animals with higher resting metabolic  
6 rates devote more energy to maintenance and have less energy available for growth.  
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10 We were not able to take into consideration the maternal condition during gestation,  
11 which can affect initial growth and size at birth. The amount of protein, for example, that the  
12 mother consumed could have affected what she deposited into the yolk. The nutritional content  
13 of the yolk could have contributed to neonatal growth. Any differences in growth between  
14 ecotypes could be influenced by maternal diet (Metcalfe and Monaghan 2001; Micke and others  
15 2011). Robbins et al. 2012, Sikes 1996). Notwithstanding, animals that grew fastest during their  
16 2<sup>nd</sup> – 4<sup>th</sup> years had lower probability of survival, despite that animals that started off larger had a  
17 higher probability of surviving their first 4 years. Although not a topic of this study, if faster  
18 growers were engaged in “catch-up” growth (for example, due to cooler rearing temperatures),  
19 then these results would suggest a downstream cost in terms of survival (see also (Marcil-Ferland  
20 and others 2013; Metcalfe and Monaghan 2003; Orizaola and others 2014) reviewed in  
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22 (Lindstrom 1999).  
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#### 43 Selection gradients: The direct link between ecomorphology and life-history

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45 One of the elements of the morphology/performance/fitness paradigm was the calculation of  
46 selection gradients and thus the ability to infer whether and how selection was operating on  
47 specific morphological/performance traits (Arnold 1983). Accomplished through regression  
48 techniques with a proxy for fitness and multiple phenotypes ((Arnold and Wade 1984a; 1984b;  
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60 Lande and Arnold 1983), the partial regression coefficients can be interpreted as the selection

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3 differentials. On the life-history side, population demography sensitivity analyses applied to a  
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5 Leslie or Lefkovitch matrix of vital rates can address how variation in vital rates affect  
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7 population growth rates; how sensitive the fitness of a phenotype is to perturbations in vital rates  
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9 (reviewed in (Caswell 2001)). At the same time, Russ Lande ((Lande 1982a; Lande 1982b) made  
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11 explicit that these estimated sensitivities are equivalent to selection differentials. Thus, selection  
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13 differentials should be a direct link between ecomorphology and life-history theory.  
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17 Selection gradients vary between these fast and slow ecotypes for survival and growth in  
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19 the pre-adult stages (Miller and others 2011). Specifically, neonatal and juvenile survival are two  
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21 vital rates that differ between M-slow and L-fast populations in their estimates of sensitivities  
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23 (selection gradients) and elasticities (contribution to population growth rates). While much is  
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25 known about the causes of neonate mortality in this system, relatively little is known about the  
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27 juvenile stage. Yet, in M-slow populations, survival increases in the juvenile stage to levels  
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29 equivalent to adults (ca. 0.80 probability of survival annually), whereas in L-fast populations,  
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31 juvenile survival is much lower (ca. 0.50 probability of annual survival). Accordingly, the  
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33 selection gradients for juvenile survival average 0.45 in M-slow populations and 0.35 in L-fast  
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35 populations. Therefore we sought to better understand the sources of variation in juvenile  
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37 survival. Figure 6 presents the contributions to juvenile survival through morphology and  
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39 performance, but also through environmental variation and ecotype variation which we use as a  
40  
41 proxy for genetic background. Strong positive effects of body size and negative effects of growth  
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43 on juvenile survival may buffer snakes from the negative effects of low-resource years and high  
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45 temperature years. The relationships among feeding behavior, hormone levels, and growth are  
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47 more complex, with larger snakes eating more and growing more, yet having lower IGF1 plasma  
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49 concentrations and negative association of growth with survival. We have argued elsewhere that  
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3 growing maximally rather than optimally is likely very costly in this system where food  
4 availability is unpredictable. In M-slow habitats, any given year is only 50% likely to have  
5 adequate snowfall for anuran breeding in the spring (the primary prey of meadow garter snakes).  
6 In L-fast habitats, although fish and leeches are the primary prey, fluctuations in water level and  
7 shoreline vegetation provide an annually changing gauntlet that snakes must traverse to get to  
8 their food source.  
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17 The results of the present study, combined with past work on the garter snakes in the  
18 Eagle Lake system, present a complex relationship between traits traditionally categorized as  
19 morphology, performance, and life history (Fig. 6). This web of mutually dependent traits belies  
20 simple categorization in a one-way causal path among different aspects of the phenotype. Rather,  
21 we see that traits feedback on each other through multiple pathways in which genetic background  
22 and environmental conditions interact to create distinct ecotypic syndromes at the population  
23 level. Importantly, our experimental results demonstrate that the hormone IGF-1 plays a central  
24 mediating role between body size and growth during the juvenile stage. Field observations  
25 complement this finding, showing that in the L-fast ecotype – where snakes grow faster and  
26 reach larger asymptotic sizes – plasma IGF-1 concentrations are correlated with size.  
27  
28 Nonetheless, this relationship is not entirely straightforward and may shift across ontogeny,  
29 especially at life-history transitions when energetic allocation to growth begins to trade-off with  
30 allocation to reproduction. What is startling about the interdependent relationships among traits  
31 within individuals is that, despite this complexity, we observe rather distinct life-history ecotypes  
32 in these natural populations. Traits of individuals within each ecotype – including metabolic and  
33 energetic functions, immune capacity, hormone titres, and life-history traits – can be categorized  
34 on opposite ends of the fast-slow pace-of-life continuum. Thus, ecotype differentiation is an  
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3 emergent property of this biological system, relying not on a single causal pathway but multiple  
4 interwoven systems that feedback on each other, resulting in suites of correlated phenotypic traits  
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6 adapted to local environments.  
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28 **Literature Cited**  
29  
30

31 Annunziata M, Granata R, Ghigo E. 2011. The IGF system. *Acta Diabetologica* 48:1-9.  
32  
33 Arnold SJ. 1983. Morphology, Performance and Fitness. *American Zoologist* 23(2):347-361.  
34  
35 Arnold SJ. 1988. Behavior, Energy and Fitness. *American Zoologist* 28(3):815-827.  
36  
37 Arnold SJ. 1992. BEHAVIORAL VARIATION IN NATURAL-POPULATIONS .6. PREY  
38  
39 RESPONSES BY 2 SPECIES OF GARTER SNAKES IN 3 REGIONS OF  
40  
41 SYMPATRY. *Animal Behaviour* 44(4):705-719.  
42  
43  
44 Arnold SJ, Bennett AF. 1984. Behavioral Variation in Natural-Populations .3. Antipredator  
45  
46 Displays in the Garter Snake *Thamnophis-Radix*. *Animal Behaviour* 32(Nov):1108-1118.  
47  
48 Arnold SJ, Wade MJ. 1984a. On the Measurement of Natural and Sexual Selection -  
49  
50 Applications. *Evolution* 38(4):720-734.  
51  
52  
53 Arnold SJ, Wade MJ. 1984b. On the Measurement of Natural and Sexual Selection - Theory.  
54  
55 Evolution 38(4):709-719.  
56  
57  
58  
59  
60

1  
2  
3 Avila-Mendoza J, Mora J, Carranza M, Luna M, Aramburo C. 2016. Growth hormone reverses  
4  
5 excitotoxic damage induced by kainic acid in the green iguana neuroretina. General and  
6  
7 Comparative Endocrinology 234:57-67.  
8  
9

10  
11 Ayres FA, Arnold SJ. 1983. Behavioral Variation in Natural-Populations .4. Mendelian Models  
12  
13 and Heritability of a Feeding Response in the Garter Snake, *Thamnophis-Elegans*.  
14  
15 Heredity 51(Aug):405-413.  
16

17  
18 Beckman BR. 2011. Perspectives on concordant and discordant relations between insulin-like  
19  
20 growth factor 1 (IGF1) and growth in fishes. General and Comparative Endocrinology  
21  
22 170:233-252.  
23

24  
25 Beckman BR, Shimizu M, Gadberry BA, Cooper KA. 2004. Response of the somatotropic axis  
26  
27 of juvenile coho salmon to alterations in plane of nutrition with an analysis of the  
28  
29 relationships among growth rate and circulating IGF-I and 41 kDa IGFBP. General and  
30  
31 Comparative Endocrinology 135(3):334-344.  
32  
33

34  
35 Bronikowski AM. 2000. Experimental evidence for the adaptive evolution of growth rate in the  
36  
37 garter snake *Thamnophis elegans*. Evolution 54(5):1760-1767.  
38

39  
40 Bronikowski AM, Arnold SJ. 1999. The evolutionary ecology of life history variation in the  
41  
42 garter snake *Thamnophis elegans*. Ecology 80(7):2314-2325.  
43  
44

45 Caswell H. 2001. Matrix Population Models. Sunderland MA: Sinauer.  
46  
47

48 Chauvigne F, Gabillard JC, Weil C, Rescan PY. 2003. Effect of refeeding on IGFI, IGFII, IGF  
49  
50 receptors, FGF2, FGF6, and myostatin mRNA expression in rainbow trout myotomal  
51  
52 muscle. General and Comparative Endocrinology 132(2):209-215.  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Cohen AA, Martin LB, Wingfield JC, McWilliams SR, Dunne JA. 2012. Physiological  
4 regulatory networks: Ecological roles and evolutionary constraints. *Trends in Ecology and Evolution*  
5 and Evolution 27:428-435.  
6  
7 Cox RM, Parker EU, Cheney DM, Liebl AL, Martin LB, Calsbeek R. 2010. Experimental  
8 evidence for physiological costs underlying the trade-off between reproduction and  
9 survival. *Functional Ecology* 24:1262-1269.  
10  
11 Dantzer B, Swanson EM. 2012. Mediation of vertebrate life histories via insulin-like growth  
12 factor-1. *Biological Reviews* 87:414-429.  
13  
14 Denley A, Cosgrove LJ, Booker GW, Wallace JC, Forbes BE. 2005. Molecular interactions of  
15 the IGF system. *Cytokine & Growth Factor Reviews* 16(4-5):421-439.  
16  
17 Duan C. 1998. Nutritional and developmental regulation of insulin-like growth factors in fish.  
18 The Journal of Nutrition 128:306S-314S.  
19  
20 Dudley SA. 1996. Differing selection on plant physiological traits in response to environmental  
21 water availability: A test of adaptive hypotheses. *Evolution* 50(1):92-102.  
22  
23 Duncan CA, Jetzt AE, Cohick WS, John-Alder HB. 2015. Nutritional modulation of IGF-1 in  
24 relation to growth and body condition in *Sceloporus* lizards. *General and Comparative*  
25 *Endocrinology*:1-9.  
26  
27 Eppler E, Caelers A, Shved N, Hwang G, Rahman AM, Maclean N, Zapf J, Reinecke M. 2007.  
28  
29 Insulin-like growth factor I (IGF-I) in a growth-enhanced transgenic (GH-  
30 overexpressing) bony fish, the tilapia (*Oreochromis niloticus*): indication for a higher  
31 impact of autocrine/paracrine than of endocrine IGF-I. *Transgenic Research* 16(4):479-  
32 489.  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Fox BK, Riley LG, Hiraono T, Grau EG. 2006. Effects of fasting on growth hormone, growth  
4  
5 hormone receptor, and insulin-like growth factor-1 axis in seawater-acclimated tilapia,  
6  
7 Oreochromis mossambicus. *General and Comparative Endocrinology* 148:340-347.  
8  
9  
10 Gaillard J-M, Pontier D, Allainé D, Lebreton JD, Trouvilliez J, Clobert J. 1989. An analysis of  
11  
12 demographic tactics in birds and mammals. *Oikos* 56:59-76.  
13  
14  
15 Gangloff EJ, Vleck D, Bronikowski AM. 2015. Developmental and immediate thermal  
16  
17 environments shape energetic trade-offs, growth efficiency, and metabolic rate in  
18  
19 divergent life-history ecotypes of the garter snake *Thamnophis elegans*. *Physiological*  
20  
21 and *Biochemical Zoology* 88(5):550-563.  
22  
23  
24 Ghalambor CK, Martin TE. 2001. Fecundity-Survival Trade-Offs and Parental Risk-Taking in  
25  
26 Birds. *Science* 292:494-497.  
27  
28  
29 Gould SJ, Lewontin RC. 1979. Spandrels of San-Marco and the Panglossian Paradigm - a  
30  
31 Critique of the Adaptationist Program. *Proceedings of the Royal Society Series B-*  
32  
33 *Biological Sciences* 205(1161):581-598.  
34  
35  
36 Hendry AP, Morbey YE, Berg OK, Wenburg JK. 2004. Adaptive variation in senescence:  
37  
38 reproductive lifespan in a wild salmon population. *Proceedings. Biological sciences / The*  
39  
40 Royal Society 271:259-266.  
41  
42  
43 Hetz JA, Menzies BR, Shaw G, Rao A, Clarke LJ, Renfree MB. 2015. Growth axis maturation is  
44  
45 linked to nutrition, growth and developmental rate. *Molecular and Cellular*  
46  
47 *Endocrinology* 411(C):38-48.  
48  
49  
50 Johnston SE, Gratten J, Berenos C, Pilkington JG, Clutton-Brock TH, Pemberton JM, Slate J.  
51  
52  
53 2013. Life history trade-offs at a single locus maintain sexually selected genetic variation.  
54  
55 *Nature* 502:93-95.  
56  
57  
58  
59  
60

1  
2  
3 Kelley KC, Arnold SJ, Glatstone J. 1997. The effects of substrate and vertebral number on  
4 locomotion in the garter snake *Thamnophis elegans*. *Functional Ecology* 11(2):189-198.  
5  
6 Kenward MG, Roger JH. 1997. Small sample inference for fixed effects from restricted  
7 maximum likelihood. *Biometrics* 53(3):983-997.  
8  
9 Ketterson ED, Nolan V. 1999. The University of Chicago Adaptation, Exaptation, and  
10 Constraint: A Hormonal Perspective. *Source: The American Naturalist* 154:4-25.  
11  
12 Killen SS. 2014. Growth trajectory influences temperature preference in fish through an effect on  
13 metabolic rate. *Journal of Animal Ecology* 83(6):1513-1522.  
14  
15 Lailvaux SP, Husak JF. 2014. The life history of whole organism performance. *The Quarterly*  
16  
17 Review of Biology

18 89:285-318.  
19  
20 Lande R. 1982a. Elements of a Quantitative Genetic Model of Life History Evolution. In: Dingle  
21 H, Hegmann JP, editors. *Evolution and Genetics of Life Histories*. New York, NY:  
22 Springer US. p. 21-29.  
23  
24 Lande R. 1982b. A Quantitative Genetic Theory of Life-History Evolution. *Ecology* 63(3):607-  
25 615.  
26  
27 Lande R, Arnold SJ. 1983. The Measurement of Selection on Correlated Characters. *Evolution*  
28  
29 37(6):1210-1226.  
30  
31 Lawson DW, Alvergne A, Gibson Ma. 2012. The life-history trade-off between fertility and  
32 child survival. *Proceedings. Biological sciences / The Royal Society* 279:4755-64.  
33  
34 Lee W-S, Monaghan P, Metcalfe NB. 2012. Experimental demonstration of the growth rate -  
35 lifespan trade-off. *Proceedings of The Royal Society B: Biological Sciences*  
36  
37 280:20122370.  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
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57  
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60

1  
2  
3 Lemaitre JF, Berger V, Bonenfant C, Douhard M, Gamelon M, Plard F, Gaillard JM. 2015.  
4  
5 Early-late life trade-offs and the evolution of ageing in the wild. Proceedings of the Royal  
6  
7 Society B-Biological Sciences 282(1806).  
8  
9  
10 Lewin N, Swanson EM, Williams BL, Holekamp KE. 2017. Juvenile concentrations of IGF-1  
11  
12 predict life-history trade-offs in a wild mammal. Functional Ecology 31(4):894-902.  
13  
14 Lindstrom J. 1999. Early development and fitness in birds and mammals. Trends in Ecology &  
15  
16 Evolution 14(9):343-348.  
17  
18  
19 Lodjak J, Magi M, Sild E, Mand R. 2017. Causal link between insulin-like growth factor 1 and  
20  
21 growth in nestlings of a wild passerine bird. Functional Ecology 31(1):184-191.  
22  
23  
24 Lodjak J, Magi M, Tilgar V. 2014. Insulin-like growth factor 1 and growth rate in nestlings of a  
25  
26 wild passerine bird. Functional Ecology 28(1):159-166.  
27  
28  
29 Losos JB. 1990. The Evolution of Form and Function - Morphology and Locomotor  
30  
31 Performance in West-Indian Anolis Lizards. Evolution 44(5):1189-1203.  
32  
33  
34 Manier MK, Seyler CM, Arnold SJ. 2007. Adaptive divergence within and between ecotypes of  
35  
36 the terrestrial garter snake, *Thamnophis elegans*, assessed with F-ST-Q(ST) comparisons.  
37  
38 Journal of Evolutionary Biology 20(5):1705-1719.  
39  
40  
41 Marcil-Ferland D, Festa-Bianchet M, Martin AM, Pelletier F. 2013. Despite Catch-Up,  
42  
43 Prolonged Growth Has Detrimental Fitness Consequences in a Long-Lived Vertebrate.  
44  
45 The American Naturalist 182(6):775-785.  
46  
47  
48 McGaugh SE, Bronikowski AM, Kuo CH, Reding DM, Addis EA, Flagel LE, Janzen FJ,  
49  
50 Schwartz TS. 2015. Rapid molecular evolution across amniotes of the IIS/TOR network.  
51  
52 Proceedings of the National Academy of Sciences of the United States of America  
53  
54 112(22):7055-7060.  
55  
56  
57  
58  
59  
60

1  
2  
3 Metcalfe NB, Monaghan P. 2001. Compensation for a bad start: grow now, pay later? Trends in  
4 Ecology & Evolution 16(5):254-260.  
5  
6 Metcalfe NB, Monaghan P. 2003. Growth versus lifespan: perspectives from evolutionary  
7 ecology. Experimental Gerontology 38(9):935-940.  
8  
9 Micke GC, Sullivan TM, McMillen IC, Gentili S, Perry VEA. 2011. Protein intake during  
10 gestation affects postnatal bovine skeletal muscle growth and relative expression of IGF1,  
11 IGF1R, IGF2 and IGF2R. Molecular and Cellular Endocrinology 332(1-2):234-241.  
12  
13 Miller DA, Clark WR, Arnold SJ, Bronikowski AM. 2011. Stochastic population dynamics in  
14 populations of western terrestrial garter snakes with divergent life histories. Ecology  
15 92:1658-1671.  
16  
17 Miller DAW, Janzen FJ, Fellers GM, Kleeman PM, Bronikowski A. 2014. Biodemography of  
18 ectothermic tetrapods provides insights into the evolution and plasticity of mortality  
19 trajectories. In: Weinstein M, Lane MA, editors. Sociality, Hierarchy, Health:  
20 Comparative Demography Advances in Biodemography: Cross-species comparisons of  
21 social environments and social behaviors, and their effects on health and longevity.  
22 Washington DC: The National Academies Press. p. 295-314.  
23  
24 Moreno-Arias Ra, Urbina-Cardona JN. 2013. Population Dynamics of the Andean Lizard *Anolis*  
25 *heterodermus*: Fast-slow Demographic Strategies in Fragmented Scrubland Landscapes.  
26 Biotropica 45:253-261.  
27  
28 Nussey DH. 2009. Plasticity in breeding time in wild vertebrates: a quantitative genetic  
29 approach. Integrative and Comparative Biology 49:E124-E124.  
30  
31 Nylin S, Gotthard K. 1998. Plasticity in Life-History Traits. Annual Review of Entomology  
32 43:63-83.  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
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48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Orizaola G, Dahl E, Laurila A. 2014. Compensatory growth strategies are affected by the  
4 strength of environmental time constraints in anuran larvae. *Oecologia* 174(1):131-137.  
5  
6 Palacios MG, Cunnick JE, Bronikowski AM. 2013. Complex Interplay of Body Condition, Life  
7 History, and Prevailing Environment Shapes Immune Defenses of Garter Snakes in the  
8 Wild. *Physiological and Biochemical Zoology* 86(5):547-558.  
9  
10 Palacios MG, Sparkman AM, Bronikowski AM. 2011. Developmental plasticity of immune  
11 defence in two life-history ecotypes of the garter snake, *Thamnophis elegans* - a  
12 common-environment experiment. *Journal of Animal Ecology* 80(2):431-437.  
13  
14 Palacios MG, Sparkman AM, Bronikowski AM. 2012. Corticosterone and pace of life in two  
15 life-history ecotypes of the garter snake *Thamnophis elegans*. *General and Comparative*  
16  
17 *Endocrinology* 175(3):443-448.  
18  
19 Pérez-Mendoza HA, Zúñiga-Vega JJ. 2014. A test of the fast – slow continuum model of life-  
20 history variation in the lizard *Sceloporus grammicus*. *Evolutionary Ecology Research*  
21  
22 16:235-248.  
23  
24 Pierce AL, Shimizu M, Beckman BR, Baker DM, Dickhoff WW. 2005. Time course of the  
25 GH/IGF axis response to fasting and increased ration in chinook salmon (*Oncorhynchus*  
26  
27 *tshawytscha*). *General and Comparative Endocrinology* 140:192-202.  
28  
29 Promislow DEL, Harvey PH. 1990. Living fast and dying young: A comparative analysis of life-  
30 history variation among mammals. *Journal of Zoology* 220:417-437.  
31  
32 Reding DM, Addis EA, Palacios MG, Schwartz TS, Bronikowski AM. 2016. Insulin-like  
33 signaling (IIS) responses to temperature, genetic background, and growth variation in  
34 garter snakes with divergent life histories. *General and Comparative Endocrinology*  
35  
36 233:88-99.  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Reid D, Armstrong JD, Metcalfe NB. 2011. Estimated standard metabolic rate interacts with  
4 territory quality and density to determine the growth rates of juvenile Atlantic salmon.  
5  
6 Functional Ecology 25(6):1360-1367.  
7  
8  
9 Ricklefs RE, Wikelski M. 2002. The physiology/life history nexus. Trends in Ecology and  
10 Evolution 17:462-469.  
11  
12  
13 Robert KA, Bronikowski AM. 2010. Evolution of Senescence in Nature: Physiological Evolution  
14 in Populations of Garter Snake with Divergent Life Histories. American Naturalist  
15 175(2):E47-159.  
16  
17 Roff. 2000. Trade-offs between growth and reproduction: an analysis of the quantitative genetic  
18 evidence. Journal of Evolutionary Biology 13:434-445.  
19  
20 Schluter D. 1995. Adaptive Radiation in Sticklebacks - Trade-Offs in Feeding Performance and  
21 Growth. Ecology 76(1):82-90.  
22  
23 Schluter D, Price TD, Rowe L. 1991. Conflicting Selection Pressures and Life History Trade-  
24 Offs. Proceedings of the Royal Society B: Biological Sciences 246:11-17.  
25  
26 Schwartz TS, Arendsee ZW, Bronikowski AM. 2015. Mitochondrial divergence between slow-  
27 and fast-aging garter snakes. Exp Gerontol 71:135-46.  
28  
29 Schwartz TS, Bronikowski AM. 2016. Evolution and Function of the Insulin and Insulin-like  
30 Signaling Network in Ectothermic Reptiles: Some Answers and More Questions.  
31 Integrative and Comparative Biology 56(2):171-184.  
32  
33 Shine R, Charnov EL. 1992. Patterns of Survival, Growth, and Maturation in Snakes and  
34 Lizards. The American Naturalist 139:1257.  
35  
36 Smith H, Kallander H, Nilsson J-A. 1989. The trade-off between offspring number and quality in  
37 the great tit *Parus major*. The Journal of Animal Ecology 58:383-401.  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

1  
2  
3 Sparkman AM, Arnold SJ, Bronikowski AM. 2007. An empirical test of evolutionary theories  
4 for reproductive senescence and reproductive effort in the garter snake *Thamnophis*  
5 elegans. *Proceedings of the Royal Society B-Biological Sciences* 274(1612):943-950.  
6  
7  
8  
9 Sparkman AM, Bronikowski AM, Billings JG, Von Borstel D, Arnold SJ. 2013. Avian Predation  
10 and the Evolution of Life Histories in the Garter Snake *Thamnophis elegans*. *American*  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
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53  
54  
55  
56  
57  
58  
59  
60  
Ecology

90(3):720-728.

Sparkman AM, Byars D, Ford NB, Bronikowski AM. 2010. The role of insulin-like growth factor-1 (IGF-1) in growth and reproduction in female brown house snakes (*Lampropeltis fuliginosus*). *General and Comparative Endocrinology* 168(3):408-414.

Sparkman AM, Schwartz TS, Madden JA, Boyken SE, Ford NB, Serb JM, Bronikowski AM. 2012. Rates of molecular evolution vary in vertebrates for insulin-like growth factor-1 (IGF-1), a pleiotropic locus that regulates life history traits. *General and Comparative Endocrinology* 178(1):164-173.

Stearns SC. 1992. *The evolution of life histories*. Oxford, UK: Oxford University Press.

Steyermark AC. 2002. A high standard metabolic rate constrains juvenile growth. *Zoology* (Jena, Germany) 105:147-151.

Wickham H. 2009. *ggplot2: elegant graphics of data analysis*. New York: Springer.

Wikelski M, Spinney L, Schelsky W, Scheuerlein A, Gwinner E. 2003. Slow pace of life in tropical sedentary birds: a common-garden experiment on four stonechat populations

1  
2  
3 from different latitudes. *Proceedings. Biological sciences / The Royal Society* 270:2383-  
4  
5 2388.  
6  
7

8 Zera aJ, Bottsford J. 2001. The endocrine-genetic basis of life-history variation: the relationship  
9  
10 between the ecdysteroid titer and morph-specific reproduction in the wing-polymorphic  
11  
12 cricket *Gryllus firmus*. *Evolution; international journal of organic evolution* 55:538-549.  
13  
14

15 Zhu X, Zhang SZ, Zhao S, Zhang R, Zhou YK, Wu XB. 2017. Insulin-like growth factor I (IGF-  
16  
17 I) in Chinese alligator, *Alligator sinensis*: Molecular characterization, tissue distribution  
18  
19 and mRNA expression changes during the active and hibernating periods. *General and*  
20  
21 *Comparative Endocrinology* 242:74-82.  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
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## Figure Legends

**Figure 1.** Scatterplots of the relationship between size (SVL in mm) and plasma IGF-1 concentration ( $\log_{10}$ -transformed ng/mL) for each life-stage by ecotype combination in field-caught *Thamnophis elegans*. Snakes were sampled from 2006-2010 at populations around Eagle Lake, CA, USA. Overall levels of IGF-1 varied across years (see Table 2), so values here are normalized with respect to year (from model of  $\log_{10}$ -transformed IGF-1 concentrations with year as a categorical fixed effect). The relationship between SVL and IGF-1 concentrations is significant for L-fast adults ( $R^2 = 0.14$ ,  $P < 0.001$ ).

**Figure 2.** Least-square means from the model of lifetime size (SVL in mm; see text for model details) for *T. elegans* juveniles raised in captivity, through age 4 years. Both ecotype and rearing treatment influenced lifetime growth. Error bars represent  $\pm$  SE.

**Figure 3.** (A) Least-square means from the model of lifetime growth (SVL in mm; see text for model details) for *T. elegans* juveniles raised in captivity, from birth to age 4 years. Both ecotype and rearing treatment were significant factors in determining lifetime growth. (B) Least-square means from the model of lifetime feeding (amount consumed in g; see text for model details) for *T. elegans* juveniles raised in captivity, from birth to age 4 years. Lwarm snakes ate more than Lcool, Mwarm, or Mcool snakes (all adjusted pairwise comparisons significant with  $p < 0.01$ ). Error bars for both panels represent  $\pm$  SE.

**Figure 4.** Survival curves for captive-born *Thamnophis elegans* by treatment group within each sex. Plots demonstrate the significant influence of ecotype, treatment, and sex on survivorship

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3 based on the semi-parametric Cox Proportional hazard model (see Table 6). Horizontal dotted  
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5 line shows 50% survivorship. Estimates are median residual lifespans (in months)  $\pm$  SE.  
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10 **Figure 5.** (A) Growth (change in SVL, mm) for captive-born *T. elegans* over the three intervals.  
11 Estimates are shown on the last day of an interval; the number of days in each interval varied:  
12 Interval 1 = 178 days; Interval 2 = 48 days; Interval 3 = 72 days. The significant effect of  
13 temperature  $\times$  interval is seen in that during interval 1, warm snakes were growing fastest,  
14 whereas in interval 3, cooler snakes were growing fastest. The significant effect of ecotype  $\times$   
15 time can be in that during interval 1, L snakes were growing fastest, whereas in interval 3, cooler  
16 snakes were growing fastest. (B) Least-square means from the model of plasma IGF-1  
17 concentration ( $\log_{10}$ -transformed ng/mL). IGF-1 levels were assayed from samples collected  
18 from the final day of 3 intervals, with first measurement 267 days after birth. The significant 3-  
19 way interaction of ecotype  $\times$  treatment  $\times$  time can be seen in the deviation of the Mcool values  
20 from the others, Note that Mcool animals grew fastest, and faster than other categories, during  
21 interval 3. Error bars for both panels represent  $\pm$  SE.  
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41 **Figure 6.** The complex interplay of phenotypic traits involved in the morphology–performance–  
42 life-history web. Black arrows represent relationships between aspects of the phenotype, with  
43 solid lines indicating positive relationships and dashed lines indicating negative. Solid gray  
44 arrows represent the direct impact of environmental characteristics on traits. Zig-zag lines show  
45 traits in which genetic canalization has resulted in differences between distinct life-history  
46 ecotypes. Traits and factors shown in bold black type are included in the present study; traits and  
47 factors in gray type have been established in previous studies (Bronikowski & Arnold 1999,  
48 Sparkman and others 2007, Miller and others 2011, Sparkman and others 2013).  
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3 **Table 1.** Sample Sizes and measuring dates for full colony of *T. elegans* neonates raised in captivity.  
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6 Study	7 N neonates	8 Ecotype (litters, neonates)	9 Female- warm	Female-cool	Male-warm	Male-cool
<b>Cohort morphology, growth, feeding, survival study</b>						
<sup>1</sup> Birth	257 (65 released)					
<sup>2</sup> Assigned a treatment	192	M-slow (19, 72) L-fast (21, 120)	18 31	20 29	18 30	16 30
29 Nov. 2010	189					
<sup>3</sup> 1 Nov. 2011	153					
15 Nov. 2012	66					
19 Aug. 2013	56					
10 Dec. 2014	28					
<sup>4</sup> IGF1-growth study	130	M-slow (18, 47) L-fast (21, 83)	10 22	10 20	15 21	12 20

27 <sup>1</sup> Birth occurred 12 Aug – 19 Sept 2010.  
28

29 <sup>2</sup> 65 neonates were released shortly after birth into their natal populations from very large litters.  
30

31 <sup>3</sup> Survival analysis was conditioned on survival to one-year (i.e., survival through the neonate stage) and used these 153 1-year olds.  
32

33 <sup>4</sup> IGF1-growth study initiated 14 Sept 2011.  
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3 **Table 2. Generalized linear model results of  $\log_{10}$ -transformed field IGF-1 plasma  
4 concentration measures collected from male and non-gravid female *T. elegans* from 2006-  
5 2010.**  
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Source of Variation	Estimate	df <sub>1</sub> , df <sub>2</sub>	F	Pr > F	Direction of significant factors
SVL	-0.00059	1, 539	1.40	0.24	
Ecotype	-0.33	1, 539	8.16	<b>0.0044**</b>	L-fast > M-slow
Life stage	-0.25	1, 539	3.78	0.052	
Year	--	4, 539	8.52	< <b>0.0001**</b>	2006 > 2007 > 2009 > 2010 > 2008
SVL × Ecotype	0.00097	1, 539	7.63	<b>0.0059**</b>	See Figure 1
SVL × Life stage	0.00081	1, 539	3.83	0.051	See Figure 1
SVL × Year	--	4, 539	1.21	0.31	
SVL × Ecotype × Life Stage	-0.00035	1, 539	9.66	<b>0.0020**</b>	See Figure 1

32 **Table 3: Mixed linear model analysis of *T. elegans* body size from Birth—Dec. 2014 (SVL in  
33 mm).**

Source of Variation	Estimate	df <sub>n</sub> , df <sub>d</sub>	F	Pr > F	Direction of significant factors
Ecotype	10.35	1, 31.6	14.33	<b>0.0006**</b>	L-fast > M-slow
Rearing treatment	-4.16	1, 137	1.36	0.25	--
Age	0.086	1, 140	1047.5	< <b>0.0001**</b>	Older > Younger
Sex	-4.39	1, 140	9.70	<b>0.0022*</b>	Males > Females
Ecotype × Rearing treatment	5.24	1, 137	3.93	<b>0.050*</b>	See Figure 2
Ecotype × Age	0.04	1, 140	15.70	<b>0.0001**</b>	See Figure 2
Rearing treatment × Age	0.038	1, 140	12.35	<b>0.0006**</b>	See Figure 2
Ecotype × Treatment × Age	-0.023	1, 140	2.50	0.12	--
Population(Ecotype)	--	5, 31.4	1.53	0.21	--

5  
6 **Table 4: Mixed linear model analysis of *T. elegans* growth from Nov. 2010—Dec. 2014**  
7 (change in SVL in mm).

Source of Variation	Estimate	df <sub>n</sub> , df <sub>d</sub>	F	Pr > F	Direction of significant factors
Growth Days	0.13	1, 137	535.63	< 0.0001**	More days > Less days
Initial size	-0.025	1, 84.4	0.01	0.91	--
Ecotype	5.29	1, 37.8	9.98	0.0031**	L-fast > M-slow, See Fig. 3A
Rearing treatment	33.82	1, 126	46.06	< 0.0001**	Warm > Cool, See Fig. 3A
Sex	12.18	1, 138	6.97	0.0092**	Females > Males
Ecotype×Rearing treatment	-6.19	1, 126	0.46	0.50	--
Population(Ecotype)	--	5, 27.1	1.48	0.23	--

30 **Table 5: Mixed linear model analysis of *T. elegans* feeding from Nov. 2010—Dec. 2014**  
31 (weight of frozen/thawed pinky mice, in g).

Source of Variation	Estimate	df <sub>n</sub> , df <sub>d</sub>	F	Pr > F	Direction of significant factors
Feeding Days	0.15	1, 173	1632.79	< 0.0001**	More days > Less days
Initial size	0.18	1, 112	1.29	0.26	--
Ecotype	18.58	1, 41.6	5.87	0.020*	L-fast > M-slow
Rearing treatment	6.64	1, 150	25.15	< 0.0001**	Warm > Cool
Sex	7.75	1, 170	5.65	0.019*	Females > Males
Ecotype×Rearing treatment	17.86	1, 149	8.25	0.0047**	See Fig. 3B
Population(Ecotype)	--	5, 31.1	1.20	0.33	--

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5 **Table 6. Results of semi-parametric Cox proportional hazards model of survivorship from**  
6 **age 1 to age 4 years in captive-born *T. elegans*.**  
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Source	$\chi^2$	Adj. df	p	Direction of significant factors
Size at 1 year	5.23	0.88	<b>0.018*</b>	Larger snakes > Smaller snakes
Body Condition at 1 year	0.34	0.85	0.49	
Lifetime feeding Rate	0.29	0.89	0.54	
Lifetime growth rate	20.23	0.88	< 0.0001**	Slow growers > Fast growers
Sex	2.63	0.84	0.083	
Ecotype	0.63	0.41	0.18	
Rearing treatment	1.28	0.90	0.23	
Sex × Ecotype	11.67	0.88	<b>0.0005**</b>	Female L-fast > Male L-fast Male M-slow > Female M-slow
Sex × Rearing treatment	1.52	0.93	0.20	
Sex × Ecotype × Rearing treatment	3.81	0.93	<b>0.046*</b>	See Figure 4
Population(Ecotype)	3.19	2.38	0.26	

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3 **Table 7. Repeated measures mixed model analysis of *T. elegans* growth over three intervals measured**  
4 **as change-in-SVL (in mm).**

Source of Variation	Estimate	df <sub>n</sub> , df <sub>d</sub>	F	Pr > F	Direction of significant factors
Size at start of interval	-0.071	1, 92.2	10.95	<b>0.0013**</b>	Larger > Smaller
Sex	3.26	1, 50.7	9.30	<b>0.0036**</b>	Females > Males
Interval	--	2, 109	257	< <b>0.0001**</b>	Interval 1 > 3 > 2
Ecotype	-2.15	1, 50.6	8.33	<b>0.0057**</b>	L-fast > M-slow
Rearing treatment	-6.22	1, 41	25.6	< <b>0.0001**</b>	Warm > Cool
Ecotype×Interval	--	2, 80.9	3.99	<b>0.0222*</b>	See Figure 4
Rearing Treatment×Interval	--	2, 82.9	21.4	< <b>0.0001**</b>	See Figure 4
Ecotype×Rearing Treatment	6.11	1, 38.3	0.46	0.50	--
Ecotype×Rearing*Interval	--	2, 81.7	0.62	0.54	--
Population(Ecotype)	--	3, 20.8	0.13	0.94	--

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29 **Table 8. Repeated measures mixed model analysis for *T. elegans* plasma log<sub>10</sub>IGF-1 with**  
30 **prior growth (growth over the preceding interval) as a covariate across three time points.**

Source of Variation	Estimate	df <sub>n</sub> , df <sub>d</sub>	F	Pr > F	Direction of significant factors
Prior interval growth	-0.0014	1, 271	4.88	<b>0.0280*</b>	Slower > Faster
Sex	-0.053	1, 93.9	7.17	<b>0.0087**</b>	Males > Females
Time	--	2, 217	10.12	< <b>0.0001**</b>	
Ecotype	0.14	1, 106	3.58	0.06	
Rearing Treatment	0.15	1, 112	6.15	<b>0.0146*</b>	Warmer > Cooler
Ecotype×Time	--	2, 178	1.26	0.28	--
Rearing Treatment×Time	--	2, 193	1.73	0.18	--
Ecotype×Rearing Treatment	-0.13	1, 102	2.18	0.14	--
Ecotype×Rearing*Time	--	2, 183	2.49	0.08	See Figure 6
Population(Ecotype)	--	3, 100	0.51	0.67	--

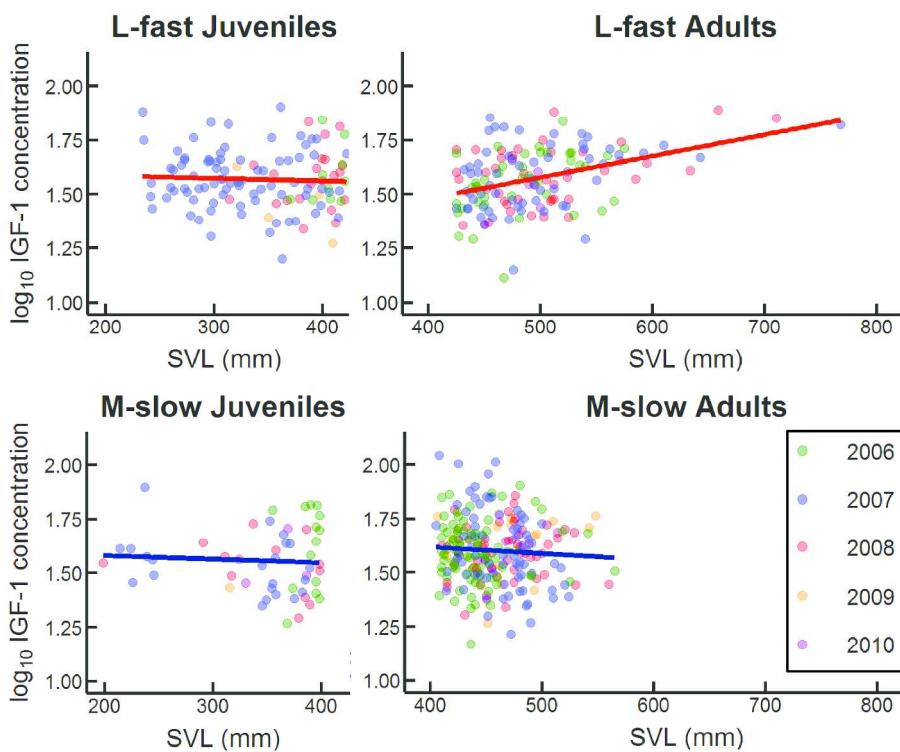


Figure 1. Scatterplots of the relationship between size (SVL in mm) and plasma IGF-1 concentration ( $\log_{10}$ -transformed ng/mL) for each life-stage by ecotype combination in field-caught *Thamnophis elegans*. Snakes were sampled from 2006-2010 at populations around Eagle Lake, CA, USA. Overall levels of IGF-1 varied across years (see Table 2), so values here are normalized with respect to year (from model of  $\log_{10}$ -transformed IGF-1 concentrations with year as a categorical fixed effect). The relationship between SVL and IGF-1 concentrations is significant for L-fast adults ( $R^2 = 0.14$ ,  $P < 0.001$ ).

292x215mm (300 x 300 DPI)

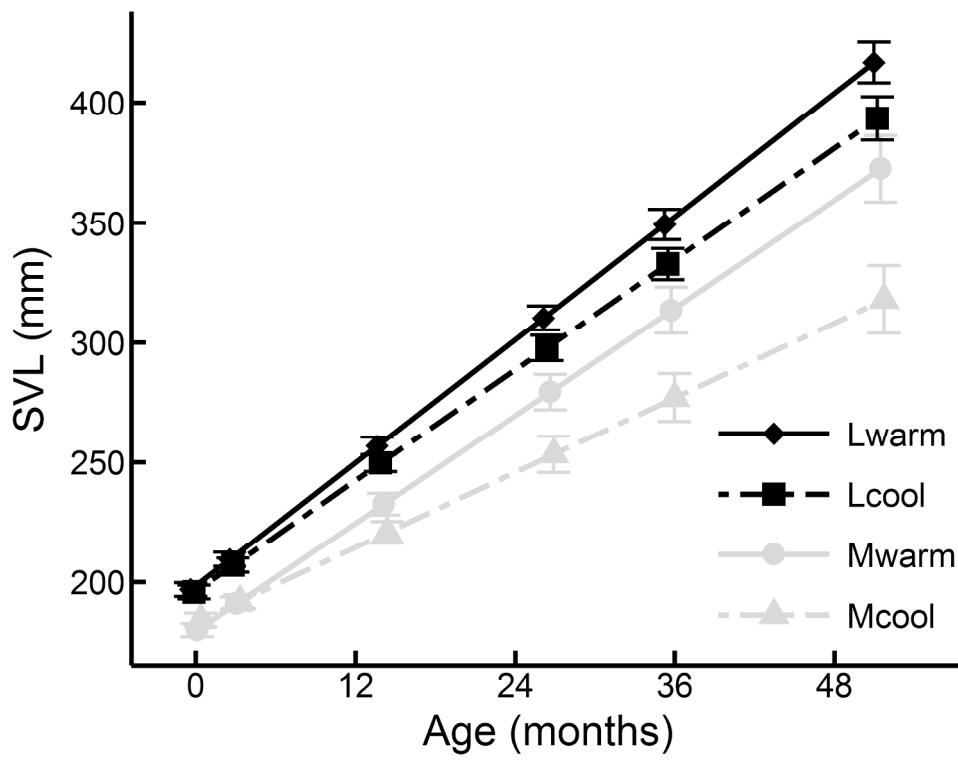


Figure 2 . Least-square means from the model of lifetime size (SVL in mm; see text for model details) for *T. elegans* juveniles raised in captivity, through age 4 years. Both ecotype and rearing treatment influenced lifetime growth. Error bars represent  $\pm$  SE.

203x152mm (300 x 300 DPI)

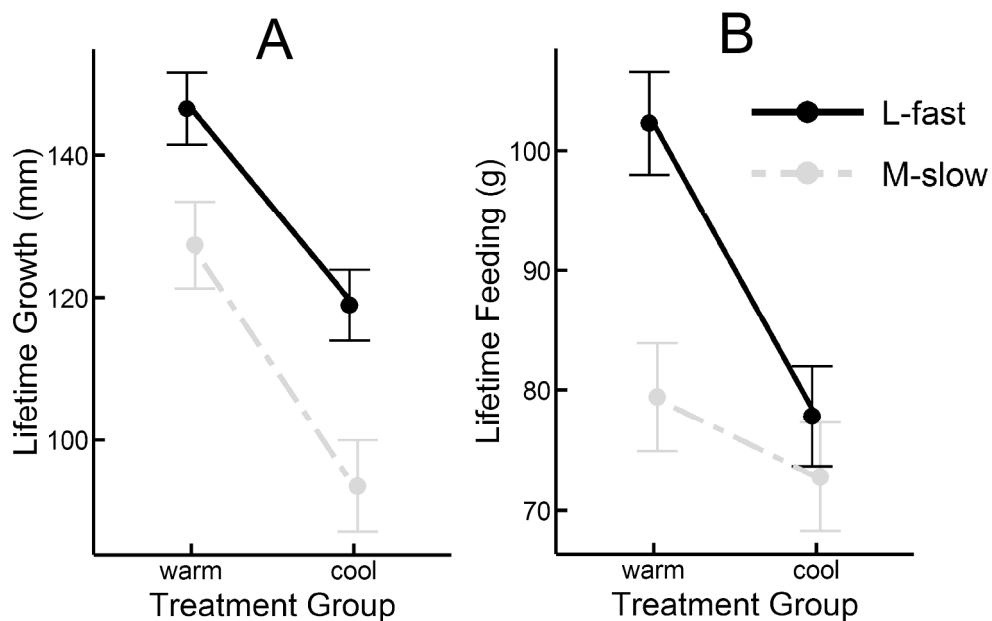


Figure 3. (A) Least-square means from the model of lifetime growth (SVL in mm; see text for model details) for *T. elegans* juveniles raised in captivity, from birth to age 4 years. Both ecotype and rearing treatment were significant factors in determining lifetime growth. (B) Least-square means from the model of lifetime feeding (amount consumed in g; see text for model details) for *T. elegans* juveniles raised in captivity, from birth to age 4 years. Lwarm snakes ate more than Lcool, Mwarm, or Mcool snakes (all adjusted pairwise comparisons significant with  $p < 0.01$ ). Error bars for both panels represent  $\pm$  SE.

254x158mm (300 x 300 DPI)

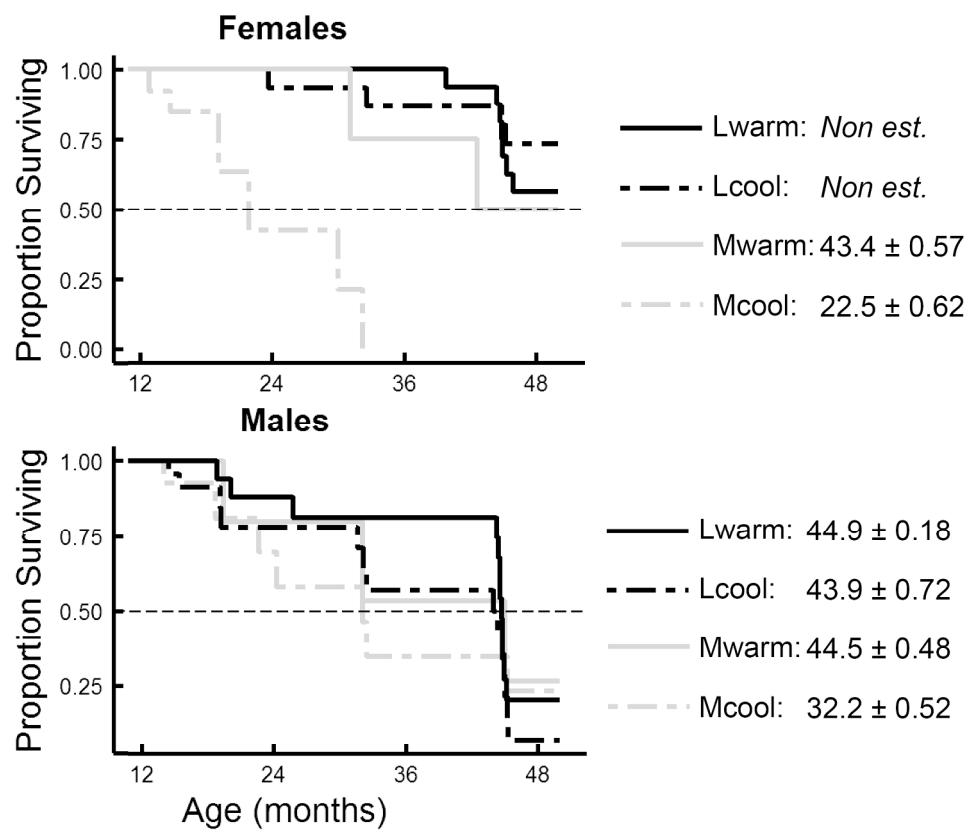


Figure 4. Survival curves for captive-born *Thamnophis elegans* by treatment group within each sex. Plots demonstrate the significant influence of ecotype, treatment, and sex on survivorship based on the semi-parametric Cox Proportional hazard model (see Table 6). Horizontal dotted line shows 50% survivorship. Estimates are median residual lifespans (in months)  $\pm$  SE.

207x177mm (300 x 300 DPI)



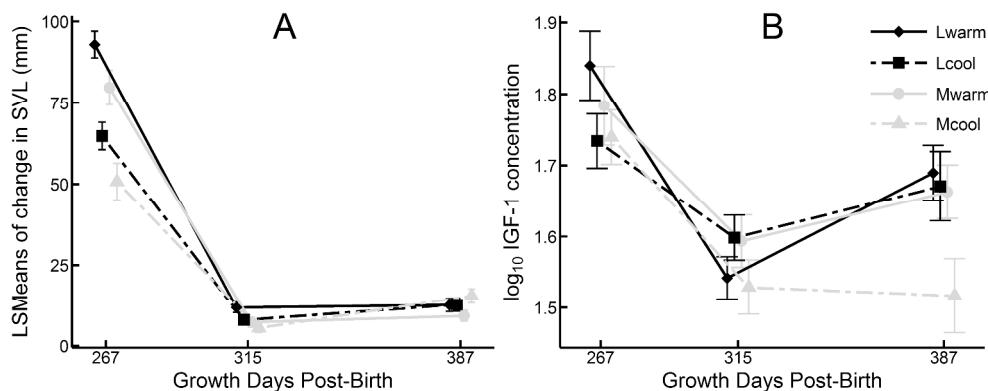


Figure 5 . (A) Growth (change in SVL, mm) for captive-born *T. elegans* over the three intervals. Estimates are shown on the last day of an interval; the number of days in each interval varied: Interval 1 = 178 days; Interval 2 = 48 days; Interval 3 = 72 days. The significant effect of temperature  $\times$  interval is seen in that during interval 1, warm snakes were growing fastest, whereas in interval 3, cooler snakes were growing fastest. The significant effect of ecotype  $\times$  time can be seen in that during interval 1, L snakes were growing fastest, whereas in interval 3, cooler snakes were growing fastest. (B) Least-square means from the model of plasma IGF-1 concentration ( $\log_{10}$ -transformed ng/mL). IGF-1 levels were assayed from samples collected from the final day of 3 intervals, with first measurement 267 days after birth. The significant 3-way interaction of ecotype  $\times$  treatment  $\times$  time can be seen in the deviation of the Mcool values from the others, Note that Mcool animals grew fastest, and faster than other categories, during interval 3. Error bars for both panels represent  $\pm$  SE.

400x158mm (300 x 300 DPI)

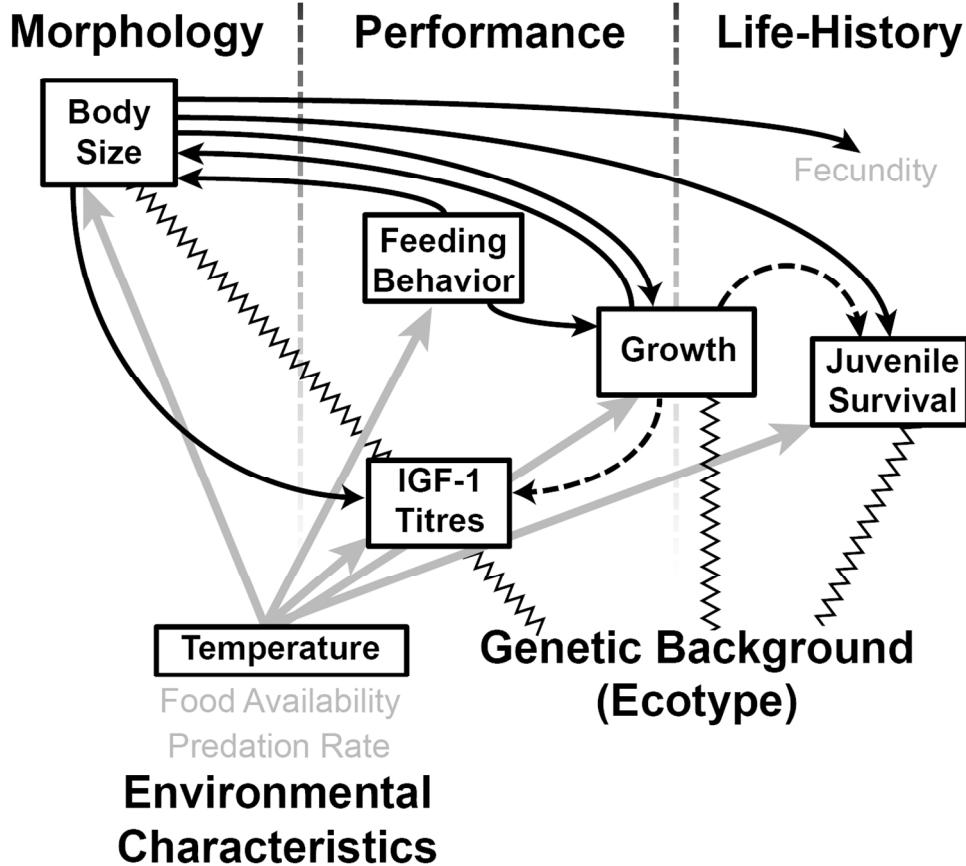


Fig. 6. The complex interplay of phenotypic traits involved in the morphology–performance–life-history web. Black arrows represent relationships between aspects of the phenotype, with solid lines indicating positive relationships and dashed lines indicating negative. Solid gray arrows represent the direct impact of environmental characteristics on traits. Zig-zag lines show traits in which genetic canalization has resulted in differences between distinct life-history ecotypes. Traits and factors shown in bold black type are included in the present study; traits and factors in gray type have been established in previous studies (Bronikowski & Arnold 1999, Sparkman and others 2007, Miller and others 2011, Sparkman and others 2013).

126x113mm (300 x 300 DPI)