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RESEARCH ARTICLE

The Arabidopsis MOS4-associated Complex Promotes MicroRNA Biogenesis and Precursor Messenger RNA Splicing

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ABSTRACT

In *Arabidopsis thaliana*, the MOS4-ASSOCIATED COMPLEX (MAC) is required for defense and development. The evolutionarily conserved, putative RNA helicase MAC7 is a component of the Arabidopsis MAC and the human MAC7 homolog, Aquarius, is implicated in pre-mRNA splicing. Here, we show that *mac7-1*, a partial loss-of-function mutant in *MAC7*, and two other MAC subunit mutants, *mac3a mac3b* and *prl1 prl2* (*pleiotropic regulatory locus*), exhibit reduced microRNA (miRNA) levels, indicating that MAC promotes miRNA biogenesis. The *mac7-1* mutant shows reduced primary miRNA (pri-miRNA) levels without affecting miRNA gene (*MIR)* promoter activity or the half-life of pri-miRNA transcripts. As a nuclear protein, MAC7 is not concentrated in dicing

bodies, but it affects the localization of HYPONASTIC LEAVES1 (HYL1), a key protein in pri-miRNA processing, to dicing bodies. Immunoprecipitation of HYL1 retrieved eleven known MAC subunits, including MAC7, indicating association between HYL1 and MAC. We propose that MAC7 links *MIR* transcription to pri-miRNA processing. RNA-seq analysis showed that down-regulated genes in MAC subunit mutants are mostly involved in plant defense and stimulus responses, confirming a role of MAC in biotic and abiotic stress responses. We also discovered global intron retention defects in mutants in three subunits of MAC, thus linking MAC function to splicing in Arabidopsis.

INTRODUCTION

MicroRNAs (miRNAs) are a class of small RNAs that are approximately 20 to 24 nucleotides in length and act as post-transcriptional regulators of gene expression in both animals and plants. MiRNAs are processed from hairpin-containing precursors, primary miRNAs (pri-miRNAs), by RNase III family enzymes. A mature miRNA is loaded into an Argonaute protein to form a silencing complex and guides this silencing complex to target RNAs through sequence complementarity with target RNAs to result in their degradation or translational repression (Rogers and Chen, 2013).

The plant miRNA pathway has been intensively studied in the past decade. Early studies identified key proteins with catalytic activities, including RNA polymerase II (Pol II) that transcribes miRNA genes (*MIR*) (Xie et al., 2005; Zheng et al., 2009), DICER-LIKE1 (DCL1), an RNase III family enzyme excising miRNAs from stem-loop precursors, HUA ENHANCER1 (HEN1), a methyltransferase stabilizing miRNAs by 2'-*O*-methylation (Park et al., 2002; Li et al., 2005; Yu et al., 2005; Yang et al., 2006b; Yu et al., 2010), and ARGONAUTE 1 (AGO1), the enzyme mediating miRNA-guided target RNA cleavage (Baumberger and Baulcombe, 2005; Ji et al., 2011; Carbonell et al., 2012; Arribas-Hernandez et al., 2016). Many factors that assist in the transcription and processing steps of miRNA biogenesis have been identified (Rogers and Chen, 2013; Achkar et al., 2016). Pol II-mediated *MIR* transcription requires the transcriptional coactivator Mediator (Kim et al., 2011) and the transcription factor NEGATIVE ON TATA

LESS 2 (NOT2) (Wang et al., 2013), and is regulated by CYCLIN-DEPENDENT KINASES (CDKs) (Hajheidari et al., 2012). In addition, the DNA binding protein CELL DIVISION CYCLE 5 (CDC5) (Zhang et al., 2013b) and Elongator (Fang et al., 2015) also regulate *MIR* transcription by Pol II.

While interacting with Pol II, NOT2, CDC5 and Elongator also interact with DCL1 and several DCL1-interacting proteins, and probably recruit them to pri-miRNAs to facilitate their processing (Wang et al., 2013; Zhang et al., 2013b; Fang et al., 2015). During the processing of miRNA precursors, DCL1 forms a dicing complex with the dsRNA binding domain protein HYPONASTIC LEAVES1 (HYL1/DRB1), and the zinc finger protein SERRATE (SE) (Han et al., 2004; Kurihara and Watanabe, 2004; Kurihara et al., 2006; Yang et al., 2006a; Dong et al., 2008). DCL1 and HYL1 are enriched in subnuclear bodies, referred to as dicing bodies, which are considered to be sites of miRNA precursor processing (Han et al., 2004; Fang and Spector, 2007; Song et al., 2007). Many other proteins also interact with DCL1 directly or indirectly in miRNA biogenesis, such as Cap-Binding Proteins (CBPs) (Laubinger et al., 2008; Raczynska et al., 2014), the forkhead-associated domain containing protein DAWDLE (DDL) (Yu et al., 2008), the G-patch domain containing RNA binding protein TOUGH (TGH) (Ren et al., 2012), and the WD-40 protein PLEIOTROPIC REGULATORY LOCUS 1 (PRL1) (Zhang et al., 2014). In addition, several proteins act in miRNA biogenesis through the regulation of HYL1 phosphorylation, such as C-TERMINAL DOMAIN PHOSPHATASE-LIKE (CPL) proteins and a K homology (KH) domain protein REGULATOR OF CBF GENE EXPRESSION 3 (RCF3) (Manavella et al., 2012; Karlsson et al., 2015). Several other factors, including two core members of the THO/TREX complex THO2 and EMU (Furumizu et al., 2010; Francisco-Mangilet et al., 2015), and the RNA binding protein MODIFIER OF SNC1, 2 (MOS2) (Wu et al., 2013) do not seem to interact with any known dicing complex components, but still affect miRNA biogenesis.

Among the known miRNA biogenesis factors, CDC5 and PRL1 belong to the same complex, the MOS4-associated complex (MAC). MAC is a highly conserved complex among eukaryotes, with its orthologs known as the NINETEEN COMPLEX (NTC) or Prp19 complex (Prp19C) in yeast and humans. The *Arabidopsis thaliana* MAC, yeast and human NTC/Prp19C all associate with the spliceosome and are predicted to share conserved functions in splicing in all three systems (Monaghan et al., 2009; Johnson et al., 2011; Koncz et al., 2012; Deng et al., 2016). Arabidopsis CDC5 is a MYB-related transcription factor and PRL1 is a conserved nuclear WD-40 protein. As core components of MAC, they regulate plant development and immunity through molecular mechanisms that remain unclear (Nemeth et al., 1998; Lin et al., 2007; Palma et al., 2007). They both promote miRNA biogenesis but may have distinct molecular functions. CDC5 binds to *MIR* promoters and interacts with DCL1 and SE to enhance miRNA biogenesis (Zhang et al., 2013b), while PRL1 may stabilize pri-miRNAs through its RNA binding activity and enhance DCL1 activity (Zhang et al., 2014). Other Arabidopsis MAC subunits include two homologous proteins MAC3A and MAC3B (MAC3B was shown to have E3 ligase activity *in vitro*), MAC7/Aquarius (an RNA helicase), and more than ten other members (Wiborg et al., 2008; Monaghan et al., 2009; Monaghan et al., 2010; Koncz et al., 2012).

Arabidopsis MAC7 is a putative RNA helicase, which is highly conserved in eukaryotes. The human MAC7 homolog is the intron-binding protein Aquarius (IBP160/AQR), which has ATPase and RNA helicase activities. It associates with the spliceosome and contributes to efficient precursor-mRNA splicing *in vitro* (Hirose et al., 2006; De et al., 2015). Recent publications show that EMB-4, the *Caenorhabditis elegans* homolog of MAC7, physically interacts with germline AGOs. It participates in the nuclear RNAi pathway and maintains the homeostasis of germline transcriptome in worms (Akay et al., 2017; Tyc et al., 2017). *MAC7* was predicted to be an essential gene for embryo development (*EMB* gene) in Arabidopsis based on sequence similarity with *EMB* genes found

in other eukaryotes (Tzafrir et al., 2004). However, the molecular function of *MAC7* remains unknown.

In this study, we performed a genetic screen using a miRNA activity reporter line, the *pSUC2:amiR-SUL* (*amiR-SUL*) transgenic line (de Felippes et al., 2011). The expression of *SUCROSE-PROTON SYMPORTER 2 (SUC2)* promoter-driven artificial miRNA targeting the *CHLORINA42 (CH42)* gene creates a bleached phenotype along the leaf veins. This bleaching of mesophyll cells caused by the silencing effects of the artificial miRNA results in an easily scorable phenotype reflecting miRNA activities in plants (de Felippes et al., 2011). From this screen, we identified a point mutation in *MAC7*, *mac7-1*, as a miRNA activity suppressor mutant. We showed that MAC7 affects miRNA accumulation through promotion of pri-miRNA biogenesis in Arabidopsis. We found that in other MAC subunit mutants, including *mac3a mac3b* and *prl1 prl2* double mutants, miRNA biogenesis is also compromised, indicating that MAC participates in miRNA biogenesis as a complex. Consistent with this, HYL1 immunoprecipitation mass spectrometry analyses revealed that HYL1 associates with MAC *in vivo*. In addition, we uncovered global intron retention defects in *mac7-1*, *mac3a mac3b* and *prl1 prl2* mutants through RNA-seq analysis. Our molecular characterization of MAC7 and its associated MAC components revealed their functions in miRNA biogenesis and pre-mRNA splicing, which could possibly explain their roles in plant development and stress responses.

RESULTS

A silencing suppressor mutant exhibits pleiotropic phenotypes and reduced miRNA levels

To identify new players of the miRNA pathway, we performed an ethylmethane sulfonate (EMS) mutagenesis screen using the *amiR-SUL* line (de Felippes et al., 2011). From this screen, we identified a mutant, which we named *mac7-1* based on subsequent characterization (*amiR-SUL mac7-1*) with a reduced area of bleaching along the veins, indicating compromised miRNA activity (Figure 1A).

The mutant has pleiotropic phenotypes, such as pointed leaves, reduced root length, reduced number of lateral roots, smaller plant stature and reduced fertility (Figure 1B, C and Supplemental Figure 1). We crossed *amiR-SUL mac7-1* with wild type (Col-0) plants to remove the *amiR-SUL* transgene. We found that the *mac7-1* mutant shows the same range of phenotypes in the Col-0 background as in the *amiR-SUL* background (Figure1B, C and Supplemental Figure 1).

Because the mutant shows compromised amiR-SUL activities, we speculated that reduced miRNA accumulation could be a reason. We performed RNA gel blot analyses to detect amiR-SUL as well as many endogenous miRNAs. We found that amiR-SUL, miR156, miR171, miR390 and many other miRNAs showed reduced accumulation in both inflorescences and seedlings. The reduction in miRNA abundance was small but reproducible in several biological replicates (Figure 1D and Supplemental Figure 2). To assess the global influence of the mutation on small RNAs, we also performed small RNA-seq with Col-0 and *mac7-1* seedlings. We found that 21-nt and 24-nt small RNAs, which represent the two most abundant small RNA size classes, showed a significant global reduction in the *mac7-1* mutant. miRNAs showed a slight global reduction in the *mac7-1* mutant but the reduction was not statistically significant (Figure 1E and Supplemental Data set 1). Most miRNAs that were found to show reduced abundance by RNA gel blotting also had lower levels in *mac7-1* in small RNA-seq, although a few miRNAs did not (e.g. miR156, miR164) (Supplemental Figure 2C). The minor inconsistency between RNA gel blotting and small RNA-seq could be caused by technical limitations in small RNA-seq, e.g. bias in RNA adaptor ligation or PCR amplification. Since small RNA-seq entails more procedures that are prone to bias, we believe that the RNA gel blotting results are more accurate.

We examined the expression of *CH42* and eight known miRNA target genes in *amiR-SUL* and *amiR-SUL mac7-1*. Opposite to the reduced miRNA accumulation,

the expression levels of *CH42* and seven endogenous miRNA targets were upregulated in *amiR-SUL mac7-1* (Figure 1F).

The reduced bleaching phenotype in *amiR-SUL mac7-1* might be attributed to other factors other than, or in addition to, reduced amiR-SUL activity, e.g. enhanced transcription of *CH42*. We performed RT-qPCR and immunoblotting to examine *CH42* mRNA and protein levels, respectively, in Col-0 and *mac7-1* to determine whether an amiR-SUL-independent effect of the *mac7-1* mutation was present. While CH42 mRNA and protein levels were both increased in *amiR-SUL mac7-1* in comparison to *amiR-SUL*, no difference was found between Col-0 and *mac7-1* (Supplemental Figure 3), indicating that the reduced bleaching phenotype and elevated *CH42* expression in *amiR-SUL mac7-1* was most likely caused by changes in amiR-SUL activity only.

A point mutation in *MAC7* is responsible for the suppression of amiR-SUL-induced silencing

Through genome re-sequencing of pooled F2 mutants from a backcross between *amiR-SUL mac7-1* and the parental *amiR-SUL* line, we found a C-to-T nucleotide transition that changes a conserved glutamic acid to lysine (E1131K) in the open reading frame (ORF) of At2g38770 (*MAC7/EMB2765*) (Figure 2A). To determine whether this mutation was the causal mutation that suppressed *amiR-SUL*, we first examined the linkage between the mutation and the visible phenotype. In the F2 population of the backcross, we identified 84 plants with the *mac7-1* phenotype, i.e., reduced vein-centered leaf bleaching. These plants were genotyped for the C-to-T mutation in *MAC7*. All 84 plants were found to be homozygous for this mutation, and thus the *amiR-SUL* suppressor phenotype was linked with this mutation.

Next, to confirm that the causal mutation of the suppressor was in *MAC7*, we generated *MAC7* promoter driven *MAC7-GFP* and *MAC7-mCherry* fusion constructs, and introduced them into *mac7-1* in *amiR-SUL* and Col-0

backgrounds. *pMAC7:MAC7-GFP* or *pMAC7:MAC7-mCherry* fully complemented the morphological defects (Figure 2B) and *pMAC7:MAC7-mCherry* restored the accumulation of miRNAs of the mutant in both inflorescences and seedlings (Figure 2C, D). Thus, the causal mutation in the suppressor was in *MAC7* and we named this mutation *mac7-1*.

To obtain another *mac7* allele, we ordered a T-DNA insertion line, SALK_129044 (which we named *mac7-2*) (Supplemental Figure 4A). Genotyping *mac7-2* in the progeny of a selfed *mac7-2/+* plant revealed 2:1 segregation between heterozygous and wild-type plants (Supplemental Table 1), suggesting that *mac7-2* homozygous plants were embryo lethal. Unlike wild-type siliques that contained only normal-looking green seeds, siliques from heterozygous (*mac7-2/+)* plants had aborted seeds that appeared pale white (Supplemental Figure 4B). An approximate 3:1 ratio between normal seeds and aborted seeds were found (Supplemental Table 1), which was a strong indication of embryo lethality of the homozygous mutant. In addition, the viability of pollen from *mac7-2/+* plants appeared normal (Supplemental Figure 4C). The above evidence demonstrated that *mac7-2* is a recessive, embryo-lethal mutation. As *mac7-1* homozygous plants are viable, this mutation is likely a partial loss-of-function allele.

The *MAC7* gene encodes an RNA helicase conserved from yeast to animals and plants. The structure of the human MAC7 ortholog, Aquarius, has been determined (De et al., 2015). Based on homology modeling, we predict that the plant MAC7 also has a long N-terminal domain composed of armadillo repeats (ARM), which is crucial for protein–protein interactions, a stalk and a β-barrel domain, which has an architectural role, a thumb and a pointer domain, which may participate in interactions with other proteins, and two RecA-like domains (RecA1 and RecA2), which form a motor module required for ATP hydrolysis, RNA unwinding and the coupling of these two processes. The *mac7-1* mutation leads to a change of a highly conserved amino acid in the RecA1 domain, which

very likely affects the core functions of this protein in ATP-binding/hydrolysis and nucleic acid unwinding (Figure 2A and Supplemental Figure 5) (De et al., 2015; Ozgur et al., 2015).

MAC7 promotes pri-miRNA levels without affecting *MIR* promoter activities or pri-miRNA transcript half-life

To determine the molecular mechanisms of MAC7 in miRNA biogenesis, we examined the levels of pri-miRNAs. We found that the levels of pri-miRNAs from many *MIR* genes were reduced in the *mac7-1* mutant (Figure 3A). This reduction could be attributed to reduced transcription of *MIR* genes or enhanced pri-miRNA degradation or processing. We first examined whether *mac7-1* affects *MIR* promoter activities. We crossed *mac7-1* to a miRNA promoter reporter line, *pMIR167a:GUS*, in which the transgene was inserted into a single genomic locus. We then examined GUS activity by staining and *GUS* transcripts level by real-time RT-PCR in wild-type and *mac7-1* plants, in which the transgene was homozygous. There was no detectable difference between wild type and *mac7-1* in terms of GUS activity or *GUS* transcripts level, indicating that *mac7-1* did not affect miRNA promoter activities (Figure 3B). We also examined whether the reduction of amiR-SUL level in *mac7-1* was due to reduced *SUC2* promoter activity. If this was the case, we would expect the endogenous *SUC2* RNA to be reduced in abundance in *mac7-1*. Real-time RT-PCR revealed that *SUC2* mRNA levels did not change significantly in *mac7-1*, implying that *SUC2* promoter activity was not affected by *mac7-1* and the reduction in amiR-SUL levels was not due to impaired *SUC2* promoter activities in the mutant (Supplemental Figure 6A).

Next, we measured the half-lives of pri-miRNAs in *amiR-SUL* and *mac7-1 amiR-SUL*. Seedlings were treated with the transcription inhibitor cordycepin, and RNAs were isolated at different time points. The levels of pri-amiR-SUL, pri-miR167a, and pri-miR172a, were determined by real-time RT-PCR. Similar half-lives were found for these pri-miRNAs in the two genotypes (Figure 3C).

It is also possible that MAC7 affects miRNA biogenesis indirectly through affecting the expression of key miRNA biogenesis factors. To test this possibility, we determined transcript and protein levels of several key miRNA biogenesis factors [i.e., DCL1, HYL1, SE, AGO1, HEN1 (transcript only)] in *amiR-SUL* and *amiR-SUL mac7-1*. No differences were detected for any of the genes in the two genotypes (Supplemental Figure 6B, C, D).

MAC7 is a nuclear protein interacting with other MAC components, which are also required for miRNA biogenesis

To further characterize the molecular functions of the MAC7 protein, we examined the subcellular localization of fluorescent protein tagged MAC7. Transgenic plants expressing *p35S:YFP-MAC7* or *pMAC7:MAC7-GFP* exhibited nuclear GFP signals (Figure 4A), which is consistent with the presence of a predicted Nuclear Localization Signal (NLS) in MAC7 (Figure 2A). To determine if MAC7 localized in dicing bodies, we transiently expressed YFP-MAC7 and DCL1-YFP in tobacco leaves and compared their expression patterns. While DCL1-YFP showed weak nucleoplasmic signals and strong dicing body signals, YFP-MAC7 exhibited dispersed distribution in the nucleoplasm and was absent from the nucleolus (Supplemental Figure 7A). When YFP-MAC7 was co-expressed with TagRFP-HYL1, signals from the two proteins overlapped in the nucleoplasm while only TagRFP-HYL1 was found concentrated in dicing bodies (Supplemental Figure 7B). Immunoblot analyses confirmed the expression of GFP- or YFP-tagged MAC7 in *Arabidopsis* transgenic lines (Supplemental Figure 7C) and in infiltrated tobacco leaves (Supplemental Figure 7D). These data suggest that MAC7 does not display the dicing body patterns as DCL1 and HYL1 do. The dispersed nucleoplasmic distribution of MAC7 suggests broader roles than miRNA biogenesis.

To uncover interacting partners of MAC7 and to determine whether MAC7 interacts with any known miRNA pathway proteins, we immunoprecipitated (IP)

MAC7 and performed mass spectrometry (MS) analyses. In one experiment, IP was performed with Col-0 plants using anti-MAC7 antibodies with pre-immune IgG as a negative control. In another experiment, IP was performed using Chromotek-RFP-Trap with a *pMAC7:MAC7-mCherry mac7-1* line in which the transgene rescued the mutant phenotypes; IP was performed using the same RFP-Trap with Col-0 as a negative control. The two independent experiments consistently pulled down all major MAC components, demonstrating that MAC7 was part of the MAC (Supplemental Table 2). An overlapping set of proteins was identified from our MAC7 IP-MS and from published MOS4 IP-MS (Figure 4B and Supplemental Table 2) (Monaghan et al., 2009). Besides the known MAC subunits (Monaghan et al., 2009), seven additional proteins were discovered as potential MAC7-associated proteins (Supplemental Table 2), but the association between these proteins and MAC7, or whether they also belong to MAC, needs to be further investigated.

Previous studies showed that the MAC subunits CDC5 and PRL1 promote miRNA biogenesis (Zhang et al., 2013b; Zhang et al., 2014). To further confirm the interactions between MAC7 with these two MAC subunits that are also miRNA biogenesis factors, we performed Bimolecular Fluorescence Complementation (BiFC) analysis, and found that MAC7 interacted with CDC5 and also interacted, albeit weakly, with PRL1 (Figure 4C). Similar BiFC studies did not reveal interactions between MAC7 and HYL1 or DCL1 (Figure 4C).

These results and our work on MAC7 raise the possibility that MAC plays a role in miRNA biogenesis, acting as a complex. To test this hypothesis, we examined whether core subunits of MAC are required for miRNA biogenesis. Arabidopsis MAC3B is a U-box E3 ubiquitin ligase, and is a core MAC component (Monaghan et al., 2009). MAC3B has a homolog, MAC3A, which shares 82% identity with MAC3B at the amino acid level (Monaghan et al., 2009). PRL1 is also a core member of MAC. It has a homolog PRL2, which is expressed at a much lower level than PRL1 (Weihmann et al., 2012). Previous studies showed that a *prl1*

single mutant has reduced miRNA levels but a *mac3b* mutant (SALK_050811) does not (Zhang et al., 2014). Several siRNAs, ta-siRNA255 and miRNA171 were found to show reduced accumulation in the *mac3a mac3b* (SALK_089300 and SALK_050811) double mutant (Zhang et al., 2013a). Here we examined the morphological and molecular phenotypes of two double mutants, *mac3a mac3b* and *prl1 prl2*. Similar to other MAC subunit mutants, *mac3a mac3b* and *prl1 prl2* exhibited pleiotropic developmental phenotypes (Figure 5A). RNA gel blot analyses showed that miR156, miR166, and miR171 all exhibited reduced accumulation in these double mutants (Figure 5B). We also performed small RNA-seq with Col-0 and *prl1 prl2* seedlings to assess the global changes of small RNAs in *prl1 prl2*. Similar to those of *mac7-1*, 21-nt and 24-nt small RNAs showed a global reduction in *prl1 prl2*. Unlike in *mac7-1*, a significant global reduction in miRNA levels was also found in *prl1 prl2*. (Figure 5C and Supplemental Data Set 1). This is consistent with *mac7-1* being a weak allele representing a partial compromise in MAC function. RT-qPCR analyses revealed that the levels of pri-miRNAs were also reduced in *mac3a mac3b* and *prl1 prl2* (Figure 5D). Thus, the molecular phenotypes of *mac3a mac3b* and *prl1 prl2* mutants were similar to those of *mac7-1*.

MAC interacts with HYL1

As CDC5 and PRL1 interact with dicing complex components (Zhang et al., 2013b; Zhang et al., 2014), it is possible that MAC7, or the entire MAC, associates with the dicing complex. To explore the interactions between the dicing complex and MAC, we performed HYL1 IP-MS. In one experiment, IP was performed with a *p35S:HYL1-YFP* line and the negative control *p35S:YFP* transgenic line and Col-0 using Chromotek-GFP-Trap. In another experiment, IP was performed with Col-0 and the negative control *hyl1-2* using anti-HYL1 antibodies. Eleven MAC subunits, including MAC7, were found in both IP-MS experiments (Supplemental Table 3), indicating that HYL1 associates with MAC *in vivo*.

The interactions between HYL1 and MAC raised the possibility that MAC7 could be involved in pri-miRNA processing through affecting the dicing body localization of HYL1. We crossed *mac7-1* to a *HYL1-YFP* transgenic line and quantified dicing body numbers in wild type and *mac7-1*. The number of HYL1-YFP-positive dicing bodies was significantly reduced in *mac7-1* compared with wild type plants (Figure 6A,B), demonstrating that *MAC7* is required for the proper localization of HYL1 in dicing bodies, which might partially explain the compromised miRNA levels in the *mac7-1* mutant.

Down-regulated genes in *mac* mutants are significantly related to stress responses

We also explored whether MAC plays a role in RNA metabolism in general. We performed RNA-seq with Col-0, *mac7-1*, *mac3a mac3b*, and *prl1 prl2* seedlings in two biological replicates. Differentially expressed genes (DEGs) were identified between each mutant and wild type with fold change of 1.5 or more. Among the three mutants, we identified 2,007 and 2,268 down-regulated (hypo-DEGs) and up-regulated (hyper-DEGs) genes, respectively. 189 hypo-DEGs and 222 hyper-DEGs were commonly found among *mac7-1*, *mac3a mac3b* and *prl1 prl2* (Figure 7 and Supplemental Data Set 2). The overlap of the DEGs among these three mutants was significant (SuperExactTest, $p = 0$), indicating that these MAC components function as a complex and regulate the same group of genes. The large portion of non-overlapped DEGs indicates that each subunit may also have its own function independent of MAC.

To understand the biological functions of MAC, we examined the Gene Ontology terms enriched in the common DEGs (Gene Ontology, 2015). An enrichment of genes involved in stress responses, including stimulus responses, plant defense or immune responses was found in the hypo-DEGs. As for the hyper-DEGs, GO terms in various small molecule biosynthetic and metabolic processes were enriched (Figure 7 and Supplemental Data Set 3). Many more GO terms related to stress responses were identified from the hypo-DEGs than the hyper-DEGs,

implying that MAC tends to activate the expression of stress response genes. The results are consistent with previous findings showing that *mac3a mac3b* and *prl1 prl2* double mutants are more susceptible to pathogen infection (Monaghan et al., 2009; Weihmann et al., 2012). To test whether *MAC7* is also required for plant immunity, *mac7-1* was infected with *Pseudomonas syringae* p.v. *maculicola* (*P.s.m.*) strain ES4326 together with Col-0 and *prl1-2*. The *prl1* mutants are more susceptible to pathogen infection and therefore served as a positive control (Weihmann et al., 2012). Pathogen growth was assayed three days after bacterial inoculation. While *P.s.m.* ES4326 accumulated to higher levels in *prl1-2* than in wild type, *mac7-1* was similar to wild type in terms of bacterial titer (Supplemental Figure 8A). Perhaps this was due to *mac7-1* being a weak allele, in which the magnitude of downregulation of the hypo-DEGs was small compared to that in other *mac* mutants (Supplemental Figure 8B).

MAC subunits affect pre-mRNA splicing

Homologs of MAC in yeast and mammals play a critical role in pre-mRNA splicing. In Arabidopsis, we lack evidence for a widespread role of MAC in splicing; only the splicing patterns of several genes, such as *SUPPRESSOR OF NPR1-1 CONSTITUTIVE 1 (SNC1)* and *RESISTANCE TO PSEUDOMONAS SYRINGAE4 (RPS4)*, were shown to be altered in several MAC subunit mutants (Xu et al., 2012; Zhang et al., 2013a).

Since intron retention is a major form of alternative splicing in Arabidopsis (Ner-Gaon et al., 2004), we examined whether *mac7-1*, *mac3a mac3b*, and *prl1 prl2* mutants had global intron retention defects. The ratio of RNA-seq reads mapping to introns (including 5'/3' splice sites) and those mapping to exons only was used as a measure of intron retention (see Methods for details). All annotated transcripts were considered in sum as long as the read counts passed an abundance filter. All three mutants exhibited significantly higher levels of intron retention compared to Col-0 (Wilcoxon test, $p < 2.2e-16$) (Figure 8A and Supplemental Data Set 4). Next, using the ratio of intron reads vs. exon reads as

448 a measure of intron retention levels, we identified genes with intron retention
449 defects in each mutant as compared to wild type (see Methods for details). 2819,
450 1466 and 298 genes were found to have intron retention defects in *mac3a mac3b*,
451 *prl1 prl2* and *mac7-1*, respectively (Figure 8B, C and Supplemental Data Set 4).
452 Significant overlap was found among the genes with intron retention defects in
453 the three mutants (SuperExactTest, $p < 0.05$), indicating that MAC works as a
454 complex in pre-mRNA splicing (Figure 8C). Two examples of intron retention
455 events in these mutants are shown in Figure 8D.
456
457 We next examined whether there was any correlation between splicing defects
458 and gene expression. We compared the expression levels of genes with intron
459 retention defects in each mutant vs. wild type. As a reference, all genes that
460 passed a minimum intron read coverage filter (see Figure 8 legend) were
461 analyzed. There was no correlation between intron retention defects and the
462 status of their differential expression in the *mac* mutants. Like all analyzed genes,
463 genes with intron retention defects were increased, reduced, or unchanged in
464 expression levels in each mutant as compared to wild type (Figure 9A and
465 Supplemental Data Set 5). In addition, we examined intron vs. exon ratios in total
466 genes, up-regulated genes and down-regulated genes in each mutant. Both up-
467 regulated and down-regulated genes showed significant intron retention as total
468 genes (Wilcoxon test, $p < 2.2e-16$), again indicating that the intron vs. exon ratio
469 has no correlation with gene expression changes (Figure 9B and Supplemental
470 Data Set 4).
471
472 Given the intron retention defects and miRNA accumulation defects in the *mac*
473 mutants, we asked whether intron retention in pri-miRNAs contributed to the
474 miRNA accumulation defects. Many *MIR* genes were shown to have introns,
475 which are spliced out in pri-miRNAs (Laubinger et al., 2008; Zhan et al., 2012;
476 Zielezinski et al., 2015). We performed RT-PCR with intron-flanking primers to
477 detect unspliced miRNA precursors, including pri-miR163, pri-miR156, pri-
478 miR166, pri-miR168, and pri-miR172. Genomic DNA was amplified with the

same sets of primers to indicate the sizes of intron-containing fragments. No intron retention was observed in these miRNA precursors in *mac7-1* (Supplemental Figure 9).

Thus, intron retention and differential expression of genes (including *MIR* genes) are not linked. MAC seems to regulate gene expression and RNA splicing independently.

DISCUSSION

MAC7 is an evolutionarily conserved protein across eukaryotes, however little is known about the molecular and biological functions of MAC7 or its orthologs. The human ortholog Aquarius is an RNA helicase with ATPase activity, and it binds to introns to assist intron splicing *in vitro* (Hirose et al., 2006; De et al., 2015). The *C. elegans* ortholog EMB-4 was reported to act in the nuclear RNAi pathway, where it interacts with nuclear AGOs and functions in germline-specific chromatin remodeling (Checchi and Kelly, 2006; Akay et al., 2017; Tyc et al., 2017). In Arabidopsis, MAC7 is involved in plant defense and was predicted to be an essential gene (Tzafrir et al., 2004; Monaghan et al., 2009). In this study, we showed that a T-DNA insertion mutant of *MAC7* is indeed embryo lethal, and we isolated the first viable *mac7* mutant, *mac7-1,* as a miRNA biogenesis-defective mutant. In this mutant, the artificial miRNA, amiR-SUL, and many endogenous miRNAs show reduced accumulation. MAC7 was identified as a MAC subunit through MOS4 IP-MS (Monaghan et al., 2009); our MAC7 IP-MS confirmed this. Two other MAC subunits, CDC5 and PRL1, were previously shown to promote miRNA biogenesis (Zhang et al., 2013b; Zhang et al., 2014). In this study, we showed that MAC3 also has a similar role. Thus, it is likely that MAC as a complex promotes miRNA biogenesis in general. Based on small RNA-seq analysis, MAC may promote the biogenesis of not only miRNAs but also siRNAs. Both 21-nt and 24-nt small RNAs, the two most abundant small RNA size classes in Arabidopsis, are reduced in *mac7-1* and *prl1 prl2* mutants. A role of MAC in

promoting siRNA biogenesis is also supported by previous findings on CDC5, MAC3 and PRL1 (Zhang et al., 2013a; Zhang et al., 2013b; Zhang et al., 2014).

How does MAC7 promote miRNA biogenesis? In *mac7-1*, the reduced accumulation of miRNAs correlated with reduced levels of pri-miRNAs. Thus, MAC7 probably acts in miRNA biogenesis by promoting *MIR* transcription or pri-miRNA stability. However, the activity of a promoter driving *MIR* expression was not affected in *mac7-1*, nor was the endogenous *SUC2* promoter activity. The half-lives of pri-miRNAs were not affected either. Although we cannot exclude the possibility that MAC7 promotes pri-miRNA processing into pre- or mature miRNAs and prevents pri-miRNA decay at the same time and therefore pri-miRNA half-lives appeared unchanged in the mutant, we prefer to hypothesize that MAC7 plays a role in transcription elongation and/or maturation of pri-miRNAs, considering the interactions between Pol II and MAC subunits CDC5 and PRL1, as well as the functions of yeast NTC in transcription elongation (Kuraoka et al., 2008; Chanarat and Strasser, 2013; Zhang et al., 2013b; Zhang et al., 2014). Given the role of MAC7 in pre-mRNA splicing discovered before and in this study (Xu et al., 2012; Zhang et al., 2013a), we considered the possibility that MAC7 acts in miRNA biogenesis by promoting the splicing of pri-miRNAs. However, we did not detect intron retention in several intron-containing pri-miRNAs in *mac7-1*, although the mature miRNAs from those pri-miRNAs are reduced. In addition, *MIR* genes without any introns (e.g. the *pSUC2:amiR-SUL* transgene, *MIR159a*, and *MIR167a*) (Szarzynska et al., 2009; de Felippes et al., 2011) were also affected in *mac7-1*. Thus, the miRNA biogenesis defects of *mac7-1* could not be attributed to defects in splicing. Intriguingly, we observed that MAC7 affects HYL1 localization to dicing bodies. Little is known about how dicing bodies form. MAC7 may recruit the dicing complex to pri-miRNAs through protein–protein interactions to form dicing bodies. Alternatively, dicing complex proteins are recruited by pri-miRNAs to dicing bodies, and therefore the reduced number of dicing bodies in *mac7-1* could be a consequence of reduced pri-miRNA levels in the mutant. We favor the second hypothesis because MAC7

plays a more general role in transcription and RNA metabolism (such as splicing), while the dicing complex acts more specifically on miRNA precursors and is presumably not recruited to other RNAs that MAC7 may also act on.

CDC5 and PRL1 were shown to interact with dicing complex proteins (e.g. DCL1, SERRATE) *in vivo* through co-IP or BiFC analyses (Zhang et al., 2013b; Zhang et al., 2014). We did not detect interactions between MAC7 and the dicing complex through MAC7 IP-MS or BiFC analyses. However, many MAC components including MAC7 were found in HYL1 IP-MS, which clearly indicates association between the dicing complex and MAC *in vivo*. The possible reasons for the inability of MAC7 to pull down the dicing complex proteins are: 1) The interaction between MAC7 and the dicing complex is indirect or weak, and bridged through CDC5 or PRL1; 2) only a small portion of MAC7 proteins interacts with the dicing complex. The second hypothesis is consistent with MAC7 having broader functions beyond miRNA biogenesis. We hypothesize that most of HYL1 or the dicing complex is associated with MAC, but not the other way around, which explains the recovery of MAC in HYL1 IP-MS.

MAC has functions beyond miRNA biogenesis in Arabidopsis. In our RNA-seq analysis, stress response-related GO terms were significantly enriched in down-regulated genes in three MAC subunit mutants. This is consistent with previous studies showing that many *mac* mutants are more susceptible to pathogen infection (Monaghan et al., 2009; Monaghan et al., 2010; Weihmann et al., 2012; Xu et al., 2012). Although the *mac7-1* hypo-DEGs are enriched in defense related GO terms, the genes show the smallest reduction magnitude compared to other *mac* mutants, which could be the reason why *mac7-1* is not susceptible to *P. syringae* infection as *prl1-2* and *mac3a mac3b* mutants are (Monaghan et al., 2009; Weihmann et al., 2012).

It has been suspected that MAC is also involved in splicing in plants, like its orthologs in human and yeast (Johnson et al., 2011; Koncz et al., 2012). Indeed,

our RNA-seq analyses uncovered intron retention defects in three MAC subunit mutants, thus linking MAC with pre-mRNA splicing. However, there were no significant correlations between intron retention and changes in gene expression. Genes with intron retention were up-regulated, down-regulated, or unchanged in the *mac* mutants. We also did not detect intron retention for intron-containing pri-miRNAs in *mac7-1*. It is likely that MAC has separate roles in RNA splicing and gene expression regulation (including the regulation of *MIR* genes). Thus, MAC has broad functions in nuclear RNA metabolism.

We speculate that a common theme of MAC's role in nuclear RNA metabolism is linking RNA processing to transcription (Figure 10). In yeast, the NTC promotes transcription elongation (Chanarat et al., 2011; Chanarat and Strasser, 2013). In both yeast and human, NTC or Prp19C associates with spliceosomes *in vivo*, although this association may entail NTC subcomplexes in human (Chanarat et al., 2012; De et al., 2015; Yan et al., 2015). In Arabidopsis, two MAC subunits, CDC5 and PRL1, have been shown to interact with Pol II *in vivo* (Zhang et al., 2013b; Zhang et al., 2014). IP-MS of an Arabidopsis spliceosome subunit also pulled down multiple subunits of MAC (Deng et al., 2016). Thus, it is possible that MAC promotes co-transcriptional splicing through its interactions with both Pol II and the spliceosome. Similarly, in miRNA biogenesis, MAC may promote co-transcriptional pri-miRNA processing through its interactions with both Pol II and HYL1.

METHODS

Plant materials and growth conditions

The *pSUC2:amiR-SUL* transgenic line is a gift from Dr. Detlef Weigel (de Felippes et al., 2011). *mac7-1* is a new allele isolated from our EMS mutagenesis screen with the *pSUC2:amiR-SUL* transgenic line. SALK_120944 (*mac7-2*) is a T-DNA insertion line obtained from the Arabidopsis Biological Resource Center (ABRC). The following published transgenic lines or mutants were used: *pMIR167a:GUS*, and *prl1-2* (SALK_008466) (Zhang et al., 2013b; Zhang et al.,

2014); *p35S:HYL1-YFP* (Qiao et al., 2015); *p35S:YFP* (Zhang et al., 2014); *mac3a mac3b* (*mac3a* is SALK_089300; *mac3b* is SALK_050811) and *prl1 prl2* (*prl1* is SALK_008466; *prl2* is SALK_075970) double mutants (Monaghan et al., 2009; Weihmann et al., 2012).

In *pMIR167a:GUS* or *p35S:HYL1-YFP* transgenic lines, the transgene was confirmed to be homozygous by Basta selection (Phosphinothricin 25mg/L, Gold Biotechnology) and by examining YFP signals under fluorescence microscopy, respectively, in multiple individuals. The *mac7-1* mutant was crossed into *pMIR167a:GUS* or *p35S:HYL1-YFP,* and F2 plants containing homozygous transgenes were confirmed in the F3 generation.

Genotyping primers used are listed in Supplemental Table 4. Plants were grown in a plant growth chamber at 23°C for 16h light (Cool white fluorescent lamps, 25-watt Sylvania 21942 FO25/741/ECO T8 linear tube) and 8 h dark cycles.

Mutagenesis screening and mapping of *MAC7*

The *amiR-SUL mac7-1* M2 mutant was backcrossed with the parental line *pSUC2:amiR-SUL*. Genomic DNA was extracted from 100 pooled F2 mutants and used in library construction. The library was paired-end sequenced on Illumina's HiSeq 2000 at ~30x coverage, and the reads were mapped to the TAIR 10 genome using the Burrows-Wheeler Alignment tool (BWA) (Li and Durbin, 2009). SamTools (Li et al., 2009) was used to identify EMS-induced single nucleotide polymorphisms (SNPs). The SNP calls generated by SamTools were processed by the Next-Generation EMS mutation mapping (NGM) website tools (Li et al., 2008). Two candidate mutations with 100% mutation rate were identified as EMS-typical C:G to T:A transitions that are predicted to cause nonsynonymous substitutions in the coding region of genes. Because one candidate mutation was only supported by three reads, we focused on the one in At2g38770 (*MAC7/EMB2765*), which was supported by 29 reads. A Derived Cleaved Amplified Polymorphic Sequences (dCAPS) marker was designed to

genotype this mutation (see Supplemental Table 4 for primers). The PCR products from wild type can be digested by *Eco*R1, whereas those from *mac7-1* could not. Linkage analysis was performed on 84 individual mutant plants in the F2 population of the backcross using this dCAPS marker to assess linkage between the mutation and the *mac7-1* mutant phenotype.

DNA constructs and complementation

The *MAC7* genomic region without the stop codon was amplified and cloned into pENTR/D-TOPO (Invitrogen) and then introduced to a modified pEarleyGate 301 vector (Earley et al., 2006) to generate *pMAC7:MAC7-mCherry* via LR reaction. The *MAC7* genomic region without the stop codon was amplified and cloned into the pMDC107 gateway vector (Curtis and Grossniklaus, 2003) to generate *pMAC7:MAC7-GFP* with the ClonTech In-Fusion HD Cloning Kit. The *MAC7* coding region was cloned and then introduced into the pEarleyGate104 vector (Earley et al., 2006) via pENTR/D-TOPO (Invitrogen) through LR reactions. The above plasmids were used to transform *mac7-1* plants through the *Agrobacterium*-mediated floral dip method. Primers used are listed in Supplemental Table 4.

Small RNA gel blotting, RT-PCR and quantitative RT-PCR

Total RNA from 2 to 3-week old seedlings (aerial part) or inflorescences was extracted with TRI reagent (Molecular Research Center). RNA gel blotting for detection of miRNAs was performed as described (Pall and Hamilton, 2008). 10 µg total RNA from aerial part of seedlings or inflorescences was used in RNA gel blotting. 5' end ^{32}P-labelled antisense DNA oligonucleotides were used to detect miRNAs. Oligonucleotide probes used are listed in Supplemental Table 4.

To perform RT-PCR, total RNA was first treated with DNase I (Roche) followed by reverse transcription using RevertAid Reverse Transcriptase (Thermo Fisher Scientific) with oligo-d(T) primers according to manufacturer's instructions. Quantitative RT-PCR was carried out in triplicate using iQ SYBRGreen Supermix

(BioRad) on the BioRad CFX96 system. Primers used are listed in Supplemental Table 4.

4

RNA half-life measurements

RNA half-life measurements were performed as described (Lidder et al., 2005) with minor modifications. 12-day-old Col and *mac7-1* whole seedlings were transferred from Murashige and Skoog medium (PhytoTechnology Lab) agar plates to 6-well-plates with 1/2 MS medium and incubated overnight. The next day, cordycepin (Sigma) was added to a final concentration of 0.6 mM and the seedlings were collected for RNA extraction at 0, 30, 90, and 120 min after cordycepin addition. RT-qPCR was then performed to determine pri-miRNA levels. *UBQ5* was used as an internal control.

Small RNA-seq library construction and data analysis

To construct small RNA libraries, the aerial parts of 12-day-old Col-0, *mac7-1, and prl1 prl2* seedlings grown on plates were harvested for total RNA extraction. Two biological replicates were included: plants grown on different plates under the same conditions were collected at the same time into two separate samples for RNA extraction and subsequent procedures. To isolate small RNAs from total RNA, 50 µg of total RNA from each sample was resolved on 15% urea-PAGE gel, and the 18–30-nt region was excised from the gel. Small RNAs were recovered by soaking the smashed gel in 0.3 M NaCl overnight, followed by ethanol precipitation. Small RNA libraries were constructed following instructions from the NEBNext Multiplex Small RNA Library Prep Set for Illumina (E7300). The libraries were sequenced on an Illumina Hiseq 2500 at the UC Riverside Genomics core facility.

Reads from small RNA-seq were first processed to remove the adaptor sequences by cutadapt (sequence: AGATCGGAA) (Martin, 2011). The reads were mapped to the TAIR10 genome using ShortStack with default parameters (Johnson et al., 2016). Normalization was performed by calculating the RPMR

(reads per million of 45S rRNA reads) value (Li et al., 2016). Only one biological replicate for Col-0 was included in this analysis since the other Col-0 sample had very few reads caused by unknown problems in library construction or sequencing.

RNA-seq library construction and data analysis

Polyadenylated RNA was isolated from total RNA extracted from 12-day-old seedlings (aerial part) of Col-0, *mac7-1*, *mac3a mac3b*, and *prl1 prl2* using the Magnetic mRNA Isolation Kit (New England Biolabs), with two biological replicates for each genotype. For the biological replicates, plants grown on different plates under the same conditions were collected at the same time into two separate samples for RNA extraction and subsequent procedures. RNA-seq libraries were prepared using NEBNext mRNA Library Prep Reagent Set for Illumina (New England Biolabs) and sequenced on an Illumina Hiseq 2500 platform to generate high-quality single-end reads of 101bp in length. Data analysis was performed with the pRNASeqTools pipeline (https://github.com/grubbybio/RNASeqTools). Firstly, the raw reads were aligned to the TAIR10 genome using HISAT2 (Kim et al., 2015). Secondly, exonic and intronic reads were classified and quantified as follows. Reads with five or more nucleotides overlapping with intronic regions (intronic regions defined in all isoforms, including splicing donor/acceptor sites) were counted as intronic reads, and reads that mapped exclusively to exonic regions were counted as exonic reads. Transcript levels were measured in reads per million total read counts. Differentially expressed genes were identified using DEseq2 with fold change of 1.5 and $p < 0.01$ as the parameters (Love et al., 2014). To identify genes that exhibited significantly higher levels of intron retention compared to Col-0, the intron/exon ratio was calculated as (I1 + I2)/2 over (E1+E2)/2 (I: Intronic reads, E: Exonic reads, 1: biological replicate 1, 2: biological replicate 2) while applying an abundance cutoff (raw intronic read number ≥ 2 and exonic read number ≥ 5). We only considered intron/exon (I/E) ratios between 0 to 1. The genes with intron retention were identified using DEseq2 based on values of $(I_{mutant}/E_{mutant})/(I_{Col\text{-}0}/E$

726 $_{Col-0}$) and fold changes ≥ 2 and FDR < 0.01 as parameters (Love et al., 2014).
727 SuperExactTest was employed to access the statistical significance of DEGs or
728 intron retention gene overlaps among *mac7-1*, *mac3a mac3b* and *prl1 prl2*
729 (Wang et al., 2015).
730
731 **Protein sequence alignment**
732 Database searching of MAC7 homologs was performed at National Center for
733 Biotechnology Information (NCBI) (www.ncbi.nlm.nih.gov/). Alignment of protein
734 sequence was performed with Muscle (Edgar, 2004) and the alignments were
735 edited with Jalview (Waterhouse et al., 2009).
736
737 **Antibody generation and immunoblotting**
738 To generate anti-MAC7 antibodies, a 5' portion of the coding region of *MAC7*
739 corresponding to the first 416 amino acids of the protein was amplified (primers
740 listed in Supplemental Table 4) and inserted into pMCSG7-His-MBP and
741 pSUMO-His vectors, respectively. The constructs were then transformed into the
742 *E. coli* strain BL21 for protein expression. The recombinant proteins were purified
743 by AKTA fast protein liquid chromatography (GE Healthcare) using the MBP-Trap
744 or His-Trap column. The purified MBP-tagged protein was used as antigens to
745 raise polyclonal antibodies in rabbits as described (Peterson et al., 2010). The
746 anti-serum was affinity-purified using a MAC7-SUMO-His conjugated column.
747 The purified antibodies were used in immunoblotting and IP-MS experiments.
748 Similar approach was employed to generate the anti-CH42/SUL antibody. The
749 full-length CH42 protein fused with SUMO-His tag was expressed and purified to
750 immunize two rabbits. Affinity purified antibodies were used in immunoblotting
751 analysis.
752
753 Other antibodies used in immunoblotting experiments include anti-GFP (Roche,
754 Cat. No. 11814460001), anti-AGO1 (Agrisera, AS09 527), anti-HYL1 (Agrisera,
755 AS06 136), anti-SERRATE (Agrisera, AS06 136), anti-DCL1 (Agrisera, AS12
756 2102), and anti-Tubulin (Sigma, T9026).

Proteomic Analysis

Total proteins from 12-day-old seedlings were extracted and immunoprecipitated with indicated antibodies. The IP products were resolved in SDS-PAGE. The antibody bands were removed and the samples were subjected to mass spectrometry as described (Sleat et al., 2006; Deng et al., 2016). Two biological replicated were performed, and the identified interacting proteins were those represented by peptides with high hits from both biological replicates.

Transient expression of fluorescent fusion proteins in tobacco leaf epidermal cells

The CDS of *HYL1* was amplified and cloned into pENTR/D-TOPO (Invitrogen) and then introduced into the pGWB661 gateway vector (Nakamura et al., 2010) to generate *p35S:TagRFP-HYL1. p35S:DCL1-YFP* in the pEG101 vector was from a published study (Zhang et al., 2013b). The generation of *p35S:YFP-MAC7* was described above. *Agrobacterium* GV3101::mp90 transformed with *p35S:YFP-MAC7, p35S:DCL1-YFP*, or *p35S:TagRFP-HYL1* was used to infiltrate tobacco leaves as described (Sparkes et al., 2006). The expression of fluorescent fusion proteins was observed using a Lecia SP5 confocal laser-scanning microscope.

BiFC analysis

BiFC analysis was performed as described (Walter et al., 2004). Paired cCFP and nVenus constructs were co-infiltrated into *Nicotiana benthamiana* leaves. After 48 h, YFP signals and chlorophyll auto fluorescence signals were excited at 488 nm and detected by Olympus Fluoview 500 confocal microscopy with a narrow band pass filter (BA505–525 nm).

Histochemical GUS assay and Alexander's staining of pollen

GUS staining was performed as described (Kim et al., 2011). Briefly, 12-day-old seedlings from Col-0 and *mac7-1* harboring a homozygous *pMIR167a:GUS*

transgene were vacuum infiltrated for 10 min and then incubated in GUS staining solution at 37°C for several hours until blue color became visible. Tissue clearing was performed with 70% ethanol for 1 to 2 days before imaging.

Alexander's staining of pollen was performed as described (Peterson et al., 2010). The stained pollen grains were observed under a microscope equipped with a charge-coupled device camera (Olympus).

Accession numbers

Genes referred to in this study correspond to the following Arabidopsis Genome Initiative locus identifiers: *MAC7/EMB2765*, AT2G38770; *SUL/CHLORINA42*, AT4G18480; *MICRORNA156A*, AT2G25095; *MICRORNA159A*, AT1G73687; *MICRORNA163* AT1G66725; *MICRORNA166A* AT2G46685; *MICRORNA167A*, AT3G22886; *MICRORNA168A*, AT4G19395; *MICRORNA171A*, AT3G51375; *MICRORNA172A*, AT2G28056; *MICRORNA390B*, AT5G58465; *MICRORNA394B*, AT1G76135; *MICRORNA396B*, AT5G35407; *MICRORNA397A*, AT4G05105; *ACTIN8*, AT1G49240; *UBIQUITIN5*, AT3G62250; *TUBULIN3*, AT5G62700; *EIF4A1*, AT3G13920; *IPP2*, AT3G02780 ;*CDC5*, AT1G09770; *PRL1*, AT4G15900; *PRL2*, AT3G16650; *MAC3A*, AT1G04510; *MAC3B*, AT2G33340 ; *HYL1*, AT1G09700; *DCL1*, AT1G01040; *SERRATE*, AT2G27100; *AGO1*, AT1G48410; *HEN1*, AT4G20910; *MYB33* AT5G06100; *SCL6-IV* AT4G00150; *SPL10* AT1G27370; *MYB65*, AT3G11440; *ARF17*, AT1G77850; *ARF8*, AT5G37020; *SCL6-III* At3g60630; *CUC2* AT5G53950.

Protein sequences of MAC7 homologs in other species correspond to the following NCBI references: *Homo sapiens* intron-binding protein aquarius, NP_055506.1; *Mus musculus* intron-binding protein aquarius, NP_033832.2; *Danio rerio* intron-binding protein aquarius, NP_956758; *Drosophila melanogaster* CG31368, NP_996198.2; *Caenorhabditis elegans* EMB-4, NP_001256831.1; *Schizosaccharomyces pombe Cwf11*, NP_595360.1.

RNA sequencing data are available from NCBI Gene Expression Omnibus (GEO) under the following reference numbers: Col-0_1, GSM2585832; Col-0_2, GSM2585833; mac7-1_1, GSM2585834; mac7-1_2, GSM2585835; mac3amac3b_1, GSM2585836; mac3amac3b_2, GSM2585837; prl1prl2_1, GSM2585838; prl1prl2_2, GSM2585839; Col-0_1_sRNA GSM2771029; mac7-1_1_sRNA GSM2771030; mac7-1_2_sRNA GSM2771031, prl1 prl2_1_sRNA GSM2771032, prl1 prl2_2_sRNA GSM2771033.

Supplemental Data

Supplemental Figure 1. The *mac7-1* mutant shows pleiotropic developmental phenotypes. (Supports Figure 1.)

Supplemental Figure 2. Reduced miRNA accumulation in both seedlings and inflorescences in the *mac7-1* mutant. (Supports Figure 1.)

Supplemental Figure 3. The *mac7-1* mutation in the Col-0 background does not affect *CH42* expression at either mRNA or protein levels. (Supports Figure 1.)

Supplemental Figure 4. The *MAC7* T-DNA insertion line, *mac7-2*, is a single-locus, recessive embryo-lethal mutant. (Supports Figure 2.)

Supplemental Figure 5. Amino acid sequence alignment of *Arabidopsis* MAC7 orthologs. (Supports Figure 2.)

Supplemental Figure 6. MAC7 does not affect the expression of the endogenous *SUC2* gene or key miRNA biogenesis factors. (Supports Figure 3.)

Supplemental Figure 7. MAC7 shows dispersed distribution in the nucleoplasm, while DCL1 and HYL1 concentrate in dicing bodies in the nuclei. (Supports Figure 4.)

Supplemental Figure 8. Growth of virulent *P.s.m.* ES4326 at 0 and 3 days post-inoculation. (Supports Figure 7.)

Supplemental Figure 9. Intron-containing pri-miRNAs show similar splicing patterns in Col-0 and *mac7-1*. (Supports Figure 8 and Figure 9.)

Supplemental Table 1. Phenotypic and genotypic segregation in the progeny of selfed heterozygous *mac7-2* plants. The *mac7-2* allele harbors a T-DNA insertion (SALK_129044).

Supplemental Table 2. MAC7-associated proteins identified by immunoprecipitation followed by mass spectrometry.

Supplemental Table 3. HYL1-associated MAC subunits identified by immunoprecipitation followed by mass spectrometry.

Supplemental Table 4. Oligonucleotide sequences.

Supplemental Data Set 1. Levels of 21-nt small RNAs, 24-nt small RNAs and miRNAs in Log_2(reads per million of 45S rRNA reads) in Col-0, *mac7-1*, and *prl1 prl2* as determined by small RNA-seq.

Supplemental Data Set 2. Differentially expressed genes in *mac7-1, mac3a mac3b,* and *prl1 prl2* mutants as determined by RNA-seq analysis.

Supplemental Data Set 3. Gene Ontology analysis of overlapped DEGs among *mac7-1, mac3a mac3b,* and *prl1 prl2* mutants.

Supplemental Data Set 4. Intron/exon ratio in Col-0, *mac7-1*, *mac3a mac3b*, and *prl1 prl2*. Genes with significantly higher intron/exon ratio as well as differentially expressed genes in the mutants are shown.

Supplemental Data Set 5. Relative expression levels of genes that passed the intronic and exonic read counts filter in *mac7-1*, *mac3a mac3b*, and *prl1 prl2* compared to Col-0, and relative expression levels of genes with intron retention defects in indicated mutants compared to Col-0.

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AUTHOR CONTRIBUTIONS

T.J. and X.C. designed the experiments. T.J., B.Z., S.L., Y.Z., K.J., B.Y. and X.L. performed the experiments. T.J., C.Y. and L.Z. analyzed the data. T.J., C.Y., and X.C. wrote the manuscript.

FIGURE LEGENDS

Figure 1. A silencing suppressor mutant *mac7-1* exhibits pleiotropic phenotypes and compromised miRNA accumulation.

(A) Differences in vein-centered bleaching in 3- to 4-week-old *pSUC2:amiR-SUL* (*amiR-SUL*) and *amiR-SUL mac7-1* seedlings. (B and C) Morphological phenotypes of *amiR-SUL*, *amiR-SUL mac7-1*, Col-0 and *mac7-1* plants. Images of rosettes and roots were taken from 2- to 3-week-old and 1-week old plants, respectively. (D) RNA gel blotting analysis of miRNAs from *amiR-SUL* and *amiR-SUL mac7-1* inflorescences. The miRNA signals were quantified and normalized to those of U6, and values were relative to *amiR-SUL* (set to 1). Two biological replicates of inflorescences collected from plants grown separately but under the same conditions were processed and shown. (E) Global abundance of 21-nt and 24-nt small RNAs and miRNAs in Col-0 and *mac7-1* as determined by small RNA-seq. Small RNA libraries were generated from 12-day-old seedlings growing on MS plates. The normalization of small RNAs was against 45S rRNA reads and abundance was expressed as RPMR (reads per million of 45S rRNA reads), and $\log_2$ ratios of *mac7-1*/Col-0 were plotted. Asterisks indicate that the mean is significantly below 0 (Wilcoxon test, $p < 2.2e\text{-}16$). (F) Determination of miRNA target mRNA levels in *amiR-SUL* and *amiR-SUL mac7-1* using 12-day-old seedlings by quantitative RT-PCR (RT-qPCR). The housekeeping gene *IPP2* was included as a control. Expression levels were normalized to those of *UBIQUITIN5 (UBQ5)* and compared with those in *amiR-SUL* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks indicate significant difference between Col-0 and *mac7-1* (t-test, $p < 0.05$).

Figure 2. A point mutation in *MAC7* is responsible for the morphological and molecular phenotypes in the *mac7-1* mutant.

(A) A diagram of the MAC7 protein showing various domains, the predicted nuclear localization signal (NLS), and the E1131K mutation in the *mac7-1* mutant. A sequence alignment of MAC7 and its orthologs in the region containing the E1131K mutation in the *mac7-1* mutant is also shown. Abbreviations for species are as follows: *Arabidopsis thaliana* (A.t.), *Homo sapiens* (H.s.), *Mus musculus* (M.m.), *Danio rerio* (D.r.), *Drosophila melanogaster* (D.m.), *Caenorhabditis elegans* (C.e.), and *Schizosaccharomyces pombe* (S.p.). N: N-terminus; ARM: armadillo domain; RecA: RecA-like domains; C: C-terminus. The point mutation site is labeled by a triangle. (B) Morphological phenotypes of 3- to 4-week-old seedlings of the indicated genotypes. *pMAC7:MAC7-GFP* and *pMAC7:MAC7-mCherry* were transformed into *amiR-SUL mac7-1* and *mac7-1*, respectively. (C and D) RNA gel blotting analysis of miRNAs from Col-0, *mac7-1*, and the complementation line *mac7-1 pMAC7:MAC7-mCherry* using inflorescences (C) and 12-day-old seedlings (D). The miRNA signals were quantified as described in Figure 1D. RNA markers (NEB, N2102S) shown in (D) were resolved in the same gel as miRNAs and probed separately by a DNA probe complementary to the marker sequences.

Figure 3. *MAC7* promotes pri-miRNA production.

(A) Determination of pri-miRNA levels in *amiR-SUL* and *amiR-SUL mac7-1* inflorescences by RT-qPCR. The housekeeping gene *IPP2* was included as a control. Expression levels were normalized to those of *UBQUITIN5 (UBQ5)* and compared with those in *amiR-SUL* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks indicate significant difference between *amiR-SUL* and *amiR-SUL mac7-1* (t-test, $p < 0.05$). (B) Representative GUS staining images of *pMIR167a:GUS* and *mac7-1 pMIR167a:GUS* seedlings. The transcript levels of *GUS* and endogenous *pri-miR167a* in *pMIR167a:GUS* and *mac7-1 pMIR167a:GUS* seedlings were determined by RT-qPCR. Expression levels were normalized to those of *UBIQUITIN5 (UBQ5)* and compared with those in *pMIR167a:GUS* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks, t-test $p < 0.05$. (C) Half-life measurements for pri-amiR-SUL, pri-miR167a, pri-miR172a and *EIF4A* mRNA. Two-week-old *amiR-SUL* and *amiR-SUL mac7-1* seedlings were treated with 0.6 mM cordycepin and harvested at various time points. RT-qPCR was performed to determine the levels of various pri-miRNAs and *EIF4A* mRNA. *UBQ5* served as an internal control. Values at time 0 were set to 1. Error bars indicate standard deviation from three technical replicates. Two biological replicates were performed and similar results were obtained.

Figure 4. MAC7 is a nuclear protein associated with other MAC subunits.
(A) Subcellular localization of N-terminal (*p35S:YFP-MAC7*) or C-terminal (*pMAC7:MAC7-GFP*) fluorescent protein tagged MAC7 in young leaves of *Arabidopsis* transgenic lines. Nuclei were stained with DAPI and pseudo-colored in cyan. Enlarged nuclei are shown in the insets. (B) MAC subunits identified from both MAC7 and MOS4 immunoprecipitation followed by mass spectrometry analysis (IP-MS). (C) BiFC analysis of MAC7 with CDC5, PRL1, HYL1 and DCL1. Paired cCFP- and nVenus-fusion proteins were co-infiltrated into tobacco leaves. The BiFC signal (YFP) was detected at 48 h after infiltration by confocal microscopy, and was pseudo-colored in green. Magenta: auto fluorescence of chlorophyll.

Figure 5. The MAC subunit genes *MAC3a*, *MAC3b*, *PRL1*, and *PRL2* also promote miRNA biogenesis.
(A) Morphological phenotypes of 4 to 5-week old plants of the indicated genotypes. (B) MiRNA levels in wild type (Col-0) and indicated mutants as determined by northern blotting. The miRNA signals were quantified and normalized to those of U6, and values were relative to Col-0 (arbitrarily set to 1). The RNA used in northern blotting was extracted from the aerial parts of 12-day-old seedlings growing on MS agar plates. (C) Global abundance of 21-nt and 24-nt small RNAs and miRNAs in Col-0 and *prl1 prl2* as determined by small RNA-seq. Small RNA libraries were generated from 12-day-old seedlings growing on MS plates. The normalization of small RNAs was against 45S rRNA reads and abundance was expressed as RPMR (reads per million of 45S rRNA reads), and $\log_2$ ratios of *prl1 prl2*/Col-0 were plotted. Asterisks indicate that the mean is significantly below 0 (Wilcoxon test, $p < 2.2e\text{-}16$). (D) Pri-miRNA levels in plants

of the indicated genotypes as determined by RT-qPCR. The housekeeping gene *IPP2* was included as a control. *UBQ5* was used as an internal control and values in Col-0 were set to 1. Error bars indicate standard deviation from three technical replicates, and asterisks indicate significant difference between Col-0 and the mutants (t-test, $p < 0.05$).

Figure 6. MAC7 affects HYL1 subcellular localization.
(A) Images of nuclei in root cells of 7 to 10-day-old seedlings of the indicated genotypes. Images (2) and (4) show five times enlarged images cropped from (1) and (3), respectively. Dicing bodies are indicated by triangles. (B) The percentage of cells containing HYL1-positive dicing bodies in wild type and *mac7-1*. The quantification was performed by observing more than 1000 cells from 27 roots for each genotype. The asterisk indicates significant difference between the samples (t-test, $p < 0.05$).

Figure 7. Down-regulated genes in the *mac* mutants are significantly related to stress responses, while up-regulated genes are involved in various biosynthetic and metabolic processes.
(A) GO enrichment analysis of 189 commonly down-regulated genes in *mac7-1, mac3a mac3b* and *prl1 prl2*. The degree of overlap among down-regulated genes in *mac7-1*, *mac3a mac3b* and *prl1 prl2* is shown in the Venn diagram. (B) GO enrichment analysis of 222 commonly up-regulated genes *mac7-1, mac3a mac3b* and *prl1 prl2*. The degree of overlap among up-regulated genes in *mac7-1*, *mac3a mac3b* and *prl1 prl2* is shown in the Venn diagram.

Figure 8. MAC subunits affect pre-mRNA splicing.
(A) Box plot of intron/exon ratios in the indicated genotypes. All genes that pass an abundance filter in expression were included in this analysis. Asterisks indicate significant difference between the mutant and wild type (Col-0) (Wilcoxon test, $p < 2.2e\text{-}16$). (B) The intron/exon ratio per gene in Col-0 vs. the indicated mutants. Black dots represent all genes with raw intronic read number ≥ 2, exonic read number ≥ 5, and final intron/exon ratio between 0 to 1. Red dots represent genes with significantly higher intron/exon ratio relative to wild type (fold change ≥ 2 and FDR < 0.01). (C) A Venn diagram showing the degree of overlap among genes with intron retention defects in *mac7-1*, *mac3a mac3b* and *prl1 prl2*. (D) Examples of genes with intron retention defects. RPKM: Reads Per Kilobase per Million mapped reads. Two biological replicates for each genotype are shown. The rectangles mark introns with higher retention in the mutants.

Figure 9. MAC affects pre-mRNA splicing and gene expression independently.
(A) Volcano plots illustrating fold changes of gene expression levels in the indicated mutants compared to Col-0. Thresholds for fold change of 1.5 and $p \leq 0.01$ are shown in the plots as gray dashed lines. Black dots and red dots represent the same genes as in Figure 8B. (B) Box plots of intron/exon ratios in total genes, up-regulated and down-regulated genes in the indicated genotypes.

Only genes that pass an abundance filter (raw intronic read number ≥ 2 and exonic read number ≥ 5) were included in this analysis. Asterisks indicate significant difference between the mutant and wild type (Col-0) (Wilcoxon test, $p < 2.2e-16$).

Figure 10. A model for MAC's functions in miRNA biogenesis and pre-mRNA processing.

MAC affects miRNA biogenesis through influencing Pol II transcription and interacting with the pri-miRNA processing factor HYL1. MAC also plays a role in pre-mRNA splicing through interactions with the spliceosome. MAC seems to have separate roles in miRNA biogenesis and RNA splicing but a common theme appears to be co-transcriptional RNA processing.

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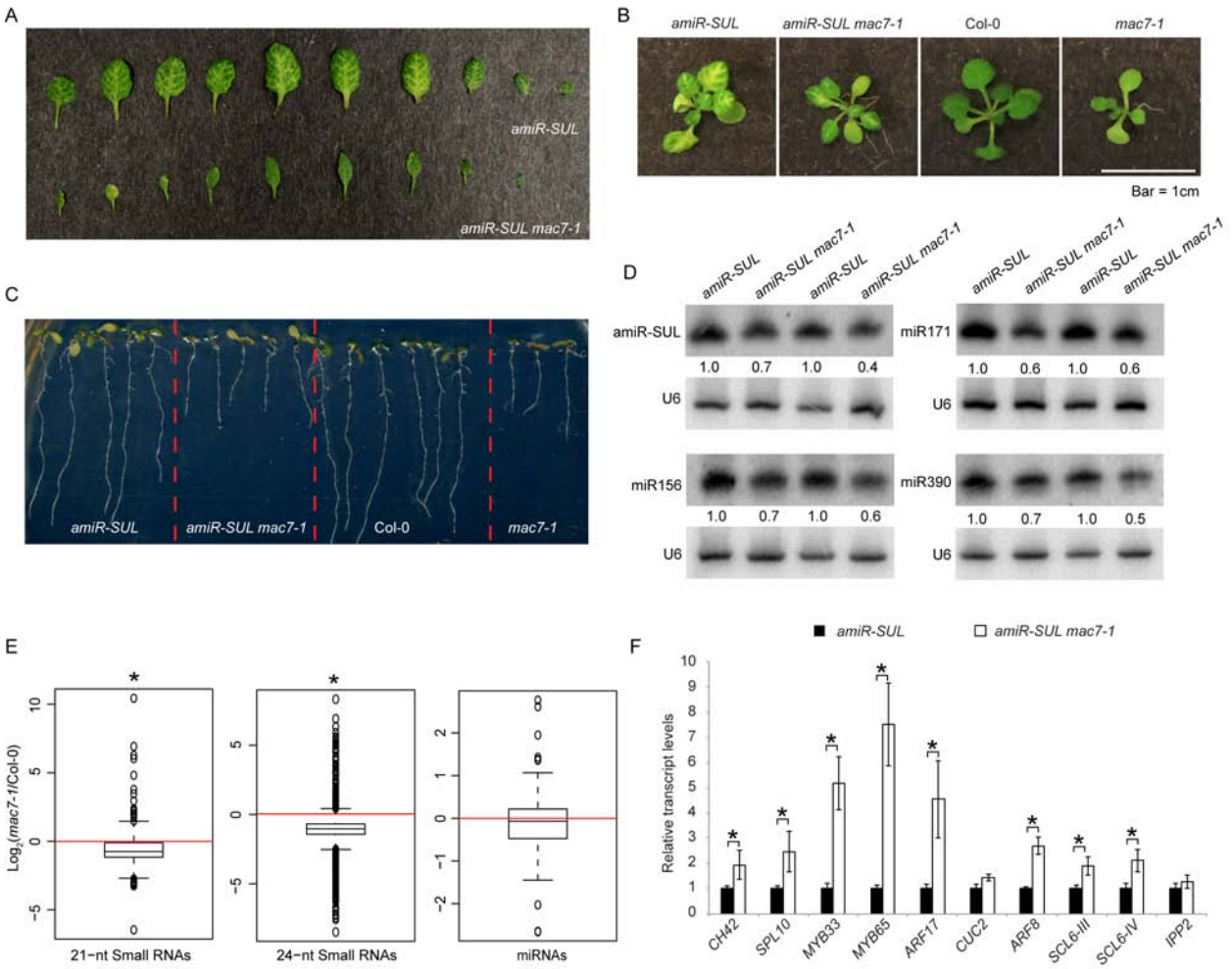

Figure 1. A silencing suppressor mutant *mac7-1* exhibits pleiotropic phenotypes and compromised miRNA accumulation.

(A) Differences in vein-centered bleaching in 3 to 4-week old *pSUC2:amiR-SUL* (*amiR-SUL*) and *amiR-SUL mac7-1* seedlings. (B and C) Morphological phenotypes of *amiR-SUL*, *amiR-SUL mac7-1*, Col-0 and *mac7-1* plants. Images of rosettes and roots were taken from 2 to 3-week old and 1-week old plants, respectively. (D) Northern blotting analysis of miRNAs from *amiR-SUL* and *amiR-SUL mac7-1* inflorescences. The miRNA signals were quantified and normalized to those of U6, and values were relative to *amiR-SUL* (arbitrarily set to 1). Two biological replicates of inflorescences collected from plants grown separately but under the same conditions were processed and shown. (E) Global abundance of 21-nt and 24-nt small RNAs and miRNAs in Col-0 and *mac7-1* as determined by small RNA-seq. Small RNA libraries were generated from 12-day-old seedlings growing on MS plates. The normalization of small RNAs was against 45S rRNA reads and abundance was expressed as RPMR (reads per million of 45S rRNA reads), and $\log_2$ ratios of *mac7-1*/Col-0 were plotted. Asterisks indicate that the mean is significantly below 0 (Wilcoxon test, $p < 2.2e\text{-}16$). (F) Determination of miRNA target mRNA levels in *amiR-SUL* and *amiR-SUL mac7-1* using 12-day-old seedlings by quantitative RT-PCR (RT-qPCR). The housekeeping gene *IPP2* was included as a control. Expression levels were normalized to those of *UBQUITIN5 (UBQ5)* and compared with those in *amiR-SUL* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks indicate significant difference between Col-0 and *mac7-1* (t-test, $p < 0.05$).

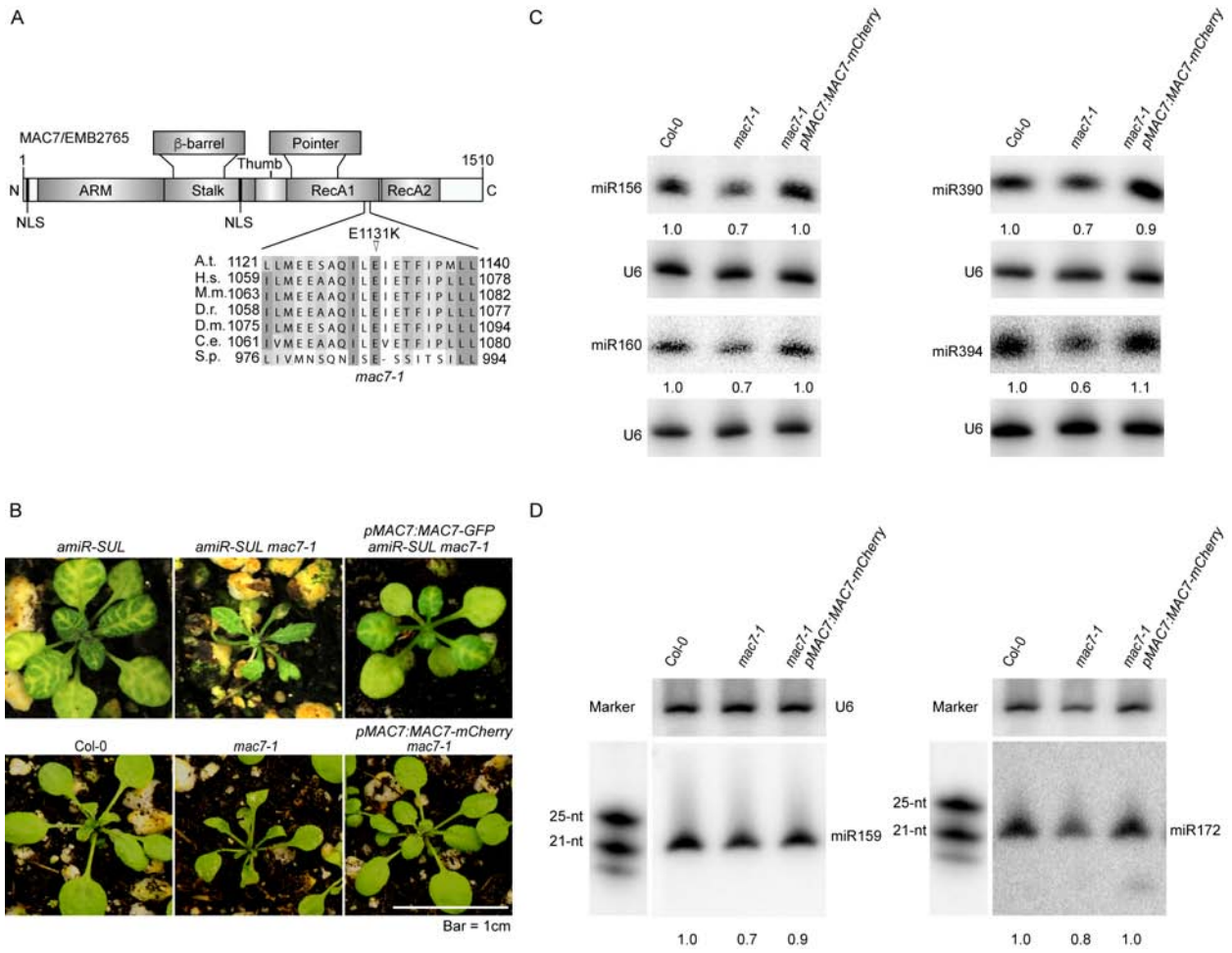

Figure 2. A point mutation in *MAC7* is responsible for the morphological and molecular phenotypes in the *mac7-1* mutant.

(A) A diagram of the MAC7 protein showing various domains, the predicted nuclear localization signal (NLS), and the E1131K mutation in the *mac7-1* mutant. A sequence alignment of MAC7 and its orthologs in the region containing the E1131K mutation in the *mac7-1* mutant is also shown. Abbreviations for species are as follows: *Arabidopsis thaliana* (A.t.), *Homo sapiens* (H.s.), *Mus musculus* (M.m.), *Danio rerio* (D.r.), *Drosophila melanogaster* (D.m.), *Caenorhabditis elegans* (C.e.), and *Schizosaccharomyces pombe* (S.p.). N: N-terminus; ARM: armadillo domain; RecA: RecA-like domains; C: C-terminus. The point mutation site is labeled by a triangle. (B) Morphological phenotypes of 3 to 4-week old seedlings of the indicated genotypes. *pMAC7:MAC7-GFP* and *pMAC7:MAC7-mCherry* were transformed into *amiR-SUL mac7-1* and *mac7-1*, respectively. (C and D) Northern blotting analysis of miRNAs from Col-0, *mac7-1*, and the complementation line *mac7-1 pMAC7:MAC7-mCherry* using inflorescences (C) and 12-day-old seedlings (D). The miRNA signals were quantified as described in Figure 1D. RNA markers (NEB, N2102S) shown in (D) were resolved in the same gel as miRNAs and probed separately by a DNA probe complementary to the marker sequences.

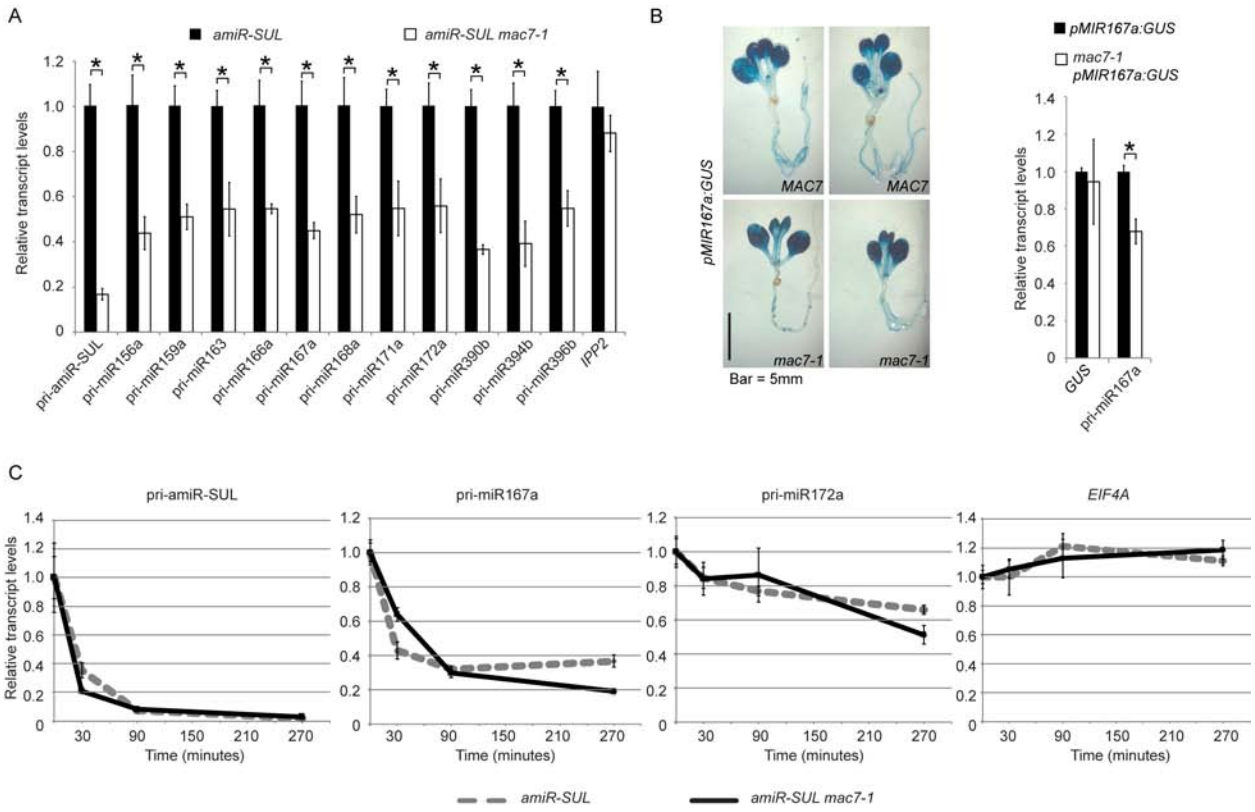

Figure 3. *MAC7* promotes pri-miRNA production.
(A) Determination of pri-miRNA levels in *amiR-SUL* and *amiR-SUL mac7-1* inflorescences by RT-qPCR. The housekeeping gene *IPP2* was included as a control. Expression levels were normalized to those of *UBQUITIN5 (UBQ5)* and compared with those in *amiR-SUL* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks indicate significant difference between *amiR-SUL* and *amiR-SUL mac7-1* (t-test, $p < 0.05$). (B) Representative GUS staining images of *pMIR167a:GUS* and *mac7-1 pMIR167a:GUS* seedlings. The transcript levels of *GUS* and endogenous *pri-miR167a* in *pMIR167a:GUS* and *mac7-1 pMIR167a:GUS* seedlings were determined by RT-qPCR. Expression levels were normalized to those of *UBQUITIN5 (UBQ5)* and compared with those in *pMIR167a:GUS* (set to 1). Error bars indicate standard deviation from three technical replicates. Asterisks, t-test $p < 0.05$. (C) Half-life measurements for pri-amiR-SUL, pri-miR167a, pri-miR172a and *EIF4A* mRNA. Two-week-old *amiR-SUL* and *amiR-SUL mac7-1* seedlings were treated with 0.6 mM cordycepin and harvested at various time points. RT-qPCR was performed to determine the levels of various pri-miRNAs and *EIF4A* mRNA. *UBQ5* served as an internal control. Values at time 0 were set to 1. Error bars indicate standard deviation from three technical replicates. Two biological replicates were performed and similar results were obtained.

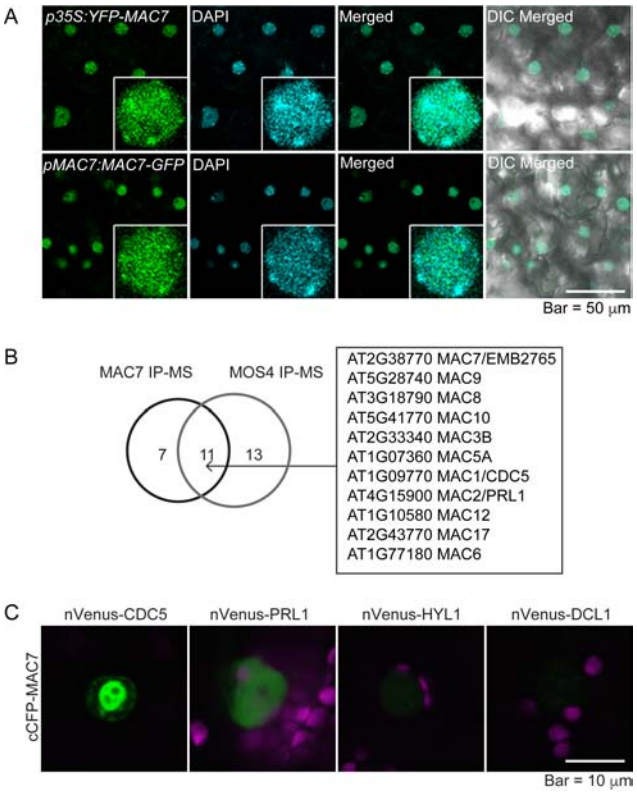

Figure 4. MAC7 is a nuclear protein associated with other MAC subunits.
(A) Subcellular localization of N-terminal (*p35S:YFP-MAC7*) or C-terminal (*pMAC7:MAC7-GFP*) fluorescent protein tagged MAC7 in young leaves of *Arabidopsis* transgenic lines. Nuclei were stained with DAPI and pseudo-colored in cyan. Enlarged nuclei are shown in the insets. (B) MAC subunits identified from both MAC7 and MOS4 immunoprecipitation followed by mass spectrometry analysis (IP-MS). (C) BiFC analysis of MAC7 with CDC5, PRL1, HYL1 and DCL1. Paired cCFP- and nVenus-fusion proteins were co-infiltrated into tobacco leaves. The BiFC signal (YFP) was detected at 48 h after infiltration by confocal microscopy, and was pseudo-colored in green. Magenta: auto fluorescence of chlorophyll.

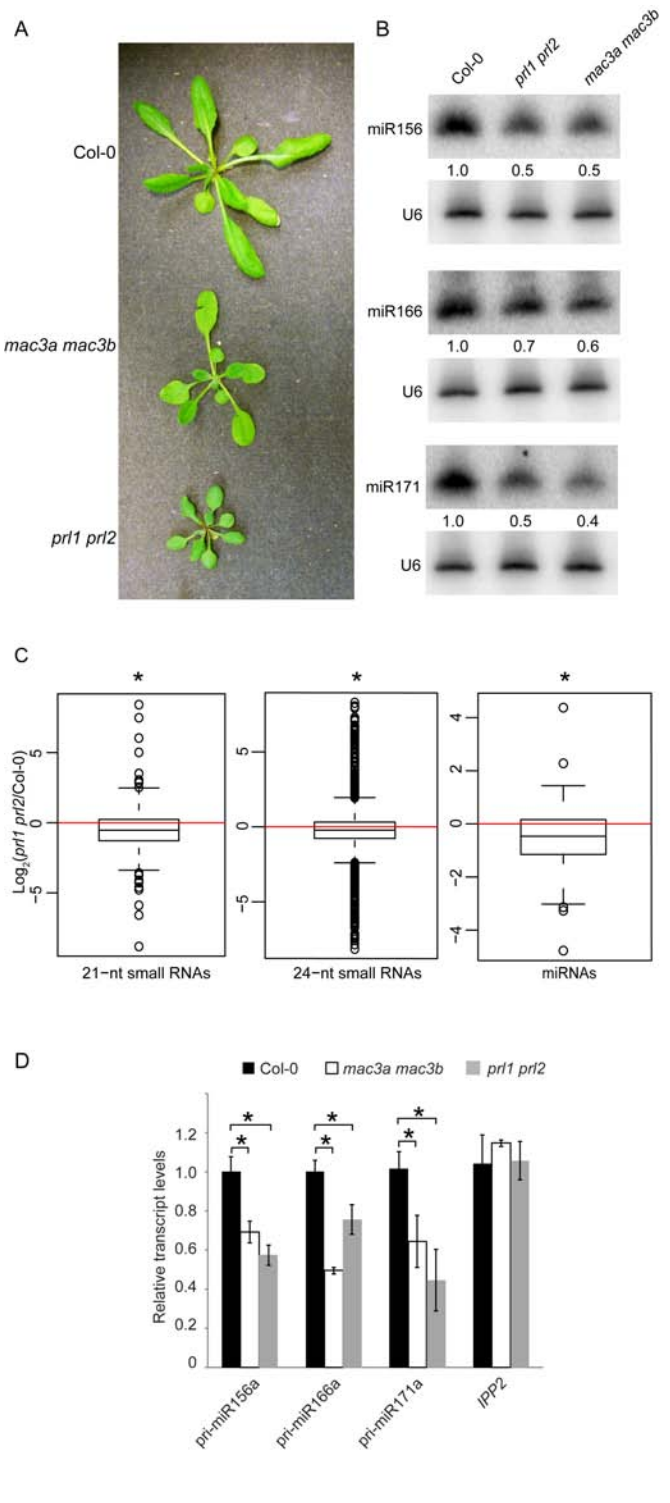
A
Col-0
mac3a mac3b
prl1 prl2
B
Col-0
prl1 prl2
mac3a mac3b
miR156
1.0
0.5
0.5
U6
miR166
1.0
0.7
0.6
U6
miR171
1.0
0.5
0.4
U6
C
Log2(prl1 prl2/Col-0)
21−nt small RNAs
24−nt small RNAs
miRNAs
D
Col-0
mac3a mac3b
prl1 prl2
Relative transcript levels
pri-miR156a
pri-miR166a
pri-miR171a
IPP2

Figure 5. The MAC subunit genes *MAC3a*, *MAC3b*, *PRL1*, and *PRL2* also promote miRNA biogenesis.

(A) Morphological phenotypes of 4 to 5-week old plants of the indicated genotypes. (B) MiRNA levels in wild type (Col-0) and indicated mutants as determined by northern blotting. The miRNA signals were quantified and normalized to those of U6, and values were relative to Col-0 (arbitrarily set to 1). The RNA used in northern blotting was extracted from the aerial parts of 12-day-old seedlings growing on MS agar plates. (C) Global abundance of 21-nt and 24-nt small RNAs and miRNAs in Col-0 and *prl1 prl2* as determined by small RNA-seq. Small RNA libraries were generated from 12-day-old seedlings growing on MS plates. The normalization of small RNAs was against 45S rRNA reads and abundance was expressed as RPMR (reads per million of 45S rRNA reads), and $\log_2$ ratios of *prl1 prl2*/Col-0 were plotted. Asterisks indicate that the mean is significantly below 0 (Wilcoxon test, $p < 2.2e-16$). (D) Pri-miRNA levels in plants of the indicated genotypes as determined by RT-qPCR. The housekeeping gene *IPP2* was included as a control. *UBQ5* was used as an internal control and values in Col-0 were set to 1. Error bars indicate standard deviation from three technical replicates, and asterisks indicate significant difference between Col-0 and the mutants (t-test, $p < 0.05$).

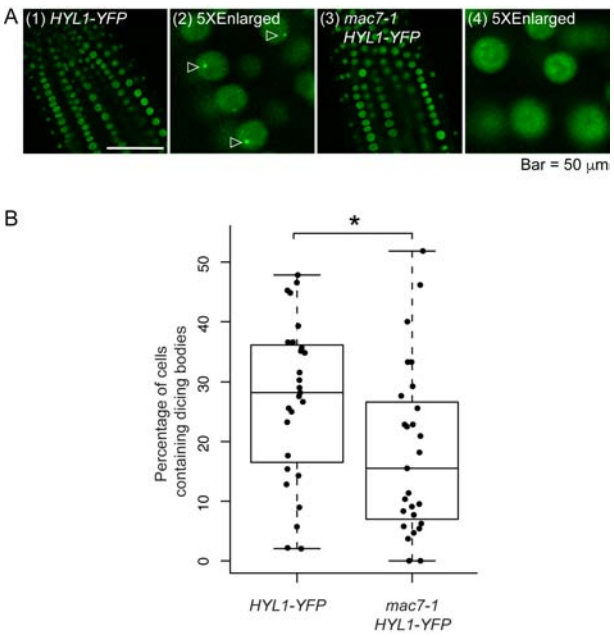

Figure 6. MAC7 affects HYL1 subcellular localization.
(A) Images of nuclei in root cells of 7 to 10-day-old seedlings of the indicated genotypes. Images (2) and (4) show five times enlarged images cropped from (1) and (3), respectively. Dicing bodies are indicated by triangles. (B) The percentage of cells containing HYL1-positive dicing bodies in wild type and *mac7-1*. The quantification was performed by observing more than 1000 cells from 27 roots for each genotype. The asterisk indicates significant difference between the samples (t-test, $p < 0.05$).

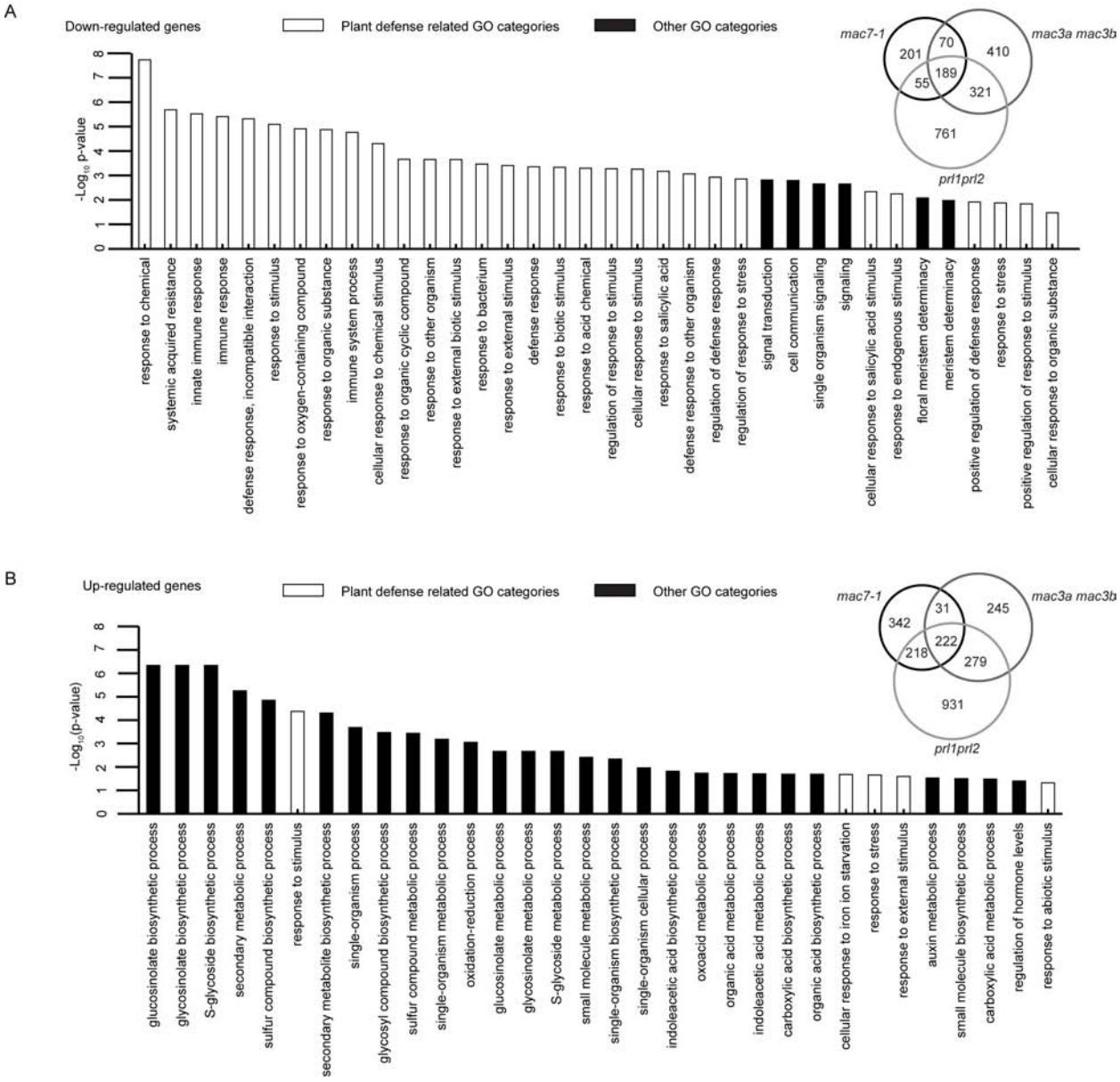

Figure 7. Down-regulated genes in the *mac* mutants are significantly related to stress responses, while up-regulated genes are involved in various biosynthetic and metabolic processes.

(A) GO enrichment analysis of 189 commonly down-regulated genes in *mac7-1, mac3a mac3b* and *prl1 prl2*. The degree of overlap among down-regulated genes in *mac7-1*, *mac3a mac3b* and *prl1 prl2* is shown in the Venn diagram. (B) GO enrichment analysis of 222 commonly up-regulated genes *mac7-1, mac3a mac3b* and *prl1 prl2*. The degree of overlap among up-regulated genes in *mac7-1*, *mac3a mac3b* and *prl1 prl2* is shown in the Venn diagram.

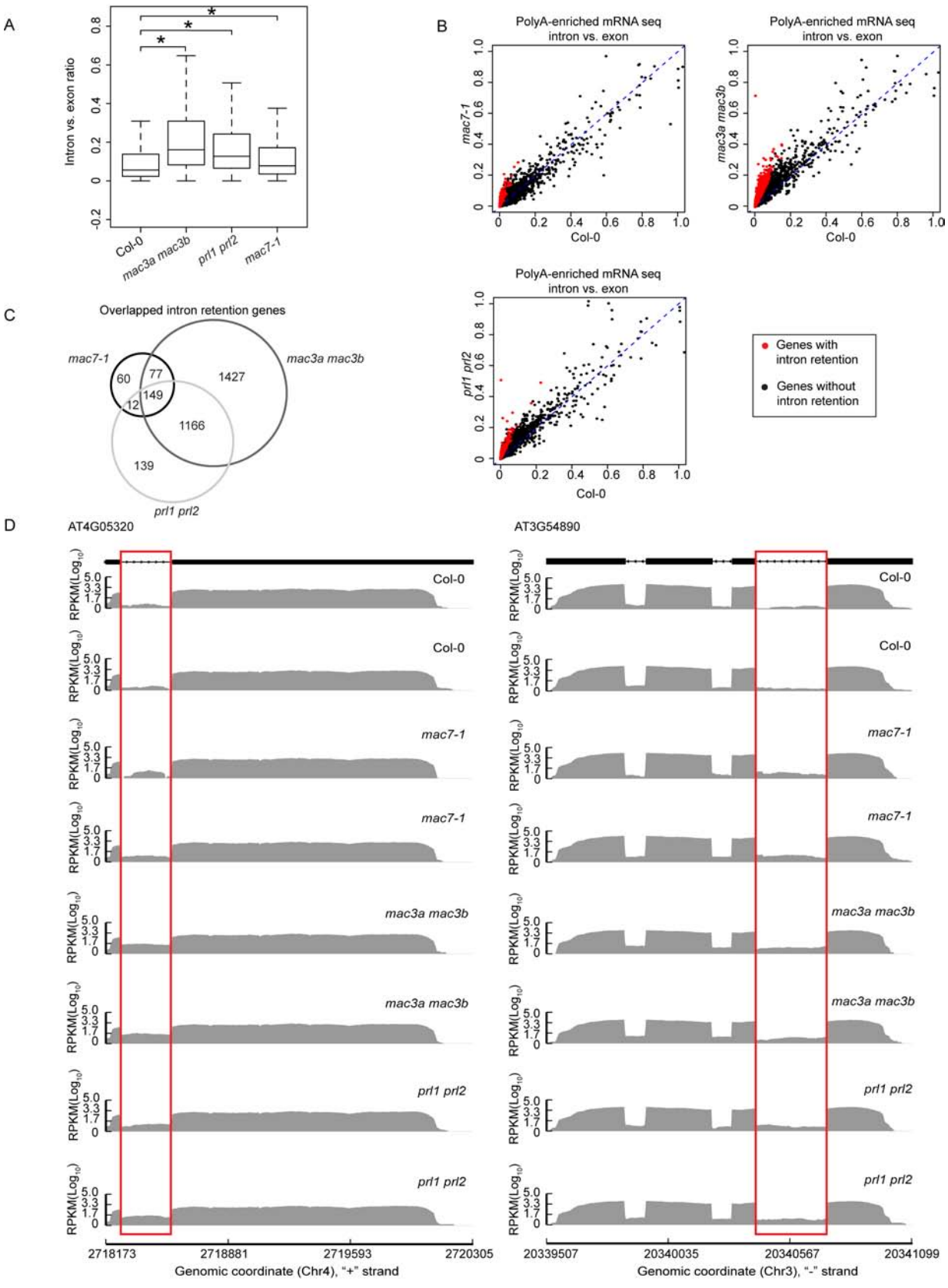
A
Intron vs. exon ratio
Col-0
mac3a mac3b
prl1 prl2
mac7-1
B
PolyA-enriched mRNA seq
intron vs. exon
mac7-1
mac3a mac3b
prl1 prl2
Col-0
Genes with intron retention
Genes without intron retention
C
Overlapped intron retention genes
mac7-1
mac3a mac3b
prl1 prl2
60
77
1427
12
149
1166
139
D
AT4G05320
AT3G54890
RPKM(Log10)
Col-0
mac7-1
mac3a mac3b
prl1 prl2
2718173
2718881
2719593
2720305
Genomic coordinate (Chr4), "+" strand
20339507
20340035
20340567
20341099
Genomic coordinate (Chr3), "-" strand

Figure 8. MAC subunits affect pre-mRNA splicing.
(A) Box plot of intron/exon ratios in the indicated genotypes. All genes that pass an abundance filter in expression were included in this analysis. Asterisks indicate significant difference between the mutant and wild type (Col-0) (Wilcoxon test, $p < 2.2e-16$). (B) The intron/exon ratio per gene in Col-0 vs. the indicated mutants. Black dots represent all genes with raw intronic read number ≥ 2, exonic read number ≥ 5, and final intron/exon ratio between 0 to 1. Red dots represent genes with significantly higher intron/exon ratio relative to wild type (fold change ≥ 2 and FDR < 0.01). (C) A Venn diagram showing the degree of overlap among genes with intron retention defects in *mac7-1*, *mac3a mac3b* and *prl1 prl2*. (D) Examples of genes with intron retention defects. RPKM: Reads Per Kilobase per Million mapped reads. Two biological replicates for each genotype are shown. The rectangles mark introns with higher retention in the mutants.

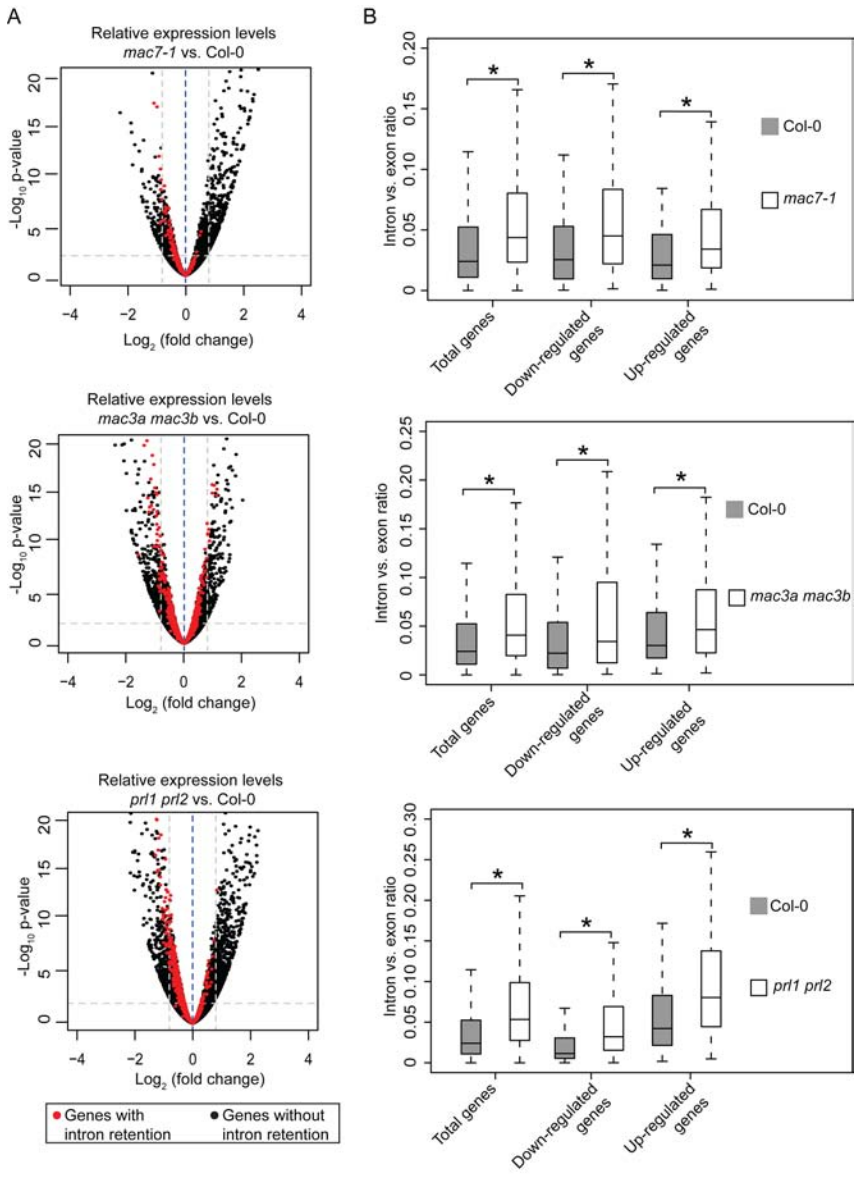

Figure 9. MAC affects pre-mRNA splicing and gene expression independently.
(A) Volcano plots illustrating fold changes of gene expression levels in the indicated mutants compared to Col-0. Thresholds for fold change of 1.5 and $p \leq 0.01$ are shown in the plots as gray dashed lines. Black dots and red dots represent the same genes as in Figure 8B. (B) Box plots of intron/exon ratios in total genes, up-regulated and down-regulated genes in the indicated genotypes. Only genes that pass an abundance filter (raw intronic read number ≥ 2 and exonic read number ≥ 5) were included in this analysis. Asterisks indicate significant difference between the mutant and wild type (Col-0) (Wilcoxon test, $p < 2.2e\text{-}16$).

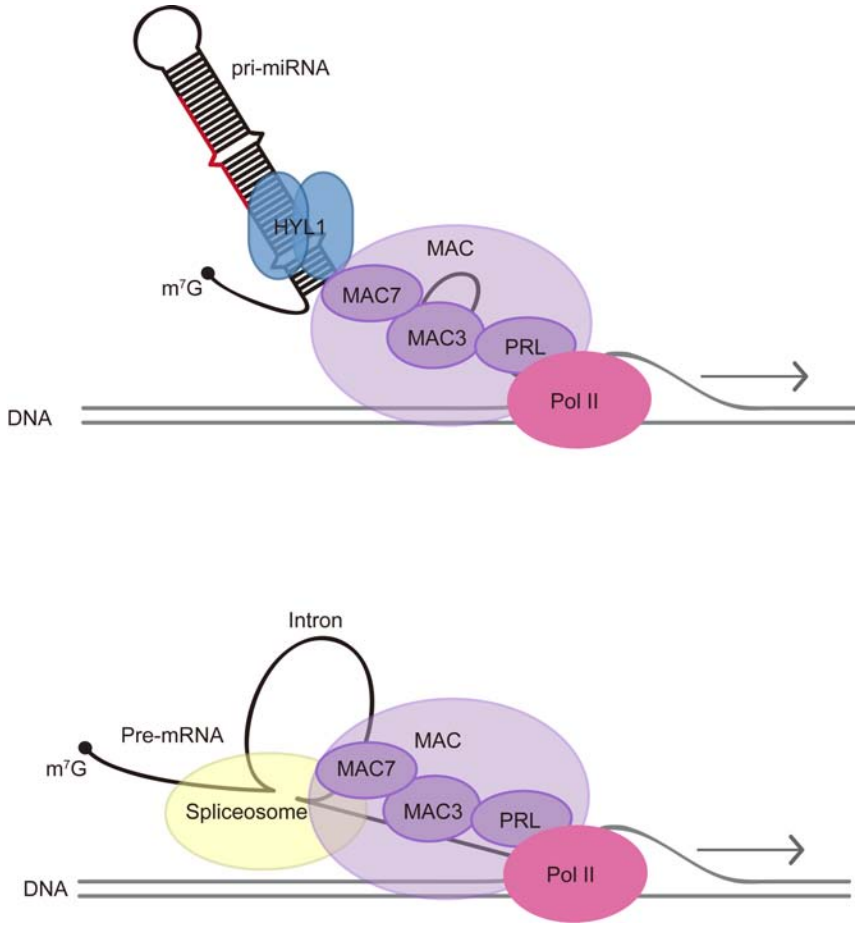

Figure 10. A model for MAC's functions in miRNA biogenesis and pre-mRNA processing. MAC affects miRNA biogenesis through influencing Pol II transcription and interacting with the pri-miRNA processing factor HYL1. MAC also plays a role in pre-mRNA splicing through interactions with the spliceosome. MAC seems to have separate roles in miRNA biogenesis and RNA splicing but a common theme appears to be co-transcriptional RNA processing.

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