

Short-term responses to warming vary between native vs. exotic species and with latitude in an
early successional plant community

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Abstract

Climate change is expected to favor exotic plant species over native species because exotics tend to have wider climatic tolerances and greater phenological plasticity, and also because climate change may intensify enemy release. Here, we examine direct effects of warming (+1.8°C above ambient) on plant abundance and phenology, as well as indirect effects of warming propagated through herbivores, in two heavily invaded plant communities in Michigan, USA separated by approximately three degrees latitude. At the northern site, warming increased exotic plant abundance by 19% but decreased native plant abundance by 31%, indicating that exotic species may be favored in a warmer world. Warming also resulted in earlier spring green-up (1.65 ± 0.77 days), earlier flowering (2.18 ± 0.92 days), and greater damage by herbivores (2-fold increase), affecting exotic and native species equally. Contrary to expectations, native and exotic plants experienced similar amounts of herbivory. Warming did not have strong ecological effects at the southern site, only resulting in a delay of flowering time by 2.42 ± 0.83 days for both native and exotic species. Consistent with the enemy release hypothesis, exotic plants experienced less herbivory than native plants at the southern site. Herbivory was lower under warming for both exotic and native species at the southern site. Thus, climate warming may favor exotic over native plant species, but the response is likely to depend on additional environmental and individual species' traits.

Keywords: climate-change, global warming, invasive species, open-top chamber, plant community ecology

Introduction

Understanding the mechanisms for the success of exotic plants in their introduced range is a longstanding ecological question (Tilman 1985; Davies 2011; Simberloff 2013). The rapid spread and eventual dominance of exotic plants may result from their ability to take advantage of vacant spatial or temporal niche space (Elton 1958), especially following disturbance (Herbold and Moyle 1986). A second proposed mechanism is enemy release, in which exotic plants experience reduced herbivore, disease, and pest pressure compared to their native counterparts (Williamson 1996; Keane and Crawley 2002). However, anthropogenic climate change is likely to interact with each of these mechanisms through extended growing season length and rapidly warming temperatures (Wolkovich and Cleland 2011). These changes are also expected to shift plant-herbivore relationships (Dijkstra et al. 2012). Understanding the mechanisms that influence exotic plants' success under warmer temperatures in their introduced range is critical for understanding and forecasting plant community dynamics.

Warmer temperatures driven by climate change are known to influence the abundance of exotic species (Dukes 2010). For example, warmer temperatures could reduce cold constraints that currently limit exotic survival and open potential opportunities for range expansion (Dukes and Mooney 1999). Further, native species that are stressed by a rapidly changing climate may have reduced competitive abilities and therefore, allow opportunities for exotic establishment or growth in the introduced range (Alpert et al. 2000; Chambers et al. 2016; Ma et al. 2017). Warmer temperatures also lead to extended growing seasons and may create open niche space for exotic species that are able to establish earlier in the season or extend their life cycle past those of native species (Wolkovich and Cleland 2011), and has been seen in both terrestrial and aquatic systems (Stachowicz et al. 2002; Fridley 2012). Thus, exotic species may also be favored

in their introduced range due to high phenological plasticity (Richards et al. 2006). Accordingly, these species may have an advantage in a rapidly changing environment (Willis et al. 2008; Matesanz et al. 2010). However, some evidence exists that climate change may lead to neutral and potentially negative responses for exotic and invasive species, suggesting their success may depend on abiotic and biotic interactions (Williams et al. 2007; Bradley et al. 2010).

The success of exotic species under climate change is also likely to depend on their interactions with herbivores. Exotic plants benefit from enemy release when herbivores prefer native species and therefore indirectly enable exotic species to thrive under relatively less consumptive pressure (Bremm et al. 2016). Climate change may moderate the magnitude and direction of herbivore impacts on exotic plant success by warmer temperatures directly increasing insect metabolic rate (Bale et al. 2002; Fey and Herren 2014). The warming induced reduction of plant food quality may also require herbivores to consume greater quantities to meet nutritional demands (Hillebrand et al. 2009; Sardans et al. 2012). If native species experience increased herbivory due to warming, but this increase is lower in magnitude for exotic species (i.e. enemy release), then exotic species would experience greater success under warmer climate conditions. Alternatively, herbivores may limit the growth of competitive plant species and therefore promote community stability (Van der Putten et al. 2010; Blois et al. 2013; Post 2013; Borer et al. 2014). Thus, forecasting community consequences of exotic plant success may depend on the influence of herbivores in a warmer world.

Given these direct and indirect responses to climate change, exotic species may be favored in their introduced ranges (Dukes and Mooney 1999; Hellmann et al. 2008). However, these predictions are largely based on theory and observational studies rather than manipulative experiments (Wolkovich and Cleland 2011; Sandel and Dangremond 2012; Wolkovich et al.

2013). Further, experiments that test the differential effects of warming on exotic and native species often use simplified biotic communities, which may reduce the ability to make predictions under the full complexity of realistic communities (Lu et al. 2015; Munier et al. 2010). Also, *in situ* field-based warming experiments in plant communities have been almost entirely limited to high-latitude and high-elevation ecosystems with low-stature plant communities, and we lack sufficient understanding in mid-latitude and lower-elevation ecosystems (Arft et al. 1999). Finally, in order to best examine the effects of warming on exotic success we must examine ecosystems where introductions of exotics are most prevalent (Simberloff and Von Holle 1999; Turbelin et al. 2017).

We addressed these research gaps with an *in situ* field experiment that manipulated warming in highly invaded early successional, mid-latitude plant communities. We examined the effects of warming exotic vs. native species for one annual cycle to test the following hypotheses:

- 1) *Warming will increase exotic species cover more than co-occurring native species cover.*
- 2) *Warming will advance exotic species phenology more than co-occurring native species phenology.*
- 3) *Warming will indirectly favor exotic species through increased leaf herbivory compared to co-occurring native species.*

Methods

Site Description

We examined two invaded, early successional plant communities in Michigan, USA separated by 354 km and approximately three degrees latitude (Appendix A) from May 2015 through October 2016. The northern site is a forest clearing at The University of Michigan's

Biological Station (UMBS) located at 45.56° N, -84.71° W and 239 meters above sea level (Appendix A). The 1.53 km² clearing was clear-cut in 1994 and is dominated by *Danthonia spicata*, *Centaurea stoebe*, and *Poa pratensis* (Table 1). The site was maintained through mowing to prevent forest succession until the beginning of the experiment. The mean annual temperature in Emmet County where the site is located is 5.4 °C with the mean monthly temperature for the hottest month (July) reaching 17.0 °C (1981-2010 normals, Menne et al. 2012a; Menne et al. 2012b). Mean annual rainfall and snowfall in Emmet County are 76.7 cm and 244 cm, respectively (1981-2010 normals, Menne et al. 2012a; Menne et al. 2012b). The southern site is an early successional grassland (previously agriculture field) at Kellogg Biological Station's Long Term Ecological Research Site (KBS-LTER) located at 42.40° N, -85.40° W and 289 meters above sea level (Appendix A). The site is dominated by *Solidago canadensis*, *Poa pratensis*, and *Hieracium pratense* (Table 1) and was maintained through mowing to prevent succession prior to the beginning of the experiment. The mean annual temperature in Kalamazoo County where the site is located is 10.1 °C with the mean monthly temperature for the hottest month (July) reaching 22.9 °C (1981-2010 normals, Menne et al. 2012a; Menne et al. 2012b). Mean annual rainfall and snowfall at this site are 100.4 cm and 127 cm, respectively (1981-2010 climate normal, Menne et al. 2012a; Menne et al. 2012b).

Experimental Design

At each site, we established 24 1m² plots within a 25 meter by 36 meter area surrounded by a 3 meter tall fence to prevent deer browsing. Plots were separated by at least 4 m of buffer area to prevent edge impacts from chambers (e.g. shading) in neighboring plots and to serve as walkways for sampling. As part of a larger, long-term experiment, the plots were treated with a

randomized, fully factorial design that included insect reduction and small mammal exclosure in addition to the warming treatment (Appendix B).

We warmed the plots year-round using a passive, open-top chamber design commonly used in tundra ecosystems (Molau and Mølgaard 1996), modified to warm taller stature (≤ 1.5 m) plant communities and to allow for migration of flying and ground dwelling organisms in and out of the chambers (Welshofer et al. 2018). We used clear, UV-transmitting, 1/8" Lexan Polycarbonate sheets to elevate temperatures using sunlight (ePlastics, San Diego, CA; Appendix A). We installed the chambers during spring 2015 at both sites. The chambers warmed plots during the daytime, on average, by $(+1.84 \pm 0.79$ °C (SE) and $+1.73 \pm 0.19$ °C (SE)) in the forest clearing and old agriculture field, respectively, with no evidence for differences in relative humidity. Warming varied with irradiance and therefore exhibited the hottest temperatures (up to approximately 45 °C) during the daytime in the growing season. At the southern site, the chambers led to drier soil by 3.66 ± 0.57 % (SE), but this trend was not observed at the northern site where moisture drained quickly from the sandy soil composition (Welshofer et al. 2018). Further, the chambers reduced winter soil temperature, presumably due to reduced snowpack insulation, and resulted in greater variability in soil freeze/thaw cycles (Welshofer et al. 2018). Here we report ecological data collected during the second growing season (2016), after nearly a full year of the warming treatment.

Data collection

Because this study was part of a longer-term experiment, we used non-destructive sampling methods. We visually estimated percent cover for each species in each 1m² plot taken at the beginning of the experiment (Spring 2015) and at the end of the second growing season (August 2016). KW performed all estimates to prevent bias between plots or sampling occasions.

The origin of each species was classified as either 'native' or 'exotic' according to the PLANTS Database (USDA, NRCS 2006).

We monitored the phenology in each plot every 3-4 days from March through October 2016. We recorded 'green-up' for each species in each plot by visually estimating the percent of active plant cover in each 1m² plot during the time period (March-May). We recorded a species as 'flowering' as the period between flower bud break and flower senescence. 'Seed set' was determined when an individual exhibited withering of the stigma along with swelling of the ovaries. For both flowering and seed set, we recorded the date when at least one individual for each species in each plot exhibited the phenological stage.

We visually estimated the percent of leaf tissue missing (presumably due to herbivory) at the end of the second growing season to capture cumulative damage throughout the season (Schultheis et al. 2015). We haphazardly selected four leaves vertically dispersed along the stem on three individuals of each species within each plot (n=12 leaves per species per plot). When three individuals of that species were not present within a plot, we sampled the maximum number of individuals present. We sampled all species except for *Achillea millefolium*, because visual estimates of tissue damage on its feather-like leaves were difficult to assess. For graminoid species, many ends of the blade were removed making it difficult to determine the amount of tissue eaten. To standardize our results between treatments, we used the average length of undamaged blades for each species as a reference for each leaf herbivory estimate. We only recorded the percent of missing plant tissue, as browning of the tissue could have been a response to abiotic stress or fungal infections (Green et al. 1990). To prevent bias, KW completed all estimates with visual aids depicting species-specific examples of exact herbivory percentages that were calculated using ImageJ software (Schneider et al. 2012). To further

determine the probability of leaf attack, we later calculated a binomial response as to whether the leaf experienced any herbivory (1), or no herbivory (0).

Statistical Analysis

We took several steps to prepare the data for analysis. We natural log transformed each percent cover value to conform to the assumptions of general linear regression. As an index of spring green-up phenology, we computed the date at which each species reached 50% of its maximum cover value per plot. For time of flowering and seed set, we computed the minimum date that each species was observed in each phenological stage in each plot. For the leaf herbivory models, we did not consider plots with the reduced insect treatment (n=12 per site).

We fit mixed-effect ANOVA models to evaluate the separate and interactive effects of warming and origin (native vs. exotic) on plant percent cover, spring green-up phenology, flowering phenology, seed set phenology, and leaf herbivory. For each response variable at each site, we used a 2-step process to compare complex and reduced models using likelihood ratio tests ($\alpha=0.05$) to select the most parsimonious model. To account for the additional treatments applied as part of the long-term experiment, even though they are not a focus of this study, we first compared the full models including terms for insect reduction and small mammal exclusion treatments to models without those terms. Next, we compared the model including an interaction between warming and origin to a model without the interaction to test for interactive effects of warming and plant species origin on each response variable. A significant interaction term would indicate that the change in the response variable caused by warming depends on plant species origin. Plant species identity was included as a random intercept in each model to account for additional species-specific variation within all response variables. For leaf herbivory models, the data did not conform to a normal distribution and could not be transformed as such; therefore, we

used a generalized linear regression model with Poisson error to determine the amount of the leaf eaten and a binomial error to determine the probability of herbivore attack. For both herbivory models the identity of individual plants was also included as a random intercept because multiple observations were made on each individual. 95% confidence intervals were calculated for each parameter estimate using bootstrap methods (number of simulations=999). We calculated the denominator degrees of freedom using the Satterthwaite approximation method (Bolker et al. 2009). We used Likelihood Ratio Tests ($\alpha=0.05$) to evaluate whether initial differences in species cover between warmed and ambient plots existed prior to the start of the experiment, and completed all analyses using R version 3.1.2 (R Development Core Team 2008) and the R package lme4 (Pinheiro and Bates 2000). There were no initial differences between warmed and ambient plots in average percent cover across species at either site in 2015 at the beginning of the experiment (Northern Site: ($\chi^2=0.84$, $df=1$, $p=0.36$); Southern Site: ($\chi^2=0.35$, $df=1$, $p=0.55$)).

Results

At the northern site, the response of plant cover to warming depended on origin (Table 2, Figure 1A). Warming increased exotic plant cover by 19% and decreased native plant cover by 31% relative to the average cover in ambient plots ($((\bar{x}_{\text{warmed}} - \bar{x}_{\text{ambient}}) / \bar{x}_{\text{ambient}}) * 100$). The greatest increases in cover were found in *Rumex acetosella*, *Poa pratensis*, and *Hypericum perforatum* (all exotic). However, we did not find evidence that warming influenced plant cover for either native or exotic species in the southern old agriculture field (Table 2, Figure 1B). We also did not find evidence for an interactive effect between warming and origin on average species cover at this site (Appendix C). However, we observed the greatest increases in cover

due to warming of *Trifolium* sp. (exotic) and *Euthamia graminifolia* (native), with the greatest decrease in *Poa pratensis* (exotic). Responses of individual species are reported in Appendix D.

At the northern forest clearing, plant species greened-up an average of 1.61 ± 0.57 (SE) days earlier in warmed plots than ambient plots (Table 2, Figure 2A). Native and exotic plants shifted green-up phenology by the same amount of time, on average (i.e. the interaction of warming and origin was not present in the most parsimonious model, Appendix C). Plants flowered 2.18 ± 0.92 days earlier in the warmed plots than plants in the ambient plots at the northern site (Table 2, Figure 2C), and the magnitude of this shift was also indistinguishable between native and exotic species (Appendix C). We found that plots at the northern site without the insecticide treatment flowered 2.13 ± 0.86 days later than plots with the insecticide treatment (Table 2). The timing of seed set did not vary according to warming treatment or origin at the northern site (Table 2, Figure 2E). At the southern old agriculture field, warming did not affect spring green-up phenology for native or exotic plants (Table 2, Figure 2B), but exotic plants greened-up an average of 10.35 ± 4.34 days earlier than native plants, regardless of warming. Warming delayed plant flowering by 2.42 ± 0.83 days for both native and exotic species (Table 2, Figure 2D). At this site, exotic plants flowered 37.14 ± 16.57 days earlier than native plants, regardless of warming (Figure 2D). The timing of seed set also did not vary according to warming treatment or origin at the southern site (Table 2, Figure 2F). The interaction between warming and origin was not present in the most parsimonious model for any of the phenology responses at the northern or southern study site (Appendix C).

Warming increased the amount of leaf herbivory approximately two-fold at the northern site (Table 3, Figure 3). Native and exotic species experienced similar amounts of herbivory (Table 3, Figure 3), but the way herbivory varied according to warming treatment was

statistically indistinguishable for native vs. exotic species (Appendix C). Warming decreased the amount of leaf herbivory in the southern old agriculture field by approximately half, and native species experienced about 6-fold greater herbivory than exotic species (Table 3). Similarly, at the southern site, the probability of an herbivore attacking a leaf decreased in warmed plots compared to ambient plots and native species experienced a higher probability of attack than exotic species (Table 3). However, the probability of herbivore attack at the northern site did not differ between warmed and ambient treatments, nor by species origin (Table 3). The most parsimonious models for both the amount of leaf eaten and the probability of herbivore attack at both sites did not contain a term for the interaction of warming and herbivory (Appendix C).

Discussion

We found evidence that warming favored exotic species over native species at the northern forest clearing site, where the effects of warming were largely consistent with our predictions. However, the same level of warming did not have strong ecological effects at the southern old agricultural field site. We also found evidence for enemy release at the southern old agriculture field site where native plant species were more likely to experience leaf herbivory and a greater amount of tissue damage than exotic species. However, these trends were not observed at the northern forest clearing site.

The moderate ecological effects observed at the northern forest clearing site were consistent with our expectations. Warmed plots had greater plant cover compared to ambient plots, consistent with experimental studies that found warming increases plant productivity (Rustad et al. 2001) or increases the abundance of more productive species (Fridley et al. 2016). Also consistent with predictions, the change in cover was positive for exotic species but negative

for native species. The observed shifts in earlier green-up and flowering time at the northern site are consistent with the literature, as plants respond to warmer temperature cues earlier in the season (Dunnell and Travers 2011; Calinger et al. 2013). Warmed plants also experienced greater amounts of herbivory than those in ambient conditions. This could be due to lower food quality (as a result of increased C:N ratios) and therefore increased herbivore demand (Hillebrand et al. 2009; Sardans et al. 2012). Additionally, warmer temperatures may have increased the metabolism of insects within the warmed plots, in turn increasing the amount of leaf tissue required by herbivores (Gillooly 2001). Exotic species greened-up an entire week earlier than native species, indicating their broad climatic tolerance early in the growing season, but the reproductive phenology of native and exotic species responded similarly to warming.

Responses at the southern old agriculture field site were not consistent with our original predictions. We observed delayed flowering phenology and a slight but statistically insignificant decrease in plant cover in the warmed plots. We suggest that physiological stress may have prevented both native and exotic plants from benefiting from warming at the southern site with the extreme temperatures (+44 days where temperatures reached 35°C) exacerbated by lower soil moisture conditions ($-3.66 \pm 0.57\%$) in the warmed plots (Welshofer et al. 2017). Although there is considerable variability among plant species in the conditions that cause physiological stress (Wahid et al. 2007), combined heat and drought conditions have shown to delay flowering phenology (Savin and Nicolas 1996; Kazan and Lyons 2016) and decrease aboveground plant biomass (Vile et al. 2011). Additionally, lower soil moisture in the warmed plots could have resulted from increased transpiration at higher temperatures and contributed to plant stress (Pandey et al. 2015). Hence, such extreme conditions could explain these observed plant responses to warming at the southern site.

Overall, we found limited evidence that exotic plant species will respond to climate change differently than native species, with the exception that warming caused a significantly greater increase in exotic cover relative to native cover at the northern site. The lack of differential effects of warming between native or exotic species may be due to similar ruderal traits shared by species of both origins. Because these systems are both heavily disturbed (clear-cut and agriculture), weedy species dominate the plant assemblage at both sites. Weedy species that readily establish in disturbed environments are likely to have broad climatic tolerances and be relatively insensitive to moderate warming, regardless of their continent of origin (Theoharides and Dukes 2007). For example, plant species that have an affinity for warmer soils and that produce wind-dispersed seeds tended to be the best colonizers of newly available habitat patches as temperatures increased in an alpine system (Matteodo et al. 2013). Notably, warming did not produce changes in the most abundant species at each site, including exotic species of national concern *Centaurea stoebe* (USDA, NRCS 2006). Thus, it is likely that species traits, in addition to species origin, are likely to play an important role in predicting species success in a warmer climate.

This experiment suggests climate warming can benefit exotic plant species more than native species, but our results suggest the magnitude and direction of responses are also likely to depend on the environmental temperature as well as species' traits within the community. While short-term experiments such as this increase our understanding of annual ecological variation, they also suggest potential mechanisms that lead to long-term ecological dynamics (Chesson and Huntly 1989; Magnuson 1990; Shriver 2016). We suggest that future research examining the influence of warming on the physiological tolerances of native and exotic species as well as

plant-herbivore interactions will provide insight to whether these short-term direct and indirect responses to warming translate into greater success of exotic plant species over time.

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References

- Alpert, P., Bone, E., and Holzapfel, C. (2000). Invasiveness, invasibility and the role of environmental stress in the spread of non-native plants. *Perspectives in Plant Ecology, Evolution and Systematics*, 3(1), 52–66.
- Arft, A. M., Walker, M. D., Gurevitch, J., Alatalo, J. M., Bret-Harte, M. S., Dale, M., Diemer, M., Gugerli, F., Henry, G. H. R., Jones, M. H., Hollister, R. D., Jónsdóttir, I. S., Laine, K., Lévesque, E., Marion, G. M., Molau, U., Mølgaard, P., Nordenhäll, U., Raszhivin, V., Robinson, C. H., Starr, G., Stenström, A., Stenström, M., Totland, Ø., Turner, P. L., Walker, L. J., Webber, P. J., Welker, J. M. and Wookey, P. A. (1999), Responses of tundra plants to experimental warming: Metal-analysis of the International Tundra

Experiment. *Ecological Monographs*, 69: 491–511.

Bale, J. S., Masters, G. J., Hodkinson, I. D., Awmack, C., Bezemer, T. M., Brown, V. K., Butterfield, J., Buse, A., Coulson, J. C., Farrar, J., Good, J. E. G., Harrington, R., Hartley, S., Jones, T. H., Lindroth, R. L., Press, M. C., Symrnioudis, I., Watt, A. D. and Whittaker, J. B. (2002), Herbivory in global climate change research: direct effects of rising temperature on insect herbivores. *Global Change Biology*, 8: 1–16.

Blois, J. L., Zarnetske, P. L., Fitzpatrick, M. C., and Finnegan, S. (2013). Climate Change and the Past, Present, and Future of Biotic Interactions. *Science*, 341(6145), 499–504.

Bolker, B. M., Brooks, M. E., Clark, C. J., Geange, S. W., Poulsen, J. R., Stevens, M. H. H. and White, J.-S. S. (2009). Generalized linear mixed models: a practical guide for ecology and evolution. *Trends in Ecology & Evolution*, 24(3), 127–135.

Borer, E. T., Seabloom, E. W., Gruner, D. S., Harpole, W. S., Hillebrand, H., Lind, E. M., Adler, P. B., Alberti, J., Anderson, T. M., Bakker, J. D., Biederman, L., Blumenthal, D., Brown, C. S., Brudvig, L. A., Buckley, Y. M., Cadotte, M., Chu, C., Cleland, E. E., Crawley, M. J., Daleo, P., Damschen, E. I., Davies, K. F., Decrappeo, N. M., Du, G., Firn, J., Hautier, Y., Heckman, R. W., Hector, A., Hillerislambers, J., Iribarne, O., Klein, J. A., Knops, J. M. H., La Pierre, K. J., Leakey, A. D. B., Li, W., MacDougall, A. S., McCulley, R. L., Melbourne, B. A., Mitchell, C. E., Moore, J. L., Mortensen, B., O'Halloran, L. R., Orrock, J. L., Pascual, J., Prober, S. M., Pyke, D. A., Risch, A. C., Schuetz, M., Smith, M. D., Stevens, C. J., Sullivan, L. L., Williams, R. J., Wragg, P. D., Wright, J. P. and Yang, L. H. (2014). Herbivores and nutrients control grassland plant diversity via light limitation. *Nature*, 508(7497), 517–520.

Bradley, B.A., Blumenthal, D.M., Wilcove, D.S. and Ziska, L.H. (2010). Predicting plant

363 invasions in an era of global change. *Trends in Ecology and Evolution*, 25 (5): 310-318.
 364 Bremm, C., Carvalho, P. C. F., Fonseca, L., Amaral, G. A., Mezzalana, J. C., Perez, N. B.,
 365 Nabinger, C. and Laca, E. A. (2016). Diet Switching by Mammalian Herbivores in
 366 Response to Exotic Grass Invasion. *PLOS ONE*, 11(2), e0150167.
 367 Calinger, K. M., Queenborough, S., and Curtis, P. S. (2013). Herbarium specimens reveal the
 368 footprint of climate change on flowering trends across north-central North America.
 369 *Ecology Letters*, 16(8), 1037–1044.
 370 Chambers, J. C., Germino, M. J., Belnap, J., Brown, C. S., Schupp, E. W. and Clair, S. B. S.
 371 (2016). Plant Community Resistance to Invasion by Bromus Species: The Roles of
 372 Community Attributes, Bromus Interactions with Plant Communities, and Bromus Traits.
 373 In M. J. Germino, J. C. Chambers, and C. S. Brown (Eds.), *Exotic Brome-Grasses in Arid*
 374 *and Semiarid Ecosystems of the Western US* (pp. 275–304). Cham: Springer International
 375 Publishing.
 376 Chesson, P. and Huntly, N. (1989). Short-term instabilities and long-term community dynamics.
 377 *Trends in Ecology & Evolution*, 4(10), 293–298.
 378 Davies, K. W. (2011). Plant community diversity and native plant abundance decline with
 379 increasing abundance of an exotic annual grass. *Oecologia*, 167(2), 481–491.
 380 Dell, A. I., Pawar, S., and Savage, V. M. (2011). Systematic variation in the temperature
 381 dependence of physiological and ecological traits. *Proceedings of the National Academy*
 382 *of Sciences*, 108(26), 10591–10596.
 383 Dijkstra, F. A., Pendall, E., Morgan, J. A., Blumenthal, D. M., Carrillo, Y., LeCain, D. R.,
 384 Follett, R. F. and Williams, D. G. (2012). Climate change alters stoichiometry of
 385 phosphorus and nitrogen in a semiarid grassland. *New Phytologist*, 196(3), 807–815.

386 Dukes, J. S. (2010). Responses of Invasive Species to a Changing Climate and Atmosphere. In
 387 D. M. Richardson (Ed.), *Fifty Years of Invasion Ecology* (pp. 345–357). Oxford, UK:
 388 Wiley-Blackwell.

389 Dukes, J. S. and Mooney, H. A. (1999). Does global change increase the success of biological
 390 invaders? *Trends in Ecology & Evolution*, 14(4), 135–139.

391 Dunnell, K. L. and Travers, S. E. (2011). Shifts in the flowering phenology of the northern Great
 392 Plains: Patterns over 100 years. *American Journal of Botany*, 98(6), 935–945.

393 Elton, C. S. (1958). *The ecology of invasions by animals and plants* (University of Chicago Press
 394 ed). Chicago: University of Chicago Press.

395 Fey, S. B. and Herren, C. M. (2014). Temperature-mediated biotic interactions influence enemy
 396 release of nonnative species in warming environments. *Ecology*, 95(8), 2246–2256.

397 Fridley, J. D. (2012). Extended leaf phenology and the autumn niche in deciduous forest
 398 invasions. *Nature*, 485(7398), 359–362.

399 Fridley, J. D., Lynn, J. S., Grime, J. P. and Askew, A. P. (2016). Longer growing seasons shift
 400 grassland vegetation towards more-productive species. *Nature Climate Change*, 6(9),
 401 865–868.

402 Gillooly, J. F. (2001). Effects of Size and Temperature on Metabolic Rate. *Science*, 293(5538),
 403 2248–2251.

404 Green, J. L., Capizzi, J. and Maloy, O. (1990). *A Systematic Approach to Diagnosing Plant*
 405 *Damage* (Ornamentals Northwest Archive No. Vol. 13 Issue 6) (p. ii-24). Oregon State
 406 University Extension.

407 Hellmann, J. J., Byers, J. E., Bierwagen, B. G. and Dukes, J. S. (2008). Five Potential
 408 Consequences of Climate Change for Invasive Species. *Conservation Biology*, 22(3),

409 534–543.

410 Herbold, B. and Moyle, P. B. (1986). Introduced Species and Vacant Niches. *The American*
411 *Naturalist*, 128(5), 751–760.

412 Hillebrand, H., Borer, E. T., Bracken, M. E. S., Cardinale, B. J., Cebrian, J., Cleland, E. E.,
413 Elser, J. J., Gruner, D. S., Stanley Harpole, W., Ngai, J. T., Sandin, S., Seabloom, E. W.,
414 Shurin, J. B., Smith, J. E. and Smith, M. D. (2009). Herbivore metabolism and
415 stoichiometry each constrain herbivory at different organizational scales across
416 ecosystems. *Ecology Letters*, 12(6), 516–527.

417 Kazan, K. and Lyons, R. (2016). The link between flowering time and stress tolerance. *Journal*
418 *of Experimental Botany*, 67(1), 47-60.

419 Keane, R. and Crawley, M. (2002). Exotic plant invasions and the enemy release hypothesis.
420 *Trends in Ecology & Evolution*, 17(4), 164–170.

421 Lu, X., Siemann, E., Wei, H., Shao, X. and Ding, J. (2015). Effects of warming and nitrogen on
422 above- and below-ground herbivory of an exotic invasive plant and its native congener.
423 *Biological Invasions*, 17(10), 2881–2892.

424 Ma, Z., Liu, H., Mi, Z., Zhang, Z., Wang, Y., Xu, W., Jiang, L. and He, J.-S. (2017). Climate
425 warming reduces the temporal stability of plant community biomass production. *Nature*
426 *Communications*, 8, 15378.

427 Magnuson, J. J. (1990). Long-Term Ecological Research and the Invisible Present. *BioScience*,
428 40(7), 495–501.

429 Matesanz, S., Gianoli, E. and Valladares, F. (2010). Global change and the evolution of
430 phenotypic plasticity in plants: Global change and plasticity. *Annals of the New York*
431 *Academy of Sciences*, 1206(1), 35–55.

432 Matteodo, M., Wipf, S., Stöckli, V., Rixen, C. and Vittoz, P. (2013). Elevation gradient of
 433 successful plant traits for colonizing alpine summits under climate change.
 434 *Environmental Research Letters*, 8(2), 024043.

435 Menne, M. J., Durre, I., Korzeniewski, B., McNeill, S., Thomas, K., Yin, X., Anthony, S., Ray,
 436 R., Vose, R. S., Gleason, B. E. and Houston, T. G. (2012a). Global Historical
 437 Climatology Network - Daily (GHCN-Daily), Version 3. NOAA National Climatic Data
 438 Center.

439 Menne, M. J., Durre, I., Vose, R. S., Gleason, B. E. and Houston, T. G. (2012b). An Overview of
 440 the Global Historical Climatology Network-Daily Database. *Journal of Atmospheric and*
 441 *Oceanic Technology*, 29(7), 897–910.

442 Molau, U. and Mølgaard, P. (1996). *ITEX Manual* (2nd ed.). Copenhagen: Danish Polar Center.

443 Munier, A., Hermanutz, L., Jacobs, J. D. and Lewis, K. (2010). The interacting effects of
 444 temperature, ground disturbance, and herbivory on seedling establishment: implications
 445 for treeline advance with climate warming. *Plant Ecology*, 210(1), 19–30.

446 Pandey, P., Ramegowda, V., and Senthil-Kumar, M. (2015). Shared and unique responses of
 447 plants to multiple individual stresses and stress combinations: physiological and
 448 molecular mechanisms. *Frontiers in Plant Science*, 6:723.

449 Post, E. S. (2013). *Ecology of climate change: the importance of biotic interactions*. Princeton:
 450 Princeton University Press.

451 R Development Core Team. (2008). R: A language and environment for statistical computing.
 452 Vienna: R Foundation for Statistical Computing. Retrieved from [http://www.R-](http://www.R-project.org)
 453 [project.org](http://www.R-project.org)

454 Richards, C. L., Bossdorf, O., Muth, N. Z., Gurevitch, J. and Pigliucci, M. (2006). Jack of all
 455 trades, master of some? On the role of phenotypic plasticity in plant invasions. *Ecology*
 456 *Letters*, 9(8), 981–993.

457 Rustad, L., Campbell, J., Marion, G., Norby, R., Mitchell, M., Hartley, A., Cornelissen, J. and
 458 Gurevitch, J. (2001). A meta-analysis of the response of soil respiration, net nitrogen
 459 mineralization, and aboveground plant growth to experimental ecosystem warming.
 460 *Oecologia*, 126(4), 543–562.

461 Sandel, B. and Dangremond, E. M. (2012). Climate change and the invasion of California by
 462 grasses. *Global Change Biology*, 18(1), 277–289.

463 Sardans, J., Rivas-Ubach, A., and Peñuelas, J. (2012). The C:N:P stoichiometry of organisms and
 464 ecosystems in a changing world: A review and perspectives. *Perspectives in Plant*
 465 *Ecology, Evolution and Systematics*, 14(1), 33–47.

466 Savin, R. and Nicolas, M.E. (1996). Effects of Short Periods of Drought and High Temperature
 467 on Grain Growth and Starch Accumulation of Two Malting Barley Cultivars. *Australian*
 468 *Journal of Plant Physiology*. 23(2), 201-210.

469 Schneider, C. A., Rasband, W. S., and Eliceiri, K. W. (2012). NIH Image to ImageJ: 25 years of
 470 image analysis. *Nature Methods*, 9(7), 671–675.

471 Schultheis, E. H., Berardi, A. E., and Lau, J. A. (2015). No release for the wicked: enemy release
 472 is dynamic and not associated with invasiveness. *Ecology*, 96(9), 2446–2457.

473 Shriver, R. K. (2016). Quantifying how short-term environmental variation leads to long-term
 474 demographic responses to climate change. *Journal of Ecology*, 104(1), 65–78.

475 Simberloff, D. (2013). *Invasive species: what everyone needs to know*. Oxford: Oxford
 476 University Press.

477 Simberloff, D. and Von Holle, B. (1999). Positive Interactions of Nonindigenous Species:
 478 Invasional Meltdown? *Biological Invasions*, 1(1), 21–32.

479 Stachowicz, J. J., Terwin, J. R., Whitlatch, R. B. and Osman, R. W. (2002). Nonlinear partial
 480 differential equations and applications: Linking climate change and biological invasions:
 481 Ocean warming facilitates nonindigenous species invasions. *Proceedings of the National*
 482 *Academy of Sciences*, 99(24), 15497–15500.

483 Theoharides, K. A. and Dukes, J. S. (2007). Plant invasion across space and time: factors
 484 affecting nonindigenous species success during four stages of invasion. *New Phytologist*,
 485 176(2), 256–273.

486 Tilman, D. (1985). The Resource-Ratio Hypothesis of Plant Succession. *The American*
 487 *Naturalist*, 125(6), 827–852.

488 Turbelin, A. J., Malamud, B. D. and Francis, R. A. (2017). Mapping the global state of invasive
 489 alien species: patterns of invasion and policy responses: Mapping the global state of
 490 invasive alien species. *Global Ecology and Biogeography*, 26(1), 78–92.

491 USDA, NRCS. (2006). The PLANTS Database, 6 March 2006 (<http://plants.usda.gov>). National
 492 Plant Data Center, Baton Rouge, LA 70874-4490 USA.

493 Van der Putten, W. H., Macel, M. and Visser, M. E. (2010). Predicting species distribution
 494 and abundance responses to climate change: why it is essential to include biotic
 495 interactions across trophic levels. *Philosophical Transactions of the Royal Society B:*
 496 *Biological Sciences*, 365(1549), 2025-2034.

497 Vile, D., Pervent, M., Belluau, M., Vasseur, F., Bresson, J., Muller, B., Granier, C., and
 498 Simonneau, T. (2012). Arabidopsis growth under prolonged high temperature and water
 499 deficit: independent or interactive effects?. *Plant, Cell, and Environment*, 35, 702-718.

500 Wahid, A., Gelani, S., Ashraf, M., and Foolad, M.R. (2007). Heat tolerance in plants: an
 501 overview. *Environmental and Experimental Botany*, 61: 199-223.

502 Welshofer, K. B., Zarnetske, P. L., Lany, N. K. and Thompson, L. A. E. (2018). Open-
 503 top chambers for temperature manipulation in taller-stature plant communities. *Methods*
 504 *Ecology and Evolution*, 9: 254-259.

505 Williams, A. L., Wills, K. E., Janes, J. K., Vander Schoor, J. K., Newton, P. C. D. and
 506 Hovenden, M. J. (2007), Warming and free-air CO₂ enrichment alter demographics in
 507 four co-occurring grassland species. *New Phytologist*, 176: 365–374.

508 Williamson, M. H. (1996). *Biological invasions* (1st ed). London ; New York: Chapman & Hall.

509 Willis, C., Ruhfel, B., Primack, R., Abraham, J. and Davis, C. (2008). Phylogenetic patterns of
 510 species loss in Thoreau’s woods are driven by climate change. *Proceedings of the*
 511 *National Academy of Sciences*, 105(44), 17029–17033.

512 Wolkovich, E. M., Davies, T. J., Schaefer, H., Cleland, E. E., Cook, B. I., Travers, S. E. and
 513 Davis, C. C. (2013). Temperature-dependent shifts in phenology contribute to the success
 514 of exotic species with climate change. *American Journal of Botany*, 100(7), 1407–1421.

515 Wolkovich, E. M. and Cleland, E. E. (2011). The phenology of plant invasions: a community
 516 ecology perspective. *Frontiers in Ecology and the Environment*, 9(5), 287–294.

Table 1. Species list for each site, and site-level relative abundance. All origin values retrieved from USDA Plant Database at the county level. Species relative abundance based on composition estimates collected in August 2016. Relative abundance calculated for each site as the (sum of each species' cover/sum of all species)*100.

Northern Forest Clearing			Southern Old Agriculture Field		
Species	Origin	Relative Abundance (%)	Species	Origin	Relative Abundance (%)
<i>Danthonia spicata</i>	Native	38.34	<i>Solidago canadensis</i>	Native	34.27
<i>Centaurea stoebe</i>	Exotic	29.65	<i>Poa pratensis</i>	Exotic	33.74
<i>Poa pratensis</i>	Exotic	11.44	<i>Hieracium pratense</i>	Exotic	10.17
<i>Carex pensylvanica</i>	Native	6.21	<i>Phleum pratense</i>	Exotic	4.73
<i>Pteridium aquilinum</i>	Native	4.83	<i>Achillea millefolium</i>	Native	4.56
<i>Rumex acetosella</i>	Exotic	2.84	<i>Trifolium pratense</i>	Exotic	3.28
<i>Hypericum perforatum</i>	Exotic	1.11	<i>Centaurea stoebe</i>	Exotic	2.82
<i>Hieracium pilosella</i>	Exotic	<1	<i>Euthamia graminifolia</i>	Native	1.66
<i>Poa compressa</i>	Exotic	<1	<i>Rubus allegheniensis</i>	Native	1.05
<i>Quercus rubra</i>	Native	<1	<i>Potentilla recta</i>	Exotic	<1
<i>Fragaria vesca</i>	Native	<1	<i>Dactylis glomerata</i>	Exotic	<1
<i>Vaccinium angustifolium</i>	Native	<1	<i>Daucus carota</i>	Exotic	<1
<i>Solidago gigantea</i>	Native	<1	<i>Trifolium repens</i>	Exotic	<1
<i>Pilosella aurantiaca</i>	Exotic	<1	<i>Alliaria petiolata</i>	Exotic	<1
<i>Asclepias</i> sp.	Native	<1	<i>Celastrus orbiculatus</i>	Exotic	<1
<i>Betula</i> sp.	Native	<1	<i>Arrhenatherum elatius</i>	Exotic	<1
<i>Antennaria</i> sp.	Native	<1	<i>Hypericum perforatum</i>	Exotic	<1
<i>Solidago nemoralis</i>	Native	<1	<i>Elymus repens</i>	Exotic	<1
<i>Tragopogon dubius</i>	Exotic	<1	<i>Asclepias</i> sp.	Native	<1
<i>Acer rubrum</i>	Native	<1	<i>Taraxicum officinale</i>	Exotic	<1
			<i>Barbarea vulgaris</i>	Exotic	<1
			<i>Cardamine hirsuta</i>	Exotic	<1

Table 2. Parameter estimates for the most parsimonious general linear mixed-effects model based on likelihood ratio tests for species cover and phenology response variables as explained by warming and species' origin at each site. All models included species as a random intercept. All parameters significantly different from zero are bolded. Positive origin effects indicate greater values for native species relative to exotic species. Interactions and additional treatments (insects, mammals) are only included when statistically significant (Appendix B).

Response	Northern Forest Clearing			Southern Old Agriculture Field		
	Estimate	Denom. df	95% CI	Estimate	Denom. df	95% CI
<i>Species Cover</i>						
Warmed	0.62	122.14	(0.16, 1.07)	-0.17	213.81	(-0.37, 0.02)
Origin	0.36	21.47	(-0.68, 1.42)	0.82	20.49	(-0.22, 1.92)
Warmed*Origin	-0.69	122.14	(-1.38, -0.07)			
Residuals		126			225	
<i>Green-up</i>						
Warmed	-1.61	103.64	(-2.74, -0.52)	0.29	251.98	(-2.87, 3.46)
Origin	6.86	18.1	(-0.69, 15.43)	10.35	17.80	(2.23, 18.33)
Residuals		115			262	
<i>Flowering Time</i>						
Warmed	-2.18	84.19	(-4.02, -0.35)	2.42	211.24	(0.76, 3.88)
Origin	-15.17	12.88	(-42.94, 9.79)	37.14	18.94	(4.00, 70.88)
Insects	2.13	84.07	(0.41, 3.96)			
Residuals		91			225	
<i>Seed Set Time</i>						
Warmed	0.82	81.09	(-1.98, 3.70)	0.89	183.53	(-2.26, 3.79)
Origin	-8.62	11.09	(-31.26, 14.66)	33.77	16.75	(-1.14, 65.48)
Residuals		87			195	

Table 3. Parameter estimates for the most parsimonious general linear mixed-effects model based on likelihood ratio tests for the herbivory response variables as explained by warming and species' origin at each site. All models included species as a random intercept. All parameters significantly different from zero are bolded. Positive origin effects indicate greater values for native species relative to exotic species. Interactions and additional treatments (insects, mammals) are only included when statistically significant (Appendix B).

Response	Northern Forest Clearing				Southern Old Agriculture Field			
	Estimate	SE	Z-Value	P-Value	Estimate	SE	Z-Value	P-Value
<i>Amount Eaten</i>								
Warmed	0.68	±0.31	2.20	0.03	-0.70	±0.28	-2.51	0.01
Origin	1.02	±0.84	1.21	0.23	1.89	±0.63	3.00	<0.01
<i>Probability Attacked</i>								
Warmed	0.07	±0.35	0.20	0.84	-0.71	±0.23	-3.07	<0.01
Origin	1.02	±0.84	1.21	0.23	2.28	±0.54	4.22	<0.01

Figure Legends

Fig. 1 Species percent cover (mean \pm 1SE) measured at the end of summer 2016 at the (a) northern forest clearing and (b) southern old agriculture field. Shaded plots represent ambient temperatures and white plots represent the warmed treatment. Data were measured for each species in each plot at each site (n=24). Warming decreased average native cover, but increased average exotic cover at the northern site ($p < 0.05$).

Fig. 2 Phenological responses measured for each species in each plot throughout the 2016 growing season at the northern site (left column) and southern site (right column). Shaded plots represent ambient temperatures and white plots represent the warmed treatment. Boxplots display the median (central line), 25th percentile (lower bound), 75th percentile (upper bound), and smallest and largest value (ends of whiskers) no larger than 1.5x the interquartile range from the upper and lower quartiles. An ANOVA ($\alpha = 0.05$) was run on each response's most parsimonious model to determine significant differences between groups. Panels (a, b) show the date where each species reached 50% of its maximum cover where warming led to earlier green-up at the northern site, but not the southern site. Exotic species also greened-up earlier than native species at the southern site, regardless of warming. This trend was also observed at the northern site, although it lacked statistical significance. Panels (c, d) show the median date of flowering, where warming led to earlier flowering at the northern site and delayed flowering at the southern site. Native species also flowered later than exotic species at the southern site. Panels (e, f) show the median date of seed set. No evidence was found for differences between groups.

Fig. 3 Percent of leaf eaten (Mean \pm 1SE) at the end of summer 2016 at the (a) northern forest clearing and (b) southern old agriculture field. Shaded plots represent ambient temperatures and

540 white plots represent the warmed treatment. Data were measured for four leaves on three
541 individuals of each species in each plot (n=12) for each site. An ANOVA ran on the most
542 parsimonious model shows warming increased the amount of leaf herbivory at the northern site
543 and decreased the amount of leaf herbivory at the southern site. Native species also experienced
544 a greater amount of herbivory than exotic species at the southern site.

Fig. 1

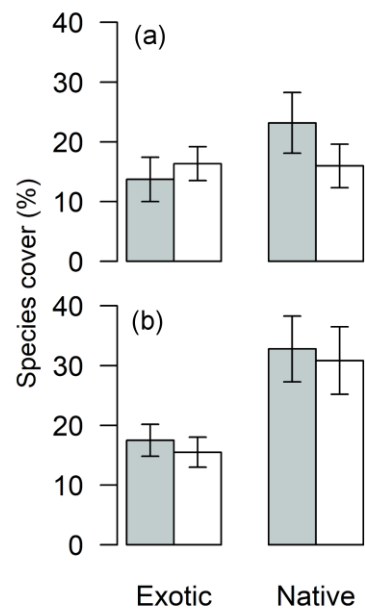


Fig. 2

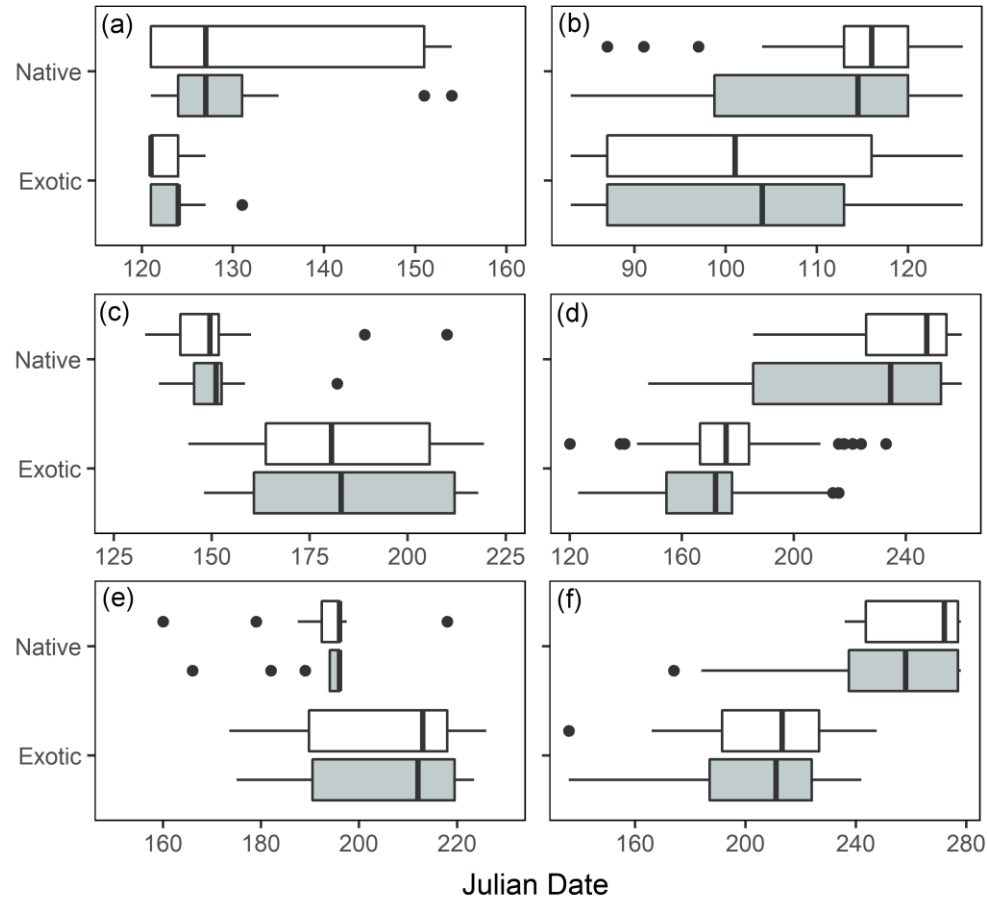
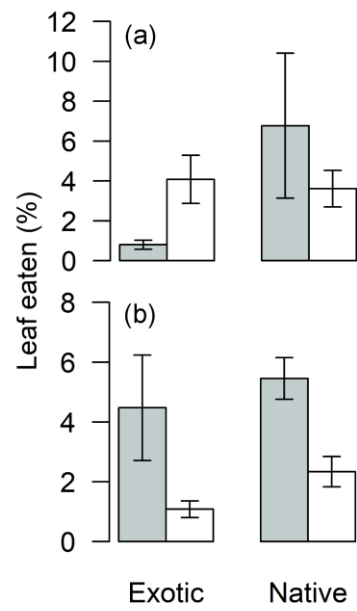


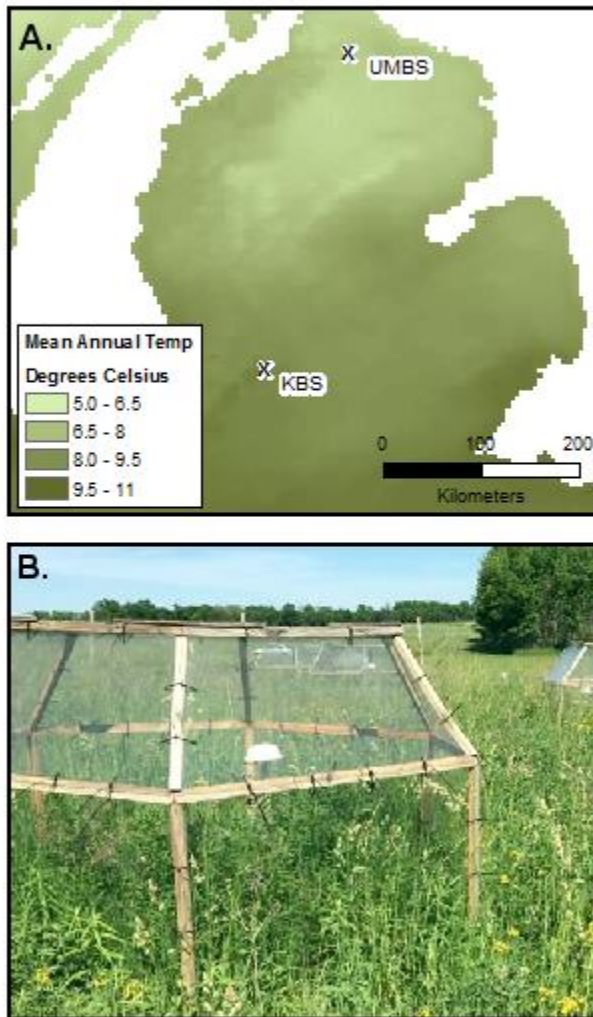
Fig. 3



Electronic Supplemental Material

Appendix A. Locations of experimental sites.

Fig. S1 A. Map of mid-western United States with study site locations and 30-year temperature normal from 1981-2010 (PRISM Climate Group 2013). B. Open-top chamber experiment in southern old-agriculture field located at Kellogg Biological Station.



Appendix B. Additional experimental treatment methods for long-term herbivory experiment.

Additional Treatment Methods

As part of the long-term experiment, we reduced insect herbivory through the application of Merit-75 WP insecticide at a concentration of 0.031 g/L with 1.2 L applied for all insect exclusion plots at each site every 13-15 days while measurements were taken during the growing season (Bayer Environmental Science, Research Triangle Park, NC). An equal amount of water (1.2 L) was applied evenly across plots where insects were not reduced within the same treatment session in order to reduce any unequal effects of added water in the insecticide to the plants. All insecticide applications were performed to dry vegetation during early morning low-wind conditions and no precipitation in the following 24 hours. Care was taken to spray all plants across sub-canopies within each plot, such that ground dwelling plants also received treatment.

We also reduced small mammal herbivory by installing 91cm tall, 1.3 cm gauge mesh hardware cloth around 12 plots at each site at the beginning of the experiment. The cloth was buried 20-30 cm beneath the ground (Brown and Davidson 1977). The fencing was reinforced in Spring 2016 by attaching a second layer of hardware cloth along the ground to reduce plot access to burrowing mammals (Nutrient Network: A Global Research Cooperative, 2008). We tested the integrity of the fences during three separate 72-hour events at each site and in each year. During each sampling event we set Sherman live traps in the fenced and unfenced plots baited with peanut butter and oats (H. B. Sherman Traps, Tallahassee, FL: 6.5 2.0 × 2.5 cm). The traps were checked for captures and reset every 8 hours.

References (Appendix A)

Brown, J. H. and Davidson, D. W. (1977). Competition between seed-eating ants and rodents in desert ecosystems. *Science*, 196(4292), 880–882.

Nutrient Network: A Global Research Cooperative. (2008, March). “Building a Fence.”

University of Minnesota Department of Ecology, Evolution, and Behavior. Retrieved from http://www.nutnet.umn.edu/files/nutnet/NutNet_FenceBuilding.pdf

Appendix C. Summary of likelihood ratio tests ($\alpha=0.05$) used to determine the most parsimonious mixed-effects model at each site.

Table S1. Model comparisons and results for Species Cover, Emergence, Flowering Time, and Timing of Seed Set. All models included species identification as a random effect. Significant results listed in bold.

Fixed Model Comparisons	Northern Forest Clearing Summary Results	Southern Old Agriculture Field Summary Results
<i>Species Cover (2015)</i>		
[M1]Cover~Warmed+Origin+Warmed*Origin+Insects+Mammals vs.	$\chi^2=0.35$; df=1;	$\chi^2=0.90$; df=1;
[M2]Cover~ Warmed+Origin+Warmed*Origin+Insects	P=0.55	P =0.34
[M1]Cover~Warmed+Origin+Warmed*Origin+Insects vs.	$\chi^2=0.30$; df=1;	$\chi^2=0.84$; df=1;
[M2]Cover~Warmed+Origin+Warmed*Origin	P =0.58	P =0.36
[M1]Cover~Warmed+Origin+Warmed*Origin vs.	$\chi^2=0.60$; df=1;	$\chi^2=2.46$; df=1;
[M2]Cover~Warmed+Origin	P =0.44	P =0.12
<i>Species Cover (2016)</i>		
[M1]Cover~Warmed+Origin+Warmed*Origin+Insects+Mammals vs.	$\chi^2=0.10$; df=1;	$\chi^2=2.05$; df=1;
[M2]Cover~ Warmed+Origin+Warmed*Origin+Insects	P =0.75	P =0.15
[M1]Cover~Warmed+Origin+Warmed*Origin+Insects vs.	$\chi^2=1.11$; df=1;	$\chi^2=0.04$; df=1;
[M2]Cover~Warmed+Origin+Warmed*Origin	P =0.29	P =0.85
[M1]Cover~Warmed+Origin+Warmed*Origin vs.	$\chi^2=4.23$; df=1;	$\chi^2=0.76$; df=1;
[M2]Cover~Warmed+Origin	P =0.04	P =0.38
<i>Emergence</i>		
[M1]Emergence ~Warmed+Origin+Warmed*Origin+Insects+Mammals vs.	$\chi^2=0.15$; df=1;	$\chi^2=1.73$; df=1;
[M2]Emergence~ Warmed+Origin+Warmed*Origin+Insects	P =0.69	P =0.19
[M1]Emergence~Warmed+Origin+Warmed*Origin+Insects vs.	$\chi^2=2.10$; df=1;	$\chi^2=1.07$; df=1;
[M2]Cover~Warmed+Origin+Warmed*Origin	P =0.15	P =0.30
[M1]Emergence~Warmed+Origin+Warmed*Origin vs.	$\chi^2=2.51$; df=1;	$\chi^2=1.75$; df=1;
[M2]Emergence~Warmed+Origin	P =0.11	P =0.19
<i>Flowering Time</i>		
[M1]Flowering Time~ Warmed+Origin+Warmed*Origin+Insects+Mammals vs.	$\chi^2=0.32$; df=1;	$\chi^2=0.33$; df=1;
[M2]Flowering Time~ Warmed+Origin+Warmed*Origin+Insects	P =0.57	P =0.57
[M1]Flowering Time~ Warmed+Origin+Warmed*Origin+Insects vs.	$\chi^2=5.40$; df=1;	$\chi^2=1.87$; df=1;
[M2]Flowering Time~Warmed+Origin+Warmed*Origin	P =0.02	P =0.17
[M1]Flowering Time~Warmed+Origin+Warmed*Origin vs.	$\chi^2=0.65$; df=1;	$\chi^2=0.37$; df=1;
[M2]Flowering Time~Warmed+Origin	P =0.42	P =0.54
<i>Timing of Seed Set</i>		
[M1]Seed Set~ Warmed+Origin+Warmed*Origin+Insects+Mammals vs.	$\chi^2=0.08$; df=1;	$\chi^2=0.05$; df=1;
[M2]Seed Set~ Warmed+Origin+Warmed*Origin+Insects	P =0.78	P =0.82
[M1]Seed Set~Warmed+Origin+Warmed*Origin+Insects vs.	$\chi^2=0.39$; df=1;	$\chi^2=0.32$; df=1;
[M2]Seed Set~Warmed+Origin+Warmed*Origin	P =0.53	P =0.57
[M1]Seed Set~Warmed+Origin+Warmed*Origin vs.	$\chi^2=0.00$; df=1;	$\chi^2=0.02$; df=1;
[M2]Seed Set ~Warmed+Origin	P =0.95	P =0.89

Table S2. Model comparisons and results for Amount of Leaf Eaten and Probability of Attack.

All models included species identification and individual plant identification as random effects.

Significant results listed in bold.

Fixed Model Comparisons	Northern Forest Clearing	Southern Old Agriculture Field
	Summary Results	Summary Results
<i>Leaf Herbivory (Amount of Leaf Eaten)</i>		
[M1]Leaf Herbivory~Warmed+Origin+Warmed*Origin+Mammals vs.	$\chi^2=2.43$; df=1;	$\chi^2=1.41$; df=1;
[M2] Leaf Herbivory~Warmed+Origin+Warmed*Origin	P =0.12	P =0.24
[M1]Leaf Herbivory~Warmed+Origin+Warmed*Origin vs.	$\chi^2=0.25$; df=1;	$\chi^2=0.76$; df=1;
[M2]Leaf Herbivory~Warmed+Origin	P =0.62	P =0.38
<i>Leaf Herbivory (Probability of Attack)</i>		
[M1]Leaf Herbivory~Warmed+Origin+Warmed*Origin+Mammals vs.	$\chi^2=0.38$; df=1;	$\chi^2=1.70$; df=1;
[M2]Leaf Herbivory~Warmed+Origin+Warmed*Origin	P =0.53	P =0.19
[M1]Leaf Herbivory~Warmed+Origin+Warmed*Origin vs.	$\chi^2=0.45$; df=1;	$\chi^2=0.05$; df=1;
[M2]Leaf Herbivory~Warmed+Origin	P =0.50	P =0.81

Appendix D. Boxplots of individual species percent cover in warmed and ambient plots after two years of treatment. Boxplots display the median (central line), 25th percentile (lower bound), 75th percentile (upper bound), and smallest and largest value (ends of whiskers) no larger than 1.5x the interquartile range from the upper and lower quartiles. Triangles represent individual data points (e.g. one point = one plot). Species are arranged by origin and from most abundant to least abundant (left → right; top → bottom) at each site.

Fig. S2 Boxplots for individual native species' cover for native at the Northern Forest Clearing located at the University of Michigan Biological Station in Pellston, Michigan, USA.

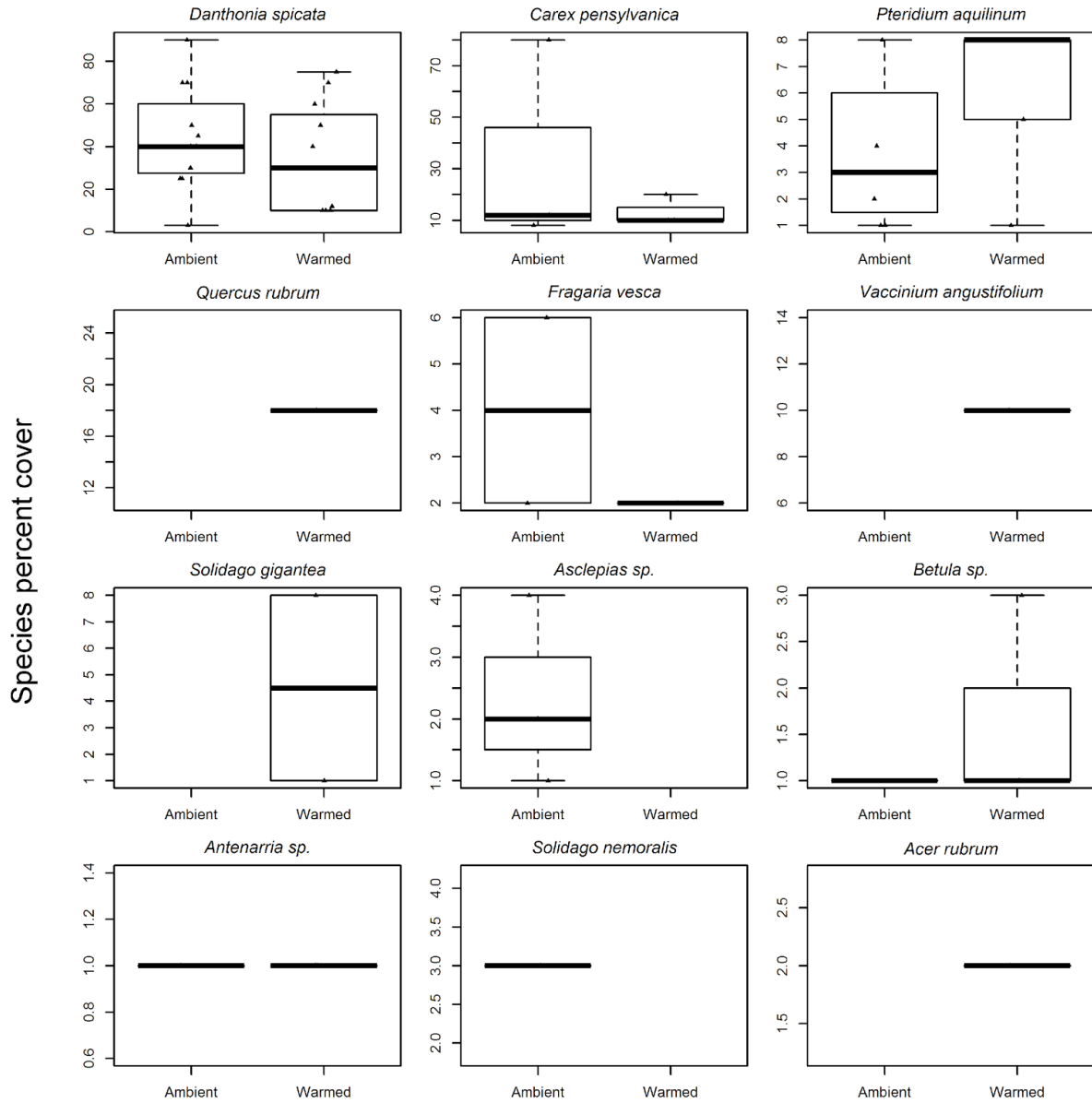


Fig. S3 Boxplots for individual exotic species' cover at the Northern Forest Clearing located at the University of Michigan Biological Station in Pellston, Michigan, USA.

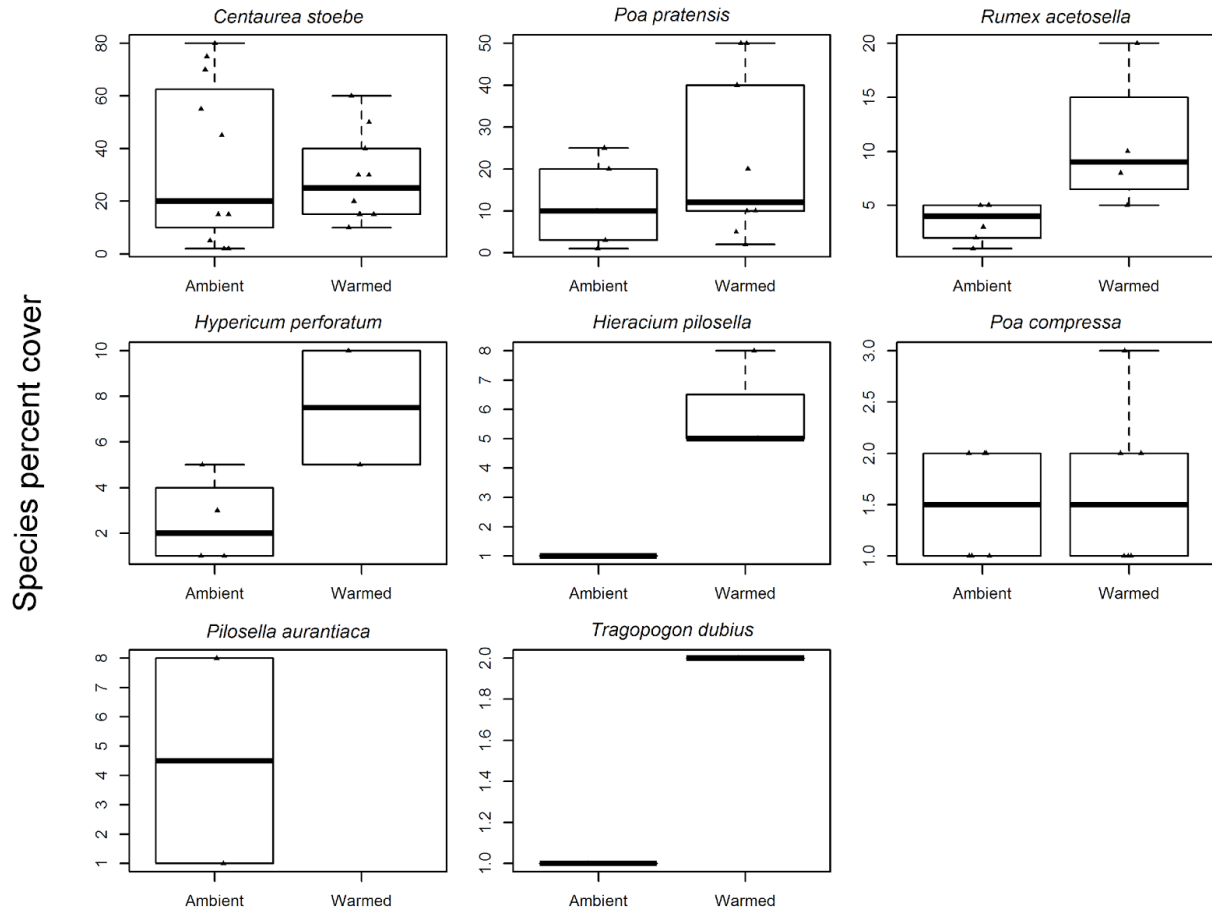


Fig. S4 Boxplots for individual native species' cover at the Old Agriculture Field located at Kellogg Biological Station in Hickory Corners, Michigan, USA.

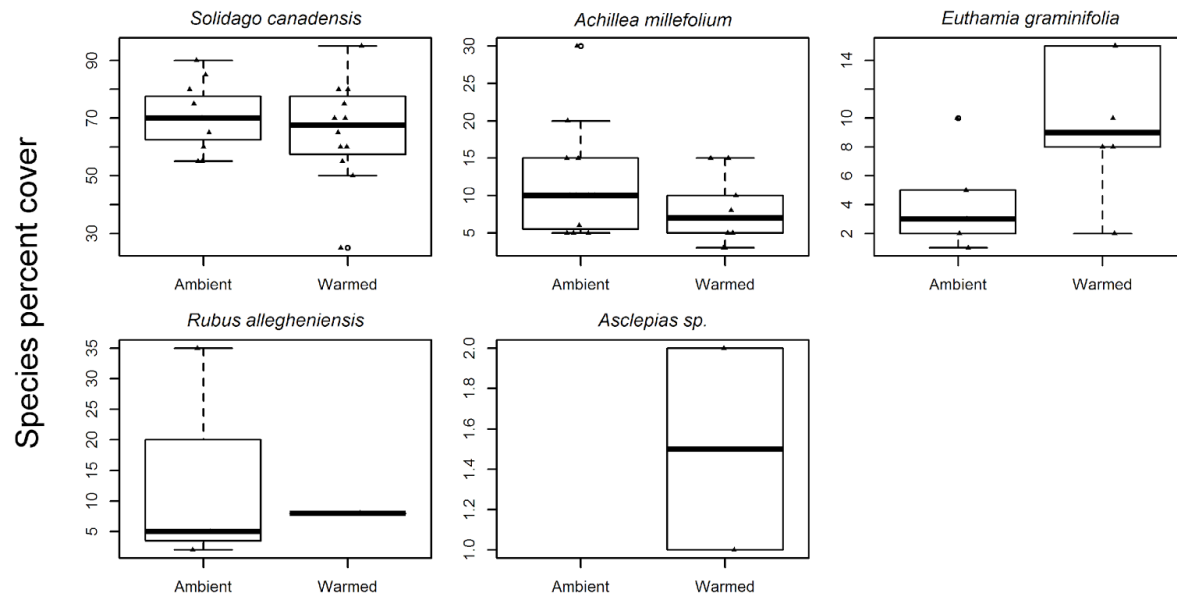


Fig. S5 Boxplots for individual exotic species' cover at the Old Agriculture Field located at Kellogg Biological Station in Hickory Corners, Michigan, USA.

