

## Highlights

- $\delta D_{\text{wax}}$  of topsoil *n*-acids was characterised 13 northern boreal sites
- Vegetation *n*-acid abundances and distributions were characterised
- Mosses dominate *n*-acids contributions to northern boreal soils
- Mean  $\epsilon_{\text{wax}/\text{MAP}}$  is  $-93 \pm 10\text{‰}$ ,  $-101 \pm 11\text{‰}$  and  $-95 \pm 11\text{‰}$  for *n*-C<sub>24,26,28</sub> acids, respectively
- Smaller  $\epsilon_{\text{wax}/\text{MAP}}$  values are observed at higher latitudes

1    **Net fractionation of hydrogen isotopes in *n*-alkanoic acids from soils**  
2    **in the northern boreal forest**

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## Abstract

Plant-derived *n*-alkyl lipids are well-preserved in sedimentary archives, and their stable hydrogen isotope ratio ( $\delta D_{\text{wax}}$ ) is a proxy for precipitation  $\delta D$  and climate. Net fractionation of H isotopes between plant source water and *n*-alkyl lipids ( $\epsilon_{\text{wax/water}}$ ) is the largest uncertainty for interpreting this proxy and depends on plant type and environment. Although popular proxies, *n*-alkanoic acids (*n*-acids) are less frequently calibrated in modern environments than *n*-alkanes. We constrain the net fractionation ( $\epsilon_{\text{wax/MAP}}$  – Mean Annual Precipitation) for the northern boreal forest based on *n*-C<sub>24,26,28</sub> acids in topsoils from a 13-site network in Yukon, Alaska and Northwest Territories (60-68°N). *n*-Acid homologue distributions for common boreal plants (gymnosperm trees, shrubs, forbs, C3 grasses and mosses) show that soil *n*-acids are dominated by mosses, but with a reduced carbon preference index compared to fresh mosses, possibly owing to post-depositional degradation. Regionally averaged  $\epsilon_{\text{wax/MAP}}$  values are  $-93 \pm 10\text{‰}$ ,  $-101 \pm 11\text{‰}$  and  $-95 \pm 11\text{‰}$  for *n*-C<sub>24,26,28</sub> acids, respectively, which are similar to values reported from low and high latitude regions. We find that  $\epsilon_{\text{wax/MAP}}$  for *n*-C<sub>26,28</sub> acids (but not *n*-C<sub>24</sub>) is significantly ( $p \leq 0.05$ ) smaller with increasing latitude and decreasing mean annual temperature, factors that covary across our network. Since latitude is typically known for ancient deposits, we parsed the dataset to obtain latitude-specific  $\epsilon_{\text{wax/MAP}}$  values for 60-64°N ( $-107 \pm 8\text{‰}$  for *n*-C<sub>26</sub>; and  $-101 \pm 8\text{‰}$  for *n*-C<sub>28</sub>) and 65-68°N ( $-94 \pm 11\text{‰}$  for *n*-C<sub>26</sub>; and  $-89 \pm 10\text{‰}$  for *n*-C<sub>28</sub>). These soil-derived values are relevant for hydroclimate reconstructions using fossil *n*-acids in paleosols from a comparable paleoflora and latitude.

**Keywords:** northern boreal forest; plant waxes; *n*-alkanoic acids; stable hydrogen isotopes; compound-specific isotope analysis; net fractionation

## 1. Introduction

Stable isotope ratios of hydrogen (D/H) and oxygen ( $^{18}\text{O}/^{16}\text{O}$ ) in precipitation are well-established tracers for hydroclimatic variables, including precipitation amount and air temperature (Dansgaard, 1964; Craig and Gordon, 1965). Hydrogen isotopes derived from meteoric waters are preserved in the long straight-chain hydrocarbons (*n*-alkanoic acids, *n*-alkanes, and *n*-alcohols) that make up the cuticular waxes of plants (Hou et al., 2008; Sachse et al., 2012; Wilkie et al., 2013), which has made fossil plant waxes a popular proxy in paleo-hydroclimate studies. Fossil plant waxes are well-preserved in sedimentary archives (e.g., relict permafrost, paleosols, lacustrine sediment cores, etc.) over geologic timescales (e.g. Yang and Huang, 2003; Tierney et al., 2008; Vonk et al., 2017) owing to their hydrophobicity, inertness, and resistance to biodegradation (Huang et al., 1997; Nguyen Tu et al., 2017). Long-chain (>20 carbon atoms) *n*-alkanoic acids (hereafter *n*-acids) with a characteristic predominance of even-number C-chains, and *n*-alkanes with a predominance of odd-number C-chains, are both derived from a common precursor (Zhou et al., 2010). Together these compounds form major constituents of the waxy coating within and on top of the cuticle of plant leaves

and, thus, both are used as biomarkers for terrestrial plants (Eglinton and Hamilton, 1963). *n*-Alkyl hydrogens do not exchange readily under 150°C (Sessions et al., 2004) allowing isotopic signals of synthesis to be retained in sedimentary storage.

The hydrogen isotopic composition of plant waxes ( $\delta D_{\text{wax}}$ ) is offset from the hydrogen isotopic composition of precipitation ( $\delta D_{\text{precip}}$ ) by a large negative fractionation. This offset is referred to as the net (or apparent) fractionation ( $\epsilon_{\text{wax/precip}}$ ) and varies regionally with climate and vegetation type (Sachse et al., 2012). Terrestrial plants absorb water from soil moisture, which is ultimately derived from precipitation. Evaporation can enrich deuterium (D) in soil water, however many plants do not use evaporatively-enriched soil water (Feakins and Sessions, 2010), although it may be a factor in some shallow rooted plants such as grasses (Smith and Freeman, 2006). Hydrogen isotopes are generally not fractionated during uptake of source water by roots (White et al., 1985; Ehleringer and Dawson, 1992; Roden et al., 2000a); however, transpiration results in D-enrichment of leaf waters (Feakins and Sessions, 2010) and the magnitude of this enrichment depends on relative humidity, isotopic composition of the ambient water vapour, and leaf geometry (Barbour et al., 2004; Kahmen et al., 2008; Sachse et al., 2012). *n*-Acid precursors inherit H isotopes from leaf water, but a series of biosynthetic fractionations then results

in D-depletion of the resulting *n*-alkyl lipids relative to leaf water (Chikaraishi et al., 2004a; Zhou et al., 2010; Sachse et al., 2012).

Large variability in  $\epsilon_{\text{wax/precip}}$  has been noted between different groups of plants and photosynthetic pathways. On average, *n*-C<sub>29</sub> alkanes produced by shrubs are more enriched in deuterium compared to trees, forbs and graminoids (Sachse et al., 2012) given the same source water. Therefore, changes in vegetation type and abundance within a catchment can result in variability in sedimentary  $\delta\text{D}_{\text{wax}}$  that is unrelated to  $\delta\text{D}_{\text{precip}}$  (Fornace et al., 2014). For  $\delta\text{D}_{\text{wax}}$ -based paleoclimate reconstructions, this underscores the need to use  $\epsilon_{\text{wax/precip}}$  values that are appropriate to the plant community and climate, to enable robust estimates of  $\delta\text{D}_{\text{precip}}$  (Feakins, 2013; Nichols et al., 2014). The  $\epsilon_{\text{wax/precip}}$  parameter has been calibrated extensively for low and mid latitude ecosystems, but there are only a few high-latitude examples, limiting our ability to interpret high-latitude  $\delta\text{D}_{\text{wax}}$  records.

Previous calibration studies have constrained  $\epsilon_{\text{wax/precip}}$  in various ways. Growth-chamber experiments have tested select species, under known conditions where irrigation water and other factors (e.g., light, humidity) are controlled, and the  $\delta\text{D}_{\text{wax}}$  of new foliage is measured and used to determine  $\epsilon_{\text{wax/water}}$  (e.g., Yang et al., 2009). Natural ecosystem surveys have compared the  $\delta\text{D}_{\text{wax}}$  of leaves and  $\delta\text{D}$  of plant water (e.g., xylem water) to calculate  $\epsilon_{\text{wax/xylem}}$  (e.g., Feakins and Sessions, 2010; Daniels et al., 2017). Other sedimentary

approaches have compared the  $\delta D_{\text{wax}}$  of integrated waxes in surface (*ca.* modern) sediments with a best estimate of local  $\delta D_{\text{precip}}$  in order to estimate  $\epsilon_{\text{wax/precip}}$  for the plant community as represented in sediments (e.g., Shanahan et al., 2013; Wilkie et al., 2013). All are valid approaches with different strengths and limitations. For example,  $\epsilon_{\text{wax/water}}$  can be directly determined for specific plants in growth-chamber experiments and ecosystem surveys, but fossil waxes in the geologic record are subject to other uncertainties related to post-depositional processes and wax inputs from other vegetation. Conversely,  $\epsilon_{\text{wax/precip}}$  estimates from sedimentary waxes (e.g., soil, lake sediment) integrate all wax inputs and pre-depositional processes, but source water  $\delta D$  cannot be directly measured and, thus, must be estimated to constrain  $\epsilon_{\text{wax/precip}}$ . The  $\delta D$  value of amount-weighted Mean Annual Precipitation ( $\delta D_{\text{MAP}}$ ) is commonly used as an approximation for source water  $\delta D$  (e.g., meta-analysis by Sachse et al., 2012). For future reference, the term ' $\epsilon_{\text{wax/MAP}}$ ' is used when referring to net fractionation estimates expressed relative to  $\delta D_{\text{MAP}}$  (measured or modelled).

In a survey of lake sediment cores in dwarf shrub-tundra catchments on Baffin Island (62-74°N), Canadian High Arctic, Shanahan et al. (2013) found a mean  $\epsilon_{\text{wax/MAP}}$  of -61‰, based on the mean  $\delta D_{\text{wax}}$  of *n*-C<sub>26</sub> and -C<sub>28</sub> acids. This value is smaller than what is typical for herbaceous plants in low and middle latitudes (Sachse et al., 2012). Shanahan et al. (2013) linked this observation to an evapotranspiration effect on leaf water that was first observed in growth

chamber experiments by Yang et al. (2009) who found *n*-alkanes from deciduous conifers under 24-hour light (simulating High Arctic conditions) were up to 40‰ enriched compared to foliage grown in diurnal light conditions.

At a comparable latitude in northeastern Siberia (67.5°N), Wilkie et al. (2013) found a  $\epsilon_{\text{wax/MAP}}$  of −95‰ for *n*-C<sub>30</sub> acids for *ca.* modern lake sediments of Lake El'gygytgyn, surrounded by tundra plants including lichen and herbaceous taxa. From lacustrine core top sediments of tundra lakes in northern Alaska (68.5°N), Daniels et al. (2017) observed larger  $\epsilon_{\text{wax/MAP}}$  values of −118‰ for *n*-C<sub>25-33</sub> alkanes, and −114‰ for *n*-C<sub>22-30</sub> acids. Sachse et al. (2004) report the largest  $\epsilon_{\text{wax/MAP}}$  values from lacustrine sediments from taiga and tundra catchments in Fennoscandia (61-69°N) with a mean of −125‰ for *n*-C<sub>29</sub> alkanes.

With the exception of the study on Baffin Island (Shanahan et al., 2013), net fractionation estimates from most high-latitude sites (>60°N) are comparable in value to observations from mid to low latitude sites with values ranging from −90 to −130‰ for both *n*-alkanes (Chikaraishi et al., 2004b; Pagani et al., 2006; Sachse et al., 2006; Smith and Freeman, 2006; Hou et al., 2007; Kahmen et al., 2013; Gao et al., 2014; Feakins et al., 2016b; Freimuth et al., 2017) and *n*-acids (Hou et al., 2007; Feakins et al., 2014, 2016b; Gao et al., 2014; Freimuth et al., 2017). Additionally, studies spanning large latitudinal gradients do not appear to support a strong relationship between latitude and



net fractionation (Sachse et al., 2006; Liu et al., 2016), which suggests that light-dependent controls on net fractionation may be unique to certain high-latitude environments. More work is needed to understand this diversity in high-latitude environments.

One high-latitude ecosystem that remains largely unstudied in the *n*-alkyl literature is the northern boreal forest. The boreal forest covers an estimated 1.2 billion hectares of the Earth, spanning large portions of North America and Eurasia (Soja et al., 2007). Furthermore, the boreal ecosystem was present in the Western sub-Arctic during the Holocene and past interglacials (Schweger et al., 2011; Kaufman et al., 2012), and at higher latitudes during the Pliocene (Csank et al., 2011, 2013), which makes it an important target for modern calibration studies to exploit fossil boreal  $\delta D_{wax}$  as a paleoenvironmental proxy.

We address this knowledge gap with a study of *n*-acids from modern vegetation and top-soils from southern Yukon to the northernmost extent of boreal treeline in Northwest Territories. We focus specifically on *n*-acids because Pinaceae gymnosperm trees (e.g., *Picea*, *Pinus* and *Larix*), which dominate the boreal canopy, are known to produce negligible amounts of *n*-alkanes (Diefendorf et al., 2011) and, thus, are likely to be unrepresented in soil *n*-alkanes (Schäfer et al., 2016). Soil surveys have been widely used for calibration of  $\delta D_{wax}$  elsewhere (Chikaraishi and Naraoka, 2006; Jia et al., 2008; Peterse et al., 2009; Rao et al., 2009; Bai et al., 2011; Ponton et al., 2014;

163 Schwab et al., 2015) to understand the plant wax signals incorporated into  
164 soils. Our research objectives are two-fold: (1) to characterize the *n*-acid chain  
165 length distributions of common boreal vegetation types in order to understand  
166 the major contributor(s) to sedimentary lipid pools; and (2) to quantify the net  
167 fractionation of *n*-acids in northern boreal soils. This work will inform future  
168 paleoclimate studies based on fossil *n*-acids sourced from boreal paleoflora.

## 2. Methods

### 2.1 Study area

Soil and vegetation samples were collected from thirteen locations in the Yukon, Northwest Territories, and Alaska (Fig. 1), spanning 60.3 to 68.3°N. The climate of the study region is cold and semi-arid, characterized by short, cool summers and long, cold winters (Fig. 2). Mean annual temperature (MAT) ranges from −8.4 in the north to −1.7°C in the south, with average daily temperatures above freezing from May to September based on ERA-Interim data (Fig. 1; Table 1; Dee et al., 2011). Average growing season temperatures range from 4 to 12°C. Daylight hours at the summer solstice range from 19 hours in southern Yukon to 24 hours in the north (Wahl, 2004). The northern half of the site network (>65°N) falls within the continuous permafrost zone, and the more southern sites are in the discontinuous or sporadic permafrost zones (Brown et al., 1997).

As our study region encompasses a large range of latitudes and mean temperatures, the start and end points of the growing season (timing and duration of leaf wax synthesis) also vary regionally. The growing season is primarily limited by minimum temperatures above the freezing mark. In the southern end of our transect (e.g., Whitehorse) minimum daily temperatures are above freezing from May 18 to September 24 (duration = 129 days), and in the northern end (e.g., Inuvik) from May 27 to September 23 (duration = 119

days) (Dee et al., 2011). However, other abiotic factors such as sunlight and ground temperatures (influencing unfrozen water availability) also influence the timing of biosynthesis during the growing season.

The Alaska Range and St. Elias Mountains represent a significant barrier to the advection of Pacific moisture into continental Yukon. Areas of low elevation receive 250-300 mm of precipitation annually, while higher elevations receive more, up to 400-500 mm, based on WorldClim 2 data (Fick and Hijmans, 2017). Over 75% of precipitation falls between May and September (Wahl, 2004). Peak summer (JJA) average relative humidity differs by 13% (62 to 74%) between sites based on CliMond data (Table 1, Fig. 1; Kriticos et al., 2012).

The study sites are mature subarctic woodlands, which are broadly representative of the northern boreal forest ecology and span a range of latitudes and climate conditions which allows us to examine the range of variability in net fractionation within this ecotype. The sites are primarily spruce (*Picea glauca* and *Picea mariana*) and moss dominated woodlands characterized by moderately open canopies. *Pinus contorta* and *Larix laricina* trees are dominant or co-dominant at two of our sites. The sub-canopy typically includes a thick (5-15 cm) moss ground cover with patches of Poaceae grass and Ericaceae forbs, and sparse shrub cover including alder (*Alnus* sp.), willow (*Salix* sp.) and dwarf birch (*Betula nana*). The mosses associated with subarctic woodlands are diverse and include *Sphagnum* hummock varieties (e.g.,

*Sphagnum fuscum* and *S. balticum*) and feathermoss varieties (e.g.,  
*Pleurozium schreberi*, *Ptilium crista-castrensis*, *Tomentypnum nitens*, and  
*Dicranum polysetum*) with the assemblage being influenced by soil moisture  
and woodland successional stage (Black and Bliss, 1978; Turetsky et al., 2012).  
Most sites are relatively flat with moist but unsaturated, organic-rich soils, and  
underlain by clastic parent material (clay, silts).

## 2.2 Sampling methods

Soil samples were collected from most sites in July 2014, and from the LC  
and SH sites in July 2016 and SP site in July 2017. Soil samples were collected  
to examine the homologue distribution and hydrogen isotope ratios of long  
chain ( $C_{24}$ ,  $C_{26}$ , and  $C_{28}$ ) *n*-acids. Soils were collected from ~30 cm soil pits in 3-  
10 cm intervals, depending on the thickness of the O-horizon; the O- and A-  
horizons were collected separately. Soil pits were dug in open areas within 2 m  
of the canopy dominant tree, and in close proximity to shrubs and forbs and  
these soil pit criteria were held constant across our site network. A single pit  
was sampled at most sites, in order to compare the O and A horizons at a  
representative location. We do not describe within-site heterogeneity such as  
may vary with plant cover, plant growth-limiting factors, or microtopography.  
Instead, we prioritized a large number of sites across a long transect to  
characterise latitudinal variability in net fractionation across the northern  
boreal forest.

Vegetation samples were collected at the same sites to examine the distribution of *n*-acid homologues and to determine which plant types are contributing most to soils. Fresh leaves or needles with minimal evidence of damage or decomposition were collected from up to three individual dominant or co-dominant shrubs, trees, mosses, grasses and forbs within a ~5 m radius of the soil pit. The isotopic composition of vegetation samples was not measured. All vegetation and soil samples were collected using nitrile gloves and stored in Whirlpak™ bags, chilled over ice in a field cooler, and shipped frozen to the laboratory for processing and analysis.

### 2.3 Vegetation sample classification

Vegetation samples were classified as tree, shrub, moss, grass or forb. An inventory of plant taxa sampled at each site is provided in the supplement (Table S1). All trees were gymnosperms. White spruce (*Picea glauca*, *n* = 33; here, ‘*n*’ refers to the number of individual *P. glauca* trees that were sampled across the regional network) was dominant at all sites except TSI, MYO, DHP174 and SP. Larch (*Larix Laricina*, *n* = 1) trees are dominant at the TSI site. Black spruce (*Picea mariana*, *n* = 3) was co-dominant with white spruce at MYO and DHP174. Lodgepole pine (*Pinus contorta*, *n* = 1) was co-dominant with white spruce at SP. Shrub cover was sparse and variable, and included willow (*Salix sp.*, *n* = 20), alder (*Alnus sp.*, *n* = 5) or dwarf birch (*Betula nana*, *n* = 7). Mosses common in northern boreal woodlands are diverse, and an

inventory of all moss taxa at each site was not attempted. Fresh mosses ( $n = 12$ ) were sampled from the top of the O horizon of our soil pits, and was mostly identified as the red-stemmed feathermoss *Pleurozium schreberi* (Hylocomiaceae); minor co-mingled fractions of *Tomentypnum nitens*, *Ptilium crista-castrensis* and *Dicranum polysetum* were also observed. Grasses in this region are Poaceae and use the C3 metabolic pathway ( $n = 13$ ). All forbs were Ericaceae, including crowberry (*Empetrum nigrum*,  $n = 1$ ), lingonberry (*Vaccinium vitis-idaea*,  $n = 2$ ), blueberry (*Vaccinium uliginosum*,  $n = 1$ ), and Labrador tea (*Rhododendron sp.*,  $n = 3$ ).

#### 2.4 Lipid extraction

Soil samples were freeze dried in a Labconco freezone 2.5 unit.  $n$ -Acid concentrations varied between soil samples. Initially, 5 g of dry, homogenized sediment was subsampled for lipid extraction. Following the initial quantification of  $n$ -acids by GC-FID (described below), additional sediment (up to 60 g) was sometimes required to yield sufficient  $n$ -acid amounts for the compound-specific isotope analysis.

Soil bound-lipids were extracted with 9:1 (v/v) Dichloromethane (DCM):Methanol (MeOH) by microwave-assisted heating (Milestone Ethos Up unit) at 70°C for 20 minutes, and continuous magnetic stirring, a method selected for optimal recovery (Chávez-Lara et al., 2018). Samples were then centrifuged at 1800 RPM for 10 minutes to settle the sediment, and the

supernatant was removed and retained. The sediment sample was rinsed three to five additional times with DCM-MeOH, centrifuged and transferred to the collection vial. Excess solvent was evaporated under a gentle stream of N<sub>2</sub> gas.

Foliage samples were dried in paper bags in an oven at 60°C for 48 hours. Chopped, dry foliage (1 g) was submerged in DCM:MeOH (9:1, v/v) and agitated by Pasteur pipette pumping and the extract removed (repeated three times) to remove the epicuticular and intracuticular waxes (similar to the methods of Feakins and Sessions, 2010; Feakins et al., 2016a).

The total lipid extract was separated by column chromatography in a 5.75” borosilicate glass Pasteur pipette packed with Phenomenex 60 Å NH<sub>2</sub> sepra stationary phase. The neutral fraction was eluted with 2:1 DCM:isopropanol, followed by the acid fraction with 4% formic acid in diethyl ether. The acid fraction was methylated to Fatty Acid Methyl Esters (FAMEs) using 5% hydrochloric acid and 95% methanol ( $\delta D_{MeOH} = -246.6\text{‰}$ , following methodology in Lee et al., 2017) at 60°C for 12 hours. The mixture was cooled and ~1 mL MilliQ water was added. Then, 1 mL hexanes was added and shaken vigorously for 30 s, to partition the FAMEs into the hexanes and this liquid-liquid extraction was repeated (three times) to ensure recovery. The extract was passed through a column of anhydrous sodium sulfate to remove any water. The extract was purified by column chromatography with 5% water-deactivated 100-200 mesh silica gel, and the FAMEs were eluted with 3 column



rinses of hexanes and then DCM. The saturated FAMES were further purified by column chromatography with silver nitrate on silica gel (+230 mesh), also eluted with hexanes and DCM.

## *2.5 Quantification and isotope analysis*

FAMES were analysed at the University of Toronto Mississauga with a Gas Chromatograph (Thermo Trace 1310) Flame Ionization Detector (GC-FID), equipped with a Programmable Temperature Vaporizing (PTV) injector and a Rxi-5 ms column (30 m x 0.25 mm, film thickness 1  $\mu$ m). The FID results were normalised to known quantities of an in-house FAME standard (*n*-C<sub>17</sub>, -C<sub>21</sub>, -C<sub>25</sub> and -C<sub>29</sub> FAMES). For all samples, we calculated the average chain length ( $ACL = \Sigma(C_n \times n) / \Sigma C_n$ , where  $C_n$  is the abundance for chain-lengths from  $n = 20$ -34), modal chain length ( $C_{max}$ ), and the carbon preference index ( $CPI = ([C_{20, 22, 24, 26, 28, 30}] + [C_{22, 24, 26, 28, 30, 32}]) / (2 \times [C_{21, 23, 25, 27, 29, 31}])$ ), the latter a measure of degradation (Bray and Evans, 1961).

Compound-specific hydrogen isotope ratios were measured at the University of Southern California using a Thermo Scientific Trace GC equipped with a PTV inlet operated in solvent split mode and a Rxi-5 ms column (30 m x 0.25 mm x 0.25  $\mu$ m), connected via a GC Isolink with pyrolysis (1400°C), via a Conflo IV to an Isotope Ratio Mass Spectrometer (Delta V IRMS). The  $H_3^+$  factor was measured daily to check for linearity and remained close to 8 ppm  $mV^{-1}$ .  $H_2$  reference peaks were injected at the beginning and end of each sample

run on the GC-IRMS, with two used for standardization between sample and standard runs. Hydrogen isotope ratios are reported in delta notation:  $\delta D = (R_{\text{sample}} - R_{\text{standard}}) / R_{\text{standard}}$ , where R is the ratio of deuterium to protium ( $^2\text{H}/^1\text{H}$ ). Precision of replicate sample injections was typically better than 1‰. Data were normalized to the VSMOW-SLAP hydrogen isotopic scale using the A3 mix alkane standard with  $\delta D$  values ranging from -233.7 to -46.3‰ (supplied by A. Schimmelmann, Indiana University, Bloomington). The RMS error of replicate analyses of the A3 mix was typically better than 4‰ over the month of analysis. Hydrogen atoms added by methylation were corrected for by mass balance:  $\delta D_{\text{wax}} = \delta D_{\text{meas}} (2i + 2 - 3 \times \delta D_{\text{MeOH}}) / (2i - 1)$ , where  $\delta D_{\text{meas}}$  is the normalized  $\delta D$  value of the FAME and  $i$  is the number of carbon atoms in the fatty acid molecule of interest.

## 2.6 Net fractionation

Net (or apparent) fractionation was calculated for each soil sample based on the general formula:  $\epsilon_{\text{wax}/\text{MAP}} = (\delta D_{\text{wax}} + 1) / (\delta D_{\text{MAP}} + 1) - 1$ , where  $\delta D_{\text{wax}}$  is a measured value from  $n$ -acids recovered from the soils and  $\delta D_{\text{MAP}}$  (amount-weighted mean annual precipitation  $\delta D$ ) is a best estimate of average source waters available to plants for each of our sites. Compared to unweighted mean annual precipitation,  $\delta D_{\text{MAP}}$  is biased to warm-season values, reflecting higher precipitation totals in summer (Fig. 2), but also accounts for significant inputs of D-depleted snowmelt to the ground in springtime (Mackay, 1983). Further

rationale for the  $\delta D_{\text{MAP}}$  assumption is discussed in Section 4.2. We used the Online Isotopes in Precipitation Calculator (OIPC; Bowen, 2018; Bowen and Wilkinson, 2002; Bowen and Revenaugh, 2003) to estimate monthly  $\delta D_{\text{precip}}$  (Fig. 2) and  $\delta D_{\text{MAP}}$  for our sites, which is common practice in other  $\delta D_{\text{wax}}$  calibration studies (e.g. Sachse et al., 2004; Shanahan et al., 2013; Freimuth et al., 2017). The OIPC provides interpolated  $\delta D_{\text{precip}}$  estimates constrained by empirical data from GNIP (Global Network for Isotopes in Precipitation) stations, including local stations at Whitehorse, Mayo, and Inuvik (Fig. 1).  $\delta D_{\text{MAP}}$  ranges from  $-160 \pm 2\text{‰}$  at Lost Chicken, to  $-176\text{‰}$  at Inuvik, with a regional, all-site average of  $-168\text{‰}$  ( $1\sigma = 6$ ) (Table 2). This region is characterized by a large seasonal range in monthly  $\delta D_{\text{precip}}$  of  $113\text{‰}$ . Minimum  $\delta D_{\text{precip}}$  occurs in January and ranges from  $-203$  to  $-252\text{‰}$  between sites, and maximum  $\delta D_{\text{precip}}$  occurs in either June or September and ranges from  $-123$  to  $-132\text{‰}$  between sites (Fig. 2).

### 3. Results

#### 3.1 Vegetation

Vegetation samples yielded long chain  $n$ -C<sub>20</sub> to -C<sub>34</sub> acids with notable differences in chain length distribution between vegetation types (Fig. 3). All vegetation types demonstrate the expected predominance of even-over-odd C-chain  $n$ -acid homologues with mean CPI values ranging from 10.5 to 14.2. Mosses and shrubs have the shortest modal chain lengths ( $C_{\max}$  = 24 and 26 for mosses and shrubs, respectively); grasses ( $C_{\max}$  = 28) and forbs and trees have a longer dominant chain length ( $C_{\max}$  = 30). Mosses and shrubs have the lowest ACL of 24.8 and forbs had the highest ACL of 29.3 (Fig. 3, Table S2). Shrubs yielded the greatest average concentration of total long-chain  $n$ -acids ( $\sum_{C20-C34}$ ) per gram of dry foliage (716  $\mu\text{g g}^{-1}$ ), followed by forbs (398.1  $\mu\text{g g}^{-1}$ ), grasses (156.6  $\mu\text{g g}^{-1}$ ), trees (132.2  $\mu\text{g g}^{-1}$ ), and mosses (60.2  $\mu\text{g g}^{-1}$ ) (Tables S1 for inventory of sampled plants, and Table S2 for  $n$ -acid data averaged by vegetation type and site). Site-averaged ACL and CPI for most plant types are not significantly correlated with site-specific climatic (temperature, relative humidity, precipitation amount) or geographic (latitude, longitude, elevation) variables, except the ACL of shrubs which is significantly correlated ( $p \leq 0.05$ ) with latitude, longitude, elevation, mean annual temperature and annual RH (see Table S3 for all correlations).

#### 3.2 Modern soils

*n*-Acids with carbon chain-lengths between C<sub>20</sub> and C<sub>30</sub> are the most abundant in the average soil sample (averaged across all sites, equal weight given to all sites) with a modal chain length of 24, and an ACL of  $24.2 \pm 0.5$  (Fig. 3; see Table S4 for site averages, and Table S5 for all samples). Most of this distribution is centered between C<sub>22</sub> and C<sub>26</sub>, and relative abundances are especially low (3% or less) for chain-lengths greater than C<sub>30</sub>. The all-site average CPI is  $2.8 \pm 0.9$  indicating even over odd chain length predominance, but is notably smaller than the CPI values observed for fresh vegetation which were greater than 10. The same homologue distribution is also observed for all-site averages calculated separately for the O and A soil horizons (Table S6). Site-averaged ACL and CPI values calculated for the O and A horizons separately are not significantly different ( $p < 0.05$ ; 2-sample t-tests). As was true for most vegetation samples, site-averaged soil ACL and CPI values are not significantly correlated with any of the climatic or geographic variables (see Table S3).

In terms of absolute abundance, total long-chain *n*-acid ( $\sum_{C_{20}-C_{34}}$ ) concentrations were highly variable between sites, ranging from 17.2 to 267.4  $\mu\text{g g}^{-1}$  (Table S4). Likewise, absolute concentrations for individual *n*-acids also varied between sites, for example, ranging from 6-94  $\mu\text{g g}^{-1}$  for n-C<sub>24</sub>, 3-25  $\mu\text{g g}^{-1}$  for n-C<sub>26</sub>, and 2-21  $\mu\text{g g}^{-1}$  for n-C<sub>28</sub>.

Site-averaged  $n$ -acid  $\delta D_{\text{wax}}$  values ranged from  $-256$  to  $-234\text{‰}$  (mean =  $-246\text{‰}$ ,  $1\sigma = 7\text{‰}$ ) for  $n\text{-C}_{24}$ ,  $-264$  to  $-241\text{‰}$  (mean =  $-253\text{‰}$ ,  $1\sigma = 7\text{‰}$ ) for  $n\text{-C}_{26}$ , and  $-260$  to  $-238\text{‰}$  (mean =  $-248\text{‰}$ ,  $1\sigma = 6\text{‰}$ ) for  $n\text{-C}_{28}$  acids (see Table S4 for site-averages, and Table S5 for individual soil samples from each site). Site-averaged  $n\text{-C}_{24}$ ,  $\text{C}_{26}$  and  $\text{C}_{28}$  acid  $\delta D_{\text{wax}}$  values are not significantly different ( $p \leq 0.05$ ; 2-sample t-test) between the O- and A- horizon (Table S6). Site-averaged  $\delta D_{\text{wax}}$  values are not significantly correlated with the climate or geographic variables, except  $n\text{-C}_{26}$   $\delta D_{\text{wax}}$  which correlates with mean annual precipitation (Table S3).

### 3.3 Net fractionation

Net fractionation values ( $\epsilon_{\text{wax}/\text{MAP}}$ ) were calculated for soil-derived  $n\text{-C}_{24}$ ,  $\text{-C}_{26}$  and  $\text{-C}_{28}$  acids for each soil sample (O and A horizons), which were then used to calculate site averages. Site averages were used to calculate regional averages (each site contributing equally to the regional average).

Site-averaged (including O and A horizon soils)  $\epsilon_{\text{wax}/\text{MAP}}$  values range from  $-110$  to  $-71\text{‰}$  for  $n\text{-C}_{24}$ ,  $-122$  to  $-80\text{‰}$  for  $n\text{-C}_{26}$ , and  $-111$  to  $-77\text{‰}$  for  $n\text{-C}_{28}$  acids (Table 2). The regional ‘all-site’ average  $\epsilon_{\text{wax}/\text{MAP}}$  values are  $-93\text{‰}$  ( $1\sigma = 10$ ),  $-101\text{‰}$  ( $1\sigma = 11$ ) and  $-95\text{‰}$  ( $1\sigma = 11$ ) for  $n\text{-C}_{24}$ ,  $\text{C}_{26}$  and  $\text{C}_{28}$  acids, respectively (Table 2). We find no significant differences between regional  $\epsilon_{\text{wax}/\text{MAP}}$  averages for the three major  $n$ -acids (t-tests,  $p \leq 0.05$ ). Further, we find no significant differences (t-tests,  $p \leq 0.05$ ) between regional  $\epsilon_{\text{wax}/\text{MAP}}$  values when calculated

for O and A horizons separately (see regional O and A horizon  $\epsilon_{\text{wax/MAP}}$  values in Table S6).

We find spatial trends in  $\epsilon_{\text{wax/MAP}}$  across the network of sites. Site-averaged  $\epsilon_{\text{wax/MAP}}$  is significantly ( $p \leq 0.05$ ) correlated with latitude for  $n\text{-C}_{26}$  and  $\text{-C}_{28}$  ( $r = 0.62$  and  $0.56$ , respectively; Fig. 4a), with MAT for  $n\text{-C}_{26}$  and  $\text{-C}_{28}$  ( $r = -0.55$  and  $-0.59$ , respectively; Fig. 4d), and with elevation but for  $n\text{-C}_{26}$  only ( $r = -0.61$ ) (Fig. 4b). The  $\epsilon_{\text{wax/MAP}}$  values are not significantly correlated with precipitation amount, relative humidity, or mean growing season temperature (Fig. 4c, 4e, and 4f).  $\epsilon_{\text{wax/MAP}}$  values are generally smaller at higher latitudes, lower mean elevations and lower MATs – factors that tend to covary across our network of sites.

## 4. Discussion

### 4.1 *n*-Acid abundance in vegetation and soils

Although *n*-alkyl homologue distributions aren't taxonomically diagnostic (Eglinton and Hamilton, 1963), some generalisations can be made between plant groups. For example, grasses, herbs and forbs produce a high proportion of high molecular weight ( $>C_{30}$ ) *n*-alkanes and -acids compared to other plants (Eglinton and Hamilton, 1963; Diefendorf et al., 2011; Bush and McInerney, 2013), while mosses produce abundant lower molecular weight *n*-alkanes ( $C_{23}$  and  $C_{25}$ ) and *n*-acids ( $C_{22}$  and  $C_{24}$ ) (Pancost et al., 2002; Bush and McInerney, 2013; Vonk et al., 2017). These tendencies are also reflected in our modern vegetation and soil samples (Fig. 3).

A comparison of *n*-acid relative abundances from our type-averaged vegetation and average soil (0-30 cm below surface, including A and O layers) shows that the distribution of *n*-acids in soils (Fig. 3f) closely resembles that of mosses (Fig. 3d), with a  $C_{24}$  modal chain-length, followed by  $C_{22}$  and then  $C_{26}$  in relative abundance. This distribution is unique to mosses (Pancost et al., 2002). We find this pattern even when the O and A horizons are examined separately (see Table S6) and, persists when we average the data from the two horizons, suggesting that mosses dominate soils throughout the profile.

Shrubs have a modal chain-length of  $C_{26}$  and high proportions of  $C_{24}$  and  $C_{28}$ , (Fig. 3b) which is inconsistent with the homologue pattern in soils. We do



find a spatial variation in the distribution of *n*-acid homologues, but only in shrubs, which have a significantly longer ACL at higher latitudes ( $r = 0.8$ ,  $p < 0.01$ ) or colder MAT ( $r = -0.77$ ,  $p < 0.01$ ) (see Table S3 for all correlations). Other studies have noted significant correlations between plant ACL and hydroclimate gradients (e.g., Bush and McInerney, 2015; Feakins et al., 2016b), which is thought to reflect a functional role long *n*-alkyl chains have in mitigating leaf-water loss. However, the fact that soil ACL across our network of sites is poorly correlated (n.s.) with the climate and geographic variables (contrary to shrubs) is further evidence that shrubs are not a major *n*-acid contributor to boreal soils at the average site.

Grasses produce mainly *n*-C<sub>26</sub> and -C<sub>28</sub> acids (Fig. 3a), which is inconsistent with the soil *n*-acid pattern, and suggests grasses are not a major *n*-acid source. This is not surprising, as grasses prefer dry, raised tussocks which represent a negligible area of the boreal forest floor.

Overall, we note that *n*-acids with chain-lengths  $>C_{30}$  are negligible in boreal soils, with no detectable *n*-C<sub>34</sub> acids at all sites, no detectable *n*-C<sub>32</sub> acids at five sites, and no detectable *n*-C<sub>30</sub> acids at three sites (Table S2). These higher molecular weight *n*-acids feature prominently in forbs and trees (Figs. 3c and 3e) but are largely absent in boreal soils (Fig. 3f). Thus, it is unlikely forbs or trees are major *n*-acid contributors to soils.

The evidence points strongly to mosses as a primary *n*-acid contributor to boreal soils. Although mosses have the lowest average *n*-acid ( $\Sigma_{C20-C34}$ ) concentration per dry mass (60.2  $\mu\text{g g}^{-1}$ ), they are spatially ubiquitous and dominate the O horizon in terms of raw biomass, which likely overcompensates for its lower absolute *n*-acid concentrations compared to other plants. The low stature and physical connection of mosses at the surface of the O-layer may also favour greater transfer (and representation) of *n*-acids in boreal soils.

Substantial inter- and intra-site variability is noted in soil *n*-acid ( $\Sigma_{C20-C34}$ ) concentrations and chain length distributions, consistent with the heterogeneous nature of soils observed elsewhere (Lehmann et al., 2008; Schäfer et al., 2016), which we attempt to correct for by averaging soils from various depths from the O and A soil horizons (1 to 9 soil samples per site). Site-averaged soil *n*-acid concentrations vary from 17.2 to 267.4.0  $\mu\text{g g}^{-1}$  between sites (Table S4), which likely reflects differences in rates of lipid production, deposition, and mineralization and degradation, including the effects of bacterial activity for example. The  $\Sigma_{C20-C34}$  concentrations are on average slightly higher for the O horizon compared to the A horizon –  $133.0 \pm 105 \mu\text{g g}^{-1}$  versus  $74.8 \pm 81 \mu\text{g g}^{-1}$ , respectively – which may indicate a higher ratio of *n*-acid accumulation to degradation in the O horizon; however, this difference is not statistically significant ( $p = 0.31$ ). The overall range of soil  $\Sigma_{C20-C34}$  *n*-acid concentrations by site (17.2 to 267.4  $\mu\text{g g}^{-1}$ ) is in line with total

soil *n*-acid concentrations from the litter and Ah soil horizons of grasslands, and conifer and deciduous forests in Western Europe which can range from ~10-200  $\mu\text{g g}^{-1}$  (Schäfer et al., 2016).

Previous studies have demonstrated that microbial activity result in substantial decreases in *n*-acid concentrations between vegetation and soil (Chikaraishi and Naraoka, 2006; Schäfer et al., 2016; Nguyen Tu et al., 2017). Preferential degradation of long chain *n*-acids ( $>\text{C}_{28}$ ) has been suggested by previous studies (Chikaraishi and Naraoka, 2006; Nguyen Tu et al., 2017). The site-averaged CPI of soils in this study ( $2.8 \pm 0.9$ ) is significantly lower (t-test,  $p < 0.05$ ) than the average CPI observed in fresh mosses ( $10.5 \pm 3.3$ ), which has the lowest CPI of all major vegetation types (Fig. 3). This suggests that some microbial degradation or other pedogenic factors may be affecting *n*-acid chain length distributions, with a disproportionate loss of the relatively abundant even chain lengths, resulting in a reduced CPI. The same effect was observed for soil *n*-acids along a transect from northern Croatia to southern Sweden (Schäfer et al., 2016). However, the processes by which this degradation occurs in soils has not been fully explored in the literature.

#### *4.2 Source water $\delta D$ and timing of lipid synthesis*

Before discussing the  $\epsilon_{\text{wax/MAP}}$  results, we briefly discuss the relevance of  $\delta D_{\text{MAP}}$  as a source water approximation for sub-Arctic plants. The relevance of this assumption depends largely on: (i) the phenology (timing and duration) of

leaf wax synthesis of the plants that contribute *n*-alkyls to soils; and (ii) mean soil water  $\delta D$  at the time of leaf wax synthesis; both of which are not well known in this region.

Most plant wax phenology studies have focused on *n*-alkanes from mid-latitude environments. *n*-Alkane synthesis in angiosperm trees (Tippie et al., 2013; Freimuth et al., 2017) and C3 grasses (Gamarra and Kahmen, 2017) is associated with leaf development, which for many plants is concentrated towards the start of the growing season; however, in temperate regions with long growing seasons (e.g., 9 months), leaf wax concentrations and homologue patterns can continue to evolve for the first half of the growing season (Sachse et al., 2015). To our knowledge, only one study has assessed *n*-acid phenology and found that *n*-acids are synthesized continuously throughout the growing season in temperate angiosperms (Freimuth et al., 2017). If *n*-acids are also produced continuously in northern boreal woodland plant types throughout the growing season, this would mean the  $\delta D$  source water assumption should also align with mean growing season soil waters (i.e., mid-May to late-September, Methods 2.1).

Average soil water  $\delta D$  in sub-Arctic regions is also not well known, but is thought to reflect changing seasonal inputs of spring snowmelt and summer rainwater, mixing with remnant pore waters from the previous year (frozen *in situ* during the freezeback), and evaporative enrichment of near-surface soil

waters in the mid-summer. In temperate alpine forests in Switzerland, which experience freeze-thaw, Brinkmann et al. (2018) demonstrated that comparable meteoric water contributions from summer (50%) and prior fall and winter months (40%) are represented in soil and xylem waters of *Fagus sylvatica* and *Picea abies* trees during the growing season. In a shrub-tundra environment in N. Alaska, Daniels et al. (2017) demonstrated that *Eriophorum vaginatum* (C3 grass) and *Betula nana* (shrub) xylem waters in the mid-growing season (mid-July) were comparable to  $\delta D_{MAP}$ , while August xylem waters were enriched relative to  $\delta D_{MAP}$  by  $\sim +10\text{‰}$ ; data were not collected for the first half of the growing season (June 3-July 17) when soil and xylem waters were likely depleted compared to  $\delta D_{MAP}$ . Both case studies offer some empirical support for the notion that mean soil and xylem waters integrate a blend of year-round precipitation in alpine and low-Arctic regions, and that  $\delta D_{MAP}$  offers a reasonable approximation for  $\delta D$  source water in cold-regions. We assume this approximation also applies to northern boreal woodlands, but acknowledge that future ecohydrology studies are needed to better substantiate this assumption.

#### 4.3 Net fractionation

Our regional average  $\epsilon_{wax/MAP}$  values calculated for  $n\text{-C}_{24,26,28}$  acids ( $-93 \pm 10\text{‰}$ ,  $-101 \pm 11\text{‰}$ , and  $-95 \pm 11\text{‰}$ , respectively) are all within error the mean  $\epsilon_{wax/MAP}$  of  $-96 \pm 8\text{‰}$  (based on  $n\text{-C}_{30}$  acids) reported by Wilkie et al. (2013) from lake sediments in N.E. Siberia, and within two standard errors of the  $\epsilon_{wax/MAP}$

of  $-114 \pm 13\text{‰}$  (based on  $n\text{-C}_{22-30}$  acids) reported by Daniels et al. (2017) from  
 lake sediments in N. Alaska (Fig. 5). All of these  $\epsilon_{\text{wax/MAP}}$  values are far more  
 negative than the mean  $\epsilon_{\text{wax/MAP}}$  of  $-61 \pm 20\text{‰}$  (based on  $n\text{-C}_{26}$  and  $\text{-C}_{28}$  acids)  
 reported by Shanahan et al. (2013) from lake sediments on Baffin Island (Fig.  
 5).  $n\text{-C}_{25-31}$  alkanes in surface sediments of northern ( $> 60^\circ\text{N}$ ) Fennoscandian  
 lakes reported by Sachse et al. (2006) are characterised by an average  $\epsilon_{\text{wax/MAP}}$   
 value of  $-125 \pm 7\text{‰}$  (Fig. 5), which is significantly larger than the  $n\text{-acid}$ -based  
 $\epsilon_{\text{wax/MAP}}$  values from Wilkie et al. (2013) and in this study. Some studies have  
 found  $n\text{-acids}$  are significantly more enriched in D by  $\sim 25 \pm 16\text{‰}$  compared to  
 $n\text{-alkanes}$  from the same leaves (Chikaraishi and Naraoka, 2007), which could  
 explain the smaller  $\epsilon_{\text{wax/MAP}}$  values in  $n\text{-acids}$  vs.  $n\text{-alkanes}$  (Fig. 5). However,  
 the average  $\epsilon_{\text{wax/MAP}}$  calculated from lake sediment  $n\text{-alkanes}$  ( $-118 \pm 9\text{‰}$ ) in N.  
 Alaska is only slightly larger than the mean  $\epsilon_{\text{wax/MAP}}$  value based on  $n\text{-acids}$  ( $-$   
 $114 \pm 13\text{‰}$ ) (Daniels et al., 2017). Therefore, factors other than compound class  
 may be responsible for the larger  $n\text{-alkane}$   $\epsilon_{\text{wax/MAP}}$  values in Fennoscandia.

At broader scales, our regional  $\epsilon_{\text{wax/MAP}}$  values are also similar to  $n\text{-acid}$  ( $n\text{-}$   
 $\text{C}_{28}$  or longer) net fractionations reported from tropical and temperate sites  
 which range from  $-90$  to  $-121\text{‰}$  (mean:  $-105\text{‰}$ ) (Hou et al., 2008; Feakins et  
 al., 2014, 2016a; Gao et al., 2014). Collectively, this argues for a similar net  
 fractionation in  $n\text{-acids}$  across broad geographic scales and ecosystem types.  
 However, our  $\epsilon_{\text{wax/MAP}}$  results show some evidence of geographic and/or climatic

dependencies, indicating that spatial factors may influence net fractionation at regional scales.

On average,  $\delta D_{\text{MAP}}$  is more negative at higher latitudes or lower MATs (Figs. 6a and c), which is typical at mid- to high-latitudes (Dansgaard, 1964), and specifically in this region (Porter et al., 2016), reflecting preferential rainout of heavy isotopologues as moist air travels away from the ocean source towards cooler, higher latitudes. Conversely,  $\delta D_{\text{wax}}$  for *n*-C<sub>26</sub> and -C<sub>28</sub> acids is slightly more positive at higher latitudes or colder MATs (Figs. 6b and 6d), although these  $\delta D_{\text{wax}}$  trends are not statistically significant ( $p \geq 0.05$ ). The combination of the opposing trends in  $\delta D_{\text{MAP}}$  and  $\delta D_{\text{wax}}$  results in smaller  $\epsilon_{\text{wax/MAP}}$  values at higher latitudes or lower MATs. However, due to collinearity between latitude and MAT ( $r = -0.91$ ,  $p \leq 0.01$ ) it is not clear which is the driving variable. A significant correlation is also observed between  $\epsilon_{\text{wax/MAP}}$  and elevation for *n*-C<sub>26</sub> acids only ( $r = -0.63$ ,  $p < 0.05$ ). However, this correlation is also confounded by collinearities with latitude ( $r = -0.82$ ,  $p \leq 0.01$ ) and MAT ( $r = 0.65$ ,  $p \leq 0.05$ ). There are no known physical linkages between  $\epsilon_{\text{wax/MAP}}$  and elevation (or dependent variables not already included in our correlation analysis; Table S3) and, thus, perhaps the most likely explanation for this correlation is random co-variability in combination with latitude and MAT collinearities. We focus the remainder of our discussion on possible linkages between latitude or MAT on  $\epsilon_{\text{wax/MAP}}$ , which have been evaluated to some extent in previous studies.

Latitude does not have a direct influence on  $\epsilon_{\text{wax/MAP}}$ . However, other variables that co-vary with latitude such as sunlight availability may have an effect (Sessions, 2006; Yang et al., 2009). Across our network of sites, June-August total sunlight hours ranges from 1608 hours (average 17.5 hours/day) at the most southern site to 1973 hours (average 21.5 hours/day) at the most northern site (USNO-AA, 2018). There are two mechanisms discussed in the literature linking sunlight and  $\epsilon_{\text{wax/MAP}}$ . Growth-chamber experiments (Yang et al., 2009) have found smaller net fractionations in plants exposed to increased light levels (simulating a high-latitude environment), an effect that is thought to be due to enhanced transpiration and D-enrichment of leaf waters, resulting in higher  $\delta D_{\text{wax}}$  and smaller  $\epsilon_{\text{wax/MAP}}$  values. This mechanism was invoked by Shanahan et al. (2013) to explain unusually small  $\epsilon_{\text{wax/MAP}}$  values (average =  $-61 \pm 20\%$ ) calculated for *n*-C<sub>26-28</sub> acids from lake sediments on Baffin Island (62-73°N). However, evidence for strong leaf water evaporative enrichment effects on  $\epsilon_{\text{wax/water}}$  in natural ecosystem surveys is generally lacking (Feakins and Sessions, 2010).

A second mechanism linking sunlight to  $\epsilon_{\text{wax/MAP}}$  relates to usage of stored carbohydrates. Under restricted light, plants use more stored carbohydrates (which are associated with an overall smaller biochemical fractionation,  $\epsilon_{\text{bio}}$ ) as feedstock for synthesizing plant waxes, leading to a smaller  $\epsilon_{\text{wax/water}}$  (Sessions, 2006; Cormier et al., 2018). However, this mechanism is inconsistent with the



sign of the latitude (sunlight)- $\epsilon_{\text{wax/MAP}}$  response observed across our network of sites, which shows smaller  $\epsilon_{\text{wax/MAP}}$  values at high latitudes where light is more abundant.

The correlation between MAT and  $\epsilon_{\text{wax/MAP}}$  could reflect a direct effect of temperature on  $\epsilon_{\text{bio}}$ . However, previous studies of tree-ring cellulose  $\delta\text{D}$  have specifically investigated this possibility and concluded that no such relation exists (Roden et al., 2000b).

Although the physical drivers of  $\epsilon_{\text{wax/MAP}}$  variability across our network of sites are not known, the reality of this variability implies that a single  $\epsilon_{\text{wax/MAP}}$  may not necessarily be appropriate for all boreal sites. This has implications for interpreting fossil  $n$ -acid  $\delta\text{D}_{\text{wax}}$  records, since the use of a single  $\epsilon_{\text{wax/MAP}}$  value to reconstruct paleo- $\delta\text{D}_{\text{precip}}$  from fossil  $\delta\text{D}_{\text{wax}}$  could introduce additional uncertainties. Rather, a latitude-specific  $\epsilon_{\text{wax/MAP}}$  (for  $n$ -C<sub>26</sub> and -C<sub>28</sub> acids only) may improve the accuracy of  $\delta\text{D}_{\text{precip}}$  reconstructions in northern boreal regions. If we parse the  $n$ -C<sub>26</sub> and -C<sub>28</sub> datasets by latitude, a variable that is generally well known for most geological applications (rather than temperature which would need to be independently reconstructed), we find that sites  $< 65^\circ\text{N}$  have a significantly larger mean  $\epsilon_{\text{wax/MAP}}$  values than sites  $> 65^\circ\text{N}$  (Fig. 4g). For the southern sites ( $< 65^\circ\text{N}$ ,  $n = 7$ ), we calculate mean  $\epsilon_{\text{wax/MAP}}$  values of  $-107 \pm 8\text{‰}$  for  $n$ -C<sub>26</sub>, and  $-101 \pm 8\text{‰}$  for -C<sub>28</sub> acids. For the northern sites ( $> 65^\circ\text{N}$ ,  $n = 6$ ),

we calculate mean  $\epsilon_{\text{wax/MAP}}$  values of  $-94 \pm 11\text{‰}$  for  $n\text{-C}_{26}$ , and  $-89 \pm 10\text{‰}$  for  $n\text{-C}_{28}$  acids.

#### 4.4 Additional uncertainties in net fractionation

While regional trends in  $\epsilon_{\text{wax/MAP}}$  are partially explained by latitude ( $r = 0.62$  or  $\sim 38\%$  of variance for  $n\text{-C}_{26}$  acids), a significant fraction of inter-site variability in  $\epsilon_{\text{wax/MAP}}$  must be related to other factors, as discussed below.

Inter-site differences in minor wax contributions from the different boreal vegetation types may contribute to soil  $\epsilon_{\text{wax/MAP}}$  variability. For example, globally,  $n$ -alkane  $\epsilon_{\text{C}_{29}/\text{MAP}}$  values for C3 grasses, forbs, trees and shrubs are  $-149\text{‰}$ ,  $-128\text{‰}$ ,  $-121\text{‰}$  and  $-99\text{‰}$ , respectively, or a  $50\text{‰}$  range between the C3 grasses and shrub end-members (Sachse et al., 2012). These differences in  $\epsilon_{\text{wax/MAP}}$  owe to plant-specific differences in biochemical fractionation and the fraction of enriched leaf water that imprints on  $n$ -alkyl precursors (Gamarra et al., 2012), as well as rooting depth which can lead to major differences between shallow versus deep-rooted plants in environments where near surface source waters are evaporatively D-enriched (Nichols et al., 2010). For a shrub-tundra environment in N. Alaska, Daniels et al. (2017) report a  $64\text{‰}$  range in  $\epsilon_{\text{wax/xylem}}$  values between C3 grasses (*Eriophorum vaginatum*;  $\epsilon_{\text{C}_{28}/\text{xylem}} = -160\text{‰}$ ) and shrubs (*Betula nana*;  $\epsilon_{\text{C}_{28}/\text{xylem}} = -96\text{‰}$ ). These Alaskan  $n$ -acid  $\epsilon_{\text{wax/xylem}}$  values are similar to global  $n$ -alkane  $\epsilon_{\text{C}_{29}/\text{MAP}}$  values for C3 grasses and shrubs (Sachse

et al., 2012); therefore, a similarly large range in  $\epsilon_{\text{wax/MAP}}$  may be expected for grasses and shrubs in the boreal forest. It is possible some inter-site  $\epsilon_{\text{wax/MAP}}$  variance in our network owes to variable wax contributions from the minor plant types (e.g., C3 grasses and shrubs). However, C3 grasses occupy a small fraction of the boreal forest understorey, restricted to micro-topographic high points that are well drained, and likely do not contribute much biomass to soils. Conversely, shrubs occupy a larger area of the boreal understory and have the highest *n*-acid concentrations in fresh foliage of all vegetation types. Therefore, it seems more probable that shrubs would have a greater influence on the soil *n*-acid pool than other minor vegetation types such as C3 grasses.

Site slope, aspect and soil character (texture, compaction, permafrost) may also contribute to soil  $\epsilon_{\text{wax/MAP}}$  variability, since these variables have some implications for drainage (Christensen et al., 2013) and, therefore, soil water  $\delta\text{D}$ . During the spring melt, winter precipitation runs off sloped, impermeable sites more rapidly, whereas sites in flat areas or depressions with permeable soils are likely to retain more winter precipitation in soil water. In turn, soil *n*-acids from poorly drained sites may be expected to have larger  $\epsilon_{\text{wax/MAP}}$  due to a greater uptake of winter precipitation compared to well-drained sites. Future insights on soil-derived  $\epsilon_{\text{wax/MAP}}$  values may benefit from investigations of site specific and microtopographic effects on source water  $\delta\text{D}$ .

Inaccurate  $\delta D_{\text{MAP}}$  estimates may also contribute to inter-site variability in  $\varepsilon_{\text{wax/MAP}}$ . We used  $\delta D_{\text{MAP}}$  estimates from the OIPC (Bowen and Revenaugh, 2003), an interpolated product that depends on available data from the local GNIP stations. There are three GNIP stations in proximity to our network of sites, including Inuvik, Mayo, and Whitehorse (Fig. 1). However, the number of monthly records, continuity and temporal coverage of these records is highly variable. For example, the Inuvik GNIP record holds 14 monthly observations spanning the years 1986-1989, most ( $n = 8$ ) of which are from 1988. Conversely, Whitehorse has the best coverage with 95 monthly records with relatively continuous coverage from 1961-1965 and 1985-1989. The spatial and temporal inequities in the GNIP network leaves open the possibility that there may be some regional bias in the  $\delta D_{\text{MAP}}$  isoscape, with implications for  $\varepsilon_{\text{wax/MAP}}$ .

Within-site  $\varepsilon_{\text{wax/MAP}}$  variability is also substantial ( $1\sigma$  ranges from 2-15 ‰ depending on the homologue and site). Increasing the number of samples per site would allow for a more robust mean  $\varepsilon_{\text{wax/MAP}}$  estimate from each site, and may reduce inter-site variability. However, considering the range of latitude and climatic conditions covered by this transect, as well as the documented heterogeneity of soils at even nanometer scales (Lehmann et al., 2008), the inter- and intra- site variability we observe is not unexpected.

#### *4.5 Potential for paleoenvironmental applications*

The contiguous region of continental Yukon and Alaska that was unglaciated during the last full glacial, collectively known as Eastern Beringia, hosts an abundance of fossil-rich sediments spanning the last ca. 2.9 Ma (Elias, 2000; Matheus et al., 2003; Matthews et al., 2003; Zazula et al., 2003; Péwé et al., 2009; Schweger et al., 2011), which offers great potential for fossil wax-based paleoclimate reconstructions. By comparison, most other sub-Arctic regions lack Pleistocene surficial deposits due to the legacy of glacial scouring. The pre-50 ka chronology of deposits in Eastern Beringia is also exceptional, owing to datable ash beds from proximal volcanic sources (Froese et al., 2009; Preece et al., 2011). Recent studies in central Yukon have documented well-preserved *n*-alkanes and -acids from steppe-tundra paleoenvironments in relict permafrost deposits dating to Late Pleistocene cold stages (Pautler et al., 2014; Porter et al., 2016). However, this proxy has not been applied to Pleistocene interglacial paleosols in this region, which are reported at numerous locales (Schweger et al., 2011). Fossil pollen and macrofossil evidence reveals that *Picea* dominated woodlands, consistent with the majority of our sites, were typical during most interglacials in this region (Schweger et al., 2011). The  $\epsilon_{\text{wax/MAP}}$  constrained in our study represents an important first step toward interpreting interglacial paleosols in Eastern Beringia.

To date, most quantitative paleoclimate estimates from this region are based on the Modern Analogue Technique using fossil pollen (Viau et al., 2008)

729 or the Mutual Climate Range approach using fossil insects (Elias, 2000; Zazula  
730 et al., 2011). However, pollen and midge reconstructions, like all paleoclimate  
731 proxies, have uncertainties and do not always produce coherent paleoclimate  
732 estimates in this region (see discussion by Porter et al., 2016). Fossil *n*-acids  
733 provide an independent proxy that can be readily applied, in concert with more  
734 traditional proxies (e.g., pollen and insect assemblages), to better resolve the  
735 Quaternary history of this region.

## 5. Conclusions

This study analyzed *n*-acids from modern vegetation and soils from a network of 13 sites in Yukon, Alaska and Northwest Territories. The goals of this study were to better understand: (1) which plant types are the primary contributors of *n*-acids to northern boreal soils, and (2) the net fractionation of hydrogen isotopes in boreal soil *n*-acids. Regional soil surveys represent a powerful approach to constraining net fractionation as they integrate a large number of uncertainties that may influence sedimentary  $\delta D_{\text{wax}}$  variability, for example, due to differences in the relative contributions of major and minor plant types, post-depositional degradation, microtopography and drainage, and a wide range of climatic and geographic variables. The net fractionation values presented here are most directly applicable for  $\delta D_{\text{MAP}}$  reconstructions based on *n*-acids from boreal paleosols, as wax transport, sources and degradation may vary between soils and other depositional systems (e.g. lake sediments; Nguyen Tu et al., 2017).

The overall *n*-acid homologue pattern observed in boreal soils (O and A horizons) in this study is closely associated with the pattern observed in fresh mosses, which suggests mosses are a primary *n*-acid contributor to the average boreal soil. The overall net fractionation of soil  $\delta D_{\text{wax}}$  relative to  $\delta D_{\text{MAP}}$  ( $\epsilon_{\text{wax/MAP}}$ ) is  $-93 \pm 10\text{‰}$  for *n*-C<sub>24</sub>,  $-101 \pm 11\text{‰}$  for -C<sub>26</sub>, and  $-96 \pm 11\text{‰}$  for -C<sub>28</sub> acids, respectively. We also observe spatial trends in  $\epsilon_{\text{wax/MAP}}$  for *n*-C<sub>26</sub> and -C<sub>28</sub> acids

(but not -C<sub>24</sub>) with smaller  $\epsilon_{\text{wax/MAP}}$  values at higher latitudes or at lower mean annual temperatures, factors that co-vary across our network. Since latitude is generally well known over recent geologic timescales, this is a more pragmatic way to parse the dataset. We define two latitude-specific  $\epsilon_{\text{wax/MAP}}$  values, one for sites < 65°N and one for sites > 65°N. For the more southern group (60-64°N),  $\epsilon_{\text{wax/MAP}}$  is  $-107 \pm 8 \text{ ‰}$  for *n*-C<sub>26</sub> and  $-101 \pm 8 \text{ ‰}$  for -C<sub>28</sub> acids, respectively. For the northern group (65-68°N),  $\epsilon_{\text{wax/MAP}}$  is  $-94 \pm 11 \text{ ‰}$  for *n*-C<sub>26</sub> and  $-89 \pm 10 \text{ ‰}$  for -C<sub>28</sub> acids, respectively. These results will help inform  $\delta D_{\text{wax}}$  interpretations from fossil boreal *n*-acids, especially from paleosol deposits that are associated with the northern boreal woodland ecology.

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## Figure Captions

Fig. 1. Regional maps of (a) annual precipitation (mm/year) (WorldClim 2) and (b) mean annual temperature (°C) (WorldClim 2), with locations of the study sites (black dots), local GNIP sites (white triangles), and the Arctic circle (thin black line), north of which sites experience 24-hr daylight at least one day per year. For colour version of figure, refer to the online version of the article.

Fig. 2. Average monthly  $\delta D_{\text{precip}}$  (OIPC), mean air temperature (ERA-Int), and precipitation (WorldClim 2). Error bars represent the range of values for each of the study sites.

Fig. 3. *n*-Acid relative abundance distributions for (a-e) type-averaged vegetation and (f) soils. The total number (*n*) of samples analysed is indicated. The *n*-acid distributions reflect regional averages for each vegetation type and soil, first averaged by site and then the site averages were used to calculate regional averages. Error bars represent  $1\sigma$ .

Fig. 4. Site-averaged  $\varepsilon_{\text{wax/MAP}}$  for *n*-C<sub>24</sub>, -C<sub>26</sub>, and -C<sub>28</sub> acids (red, grey and blue circles, respectively) versus: (a) latitude, (b) elevation (m a.s.l.), (c) annual precipitation (mm; WorldClim 2), (d) mean annual air temperature (°C; ERA-

Interim), (e) mean growing season air temperature (May-September), and (f) Summer (JJA) relative humidity (%; CliMond). Correlations significant at  $p \leq 0.05$  are indicated. Box plots (g) of site-averaged  $\epsilon_{\text{wax/MAP}}$  values for  $n\text{-C}_{24}$  (red),  $\text{-C}_{26}$  (grey), and  $\text{-C}_{28}$  (blue) acids grouped by latitude ( $< 65^\circ\text{N}$  and  $> 65^\circ\text{N}$ ); the  $n\text{-C}_{26}$  and  $\text{-C}_{28}$  group means are significantly different ( $p \leq 0.05$ ; 2-sample t-test). For colour version of figure, refer to the online version of the article.

Fig. 5. (a) Comparison of sediment-derived  $\epsilon_{\text{wax/MAP}}$  values from other studies  $> 60^\circ\text{N}$ , including  $n$ -acids and -alkanes; (b) map of corresponding site locations. The numeric codes refer to: [1] this study; [2] Wilkie et al. (2013); [3] Daniels et al. (2017); (4) Shanahan et al. (2013); [5] Sachse et al. (2006). For colour version of figure, refer to the online version of the article.

Fig. 6. OIPC- $\delta\text{D}_{\text{MAP}}$  for the study sites versus (a) latitude and (c) mean annual temperature.  $\delta\text{D}_{\text{wax}}$  ( $n\text{-C}_{24,26,28}$  acids; red, grey and blue circles, respectively) for the study sites versus (b) latitude and (d) mean annual temperature. For colour version of figure, refer to the online version of the article.

1149    **Table Captions**

1150    Table 1: Sampling site locations and mean climate

1151    Table 2: Site-averaged  $\delta D_{\text{wax}}$  and  $\delta_{\text{wax/MAF}}$  for n-C<sub>24-28</sub> acids in topsoil (0-30 cm)

1152    samples

1153

Table 1

Site		Location		Temperature (°C) <sup>a</sup>		Precipitation (mm) <sup>b</sup>		Relative Humidity (%) <sup>c</sup>	
Code	Full Name	Lat.	Long.	Elev. (m a.s.l.)	MAT	MJJAS	Annual	Annual	JJA
SP	Spindly Pine Lake	60.302	-134.992	792	-2.6	6.9	387	70.2	65.4
WH	Whitehorse	60.812	-135.211	729	-1.7	8.1	250	68.4	62.9
TKR	Takhini River	60.852	-135.718	662	-2.0	7.8	254	69.3	64.3
KL	Kluane Lake	60.965	-138.059	950	-4.5	4.2	363	74.6	74.1
MYO	Mayo	63.541	-136.162	555	-3.5	9.4	325	68.3	61.5
BRM	Brimstone	63.742	-138.849	639	-2.9	10.1	329	70.9	68.1
LC	Lost Chicken	64.068	-141.911	588	-4.9	9.2	249	70.9	68.3
SH	Scree Hill	65.071	-138.155	875	-6.7	6.7	376	69.0	65.8
DHP174	Dempster Hwy Peatland	65.212	-138.327	738	-6.4	7.0	359	69.0	65.6
EAG	Eagle Plains	66.444	-136.675	370	-6.7	8.3	333	67.9	63.6
TSI	Tsiigehtchic	68.042	-133.489	78	-7.0	9.0	241	67.5	62.5
CL	Campbell Lake	68.207	-133.406	46	-8.4	8.0	242	67.5	62.5
INU	Inuvik	68.315	-133.553	58	-8.4	8.0	238	67.5	62.5

<sup>a</sup>ERA-Interim surface temperature data (1981-2010), (~80 km spatial resolution) mean annual temperature and mean temperature May-September (Dee et al., 2011)

<sup>b</sup>WorldClim2 precipitation data, annual (30" spatial resolution) and total May-September (Fick and Hijmans, 2017)

<sup>c</sup>ClimMond humidity data (10' spatial resolution), annual and average May-September (Kriticos et al., 2012)

Table 2

Site	$\delta D_{\text{MAP}} (\text{‰})^a$	$\delta D_{\text{wax}} (\text{‰}) \pm 1\sigma (n)$				$\epsilon_{\text{wax}/\text{MAP}} (\text{‰}) \pm 1\sigma (n)$			
		$C_{24}$	$C_{26}$	$C_{28}$		$C_{24}$	$C_{26}$	$C_{28}$	
SP	-163	$-237 \pm 6.9 (9)$	$-258 \pm 7.5 (9)$	$-255 \pm 11.7 (9)$		$-88 \pm 8.3 (9)$	$-114 \pm 9.0 (9)$	$-110 \pm 14.0 (9)$	
WH	-165	$-252 (1)$	$-250 \pm 2.6 (3)$	$-244 \pm 5.0 (3)$		$-104 (1)$	$-101 \pm 3.1 (3)$	$-95 \pm 6.0 (3)$	
TKR	-164	$-240 \pm 3.9 (5)$	$-249 \pm 9.9 (5)$	$-247 \pm 6.3 (3)$		$-91 \pm 4.7 (5)$	$-102 \pm 11.8 (5)$	$-100 \pm 7.5 (3)$	
KL	-167	$-243 (1)$	$-254 (1)$	$-242 (1)$		$-92 (1)$	$-105 (1)$	$-91 (1)$	
MYO	-161	$-253 \pm 1.8 (2)$	$-264 \pm 2. (2)$	$-255 \pm 2.4 (2)$		$-110 \pm 2.1 (2)$	$-122 \pm 2.4 (2)$	$-111 \pm 2.8 (2)$	
BRM	-164	$-245 \pm 2.9 (3)$	$-246 \pm 5.9 (3)$	$-248 \pm 3.9 (3)$		$-97 \pm 3.4 (3)$	$-98 \pm 7.0 (3)$	$-101 \pm 4.7 (3)$	
LC	-160	$-242 \pm 7.8 (2)$	$-251 \pm (1)$	$-243 (1)$		$-98 \pm 9.3 (2)$	$-109 (1)$	$-99 (1)$	
SH	-174	$-253 \pm 12.6 (3)$	$-261 \pm 5.9 (2)$	$-260 \pm 5.7 (2)$		$-95 \pm 15.2 (3)$	$-105 \pm 7.1 (2)$	$-104 \pm 6.9 (2)$	
DHP174	-173	$-243 \pm 7.6 (3)$	$-255 \pm 4.2 (3)$	$-239 \pm 2.3 (3)$		$-84 \pm 9.2 (3)$	$-99 \pm 5.1 (3)$	$-80 \pm 2.8 (3)$	
EAG	-171	$-256 \pm 2.7 (4)$	$-255 \pm 8.1 (4)$	$-252 \pm 8.4 (3)$		$-102 \pm 3.2 (4)$	$-102 \pm 9.8 (4)$	$-98 \pm 10.1 (3)$	
TSI	-175	$-244 \pm 2.5 (3)$	$-241 \pm 8.7 (2)$	$-238 \pm 4.3 (2)$		$-84 \pm 3.1 (3)$	$-80 \pm 10.6 (2)$	$-77 \pm 5.2 (2)$	
CL	-176	$-253 \pm 2.2 (2)$	$-256 \pm 6.7 (2)$	$-248 \pm 8.2 (2)$		$-94 \pm 2.6 (2)$	$-97 \pm 8.1 (2)$	$-87 \pm 9.9 (2)$	
INU	-176	$-234 \pm 5.0 (2)$	$-244 \pm 6.7 (2)$	$-247 (1)$		$-71 \pm 6. (2)$	$-82 \pm 8.1 (2)$	$-87 (1)$	
Mean	-168	-246	-253	-248		-93	-101	-95	
$1\sigma$	6.0	6.8	6.6	6.4		10.0	11.3	10.7	

<sup>a</sup>OIPC  $\delta D_{\text{MAP}}$  (Bowen and Revenaugh, 2003)

Figure 1

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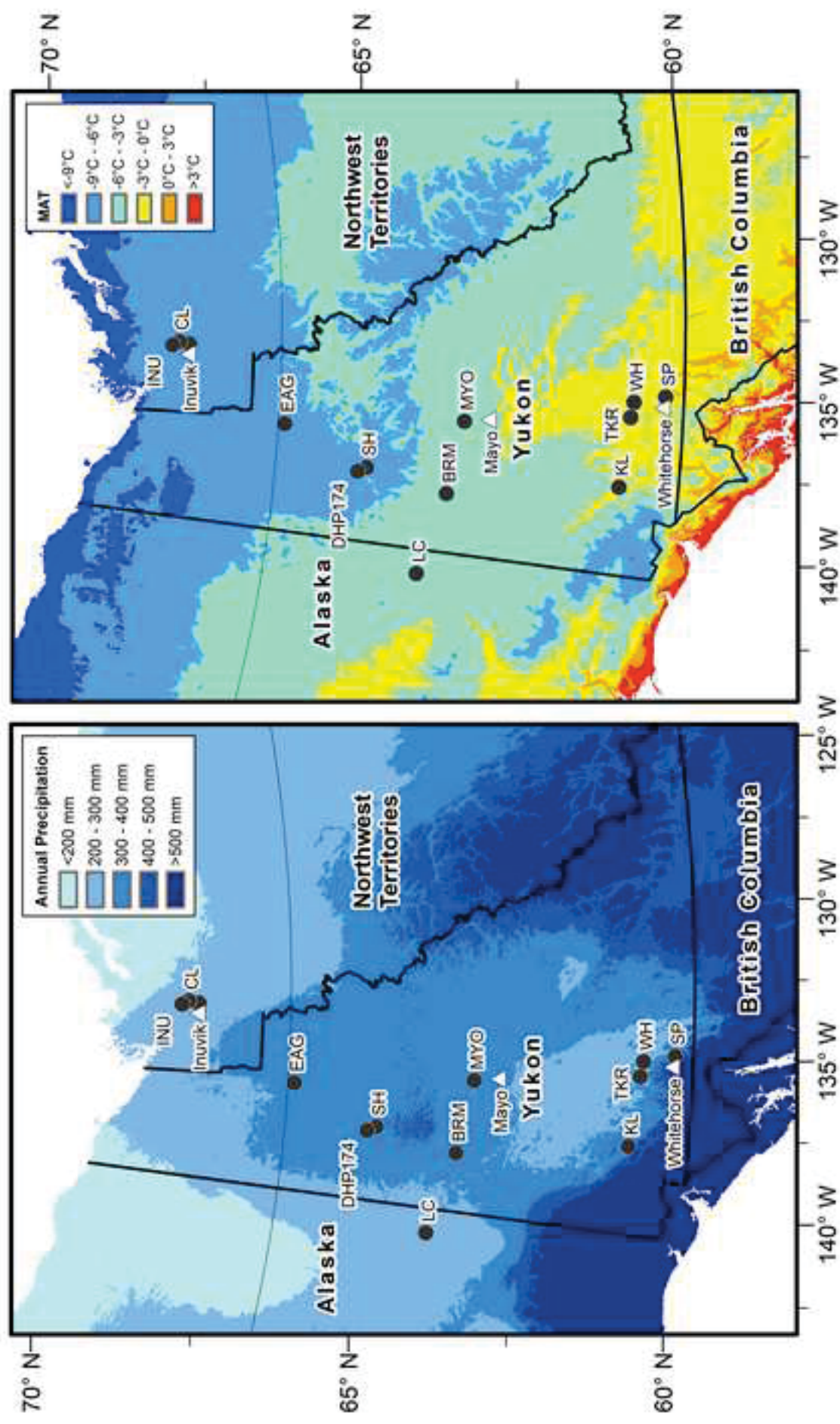
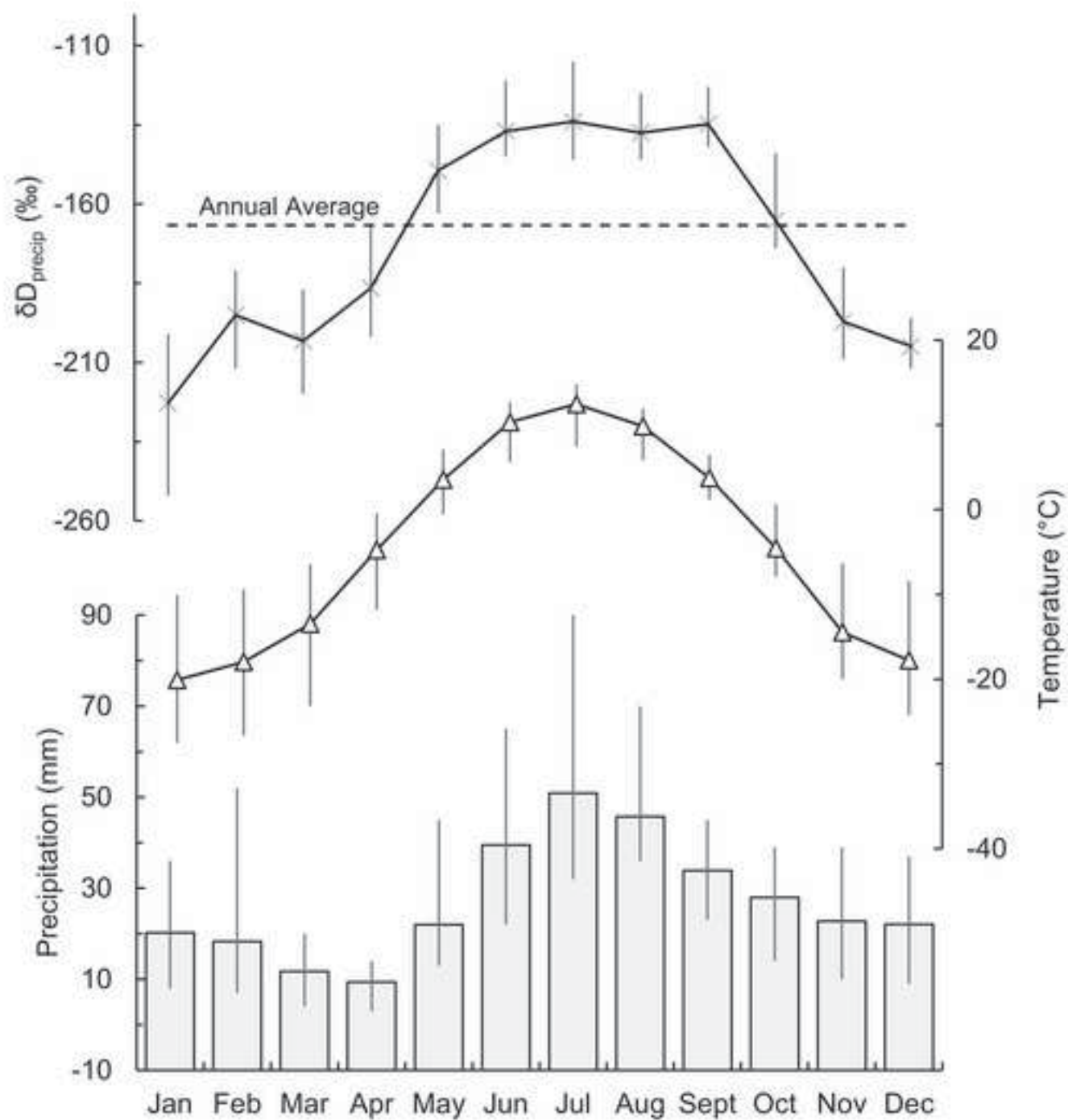


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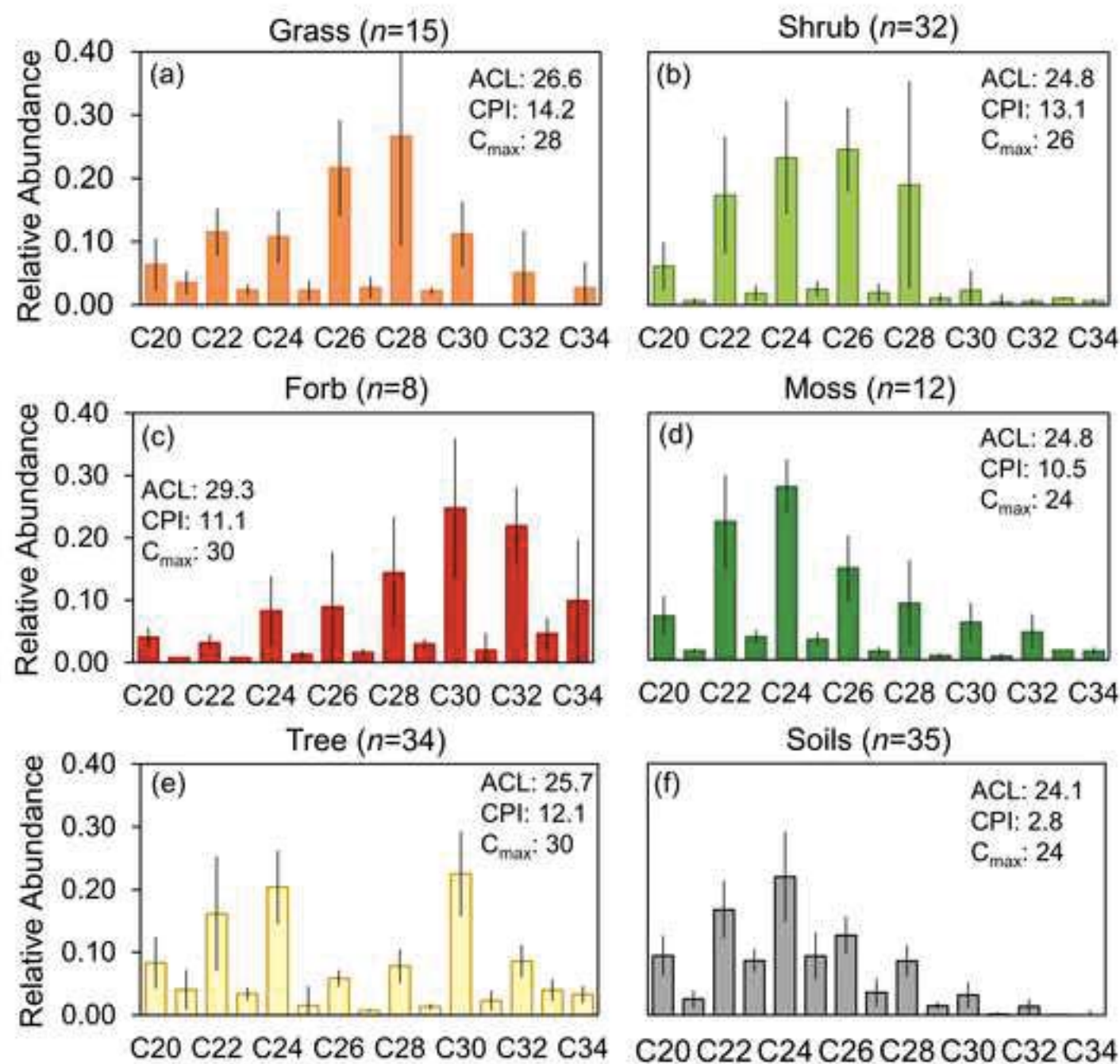




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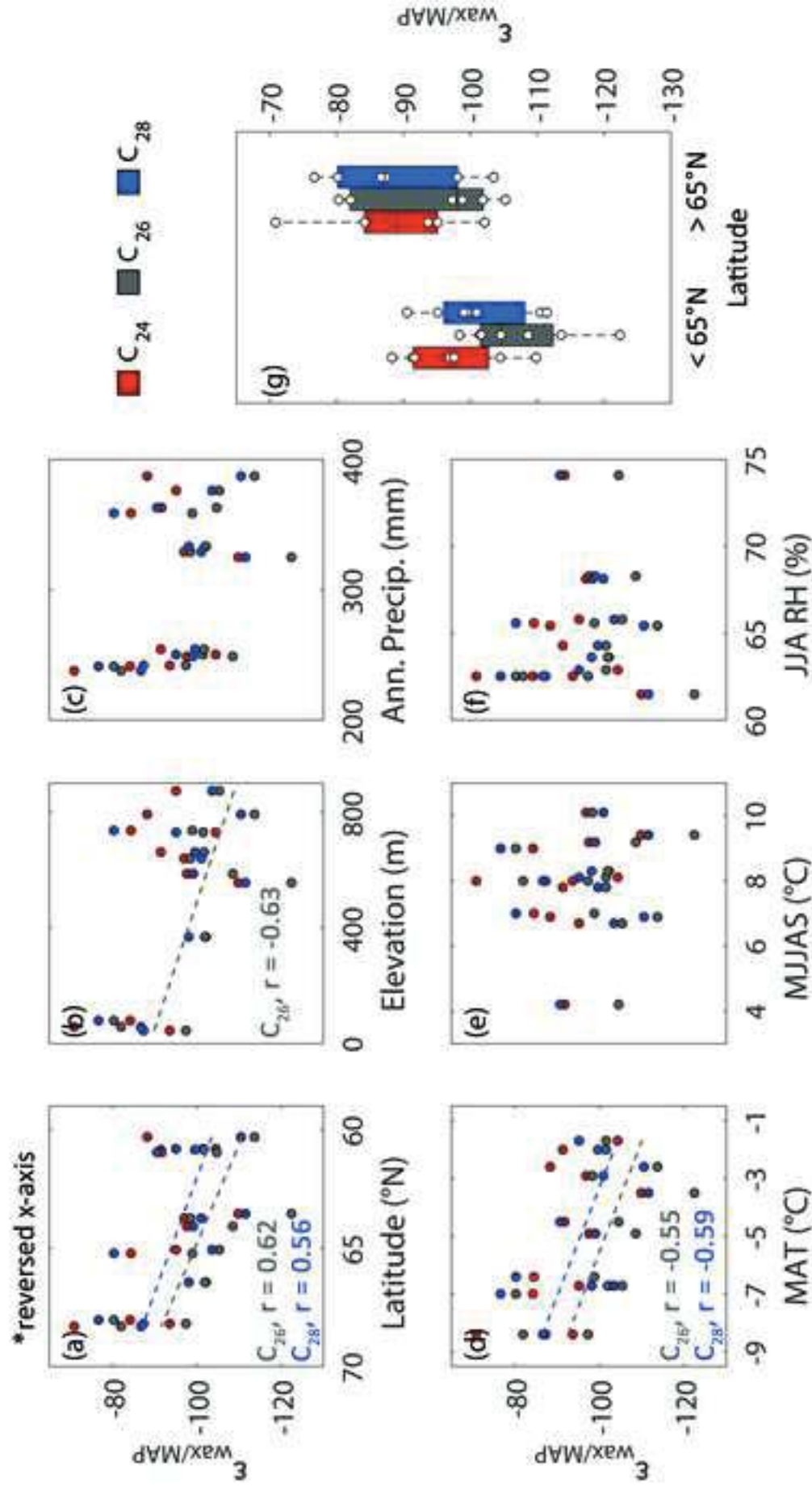


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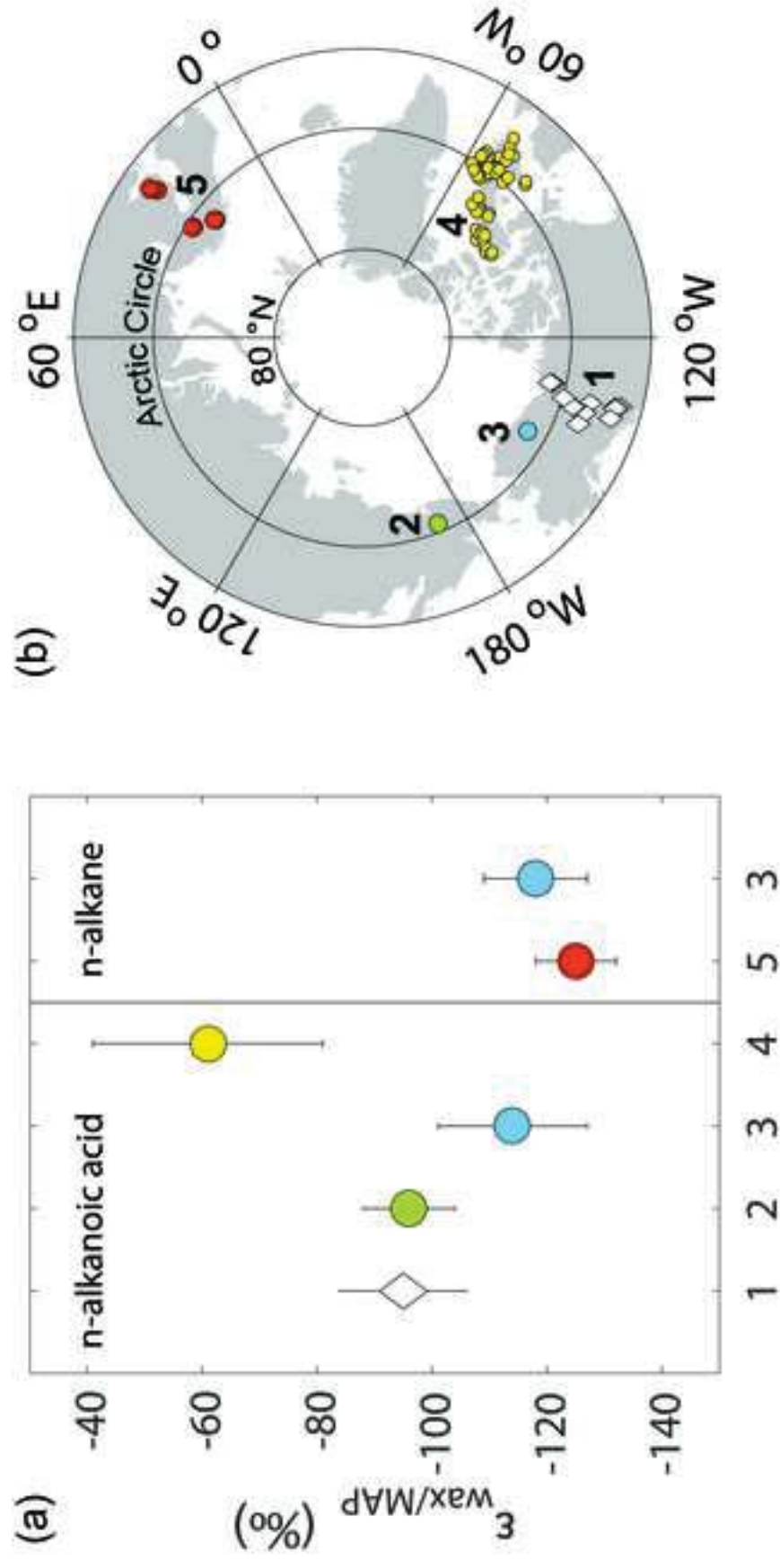


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