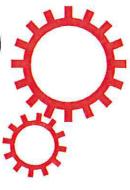


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Dissolved organic matter uptake by *Trichodesmium* in the Southwest Pacific

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The globally distributed diazotroph *Trichodesmium* contributes importantly to nitrogen inputs in the oligotrophic oceans. Sites of dissolved organic matter (DOM) accumulation could promote the mixotrophic nutrition of *Trichodesmium* when inorganic nutrients are scarce. Nano-scale secondary ion mass spectrometry (nanoSIMS) analyses of individual trichomes sampled in the South Pacific Ocean, showed significant ¹³C-enrichments after incubation with either ¹³C-labeled carbohydrates or amino acids. These results suggest that DOM could be directly taken up by *Trichodesmium* or primarily consumed by heterotrophic epibiont bacteria that ultimately transfer reduced DOM compounds to their host trichomes. Although the addition of carbohydrates or amino acids did not significantly affect bulk N₂ fixation rates, N₂ fixation was enhanced by amino acids in individual colonies of *Trichodesmium*. We discuss the ecological advantages of DOM use by *Trichodesmium* as an alternative to autotrophic nutrition in oligotrophic open ocean waters.

Nitrogen is recognized as the proximate limiting nutrient for primary production in the oceans¹. The oceanic nitrogen reservoir is controlled by a balance between fixed nitrogen gains (via dinitrogen -N₂- fixation) and losses (denitrification)². While global nitrogen budget estimations determine that denitrification exceeds N₂ fixation considerably³, recent improvements in the ¹⁵N₂ isotope tracer method used to measure biological N₂ fixation have evidenced that formerly published rates could be underestimated by a factor of ~2 to 6⁴⁻⁸, and thus could be high enough to balance denitrification on a global basis. However, the variability among N₂ fixation rates obtained when using the two different methods (adding ¹⁵N₂ as a bubble or pre-dissolved in seawater)^{4,9} can be high⁷ and at times not significant¹⁰⁻¹². A mechanistic understanding of which factors determine the degree of discrepancy between the two ¹⁵N₂ methods is currently lacking. Moreover, marine pelagic N₂ fixation had been long attributed to the tropical and subtropical latitudinal bands of the ocean, e.g.¹³, while other ecological niches such as high latitude waters, oxygen minimum zones and the vast dark realm of the ocean (below the euphotic zone) are now recognized as active N₂ fixation sites¹⁴⁻¹⁶. It is likely that the inclusion of these previously unaccounted for active N₂ fixation sites will be more important in equilibrating denitrification and N₂ fixation rates than the underestimation of rates due to discrepancies between isotopic tracer methods.

In chronically stratified open ocean regions such as the vast subtropical gyres, primary production depends largely on external fixed nitrogen inputs provided by N₂ fixation performed by prokaryotes termed 'diazotrophs'. Diazotrophic cyanobacteria are photosynthetic prokaryotes (with the exception of the photoheterotrophic *Candidatus Atelocyanobacterium thalassa* which cannot photosynthesize)¹⁷ that thrive in oligotrophic tropical and subtropical waters of the oceans where they provide an important source of fixed nitrogen for other phytoplankton¹³. Despite being classically regarded as photoautotrophs, some unicellular diazotrophic cyanobacteria like *Cyanothece* are able to take up dissolved organic matter (DOM) molecules photoheterotrophically¹⁸. As well, various filamentous diazotrophic cyanobacteria such as *Anabaena* bear genes for amino acids transport, which may be used to incorporate amino acids from the *in situ* DOM pool, or to assimilate amino acids self-produced during diazotrophic growth¹⁹.

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The filamentous diazotrophic cyanobacterium *Trichodesmium* is ubiquitous in the tropical and subtropical oceans where it is estimated to contribute 60–80% of global N₂ fixation inputs²⁰. *Trichodesmium* is limited by iron and/or phosphate availability²¹ which are often scarce in oligotrophic subtropical gyres. The concomitant accumulation of DOM in these oligotrophic gyres²² where *Trichodesmium* thrives, suggests it could benefit from organic compounds. While most of the marine DOM is composed of refractory molecules that persist in seawater for millennia, labile DOM (degraded within hours or days) accumulates preferentially at the surface ocean as a result of photosynthesis products²³. *Trichodesmium* has been shown to assimilate organic phosphorus compounds such as phosphomonoesters and phosphonates when phosphate is scarce^{24,25} in an equally efficient manner to phosphate consumption²⁶, although a minimum availability of inorganic nutrients may be needed before *Trichodesmium* can cleave the carbon-phosphorus bond of phosphonates²⁷. On the other hand, while the uptake of carbon or nitrogen-rich DOM compounds has been studied in cultures of *Trichodesmium* (e.g. refs 28 and 29), such activity has not been revisited for almost two decades. The extent to which mixotrophic nutrition facilitates the growth and/or N₂ fixation in *Trichodesmium* remains poorly known, particularly for natural colonies. Here we quantify the uptake of carbohydrates and amino acids and their effect on N₂ fixation by natural *Trichodesmium* colonies using nano-scale secondary ion mass spectrometry (nanoSIMS).

Results

Station LDA presented relatively oligotrophic conditions at the surface with inorganic nutrient concentrations below the detection limit (0.02 μM for both nitrate -NO₃⁻ and phosphate -PO₄³⁻; Table S1), but high dissolved organic carbon (DOC; 95.34 \pm 2.81 μM) and relatively high chlorophyll *a* concentrations (0.36 \pm 0.05 $\mu\text{g L}^{-1}$; Table S1) when compared to typical open ocean regional values³⁰. Station LDB was sampled in an elevated chlorophyll *a* patch (0.83 \pm 0.07 $\mu\text{g L}^{-1}$) and exhibited lower DOC concentrations (70.65 \pm 0.09 μM). Bacterial abundance was > 3-fold higher at LDB than at LDA (Table S1).

Bulk N₂ fixation rates at LDA were 2.23, 4.61 and 4.10 nmol N L⁻¹ d⁻¹ for the control, carbohydrate and amino acid treatments, respectively. At station LDB, bulk N₂ fixation rates were ~9-, 5- and 2-fold higher than at LDA (21.28, 23.44 and 10.79 nmol N L⁻¹ d⁻¹, respectively; Fig. 1a,b). At LDA, the addition of both carbohydrates and amino acids increased bulk N₂ fixation but the variability among replicates was high, resulting in non-significant differences ($p > 0.05$) as observed in previous similar experiments¹⁵. No significant enhancement of bulk N₂ fixation rates were observed at station LDB for either treatment.

At both stations LDA and LDB, nanoSIMS analyses of individual trichomes (Table S2) revealed significant ¹³C-enrichments ($p < 0.0001$) by ~1.7-fold relative to the control upon both carbohydrate and amino acid additions (Fig. 1c,d). These additions also significantly enhanced the ¹⁵N-enrichment of *Trichodesmium* by ~1.2-fold at station LDA (both $p < 0.0001$), but not at station LDB, where a high degree of variability was observed between filaments (Fig. 1c,d). When comparing both stations, we observed that per-trichome carbon and nitrogen uptake rates were ~2- and 5-fold higher at LDB than at LDA (Fig. 1e,f). NanoSIMS example images of ¹³C and ¹⁵N enriched trichomes in control, carbohydrate and amino acid treatments are shown in Fig. 2a–f, respectively. The addition of both carbohydrate or amino acids enhanced per-trichome nitrogen uptake rates at each station, although increases were only statistically significant for amino acid additions (Fig. 1e,f).

Discussion

We present evidence of carbohydrate and amino acid uptake by natural *Trichodesmium* colonies in conditions usually regarded as optimal for their autotrophic growth (Table S1). The addition of either carbohydrates or amino acids increased per-trichome N₂ fixation rates compared to the control at both LDA and LDB, but only amino acid additions induced statistically significant per-trichome N₂ fixation enhancements (Fig. 1e,f). At LDB, the enhancement of per-trichome N₂ fixation rates with respect to the control upon the addition of either carbohydrates or amino acids was ~5-fold higher than at LDA, suggesting different DOM degradation patterns at LDB. LDB was located inside a massive chlorophyll patch, which had been drifting eastwards for several months (see chlorophyll *a* satellite images time lapse, where station LDB is represented by a pink cross; de Verneil, 2015). The persistence of the patch could be maintained by a regular input of inorganic nutrients via wet deposition, which often enhances primary production due to the high nutrient and trace metal content of volcanic ashes in this highly seismic active area of the South Pacific Ocean³¹. The wet deposition of inorganic nutrients could maintain both photosynthetic and N₂ fixation activities while promoting a dynamic production and consumption of DOM. The higher DOM uptake by *Trichodesmium* at LDB (Fig. 1d,f) suggests that higher *in situ* DOM availability at LDB compared to LDA, promoted the heterotrophic nutrition of *Trichodesmium* over growth on inorganic nutrients. DOC standing stocks were lower at LDB than at LDA likely due to a tight coupling between its production and consumption, as suggested by the higher bacterial abundance (Table S1) and production at LDB than at LDA (F. Van Wambeke, personal communication). Alternatively, differences in *Trichodesmium* DOM uptake between stations LDA and LDB could be influenced by different DOM uptake genetic ability in different *Trichodesmium* strains³².

Given the high energetic cost of CO₂ and N₂ fixation in cyanobacteria^{33,34}, the alternative nutrition on DOM is thought to alleviate energy shortages. For example, unicellular cyanobacteria use glycerol as an alternative to CO₂¹⁸, but the use of organic carbon substrates such as carbohydrates by *Trichodesmium* has seldom been observed and at low rates³⁵. Although the external input of combined nitrogen is thought to preclude N₂ fixation in *Trichodesmium*^{29,36,37}, our results show a significant enhancement of per-trichome N₂ fixation rates upon the addition of amino acids, as observed in other sites where heterotrophic diazotrophs predominate, like in mesopelagic waters^{15,38,39}. Amino acids may provide a more readily accessible source of organic carbon than carbohydrates, resulting in a greater enhancement of N₂ fixation rates.

DOM utilization likely confers nutritional plasticity to *Trichodesmium* in oligotrophic environments, reinforcing the obsoleteness of the categorical division of marine microbes into autotrophs or heterotrophs. Although we

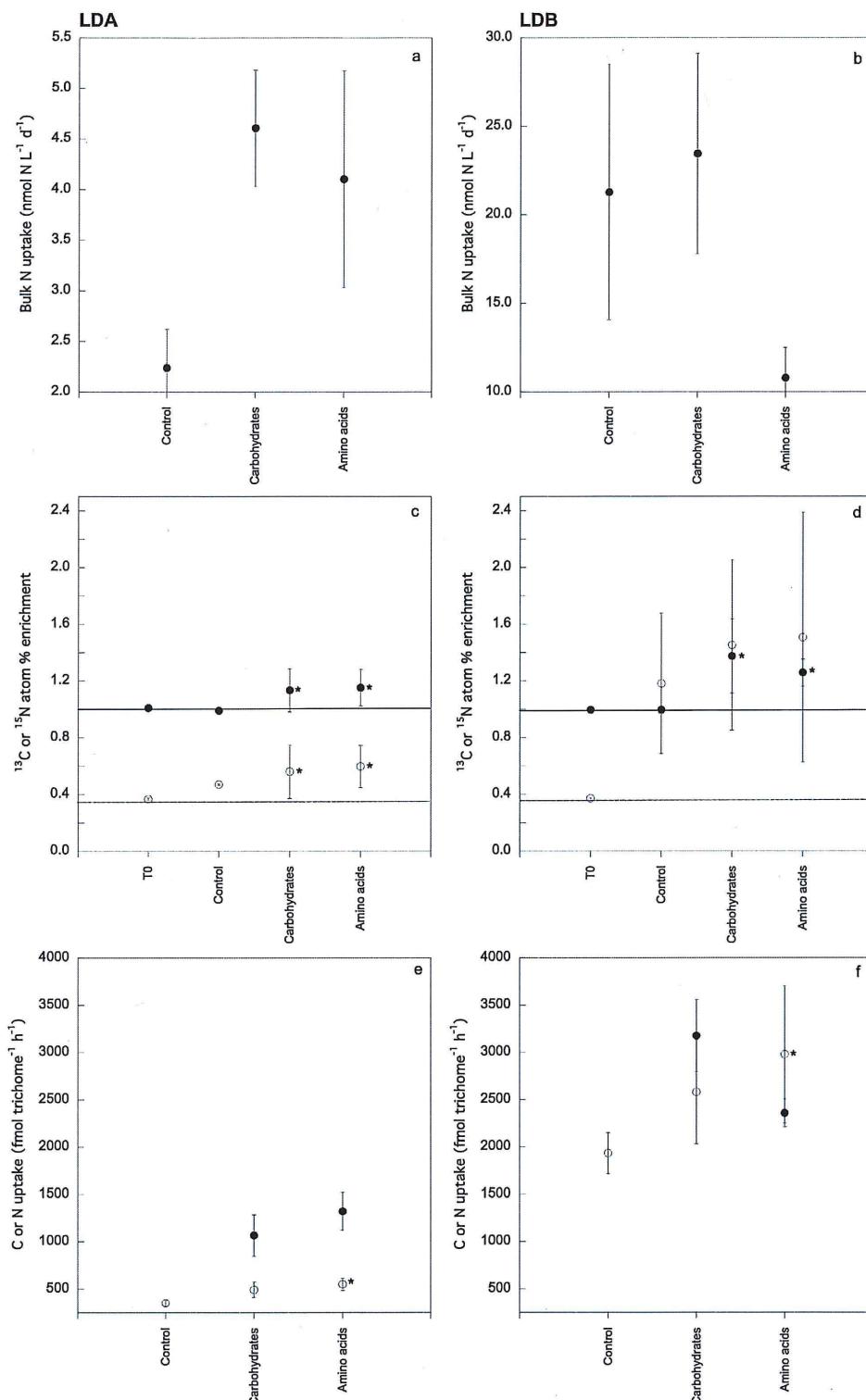


Figure 1. Bulk seawater N uptake rates at (a) station LDA, and (b) station LDB (note the different scale ranges on the y-axis). *Trichodesmium* (c,d) ¹³C-filled circles- and ¹⁵N-open circles- atom % enrichment values, and (e,f) ¹³C and ¹⁵N uptake rates at stations LDA and LDB, respectively. Error bars for C or N uptake rates (a,b,e,f) represent the standard error, while error bars for atom % graphs (c,d) represent the standard deviation of the mean. Straight and dotted lines in (c,d) indicate the average ¹³C and ¹⁵N atom % enrichment, respectively, of time zero samples (average natural abundance). Asterisks indicate statistically significant differences (Mann Whitney test, $p < 0.05$).

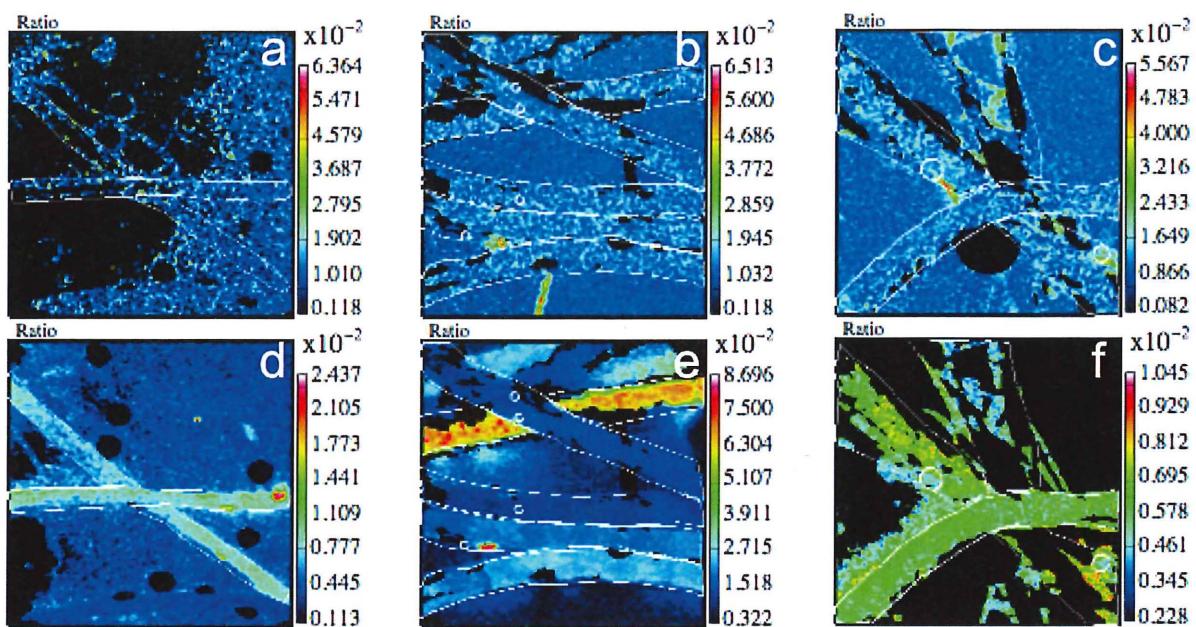


Figure 2. NanoSIMS trichome $^{13}\text{C}/^{12}\text{C}$ ratio images of (a) control, (b) carbohydrate-amended and (c) amino acid-amended samples. The corresponding $^{15}\text{N}/^{14}\text{N}$ ratio images are displayed below (d–f).

did not conduct ^{13}C -labeled bicarbonate uptake experiments during this cruise, previous experiments performed in the Southwest Pacific Ocean and in cultures of *Trichodesmium* IMS101 have shown per trichome bicarbonate uptake rates of $\sim 2\text{--}3 \times 10^6 \text{ fmol C trichome}^{-1} \text{ h}^{-1}$ ^{40,41}, which are in the same order of magnitude as the per trichome carbohydrate uptake rates measured here ($\sim 1\text{--}9 \times 10^6 \text{ fmol C trichome}^{-1} \text{ h}^{-1}$). This suggests that under certain environmental conditions, *Trichodesmium* may be able to exploit carbon comparably from inorganic and organic carbon sources.

Our results cannot however confirm whether DOM molecules were directly taken up by *Trichodesmium*, or if they were primarily reduced by epibiont bacteria and then transferred to the trichomes. For example, heterotrophic bacterial epibionts are known to facilitate dissolved organic phosphorus acquisition in *Trichodesmium* colonies⁴². Thus, the degree and/or functional diversity of epibiont bacteria colonization among sampling stations could have also influenced DOM uptake rates in our *Trichodesmium* samples⁴³. We observed bacteria appearing to be attached to trichomes in our samples (Fig. S1), and thus cannot rule out this possibility. Different incubation time span experiments are needed to discern whether DOM passes through bacteria before being taken up by *Trichodesmium*, or if *Trichodesmium* assimilates DOM directly. However, such short-term experiments would require a high isotopic enrichment of the source DOM pool, which would likely bias the measured uptake rates.

We present evidence of carbohydrate and amino acid uptake by natural *Trichodesmium* colonies. Climate change scenarios predict inorganic nutrient limitation and increased DOM retention within the photic zone⁴⁴, which will likely promote mixotrophy in *Trichodesmium*. Further studies on *Trichodesmium* organic versus inorganic nutrient acquisition are thus needed to predict how this important diazotroph will respond to climate alterations.

Methods

We sampled seawater at two stations in the Southwest Pacific (LDA: 19.21°S-164.68°E, LDB: 18.24°S-170.80°W, on 26 February and 15 March 2015, respectively) at depths receiving 50% of surface photosynthetically active radiation (corresponding to 7 and 9 m depth, respectively). The samples were incubated under *in situ* simulated conditions for 36 h with equimolar quantities of ^{13}C -labeled carbohydrates (sodium pyruvate, sodium acetate and glucose) or amino acids (alanine, leucine and glutamic acid; Sigma-Aldrich, Munich, Germany), added at concentrations of $4 \mu\text{M C}$ (final concentration for the mix of all three carbohydrates or all three amino acids³⁸). While the real marine DOM pool is molecularly highly complex and mostly refractory, these commercially available compounds were chosen as representative of carbohydrate and small organic acids typically found in marine labile DOM^{15,38,45}. Seawater was distributed into sixteen 4.3 L transparent polycarbonate bottles (Nalgene, Rochester, NY, USA). Four bottles were filtered immediately upon collection (T0), four were amended with the carbohydrate mix, and another four with the amino acids mix. The last four bottles were used as a control without amendments. All bottles were labeled with 6 mL 98.9 atom% $^{15}\text{N}_2$ gas (Cambridge Isotope Laboratories, Tewksbury, MA, USA) to assay N_2 fixation simultaneously. Of each quadruplicate set, three bottles were used to estimate bulk N_2 fixation rates (expressed as ‘N uptake’) and one bottle was used for nanoSIMS analyses (see Supplementary Information). Mann-Whitney statistical tests were used to test the significance of our results.

References

1. Falkowski, P. G. Evolution of the nitrogen cycle and its influence on the biological sequestration of CO₂ in the ocean. *Nature* **387**, 272–275 (1997).
2. Gruber, N. & Galloway, J. N. An Earth-system perspective of the global nitrogen cycle. *Nature* **451**, 293–296 (2008).
3. Codispoti, L. A. An oceanic fixed nitrogen sink exceeding 400 Tg N a⁻¹ vs the concept of homeostasis in the fixed-nitrogen inventory. *Biogeosciences* **4**, 233–253 (2007).
4. Mohr, W., Großkopf, T., Wallace, D. W. R. & LaRoche, J. Methodological Underestimation of Oceanic Nitrogen Fixation Rates. *PLOS ONE* **5**, e12583 (2010).
5. Großkopf, T. *et al.* Doubling of marine dinitrogen-fixation rates based on direct measurements. *Nature* **488**, 361–364 (2012).
6. Wilson, S. T., Böttjer, D., Church, M. J. & Karl, D. M. Comparative Assessment of Nitrogen Fixation Methodologies, Conducted in the Oligotrophic North Pacific Ocean. *Appl Environ Microbiol* **78**, 6516–6523 (2012).
7. Benavides, M. *et al.* Longitudinal variability of size-fractionated N₂ fixation and DON release rates along 24.5°N in the subtropical North Atlantic. *J. Geophys. Res. Oceans* **118**, 3406–3415 (2013).
8. Böttjer, D. *et al.* Temporal variability of nitrogen fixation and particulate nitrogen export at Station ALOHA. *Limnol Oceanogr*, doi: 10.1002/lno.10386 (2016).
9. Montoya, J. P., Voss, M., Kahler, P. & Capone, D. G. A simple, high-precision, high-sensitivity tracer assay for N₂ fixation. *Appl Environ Microbiol* **62**, 986–993 (1996).
10. Mulholland, M. R. *et al.* Rates of dinitrogen fixation and the abundance of diazotrophs in North American coastal waters between Cape Hatteras and Georges Bank. *Limnol. Oceanogr.* **57**, 1067–1083 (2012).
11. Bonnet, S., Baklouti, M., Gimenez, A., Berthelot, H. & Berman-Frank, I. Biogeochemical and biological impacts of diazotroph blooms in a low-nutrient, low-chlorophyll ecosystem: synthesis from the VAHINE mesocosm experiment (New Caledonia). *Biogeosciences* **13**, 4461–4479 (2016).
12. Shiozaki, T., Nagata, T., Ijichi, M. & Furuya, K. Nitrogen fixation and the diazotroph community in the temperate coastal region of the northwestern North Pacific. *Biogeosciences* **12**, 4751–4764 (2015).
13. Karl, D. M. *et al.* Dinitrogen fixation in the world's oceans. *Biogeochemistry* **57/58**, 47–98 (2002).
14. Loescher, C. R. *et al.* Facets of diazotrophy in the oxygen minimum zone waters off Peru. *ISME J* **8**, 2180–2192 (2014).
15. Benavides, M. *et al.* Mesopelagic N₂ Fixation Related to Organic Matter Composition in the Solomon and Bismarck Seas (Southwest Pacific). *PLOS ONE* **10**, e0143775 (2015).
16. Ratten, J.-M. *et al.* Sources of iron and phosphate affect the distribution of diazotrophs in the North Atlantic. *Deep-Sea Res Part II* **116**, 332–341 (2015).
17. Zehr, J. P. *et al.* Globally distributed uncultivated oceanic N₂-fixing cyanobacteria lack oxygenic photosystem II. *Science* **322**, 1110–1112 (2008).
18. Feng, X. *et al.* Mixotrophic and photoheterotrophic metabolism in *Cyanothece* sp. ATCC 51142 under continuous light. *Microbiology* **156**, 2566–2574 (2010).
19. Montesinos, M. L., Herrero, A. & Flores, E. Amino acid transport systems required for diazotrophic growth in the cyanobacterium *Anabaena* sp. strain PCC 7120. *Journal of Bacteriology* **177**, 3150–3157 (1995).
20. Mahaffey, C., Michaels, A. F. & Capone, D. G. The conundrum of marine N₂ fixation. *Am. J. Sci.* **305**, 546–595 (2005).
21. Bergman, B., Sandh, G., Lin, S., Larsson, J. & Carpenter, E. J. *Trichodesmium* - a widespread marine cyanobacterium with unusual nitrogen fixation properties. *FEMS Microbiol Rev* **37**, 286–302 (2012).
22. Hansell, D. A., Carlson, C. A., Repeta, D. J. & Schlitzer, R. Dissolved organic matter in the ocean: a controversy stimulates new insights. *Oceanography* **22**, 202–211 (2009).
23. Moran, M. A. *et al.* Deciphering ocean carbon in a changing world. *Proc Natl Acad Sci USA* **201514645**, doi: 10.1073/pnas.1514645113 (2016).
24. Dyhrman, S. T., Webb, E. A., Anderson, D. M., Moffett, J. W. & Waterbury, J. B. Cell-specific detection of phosphorus stress in *Trichodesmium* from the Western North Atlantic. *Limnol Oceanogr* **47**, 1832–1836 (2002).
25. Dyhrman, S. T. *et al.* Phosphonate utilization by the globally important marine diazotroph *Trichodesmium*. *Nature* **439**, 68–71 (2006).
26. Beversdorf, L. J., White, A. E., Björkman, K. M., Letelier, R. M. & Karl, D. M. Phosphonate metabolism by *Trichodesmium* IMS101 and the production of greenhouse gases. *Limnol Oceanogr* **55**, 1768–1778 (2010).
27. del Valle, D. A. & Karl, D. M. Aerobic production of methane from dissolved water-column methylphosphonate and sinking particles in the North Pacific Subtropical Gyre. *Aquat. Microb. Ecol.* **73**, 93–105 (2014).
28. Mulholland, M. R., Ohki, K. & Capone, D. G. Nitrogen utilization and metabolism relative to patterns of N₂ fixation in cultures of *Trichodesmium* NIBB1067. *J Phycol* **35**, 977–988 (1999).
29. Ohki, K., Zehr, J. P., Falkowski, P. G. & Fujita, Y. Regulation of nitrogen-fixation by different nitrogen sources in the marine non-heterocystous cyanobacterium *Trichodesmium* sp. NIBB1067. *Arch Microbiol* **156**, 335–337 (1991).
30. Aufdenkampe, A. K. *et al.* Biogeochemical controls on new production in the tropical Pacific. *Deep-Sea Research II* **49**, 2619–2648 (2002).
31. Martino, M. *et al.* Western Pacific atmospheric nutrient deposition fluxes, their impact on surface ocean productivity. *Global Biogeochemical Cycles*, doi: 10.1002/(ISSN)1944-9224 (2014).
32. Rouco, M., Joy-Warren, H., McGillicuddy, D. J., Waterbury, J. B. & Dyhrman, S. T. *Trichodesmium* sp. clade distributions in the western North Atlantic Ocean. *Limnol Oceanogr* **59**, 1899–1909 (2014).
33. Kranz, S. A., Gladrow, D. W., Nehrke, G., Langer, G. & Rosta, B. Calcium carbonate precipitation induced by the growth of the marine cyanobacteria *Trichodesmium*. *Limnol Oceanogr* **55**, 2563–2569 (2010).
34. Holl, C. M. & Montoya, J. P. Interactions between nitrate uptake and nitrogen fixation in continuous cultures of the marine diazotroph *Trichodesmium* (Cyanobacteria). *J Phycol* **41**, 1178–1183 (2005).
35. Paerl, H. W., Bebout, B. M. & Prufert, L. E. Bacterial associations with marine *Oscillatoria* sp. (*Trichodesmium* sp.) populations: ecophysiological implications. *J Phycol* **25**, 773–784 (1989).
36. Mulholland, M. R. & Capone, D. G. Nitrogen fixation, uptake and metabolism in natural and cultured populations of *Trichodesmium* spp. *Mar Ecol Prog Ser* **188**, 33–49 (1999).
37. Fu, F.-X. & Bell, P. R. Factors affecting N₂ fixation by the cyanobacterium *Trichodesmium* sp. GBRTRLI101. *FEMS Microbiol Ecol* **45**, 203–209 (2003).
38. Bonnet, S. *et al.* Aphotic N₂ Fixation in the Eastern Tropical South Pacific Ocean. *PLOS ONE* **8**, e81265 (2013).
39. Rahav, E. *et al.* Dinitrogen fixation in aphotic oxygenated marine environments. *FMICB* **4**, 1–11 (2013).
40. Berthelot, H., Bonnet, S., Grosso, O., Cornet, V. & Barani, A. Transfer of diazotroph-derived nitrogen towards non-diazotrophic planktonic communities: a comparative study between *Trichodesmium erythraeum*, *Crocospheara watsonii* and *Cyanothece* sp. *Biogeosciences* **13**, 4005–4021 (2016).
41. Bonnet, S. *et al.* Diazotroph derived nitrogen supports diatom growth in the South West Pacific: a quantitative study using nanoSIMS. *Limnol Oceanogr*, doi: 10.1002/lno.10300 (2016).
42. Van Mooy, B. A. S. *et al.* Quorum sensing control of phosphorus acquisition in *Trichodesmium* consortia. *ISME J* **6**, 422–429 (2012).
43. Rouco, M., Haley, S. T. & Dyhrman, S. T. Microbial diversity within the *Trichodesmium* holobiont. *Environ Microbiol*, doi: 10.1111/1462-2920.13513 (2016).

44. Passow, U. & Carlson, C. A. The biological pump in a high CO₂ world. *Mar Ecol Prog Ser* **470**, 249–271 (2012).
45. Dekaezemacker, J. *et al.* Evidence of active dinitrogen fixation in surface waters of the eastern tropical South Pacific during El Niño and La Niña events and evaluation of its potential nutrient controls. *Global Biogeochemical Cycles* **27**, 768–779 (2013).

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Author Contributions

Conceived and designed the experiments: M.B., S.B., S.D. Performed the experiments: M.B., S.D., H.B. Analyzed the data: M.B., H.B. Contributed reagents/ materials/ analysis tools: P.R., S.D., S.B. Wrote the paper: M.B., S.B., S.D., H.B.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

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