

1 AN EVALUATIVE COMPARISON OF LIGNOCELLULOSIC PYROLYSIS PRODUCTS
2 DERIVED FROM VARIOUS PARTS OF *POPULUS DELTOIDES* TREES AND *PANICUM*
3 *VIRGATUM* GRASS IN AN INDUCTIVELY HEATED REACTOR

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13

14 **ABSTRACT**

15 Poplar is considered a suitable resource for production of renewable fuels and chemicals due to
16 its rapid growth and tolerance to environmental stresses. Switchgrass is also extensively studied
17 for biofuel production due to its use as a resource-efficient low-input plant and ability to grow
18 and thrive in diverse weather or soil conditions. In this study, fast pyrolysis of biomass obtained
19 from various parts (main stem, secondary stems, branches) of eastern cottonwood (*Populus*
20 *deltoides*) and switchgrass (*Panicum virgatum*) was carried out in an inductively heated reactor.

21 Devolatilization rates (ranging from 450 °C to 600 °C) were initially obtained to determine their
22 decomposition kinetics and estimate process parameters (temperatures and times) suitable for
23 their pyrolysis in an inductively-heated reactor. The effect of temperature (450 °C, 500 °C, 550
24 °C) on pyrolysis product yields and composition was investigated. Results indicate that
25 activation energies ranged from 9.2 to 13.5 kJ/mol, while pre-exponential constants ranged from
26 0.23 to 0.51 s⁻¹. The maximum quantity of bio-oil of 39.8% ± 9.50 was obtained from poplar
27 stem at 450 °C whereas the least amount of 33% was obtained from poplar branch at 550 °C. The

28 highest amount of bio-oils of 34% from switchgrass was obtained at 450 °C. Water content in the
29 bio-oil obtained from switchgrass was significantly higher than those from poplar tree parts. The
30 GC-MS results showed that bio-oil compositions are similar among the various parts of poplar
31 trees, with phenols being the dominant chemical specie for all bio-oil samples analyzed. With
32 increasing temperature, an increase in furans is observed. Bio-oil fractions derived from biomass
33 pyrolysis of the trunk and stems have higher HHV than those from switchgrass, with the average
34 HHV of water-free bio-oils ranging from 17.4 - 19.4 MJ/kg. Overall process energy recovery
35 from initial biomass reached a maximum of 80.1% in the case of cottonwood main stem
36 pyrolyzed at 500 °C.

37

38 **KEYWORDS:** Cottonwood poplar; stem; branch; switchgrass; inductive pyrolysis; bioenergy

39

40 1. INTRODUCTION

41 Global challenges such as the disruption to the supply of limited fossil fuels, spikes in energy
42 costs, consistent rise in world energy demand, and the need for lower greenhouse gas (GHG)
43 emissions have informed substantial research efforts in the conversion of biomass into biofuels
44 and other beneficial renewable products [1-3]. Biomethanation, fermentation and
45 thermochemical methods are commonly used approaches to obtain biofuels from biomass
46 products [4]. For instance, conversion of biomass into biogas, ethanol, and biodiesel can be done
47 by microbial/enzymatic fermentations with the aid of physical and chemical pretreatment steps
48 [5, 6]. Bio-oil, biochar and syngas are produced from biomass using entirely thermochemical
49 conversion processes such as torrefaction, carbonization, hydrothermal liquefaction, gasification,
50 and pyrolysis. These processes yield fuels with high-energy content with minimal environmental
51 impact [3, 7]. The thermochemical conversion method to be used depends on the available raw

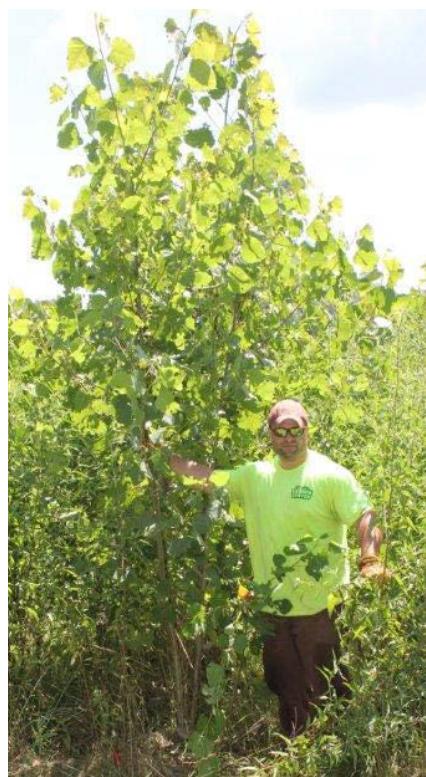
52 material and the desired product. Pyrolysis is a valuable thermochemical process which converts
53 biomass into combustible gas, bio-oil, and char in an oxygen-free environment within reactor
54 temperatures range of 300 $^{\circ}\text{C}$ and 900 $^{\circ}\text{C}$ [8].

55 Fluidized bed reactors, ablative pyrolysis reactors, and fixed bed drop-type reactors are some of
56 the several reactor systems that have been designed to optimize liquid oil reduction [9]. To
57 obtain higher quantities of liquid oil from the pyrolysis process, several features have been
58 identified as important, including rapid heating rates, high heat transfer-to-biomass rates,
59 methodical control of the reactor temperature, rapid removal of the pyrolysis vapors from the
60 reactor, and the rapid cooling of the vapors to make the bio-crude products [10]. Based on these
61 features, an induction-heating reactor has proven to be a suitable alternative to carry out the
62 pyrolysis process [9], as compared to other fast/flash pyrolysis systems due to its high-energy
63 efficiency and low pollution rates [11]. The induction-heating system is designed to heat any
64 ferromagnetic material placed within its coils by creating an oscillating magnetic field with the
65 aid of an AC power supply. Since the heat is generated inside the material itself, during induction
66 heating, and not from an external source, process objectives such as rapid heating rates and high
67 energy efficiency are readily achieved. Although induction heating mechanism has been
68 demonstrated to be effective both in the pyrolysis of biomass and subsequent bio-oil upgrading,
69 major downsides of the mechanism include low performance efficiency and minimal tolerance
70 for design flexibility or expansions [12].

71 The eastern cottonwood tree (*Populus deltoides*) is tree is one of the fastest-growing trees in
72 North America and Europe [13], as it can grow up to 30 m tall and adapt to a wide gamut of soil
73 and climatic conditions. Although eastern cottonwood is not a quality source of timber due to its
74 fragile nature, it presents potential as a bioenergy crop due to its rapid growth, high rate of

75 biomass production, and its high cellulose and low lignin contents [14]. These features are
76 significant in the production of liquid fuels because the cellulose provides the substrate for liquid
77 fuels whereas its low lignin content makes it suitable for various pretreatment processes [14]. It
78 has been established that short-rotation woody crops (SRWCs) in which trees such as
79 cottonwood are managed for rotations of 10 years or less have lower environmental footprints
80 than fossil fuels [15]. The studies of the effect of pyrolysis on various parts of the cottonwood
81 tree are limited to non-existent. In this paper, we compare the effect of induction-heating
82 pyrolysis on the various parts of cottonwood tree – stem, branch, main trunk – using a laboratory
83 scale induction heating reactor.

84 Switchgrass (*Panicum virgatum* L.) is a native warm-season grass used for hay and forage crops
85 in the United States. Switchgrass has several characteristics which makes it suitable as a biomass
86 energy crop in the US, including being native in North America, consistently growing in diverse
87 planting conditions, producing average to high biomass yields on marginal soils [16], and
88 existing as an environmental enhancement energy crop [17]. For these reasons, switchgrass is
89 well positioned to be used as a bioenergy feedstock for pyrolysis, gasification, cellulosic ethanol
90 production, and combustion for heat and electrical generation [18]. While there are several
91 studies on ethanol production from these feedstocks [19-22], there is only limited published
92 literature on the pyrolysis of cottonwood and switchgrass biomasses [23]. Moreover, there is a
93 knowledge gap in terms of induction-based pyrolysis for both feedstocks. In this study, we aim
94 to fill this gap by investigating devolatilization kinetics in inductively heated reactor to evaluate
95 the yield and quality of the residual bio-oils. The process performance for the various parts of the
96 poplar tree is critically compared, as from a practical perspective, depending on specific
97 management of the trees, only select parts of the tree may be harvested for biofuel production.



98

99 Figure 1. Left: Naturally occurring cottonwood (*Populus deltoids*) tree. Right: Various parts of
100 managed cottonwood tree – person holding main stem (largest of the shoots), secondary stems
101 shooting from the stool, and branches (twigs and branches off the main and secondary stems)

102 2. MATERIALS AND METHODS

103 2.1. Materials

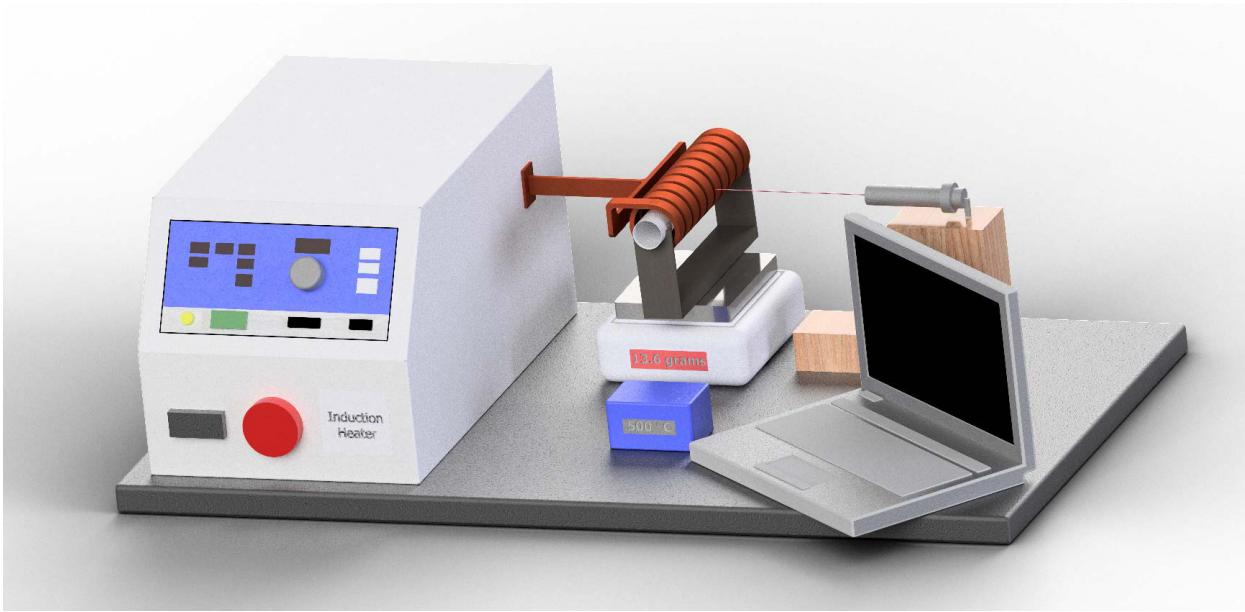
104 Eastern cottonwood and switchgrass biomass samples were collected at a research site in
105 northeastern Louisiana as part of a larger project described in previous reports [24, 25]. Both
106 biomass types were collected in the fourth year after planting at the retired agricultural site.
107 Eastern cottonwood trees were trimmed and sorted based on the part of the tree from which the
108 trimmings was collected viz-a-viz stem, branch, and main trunk. Switchgrass was cut and broken
109 by hand into smaller pieces. All samples were ground with the aid of a mill able to produce
110 particles of uniform size that are less than 1 mm in diameter. The measured particle size
111 distribution of the ground biomass, alongside the moisture content determined gravimetrically
112 according to ASTM C566 – 13 [26], was recorded. Moisture content of the biomass parts ranged

113 from 2.2% wet basis for cottonwood stem to 4.8% wet basis for switchgrass. The milled biomass
114 was placed in thin layer on trays and oven-dried overnight at 105 °C. The dried biomass was then
115 stored in sealed plastic bags at low temperatures to minimize moisture absorption during storage
116 until the experiments are ready to be run.

117 ***2.2. Equipment***

118 For the initial experiments determining the devolatilization kinetics, a free-standing stainless
119 steel reactor (a 10 cm long cylinder with an internal diameter of 2.54 cm), capped at one end and
120 with a small opening at the other, was placed in an induction coil connected to an RF generator
121 (2 kW max power, model Flex Heat 2, RDO Induction, LLC, Washington, NJ). The reactor was
122 placed on a support system consisting of an fabric insulating layer, a quartz half-cylinder, and
123 wood legs, all located on a balance (with resolution of 0.1 g) as shown in Figure 2 below. The
124 balance (model #, Ohaus, Inc, CITY, STATE) was connected to computer that recorded the mass
125 loss in real time.

126 For the second study which included gas and bio-oil collection and analysis, a low-frequency
127 induction heater (RDO Induction L.L.C., Washington, NJ) with a power output of 5 kW was
128 used to heat a 1.5-inch nominal diameter stainless steel reaction tube (which held the biomass) at
129 various temperatures [9]. An infrared Omega iR2C PID controller (Omega Engineering, Inc.,
130 Stamford, CT) was used to control the power output of the induction heating system, with the aid
131 of a 4–20 mA control signal to achieve and maintain the desired reactor temperature. The
132 reaction tube was a 316 – stainless steel tri-clamp tube, 454 mm in length, with inner and outer
133 diameters of 34.925 mm and 50.8 mm respectively. The biomass was compactly packed in the
134 reactor which was placed within a ten-loop rubber-coated copper induction coil 285 mm in
135 length and 59 mm inner diameter.



136

137 Figure 2. Autodesk Inventor drawing of pyrolysis system. The balance was only used for the
138 collection of kinetic data.

139

140 The pyrolysis vapors were collected and condensed into liquid bio-oil as condensate using a
141 system consisting of a round bottomed flask placed in an ice bath at 0 °C and an electrostatic
142 precipitator (ESP) to prevent the dissolved liquid from vaporizing and prevent secondary
143 reactions by reducing the system temperature [27, 28]. The ESP was specifically built for this
144 study from a glass recipient (Technical Glass Products INC., Painesville, OH) and operated at a
145 15-kV power level with the aid of a Gamma High Voltage power supply (Gamma High Voltage
146 Research, Ormond Beach, FL). The ESP exhaust was connected to a water-ethanol dual trap
147 system and the resulting non-condensable, non-soluble gases were sampled and weighed.

148

149 **2.3. Experimental procedure**

150 To determine devolatilization kinetics and pyrolysis times, triplicate experiments were conducted
151 without collecting products. Dried and ground cottonwood and switchgrass biomass were

152 pyrolyzed in a batch induction pyrolysis system at four different reactor bed temperatures: 450,
153 500, 550, and 600°C. Kinetic experiments indicated that the residence time after which the
154 biomass recorded no change in mass was 15 minutes at most. The electric field impulses during
155 active heating introduced transient effects in the mass balance sensor, resulting in significant
156 noise in the signal. This noise was eliminated by performing a 20-second moving average on
157 each data set and averaging the three replicates.

158 *Analysis of Reaction Kinetics*

159 The kinetic reactions were modeled using the Borchardt and Daniels method [29] which
160 describes the dependence of the rate of reaction on the amount of material present as shown in
161 Equation 1.

162
$$\frac{d\alpha}{dt} = k(T)(1 - \alpha)^n \quad (1)$$

163 Where: $\frac{d\alpha}{dt}$ = reaction rate (s^{-1}), α = fraction reacted (dimensionless), $k(T)$ = rate constant at
164 temperature T (s^{-1}), n = reaction order (dimensionless).

165 The rate constant $k(T)$ was determined by calculating the slope of the linear portion of the
166 experimental mass loss data plotted versus time. The Borchardt and Daniels model also uses the
167 Arrhenius equation to describe how the reaction rate changes as a function of temperature as
168 shown in Equation 2.

169
$$k(T) = A e^{-\frac{E_a}{RT}} \quad (2)$$

170 Where: A = Arrhenius pre-exponential factor (s^{-1}), E_a = Activation energy ($J \text{ mol}^{-1}$), T = Absolute
171 temperature (K), R = Gas constant ($8314 \text{ J mol}^{-1} \text{ K}^{-1}$). E_a was determined by plotting $k(T)$ versus

172 the \ln of $1/T$ and determining the slope of the resulting line. Multiplying the slope by $-1/R$ gave
173 E_a . A was then determined as the linear slope of a plot of $k(T)$ versus $e^{-E_a/RT}$.

174 A second set of studies was performed on the biomass in a similar, but larger reactor (30 cm
175 long, 3.5 cm internal diameter), modified to allow for more biomass loads as well as for the
176 collection of products. In order to account for the higher initial mass and the larger reactor, as
177 well as to insure a complete pyrolysis, all experiments in the secondary study were uniformly run
178 at a conservative residence time of at least 30 minutes. Based on our previously published work
179 [28, 30], it was estimated that no additional benefits are derived from processing the biomass at
180 600 °C, thus these experiments were carried out only at 450, 500, and 550 °C. The dried biomass
181 was weighed out in 25 g sizes and packed in the center of the pyrolysis reaction tube. The
182 reaction tube was then placed in the induction coil, connected to the gas inlet and outlet, and in
183 such a way that the laser light from the infrared PID controller would focus on the center of the
184 tube.

185 All apparatus including the ESP condenser tube, round bottomed flask and connector stem were
186 cleaned, dried and weighed. Nitrogen gas flowing at 1.0 L min^{-1} was bubbled through the inlet to
187 purge the system of oxygen for 25 min. After purging, the induction heating system was turned
188 on to pyrolyze the biomass while the nitrogen gas flow was maintained. The ESP, placed in an
189 ice bath, served as a conduit for condensable vapors (bio-oil) into the collection flask, while non-
190 condensable vapors were filtered out via the ethanol and water bubble outlet.

191 At the end of the experiment, the residue in the reaction tube was weighed to obtain the char
192 yield. The content of the collection flask was weighed to determine the bio-oil yield. Obtained
193 bio-oil yield was transferred to scintillation vials and stored at -20°C to prevent secondary

194 reactions until further analysis. The mass of the syngas was determined by difference according
195 to the following:

196 $\text{Mass of syngas} = \text{Mass of biomass feedstock} - (\text{Mass of bio-oil} + \text{Mass of biochar}) \quad (3)$

197

198 ***2.4. Characterization and analysis***

199 Cottonwood and switchgrass biomass pyrolysis products were qualitatively analyzed to
200 determine the chemical compositions of the various product fractions. The proximate analyses
201 of the samples were carried out according to ASTM standards [moisture content (MC): ASTM
202 E871-82; ash content (Ash): ASTM E1755-4582; volatile matter (VM): ASTM E872-82, while
203 the fixed carbon (FC) content was determined according to $\text{FC} (\%) = 100\% - [\text{MC} \% + \text{Ash} \% +$
204 $\text{VM} \%]$. The ultimate analysis was carried out according to ASTM D5373 using an elemental
205 analyzer 2400 Series 2 CHNS/O (Perkin Elmer, Inc., Waltham, MA). The solid char samples
206 were analyzed for their Carbon, Hydrogen, and Nitrogen content using the same elemental
207 analyzer. The ash content of each char resides was also tested to determine its Oxygen content.
208 The liquid fraction was analyzed using three different analysis techniques: Karl Fischer titrations
209 to determine water content, CHN elemental analysis, and GC-MS to identify the different
210 compound groups in the liquid fraction. The detailed description of these processes has been
211 provided [31]. High heating values of the liquid fractions were measured using bomb calorimeter
212 (Parr Instrument Company, Moline, IL.) according to the procedure already described elsewhere
213 [32].

214 ***2.5. Carbon and energy balances***

215 The carbon balance (a primary component of the heating value) was performed to determine the
216 carbon distribution in the pyrolysis products, which estimates the extent of temperature effects
217 on individual component yields. The CHNO content of bio-oil and bio-char were analyzed using
218 Perkin Elmer 2100 series elemental analyzer. The energy content of the biomass was obtained
219 using the equation provided by Scholze and Meier to compute the higher heating value (HHV) of
220 the liquid product using the CHN analysis, equivalent to that developed by Dulong [33]:

221
$$HHV \left(\frac{\text{MJ}}{\text{kg}} \right) = \left[338.2 * C\% + 1442.8 * \left(H\% - \frac{O\%}{8} \right) \right] * 0.001 \quad (4)$$

222 Where C, H, and O are mass fractions obtained from pyrolysis analysis.

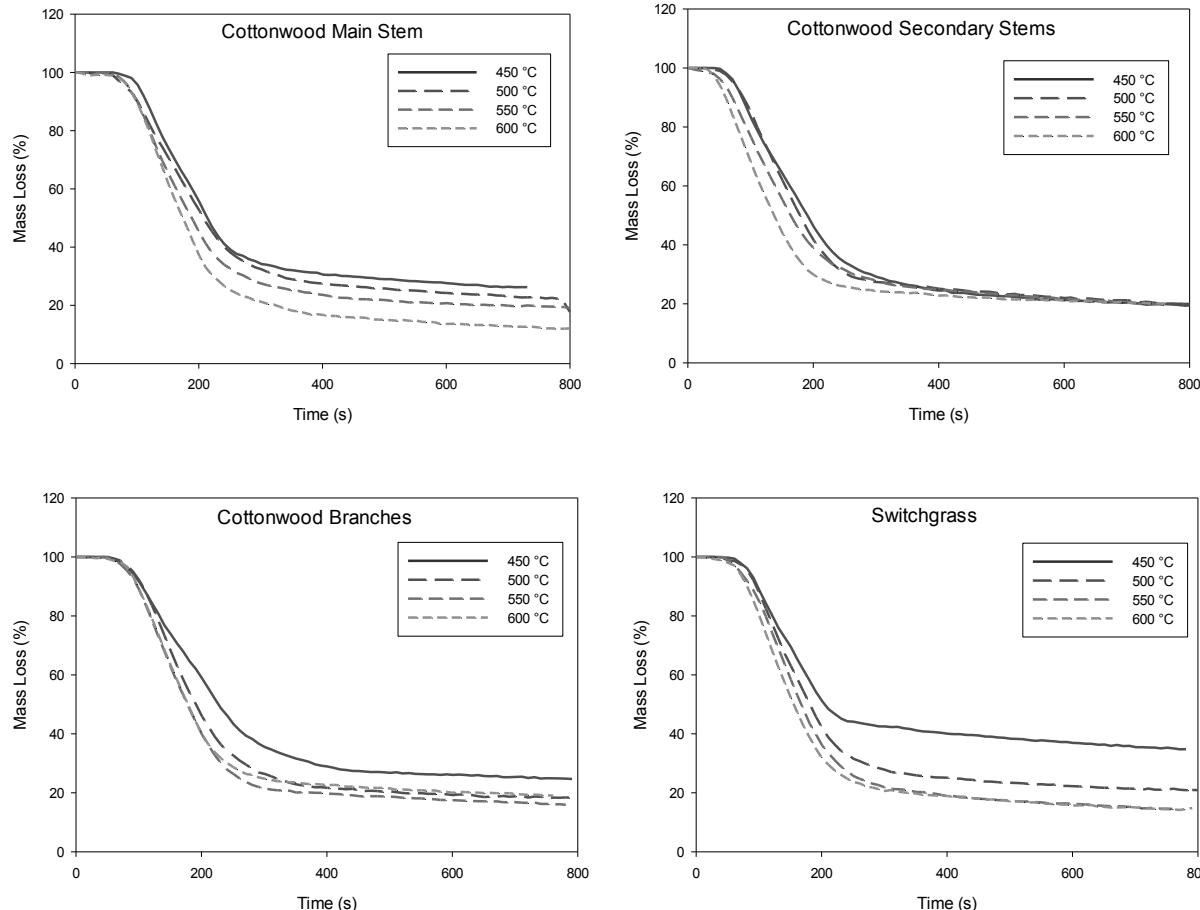
223 Likewise, the energy content of the four biomasses were determined using the equations
224 provided by Dulong and Vandalek [33], using the C, H, N, O, and ash contents of the
225 feedstocks.

226 **3. RESULTS AND DISCUSSION**

227 ***3.1. Reaction Kinetics***

228 The mass loss data (Figure #) indicated a clear relationship between the process temperature and
229 both the overall process time and the rate at which this mass loss occurs. In all cases, a higher
230 temperature resulted in a more rapid decomposition. With the exception of secondary stems,
231 which seemed to converge toward the same value (18.5% to 19%) indifferent of the process
232 temperature, the final char content decreased with an increase in temperature. For cottonwood
233 main stem the range was 12.1 to 26.2% and for branches was 16.3 to 24.6%, whereas for
234 switchgrass the range was 14.6 to 34.6%. This higher stability at lower temperatures compared to

235 cottonwood indicates the different internal structure of switchgrass, containing more ash-type
 236 compounds binding the organic carbon more tightly into the matrix and resisting decomposition.



237

238

239 *Figure #. Mass loss in the inductively heated reactor for the four biomasses under investigation.*

240 The devolatilization kinetic rates are presented in Table x. As expected, in general mass loss
 241 rates increase with temperature for all samples tested. The activation energies and pre-
 242 exponential constants calculated using the Borchardt and Daniels model are also presented in
 243 Table x. Compared to values presented in literature by Sadaka *et al.* [34] for TGA analysis, these
 244 values are markedly different (9.2 to 13.5 kJ/mol in this study vs. 234.5 to 300.3 kJ/mol in [34]).
 245 Similarly, the pre-exponential constants are multiple orders of magnitude smaller (0.23 to 0.51 s⁻¹
 246 ¹ in this study vs. 2.26E+13 to 3.94E+18 s⁻¹ in [34]). These differences are explained by the

247 specific experimental conditions. In the study by Sadaka et al. [34], the kinetic parameters were
 248 calculated using the Kissinger method [35], on an extremely small sample (milligram in size)
 249 compared to what is presented in this study (12 g). The TGA method also uses for calculation
 250 different heating rates as a parameter, rather than different temperatures. Thus, the results cannot
 251 be directly compared due to the differences in samples sizes, reactor configurations, and method
 252 of heating. When compared to similar size samples and reactors, conventionally heated in a
 253 furnace [36] (personal communications), the activation energies and pre-exponential constants
 254 are in the same range. For white poplar (a similar but different subspecies of *Populus*) E_a and A
 255 were determined to be 30.5 kJ/mol and 4.7 s^{-1} , respectively [36] (personal communications).
 256 Also, recent data on food court waste in similar size and reactor systems [37] indicate E_a of
 257 21.32 kJ/mol, A of 6.1 s^{-1} for induction heating vs. E_a of 30.75 kJ/mol and A of 10.9 s^{-1} in
 258 conventional heating.

259 Table 1: Values of the thermal decomposition rate constant $k(T)$, activation energy and
 260 Arrhenius pre-exponential factor for fractionated Cottonwood (main trunk, secondary stem, and
 261 branches) and Switchgrass undergoing fast pyrolysis in an inductively heated reactor.

Material	$k(T) (s^{-1})$				$E_a (J mol^{-1})$	$A (s^{-1})$
	450 °C	500 °C	550 °C	600 °C		
Cottonwood – Main stem	0.40	0.37	0.43	0.52	9,198	0.37
Cottonwood – Secondary stems	0.36	0.45	0.41	0.44	5,502	0.51
Cottonwood - Branches	0.32	0.46	0.48	0.48	13,507	0.23
Switchgrass	0.38	0.46	0.50	0.50	10,023	0.42

262

263 **3.2. Product yields**

264 The results show the volatile matter, fixed carbon, ash content, and moisture present (dry basis)
 265 in the raw biomasses and resulting bio chars (Table 1). These values are comparable with results
 266 presented in literature for similar biomasses [34].

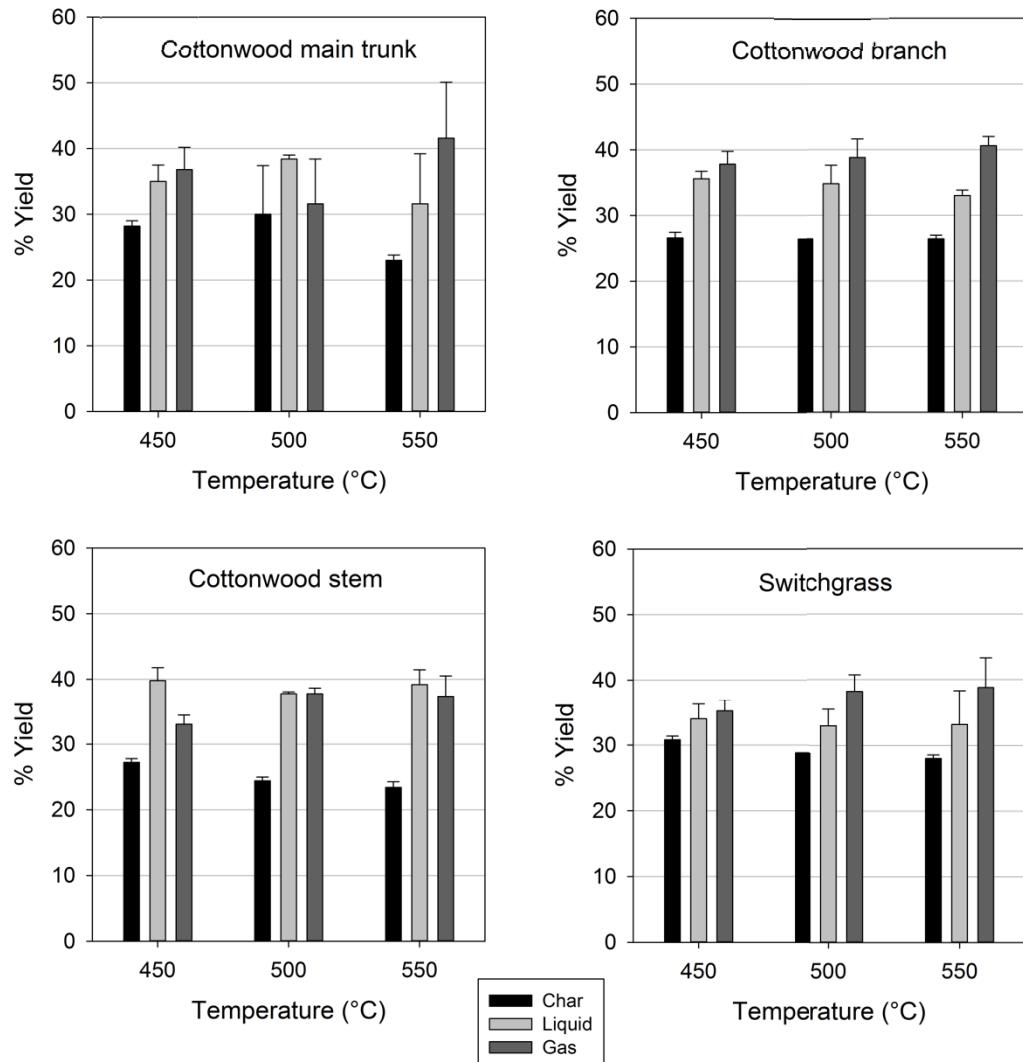
267 Table 2: Proximate and ultimate analysis of biomass (in %).

Biomass	Cottonwood stem	Cottonwood branch	Cottonwood main trunk	Switchgrass [38]
Proximate analysis	% Volatile matter	66.9±19.15	70.83±8.25	65.85±2.75
	% Fixed carbon	23.42	19.05	23.67
	% Ash content	1.53±0.19	1.75±0.13	2.14±0.29
	% Moisture	8.15±0.09	8.37±0.03	8.34±0.10
Ultimate analysis	% C	48.5	49.0	48.5
	% H	5.9	6.1	6.0
	% N	0.6	0.8	0.7
	% O	43.2	41.4	43.0

268

269 The product yields from cottonwood stem, trunk, and branches, and switchgrass were
 270 investigated to better understand the effects of reaction temperature on yields (Figure 3). All
 271 biomasses show a decrease in char yield as temperature increased from 450 °C to 550 °C.
 272 However, an unexpected spike in char yield was observed for cotton wood main stem from 450
 273 °C to 550 °C, which eventually reduced at 550 °C. For the cottonwood main trunk, char yield
 274 increased with temperature increase of 450 °C to 500 °C but reduced beyond that range. Char
 275 yield was however uniform at all experiment temperatures for cottonwood branches. Also, char
 276 yield decreased as temperature increased in the cases of cottonwood stem and switchgrass. All
 277 biomasses exhibited a decrease in char yield as the temperature increased from 450 °C to 500 °C.
 278 However, cottonwood branch displayed the lowest decrease in char yields (0.75%) with respect
 279 to temperature as compared to the other three biomasses, with the absolute percentage decrease
 280 in char yield from 450 °C to 550 °C being 18.31%, 12.0% and 9.1% for cottonwood main stem,
 281 cottonwood stem, and switchgrass respectively. The small statistical significance in char yield
 282 for cottonwood branches shows that cottonwood branches are more rapidly converted to char at
 283 lower temperatures than switchgrass other biomasses and/or the other cottonwood tree parts.
 284 Sadaka *et al.* [34] reported char yields for cottonwood in the range of 36.9 – 45.7%, and 38.7%
 285 for switchgrass, higher that was was found here, but those experiments were performed only at

286 400 °C, a lower temperature which is expected to lead to a higher char yield. The individual
 287 yields as percentages of total product for char, bio-oils and gases at different temperatures for
 288 each biomass sample are presented in Figure 3.



289
 290 Fig. 3: Yields of char, liquid bio-oil, and gas as % of total products from pyrolysis of biomasses
 291 derived from cottonwood main trunk, branch, and stem; and switchgrass.

292
 293 Bio-oil yield for poplar main stem was 38.4% at 500 °C while bio-oil yields were highest for the
 294 secondary stems, branches and switchgrass at 39.8% ± 0.019, 35.6% ± 0.011, and 34% ± 0.023

295 respectively, all at 450 °C. Bio-oil obtained from poplar secondary stems had the greatest yield
296 of $39.8\% \pm 0.019$ on average, while that obtained from poplar branch was least in bio-oil yield at
297 $34.5\% \pm 0.011$. It can thus be inferred that more quantities of bio-oil can be derived from poplar
298 stems compared to branches. Bio-oil yields decreased as temperature increased, reaching a
299 lowest value of 31.6% for cottonwood main stem and 33% for both cottonwood branch and
300 switchgrass at 550 °C. The highest bio-oil yield of 39.8% was obtained from cottonwood
301 secondary stems at 450 °C (Fig. 3). The maximum bio-oil yield for other biomasses were 38.4%
302 at 500 °C for cottonwood main stem, 35.6% at 450 °C for cottonwood branch, and 34% at 450 °C
303 for switchgrass. Published data for bio-oil yield obtained in a conventional reactor at 400 °C
304 (lower than what is reported here) indicate bio-oil yields of 33-35% for cottonwood and 29.5%
305 for switchgrass [34], which are lower than what is reported in the present work. It is therefore
306 concluded that maximum bio-oil yields are obtained at temperatures in the range of 450-500 °C.
307 Sadaka *et al.* [34] did not perform pyrolysis studies for yield determination at other temperatures,
308 except 400 °C, so a clear comparison between conventional and induction heating cannot be
309 made.

310 Volatile matter is released in larger quantities and at a higher rate as pyrolysis temperatures
311 increase up to a certain point [1, 39]. Pyrolysis at this temperature range results in optimum
312 extraction of the volatile condensable elements from biomass feedstock [40]. Beyond this
313 temperature, volatile matter undergoes secondary tar reactions such as thermal cracking [41], and
314 decomposes into incondensable gases and molecules small enough to escape at room temperature
315 or even lower, thus reducing bio-oil yield [42].

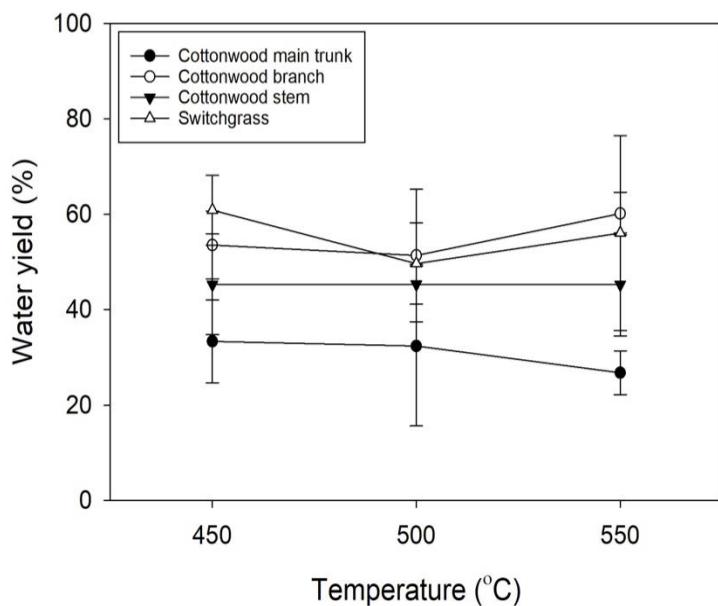
316 All biomasses showed an increase in gas yields as pyrolysis temperature increased from 450 °C
317 to 550 °C. Cottonwood stem displayed the highest percentage increase in gas yields of 13.33%

318 while cottonwood main stem (13.04%), switchgrass (11.43%), and cottonwood branch (7.41%)
 319 respectively trailed behind. This trend of increasing gas yields with temperature was expected as
 320 more gases are produced upon the more complete breakdown of biomasses at higher reaction
 321 temperatures as indicated by previous studies [30, 43-45]. The ideal operating conditions for
 322 highest overall energy efficiency are discussed in the energy balance section of this study.

323

324 **3.2. Water Content of the Liquid Fraction**

325 The water yield of the bio-oil was determined by performing Karl Fischer titrations on the liquid
 326 fractions as shown in Fig 4. Initial moisture content of the biomasses has been reported in Table
 327 2.



328

329 Fig. 4: Water content of the bio-oil as a fraction of the total liquid mass with respect to
 330 temperature for the different feedstock.

331 It was necessary to ascertain the water content in the bio-oil (both from the feedstock and
 332 pyrolysis process) due to its negative impact on the bio-oil's heating value [46]. For the

333 cottonwood main stem, the water yield reduced with temperature increase, as was the case with
334 cottonwood stem, although there was a minor spike at 500 °C. The range of water yields from the
335 various biomasses were 26.75% to 33.32% for cottonwood mainstem, 51.35% to 60.13% for
336 cottonwood branch, 45.3% on average for cottonwood stem, and 49.66% to 60.85% for
337 switchgrass. Thus, the water yields from switchgrass was notably higher than that from
338 cottonwood which agrees with the chemical compositions of both materials outlined in literature
339 [28, 47]. Lignin, which has been proven to be present in switchgrass in larger quantities
340 compared to switchgrass, produces water when dehydrated via pyrolysis reaction [48]. It was
341 also observed that cottonwood main trunk had the highest liquid yield with a corresponding low
342 water content compared to other poplar parts and switchgrass. Cottonwood main trunk harnesses
343 vascular tissues to transport water and nutrients from the soil to other parts for overall tree
344 growth. While pyrolysis temperature, heating rate, and particle size have been determined as key
345 factors influencing bio-oil yield [49, 50], this phenomenon in cotton wood main trunk is
346 indicative of its richness nutrients which are in relatively lower quantities in other tree parts such
347 as the stem or branch.

348 ***3.3. Characterization of char***

349 The elemental analyses of char yields were compared to the CHNO analysis of their respective
350 raw biomasses (Table 2). The values obtained for both raw biomass and the carbonized (char)
351 form are in the same range as those reported in literature for cottonwood and switchgrass
352 pyrolyzed in conventional reactors [34]. The minor differences can be attributed to the different
353 pyrolysis temperature (400 °C vs 450-600 °C in the present study).

354 Table 3: CHNO analysis of char yields from the induction pyrolysis of cottonwood main trunk,
355 branch, stem, and switchgrass compared to the CHNO analysis of their respective raw
356 biomasses.

Biomass	Temperature (°C)	Carbon (%)	Hydrogen (%)	Nitrogen (%)	Oxygen* (%)
Cottonwood main trunk	Raw biomass	48.5	6.0	0.7	43.0
	450	69.3 ± 4.65	3.0 ± 0.09	0.7 ± 0.30	25.3 ± 4.26
	500	74.7 ± 0.80	2.4 ± 0.24	0.4 ± 0.01	20.7 ± 0.57
	550	74.8 ± 0.25	2.7 ± 3.87	0.5 ± 0.68	20.3 ± 0.06
Cottonwood branch	Raw biomass	49.0	6.1	0.8	41.4
	450	70.5 ± 0.47	2.9 ± 0.15	0.8 ± 0.05	23.0 ± 0.37
	500	74.3 ± 1.36	2.5 ± 0.04	0.8 ± 0.13	19.7 ± 1.27
	550	75.8 ± 0.17	2.5 ± 0.13	0.8 ± 0.02	18.1 ± 0.28
Cottonwood stem	Raw biomass	48.5	5.9	0.6	43.2
	450	74.4 ± 0.88	3.1 ± 0.11	0.8 ± 0.01	19.8 ± 0.98
	500	77.0 ± 0.38	2.4 ± 0.00	0.6 ± 0.00	18.2 ± 0.38
	550	77.8 ± 0.24	2.3 ± 0.17	0.8 ± 0.06	17.2 ± 0.35
Switchgrass	Raw biomass	47.3	5.7	0.5	43.6
	450	65.2 ± 0.51	2.8 ± 0.52	1.0 ± 0.28	28.2 ± 0.29
	500	69.3 ± 1.30	2.4 ± 0.17	0.8 ± 0.06	24.7 ± 1.41
	550	66.8 ± 0.62	1.8 ± 0.02	0.9 ± 0.05	27.6 ± 0.69

* Calculated by difference on ash-free basis

The carbon content for cottonwood main stem reduced with increasing temperature from 32.22% at 450 °C to 31.58% at 550 °C. The unburned cottonwood branch sample has a carbon content of 48.95% almost twice (53.5%) that of the char when pyrolyzed at 550 °C – the highest reduction of all the biomasses. This significant reduction indicates that the oxygen content in the cottonwood branch biomass, which represents a major percentage of total unburned biomass CHN content, was significantly reduced by the pyrolysis process. Similarly, other biomasses exhibited a decrease, albeit lesser, in their carbon content from 450 to 550 °C with 31.8% and 33.8% for cottonwood main stem and stem products respectively. However, switchgrass showed a remarkable deviation – the pyrolysis process increased its carbon content from 47.29% unburned to 66.83% when processed to a 550 °C temperature, accounting for 41.3% increase in carbon content. This behavior connotes that the oxygen content of the switchgrass biomass, which represents most of the remaining mass in the CHN content of the unburned biomass, has depleted due to the pyrolysis process. Thus, its oxygen content reduced by 30.14% as against a

371 remarkable increase in oxygen content for the other biomasses. This increase in carbon content is
 372 a result of the increasingly condense carbon matrix caused by the pyrolysis process [29].

373 **3.4. Characterization of liquid fraction**

374 The bio-oil liquid fraction was dehydrated and characterized using CHN (Table 4) and GC-MS
 375 (Table 5) analyses. The liquid products from the GC-MS spectra were analyzed to quantify the
 376 peak areas of major compound groups namely: furans, ketones, alcohols, acids, phenols, and
 377 carbohydrates. The data revealed that bio-oils obtained from all biomasses yielded high
 378 concentration of phenolic compounds, which have earlier been reported. This is because
 379 phenolics naturally occur in trees which contain high lignin content. The phenolic yields are
 380 highest in the liquids derived from cottonwood main stem and stem; this can be explained by the
 381 presence of higher lignin amounts in tree stems than other parts [51]. Sadaka et al. [34] did not
 382 report the analysis of the liquid obtained from similar biomasses at 400 °C in a conventional
 383 reactor, so no comparison can be made with respect to bio-oil quality and composition.

384 Table 4: CHN analysis of the change in composition of the bio-oil fractions of cottonwood main
 385 trunk, branch, stem, and switchgrass as a function of temperature.

Biomass	Temperature (°C)	Carbon (%)	Hydrogen (%)	Nitrogen (%)	Oxygen* (%)
Cottonwood main trunk	450	32.2 ± 1.59	8.4 ± 0.63	0.2 ± 0.06	59.2 ± 1.02
	500	32.3 ± 1.97	8.7 ± 0.22	0.1 ± 0.04	58.9 ± 1.78
	550	33.1 ± 4.93	8.9 ± 1.03	0.1 ± 0.04	57.9 ± 3.95
Cottonwood secondary stems	450	29.1 ± 0.27	8.8 ± 0.48	0.1 ± 0.00	62.0 ± 0.74
	500	28.3 ± 0.69	8.3 ± 0.48	0.1 ± 0.01	63.3 ± 0.32
	550	31.7 ± 3.74	7.9 ± 0.72	0.2 ± 0.08	60.3 ± 3.91
Cottonwood branch	450	28.0 ± 1.81	9.5 ± 0.89	0.1 ± 0.01	62.3 ± 0.92
	500	22.2 ± 2.47	8.9 ± 0.07	0.1 ± 0.02	68.7 ± 2.42
	550	22.8 ± 1.87	9.4 ± 0.55	0.1 ± 0.01	67.6 ± 2.43
Switchgrass	450	28.4 ± 0.33	7.8 ± 1.13	0.9 ± 0.14	63.4 ± 0.93
	500	30.5 ± 9.28	7.9 ± 0.02	0.9 ± 0.10	60.7 ± 9.36
	550	27.6 ± 0.98	8.1 ± 0.20	1.0 ± 0.05	63.3 ± 0.96

386 * Calculated by difference

387 Table 5: Quantification of products from GC-MS analysis of the liquid fractions from
 388 cottonwood main stem (CWMS), branch (CWB), secondary stems (CWSS), and switchgrass
 389 (SG).

Biomass Temperature (°C)	CWMS			CWSS			CWB			SG		
	450	500	550	450	500	550	450	500	550	450	500	550
Furans	5.0	5.8	7.2	6.3	6.1	8.4	6.0	8.4	9.6	16.5	17.8	16.9
Ketones	5.6	9.9	2.9	6.9	8.8	3.5	8.1	4.6	8.5	5.6	5.6	3.8
Alcohols	0.0	0.0	0.0	0.0	0.0	0.0	0.0	3.8	0.0	3.9	0.0	0.0
Acids	0.0	9.5	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	1.5	0.0
Phenols	73.4	59.1	81.9	76.3	75.5	79.2	69.6	70.6	69.4	62.2	62.1	72.6
Others	16.0	15.7	8.0	10.6	9.6	8.9	16.3	12.6	12.5	11.7	13.1	6.7

390

391

392 **3.4. Energy balance**

393 Using the elemental analysis of each sample the specific energy content (Higher Heating Value –
 394 HHV and Lower Heating Value – LLV) was calculated for each feedstock and reaction product
 395 according to Scholze and Meyer equation above. For industry application the LHV is the
 396 relevant parameter, but for the purpose of this research the HHV is of interest and it is further
 397 discussed. The specific energy of the raw switchgrass is approximately 16.36 MJ/kg, whereas for
 398 the different parts of cottonwood is around 17.5 MJ/kg, with small variations between the
 399 different parts of the trees. The specific energy of the char (HHV) varies between 20.2 to 26.5
 400 MJ/kg with no significant influence of the process temperature on it. This values are similar with
 401 those obtained in other studies on similar biomass [34]. Thus, the only factor influencing the
 402 total energy delivered is the char yield as a reaction product, and not its composition. The
 403 specific energy of the oil ranges between 5.9 MJ/kg to 11.5 MJ/kg. In this case the process
 404 temperature has a strong influence on the specific energy content of the oil. For example the

405 Cottonwood branch oil has a minimum of 5.9 MJ/kg at 500°C while at 450°C the specific energy
 406 content is about 9.8 MJ/kg.

407 Table 6. Specific energy content of the initial biomass and reaction products

Parameter Sample	Temperature °C	HHV (dry basis) MJ/kg	LHV (dry basis) MJ/kg	Total Energy content kJ	Energy Ratio Products / Feedstock %
Cottonwood main stem		17.304	15.95	398.74	-
	450	23.203	22.525	159.93	40.11
	500	24.993	24.451	183.38	45.99
	550	25.532	24.922	144.55	36.25
Oil	450	12.333	10.436	91.84	23.03
	500	12.854	10.889	104.54	26.22
	550	13.593	11.583	103.09	25.85
Cottonwood secondary stems		17.124	15.792	394.80	-
	450	26.064	25.364	172.47	43.26
	500	26.222	25.68	156.65	39.29
	550	26.528	26.009	153.45	38.49
Oil	450	11.357	9.369	93.69	23.50
	500	10.13	8.256	78.43	19.67
	550	11.262	9.478	92.89	23.30
Cottonwood branches		17.906	16.529	413.22	-
	450	23.879	23.224	155.60	39.02
	500	25.182	24.618	162.48	40.75
	550	25.978	25.414	167.73	42.07
Oil	450	11.922	9.777	87.02	21.82
	500	7.941	5.931	51.60	12.94
	550	9.064	6.941	57.61	14.45
Switchgrass		16.358	15.07	376.76	-
	450	21.005	20.372	156.87	39.34
	500	22.445	21.903	157.70	39.55
	550	20.211	19.805	138.63	34.77
Oil	450	9.515	7.753	65.90	16.53
	500	10.766	8.982	74.55	18.70
	550	9.605	7.776	64.54	16.19

408
 409 The obtained energy (as reaction products) was referred to the input energy of the sample. As it
 410 may be noticed in Table 6 above, the main energy component of the reaction products is
 411 contained by the char (36 to 43% of the wood energy). The oil contains between 13 and 26% of
 412 the wood energy, while the rest is contained by the pyrolysis gas. The overall energy conversion
 413 efficiency (Table 7) varies between 61% - 80% depending both on feedstock and pyrolysis

414 temperature. The values in Table # indicate that maximum energetic value is obtained from
 415 cottonwood main stem processed at 500 °C.

416 Table 7. Energy conversion efficiency of the process

	Temperature °C	Energy of reaction products kJ	Energy conversion efficiency %
Cottonwood main stem	450	288.57	72.37
	500	319.52	80.13
	550	289.24	72.54
Cottonwood secondary stems	450	299.37	75.83
	500	273.08	69.17
	550	283.94	71.92
Cottonwood branches	450	280.62	67.91
	500	252.88	61.20
	550	266.14	64.41
Switchgrass	450	257.97	68.47
	500	270.66	71.84
	550	241.97	64.22

417

418 4. CONCLUSION

419 In this study, fast batch inductive-heating pyrolysis of cottonwood main stem, stem, branch, and
 420 switchgrass were conducted in a fixed-bed induction heating system. Kinetics experiments
 421 showed that three temperatures (450, 500, and 550 °C) could be used to perform proximate and
 422 ultimate analyses on all biomasses. It was discovered that char yield and carbon content
 423 decreased with increasing temperatures for all the biomasses except for switchgrass, in which the
 424 carbon content increased as the temperature increases. The highest bio-oil yield of 39.8% was
 425 achieved by cottonwood stem at 450 °C, while the maximum bio-oil yields for cottonwood main
 426 stem (38.4%), cottonwood branch (35.6%), and switchgrass (34%) were recorded at 500 °C, 450
 427 °C, and 450 °C respectively. Bio-oil outputs from cottonwood branch and switchgrass were
 428 significantly higher than cottonwood's stem parts because of the pyrolysis of lignin present in the
 429 latter which provide mechanical support, strength, and rigidity for the plant walls and therefore
 430 increased the bio-oil yield at a mid-range temperature of 500°C, enough to extract the liquid, but

431 not extreme to cause secondary reactions. Water content of the bio-oil yield from cottonwood
432 main stem was the lowest, for which there was a uniform reduction of water content from 450 ⁰C
433 to 550 ⁰C, as against the haphazard trend noticed for liquids from other biomasses. The GC-MS
434 showed that for all poplar biomasses, most of the bio-oils comprised of phenolic hydrocarbons
435 with minimal acid and alcohol content. The pronounced presence of phenols in the poplar
436 biomasses can be explained by the presence of lignin in those parts, as compared to switchgrass
437 with relatively lower lignin content. Rather, slightly higher content of furans was noticed in the
438 case of switchgrass which was non-existent in bio-oils from other biomasses.

439

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458 REFERENCES

- 459 1. Ronsse, F., et al., *Production and characterization of slow pyrolysis biochar: influence of*
460 *feedstock type and pyrolysis conditions.* GCB Bioenergy, 2013. **5**(2): p. 104-115.
- 461 2. Popp, J., et al., *The effect of bioenergy expansion: Food, energy, and environment.* Renewable
462 and Sustainable Energy Reviews, 2014. **32**(Supplement C): p. 559-578.
- 463 3. Müller-Hagedorn, M. and H. Bockhorn, *Pyrolytic behaviour of different biomasses (angiosperms)*
464 *(maize plants, straws, and wood) in low temperature pyrolysis.* Journal of Analytical and Applied
465 Pyrolysis, 2007. **79**(1/2): p. 136-146.
- 466 4. Verma, M., et al., *Biofuels Production from Biomass by Thermochemical Conversion*
467 *Technologies.* International Journal of Chemical Engineering, 2012. **2012**: p. 18.
- 468 5. Elshahed, M.S., *Microbiological aspects of biofuel production: Current status and future*
469 *directions.* Journal of Advanced Research, 2010. **1**(2): p. 103-111.
- 470 6. Drosig, B., et al., *Anaerobic digestion of stillage fractions – estimation of the potential for energy*
471 *recovery in bioethanol plants.* Water Science and Technology, 2012. **67**(3): p. 494-505.
- 472 7. Demirbas, A., *Effect of initial moisture content on the yields of oily products from pyrolysis of*
473 *biomass.* Journal of Analytical and Applied Pyrolysis, 2004. **71**(2): p. 803-815.
- 474 8. Fatih Demirbas, M., M. Balat, and H. Balat, *Biowastes-to-biofuels.* Energy Conversion and
475 Management, 2011. **52**(4): p. 1815-1828.
- 476 9. Muley, P.D., et al., *A critical comparison of pyrolysis of cellulose, lignin, and pine sawdust using*
477 *an induction heating reactor.* Energy Conversion and Management, 2016. **117**: p. 273-280.
- 478 10. Bridgwater, A.V., D. Meier, and D. Radlein, *An overview of fast pyrolysis of biomass.* Organic
479 *Geochemistry, 1999. **30**(12): p. 1479-1493.*
- 480 11. Meier, D. and O. Faix, *State of the art of applied fast pyrolysis of lignocellulosic materials — a*
481 *review.* Bioresource Technology, 1999. **68**(1): p. 71-77.
- 482 12. Lucia, O., et al., *Induction Heating Technology and Its Applications: Past Developments, Current*
483 *Technology, and Future Challenges.* IEEE Transactions on Industrial Electronics, 2014. **61**(5): p.
484 2509-2520.
- 485 13. Cain, K., *Trouble in Cottonwood Country, in The Cottonwood Tree: An American Champion.* 2007,
486 Johnson Books: Colorado. p. 256.
- 487 14. Gordon, J.C., *Poplars: Trees of the people, trees of the future.* The Forestry Chronicle, 2001. **77**:
488 p. 217-219.
- 489 15. Blanco-Canqui, H., *Energy Crops and Their Implications on Soil and Environment All rights*
490 *reserved. No part of this periodical may be reproduced or transmitted in any form or by any*
491 *means, electronic or mechanical, including photocopying, recording, or any information storage*
492 *and retrieval system, without permission in writing from the publisher.* Agronomy Journal, 2010.
493 **102**(2): p. 403-419.
- 494 16. Niu, L., et al., *Control of floral transition in the bioenergy crop switchgrass.* Plant, Cell &
495 Environment, 2016. **39**(10): p. 2158-2171.
- 496 17. Bouton, J., *Improvement of Switchgrass as a Bioenergy Crop, in Genetic Improvement of*
497 *Bioenergy Crops,* W. Vermerris, Editor. 2008, Springer New York: New York, NY. p. 309-345.
- 498 18. Sanderson, M.A., et al., *Switchgrass as a biofuels feedstock in the USA.* Canadian Journal of Plant
499 Science, 2006. **86**(Special Issue): p. 1315-1325.

500 19. Pimentel, D. and T.W. Patzek, *Ethanol Production Using Corn, Switchgrass, and Wood; Biodiesel*
501 *Production Using Soybean and Sunflower*. Natural Resources Research, 2005. **14**(1): p. 65-76.

502 20. English, B.C., et al., *Economic Impacts of Using Switchgrass as a Feedstock for Ethanol*
503 *Production: A Case Study Located in East Tennessee*. Economics Research International, 2013.
504 **2013**: p. 14.

505 21. Haque, M., et al., *Economic Evaluation of Switchgrass Feedstock Production Systems Tested in*
506 *Potassium-Deficient Soils*. BioEnergy Research, 2014. **7**(1): p. 260-267.

507 22. Xu, J., et al., *Sodium Hydroxide Pretreatment of Switchgrass for Ethanol Production*. Energy &
508 Fuels, 2010. **24**(3): p. 2113-2119.

509 23. Boateng, A.A., K.B. Hicks, and K.P. Vogel, *Pyrolysis of switchgrass (*Panicum virgatum*) harvested*
510 *at several stages of maturity*. Journal of Analytical and Applied Pyrolysis, 2006. **75**(2): p. 55-64.

511 24. Dipesh, K.C., et al., *Genotype influences survival and growth of eastern cottonwood (*Populus**
512 *deltoides L.*) managed as a bioenergy feedstock on retired agricultural sites of the Lower
513 *Mississippi Alluvial Valley*. New Forests, 2017. **48**(1): p. 95-114.

514 25. Blazier, M.A., et al., *Growth and Development of Switchgrass Managed for Biofuel Production in*
515 *the Mid-South USA: Influences of Variety, Cropping Systems, Fertilization Practices, and Site*
516 *Conditions*, in *Advances in Environmental Research*, J.A. Daniels, Editor. 2016, Nova Science
517 Publishers, Inc.: Hauppauge, NY. p. 101-127.

518 26. D4809, A.S., *Standard Test Method for Heat of Combustion of Liquid Hydrocarbon Fuels by Bomb*
519 *Calorimeter (Precision Method)*. 2013, ASTM International: West Conshohocken, PA, 2013.

520 27. Aguilar, G., et al., *Effects of biomass particle size on yield and composition of pyrolysis bio-oil*
521 *derived from Chinese tallow tree (*Triadica Sebifera L.*) and energy cane (*Saccharum complex*) in*
522 *an inductively heated reactor*. AIMS Energy, 2015. **3**(4): p. 838-850.

523 28. Henkel, C., et al., *Pyrolysis of energy cane bagasse and invasive Chinese tallow tree (*Triadica**

524 *sebifera L.*) biomass in an inductively heated reactor. Energy Conversion and Management, 2016. **109**: p. 175-183.

525 29. Borchardt, H.J. and F. Daniels, *Journal of the American Chemical Society*, 1957. **79**: p. 41-46.

526 30. Muley, P.D., et al., *Pyrolysis and Catalytic Upgrading of Pinewood Sawdust Using an Induction*
527 *Heating Reactor*. Energy & Fuels, 2015. **29**(11): p. 7375-7385.

528 31. Shafizadeh, F. and G.D. McGinnis, *Chemical composition and thermal analysis of cottonwood*.
529 *Carbohydrate Research*, 1971. **16**(2): p. 273-277.

530 32. Scholze, B. and D. Meier, *Characterization of the water-insoluble fraction from pyrolysis oil*
531 *(pyrolytic lignin). Part I. PY-GC/MS, FTIR, and functional groups*. Journal of Analytical and
532 *Applied Pyrolysis*, 2001. **60**(1): p. 41-54.

533 33. Jean Fidele NZIHOU, S.H., Medard Bouda, Jean Koulidiati, B. Gerard Segda, *Using dulong and*
534 *Vandalek formulas to estimate the calorific heating value of a household waste Model*. Int. J.
535 *Sci. Eng. Res.*, 2014. **5**(1): p. 1878-1883.

536 34. Sadaka, S., et al., *Pyrolysis and Combustion Kinetics of Raw and Carbonized Cottonwood and*
537 *Switchgrass Agroforests*. Bioresources, 2015. **10**(3): p. 4498-4518.

538 35. Kissinger, H.E., *Variation of Peak Temperature with Heating Rate in Differential Thermal*
539 *Analysis*. Journal of Research of the National Bureau of Standards, 1956. **57**(4): p. 217-221.

540 36. Stănciulescu, I., et al., *Reaction Constants and Activation Energies in Pyrolysis for *Populus Alba**.
541 *University Politehnica of Bucharest Scientific Bulletin-Series a-Applied Mathematics and Physics*.
542 **(under review)**.

543 37. Marculescu, C., et al., *Pyrolysis of Food Waste: Thermal Deconstruction Rates in a Regular*
544 *Calcination Oven vs. an Induction Heating Reactor*, in *2017 American Institute of Chemical*
545 *Engineers (AIChE) Annual Meeting*. 2017: Minneapolis, MN, USA.

547 38. Ogden, C.A., et al., *In-field direct combustion fuel property changes of switchgrass harvested*
548 *from summer to fall*. Fuel Processing Technology, 2010. **91**(3): p. 266-271.

549 39. Li, J., et al., *Prediction of high-temperature rapid combustion behaviour of woody biomass*
550 *particles*. Fuel, 2016. **165**: p. 205-214.

551 40. Park, H.J., et al., *Clean bio-oil production from fast pyrolysis of sewage sludge: Effects of reaction*
552 *conditions and metal oxide catalysts*. Bioresource Technology, 2010. **101**(1, Supplement): p. S83-
553 S85.

554 41. Hanif, M.U., et al., *Effects of Pyrolysis Temperature on Product Yields and Energy Recovery from*
555 *Co-Feeding of Cotton Gin Trash, Cow Manure, and Microalgae: A Simulation Study*. PLoS One,
556 2016. **11**(4): p. e0152230.

557 42. Qu, T., et al., *Experimental Study of Biomass Pyrolysis Based on Three Major Components:*
558 *Hemicellulose, Cellulose, and Lignin*. Industrial & Engineering Chemistry Research, 2011. **50**(18):
559 p. 10424-10433.

560 43. Açıkalın, K., F. Karaca, and E. Bolat, *Pyrolysis of pistachio shell: Effects of pyrolysis conditions and*
561 *analysis of products*. Fuel, 2012. **95**: p. 169-177.

562 44. Chhiti, Y., et al., *Wood Bio-Oil Noncatalytic Gasification: Influence of Temperature, Dilution by an*
563 *Alcohol and Ash Content*. Energy & Fuels, 2011. **25**(1): p. 345-351.

564 45. McKendry, P., *Energy production from biomass (Part 3): Gasification technologies*. Bioresour
565 Technol, 2002. **83**(1): p. 55-63.

566 46. Chen, D., J. Zhou, and Q. Zhang, *Effects of heating rate on slow pyrolysis behavior, kinetic*
567 *parameters and products properties of moso bamboo*. Bioresour Technol, 2014. **169**: p. 313-319.

568 47. Hu, Z., et al., *Chemical profiles of switchgrass*. Bioresource Technology, 2010. **101**(9): p. 3253-
569 3257.

570 48. Klein, M.T. and P.S. Virk, *Modeling of Lignin Thermolysis*. Energy & Fuels, 2008. **22**(4): p. 2175-
571 2182.

572 49. Gao, Y., et al., *Factors affecting the yield of bio-oil from the pyrolysis of coconut shell*.
573 SpringerPlus, 2016. **5**: p. 333.

574 50. Duarte, S.J., et al., *Effect of temperature and particle size on the yield of bio-oil, produced from*
575 *conventional coconut core pyrolysis*. International Journal of Chemical Engineering and
576 Applications, 2016. **7**(2): p. 102-108.

577 51. Wang, G., et al., *TG study on pyrolysis of biomass and its three components under syngas*. Fuel,
578 2008. **87**(4): p. 552-558.

579

580

581