

Heavy-Atom Substituted Nucleobases in Photodynamic Applications: Substitution of Sulfur with Selenium in 6-Thioguanine Induces a Remarkable Increase in the Rate of Triplet Decay in 6-Selenoguanine

Kieran M. Farrell,[†] Matthew M. Brister,[†] Michael Pittelkow,[‡] Theis I. Sølling,[‡] Carlos E. Crespo-Hernández^{†,*}

[†] Department of Chemistry, Case Western Reserve University, 10900 Euclid Ave., Cleveland, Ohio 44106

[‡] University of Copenhagen, Department of Chemistry, Universitetsparken 5, DK-2100 Copenhagen, Denmark

Supporting Information Placeholder

ABSTRACT: Sulfur substitution of carbonyl oxygen atoms of DNA/RNA nucleobases promotes ultrafast intersystem crossing and near-unity triplet yields that are being used for photodynamic therapy and structural-biology applications. Replacement of sulfur with selenium or tellurium should significantly redshift the absorption spectra of the nucleobases without sacrificing the high triplet yields. Consequently, selenium/tellurium-substituted nucleobases are thought to facilitate treatment of deeper tissue carcinomas relative to the sulfur-substituted analogues, but their photodynamics are yet unexplored. In this contribution, the photochemical relaxation mechanism of 6-selenoguanine is elucidated and compared to that of the 6-thioguanine prodrug. Selenium substitution leads to a remarkable enhancement of the intersystem crossing lifetime both to and from the triplet manifold, resulting in an efficiently populated, yet short-lived triplet state. Surprisingly, the rate of triplet decay in 6-selenoguanine increases by 835-fold compared to that in the 6-thioguanine prodrug. This appears to be an extreme manifestation of the classical heavy-atom effect in organic photochemistry, which challenges conventional wisdom.

In addition to optimizing DNA for its role as the genetic blueprint of life, evolution has tailored the structure of DNA nucleotides such that they efficiently dissipate energy absorbed from ultraviolet (UV) radiation.^{1,2} Numerous theoretical and experimental investigations have demonstrated that the naturally occurring nucleobases, nucleosides, and nucleotides recover their electronic ground states within ~ 1 ps of excitation.^{1–4} Ultrafast ground state recovery has made nucleic acids robust against the solar onslaught of one of the most ubiquitous carcinogens—UV radiation.^{5,6} Conversely, sulfur substitution of carbonyl oxygen atoms on nucleobases leads to the efficient population of long-lived, reactive triplet excited states in high yields.^{7–11} As a result, the photochemistry of sulfur-substituted DNA/RNA nucleobases (a.k.a., thiobases) have drawn increased attention because of their use in photodynamic therapy and photocrosslinking applications.^{9–19}

The high triplet yield makes the thiobases prime sensitizers for targeting DNA/RNA damage,¹⁰ however, their absorption properties limit the applications to the treatment of skin cancer cells.¹² Replacement of the sulfur atom with selenium or tellurium should

redshift the absorption spectra into the visible with the potential additional benefit of maximizing their triplet yields. Recent quantum-chemical calculations predict that selenium and tellurium substituted nucleobases are highly promising photosensitizers.^{20,21} By using femtosecond broadband transient absorption spectroscopy, we demonstrate that selenium substitution of guanine leads to a remarkable enhancement of the intersystem crossing (ISC) lifetime both to and from the triplet manifold. In particular, the rate of ISC to the ground state in 6-selenoguanine increases by 835-fold, or more than two orders of magnitude, compared to that of the 6-thioguanine prodrug. A strategy for harvesting the use of 6SeGua for photodynamic therapy and structural-biology applications is presented, which might enable heavy-atom substituted nucleobases to reach their full potential.

Figure 1 shows the absorption spectra for 6SeGua and 6tGua normalized to their lowest energy $\pi \rightarrow \pi^*$ transition. 6SeGua exhibits maxima at 357 and 209 nm, whereas 6tGua has maxima at 341, 254, and 204 nm. Selenium substitution broadens and red shifts the UVA transition by 16 nm (1314.3 cm^{-1}) relative to sulfur substitution. The combined shifting and broadening of 6SeGua's absorption give rise to a tail absorbing in the visible region. The oscillator strength of the UVA vertical transition for 6SeGua, shown in Figure 1, has very similar magnitude than that for 6tGua,²² suggesting that the molar absorptivity in the UVA region is similar for both molecules. We estimate that using 6SeGua as photosensitizer could facilitate up to 90% deeper tissue treatment by UVA radiation than 6tGua (see Figure S1 and the Supporting Information (SI) for details).

Quantum-chemical computations have shown that the N⁹ tautomer of 6SeGua is most stable in solution.^{23–26} We have performed ground state optimizations and subsequent vertical excitation energy calculations for the N⁹ tautomer of 6SeGua to characterize the excited states available to UVA excitation. A complete summary of computational results is given in Tables S1 to S3 and Figure S2. UVA excitation of 6SeGua in aqueous solution excites the S₂($\pi\pi^*$) state. Four lower-lying states are also available for electronic relaxation: namely, a dark S₁($n\pi^*$) state and three triplet states. Both the S₂($\pi\pi^*$) and T₃($\pi\pi^*$) states, as well as the S₁($n\pi^*$) and T₂($n\pi^*$) states, are isoenergetic within computational uncertainty. The S₁($n\pi^*$) and T₁($\pi\pi^*$) states are separated by an energy gap of 0.64 eV.

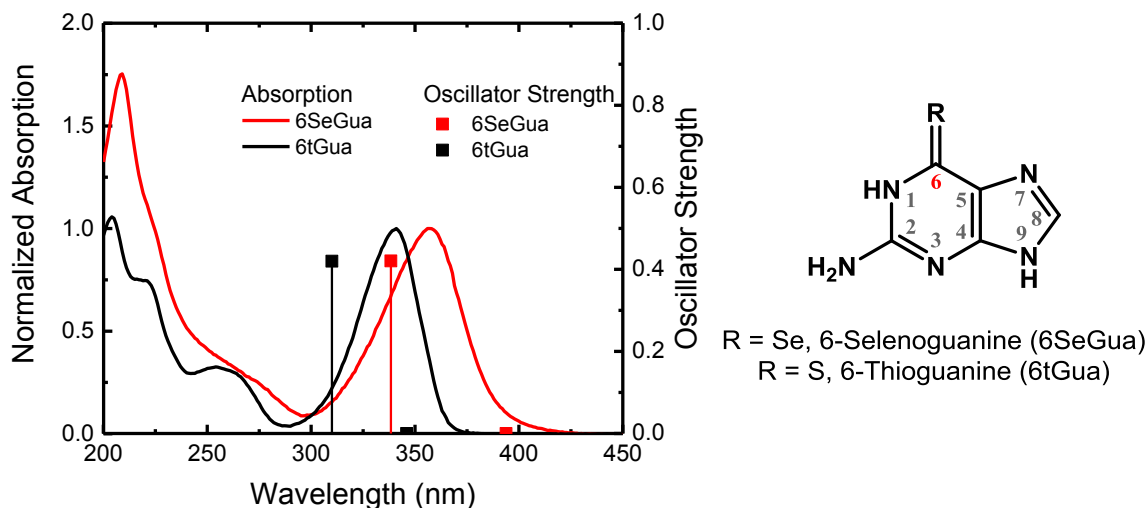


Figure 1. Ground state absorption spectra of 6SeGua and 6tGua normalized to their lowest-energy $\pi \rightarrow \pi^*$ transitions. All spectra were obtained in pH 7.4 phosphate buffer saline (PBS) at room temperature. Oscillator strengths obtained from TDDFT computations at the PBE0/IEF-PCM/6-311++G(d,p) level of theory are included (see SI for details).

Femtosecond broadband transient absorption experiments were performed to unveil the electronic decay pathways following UVA excitation. The dynamics of selenium substitution at the C⁶ position were compared to sulfur substitution by performing successive experiments on both 6SeGua and 6tGua. Figure 2 shows the transient absorption spectra (TAS) of 6SeGua and 6tGua following 352 nm excitation. For 6SeGua, two bands appear concurrently on a sub-picosecond time-scale: one with a maximum *ca.* 490 nm and another with a tail apparent at 675 nm. A negative amplitude signal is observed at short wavelengths immediately after excitation. The signal overlaps with the ground state absorption spectrum in Figure 1, and is assigned to ground state depopulation. From a delay time of 0.6 to 104 ps, the 490 nm band blue shifts 30 nm. Simultaneous to this shift, the ground state bleach increases in amplitude (Figure 2b, inset). Following the blue shift, all spectral features decay uniformly through 3 ns.

The TAS for 6tGua (Figure 2d-f) are similar to those of 6SeGua and consistent with those reported by Ashwood et al.²² A UV band *ca.* 375 nm, a visible band *ca.* 500 nm, and ground state depopulation signal are observed on a sub-picosecond timescale. The UV band increases in amplitude from a time delay of 0.77 to 68 ps, while the red edge of the visible band narrows. Like 6SeGua, a slight increase in negative amplitude signal occurs during this process. No apparent spectral changes occur from 68 ps through 3 ns.

Kinetics traces corresponding to the TAS in Figure 2 were analyzed globally. Figure 3 shows representative traces with fits obtained from three- and two-lifetime sum of exponential functions for 6SeGua and 6tGua, respectively. Lifetimes obtained from these fits are reported in Table 1. For both 6SeGua and 6tGua, τ_1 reflects ultrafast ISC responsible for growth of the $T_1(\pi\pi^*)$ absorption bands, τ_2 corresponds to spectral evolution shown in Figure 2b, and τ_3 corresponds to the uniform triplet decay.

The TAS for 6SeGua and 6tGua are qualitatively similar through the second lifetime. Concurrent to ground state depopulation, the 490 and 675 nm bands rise during τ_1 . We assign the rise of these two bands to $T_1(\pi\pi^*)$ absorption. This assignment is supported by the fact that 6tGua undergoes an ultrafast ISC event following UVA excitation, and similarly shows $T_1(\pi\pi^*)$ absorption with maxima at 375 and 500 nm.²² Heavy atom substitution enhances ISC dynamics in 6tGua relative to guanine,²² and selenium substitution should further enhance ISC relative to 6tGua. Indeed, the spin orbit coupling between the $S_1(n\pi^*)$ and $T_1(\pi\pi^*)$ states increases nearly five-fold for 6-selenoguanosine compared to 6-thioguanosine.²¹

6SeGua's ISC lifetime (τ_1), which is three times faster than that in 6tGua, may increase the triplet yield (assuming that the lifetime of any other competitive relaxation pathway is similar in both molecules), arguably rendering 6SeGua a more effective photosensitizer when incorporated in DNA/RNA.

Table 1. Global lifetimes for 6SeGua and 6tGua following excitation at 352 nm under N₂-saturated conditions

Lifetimes	τ_1 / ps	τ_2 / ps	τ_3 / ns
6SeGua	0.13 ± 0.05	31 ± 2	1.7 ± 0.1^a
6tGua	0.35 ± 0.04^b	18 ± 1	1420 ± 180^c

^a Estimated by extrapolating fit past 1 ns; ^b Same lifetime value was obtained within experimental uncertainties exciting at 342 nm; ^c Reproduced from ref. 22, where an excitation wavelength of 345 nm was used.

El-Sayed's propensity rules suggest that ultrafast ISC should occur between singlet and triplet states of different character.^{27,28} Pathways that obey this rule for 6SeGua include $S_2(\pi\pi^*) \rightarrow T_2(n\pi^*)$ and $S_1(n\pi^*) \rightarrow T_1(\pi\pi^*)$ (Figure S2). Calculations for 6-selenoguanosine and 6-thioguanosine predict that ISC predominately occurs via the $S_1(n\pi^*) \rightarrow T_1(\pi\pi^*)$ pathway.²¹ However, non-adiabatic dynamics simulations indicate that ISC in 6tGua predominately occurs from the $S_1(n\pi^*)$ minimum to the $T_2(n\pi^*)$ state.²⁹ Given the similarities between the theoretical and experimental results for 6SeGua and 6tGua herein, the $S_1(n\pi^*) \rightarrow T_2(n\pi^*)$ ISC pathway cannot be excluded. Collectively, the theoretical results suggest that after excitation of 6SeGua to the $S_2(\pi\pi^*)$ state, electron population internally converts to the $S_1(n\pi^*)$ state before ISC to $T_2(n\pi^*)$ and/or $T_1(\pi\pi^*)$ state occurs: $S_2 \rightarrow S_1 \rightarrow T_2 \rightarrow T_1$.

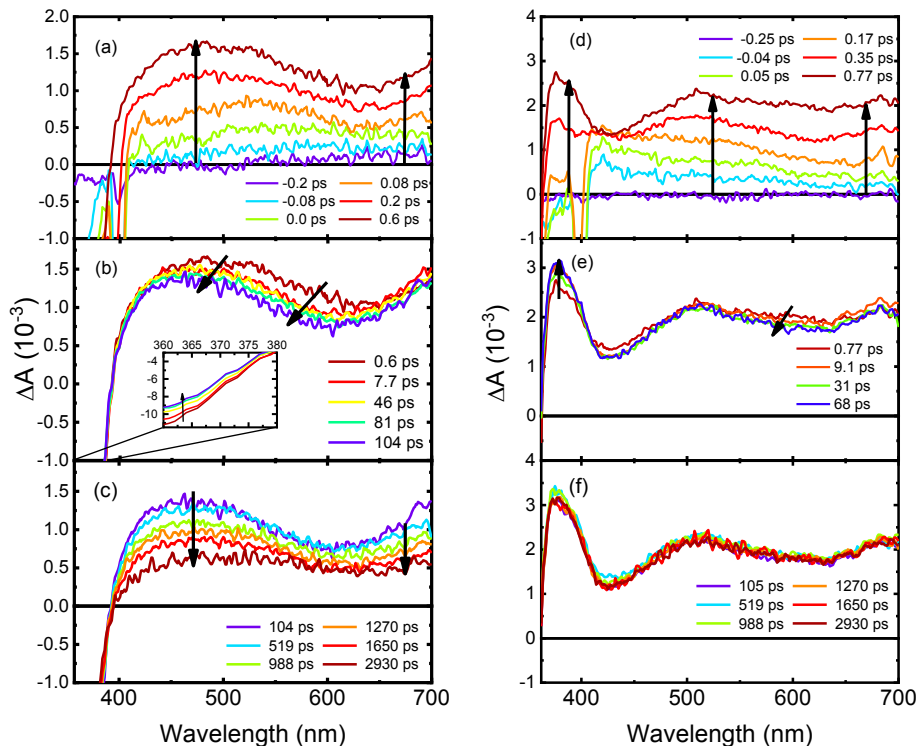


Figure 2. (a-c) TAS for 6SeGua following 352 nm excitation through a time delay of 3 ns. (2b, Inset) Ground state recovery concurrent with blue shift of 490 nm band. (d-f) TAS for 6tGua obtained back-to-back with 6SeGua. The stimulated Raman emission band of water is observed around 400 nm within the cross correlation of the pump and probe beams. TAS were obtained in pH 7.4 PBS at room temperature under N_2 -purged conditions (See SI for details).

The spectral dynamics that occur during τ_2 are also similar for both 6SeGua and 6tGua. The blue shift of the 495 nm band in 6SeGua to 460 nm and the narrowing of the visible band in 6tGua both suggest vibrational relaxation during τ_2 . Thus, the blue shift is assigned to vibrational cooling of the hot $T_1(\pi\pi^*)$ state following population *via* ultrafast ISC (see Figures S2 and S3). This assignment is supported by the observation that the shifting process is completed prior to the uniform decay of both triplet bands, and excess vibrational energy is dissipated prior to the monotonic decay of the $T_1(\pi\pi^*)$ state.^{30,31}

The negative amplitude signal at ca. 370 nm increases simultaneously to vibrational cooling of the $T_1(\pi\pi^*)$ state. This phenomenon is also observed for 6tGua,²² for which it was unclear whether the increase was due to ground state recovery or overlap with a

growing UV band (Figure 2e). For 6SeGua, this event corresponds to ground state recovery, as the absorption change at 360 nm (ca. 2 mAU) is an order of magnitude larger than the decline of the 495 nm band (ca. 0.2 mAU). This event is therefore concurrent, yet unrelated to the vibrational cooling dynamics observed in the $T_1(\pi\pi^*)$ state. Ground state recovery during τ_2 may be achieved via internal conversion of a residual $S_1(n\pi^*)$ state population to the ground state. Triplet self-quenching^{11,32,33} is unlikely because preliminary experiments in which the relative absorbance of the 6SeGua solution or the excitation intensity were varied by up to 68 and 60%, respectively, do not support a triplet quenching pathway.

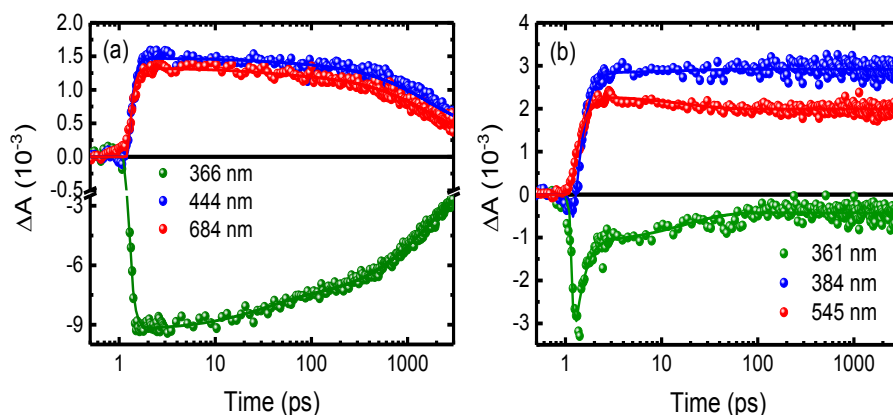


Figure 3. Kinetics decay traces for (a) 6SeGua and (b) 6tGua, featuring global fits yielded by a sum of exponentials model.

The most remarkable difference between 6SeGua and 6tGua is manifested in τ_3 . Kinetic traces for 6tGua reach an offset following τ_2 , indicating that further relaxation events occur on a nanosecond timescale. Previous work has shown that triplet decay occurs over 1420 ns under N_2 -saturated conditions.²² Due to its long-lived triplet state, photoexcited 6tGua sensitizes singlet oxygen with a 21% yield.²² 6SeGua instead exhibits immediate, uniform decay of both triplet bands following τ_2 . Simultaneous recovery of ground state bleaching signal confirms that this event represents triplet decay with a lifetime of 1.7 ns (Figure 3). In addition to the stronger T_1/S_0 spin-orbit coupling in 6SeGua than in 6tGua, we argue that there is a smaller energy barrier to access the region of the T_1 potential energy surface where the ISC event occurs, explaining the 835-fold faster triplet decay in 6SeGua.³⁴ Given the sub-2 ns triplet decay lifetime, the yield of singlet oxygen generation for 6SeGua should be smaller than for 6tGua because diffusional encounter to sensitize molecular oxygen becomes less competitive as the rate of triplet decay increases.

The incorporation of 6SeGua into DNA and RNA has been demonstrated,^{35–37} and molecular dynamics simulations have shown that 6SeGua-substituted DNA duplexes are stable.^{38,39} Hence, regardless of the sub-2 ns triplet decay lifetime, the incorporation of 6SeGua into DNA/RNA duplexes should increase the probability that a reaction between the triplet state of 6SeGua and an adjacent nucleobase may occur. The close contact of 6SeGua to an adjacent π -stacked canonical nucleobase in a DNA/RNA duplex relaxes the requirement of diffusional encounter for a reaction to occur, enabling 6SeGua to photosensitize damage to DNA before triplet state decay. 6SeGua is a known chemotherapeutic agent.⁴⁰ The results presented in this study further show that it is prudent to investigate the photosensitization mechanism of 6SeGua-substituted DNA/RNA duplexes prior to studying tellurium-substituted nucleobases. The substitution with tellurium atoms is expected to further increase the rate of triplet decay to the ground state, which may further decrease the probability of a photosensitized reaction with an adjacent nucleobase compared to a selenium-substituted nucleobase.

ASSOCIATED CONTENT

Supporting Information

Materials and computational and experimental methods; data analysis and supporting results; single-electron transitions and character composition of computed excited states; Kohn-Sham orbitals corresponding to vertical electronic transitions; Jablonski diagrams; plot of depth of tissue penetration; decay associated spectra

obtained from sum of exponential model; procedure for synthesis of 6-selenoguanine; NMR spectra.

AUTHOR INFORMATION

Corresponding Author

* carlos.crespo@case.edu

ACKNOWLEDGMENT

The authors acknowledge funding from the National Science Foundation (Grant No. CHE-1800052). The Faculty Early Career Development Program from NSF (Grants No. CHE-1255084 and CHE-1539808) is also acknowledge for initial support of this work. The authors also thank Mr. Sean Hoehn and Mr. Luis Rodríguez-Ortiz for performing preliminary TAS experiments at different excitation intensities and concentrations for 6SeGua.

REFERENCES

- (1) Crespo-Hernández, C. E.; Cohen, B.; Hare, P. M.; Kohler, B. Ultrafast Excited-State Dynamics in Nucleic Acids. *Chem. Rev.* **2004**, *104*, 2077–2019.
- (2) Middleton, C. T.; de La Harpe, K.; Su, C.; Law, Y. K.; Crespo-Hernández, C. E.; Kohler, B. DNA Excited-State Dynamics: From Single Bases to the Double Helix. *Annu. Rev. Phys. Chem.* **2009**, *60*, 217–239.
- (3) Improta, R.; Santoro, F.; Organo, C.; Blancafort, L. Quantum Mechanical Studies on the Photophysics and the Photochemistry of Nucleic Acids and Nucleobases. *Chem. Rev.* **2016**, *116*, 3540–3593.
- (4) Beckstead, A. A.; Zhang, Y.; de Vries, S.; Kohler, B. Life in the Light: Nucleic Acid Photoproperties as a Legacy of Chemical Evolution. *Phys. Chem. Chem. Phys.* **2016**, *18*, 24228–24238.
- (5) The International Agency for Research on Cancer, in *Monographs on the Evaluation of Carcinogenic Risks to Humans*; **2012**; *110D*, 35–101.
- (6) World Health Organization. (2018). *The Known Health Effects of UV*. [online] Available at: <http://www.who.int/uv/fac/uvhealthfac/en/> [Accessed 15 Jul. 2018].
- (7) Reichardt, C.; Guo, C.; Crespo-Hernández, C. E. Excited-State Dynamics in 6-Thioguanosine from the Femtosecond to Microsecond Time Scale. *J. Phys. Chem. B.* **2011**, *115*, 3263–3270.
- (8) Taras-Goślińska, K.; Burdziński, G.; Wenska, G. Relaxation of the T_1 Excited State of 2-Thiothymine, its Riboside and Deoxyribose-Enhanced Nonradiative Decay Rate Induced by Sugar Substituent. *J. Photochem. Photobiol. A* **2014**, *275*, 89–95.
- (9) Pollum, M.; Jockusch, S.; Crespo-Hernández, C. E. 2,4-Dithiothymidine as a Potent UVA Chemotherapeutic Agent. *J. Am. Chem. Soc.* **2014**, *136*, 17930–17933.
- (10) Ashwood, B.; Pollum, M.; Crespo-Hernández, C. E. Photochemical and Photodynamical Properties of Sulfur-Substituted Nucleic Acid Bases. *Photochem. Photobiol.* **2018**, DOI: 10.1111/php.12975.

- (11) Pollum, M.; Martínez-Fernández, L.; Crespo-Hernández, C. In *Photoinduced Phenomena in Nucleic Acids I, Topics in Current Chemistry*; Springer, Cham, **2015**, vol. 355, pp 245–327.
- (12) Pollum, M.; Lam, M.; Jockusch, S.; Crespo-Hernández, C. E. Dithionated Nucleobases as Effective Photodynamic Agents against Human Epidermoid Carcinoma Cells. *Chem. Med. Chem.* **2018**, *13*, 1044–1050.
- (13) Pollum, M.; Jockusch, S.; Crespo-Hernández, C. E. Increase in Photoreactivity of Uracil Derivatives by Doubling Thionation. *Phys. Chem. Chem. Phys.* **2015**, *17*, 27851–27861.
- (14) Reelfs, O.; Karran, P.; Young, A. R. 4-Thiothymidine Sensitization of DNA to UVA Offers Potential for a Novel Photochemotherapy. *Photochem. Photobiol. Sci.* **2012**, *11*, 148–154.
- (15) Pridgeon, S. W.; Heer, R.; Taylor, G. A.; Newell, D. R.; O'Toole, K.; Robinson, M.; Xu, Y.-Z.; Karran, P.; Boddy, A. V. Thiothymidine Combined with UVA as a Potential Novel Therapy for Bladder Cancer. *Brit. J. Cancer* **2011**, *104*, 1869–1876.
- (16) Brem, R.; Daehn, I.; Karran, P. Efficient DNA Interstrand Crosslinking by 6-Thioguanine and UVA Radiation. *DNA Repair* **2011**, *10*, 869–876.
- (17) Favre, A.; Saintomé, C.; Fourrey, J.; Clivio, P.; Laugãa, P. New Trends in Photobiology Thionucleobases as Intrinsic Photoaffinity Probes of Nucleic Acid Structure and Nucleic Acid-Protein Interactions. *J. Photochem. Photobiol. B* **1998**, *42*, 109–124.
- (18) Favre, A. In *Bioorganic Photochemistry, Volume 1: Photochemistry and the Nucleic Acids*; 1990; pp 379–425.
- (19) Gemenetzidis, E.; Shavorskaya, O.; Xu, Y.-Z.; Trigiante, G. Topical 4-Thiothymidine is a Viable Photosensitizer for the Photodynamic Therapy of Skin Malignancies. *J. Dermatol. Treat.* **2013**, *24*, 209–214.
- (20) Pirillo, J.; de Simon, B. C.; Russo, N. Photophysical Properties Prediction of Selenium and Tellurium Substituted Thymidine as Potential UVA Chemotherapeutic Agents. *Theor. Chem. Acc* **2016**, *135*, 1–5.
- (21) Pirillo, J.; Mazzone, G.; Russo, N.; Bertini, L. Photophysical Properties of S, Se and Te-Substituted Deoxyguanosines: Insight into Their Ability to Act as Chemotherapeutic Agents. *J. Chem. Inf. Mod.* **2017**, *57*, 234–242.
- (22) Ashwood, B.; Jockusch, S.; Crespo-Hernández, C. E. Excited-State Dynamics of the Thiopurine Prodrug 6-Thioguanine: Can N9-Glycosylation Affect Its Phototoxic Activity? *Molecules* **2017**, *22*, 379.
- (23) Venkateswarlu, D.; Leszczynski, J. Tautomerism and Proton Transfer in 6-Selenoguanine: A Post Hartree-Fock Level ab Initio SCF-MO Investigation. *J. Phys. Chem. A* **1998**, *102*, 6161–6166.
- (24) Leszczynski, J. Guanine, 6-Thioguanine and 6-Selenoguanine: Ab Initio HF/DZP and MP2/DZP Comparative Studies. *J. Mol. Struct.* **1994**, *311*, 37–44.
- (25) Karthika, L.; Senthikumar, R.; Kanakaraju, M. Hydrogen-Bond Interactions in Hydrated 6-Selenoguanine Tautomers: A Theoretical Study. *Struct. Chem.* **2014**, *25*, 197–213.
- (26) Kim, Y.; Jang, Y. H.; Cho, H.; Hwang, S. Tautomers and Acid Dissociation Constants of 6-Selenoguanine from Density Functional Theoretical Calculations. *Bull. Korean Chem. Soc.* **2010**, *31*, 3013–3016.
- (27) El-Sayed, M. A. Spin—Orbit Coupling and the Radiationless Processes in Nitrogen Heterocyclics. *J. Chem. Phys.* **1963**, *38*, 2834–2838.
- (28) El-Sayed, M. A. The Radiationless Processes Involving Change of Multiplicity in the Diazenes. *J. Chem. Phys.* **1962**, *573*, 573–574.
- (29) Martínez-Fernández, L.; Corral, I.; Granucci, G.; Persico, M. Competing Ultrafast Intersystem Crossing and Internal Conversion: A Time Resolved Picture for the Deactivation of 6-Thioguanine. *Chem. Sci.* **2014**, *5*, 1336–1347.
- (30) Kasha, M. Characterization of Electronic Transitions in Complex Molecules. *Discuss. Faraday Soc.* **1950**, *9*, 14–19.
- (31) Kasha, M.; McGlynn, S. P. Molecular Electronic Spectroscopy. *Annu. Rev. Phys. Chem.* **1956**, *7*, 403–424.
- (32) Reichardt, C.; Crespo-Hernández, C. E. Ultrafast Spin Crossover in 4-Thiothymidine in an Ionic Liquid. *Chem. Commun.* **2010**, *46*, 5963–5965.
- (33) Reichardt, C.; Crespo-Hernández, C. E. Room-Temperature Phosphorescence of the DNA Monomer Analogue 4-Thiothymidine in Aqueous Solutions after UVA Excitation. *J. Phys. Chem. Lett.* **2010**, *1*, 2239–2243.
- (34) It is possible that the presence of a deprotonated anionic species of 6SeGua in equilibrium with the neutral form²⁶ at pH 7.4 may contribute to the observed dynamics. Additional experiments in which the pH of the solution is systematically varied are necessary in order to examine the putative extent of participation of deprotonated species in the dynamics under physiological conditions. Such an investigation is currently underway in our group, but outside of the scope of the present communication.
- (35) Salon, J.; Gan, J.; Abdur, R.; Liu, H.; Huang, Z. Synthesis of 6-Se-Guanosine RNAs for Structural Study. *Org. Lett.* **2013**, *15*, 3934–3937.
- (36) Salon, J.; Jiang, J.; Sheng, J.; Gerlits, O. O.; Huang, Z. Derivatization of DNAs with Selenium at 6-position of Guanine for Function and Crystal Structure Studies. *Nucleic Acids Res.* **2008**, *36*, 7009–7018.
- (37) Kuar, M.; Rob, A.; Caton-Williams, J.; Huang, Z. In *Biochalcogen Chemistry: The Biological Chemistry of Sulfur, Selenium, and Tellurium*; 2013; pp 89–126.
- (38) Salon, J.; Sheng, J.; Jiang, J.; Chen, G.; Caton-Williams, J.; Huang, Z. Oxygen Replacement with Selenium at the Thymidine 4-Position for the Se Base Pairing and Crystal Structure Studies. *J. Am. Chem. Soc.* **2007**, *129*, 4862–4863.
- (39) Faustino, I.; Curutchet, C.; Luque, F. J.; Orozco, M. The DNA-Forming Properties of 6-Selenoguanine. *Phys. Chem. Chem. Phys.* **2014**, *16*, 1101–1110.
- (40) Ross, A. F.; Agarwal, K. C.; Chu, S. H.; Parks, R. E. Studies on the Biochemical Actions of 6-Selenoguanine and 6-Selenoguanosine. *Biochem. Pharmacol.* **1973**, *22*, 141–154.

