

Anti-fatigue-fracture hydrogels by designing crystalline domains

Shaoting Lin,^{1*} Xinyue Liu,^{1*} Ji Liu,^{1*} Hyunwoo Yuk,¹ Hyun-Chae Loh,² German A. Parada,^{1,3}

Charles Settens,⁴ Jake Song,⁵ Admir Masic,² Gareth H. McKinley,¹ Xuanhe Zhao^{1,2,†}

¹Department of Mechanical Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

²Department of Civil and Environmental Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

³Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, USA;

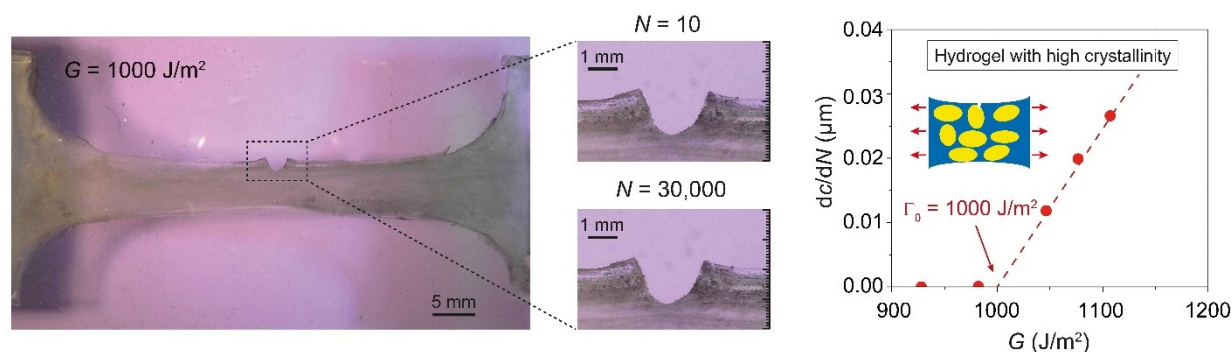
⁴Center for Materials Science and Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, USA;

⁵Department of Materials Science and Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139, USA;

Shaoting Lin, Xinyue Liu and Ji Liu contributed equally to this work.

†To whom correspondence should be addressed. Email: zhaox@mit.edu

One sentence summary: Designing nano-crystalline domains gives extremely anti-fatigue-fracture hydrogels for artificial cartilages and soft robots.



21 **Abstract**

22
23 The emerging applications of hydrogels in devices and machines require these soft materials to
24 maintain robustness under cyclic mechanical loads. Whereas hydrogels have been made tough to
25 resist fracture under a single cycle of mechanical load, these toughened gels still suffer from fatigue
26 fracture under multiple cycles of loads. The reported fatigue threshold (i.e., the minimal fracture
27 energy at which crack propagation occurs under cyclic loads) for synthetic hydrogels is on the order
28 of 1–100 J/m², which is primarily associated with the energy required to fracture a single layer of
29 polymer chains per unit area. Here, we demonstrate that the controlled introduction of crystallinity
30 in hydrogels can significantly enhance their fatigue thresholds, since the process of fracturing
31 crystalline domains for fatigue-crack propagation requires much higher energy than fracturing a
32 single layer of polymer chains. The fatigue threshold of polyvinyl alcohol (PVA) with a crystallinity
33 of 18.9 wt.% in the swollen state can exceed 1,000 J/m². We further develop a strategy to enhance
34 the anti-fatigue-fracture properties of PVA hydrogels, but still maintain their high water contents
35 and low moduli by patterning highly-crystalline regions in the hydrogels. The current work not only
36 reveals an anti-fatigue-fracture mechanism in hydrogels but also provides a practical method to
37 design anti-fatigue-fracture hydrogels for diverse applications.

38

39

40

41 **Introduction**

42
43 As polymer networks infiltrated with water, hydrogels have been widely used as scaffolds for tissue
44 engineering (1), vehicles for drug delivery (2) and model platforms for biological studies (3). More
45 recently, hydrogels have been explored for applications in devices and machines, including
46 wearable electronics (4, 5), soft robotics (6), adhesives (7-9), and hydrogel-based soft machines
47 (10, 11). The use of hydrogels in devices and machines requires them to maintain robustness under
48 cyclic mechanical loads. Following the pioneering work by Gong et al. (12), hydrogels have been
49 made tough to resist crack propagation under a single cycle of mechanical load (13-16). The
50 toughening of hydrogels is achieved by integrating mechanisms for dissipating mechanical energy
51 such as fracture of short polymer chains and reversible crosslinks into stretchy polymer networks
52 (17, 18). However, existing tough hydrogels still suffer from fatigue fracture under multiple cycles
53 of mechanical loads (10, 19, 20), because the resistance to fatigue-crack propagation after
54 prolonged cycles of loads is the energy required to fracture a single layer of polymer chains (i.e.,
55 the intrinsic fracture energy of the hydrogel), which is unaffected by the additional dissipation
56 mechanisms introduced in tough hydrogels (19, 20). The reported fatigue thresholds of various
57 tough hydrogels are 8.4 J/m² for polyacrylamide (PAAm)-polyvinyl alcohol (PVA) (20) and 53.2
58 J/m² for PAAm-alginate (19), the same order as their intrinsic fracture energies. The highest fatigue
59 threshold for hydrogels reported so far is 418 J/m² for a double network hydrogel, poly(2-
60 acrylamido-2-methyl-1-propanesulfonic acid) (PAMPS)-PAAm, which possibly can be attributed
61 to the PAAm network with very long polymer chains and thus high intrinsic fracture energy (12,
62 21). A general strategy towards the design of anti-fatigue-fracture hydrogels has remained as a
63 critical need and a central challenge for long-term applications of hydrogels in devices and
64 machines.

65 In contrast to synthetic hydrogels, biological tissues such as cartilages, tendons, muscles
66 and heart valves show extraordinary anti-fatigue **properties**. For example, the knee joint of an

average person needs to sustain peak stresses of 4–9 MPa for 1 million cycles per year, and its fracture energy after prolonged cycles of loads is above 1,000 J/m² (22, 23). The anti-fatigue property of biological tissues possibly arises from the inherent highly ordered and partially crystalline structures of collagen fibers in the tissues (24). Inspired by anti-fatigue biological tissues, here we hypothesize that the increase of crystallinity in synthetic hydrogels can significantly enhance their fatigue thresholds, due to the need to fracture crystalline domains for fatigue-crack propagation (Fig. 1). The energy per unit area required to fracture crystalline domains of a polymer is much higher than that required to fracture a single layer of amorphous chains of the same polymer (25). To test the hypothesis, we select PVA as a model hydrogel with tunable crystallinity. We increase the annealing time (after freeze-thawing and air-drying) of the PVA hydrogel to give higher crystallinity, larger crystalline domain size, and smaller average distance between adjacent domains in the hydrogel (Fig. 1A). We then measure the fatigue thresholds of PVA hydrogels with various crystallinities (Fig. 1, B and C). We find that the increase of crystallinity can significantly enhance the fatigue thresholds of PVA hydrogels. In particular, the fatigue threshold can exceed 1,000 J/m² when the crystallinity of PVA in the swollen state reaches 18.9 wt.%. By annealing selected regions in hydrogels, we further demonstrate a general strategy to pattern highly-crystalline regions in the PVA hydrogels to render them resistant to fatigue-fracture but still maintain their high water content and low modulus. With this strategy, we create *kirigami hydrogel sheets* that are highly stretchable and resistant to fatigue-fracture by introducing patterned cuts into hydrogel sheets and then reinforcing the cut tips. The current work not only reveals a new anti-fatigue-fracture mechanism in hydrogels but also provides a practical method to design anti-fatigue-fracture hydrogels for diverse applications.

Results

To validate the hypothesis that the increase of crystallinity in hydrogels can significantly enhance their fatigue thresholds, we use PVA hydrogels as a model material system with tunable

92 crystallinity. We first freeze a solution of uncrosslinked PVA at -20 °C for 8 h and thaw it at 25 °C
93 for 3 h, to form a hydrogel crosslinked by crystalline domains (26). The freeze-thawed PVA
94 hydrogel is further dried in an incubator at 37 °C and then annealed at 100 °C for various times
95 ranging from 0 min to 90 min (27). (The hydrogel dry-annealed for 0 min means no annealing
96 process.) The crystallinity of the hydrogel can be tuned by drying and annealing it for different
97 times. As a control sample, we also fabricate a chemically-crosslinked PVA as a reference hydrogel
98 composed of an entirely amorphous polymer network (See Materials and Methods for details on
99 fabrication of the samples).

100 *Characterization of the crystalline morphology in PVA hydrogels*

101 We first measure the crystallinities of the resultant PVA hydrogels in their dry state by
102 differential scanning calorimetry (DSC). For the chemically-crosslinked and freeze-thawed PVA
103 hydrogels, we use excess chemical crosslinks to fix the amorphous polymer chains before air-drying
104 them in an incubator at 37 °C (see Materials and Methods for details). The excess chemical
105 crosslinks minimize the formation of further crystalline domains during the air-drying process (28,
106 29). As shown in Fig. 2A, both the chemically-crosslinked and freeze-thawed PVA hydrogels show
107 negligible endothermic peaks with measured crystallinities in the dry state of 0.2 wt.% and 2.1
108 wt.%, respectively. However, when the freeze-thawed PVA (without excess chemical crosslinks)
109 is dried in air, the crystallinity in the dry state increases to 37.7 wt.% (see Fig. 2A). The increase of
110 the crystallinity implies that substantially more crystalline domains nucleate during the air-drying
111 process (27). The crystallinity further increases gradually by increasing the annealing time. When
112 the sample is annealed for 90 min at 100 °C, the crystallinity in the dry state reaches 47.3 wt.%. In
113 addition to the crystallinities in the dry state measured from DSC, we further measure the water
114 contents in the fully swollen samples shown in Fig. 2B and calculate the corresponding
115 crystallinities in the swollen state shown in Fig. 2C. It is well-known that the as-prepared dry PVA
116 samples may contain residual water bonded with polymer chains (29). The amount of residual water

117 can be calculated from the endothermic transition ranging from 60 °C to 180 °C on the DSC curves
118 (See Fig. 2A and fig. S1) (26). The above reported crystallinities in the dry state (without residual
119 water), water contents and crystallinities in the swollen state have been corrected to account for the
120 weights of residual water in the as-prepared dry samples (see Materials and Methods, table S1 and
121 fig. S1 for details).

122 To quantify the evolution of crystalline morphology, we measure the average distance
123 between adjacent crystalline domains L through small-angle X-ray scattering (SAXS) and the
124 average size of crystalline domains D through wide-angle X-ray scattering (WAXS) (see details in
125 Materials and Methods). We first perform SAXS measurements on the samples in the swollen state
126 after subtracting the water background, measuring the scattering intensity $I(q)$ versus the scattering
127 vector q . To identify the location of the peak intensity, we correct the intensity by multiplying the
128 scattering intensity with the scattering vector q (30). As shown in Fig. 2D, there is no peak in the
129 plot of the corrected intensity Iq versus the scattering vector q for the freeze-thawed hydrogel,
130 which implies negligible interference between adjacent crystalline domains. For the hydrogel dry-
131 annealed for 0 min, there is a slight shoulder shown in the corrected intensity curve, which indicates
132 stronger interference between adjacent domains. The average distance between adjacent crystalline
133 domains L can be estimated from the critical vector corresponding to the peak intensity q_{max} ,
134 following the Bragg expression $L = 2\pi / q_{max}$ (30). The average distance for the hydrogel dry-
135 annealed for 0 min is estimated to be 21 nm in the swollen state. As the annealing time increases to
136 90 min, the average distance L decreases to 13 nm in the swollen state (Fig. 2G). As a control case,
137 we also measure SAXS profiles of the hydrogel dry-annealed 90 min in the dry state. As shown in
138 Fig. 2F, the average distance between adjacent crystalline domains in the dry state is around 9 nm,
139 smaller than the distance in the same hydrogel in the swollen state (i.e., 13 nm). This is because
140 swelling of the interstitial amorphous polymer chains increases the distance between adjacent
141 crystalline domains.

We further perform WAXS measurements on the hydrogels in their dry state using Ni-filtered $\text{CuK}\alpha 1$ radiation with X-ray wavelength $\lambda = 1.54 \text{ \AA}$. As shown in Fig. 2E, all dry-annealed PVA hydrogels show a strong diffraction peak at $2\theta = 19.7^\circ$, which corresponds to the typical reflection plane of $(10\bar{1})$ in semi-crystalline PVA (31). In addition, small peaks at $2\theta = 11.5^\circ$ and 23.1° are also observed in the hydrogel dry-annealed for 90 min, suggesting a high crystallinity in the hydrogel, which is consistent with the DSC measurement. By identifying the half-width of the maximum diffraction peak β , the average size of crystalline domains D can be approximately calculated using Scherrer's equation $D = k\lambda / (\beta \cos \theta)$ (32), where k is a dimensionless shape factor varying with the actual shape of the crystalline domain; λ is the wavelength of X-ray diffraction; and θ is the Bragg angle. Here, β is identified after subtracting the instrumental line broadening; and the dimensionless shape factor k is set as 1, approximating the spherical shape of the crystalline domains (33). As shown in Fig. 2G, by increasing the annealing time from 0 min to 90 min, the average size of the crystalline domains increases from 3.8 nm to 6.5 nm. This trend is consistent with the decrease of the average distance between adjacent crystalline domains with annealing time, since the growth of the crystalline domains consumes the interstitial amorphous polymer chains.

To further validate the tuning of crystalline domains in the PVA hydrogel with annealing time, we use tapping-mode atomic force microscopy (AFM) to obtain phase images of the hydrogels dry-annealed for 0 min and 90 min, respectively. The bright areas in Fig. 2H correspond to the regions with relatively high modulus (mainly crystalline domains), whereas the dark areas represent the regions with relatively low modulus (mainly amorphous domains). As shown in Fig. 2H, the morphology of isolated crystalline domains is observed in the hydrogel dry-annealed for 0 min, while the hydrogel dry-annealed for 90 min shows larger aggregated crystalline domains.

Characterization of fatigue fracture properties of hydrogels

To measure the fatigue threshold of PVA hydrogels, we adopt the single-notch method, which has been widely used in fatigue tests of rubbers (34, 35). Notably, all fatigue tests in this study are performed on fully-swollen hydrogels immersed in a water bath, to prevent the dehydration-induced crack propagation (fig. S2). We use **dogbone-shaped** samples and perform cyclic tensile tests on both notched and unnotched samples, which are otherwise the same (Fig. 1, B and C). The nominal stress versus stretch curves (i.e., S vs. λ) of unnotched samples are obtained over N cycles of applied stretch λ^A . The strain energy density W of the unnotched sample under the N_{th} cycle of applied stretch λ^A can be calculated as

$$W(\lambda^A, N) = \int_1^{\lambda^A} S d\lambda \quad (1)$$

where S and λ are the measured nominal stress and stretch, respectively. Thereafter, the same cyclic stretch λ^A is applied on the notched sample, measuring the evolution of the cut length in undeformed state c with the cycle number N . The applied energy release rate G in the notched sample under the N_{th} cycle of applied stretch λ^A can be calculated as (34, 35)

$$G(\lambda^A, N) = 2k(\lambda^A) \cdot c(N) \cdot W(\lambda^A, N) \quad (2)$$

where k is a slowly varying function of the applied stretch expressed as $k = 3 / \sqrt{\lambda^A}$, c the current crack length at undeformed configuration, and W the strain energy density measured in the unnotched sample (Eq. 1). By systematically varying the applied stretch λ^A , we can obtain a plot of crack extension per cycle versus the applied energy release rate (i.e., dc / dN vs. G). By linearly extrapolating the curve of dc / dN vs. G to the intercept with the **abscissa**, we can approximately obtain the critical energy release rate G_c , below which the fatigue crack will not propagate under infinite cycles of loads. By **definition**, the fatigue threshold Γ_0 is equal to the critical energy release rate G_c . To validate that this extrapolated value G_c is indeed the fatigue threshold Γ_0 , we further apply G_c to the notched sample over 30,000 cycles (to approximate infinite cycles of loads) and

observe no crack extension (Fig. 3H). In addition, we also measure the fatigue thresholds using the pure-shear test (19) to validate the results from the single-notch test (fig. S6).

For cyclic tensile tests on unnotched samples (Fig. 3, A and B and fig. S3), both chemically-crosslinked PVA and freeze-thawed PVA show negligible Mullins effect, and their S vs. λ curves reach steady states after only a few cycles (i.e., 10 for chemically-crosslinked PVA at an applied stretch of 1.6, and 200 for freeze-thawed PVA at an applied stretch of 2.2). On the other hand, the dry-annealed PVA hydrogels exhibit a more significant Mullins effect, due to mechanical dissipation caused by melting and reorientation of crystalline domains (30). As the stretch further increases, the crystalline domains may transform into aligned fibrils along the loading direction (30). The energy required to damage the crystalline domains and fibrils is much higher than that to fracture a single layer of the same polymer in amorphous state. The hydrogel dry-annealed 90 min reaches steady state after 1,000 cycles of applied stretches of $\lambda^A = 2$ (Fig. 3C). Despite the Mullins effect, the steady-state maximum nominal stress of the hydrogel dry-annealed for 90 min is much higher than that of both the chemically-crosslinked PVA and the freeze-thawed PVA at the same applied stretch of $\lambda^A = 2$ (e.g., 2.3 MPa for hydrogel dry-annealed 90 min, 15 kPa for chemically-crosslinked PVA, and 3 kPa for freeze-thawed PVA in fig. S4).

For cyclic tensile tests on notched samples, a pre-crack is cut using a razor blade with tip radius of around 200 μm and initial crack length around 1 mm, smaller than one-fifth of the width of the sample (35). A digital microscope (AM4815ZT, Dino-Lite, resolution 20 $\mu\text{m}/\text{pixel}$) is used to record the cut length under cyclic loads (Materials and Methods, fig. S5). We first apply cyclic loads with a small applied stretch (i.e., $\lambda^A = 1.3$) on a notched sample. If the crack remains quasi-stationary with crack extension per cycle (dc/dN) smaller than 20 nm/cycle (i.e., no detectable crack extension in 1,000 cycles), the applied cyclic stretch is increased by the increment of $\Delta\lambda^A = 0.1$ for other notched samples until crack propagation greater than 20 nm/cycle is captured. As shown in Fig. 3, D and E, the fatigue thresholds of chemically-crosslinked PVA and freeze-

214 thawed PVA are measured to be 10 J/m² and 23 J/m², respectively. The fatigue threshold for the
215 hydrogel dry-annealed 0 min increases to 110 J/m². As the annealing time increases, the fatigue
216 threshold further increases. In particular, for the hydrogel dry-annealed 90 min, the fatigue
217 threshold can achieve 1000 J/m² (Fig. 3F and fig. S5). The measured fatigue threshold of the
218 hydrogel dry-annealed 90 min from the pure-shear test is 918 J/m², consistent with the single-notch
219 test (fig. S6). The dependence of fatigue threshold on the crystallinity is summarized in Fig. 3G.
220 The fatigue threshold increases with the crystallinity and demonstrates a sharp jump when the
221 crystallinity in the swollen state reaches approximate 15 wt.%.

222 In addition to fatigue tests, we also measure the nominal stress versus stretch curves of all
223 hydrogels to obtain their Young's moduli and tensile strengths (fig. S7). As shown in Fig. 4, A and
224 B, both the Young's modulus and tensile strength increase with the hydrogels' crystallinity and
225 show dramatic enhancements when the crystallinity in the swollen state reaches approximate 15
226 wt.% (36). This sharp jump in Young's modulus and tensile strength is consistent with the dramatic
227 increase of fatigue threshold of the hydrogel at approximate 15 wt.% crystallinity in the swollen
228 state (Fig. 3G).

229

230 ***Patterning of highly-crystalline regions in hydrogels***

231 Whereas annealing the whole PVA hydrogel can significantly enhance its fatigue threshold, the
232 annealing treatment also increases the modulus and decreases the water content of the hydrogel
233 (Fig. 4C and fig. S8). However, for many applications, it is desirable to maintain the relatively low
234 modulus and high water content of the hydrogels. Here we propose a strategy to introduce
235 programed highly-crystalline regions in the hydrogels. We use the computer-aided design of
236 electrical circuits to induce localized heat treatment for annealing selected regions of the hydrogels
237 (See details in Materials and Methods and fig. S9). The chemically-crosslinked PVA hydrogel is
238 used as the pristine sample with low fatigue threshold of 15 J/m², low Young's modulus of 114 kPa

239 and high water content of 88 wt.%. Three examples of programmed annealing patterns on the pristine
240 PVA hydrogels are demonstrated with enhanced fatigue thresholds.

241 We first locally introduce a highly-crystalline ring-shaped region around a crack tip (Fig.
242 5A), leading to a fatigue threshold over 236 J/m² (Fig. 5B). Despite the small area of the highly-
243 crystalline region, this local conditioning can greatly reinforce the crack tip, delaying crack
244 propagation. Meanwhile, the measured overall Young's modulus and water content of the sample
245 are maintained at 114 kPa and 87 wt.% (Fig. 5, E and F), respectively, which are almost unaffected
246 by the local annealing around the crack tip. As a second example, we pattern mesh-like highly-
247 crystalline regions on the pristine hydrogel (Fig. 5C). Compared with the pristine sample, the
248 fatigue threshold of the mesh-reinforced sample increases to 290 J/m² (Fig. 5D), and its Young's
249 modulus remains relatively low (627 kPa) and its water content stays relatively high as 83 wt.%
250 (Fig. 5, E and F).

251 The strategy of patterning highly-crystalline regions can be applied to various structures of
252 hydrogels for improving anti-fatigue performances as well. For example, the kirigami structure is
253 commonly adopted to enhance the stretchability (37) and effective adhesion (38) of films. As a third
254 example, we demonstrate a kirigami hydrogel sheet with improved anti-fatigue performance by
255 patterning highly-crystalline regions around the cut tips. As shown in Fig. 5G, we first generate a
256 kirigami pattern on a pristine sample by introducing parallel periodic cuts with equal length and
257 equal distance between adjacent cuts. The ultimate stretch and effective nominal stress (i.e., force
258 divided by cross-section area) of the pristine kirigami hydrogel sheet under a single cycle of load
259 are measured to be 1.7 and 0.9 kPa, respectively. Thereafter, the pristine kirigami sheet is reinforced
260 around the cut tips with local annealing. The reinforced kirigami sheet can sustain cyclic tensile
261 loads at an applied stretch of $\lambda^A = 2.1$ without detectable fatigue-fracture even when the cycle
262 number approaches 3,000. Moreover, the effective nominal stress after prolonged cycles at the

263 applied stretch of $\lambda^A = 2.1$ can maintain the plateau of 13 kPa, which is 14.4 times the strength of
264 the pristine kirigami sheet (Fig.5, H and I).

265 Figure 5J and K compare the fatigue thresholds, water contents and the Young's moduli of
266 reported hydrogels in literature (19-21, 39, 40). We show that by patterning highly-crystalline
267 regions, both tip-reinforced and mesh-reinforced PVA hydrogels outperform existing synthetic
268 hydrogels in terms of **fatigue thresholds**, and they can still maintain relatively high water contents
269 and relatively low Young's moduli.

270

271 Discussion

272 We **have demonstrated** that the fatigue threshold of hydrogels can be greatly enhanced by designing
273 crystalline domains in the hydrogels. We use PVA hydrogels as a model material to validate this
274 new mechanism for designing anti-fatigue-fracture hydrogels. The fatigue threshold of a PVA
275 hydrogel with a crystallinity of 18.9 wt.% in the swollen state can achieve over 1000 J/m². We
276 further develop a strategy to make PVA hydrogels anti-fatigue-fracture but still maintain their high
277 water content and low modulus by patterning highly-crystalline regions in the hydrogels. The
278 reported mechanism and strategy for designing anti-fatigue-fracture hydrogels can be extended to
279 hydrogel composites with fillers such as nanoclay, nanocellulose and nanofibers.

280 The capability to enhance the anti-fatigue-fracture performance of synthetic hydrogels
281 makes a number of future research directions and applications possible. For example, anti-fatigue
282 hydrogels can be used for hydrogel-based gastric-retentive devices and implantable tissue
283 replacements of meniscus, intervertebral disk and cartilage, **which require** long-term mechanical
284 robustness when interacting with the human body.

285

286 Materials and Methods

287 Synthesis of PVA hydrogels

288 All types of our PVA hydrogels are synthesized from 10 wt.% poly(vinyl alcohol) (PVA; Mw
289 146,000-186,000, 99+% hydrolyzed; Sigma-Aldrich, 363065) solution. The solution was heated in
290 a water bath at 100 °C with stirring for 5 h. To synthesize chemically-crosslinked PVA hydrogels,
291 we added 10 µL glutaraldehyde (25 vol.%, Sigma-Aldrich, G6257) as a crosslinker to 1 mL 10
292 wt.% PVA solution, and added 10 µL hydrochloric acid (36.5-38 wt.%, J.T. Baker, 9535-02) as an
293 accelerator into the other 1 mL 10 wt.% PVA solution. We then mixed and defoamed each of them
294 by using a centrifugal mixer (AR-100; Thinky). The final mixtures, obtained by mixing and
295 defoaming the two solutions together, were then cast into a mold and allowed to cure for 2 h. The
296 chemically-crosslinked PVA hydrogels were immersed in deionized water for two days to remove
297 unreacted chemicals. To fabricate freeze-thawed PVA hydrogels, 10 wt.% PVA solutions after
298 mixing and defoaming were poured into a mold, frozen at -20 °C for 8 h and thawed at 25 °C for 3
299 h. The freeze-thawed hydrogels were further dried in an incubator (New Brunswick Scientific, C25)
300 at 37 °C for 2 h, and then annealed at 100 °C for a controlled time (i.e., 0, 1, 3, 5, 10, or 90 min).
301 All as-prepared PVA hydrogels were immersed in water to achieve their equilibrium-swollen state.

302

303 **Measurement of residual water and crystallinity in dry samples**

304 We measured the crystallinities of the resultant PVA hydrogels in their dry state by differential
305 scanning calorimetry (DSC/cell: RCS1-3277 Cooling System: DSC1-0107). For as-prepared
306 chemically-crosslinked PVA and freeze-thawed PVA, we used excess chemical crosslinks to fix
307 the amorphous polymer chains before air-drying, minimizing the formation of further crystalline
308 domains during the air-drying process. We first soaked the samples (thickness of 1mm) in the
309 aqueous solution consisting of 10 mL glutaraldehyde (25 vol.%, Sigma-Aldrich, G6257), 500 µL
310 hydrochloric acid (36.5-38 wt.%, J.T. Baker, 9535-02) and 50 mL DI water for 2 h. Thereafter, we
311 soaked the samples in a DI water bath for 2 h to remove the residual hydrochloric acid. The samples
312 were further dried in an incubator (New Brunswick Scientific, C25) at 37 °C for 2 h.

Thereafter, we measured the mass of residual water $m_{residual}$, the mass of crystalline domains $m_{crystalline}$ and the total mass of the dry samples (with residual water) m using DSC. In a typical DSC measurement, we first weighed the total mass of the dry sample (with residual water) m . The sample were thereafter placed in a Tzero-pan and heated up from 50 °C to 250 °C at the rate of 20 °C/min under a nitrogen atmosphere with flow rate of 30 mL/min. The curve of heat flow shows a broad peak from 60 °C to 180 °C, indicating that the sample contains a small amount of residual water. The integration of the endothermic transition ranging from 60 °C to 180 °C gives the enthalpy for evaporation of the residual water per unit mass of the dry sample (with residual water) $H_{residual}$. Therefore, the mass of the residual water $m_{residual}$ can be calculated as

$$m_{residual} = m \cdot \frac{H_{residual}}{H_{water}^0} \quad (3)$$

where $H_{water}^0 = 2260$ J/g is the latent heat of water evaporation. The curve of heat flow shows another narrow peak ranging from 200 °C to 250 °C corresponding to melting of the crystalline domains. The integration of the endothermic transition ranging from 200 °C to 250 °C gives the enthalpy for **melting** the crystalline domains per unit mass of the dry sample (with residual water) $H_{crystalline}$. Therefore, the mass of the crystalline domains $m_{crystalline}$ can be calculated as

$$m_{crystalline} = m \cdot \frac{H_{crystalline}}{H_{crystalline}^0} \quad (4)$$

where $H_{crystalline}^0 = 138.6$ J/g is the enthalpy of fusion of 100 wt.% crystalline PVA measured at the equilibrium melting point T_m^0 (29). Therefore, the crystallinity in the ideally dry sample X_{dry} (without residual water) can be calculated as

$$X_{dry} = \frac{m_{crystalline}}{m - m_{residual}} \quad (5)$$

Measurement of water content and crystallinity in swollen samples

335 The swollen hydrogels weighing $m_{swollen}$ were placed in an incubator (New Brunswick Scientific,
 336 C25) at 37 °C for 2 h, and weighed m after air-drying. The mass of the residual water in the as-
 337 prepared dry samples $m_{residual}$ was measured in the previous section. Therefore, the water content in
 338 the swollen state can be calculated as $(m_{swollen} - m + m_{residual}) / m_{swollen}$ and the polymer content in
 339 the swollen state can be calculated as $(m - m_{residual}) / m_{swollen}$. In addition, the crystallinity in the
 340 swollen sample $X_{swollen}$ can be calculated as

$$341 \quad X_{swollen} = \frac{m_{crystalline}}{m_{swollen}} = X_{dry} \cdot \frac{m - m_{residual}}{m_{swollen}} \quad (6)$$

342 where $m_{crystalline}$ and X_{dry} were measured as described in the previous section.

343 **AFM phase imaging**

344 AFM phase images were acquired by atomic force microscope (MFP-3D, Asylum Research) in
 345 tapping mode. Dry free-standing PVA films were directly attached onto sample stage by double-
 346 sided carbon tape. The probe lightly taps on the sample surface with recorded phase shift angle of
 347 the probe motion relative to a driving oscillator. The bright regions with high phase angle
 348 correspond to regions with relatively high modulus; and the dark regions with low phase angle
 349 correspond to regions with relatively low modulus.

350 **X-ray scattering**

351 The X-ray scattering measurement was performed with Pilatus3R 300K detector Bruker Nanostar
 352 SAXS in X-ray Diffraction Shared Experimental Facility at MIT. We used small angle 2 mm
 353 beamstop with sample-detector distance of 1059.1 mm for SAXS measurements and wide angle
 354 2mm beamstop with sample-detector distance of 109.1 mm for WAXS measurements. The
 355 exposure time was set as 300 s. Raw SAXS and WAXS patterns were processed with corrections
 356 by MATLAB-based GIXSGUI software before analysis.

357 **Raman spectroscopy**

Each sample was hydrated for more than 2 h and then pressed between a glass slide and a cover slip in order to ensure a flat surface. The cover slip was then sealed at the edges with nail polish to prevent the hydrogel from drying. A confocal Raman microscope (Alpha300RA; WiTec, Germany) with 20× objective (Zeiss, Germany) was used. Nd:YAG laser (532 nm) was used as the excitation source with the maximum power of 75mW. Data was collected with a CCD detector (DU401A-BV; Andor, UK) behind a 600 g/mm grating spectrometer (UHTS 300; WiTec, Germany). A 20- μ m resolution Raman map of 4×3 mm scan area was acquired with an accumulation time of 1 second per point. Each point was pre-bleached for 400 ms to decrease the effect of fluorescence. Cosmic ray removal and background subtraction were performed to clean the spectra. The intensity of O-H bond within the PVA and water were calculated by integrating the spectra in the range of 2,800-3,000 cm^{-1} and 3,075-3,625 cm^{-1} , respectively. The ratio of PVA and water was then calculated and plotted as a heatmap shown in Fig. 5A.

Measurement of the fatigue threshold

All the mechanical tests were performed in a water bath at 25°C with a U-stretch testing device (CellScale, Canada). For mechanically weak samples (e.g., the Ch and FT hydrogels), a load cell with maximum force of 4.4 N was used; for mechanically strong samples (e.g., hydrogel dry-annealed 90 min), a load cell with maximum force of 44 N was used. The dogbone-shaped sample had the dimensions at as-prepared state with the width of 5mm, thickness of 0.8mm and gauge length of 10mm. The nominal stress S was measured from the recorded force F divided by width W and thickness t in the swollen state. To measure the applied stretch λ^A in gauge length, we adopted the digital image correlation method and calibrated the correlation between λ^A and loading distance d at gripping points (fig. S2). A digital microscope (AM4815ZT; Dino-Lite, resolution 20 μ m/pixel) was used to record the crack extension. Since dry annealed PVA is transparent when immersing in water, we spread a small amount of graphite powder on the surface of the sample for visualization.

383 **Preparation of samples with programmable crystalline domains**

384 A copper tape was first loaded in a paper cutting machine (Silhouette CAMEO 3). The electrical
385 circuits were first designed with CAD and loaded to the machine via Silhouette Studio software.
386 After completion of the cutting, the copper tape was placed on a flat acrylic plate and unwanted
387 parts were carefully peeled off. Two additional conductive wires were soldered on the copper tape
388 as two electrodes. An electrical generator (Instek PSB-2400L2; Tequipment) was used to apply
389 controlled current around 2-5 A on the designed circuits. To ensure achieving the designed
390 temperature, a thermal imager (Seek thermal XR imager) was first used to measure the local
391 temperature from the circuit (see [fig. S9](#)). By adjusting the current to control the designed
392 temperature, the air-dried PVA hydrogel was placed on the targeted circuits for local heating. A
393 gripper was used to ensure fully contact between the PVA film and copper circuits.

394 **Supplementary Material**

395 table S1. [Crystallinities and water contents in chemically-crosslinked \(Ch\), freeze-thawed \(FT\)](#)
396 [and dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min.](#)

397 [fig. S1. Measurement of the mass of freeze-thawed PVA during air-drying and the amount of](#)
398 [residual water in the sample after air-drying.](#)

399 [fig. S2. Experiment method for measuring fatigue threshold.](#)

400 [fig. S3. Shakedown softening of three types of PVA hydrogels.](#)

401 [fig. S4. Steady-state nominal stress versus stretch curves of PVA hydrogels with various](#)
402 [crystallinities.](#)

403 [fig. S5. Validation of high fatigue threshold with single-notch test.](#)

404 [fig. S6. Validation of high fatigue threshold with pure shear test.](#)

405 [fig. S7. Mechanical characterization of PVA hydrogels with various crystallinities.](#)

406 [fig. S8. Raman spectroscopy of freeze-thawed PVA and 90-min dry-annealed PVA.](#)

fig. S9. Electrical circuit and thermal mapping for programmable crystalline domains.

movie S1. Tension of the pristine notched sample and the tip-reinforced sample.

movie S2. Cyclic loading of the reinforced kirigami hydrogel sheet.

References and Notes

1. K. Y. Lee, D. J. Mooney, Hydrogels for tissue engineering. *Chem. Rev.* **101**, 1869-1880 (2001).
2. J. Li, D. J. Mooney, Designing hydrogels for controlled drug delivery. *Nat. Rev. Mater.* **1**, 16071 (2016).
3. L. Casares, R. Vincent, D. Zalvidea, N. Campillo, D. Navajas, M. Arroyo, X. Trepas, Hydraulic fracture during epithelial stretching. *Nat. Mater.* **14**, 343 (2015).
4. S. Lin, H. Yuk, T. Zhang, G. A. Parada, H. Koo, C. Yu, X. Zhao, Stretchable hydrogel electronics and devices. *Adv. Mater.* **28**, 4497-4505 (2016).
5. X. Liu, T.-C. Tang, E. Tham, H. Yuk, S. Lin, T. K. Lu, X. Zhao, Stretchable living materials and devices with hydrogel-elastomer hybrids hosting programmed cells. *Proc. Natl. Acad. Sci.* **114**, 2200-2205 (2017).
6. H. Yuk, S. Lin, C. Ma, M. Takaffoli, N. X. Fang, X. Zhao, Hydraulic hydrogel actuators and robots optically and sonically camouflaged in water. *Nat. Commun.* **8**, 14230 (2017).
7. H. Yuk, T. Zhang, S. Lin, G. A. Parada, X. Zhao, Tough bonding of hydrogels to diverse non-porous surfaces. *Nat. Mater.* **15**, 190 (2016).
8. S. Lin, H. Yuk, T. Zhang, G. A. Parada, H. Koo, C. Yu, X. Zhao, Stretchable hydrogel electronics and devices. *Advanced Materials* **28**, 4497-4505 (2016).
9. Q. Liu, G. Nian, C. Yang, S. Qu, Z. Suo, Bonding dissimilar polymer networks in various manufacturing processes. *Nat. Commun.* **9**, 846 (2018).
10. C. Yang, Z. Suo, Hydrogel ionotronics. *Nat. Rev. Mater.*, 1 (2018).
11. D. Wirthl, R. Pichler, M. Drack, G. Kettlhuber, R. Moser, R. Gerstmayr, F. Hartmann, E. Bradt, R. Kaltseis, C. M. Siket, Instant tough bonding of hydrogels for soft machines and electronics. *Sci. Adv.* **3**, e1700053 (2017).
12. J. P. Gong, Y. Katsuyama, T. Kurokawa, Y. Osada, Double-network hydrogels with extremely high mechanical strength. *Adv. Mater.* **15**, 1155-1158 (2003).
13. J.-Y. Sun, X. Zhao, W. R. Illeperuma, O. Chaudhuri, K. H. Oh, D. J. Mooney, J. J. Vlassak, Z. Suo, Highly stretchable and tough hydrogels. *Nature* **489**, 133 (2012).
14. T. L. Sun, T. Kurokawa, S. Kuroda, A. B. Ihsan, T. Akasaki, K. Sato, M. A. Haque, T. Nakajima, J. P. Gong, Physical hydrogels composed of polyampholytes demonstrate high toughness and viscoelasticity. *Nat. Mater.* **12**, 932 (2013).
15. Q. Chen, L. Zhu, C. Zhao, Q. Wang, J. Zheng, A robust, one-pot synthesis of highly mechanical and recoverable double network hydrogels using thermoreversible sol-gel polysaccharide. *Adv. Mater.* **25**, 4171-4176 (2013).
16. H. Chen, F. Yang, Q. Chen, J. Zheng, A Novel Design of Multi-Mechanoresponsive and Mechanically Strong Hydrogels. *Adv. Mater.* **29**, 1606900 (2017).
17. X. Zhao, Multi-scale multi-mechanism design of tough hydrogels: building dissipation into stretchy networks. *Soft Matter* **10**, 672-687 (2014).
18. J. P. Gong, Why are double network hydrogels so tough? *Soft Matter* **6**, 2583-2590 (2010).

19. R. Bai, Q. Yang, J. Tang, X. P. Morelle, J. Vlassak, Z. Suo, Fatigue fracture of tough hydrogels. *Extreme Mech. Lett.* **15**, 91-96 (2017).
20. R. Bai, J. Yang, X. P. Morelle, C. Yang, Z. Suo, Fatigue fracture of self-recovery hydrogels. *ACS Macro Lett.* **7**, 312-317 (2018).
21. W. Zhang, X. Liu, J. Wang, J. Tang, J. Hu, T. Lu, Z. Suo, Fatigue of double-network hydrogels. *Eng. Fract. Mech.* **187**, 74-93 (2018).
22. D. Taylor, N. O'Mara, E. Ryan, M. Takaza, C. Simms, The fracture toughness of soft tissues. *J. Mech. Behav. Biomed. Mater.* **6**, 139-147 (2012).
23. C. Creton, 50th anniversary perspective: networks and gels: soft but dynamic and tough. *Macromolecules* **50**, 8297-8316 (2017).
24. R. Berisio, L. Vitagliano, L. Mazzarella, A. Zagari, Crystal structure of the collagen triple helix model [(Pro-Pro-Gly) 10] 3. *Protein Sci.* **11**, 262-270 (2002).
25. A. J. Kinloch, *Fracture behaviour of polymers*. (Springer Science & Business Media, 2013).
26. C. M. Hassan, N. A. Peppas, Structure and morphology of freeze/thawed PVA hydrogels. *Macromolecules* **33**, 2472-2479 (2000).
27. J. Y. Chang, D. Godovsky, M. Han, C. Hassan, J. Kim, B. Lee, Y. Lee, N. Peppas, R. Quirk, T. Yoo, *Biopolymers: PVA Hydrogels Anionic Polymerisation Nanocomposites*. (Springer Science & Business Media, 2000), vol. 153.
28. K. C. Figueiredo, T. L. Alves, C. P. Borges, Poly (vinyl alcohol) films crosslinked by glutaraldehyde under mild conditions. *J. Appl. Polym. Sci.* **111**, 3074-3080 (2009).
29. N. A. Peppas, E. W. Merrill, Differential scanning calorimetry of crystallized PVA hydrogels. *J. Appl. Polym. Sci.* **20**, 1457-1465 (1976).
30. R. Zhang, Q. Zhang, Y. Ji, F. Su, L. Meng, Z. Qi, Y. Lin, X. Li, X. Chen, F. Lv, Stretch-induced complexation reaction between poly (vinyl alcohol) and iodine: an in situ synchrotron radiation small-and wide-angle X-ray scattering study. *Soft matter* **14**, 2535-2546 (2018).
31. R. Ricciardi, F. Auriemma, C. De Rosa, F. Lauprêtre, X-ray diffraction analysis of poly (vinyl alcohol) hydrogels, obtained by freezing and thawing techniques. *Macromolecules* **37**, 1921-1927 (2004).
32. A. Patterson, The Scherrer formula for X-ray particle size determination. *Phys. Rev.* **56**, 978 (1939).
33. E. Otsuka, S. Komiya, S. Sasaki, J. Xing, Y. Bando, Y. Hirashima, M. Sugiyama, A. Suzuki, Effects of preparation temperature on swelling and mechanical properties of PVA cast gels. *Soft Matter* **8**, 8129-8136 (2012).
34. G. Lake, P. Lindley, Cut growth and fatigue of rubbers. II. Experiments on a noncrystallizing rubber. *J. Appl. Polym. Sci.* **8**, 707-721 (1964).
35. A. Gent, P. Lindley, A. Thomas, Cut growth and fatigue of rubbers. I. The relationship between cut growth and fatigue. *J. Appl. Polym. Sci.* **8**, 455-466 (1964).
36. S. M. Liff, N. Kumar, G. H. McKinley, High-performance elastomeric nanocomposites via solvent-exchange processing. *Nature Materials* **6**, 76 (2007).
37. T. C. Shyu, P. F. Damasceno, P. M. Dodd, A. Lamoureux, L. Xu, M. Shlian, M. Shtein, S. C. Glotzer, N. A. Kotov, A kirigami approach to engineering elasticity in nanocomposites through patterned defects. *Nature Materials* **14**, 785 (2015).
38. R. Zhao, S. Lin, H. Yuk, X. Zhao, Kirigami enhances film adhesion. *Soft Matter* **14**, 2515-2525 (2018).
39. J. Tang, J. Li, J. J. Vlassak, Z. Suo, Fatigue fracture of hydrogels. *Extreme Mechanics Letters* **10**, 24-31 (2017).
40. E. Zhang, R. Bai, X. P. Morelle, Z. Suo, Fatigue fracture of nearly elastic hydrogels. *Soft Matter* **14**, 3563-3571 (2018).

41. D. Taylor, N. O'Mara, E. Ryan, M. Takaza, C. Simms, The fracture toughness of soft tissues. *Journal of the mechanical behavior of biomedical materials* **6**, 139-147 (2012).

Acknowledgments

The authors thank Dr. A. F. Schwartzman in the Nano Mechanical Technology Laboratory at MIT for his help with AFM phase imaging, J. Zhou at MIT for discussion on DSC measurements and Z. Jiang at Argonne National Laboratory for discussion on SAXS results. **Funding:** This work is supported by National Science Foundation (CMMI-1661627), Office of Naval Research (N00014-17-1-2920), National Institutes of Health (EB000244), and MIT Institute for Soldier Nanotechnologies (CMMI-1253495). H.Y. acknowledges the financial support from Samsung Scholarship. **Author Contributions:** S.L. and X.Z. conceived the idea, designed the study and interpreted the results. S.L. performed DSC, SAXS and AFM measurements. S.L. and X.L. performed mechanical characterization and fatigue tests. S.L., C.S. and J. S. interpreted the SAXS results. H.L. performed Raman spectroscopy. S.L., X.L., H.Y. and J.L. performed the demonstration of patterning highly crystalline regions. S.L., X.L., H.Y., J.L., G.A.P, C.S., G.H.M., and X.Z. analyzed and interpreted the results. S.L. and X.Z. drafted the manuscript with inputs from all other authors. X.Z. supervised the study. **Competing interests:** The authors declare that they have no competing interests. **Data and materials availability:** The data that support the findings of this study are available from the corresponding author upon request.

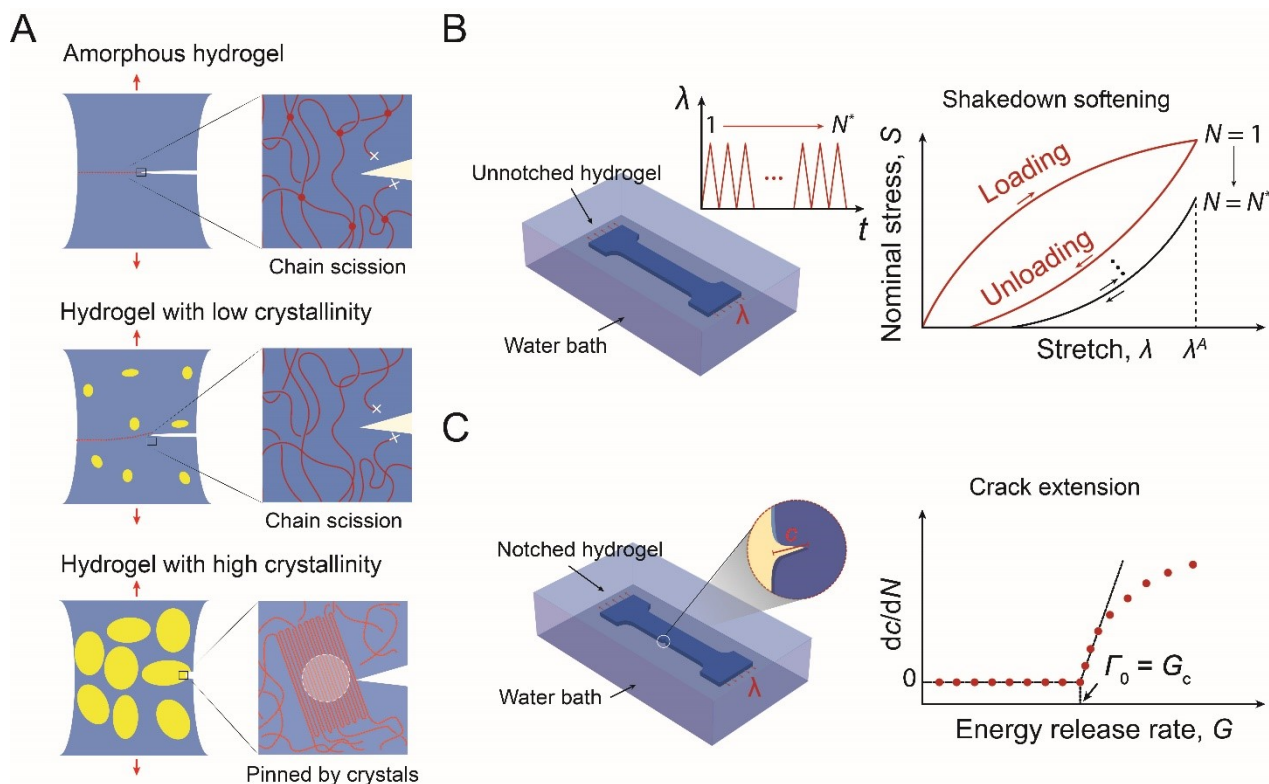


Fig. 1. The design strategy for anti-fatigue-fracture hydrogels. (A) Illustration of fatigue-crack propagation in an amorphous hydrogel and in hydrogels with low and high crystallinities under cyclic loads. The yellow areas represent crystalline domains and the blue areas denote amorphous domains. In the amorphous hydrogel and the hydrogel with low crystallinity, the fatigue threshold can be attributed to the energy required to fracture a single layer of polymer chains per unit area. In the hydrogel with high crystallinity, the fatigue-crack propagation requires fracture of crystalline domains. (B) Illustration of measuring nominal stress S vs. stretch λ curves over N cycles of the applied stretch λ^A . The stress-stretch curve reaches steady state as N reaches a critical value N^* . (C) Illustration of measuring crack extension per cycle dc/dN vs. energy release rate G curves. By linearly extrapolating the curve to intercept with the abscissa, we can approximately obtain the critical energy release rate G_c , below which the fatigue crack will not propagate under infinite cycles of loads. By definition, the fatigue threshold Γ_0 is equal to the critical energy release rate G_c .

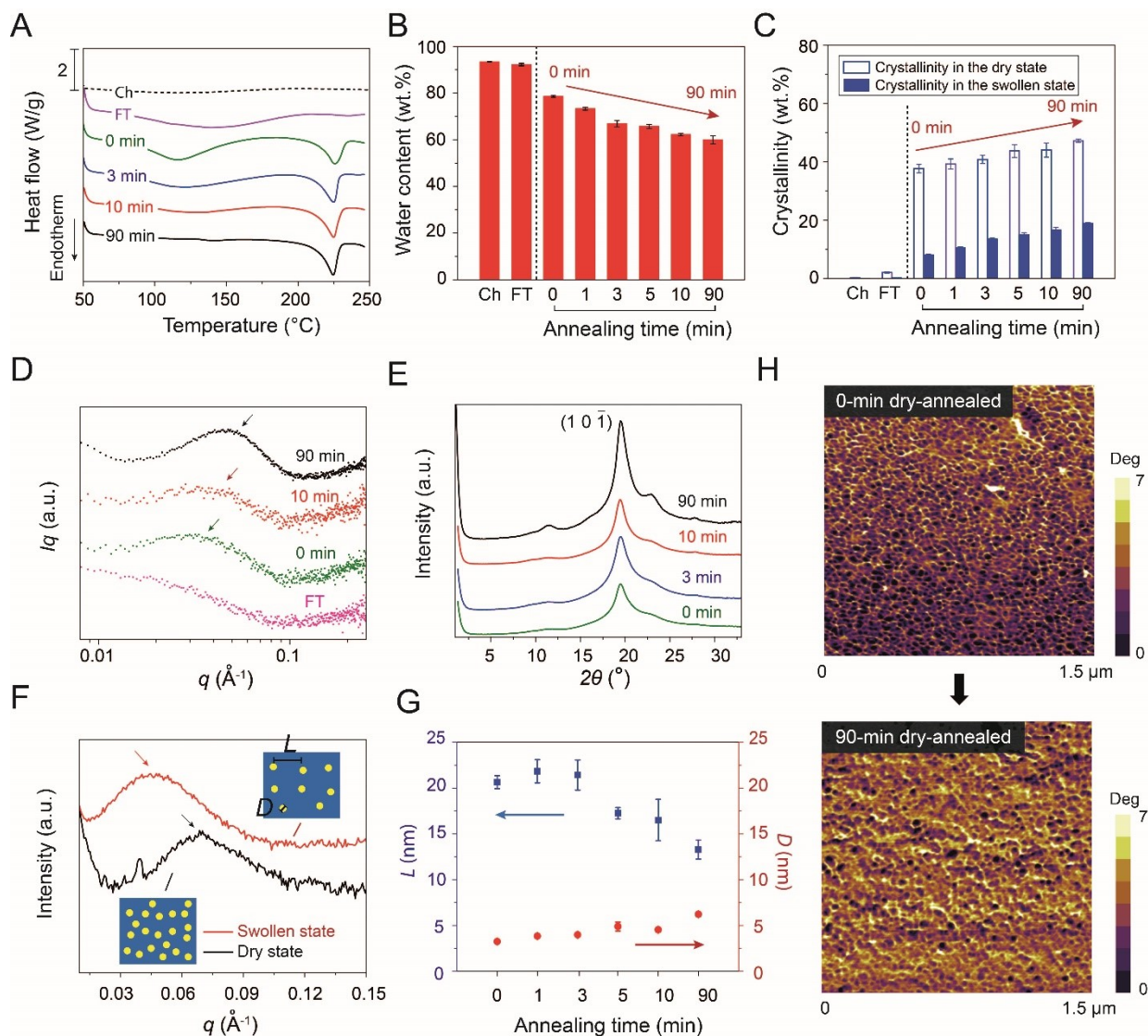


Fig. 2. Characterization of crystalline domains in PVA hydrogels. (A) Representative DSC thermographs of chemically-crosslinked (Ch), freeze-thawed (FT) and dry-annealed PVA with annealing time of 0, 3, 10, 90 min. (B) Water contents of chemically-crosslinked (Ch), freeze-thawed (FT) and dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min. (C) Measured crystallinity in the dry and swollen states of Ch, FT and dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min. (D) Representative SAXS profiles of FT hydrogel and dry-annealed PVA with annealing time of 0, 10, and 90 min. (E) Representative WAXS profiles of annealed PVA with annealing time of 0, 3, 10, and 90 min. (F) SAXS profiles of 90-min dry-annealed PVA in the dry state and the swollen state. The insets illustrate the increase of the distance between adjacent crystalline domains due to swelling of amorphous polymer chains. (G) The estimated average distance between adjacent crystalline domains L and average crystalline domain size D of dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min. (H) AFM phase images of dry-annealed hydrogel with annealing time of 0 min and 90 min. Data in B, C and G are means \pm SD, $n = 3$.

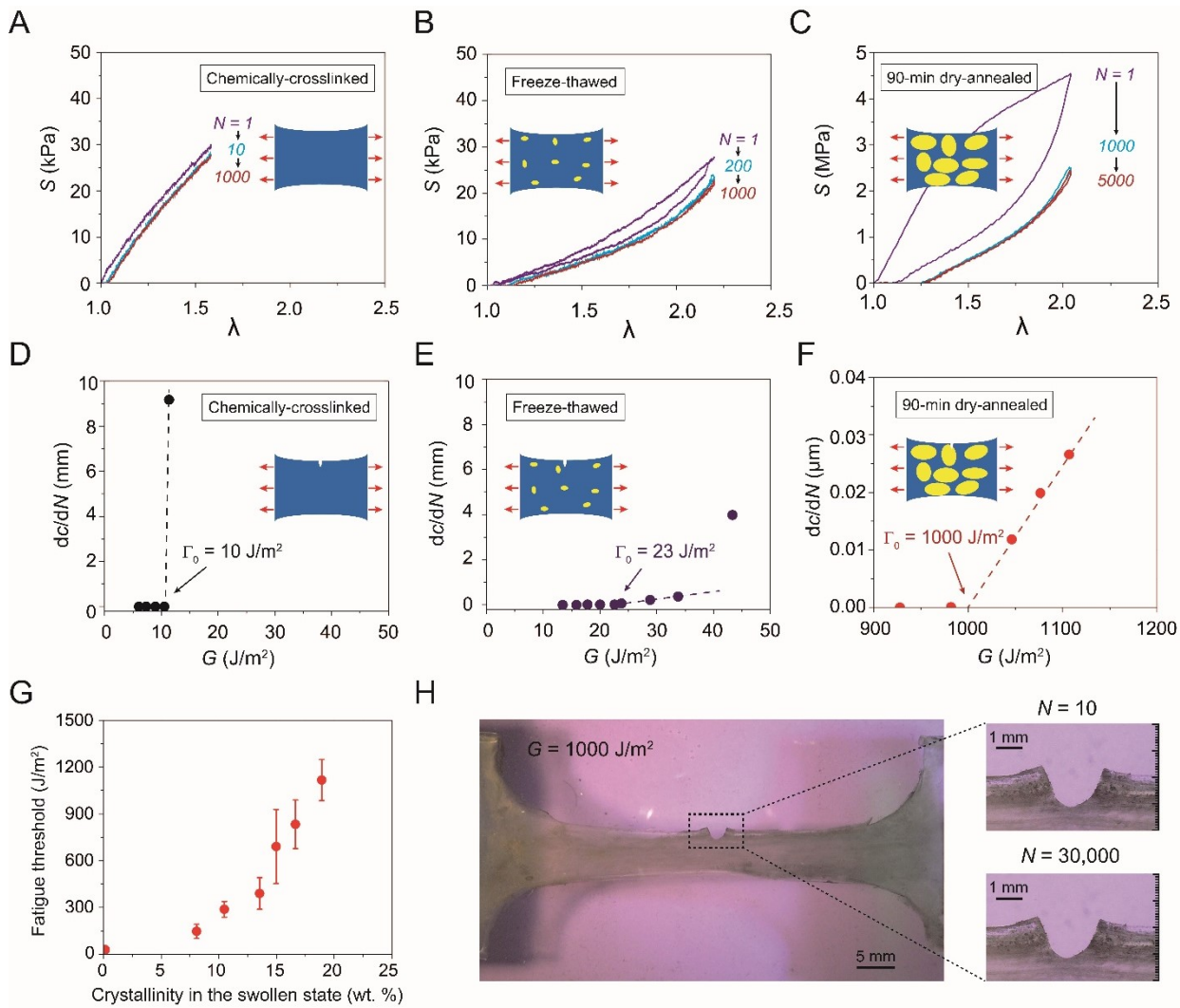


Fig. 3. Measurement of fatigue thresholds of PVA hydrogels. Nominal stress S vs. stretch λ curves over cyclic loads for (A) chemically-crosslinked hydrogel at an applied stretch of $\lambda^A = 1.6$, (B) freeze-thawed hydrogel at an applied stretch of $\lambda^A = 2.2$, and (C) 90-min dry-annealed hydrogel at an applied stretch of $\lambda^A = 2.0$. Crack extension per cycle dc/dN vs. applied energy release rate G for (D) chemically-crosslinked hydrogel, (E) freeze-thawed hydrogel, and (F) dry-annealed hydrogel with annealing time of 90 min. (G) The fatigue threshold increases with the crystallinity of the hydrogel in the swollen state. (H) Validation of fatigue threshold as high as 1000 J/m² in 90-min dry-annealed hydrogel using the single-notch test. Data in G are means \pm SD, $n = 3$. Scale bars are 5 mm and 1 mm for left and right images in H.

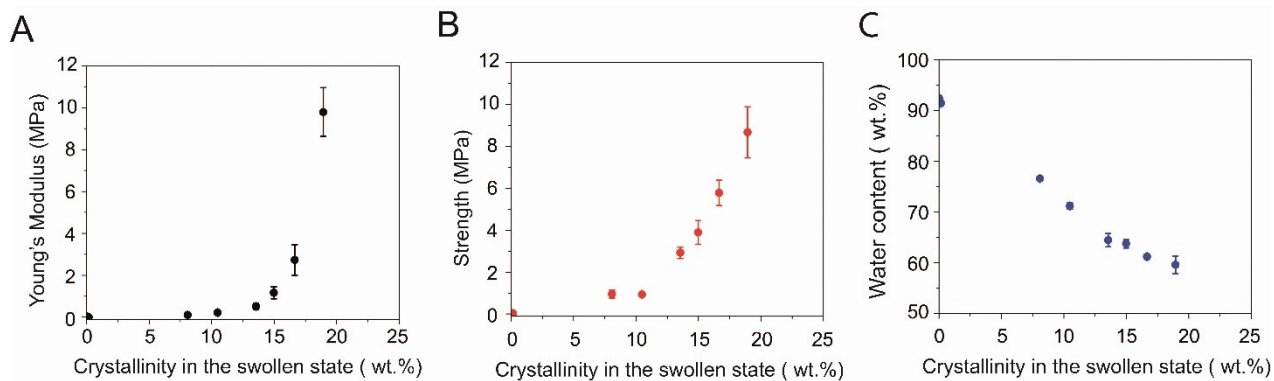


Fig. 4. Young's moduli, tensile strengths and water contents of PVA hydrogels. (A) Young's modulus versus crystallinity in the swollen state. **(B)** Tensile strength versus crystallinity in the swollen state. **(C)** Water content versus crystallinity in the swollen state. Data in **A**, **B** and **C** are means \pm SD, $n = 3$.

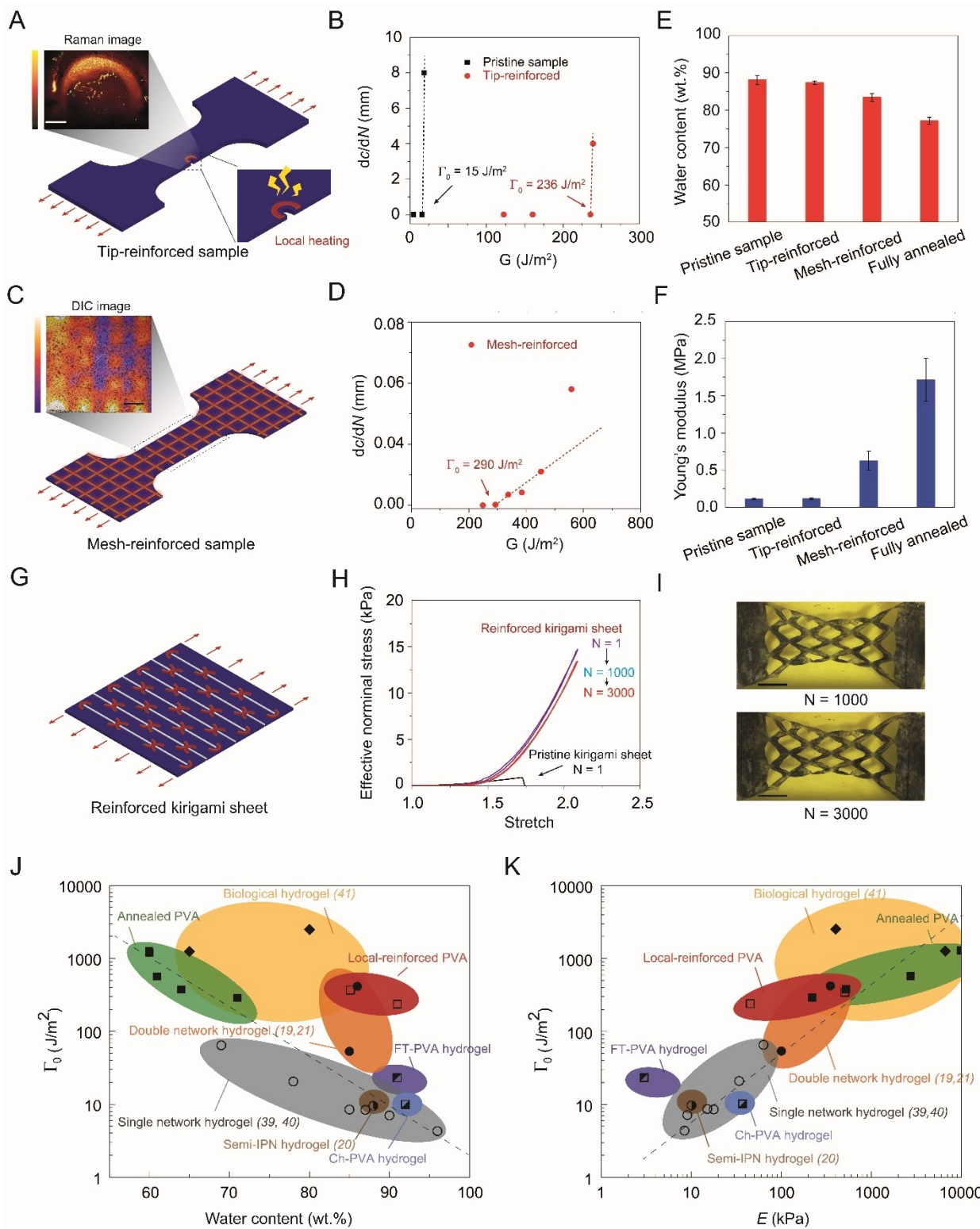


Fig. 5. Patterning highly-crystalline regions in PVA hydrogels. (A) Illustration of introducing a highly-crystalline region around crack tip. Inset: Raman spectroscopy with bright color representing low water content and dark color representing high water content (see details in Materials and Methods). (B) Comparison of crack extension per cycle dc/dN vs. applied energy releasing rate G between the pristine sample and the tip-reinforced sample. The fatigue thresholds of the pristine sample and the tip-reinforced sample are $15 J/m^2$ and $236 J/m^2$, respectively. (C) Illustration of introducing mesh-like highly-crystalline regions. Inset: Digital image correlation method (DIC)

shows large deformation in low-crystallinity regions and small deformation in high-crystallinity regions. **(D)** Crack extension per cycle dc/dN vs. applied energy releasing rate G of the mesh-reinforced sample. The fatigue threshold of the mesh-reinforced sample is 290 J/m^2 . **(E)** Water contents of the pristine sample, the tip-reinforced sample, the mesh-reinforced sample and the fully-annealed PVA hydrogels. **(F)** Young's moduli of the pristine sample, the tip-reinforced sample, the mesh-reinforced sample and the fully-annealed PVA hydrogels. **(G)** Illustration of introducing highly-crystalline regions around cut tips in a pristine kirigami sheet. **(H)** Effective nominal stress vs. stretch curves of the reinforced kirigami sheet under cyclic loading. Effective nominal stress vs. stretch curve of the pristine kirigami sheet under a single cycle of load. **(I)** Images of the reinforced kirigami sheet under 1000^{th} cycle and under 3000^{th} cycle. **(J)** Comparison of fatigue thresholds and water contents among reported synthetic hydrogels (19-21, 39, 40), PVA hydrogels with patterned highly-crystalline regions and biological tissues (41). **(K)** Comparison of fatigue thresholds and Young's moduli among reported synthetic hydrogels, PVA hydrogels with patterned highly-crystalline regions and biological tissues. Data in **E** and **F** are means \pm SD, $n = 3$. Scale bars are $800 \mu\text{m}$ for **A**, 1 mm for **C**, and 40 mm for **I**.