

Manuscript Template

Anti-fatigue-fracture hydrogels by designing crystalline domains

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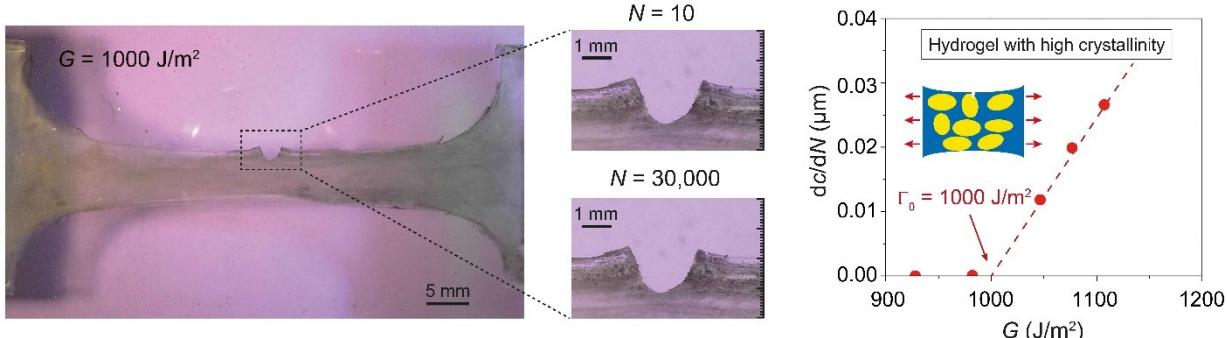
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One sentence summary: Designing nano-crystalline domains gives extremely anti-fatigue-fracture hydrogels for artificial cartilages and soft robots.



21 **Abstract**

22
23 The emerging applications of hydrogels in devices and machines require **these soft materials** to
24 maintain robustness under cyclic mechanical loads. Whereas hydrogels have been made tough to
25 resist fracture under a single cycle of mechanical load, these **toughened gels** still suffer from fatigue
26 fracture under multiple cycles of loads. The reported fatigue threshold (i.e., the minimal fracture
27 energy **at which** crack propagation **occurs** under cyclic loads) **for** synthetic hydrogels is on the order
28 of 1–100 J/m², which is **primarily** associated with the energy required to fracture a single layer of
29 polymer chains per unit area. Here, we demonstrate that the controlled introduction of crystallinity
30 in hydrogels can significantly enhance their fatigue thresholds, since the process of fracturing
31 crystalline domains for fatigue-crack propagation requires much higher energy than fracturing a
32 single layer of polymer chains. The fatigue threshold of polyvinyl alcohol (PVA) with a crystallinity
33 of 18.9 wt.% in the swollen state can exceed 1,000 J/m². We further develop a strategy to enhance
34 the anti-fatigue-fracture properties of PVA hydrogels, but still maintain their high water contents
35 and low moduli by patterning highly-crystalline regions in the hydrogels. The current work not only
36 reveals an anti-fatigue-fracture mechanism in hydrogels but also provides a practical method to
37 design anti-fatigue-fracture hydrogels for diverse applications.

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41 **Introduction**

42
43 As polymer networks infiltrated with water, hydrogels have been widely used as scaffolds for tissue
44 engineering (1), vehicles for drug delivery (2) and model platforms for biological studies (3). More
45 recently, hydrogels have been explored for applications in devices and machines, including
46 wearable electronics (4, 5), soft robotics (6), adhesives (7-9), and hydrogel-based soft machines
47 (10, 11). The use of hydrogels in devices and machines requires them to maintain robustness under
48 cyclic mechanical loads. Following the pioneering work by Gong et al. (12), hydrogels have been
49 made tough to resist crack propagation under a single cycle of mechanical load (13-16). The
50 toughening of hydrogels is achieved by integrating mechanisms for dissipating mechanical energy
51 such as fracture of short polymer chains and reversible crosslinks into stretchy polymer networks
52 (17, 18). However, existing tough hydrogels still suffer from fatigue fracture under multiple cycles
53 of mechanical loads (10, 19, 20), because the resistance to fatigue-crack propagation after
54 prolonged cycles of loads is the energy required to fracture a single layer of polymer chains (i.e.,
55 the intrinsic fracture energy of the hydrogel), which is unaffected by the additional dissipation
56 mechanisms introduced in tough hydrogels (19, 20). The reported fatigue thresholds of various
57 tough hydrogels are 8.4 J/m² for polyacrylamide (PAAm)-polyvinyl alcohol (PVA) (20) and 53.2
58 J/m² for PAAm-alginate (19), the same order as their intrinsic fracture energies. The highest fatigue
59 threshold for hydrogels reported so far is 418 J/m² for a double network hydrogel, poly(2-
60 acrylamido-2-methyl-1-propanesulfonic acid) (PAMPS)-PAAm, which possibly can be attributed
61 to the PAAm network with very long polymer chains and thus high intrinsic fracture energy (12,
62 21). A general strategy towards the design of anti-fatigue-fracture hydrogels has remained as a
63 critical need and a central challenge for long-term applications of hydrogels in devices and
64 machines.

65 In contrast to synthetic hydrogels, biological tissues such as cartilages, tendons, muscles

66 and heart valves show extraordinary anti-fatigue **properties**. For example, the knee joint of an

67 average person needs to sustain peak stresses of 4–9 MPa for 1 million cycles per year, and its
68 fracture energy after prolonged cycles of loads is above 1,000 J/m² (22, 23). The anti-fatigue
69 property of biological tissues possibly arises from the inherent highly ordered and partially
70 crystalline structures of collagen fibers in the tissues (24). Inspired by anti-fatigue biological
71 tissues, here we hypothesize that the increase of crystallinity in synthetic hydrogels can significantly
72 enhance their fatigue thresholds, due to the need to fracture crystalline domains for fatigue-crack
73 propagation (Fig. 1). The energy per unit area required to fracture crystalline domains of a polymer
74 is much higher than that required to fracture a single layer of amorphous chains of the same polymer
75 (25). To test the hypothesis, we select PVA as a model hydrogel with tunable crystallinity. We
76 increase the annealing time (after freeze-thawing and air-drying) of the PVA hydrogel to give
77 higher crystallinity, larger crystalline domain size, and smaller average distance between adjacent
78 domains in the hydrogel (Fig. 1A). We then measure the fatigue thresholds of PVA hydrogels with
79 various crystallinities (Fig. 1, B and C). We find that the increase of crystallinity can significantly
80 enhance the fatigue thresholds of PVA hydrogels. In particular, the fatigue threshold can exceed
81 1,000 J/m² when the crystallinity of PVA in the swollen state reaches 18.9 wt.%. By annealing
82 selected regions in hydrogels, we further demonstrate a general strategy to pattern highly-crystalline
83 regions in the PVA hydrogels to render them resistant to fatigue-fracture but still maintain their
84 high water content and low modulus. With this strategy, we create *kirigami hydrogel sheets* that are
85 highly stretchable and resistant to fatigue-fracture by introducing patterned cuts into hydrogel
86 sheets and then reinforcing the cut tips. The current work not only reveals a new anti-fatigue-
87 fracture mechanism in hydrogels but also provides a practical method to design anti-fatigue-fracture
88 hydrogels for diverse applications.

89 **Results**

90 To validate the hypothesis that the increase of crystallinity in hydrogels can significantly
91 enhance their fatigue thresholds, we use PVA hydrogels as a model material system with tunable

92 crystallinity. We first freeze a solution of uncrosslinked PVA at -20 °C for 8 h and thaw it at 25 °C
93 for 3 h, to form a hydrogel crosslinked by crystalline domains (26). The freeze-thawed PVA
94 hydrogel is further dried in an incubator at 37 °C and then annealed at 100 °C for various times
95 ranging from 0 min to 90 min (27). (The hydrogel dry-annealed for 0 min means no annealing
96 process.) The crystallinity of the hydrogel can be tuned by drying and annealing it **for** different
97 times. As a control sample, we also fabricate a chemically-crosslinked PVA as a reference hydrogel
98 **composed of an entirely** amorphous polymer network (See Materials and Methods for details on
99 fabrication of the samples).

100 ***Characterization of the crystalline morphology in PVA hydrogels***

101 We first measure the crystallinities of the resultant PVA hydrogels in their dry state by
102 differential scanning calorimetry (DSC). For the chemically-crosslinked and **freeze-thawed** PVA
103 hydrogels, we use excess chemical crosslinks to fix the amorphous polymer chains before air-drying
104 them in an incubator at 37 °C (see Materials and Methods for details). The excess chemical
105 crosslinks minimize the formation of further crystalline domains during **the** air-drying process (28,
106 29). As shown in Fig. 2A, both the chemically-crosslinked and freeze-thawed PVA hydrogels show
107 negligible **endothermic** peaks with measured crystallinities in the dry state of 0.2 wt.% and 2.1
108 wt.%, respectively. However, when the freeze-thawed PVA (without excess chemical crosslinks)
109 is dried in air, the crystallinity in the dry state increases to 37.7 wt.% (see Fig. 2A). The increase of
110 the crystallinity implies that substantially more crystalline domains nucleate during the air-drying
111 process (27). The crystallinity further increases gradually by increasing the annealing time. When
112 the sample is annealed for 90 min at 100 °C, the crystallinity in the dry state reaches 47.3 wt%. In
113 addition to the crystallinities in the dry state measured from DSC, we further measure the water
114 contents in the fully swollen samples shown in Fig. 2B and calculate the corresponding
115 crystallinities in the swollen state shown in Fig. 2C. It is well-known that the as-prepared dry PVA
116 samples may contain residual water bonded with polymer chains (29). The amount of residual water

117 can be calculated from the endothermic transition ranging from 60 °C to 180 °C on the DSC curves
118 (See Fig. 2A and fig. S1) (26). The above reported crystallinities in the dry state (without residual
119 water), water contents and crystallinities in the swollen state have been corrected to account for the
120 weights of residual water in the as-prepared dry samples (see Materials and Methods, table S1 and
121 fig. S1 for details).

122 To quantify the evolution of crystalline morphology, we measure the average distance
123 between adjacent crystalline domains L through small-angle X-ray scattering (SAXS) and the
124 average size of crystalline domains D through wide-angle X-ray scattering (WAXS) (see details in
125 Materials and Methods). We first perform SAXS measurements on the samples in the swollen state
126 after subtracting the water background, measuring the scattering intensity $I(q)$ versus the scattering
127 vector q . To identify the location of the peak intensity, we correct the intensity by multiplying the
128 scattering intensity with the scattering vector q (30). As shown in Fig. 2D, there is no peak in the
129 plot of the corrected intensity I_q versus the scattering vector q for the freeze-thawed hydrogel,
130 which implies negligible **interference** between adjacent crystalline domains. For the hydrogel dry-
131 annealed for 0 min, there is a slight **shoulder** shown in the corrected intensity curve, which indicates
132 stronger **interference** between adjacent domains. The average distance between adjacent crystalline
133 domains L can be estimated from the critical vector corresponding to the peak intensity q_{max} ,
134 following the Bragg expression $L = 2\pi / q_{max}$ (30). The average distance for the hydrogel dry-
135 annealed for 0 min is estimated to be 21 nm in the swollen state. As the annealing time increases to
136 90 min, the average distance L decreases to 13 nm in the swollen **state** (Fig. 2G). As a control case,
137 we also measure SAXS profiles of the hydrogel dry-annealed 90 min in the dry state. As shown in
138 Fig. 2F, the average distance between adjacent crystalline domains in the dry state is around 9 nm,
139 smaller than the distance in the same hydrogel in the swollen state (i.e., 13 nm). This is because
140 swelling of the interstitial amorphous polymer chains increases the distance between adjacent
141 crystalline domains.

142 We further perform WAXS measurements on the hydrogels in their dry state using Ni-
143 filtered **CuK α 1** radiation with X-ray wavelength $\lambda = 1.54$ Å. As shown in Fig. 2E, all dry-annealed
144 PVA hydrogels show a strong diffraction peak at $2\theta = 19.7$ °, which corresponds to the typical
145 reflection plane of $(10\bar{1})$ in semi-crystalline PVA (31). In addition, small peaks at $2\theta = 11.5$ ° and
146 23.1 ° are also observed in the hydrogel dry-annealed for 90 min, suggesting a high crystallinity in
147 the hydrogel, which is consistent with the DSC measurement. By identifying the half-width of the
148 maximum diffraction peak β , the average size of crystalline domains D can be approximately
149 calculated using Scherrer's equation $D = k\lambda / (\beta \cos \theta)$ (32), where k is a dimensionless shape
150 factor varying with the actual shape of the crystalline domain; λ is the wavelength of X-ray
151 diffraction; and θ is the Bragg angle. Here, β is identified after subtracting the instrumental line
152 broadening; and the dimensionless shape factor k is set as 1, approximating the spherical shape of
153 the crystalline domains (33). As shown in Fig. 2G, by increasing the annealing time from 0 min to
154 90 min, the average size of the crystalline domains increases from 3.8 nm to 6.5 nm. This trend is
155 consistent with the decrease of the average distance between adjacent crystalline domains with
156 annealing time, since the growth of the crystalline domains consumes the interstitial amorphous
157 polymer chains.

158 To further validate the tuning of crystalline domains in the PVA hydrogel with annealing
159 time, we use tapping-mode atomic force microscopy (AFM) to obtain phase images of the
160 hydrogels dry-annealed for 0 min and 90 min, respectively. The bright areas in Fig. 2H correspond
161 to the regions with **relatively** high modulus (mainly crystalline domains), whereas the dark areas
162 represent the regions with **relatively** low modulus (mainly amorphous domains). As shown in Fig.
163 2H, the morphology of isolated crystalline domains is observed in the hydrogel dry-annealed for 0
164 min, while the hydrogel dry-annealed for 90 min shows larger aggregated crystalline domains.

165 ***Characterization of fatigue fracture properties of hydrogels***

166 To measure the fatigue threshold of PVA hydrogels, we adopt the single-notch method,
167 which has been widely used in fatigue tests of rubbers (34, 35). Notably, all fatigue tests in this
168 study are performed on fully-swollen hydrogels immersed in a water bath, to prevent the
169 dehydration-induced crack propagation (fig. S2). We use **dogbone-shaped** samples and perform
170 cyclic tensile tests on both notched and unnotched samples, which are otherwise the same (Fig. 1,
171 B and C). The nominal stress versus stretch curves (i.e., S vs. λ) of unnotched samples are
172 obtained over N cycles of applied stretch λ^A . The strain energy density W of the unnotched sample
173 under the N_{th} cycle of applied stretch λ^A can be calculated as

$$174 \quad W(\lambda^A, N) = \int_1^{\lambda^A} S d\lambda \quad (1)$$

175 where S and λ are the measured nominal stress and stretch, respectively. Thereafter, the same
176 cyclic stretch λ^A is applied on the notched sample, measuring the evolution of the cut length in
177 undeformed state c with the cycle number N . The applied energy release rate G in the notched
178 sample under the N_{th} cycle of applied stretch λ^A can be calculated as (34, 35)

$$179 \quad G(\lambda^A, N) = 2k(\lambda^A) \cdot c(N) \cdot W(\lambda^A, N) \quad (2)$$

180 where k is a slowly varying function of the applied stretch expressed as $k = 3 / \sqrt{\lambda^A}$, c the current
181 crack length at undeformed configuration, and W the strain energy density measured in the
182 unnotched sample (Eq. 1). By systematically varying the applied stretch λ^A , we can obtain a plot
183 of crack extension per cycle versus the applied energy release rate (i.e., dc / dN vs. G). By linearly
184 extrapolating the curve of dc / dN vs. G to the intercept with the **abscissa**, we can approximately
185 obtain the critical energy release rate G_c , below which the fatigue crack will not propagate under
186 infinite cycles of loads. By **definition**, the fatigue threshold Γ_0 is equal to the critical energy release
187 rate G_c . To validate that this extrapolated value G_c is indeed the fatigue threshold Γ_0 , we further
188 apply G_c to the notched sample over 30,000 cycles (to approximate infinite cycles of loads) and

189 observe no crack extension (Fig. 3H). In addition, we also measure the fatigue thresholds using the
190 pure-shear test (19) to validate the results from the single-notch test (fig. S6).

191 For cyclic tensile tests on unnotched samples (Fig. 3, A and B and fig. S3), both chemically-
192 crosslinked PVA and freeze-thawed PVA show negligible Mullins effect, and their S vs. λ curves
193 reach steady states after only a few cycles (i.e., 10 for chemically-crosslinked PVA at an applied
194 stretch of 1.6, and 200 for freeze-thawed PVA at an applied stretch of 2.2). On the other hand, the
195 dry-annealed PVA hydrogels exhibit a more significant Mullins effect, due to mechanical
196 dissipation caused by melting and reorientation of crystalline domains (30). As the stretch further
197 increases, the crystalline domains may transform into aligned fibrils along the loading direction
198 (30). The energy required to damage the crystalline domains and fibrils is much higher than that to
199 fracture a single layer of the same polymer in amorphous state. The hydrogel dry-annealed 90 min
200 reaches steady state after 1,000 cycles of applied stretches of $\lambda^A = 2$ (Fig. 3C). Despite the Mullins
201 effect, the steady-state maximum nominal stress of the hydrogel dry-annealed for 90 min is much
202 higher than that of both the chemically-crosslinked PVA and the freeze-thawed PVA at the same
203 applied stretch of $\lambda^A = 2$ (e.g., 2.3 MPa for hydrogel dry-annealed 90 min, 15 kPa for chemically-
204 crosslinked PVA, and 3 kPa for freeze-thawed PVA in fig. S4).

205 For cyclic tensile tests on notched samples, a pre-crack is cut using a razor blade with tip
206 radius of **around 200** μm and initial crack length around 1 mm, smaller than one-fifth of the width
207 of the sample (35). A digital microscope (AM4815ZT, Dino-Lite, resolution 20 $\mu\text{m}/\text{pixel}$) is used
208 to record the cut length under cyclic loads (Materials and Methods, fig. S5). We first apply cyclic
209 loads with a small applied stretch (i.e., $\lambda^A = 1.3$) on a notched sample. If the crack remains quasi-
210 stationary with crack extension per cycle (dc/dN) smaller than 20 nm/cycle (i.e., no detectable
211 crack extension in 1,000 cycles), the applied cyclic stretch is increased by the increment of
212 $\Delta\lambda^A = 0.1$ for other notched samples until crack propagation greater than 20 nm/cycle is captured.
213 As shown in Fig. 3, D and E, the fatigue thresholds of chemically-crosslinked PVA and freeze-

214 thawed PVA are measured to be 10 J/m^2 and 23 J/m^2 , respectively. The fatigue threshold for the
215 hydrogel dry-annealed 0 min increases to 110 J/m^2 . As the annealing time increases, the fatigue
216 threshold further increases. In particular, for the hydrogel dry-annealed 90 min, the fatigue
217 threshold can achieve 1000 J/m^2 (Fig. 3F and fig. S5). The measured fatigue threshold of the
218 hydrogel dry-annealed 90 min from the pure-shear test is 918 J/m^2 , consistent with the single-notch
219 test (fig. S6). The dependence of fatigue threshold on the crystallinity is summarized in Fig. 3G.
220 The fatigue threshold increases with the crystallinity and demonstrates a sharp jump when the
221 crystallinity in the swollen state reaches approximate 15 wt.%.

222 In addition to fatigue tests, we also measure the nominal stress versus stretch curves of all
223 hydrogels to obtain their Young's moduli and tensile strengths (fig. S7). As shown in Fig. 4, A and
224 B, both the Young's modulus and tensile strength increase with the hydrogels' crystallinity and
225 show dramatic enhancements when the crystallinity in the swollen state reaches approximate 15
226 wt.% (36). This sharp jump in Young's modulus and tensile strength is consistent with the dramatic
227 increase of fatigue threshold of the hydrogel at approximate 15 wt.% crystallinity in the swollen
228 state (Fig. 3G).

229

230 *Patterning of highly-crystalline regions in hydrogels*

231 Whereas annealing the whole PVA hydrogel can significantly enhance its fatigue threshold, the
232 annealing treatment also increases the modulus and decreases the water content of the hydrogel
233 (Fig. 4C and fig. S8). However, for many applications, it is desirable to maintain the relatively low
234 modulus and high water content of the hydrogels. Here we propose a strategy to introduce
235 programed highly-crystalline regions in the hydrogels. We use the computer-aided design of
236 electrical circuits to induce localized heat treatment for annealing selected regions of the hydrogels
237 (See details in Materials and Methods and fig. S9). The chemically-crosslinked PVA hydrogel is
238 used as the pristine sample with low fatigue threshold of 15 J/m^2 , low Young's modulus of 114 kPa

239 and high water content of 88 wt.%. Three examples of programed annealing patterns on the pristine
240 PVA hydrogels are demonstrated with enhanced fatigue thresholds.

241 We first locally introduce a highly-crystalline ring-shaped region around a crack tip (Fig.
242 5A), leading to a fatigue threshold over 236 J/m² (Fig. 5B). Despite the small area of the highly-
243 crystalline region, this local conditioning can greatly reinforce the crack tip, delaying crack
244 propagation. Meanwhile, the measured overall Young's modulus and water content of the sample
245 are maintained at 114 kPa and 87 wt.% (Fig. 5, E and F), respectively, which are almost unaffected
246 by the local annealing around the crack tip. As a second example, we pattern mesh-like highly-
247 crystalline regions on the pristine hydrogel (Fig. 5C). Compared with the pristine sample, the
248 fatigue threshold of the mesh-reinforced sample increases to 290 J/m² (Fig. 5D), and its Young's
249 modulus remains relativiely low (627 kPa) and its water content stays relatively high as 83 wt.%
250 (Fig. 5, E and F).

251 The strategy of patterning highly-crystalline regions can be applied to various structures of
252 hydrogels for improving anti-fatigue performances as well. For example, the kirigami structure is
253 commonly adopted to enhance the stretchablity (37) and effective adhesion (38) of films. As a third
254 example, we demonstrate a kirigami hydrogel sheet with improved anti-fatigue performance by
255 patterning highly-crystalline regions around the cut tips. As shown in Fig. 5G, we first generate a
256 kirigami pattern on a pristine sample by introducing parallel periodic cuts with equal length and
257 equal distance bewteen adjacent cuts. The ultimate stretch and effective nominal stress (i.e., force
258 divided by cross-section area) of the pristine kirigami hydrogel sheet under a single cycle of load
259 are measured to be 1.7 and 0.9 kPa, respectively. Thereafter, the pristine kirigami sheet is reinforced
260 around the cut tips with local annealing. The reinforced kirigami sheet can sustain cyclic tensile
261 loads at an applied stretch of $\lambda^A = 2.1$ without detectable fatigue-fracture even when the cycle
262 number approaches 3,000. Moreover, the effective nominal stress after prolonged cycles at the

263 applied stretch of $\lambda^A = 2.1$ can maintain the plateau of 13 kPa, which is 14.4 times the strength of
264 the pristine kirigami sheet (Fig.5, H and I).

265 Figure 5J and K compare the fatigue thresholds, water contents and the Young's moduli of
266 reported hydrogels in literature (19-21, 39, 40). We show that by patterning highly-crystalline
267 regions, both tip-reinforced and mesh-reinforced PVA hydrogels outperform existing synthetic
268 hydrogels in terms of **fatigue thresholds**, and they can still maintain relatively high water contents
269 and relatively low Young's moduli.

270

271 **Discussion**

272 We **have demonstrated** that the fatigue threshold of hydrogels can be greatly enhanced by designing
273 crystalline domains in the hydrogels. We use PVA hydrogels as a model material to validate this
274 new mechanism for designing anti-fatigue-fracture hydrogels. The fatigue threshold of a PVA
275 hydrogel with a crystallinity of 18.9 wt.% in the swollen state can achieve over 1000 J/m². We
276 further develop a strategy to make PVA hydrogels anti-fatigue-fracture but still maintain their high
277 water content and low modulus by patterning highly-crystalline regions in the hydrogels. The
278 reported mechanism and strategy for designing anti-fatigue-fracture hydrogels can be extended to
279 hydrogel composites with fillers such as nanoclay, nanocellulose and nanofibers.

280 The capability to enhance the anti-fatigue-fracture performance of synthetic hydrogels
281 makes a number of future research directions and applications possible. For example, anti-fatigue
282 hydrogels can be used for hydrogel-based gastric-retentive devices and implantable tissue
283 replacements of meniscus, intervertebral disk and cartilage, **which require** long-term mechanical
284 robustness when interacting with the human body.

285

286 **Materials and Methods**

287 **Synthesis of PVA hydrogels**

288 All types of our PVA hydrogels are synthesized from 10 wt.% poly(vinyl alcohol) (PVA; Mw
289 146,000-186,000, 99+% hydrolyzed; Sigma-Aldrich, 363065) solution. The solution was heated in
290 a water bath at 100 °C with stirring for 5 h. To synthesize chemically-crosslinked PVA hydrogels,
291 we added 10 μ L glutaraldehyde (25 vol.%, Sigma-Aldrich, G6257) as a crosslinker to 1 mL 10
292 wt.% PVA solution, and added 10 μ L hydrochloric acid (36.5-38 wt.%, J.T. Baker, 9535-02) as an
293 accelerator into the other 1 mL 10 wt.% PVA solution. We then mixed and defoamed each of them
294 by using a centrifugal mixer (AR-100; Thinky). The final mixtures, obtained by mixing and
295 defoaming the two solutions together, were then cast into a mold and allowed to cure for 2 h. The
296 chemically-crosslinked PVA hydrogels were immersed in deionized water for two days to remove
297 unreacted chemicals. To fabricate freeze-thawed PVA hydrogels, 10 wt.% PVA solutions after
298 mixing and defoaming were poured into a mold, frozen at -20 °C for 8 h and thawed at 25 °C for 3
299 h. The freeze-thawed hydrogels were further dried in an incubator (New Brunswick Scientific, C25)
300 at 37 °C for 2 h, and then annealed at 100 °C for a controlled time (i.e., 0, 1, 3, 5, 10, or 90 min).
301 All as-prepared PVA hydrogels were immersed in water to achieve their equilibrium-swollen state.
302

303 **Measurement of residual water and crystallinity in dry samples**

304 We measured the crystallinities of the resultant PVA hydrogels in their dry state by differential
305 scanning calorimetry (DSC/cell: RCS1-3277 Cooling System: DSC1-0107). For as-prepared
306 chemically-crosslinked PVA and freeze-thawed PVA, we used excess chemical crosslinks to fix
307 the amorphous polymer chains before air-drying, minimizing the formation of further crystalline
308 domains during the air-drying process. We first soaked the samples (thickness of 1mm) in the
309 aqueous solution consisting of 10 mL glutaraldehyde (25 vol.%, Sigma-Aldrich, G6257), 500 μ L
310 hydrochloric acid (36.5-38 wt.%, J.T. Baker, 9535-02) and 50 mL DI water for 2 h. Thereafter, we
311 soaked the samples in a DI water bath for 2 h to remove the residual hydrochloric acid. The samples
312 were further dried in an incubator (New Brunswick Scientific, C25) at 37 °C for 2 h.

313 Thereafter, we measured the mass of residual water $m_{residual}$, the mass of crystalline domains
 314 $m_{crystalline}$ and the total mass of the dry samples (with residual water) m using DSC. In a typical DSC
 315 measurement, we first weighed the total mass of the dry sample (with residual water) m . The
 316 sample were thereafter placed in a Tzero-pan and heated up from 50 °C to 250 °C at the rate of 20
 317 °C/min under a nitrogen atmosphere with flow rate of 30 mL/min. The curve of heat flow shows a
 318 broad peak from 60 °C to 180 °C, indicating that the sample contains a small amount of residual
 319 water. The integration of the endothermic transition ranging from 60 °C to 180 °C gives the
 320 enthalpy for evaporation of the residual water per unit mass of the dry sample (with residual water)
 321 $H_{residual}$. Therefore, the mass of the residual water $m_{residual}$ can be calculated as

$$322 \quad m_{residual} = m \cdot \frac{H_{residual}}{H_{water}^0} \quad (3)$$

323 where $H_{water}^0 = 2260 \text{ J/g}$ is the latent heat of water evaporation. The curve of heat flow shows
 324 another narrow peak ranging from 200 °C to 250 °C corresponding to melting of the crystalline
 325 domains. The integration of the endothermic transition ranging from 200 °C to 250 °C gives the
 326 enthalpy for **melting** the crystalline domains per unit mass of the dry sample (with residual water)
 327 $H_{crystalline}$. Therefore, the mass of the crystalline domains $m_{crystalline}$ can be calculated as

$$328 \quad m_{crystalline} = m \cdot \frac{H_{crystalline}}{H_{crystalline}^0} \quad (4)$$

329 where $H_{crystalline}^0 = 138.6 \text{ J/g}$ is the enthalpy of fusion of 100 wt.% crystalline PVA measured at the
 330 equilibrium melting point T_m^0 (29). Therefore, the crystallinity in the ideally dry sample X_{dry}
 331 (without residual water) can be calculated as

$$332 \quad X_{dry} = \frac{m_{crystalline}}{m - m_{residual}} \quad (5)$$

333

334 **Measurement of water content and crystallinity in swollen samples**

335 The swollen hydrogels weighing $m_{swollen}$ were placed in an incubator (New Brunswick Scientific,
336 C25) at 37 °C for 2 h, and weighed m after air-drying. The mass of the residual water in the as-
337 prepared dry samples $m_{residual}$ was measured in the previous section. Therefore, the water content in
338 the swollen state can be calculated as $(m_{swollen} - m + m_{residual}) / m_{swollen}$ and the polymer content in
339 the swollen state can be calculated as $(m - m_{residual}) / m_{swollen}$. In addition, the crystallinity in the
340 swollen sample $X_{swollen}$ can be calculated as

$$341 X_{swollen} = \frac{m_{crystalline}}{m_{swollen}} = X_{dry} \cdot \frac{m - m_{residual}}{m_{swollen}} \quad (6)$$

342 where $m_{crystalline}$ and X_{dry} were measured as described in the previous section.

343 AFM phase imaging

344 AFM phase images were acquired by atomic force microscope (MFP-3D, Asylum Research) in
345 tapping mode. Dry free-standing PVA films were directly attached onto sample stage by double-
346 sided carbon tape. The probe lightly taps on the sample surface with recorded phase shift angle of
347 the probe motion relative to a driving oscillator. The bright regions with high phase angle
348 correspond to regions with **relatively** high modulus; and the dark regions with low phase angle
349 correspond to regions with **relatively** low modulus.

350 X-ray scattering

351 The X-ray scattering measurement was performed with Pilatus3R 300K detector Bruker Nanostar
352 SAXS in X-ray Diffraction Shared Experimental Facility at MIT. We used small angle 2 mm
353 beamstop with sample-detector distance of 1059.1 mm for SAXS measurements and wide angle
354 2mm beamstop with sample-detector distance of 109.1 mm for WAXS measurements. The
355 exposure time was set as 300 s. Raw SAXS and WAXS patterns were processed with **corrections**
356 **by** MATLAB-based GIXGUI software before analysis.

357 Raman spectroscopy

358 Each sample was hydrated for more than 2 h and then pressed between a glass slide and a cover
359 slip in order to ensure a flat surface. The cover slip was then sealed at the edges with nail polish to
360 prevent the hydrogel from drying. A confocal Raman microscope (Alpha300RA; WiTec, Germany)
361 with 20 \times objective (Zeiss, Germany) was used. Nd:YAG laser (532 nm) was used as the excitation
362 source with the maximum power of 75mW. Data was collected with a CCD detector (DU401A-
363 BV; Andor, UK) behind a 600 g/mm grating spectrometer (UHTS 300; WiTec, Germany). A 20-
364 μm resolution Raman map of 4 \times 3 mm scan area was acquired with an accumulation time of 1
365 second per point. Each point was pre-bleached for 400 ms to decrease the effect of fluorescence.
366 Cosmic ray removal and background subtraction were performed to clean the spectra. The intensity
367 of O-H bond within the PVA and water were calculated by integrating the spectra in the range of
368 2,800-3,000 cm^{-1} and 3,075-3,625 cm^{-1} , respectively. The ratio of PVA and water was then
369 calculated and plotted as a heatmap shown in Fig. 5A.

370 **Measurement of the fatigue threshold**

371 All the mechanical tests were performed in a water bath at 25°C with a U-stretch testing device
372 (CellScale, Canada). For mechanically weak samples (e.g., the Ch and FT hydrogels), a load cell
373 with maximum force of 4.4 N was used; for mechanically strong samples (e.g., hydrogel dry-
374 annealed 90 min), a load cell with maximum force of 44 N was used. The dogbone-shaped sample
375 had the dimensions at as-prepared state with the width of 5mm, thickness of 0.8mm and gauge
376 length of 10mm. The nominal stress S was measured from the recorded force F divided by width
377 W and thickness t in the swollen state. To measure the applied stretch λ^A in gauge length, we
378 adopted the digital image correlation method and calibrated the correlation between λ^A and loading
379 distance d at gripping points (fig. S2). A digital microscope (AM4815ZT; Dino-Lite, resolution 20
380 $\mu\text{m}/\text{pixel}$) was used to record the crack extension. Since dry annealed PVA is transparent when
381 immersing in water, we spread a small amount of graphite powder on the surface of the sample for
382 visualization.

383 **Preparation of samples with programmable crystalline domains**

384 A copper tape was first loaded in a paper cutting machine (Silhouette CAMEO 3). The electrical
385 circuits were first designed with CAD and loaded to the machine via Silhouette Studio software.
386 After completion of the cutting, the copper tape was placed on a flat acrylic plate and unwanted
387 parts were carefully peeled off. Two additional conductive wires were soldered on the copper tape
388 as two electrodes. An electrical generator (Insteek PSB-2400L2; Tequipment) was used to apply
389 controlled current around 2-5 A on the designed circuits. To ensure achieving the designed
390 temperature, a thermal imager (Seek thermal XR imager) was first used to measure the local
391 temperature from the circuit (see [fig. S9](#)). By adjusting the current to control the designed
392 temperature, the air-dried PVA hydrogel was placed on the targeted circuits for local heating. A
393 gripper was used to ensure fully contact between the PVA film and copper circuits.

394 **Supplementary Material**

395 [table S1. Crystallinities and water contents in chemically-crosslinked \(Ch\), freeze-thawed \(FT\)](#)
396 [and dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min.](#)

397 [fig. S1. Measurement of the mass of freeze-thawed PVA during air-drying and the amount of](#)
398 [residual water in the sample after air-drying.](#)

399 [fig. S2. Experiment method for measuring fatigue threshold.](#)

400 [fig. S3. Shakedown softening of three types of PVA hydrogels.](#)

401 [fig. S4. Steady-state nominal stress versus stretch curves of PVA hydrogels with various](#)
402 [crystallinities.](#)

403 [fig. S5. Validation of high fatigue threshold with single-notch test.](#)

404 [fig. S6. Validation of high fatigue threshold with pure shear test.](#)

405 [fig. S7. Mechanical characterization of PVA hydrogels with various crystallinities.](#)

406 [fig. S8. Raman spectroscopy of freeze-thawed PVA and 90-min dry-annealed PVA.](#)

407 fig. S9. Electrical circuit and thermal mapping for programmable crystalline domains.

408 movie S1. **Tension of the pristine notched sample and the tip-reinforced sample.**

409 movie S2. Cyclic loading of the reinforced kirigami hydrogel sheet.

410

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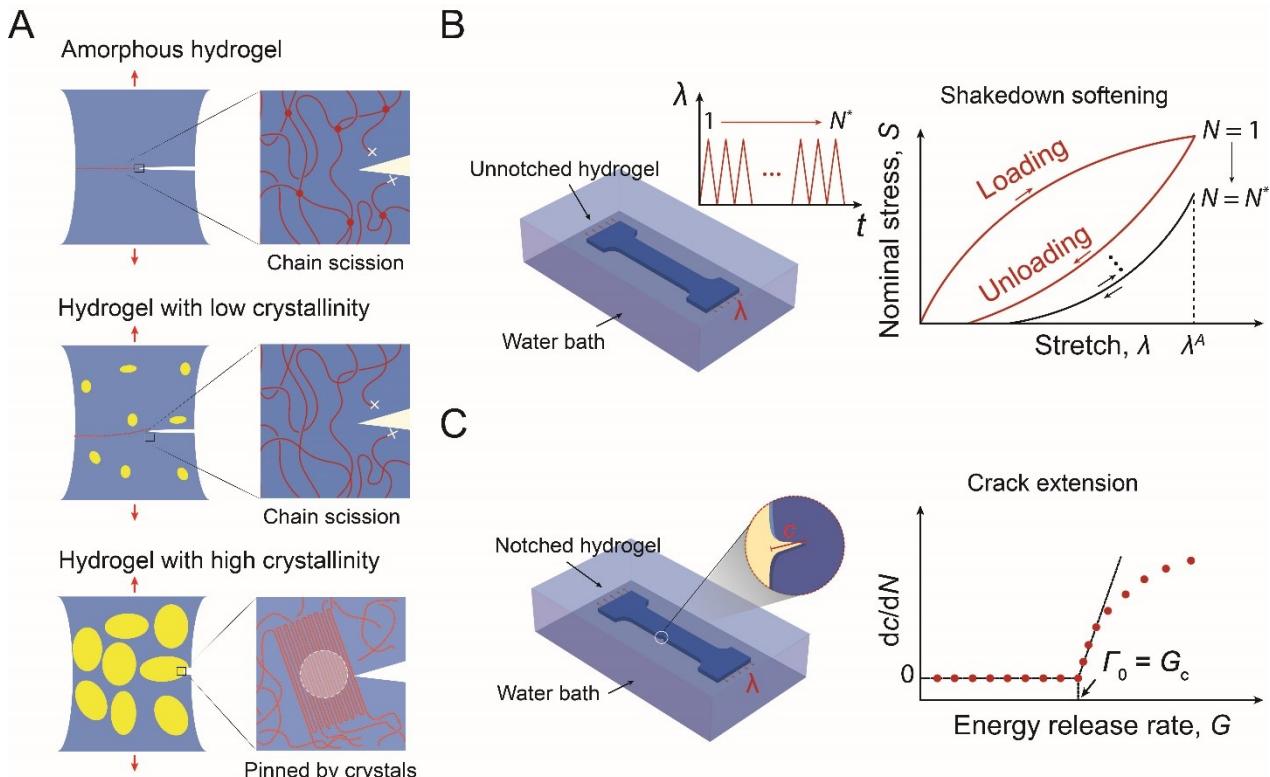
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517 results. H.L. performed Raman spectroscopy. S.L., X.L., H.Y. and J.L. performed the
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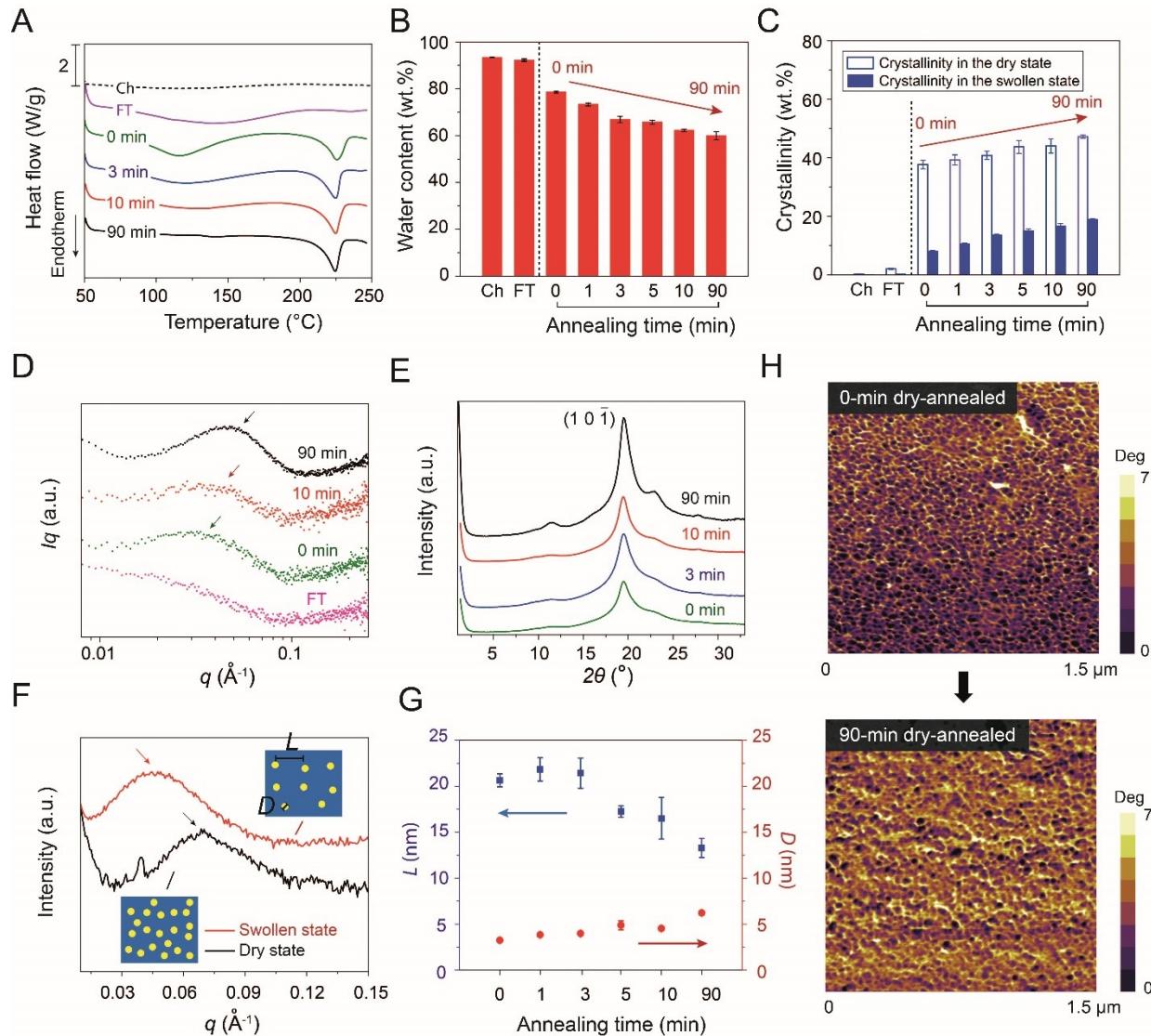
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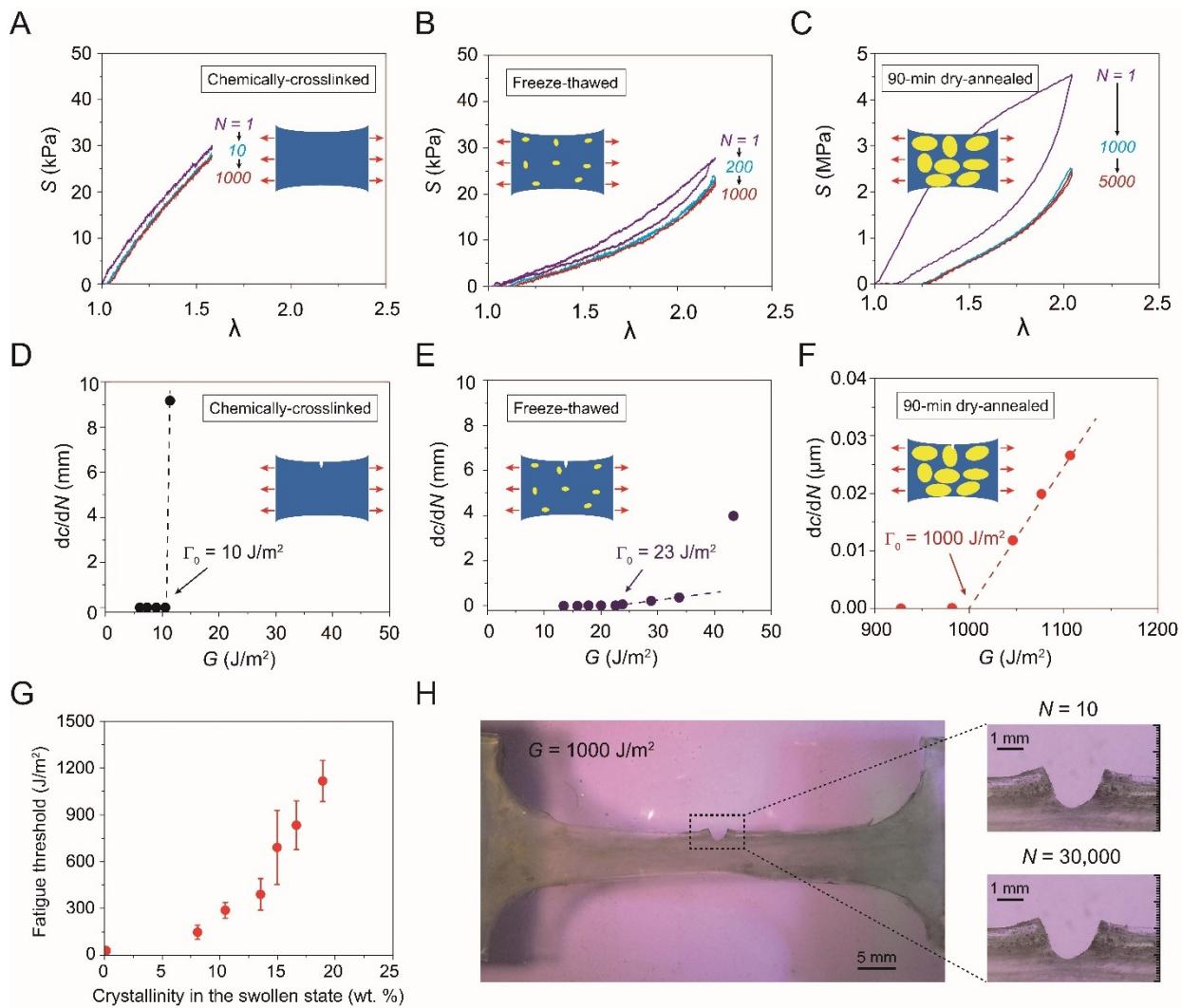
Figures and Tables



529 **Fig. 1. The design strategy for anti-fatigue-fracture hydrogels.** (A) Illustration of fatigue-crack
 530 propagation in an amorphous hydrogel and in hydrogels with low and high crystallinities under
 531 cyclic loads. The yellow areas represent crystalline domains and the blue areas denote amorphous
 532 domains. In the amorphous hydrogel and the hydrogel with low crystallinity, the fatigue threshold
 533 can be attributed to the energy required to fracture a single layer of polymer chains per unit area. In
 534 the hydrogel with high crystallinity, the fatigue-crack propagation requires fracture of crystalline
 535 domains. (B) Illustration of measuring nominal stress S vs. stretch λ curves over N cycles of the
 536 applied stretch λ^A . The stress-stretch curve reaches steady state as N reaches a critical value N^* . (C)
 537 Illustration of measuring crack extension per cycle dc/dN vs. energy release rate G curves. By
 538 linearly extrapolating the curve to intercept with the abscissa, we can approximately obtain the
 539 critical energy release rate G_c , below which the fatigue crack will not propagate under infinite cycles
 540 of loads. By definition, the fatigue threshold Γ_0 is equal to the critical energy release rate G_c .

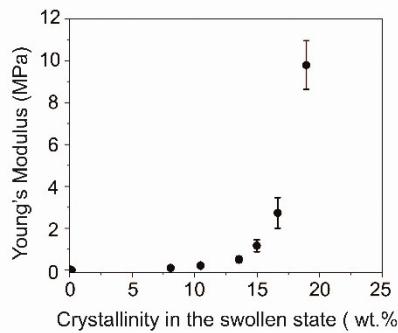


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545 **Fig. 2. Characterization of crystalline domains in PVA hydrogels.** (A) Representative DSC
546 thermographs of chemically-crosslinked (Ch), freeze-thawed (FT) and dry-annealed PVA with
547 annealing time of 0, 3, 10, 90 min. (B) Water contents of chemically-crosslinked (Ch), freeze-
548 thawed (FT) and dry-annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min. (C) Measured
549 crystallinity in the dry and swollen states of Ch, FT and dry-annealed PVA with annealing time of
550 0, 1, 3, 5, 10, 90 min. (D) Representative SAXS profiles of FT hydrogel and dry-annealed PVA
551 with annealing time of 0, 10, and 90 min. (E) Representative WAXS profiles of annealed PVA with
552 annealing time of 0, 3, 10, and 90 min. (F) SAXS profiles of 90-min dry-annealed PVA in the dry
553 state and the swollen state. The insets illustrate the increase of the distance between adjacent
554 crystalline domains due to swelling of amorphous polymer chains. (G) The estimated average
555 distance between adjacent crystalline domains *L* and average crystalline domain size *D* of dry-
556 annealed PVA with annealing time of 0, 1, 3, 5, 10, 90 min. (H) AFM phase images of dry-annealed
557 hydrogel with annealing time of 0 min and 90 min. Data in B, C and G are means \pm SD, n = 3.

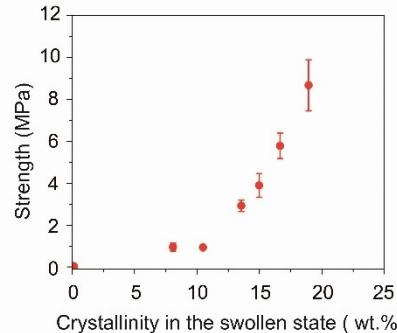


563 **Fig. 3. Measurement of fatigue thresholds of PVA hydrogels.** Nominal stress S vs. stretch λ
564 curves over cyclic loads for (A) chemically-crosslinked hydrogel at an applied stretch of $\lambda^A = 1.6$,
565 (B) freeze-thawed hydrogel at an applied stretch of $\lambda^A = 2.2$, and (C) 90-min dry-annealed
566 hydrogel at an applied stretch of $\lambda^A = 2.0$. Crack extension per cycle dc/dN vs. applied energy
567 release rate G for (D) chemically-crosslinked hydrogel, (E) freeze-thawed hydrogel, and (F) dry-
568 annealed hydrogel with annealing time of 90 min. (G) The fatigue threshold increases with the
569 crystallinity of the hydrogel in the swollen state. (H) Validation of fatigue threshold as high as 1000
570 J/m^2 in 90-min dry-annealed hydrogel using the single-notch test. Data in G are means \pm SD, $n =$
571 3. Scale bars are 5 mm and 1 mm for left and right images in H.

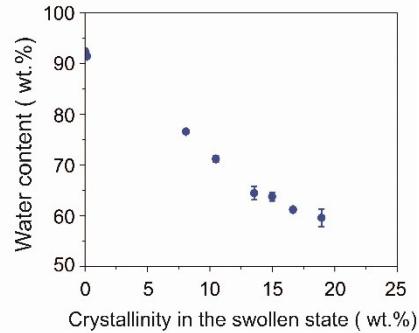
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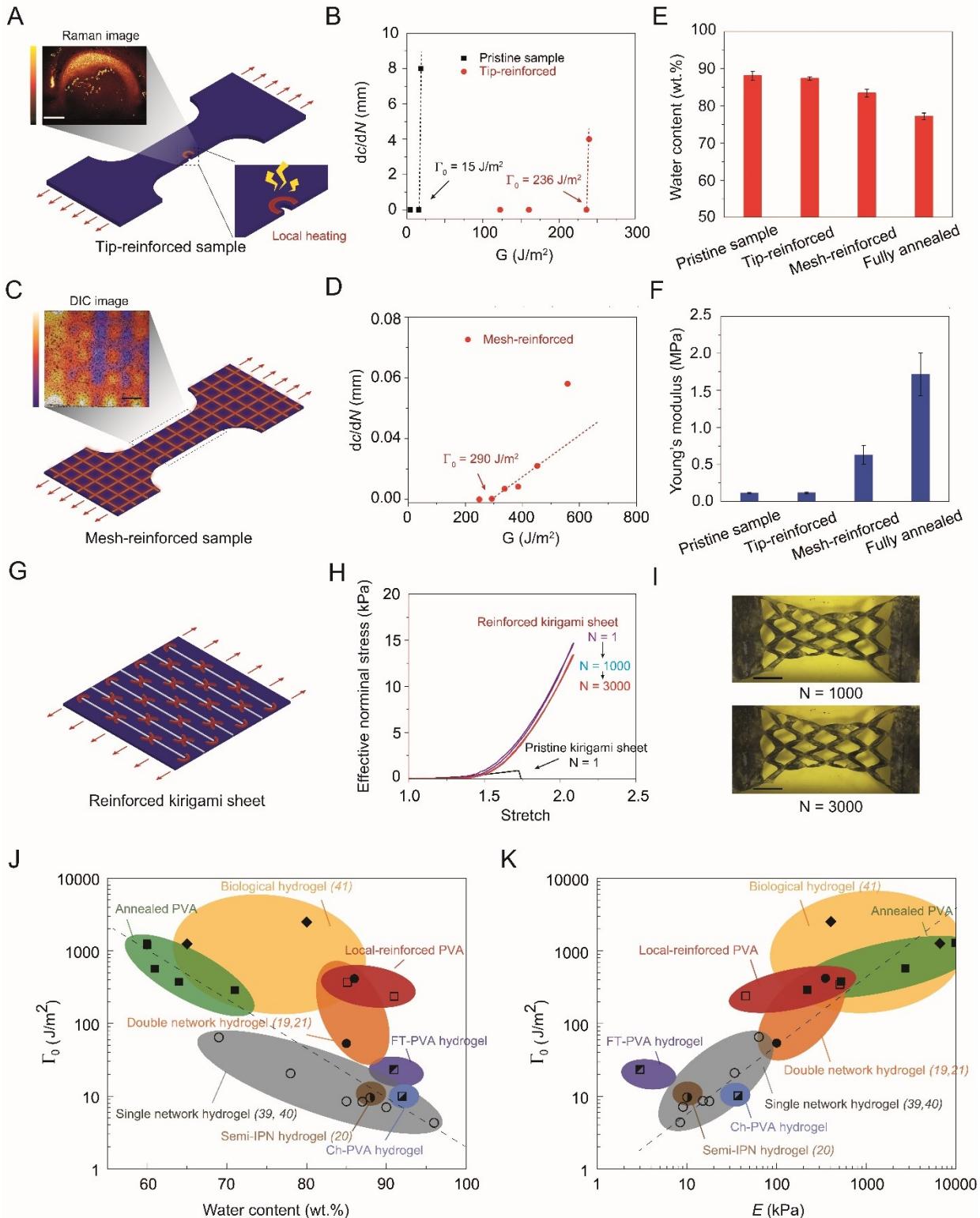


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Fig. 4. Young's moduli, tensile strengths and water contents of PVA hydrogels. (A) Young's
575 moduli versus crystallinity in the swollen state. **(B)** Tensile strength versus crystallinity in the
576 swollen state. **(C)** Water content versus crystallinity in the swollen state. Data in **A**, **B** and **C** are
577 means \pm SD, $n = 3$.
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582 **Fig. 5. Patterning highly-crystalline regions in PVA hydrogels.** (A) Illustration of introducing a
583 highly-crystalline region around crack tip. Inset: Raman spectroscopy with bright color representing
584 low water content and dark color representing high water content (see details in Materials and
585 Methods). (B) Comparison of crack extension per cycle dc/dN vs. applied energy releasing rate G
586 between the pristine sample and the tip-reinforced sample. The fatigue thresholds of the pristine
587 sample and the tip-reinforced sample are 15 J/m^2 and 236 J/m^2 , respectively. (C) Illustration of
588 introducing mesh-like highly-crystalline regions. Inset: Digital image correlation method (DIC)

589 shows large deformation in low-crystallinity regions and small deformation in high-crystallinity
590 regions. **(D)** Crack extension per cycle dc/dN vs. applied energy releasing rate G of the mesh-
591 reinforced sample. The fatigue threshold of the mesh-reinforced sample is 290 J/m^2 . **(E)** Water
592 contents of the pristine sample, the tip-reinforced sample, the mesh-reinforced sample and the fully-
593 annealed PVA hydrogels. **(F)** Young's moduli of the pristine sample, the tip-reinforced sample, the
594 mesh-reinforced sample and the fully-annealed PVA hydrogels. **(G)** Illustration of introducing
595 highly-crystalline regions around cut tips in a pristine kirigami sheet. **(H)** Effective nominal stress
596 vs. stretch curves of the reinforced kirigami sheet under cyclic loading. Effective nominal stress vs.
597 stretch curve of the pristine kirigami sheet under a single cycle of load. **(I)** Images of the reinforced
598 kirigami sheet under 1000^{th} cycle and under 3000^{th} cycle. **(J)** Comparison of fatigue thresholds and
599 water contents among reported synthetic hydrogels (19-21, 39, 40), PVA hydrogels with patterned
600 highly-crystalline regions and biological tissues (41). **(K)** Comparison of fatigue thresholds and
601 Young's moduli among reported synthetic hydrogels, PVA hydrogels with patterned highly-
602 crystalline regions and biological tissues. Data in **E** and **F** are means \pm SD, $n = 3$. Scale bars are
603 $800 \mu\text{m}$ for **A**, 1 mm for **C**, and 40 mm for **I**.