

1 Title: Seagrass restoration reestablishes the coastal nitrogen filter through enhanced burial

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Abstract

Seagrass meadows perform an important ecological function as filters for incoming nutrients from surrounding watersheds, especially nitrogen (N). By enhancing N removal processes, including N burial in sediments and denitrification, seagrass meadows improve water quality. With accelerating losses of seagrass meadows worldwide, seagrass restoration plays a key role in reestablishing these coastal ecosystem functions. However, few measurements exist of N burial rates in temperate seagrass meadows and none have been published for restored meadows. In this study, we measured N burial rates in a large (6.9 km²) restored eelgrass (*Zostera marina*) meadow and compared N removal through burial to previous measurements of removal via denitrification. We also compared N removal to inputs from external loading and fixation and to N assimilation in seagrass biomass. We found that, in this meadow, burial was the dominant process of N removal; the burial rate of 3.52 g N m⁻² y⁻¹ was comparable to rates in natural meadows within 10 years after seeding and was more than 20x the rate in adjacent bare sediments (0.17 g N m⁻² y⁻¹). We also found that the high rates of N assimilation (2.62 g N m⁻² y⁻¹) created a substantial though temporary sink for nitrogen during the growing season. Our results highlight how seagrass meadows mediate N cycling through high rates of burial, which to date has been understudied in the literature. The successful return of the N filter function after restoration, shown here for the first time, can motivate continued efforts for seagrass restoration and conservation.

Introduction

Seagrass meadows are highly productive ecosystems that provide a wide array of ecosystem services, ranging from supporting tourism and recreation to regulating climate via carbon sequestration (Barbier et al. 2011). Through their high productivity, seagrass can strongly influence the nitrogen (N) cycle in coastal waters. Large amounts of N are assimilated into seagrass biomass, which turns over slowly compared to algal biomass, leading to temporary retention of nitrogen (on the order of weeks to months); retention of N in seagrass reduces N availability to ephemeral algae species (Banta et al. 2004; McGlathery et al. 2007). Seagrass meadows can also remove nitrogen from coastal waters by enhancing two removal processes, burial and denitrification, in seagrass sediments. Seagrass increase N burial both by the physical processes of increased sedimentation (due to wave and current attenuation by the seagrass canopy) and sediment stabilization by roots and rhizomes (van der Heide et al. 2011; Hansen and Reidenbach 2012), and by the biological process of accumulation of recalcitrant seagrass tissue in the sediment (Middelburg et al. 2004). N can remain buried in seagrass sediments over long time scales (from decades to centuries) (Mateo et al. 1997). Seagrass roots also exude oxygen and labile carbon into the sediment, which can stimulate coupled nitrification-denitrification, leading to permanent N removal through the microbial transformation of biologically available nitrate into inert dinitrogen gas (Iizumi et al. 1980; Aoki and McGlathery 2018). Taken together, these retention and removal processes form a ‘filter’ for nitrogen that increases coastal water quality and slows the movement of nitrogen inputs from watersheds to the open ocean.

The seagrass filter function promotes a positive feedback for seagrass growth. By reducing particulates in the water column (i.e. through wave attenuation and sediment stabilization), the presence of the meadow can increase light availability deeper in the water

column, supporting greater seagrass growth (Carr et al. 2010). In addition, uptake of dissolved N into seagrass biomass limits N availability for algal growth that can shade the seagrass (Gurbisz et al. 2017). This positive feedback highlights the role of seagrass as an ecosystem engineer that modifies its habitat to better suit its needs, creating a stabilizing effect that allows the seagrass ecosystem to flourish. The restoration of the N filter function to levels observed in natural systems may therefore be a useful measure of the overall success of a seagrass restoration.

While some N cycling processes in seagrass meadows have been widely studied, our understanding of N removal as an ecosystem function remains incomplete. Previous measurements of N removal in seagrass meadows have focused primarily on denitrification. Recent studies have generally found elevated rates of denitrification in seagrass meadows compared to other estuarine habitats (Eyre et al. 2011, 2013; Smyth et al. 2013), but in some cases, rates were equivalent between seagrass and unvegetated sediments (Russell et al. 2016; Zarnoch et al. 2017). In contrast, measurements of N burial in seagrass meadows are few and focus mainly on peat-forming *Posidonia* species (Romero et al. 1994; Mateo and Romero 1997; Gacia et al. 2002, see Table 1). However, a recent study by Eyre et al. (2016) found elevated rates of both denitrification and N burial within *Zostera* seagrass communities relative to subtidal flats in three sub-tropical lagoons; the seagrass community N removal rates contributed significantly to lagoon-wide N removal. Further work is needed to better characterize N burial in temperate seagrass meadows and to assess the relative contributions of burial and denitrification to net N removal.

Retention in seagrass biomass, the temporary portion of the N filter function, is better studied than N removal and is generally found to be a dominant flux of nitrogen in seagrass meadows (McGlathery 2008). In non-eutrophic estuarine systems, N demand typically exceeds

N loading (Pedersen et al. 2004), indicating that most N entering these systems passes through the primary producer pool (McGlathery et al. 2007). Assimilation rates typically peak during the summer growing season in temperate meadows (Pedersen and Borum 1993; Risgaard-Petersen et al. 1998; Park et al. 2013). Seagrass tissue is more recalcitrant than algal biomass (Banta et al. 2004), indicating that the temporary N retention during the growing season reduces N availability compared to algae-dominated systems, thus contributing to the N filter and supporting the positive feedback for seagrass growth. Given that N assimilation fluxes are large relative to biogeochemical N processes, the N retention component of the N filter function is significant, despite the short-term impact. Comparing the relative importance of N retention in seagrass biomass to N removal through burial and denitrification will clarify the different ecological dimensions of the seagrass N filter function.

The coastal N filter function is lost when seagrass meadows decline due to anthropogenic impacts, including dredging, eutrophication, and marine heatwaves. Globally, seagrass loss is accelerating (Orth et al. 2006a, Waycott et al. 2009). Habitat restoration reestablishes natural meadow processes (Orth and McGlathery 2012; Orth et al. 2012), including carbon sequestration at rates on par with natural, undisturbed meadows (Greiner et al. 2013, Oreska et al. 2017a). We expect that restoration also reestablishes the coastal N filter function of seagrass meadows to a level comparable to natural meadows; however, this effect has yet to be examined.

The purpose of this study was to assess the return of the coastal nitrogen filter in a restored *Zostera marina* (eelgrass) meadow located in a shallow coastal lagoon in Virginia, USA, and to compare the magnitude of N filtering processes to both external N loading from watershed sources and to internal N loading from N fixation within the meadow. Historically, eelgrass dominated the Virginia coastal bays but a combination of wasting disease and the

impacts of a hurricane in 1933 lead to local extinction (Orth and McGlathery 2012). Restoration by seeding was begun in 2001 by the Virginia Institute of Marine Sciences (VIMS) leading to the establishment of >25 km² of mature eelgrass meadows (<http://web.vims.edu/bio/sav/>). This restoration provides a unique opportunity to assess the impact of seagrass presence on retention and removal of nitrogen in the lagoon. By comparing measurements made in the mature restored meadow to measurements in adjacent, unrestored (“bare”) sediments, we were able to directly assess the nitrogen cycle in the lagoon with and without seagrass. In previous studies, we have measured the effect of this seagrass restoration on biogeochemical fluxes, including N fixation and denitrification (Aoki and McGlathery 2018, 2019). Here, we collected detailed measurements of N burial and N assimilation and compared these components to N fixation, denitrification, and external N loading rates, in order to understand how restoration affected the coastal N filter.

Methods

Study site

This study was conducted at the restored *Z. marina* meadow in South Bay, a shallow lagoon located on the Eastern Shore of Virginia, along the USA mid-Atlantic coast (37.269852, -75.810716). The South Bay meadow is one of four restored seagrass meadows in Virginia that were seeded as part of a landscape-scale restoration project (Orth et al. 2012). In South Bay, seeds were broadcast in 0.4 ha plots in 2001; these original seed plots eventually coalesced into a contiguous meadow that has continued to spread ([Figure 1](#)). In 2015, the restored meadow was approximately 6.9 km² (<http://web.vims.edu/bio/sav/>).

Nitrogen cycle processes

To assess the magnitude of the coastal filter in the restored seagrass meadow, we compared N inputs to N removal and retention measured in the meadow and in adjacent bare sediments. N inputs included loading from terrestrial and atmospheric sources and N fixation occurring within the meadow, including the contributions from sediment N fixation and epiphytic N fixation occurring on seagrass leaves (potentially an important component of total N fixation, Moriarty and O'Donohue 1993; Cole and McGlathery 2012). N removal consisted of burial and denitrification, and N retention was estimated by measuring N assimilation in seagrass biomass. Burial and assimilation rates were measured directly in this study, and fixation and denitrification data were compiled from recent studies in the South Bay meadow (Aoki and McGlathery 2018, 2019). N loading data were adapted from a nitrogen budget prepared by Anderson et al. (2010) for a neighboring coastal lagoon with similar watershed land use. Fixation, denitrification, and assimilation were measured seasonally in spring, summer, and fall; in order to determine annual rates, winter rates were estimated as half of the rates during fall, based on ratios of fall and winter measurements in other *Zostera* systems (Risgaard-Petersen et al. 1998; Eyre et al. 2011) and low winter shoot densities in this system (unpublished data). The annual rates were compared in units of $\text{g N m}^{-2} \text{ y}^{-1}$; standard errors and degrees of freedom for each rate were propagated using the approach and equations described by Lehrter & Cebrian (2010) in order to calculate confidence intervals for each rate. Bare measurements of N burial were made in sediments outside of the established seagrass meadow, approximately 150 m from the meadow edge. For denitrification and fixation, we relied on rates reported in earlier studies from this site (Aoki and McGlathery 2018, 2019). As reported in those studies, bare measurements of denitrification and N fixation were made in experimentally cleared sub-plots (4 m^2) in the meadow interior in order to isolate the direct effects of seagrass growth, i.e. oxygen

and carbon exudation, and to eliminate the effects of confounding factors outside the meadow, such as varying water depths and flow regimes, on sediment biogeochemistry.

N loading

Inputs of nitrogen to South Bay from terrestrial and atmospheric sources were assessed based on an earlier assessment of nitrogen cycling in Hog Island Bay, a coastal lagoon located approximately 20 km north of South Bay, with a similar lagoon area, ratio of watershed area to lagoon area, and level of development and land use in the watershed (Anderson et al. 2010). Terrestrial and atmospheric inputs to Hog Island Bay were therefore considered to be broadly similar to South Bay (Cole 2011). The nitrogen sources evaluated in the Anderson study were base flow, surface water run-off, groundwater discharge, and atmospheric deposition. Base flow was calculated from stream discharge and nitrate concentrations; N loading from run-off and groundwater discharge were estimated as 40% and 33% of base flow loading respectively, based on a combination of direct measurements and modeling (see Anderson et al. 2010 for details). Atmospheric deposition (wet and dry) was calculated from samples collected at a meteorological tower located on Hog Island, the barrier island adjacent to Hog Island Bay, from 1990-1999, and corrected for sample preservation error. For the South Bay meadow budget, the loading rates determined in the Anderson study ($\text{kg N ha}^{-1} \text{ y}^{-1}$) were scaled based on the area of the South Bay lagoon and watershed, as determined by Hayden & Porter (2001).

N fixation and denitrification

Daily rates of N fixation and denitrification were calculated from hourly rates previously measured in this system, and full details of the incubation methods are available elsewhere (Aoki and McGlathery 2018 and Aoki and McGlathery 2019). Briefly, hourly acetylene reduction rates in sediments and by epiphytes on seagrass blades were measured from June 2016-August 2017

using jar incubations; rates were converted to N fixation rates using the theoretical 3:1 ratio (Seitzinger and Garber 1987; Welsh 2000). To scale hourly N fixation rates to annual rates, dark and light incubation rates were pooled, as there was no significant effect of light on either sediment or epiphyte N fixation (see Aoki and McGlathery 2019), and the pooled rates were integrated over 24 hours to calculate daily rates for each season. Total daily N fixation was calculated as the sum of sediment and epiphyte rates; epiphyte N fixation was a maximum of 20% of total N fixation in summer. Daily seasonal rates were then integrated over each season to calculate annual rates.

Hourly denitrification rates in sediments were measured from June 2014-July 2015 using an in situ, push-pull, isotope pairing method; the method is described in detail in Aoki and McGlathery (2017). Using the push-pull method, the hourly rate measurements were made under variable in situ daylight conditions; extrapolating from hourly to daily rates therefore required assumptions about sediment denitrification under dark conditions. Under light conditions, the denitrification rates were characterized by “hotspots” where maximum rates exceeded median rates by an order of magnitude. We attributed these hotspot rates to the effects of seagrass root exudation (see Aoki and McGlathery 2018), and assumed the hotspots would not occur under dark conditions. We therefore calculated a dark rate based on the median of the light rates excluding the hotspot rates, and extrapolated to daily rates by scaling the light and dark rates for 12 hours each. There is some evidence that denitrification rates in seagrass meadow sediments may be greater under dark conditions compared to light conditions, due to the lack of competition for nitrate from autotrophs (Welsh et al. 2000; Eyre et al. 2011, 2013). The daily rates reported here may therefore underestimate daily denitrification. Fall and spring rates of denitrification in the bare sediments were estimated based on the ratio of bare rates to seagrass

rates during summer. Daily seasonal rates were integrated by each season to calculate annual rates.

Burial

N burial rates in seagrass sediments and bare sediments were calculated from sediment accretion rates and the N content of the sediment. Accretion rates (cm yr^{-1}) were provided in previous work at the site by Greiner et al. (2013) that used ^{210}Pb dating of sediment cores to a 20-cm depth; the accretion rates were confirmed in a later study by Oreska et al. (2017c). To determine N content, small sediment cores (2.5 cm diameter, 6 cm depth) were collected in 2013 from 16 sites distributed across the meadow and 2 bare sediment sites outside the meadow, with 4 replicate cores per site. An additional 6 sites clustered in the interior of the meadow and an additional bare site outside the meadow were sampled to a depth of 5 cm in 2014, with 5 replicate cores per site. A total of 22 seagrass sites and 3 bare sites were analyzed for N content. Sediments were dried to a constant weight at 60°C , and bulk density was calculated from the dry weight and wet volume of the sample. N content (%N) was measured using a Carlo Erba NA 2500 Elemental Analyzer, bulk N content (mg cm^{-3}) was calculated from %N and bulk density, and N burial rates ($\text{g N m}^{-2} \text{ y}^{-1}$) were calculated from bulk N content and accretion rates.

As the South Bay meadow has expanded from the original seed plots, different areas of the meadow have been restored for varying amounts of time, i.e., the meadow age varies spatially. Long-term monitoring has shown a 5-year lag between the initial seeding and changes to sediment characteristics and has shown that after 9 years, shoot densities in the meadow stabilized around $500 \text{ shoots m}^{-2}$ (McGlathery et al. 2012). The accretion rates measured by Greiner et al. (2013) showed the same pattern of a slight increase in accretion over the first five years of restoration, followed by a more rapid increase over years 5-9. We therefore calculated

separate burial rates for three age brackets of restored meadow: 1-5 years, 5-9 years, and >9 years. Aerial photographs from an annual survey by VIMS (<http://web.vims.edu/bio/sav/>) allowed us to determine the age of the 22 plots sampled for burial within the meadow and to calculate mean N content and burial rates for each age bracket. We also used the aerial photography (digitized in ArcGIS) to determine the extent of meadow coverage within each age bracket in 2015 in order to calculate a weighted average burial rate for the entire meadow.

Assimilation

Assimilation of N into seagrass biomass was calculated from seagrass productivity rates and seagrass tissue N content. Productivity was measured using the leaf-marking technique (Short and Duarte 2001) in June, August, and October 2016 and in April and June 2017. Replicate 20 x 10 cm wire frames were anchored in the seagrass sediment, and all shoots within the frame were marked by puncturing the sheath bundle with a 25.5-gauge needle. After 10-15 days, the shoots, including rhizomes, were carefully harvested. Each shoot was separated into new growth (unmarked new leaves and leaf tissue below the needle scar) and old growth (tissue above the scar); new and old aboveground tissue was dried to a constant weight to determine shoot specific growth rates over the marking period ($\text{g shoot}^{-1} \text{d}^{-1}$). The plastochrone interval, or the time period between the production of each consecutive leaf, was calculated based on the number of new leaves that appeared during the marking period. The total rhizome length and average internode length were measured and rhizomes were also dried to a constant weight in order to calculate a length-to-weight ratio for belowground biomass. Because *Zostera marina* produces a new rhizome node for each new leaf, the belowground biomass growth rates ($\text{g shoot}^{-1} \text{d}^{-1}$) could be calculated from the average internode length, length-to-weight ratio, and the plastochrone interval. Four productivity frames were deployed at each of 3 plots in each

sampling month except in June when frames were deployed at 6 plots; on average, 68 marked shoots were recovered during each marking period. Above- and belowground productivity rates were converted to N assimilation rates using the N content of triplicate seagrass biomass samples that were collected simultaneously with the productivity samples at two plots using 15 cm diameter, 15 cm depth cores. Biomass samples were sorted into live and dead above and belowground biomass, dried to a constant weight at 60°C, pulverized using a Biospec Products MiniBeadBeater, and analyzed for N content on a Carlo Erba NA 2500 Elemental Analyzer. Seagrass shoot density was also measured concurrently with productivity in 8-10 replicate 0.25 m² quadrats scattered haphazardly at each plot, and shoot-specific assimilation rates were scaled to areal assimilation rates by shoot density.

Results

N loading

Table 2 shows the N loading values from Anderson et al. 2010 scaled to the lagoon and watershed areas of South Bay in order to calculate the total N load to the South Bay meadow. On an areal basis, the lagoon received a nitrogen load of 1.23 g N m⁻² y⁻¹ (95% confidence interval (CI) 0.39 g N m⁻² y⁻¹). Atmospheric deposition was the dominant source of nitrogen, accounting for 77% of the external N inputs. Since these external inputs were not affected by the presence of seagrass in the lagoon, the external N load was equivalent for both the seagrass and bare sediment.

N fixation and denitrification

Daily rates of both N fixation and denitrification, scaled from the hourly rates reported previously by Aoki and McGlathery (2018; 2019), peaked during summer (Figure 2). Total N fixation rates, combining sediment and epiphyte rates, were greater than denitrification during

summer and similar during other seasons. Bare and seagrass sediments had comparable N fixation rates, likely due to the similar organic matter content in the seagrass sediments and in the experimentally cleared bare plots within the meadow (Aoki and McGlathery 2019). The annual N fixation rate was $0.93 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.17 \text{ g N m}^{-2} \text{ y}^{-1}$) in the seagrass meadow and $0.74 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.16 \text{ g N m}^{-2} \text{ y}^{-1}$) in the bare sediment. Daily rates of summer denitrification were on average 4x greater in seagrass sediments than in bare sediments (Figure 2; note that denitrification rates were measured in both experimentally cleared plots within the meadow and in bare sediments outside the meadow, see Aoki and McGlathery 2018 for details). The annual denitrification rate was $0.62 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.21 \text{ g N m}^{-2} \text{ y}^{-1}$) in seagrass sediments and $0.16 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.05 \text{ g N m}^{-2} \text{ y}^{-1}$) in bare sediments.

N burial

As the seagrass meadow has matured over time, burial rates have increased due to increased sedimentation rates and increased sediment N content (Figure 3). Burial rates within each age bracket ranged from $0.17 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.14 \text{ g N m}^{-2} \text{ y}^{-1}$) in bare sediments to $3.52 \text{ g N m}^{-2} \text{ y}^{-1}$ (95% CI $0.69 \text{ g N m}^{-2} \text{ y}^{-1}$) in the oldest (>9 years old) meadow sediments (Table 3). Compared across age brackets, burial rates varied significantly (ANOVA, $F_{3,21}=5.22$, $p<0.05$), and post-hoc contrasts showed that the >9 year old sediments had significantly higher rates than both the bare and recently colonized (1-5 year old) sediments (Tukey test, $p<0.05$). The total area of the seagrass meadow has also expanded steadily through 2015 (Figure 4), and the oldest areas of the meadow accounted for only a small proportion of the total meadow area in 2015 (Table 3). The average burial rate over the total 2015 meadow area, weighted based on the proportion of the meadow area in each age bracket, was $1.95 \text{ g N m}^{-2} \text{ y}^{-1}$.

N retention

Shoot-specific aboveground seagrass productivity peaked in June (Figure 5), leading to peak N assimilation rates in summer (Figure 6). Shoot counts were highest in June 2017; however, N content of aboveground tissue was highest in October 2016. Belowground tissue N content and belowground biomass growth rates showed little variation across seasons. Assuming a winter assimilation rate of half the fall assimilation rate (based on winter shoot densities measured in previous years, data not shown), the annual assimilation rate was $2.62 \text{ g N m}^{-2} \text{ y}^{-1}$.

Discussion

Burial dominates N removal

Net N removal, the combination of burial and denitrification, was more than 12x greater in the mature meadow sediments compared to bare sediments (4.14 compared to $0.33 \text{ g N m}^{-2} \text{ y}^{-1}$), demonstrating that the seagrass restoration successfully reinstated the N filter function. Burial was the major component of N removal, accounting for 85% of the total. Recent work using staple isotopes to characterize the sediment organic carbon (SOC) in the South Bay seagrass meadow showed that approximately 50% of the SOC derived from benthic microalgae, 40% from seagrass, and 10% from marsh grass (*Spartina alterniflora*) (Oreska et al. 2017b). A similar breakdown of the sediment organic nitrogen pool would suggest that both the seagrass and benthic microalgae were important sources of nitrogen buried in the seagrass sediments. Burial is therefore one important link between the high productivity of the seagrass and the filter function. The meadow assimilates large amounts of dissolved nitrogen into biomass during the growing season and a portion of that biomass remains buried in the sediments, reducing the N available for immediate recycling. Productivity is also linked to burial by the physical structure of above and belowground seagrass biomass that directly enhances burial through wave attenuation and sediment stabilization.

Our analysis showed that N burial increased with time since restoration (Table 3). The maximum burial rate measured at the mature meadow sites (>9 years old) was $3.52 \text{ g N m}^{-2} \text{ y}^{-1}$, which falls within the range of previously reported rates in *Zostera* meadows ($2.7\text{-}3.9 \text{ g N m}^{-2} \text{ y}^{-1}$, Eyre et al. 2016, Table 1). Thus, within 10 years after seeding, the restoration successfully reestablished N removal via burial to rates comparable to natural meadows. The meadow-wide burial rate in 2015, calculated as a weighted average of burial rates in the different age classes (time since restoration) of the meadow, was lower, $1.95 \text{ g N m}^{-2} \text{ y}^{-1}$. If we extrapolate this current meadow-wide burial rate to the full area of the meadow in 2015, 6.9 km^2 , we would expect the meadow to remove 13.6 t N y^{-1} through burial, while an equivalent area of bare sediment would bury an order of magnitude less (1.2 t N y^{-1}). As the restored meadow continues to mature, we expect that the burial rate will increase in younger areas of the meadow, leading to a meadow-wide burial rate on par with natural systems. This finding mirrors earlier work demonstrating that carbon burial in the restored meadow sediments was comparable to natural systems within ten years of restoration (Greiner et al. 2013; Oreska et al. 2017a).

Denitrification was a minor component of the N filter in the seagrass sediments (15% of net N removal), but denitrification rates in seagrass sediments were enhanced by approximately 4x compared to bare sediments. Many studies of N removal in seagrass meadows to date have focused on enhanced denitrification; however, our results show that burial can be the more important factor. The hourly rates used in this study agree well with numerous measurements of denitrification in seagrass sediments, including both isotope pairing (e.g. Welsh et al. 2000, Russell et al. 2016) and $\text{N}_2:\text{Ar}$ studies (Zarnoch et al. 2017). However, the extrapolated annual rate of denitrification ($0.62 \text{ g N m}^{-2} \text{ y}^{-1}$) is low compared to two recent studies using dark $\text{N}_2:\text{Ar}$ rates ($7\text{-}19 \text{ g N m}^{-2} \text{ y}^{-1}$; Smyth et al. 2013; Eyre et al. 2016). While the denitrification rates in this

system were likely limited by the very low nitrate availability (undetectable in surface waters throughout the year) and relatively low sediment organic matter content (2.6%), it is not immediately clear why the rates differ so dramatically between locations. Factors such as residence time and benthic microalgae presence also influence denitrification rates (Cornwell et al. 1999; Welsh et al. 2000), and methodological differences likely contribute to the wide range of denitrification rates reported in the literature (see Eyre et al. (2013) and Aoki and McGlathery (2017) for more discussion of how methodology affects denitrification measurements). Further investigation of denitrification in seagrass meadows is needed to clarify the range of rates in natural systems under different conditions and to establish the importance of denitrification relative to N burial.

Significant N retention in seagrass biomass

Over an annual time scale, seagrass N assimilation was a major flux, comparable to 75% of the burial rate in the most mature areas (Table 4). In fact, this N assimilation rate likely underestimates total N demand by the seagrass, since the leaf-marking technique used in this study does not include leaf maturation (thickening and widening of the leaf) above the needle scar in the growth rate (Park et al. 2010). The large N assimilation rate indicates how seagrass biomass mediates N cycling in the lagoon. By retaining nitrogen in recalcitrant seagrass tissue during the growing season, the meadow limits rapid recycling of nutrients that can fuel phytoplankton and macroalgae blooms (Banta et al. 2004). Temporary N retention in seagrass biomass thus provides an additional filter mechanism during the growing season that helps to maintain water quality, creating an environment conducive to seagrass growth.

Although seagrass biomass is more persistent than algae, a portion of the N assimilated into seagrass tissue will eventually be recycled. N recycling includes both remineralization of

seagrass-derived N in meadow sediments and internal recycling of N within seagrass shoots, sometimes known as N reclamation. We did not measure remineralization or reclamation, but previous studies indicate that each of these processes may meet as much as 50% of the seagrass N demand (Romero et al. 2006 and references therein), and previous work has demonstrated that remineralization is important to meet benthic microalgal demand in this system (Anderson et al. 2003). Generally, tightly linked recycling processes allow seagrass to achieve extremely high rates of productivity in nutrient-poor waters, as was likely the case in our study system. The high N assimilation rate measured here underscores the duality of seagrass meadows. As productivity hotspots, they drive nutrient cycling, including the release of nutrients through remineralization. At the same time, the meadows filter nitrogen through burial, stimulation of denitrification, and retention in biomass. By creating a positive feedback for seagrass growth, the coastal filter function supports the many other ecosystem services provided by seagrass meadows, such as habitat provisioning and carbon sequestration, in addition to limiting water column nitrogen.

The fate of nitrogen assimilated into seagrass biomass is largely unknown. While a portion of the nitrogen that accumulates in seagrass biomass is buried in seagrass sediments, an additional portion is exported from the ecosystem. Leaves and shoots are lost throughout the growing season, and, in this system, accumulations of seagrass wrack have been observed on the ocean side of the barrier islands. Seagrass biomass may contribute to sediment organic matter in the neighboring marshes, and a recent study suggests that seagrass wrack may be buried on the continental shelf (Duarte and Krause-Jensen 2017). This physical transport of biomass nitrogen out of the seagrass meadow, coupled with the low rates of external N inputs from the watershed, underscores the importance of internal N loading from enhanced N fixation and remineralization.

Low N inputs from loading and fixation

The N inputs from watershed loading were low compared to the N removal and retention fluxes (Table 4). Previous measurements have shown that the Virginia coastal bays experience some of the lowest rates of watershed N loading compared to coastal lagoons worldwide (McGlathery et al. 2007) so the low rates shown here were expected. N fixation, an internal source of N within the meadow, was also low compared to total N removal and retention fluxes, although fixation did outweigh denitrification, both during peak summer rates and in the integrated annual rate. The low rates of N loading likely contributed to the dominance of N fixation over denitrification; NO_3^- concentrations in surface waters in South Bay were often undetectable (unpublished data), limiting the supply of nitrate to support denitrification. The underlying hourly rates of N fixation used in this analysis were similar to rates measured in other *Zostera* meadows (Welsh et al. 1996; McGlathery et al. 1998; Russell et al. 2016). Few authors have scaled hourly rates to annual, areal rates, although Russell et al. (2016) recently found that N fixation dominated denitrification both hourly and over an annual time-scale in intertidal *Z. muelleri* flats. Overall, the low rates of N fixation were in line with expectations from the literature.

Given the high rates of burial and assimilation and low rates of N loading and fixation measured in this study, it is clear that additional sources provided nitrogen to the meadow. As mentioned above, internal recycling (i.e. remineralization and N reclamation) were likely important sources of N to the seagrass meadow. However, there is also evidence to suggest that the N fixation rates used in this study were underestimated. Acetylene reduction, the traditional method used here to measure N fixation in sediments, is an indirect method with several known drawbacks (Welsh 2000; Fulweiler et al. 2015). Recent studies using direct measurement techniques in estuarine sediments have found substantially higher N fixation rates compared to

published acetylene reduction values in both unvegetated sediments (Gardner et al. 2006; Fulweiler et al. 2007; Newell et al. 2016), and in the South Bay meadow (Aoki and McGlathery 2019). Additional direct measurements of N fixation are needed to explore these patterns and to fully account for internal N inputs that support the high rates of N assimilation in the meadow.

Motivation for restoration and conservation

Seagrass meadows around the world are threatened by human activity. Both restoration of lost meadows and conservation of existing meadows are needed to preserve these important ecosystems and the many functions they perform (Unsworth et al. 2018). This study shows that seagrass restoration can recover the N filter function within ten years of seeding, reestablishing positive feedbacks that support continued seagrass growth. While the exact rates measured here may not translate to other restored systems, the relative importance of different N filter processes is instructive.

Overall, our results indicate that burial was by far the dominant process removing N from the system, accounting for 85% of total removal. Seagrass productivity was the second most important component of the N filter function, temporarily retaining N during the growing season and improving water quality. These two processes are linked through the physical structure of the canopy and the contribution of seagrass tissue to the sediment N pool. However, burial rates in the most mature meadow sites, i.e. sites comparable to an established natural meadow, exceeded N assimilation into seagrass biomass by 1.3x. Rates of both burial and assimilation measured here were likely low compared to other eelgrass systems. The Virginia coastal bays are near the southern end of the geographic range of *Z. marina* (Moore and Short 2006), and eelgrass meadows in more northern climates have shown even greater productivity rates (Pedersen and Borum 1993; Risgaard-Petersen et al. 1998). Leaf N content also indicated that the eelgrass in

South Bay were N limited at least part of the year (Figure 5B, values below 1.8% indicate nitrogen limitation, Duarte 1990). In systems with greater N loading that relieves this limitation, seagrass assimilation rates would be somewhat greater. Burial rates could also be much greater than the rate reported here in more eutrophied systems with greater seston concentrations; under eutrophied conditions, burial would likely exceed assimilation by even more than in this oligotrophic system. To date, burial rates in *Zostera* meadows have been measured only under oligotrophic conditions (Eyre et al. 2016 and this study), and more measurements of N burial in temperate seagrass meadows are needed to quantify the range of burial rates across different environmental conditions.

The importance of seagrass ecosystems as sites for N burial can motivate both restoration and conservation. In the restored meadow studied here, the return of mature seagrass increased the capacity of the system to remove N by 20x compared to unvegetated sediments. This large N removal capacity in turn could potentially limit negative effects from external N loading. Conversely, in areas where seagrass meadows are lost, the system would have a diminished capacity to retain N, leading to an increase in available N that is independent of increases in external N loading rates. The results of this study therefore emphasize the importance of both restoration and conservation of seagrass meadows in order to maintain the coastal N filter function. Better quantification of N removal rates in both natural and restored meadows is needed to add to the results presented here and to further our understanding of how seagrass influence N removal rates.

444 **References**

- 445 Anderson, I. C., K. J. McGlathery, and A. C. Tyler. 2003. Microbial mediation of “reactive”
 446 nitrogen transformations in a temperate lagoon. *Marine Ecology Progress Series* **246**: 73–
 447 84.
- 448 Anderson, I. C., J. W. Stanhope, A. K. Hardison, and K. J. McGlathery. 2010. Sources and fates
 449 of nitrogen in Virginia coastal bays, p. 43–72. *In* *Coastal Lagoons: Critical habitats of*
 450 *environmental change*. CRC Press.
- 451 Aoki, L., and K. McGlathery. 2018. Restoration enhances denitrification and DNRA in
 452 subsurface sediments of *Zostera marina* seagrass meadows. *Marine Ecology Progress*
 453 *Series* **602**: 87–102. doi:10.3354/meps12678
- 454 Aoki, L. R., and K. J. McGlathery. 2017. Push-pull incubation method reveals the importance of
 455 denitrification and dissimilatory nitrate reduction to ammonium in seagrass root zone.
 456 *Limnology and Oceanography: Methods* **15**: 766–781. doi:10.1002/lom3.10197
- 457 Aoki, L. R., and K. J. McGlathery. 2019. High rates of N fixation in seagrass sediments
 458 measured via a direct ³⁰N₂ push-pull method. *Marine Ecology Progress Series*.
 459 doi:https://doi.org/10.3354/meps12961p
- 460 Banta, G. T., M. F. Pedersen, and S. L. Nielsen. 2004. Decomposition of marine primary
 461 producers: Consequences for nutrient recycling and retention in coastal ecosystems, p.
 462 187–216. *In* *Estuarine nutrient cycling: the influence of primary producers*. Kluwer
 463 Academic Publishers.
- 464 Barbier, E. B., S. D. Hacker, C. Kennedy, E. W. Koch, A. C. Stier, and B. R. Silliman. 2011. The
 465 value of estuarine and coastal ecosystem services. *Ecological Monographs* **81**: 169–193.
- 466 Carr, J., P. D’Odorico, K. McGlathery, and P. Wiberg. 2010. Stability and bistability of seagrass
 467 ecosystems in shallow coastal lagoons: Role of feedbacks with sediment resuspension
 468 and light attenuation. *Journal of Geophysical Research* **115**. doi:10.1029/2009JG001103
- 469 Cole, L. 2011. Inputs and fluxes of nitrogen in the Virginia coastal bays: Effects of newly-
 470 restored seagrasses on the nitrogen cycle. Ph.D. University of Virginia.
- 471 Cole, L., and K. McGlathery. 2012. Nitrogen fixation in restored eelgrass meadows. *Marine*
 472 *Ecology Progress Series* **448**: 235–246. doi:10.3354/meps09512
- 473 Cornwell, J. C., W. M. Kemp, and T. M. Kana. 1999. Denitrification in coastal ecosystems:
 474 methods, environmental controls, and ecosystem level controls, a review. *Aquatic*
 475 *Ecology* **33**: 41–54. doi:10.1023/A:1009921414151
- 476 Duarte, C. M. 1990. Seagrass nutrient content. *Marine Ecology Progress Series* **67**: 201–207.
- 477 Duarte, C. M., and D. Krause-Jensen. 2017. Export from seagrass meadows contributes to
 478 marine carbon sequestration. *Frontiers in Marine Science* **4**: 13.
 479 doi:10.3389/fmars.2017.00013
- 480 Eyre, B. D., A. J. P. Ferguson, A. Webb, D. Maher, and J. M. Oakes. 2011. Denitrification, N-
 481 fixation and nitrogen and phosphorus fluxes in different benthic habitats and their
 482 contribution to the nitrogen and phosphorus budgets of a shallow oligotrophic sub-
 483 tropical coastal system (southern Moreton Bay, Australia). *Biogeochemistry* **102**: 111–
 484 133. doi:10.1007/s10533-010-9425-6
- 485 Eyre, B. D., D. T. Maher, and C. Sanders. 2016. The contribution of denitrification and burial to
 486 the nitrogen budgets of three geomorphically distinct Australian estuaries: Importance of
 487 seagrass habitats. *Limnology and Oceanography* **61**: 1144–1156. doi:10.1002/lno.10280

- Eyre, B. D., D. T. Maher, and P. Squire. 2013. Quantity and quality of organic matter (detritus) drives N₂ effluxes (net denitrification) across seasons, benthic habitats, and estuaries. *Global Biogeochemical Cycles* **27**: 1083–1095. doi:10.1002/2013GB004631
- Fulweiler, R. W., E. M. Heiss, M. K. Rogener, S. E. Newell, G. R. LeClerc, S. M. Kortebein, and S. W. Wilhelm. 2015. Examining the impact of acetylene on N-fixation and the active sediment microbial community. *Frontiers in Microbiology* **6**: 418. doi:10.3389/fmicb.2015.00418
- Fulweiler, R. W., S. W. Nixon, B. A. Buckley, and S. L. Granger. 2007. Reversal of the net dinitrogen gas flux in coastal marine sediments. *Nature* **448**: 180–182. doi:10.1038/nature05963
- Gacia, E., C. M. Duarte, and J. J. Middelburg. 2002. Carbon and nutrient deposition in a Mediterranean seagrass (*Posidonia oceanica*) meadow. *Limnology and Oceanography* **47**: 23–32.
- Gardner, W. S., M. J. McCarthy, S. An, D. Sobolev, K. S. Sell, and D. Brock. 2006. Nitrogen fixation and dissimilatory nitrate reduction to ammonium (DNRA) support nitrogen dynamics in Texas estuaries. *Limnology and Oceanography* **51**: 558–568. doi:10.4319/lo.2006.51.1_part_2.0558
- Greiner, J. T., K. J. McGlathery, J. Gunnell, and B. A. McKee. 2013. Seagrass restoration enhances “blue carbon” sequestration in coastal waters J. Cebrian [ed.]. *PLoS ONE* **8**: e72469. doi:10.1371/journal.pone.0072469
- Gurbisz, C., W. M. Kemp, J. C. Cornwell, L. P. Sanford, M. S. Owens, and D. C. Hinkle. 2017. Interactive effects of physical and biogeochemical feedback processes in a large submersed plant bed. *Estuaries and Coasts* **40**: 1626–1641. doi:10.1007/s12237-017-0249-7
- Hansen, J. C. R., and M. A. Reidenbach. 2012. Wave and tidally driven flows in eelgrass beds and their effect on sediment suspension. *Marine Ecology Progress Series* **448**: 271–287. doi:10.3354/meps09225
- Hayden, B., and J. Porter. 2001. Terrestrial-marine watershed boundaries on the Delmarva peninsula of Virginia 2001. knb-lter-vcr.88.22. knb-lter-vcr.88.22 Virginia Coast Reserve Long-Term Ecological Research Project Data Publication.
- van der Heide, T., E. H. van Nes, M. M. van Katwijk, H. Olff, and A. J. P. Smolders. 2011. Positive Feedbacks in Seagrass Ecosystems – Evidence from Large-Scale Empirical Data T. Romanuk [ed.]. *PLoS ONE* **6**: e16504. doi:10.1371/journal.pone.0016504
- Iizumi, H., A. Hattori, and C. P. McRoy. 1980. Nitrate and nitrite in interstitial waters of eelgrass beds in relation to the rhizosphere. *Journal of Experimental Marine Biology and Ecology* **47**: 191–201.
- Lehrter, J. C., and J. Cebrian. 2010. Uncertainty propagation in an ecosystem nutrient budget. *Ecological Applications* **20**: 508–524.
- Mateo, M. A., and J. Romero. 1997. Detritus dynamics in the seagrass *Posidonia oceanica*: Elements for an ecosystem carbon and nutrient budget. *Marine Ecology Progress Series* **151**: 43–53.
- Mateo, M. A., J. Romero, M. Pérez, M. M. Littler, and D. S. Littler. 1997. Dynamics of millenary organic deposits resulting from the growth of the Mediterranean seagrass *Posidonia oceanica*. *Estuarine, Coastal and Shelf Science* **44**: 103–110. doi:10.1006/ecss.1996.0116

- McGlathery, K. J. 2008. Seagrass habitats, p. 1037–1072. *In* Nitrogen in the marine environment. Elsevier.
- McGlathery, K. J., L. K. Reynolds, L. W. Cole, R. . Orth, S. R. Marion, and A. Schwarzschild. 2012. Recovery trajectories during state change from bare sediment to eelgrass dominance. *Marine Ecology Progress Series* **448**: 209–221. doi:10.3354/meps09574
- McGlathery, K. J., N. Risgaard-Petersen, and P. B. Christensen. 1998. Temporal and spatial variation in nitrogen fixation activity in the eelgrass *Zostera marina* rhizosphere. *Marine Ecology Progress Series* **168**: 245–258.
- McGlathery, K., K. Sundbäck, and I. Anderson. 2007. Eutrophication in shallow coastal bays and lagoons: The role of plants in the coastal filter. *Marine Ecology Progress Series* **348**: 1–18. doi:10.3354/meps07132
- Middelburg, J. J., K. Soetaert, P. M. J. Herman, H. T. S. Boschker, and C. H. R. Heip. 2004. Burial of nutrients in coastal sediments: The role of primary producers, p. 217–231. *In* Estuarine Nutrient Cycling: The Influence of Primary Producers. Kluwer Academic Publishers.
- Moore, K. A., and F. T. Short. 2006. *Zostera*: Biology, Ecology, and Management, p. 361–386. *In* Seagrasses: Biology, Ecology, and Conservation. Springer.
- Moriarty, D. J. W., and M. J. O'Donohue. 1993. Nitrogen fixation in seagrass communities during summer in the Gulf of Carpentaria, Australia. *Marine and Freshwater Research* **44**: 117–127.
- Newell, S. E., M. J. McCarthy, W. S. Gardner, and R. W. Fulweiler. 2016. Sediment nitrogen fixation: A call for re-evaluating coastal N budgets. *Estuaries and Coasts* **39**: 1626–1638. doi:10.1007/s12237-016-0116-y
- Oreska, M. P. J., K. J. McGlathery, and J. H. Porter. 2017a. Seagrass blue carbon spatial patterns at the meadow-scale J. Cebrian [ed.]. *PLOS ONE* **12**: e0176630. doi:10.1371/journal.pone.0176630
- Oreska, M. P. J., G. M. Wilkinson, K. J. McGlathery, M. Bost, and B. A. McKee. 2017b. Non-seagrass carbon contributions to seagrass sediment blue carbon. *Limnology and Oceanography* **S3–S18**. doi:10.1002/lno.10718
- Orth, R. ., K. A. Moore, S. R. Marion, D. . Wilcox, and D. B. Parrish. 2012. Seed addition facilitates eelgrass recovery in a coastal bay system. *Marine Ecology Progress Series* **448**: 177–195. doi:10.3354/meps09522
- Orth, R. J., T. J. B. Carruthers, W. C. Dennison, and others. 2006. A global crisis for seagrass ecosystems. *BioScience* **56**: 987–996. doi:10.1641/0006-3568(2006)56[987:AGCFSE]2.0.CO;2
- Orth, R., and K. McGlathery. 2012. Eelgrass recovery in the coastal bays of the Virginia Coast Reserve, USA. *Marine Ecology Progress Series* **448**: 173–176. doi:10.3354/meps09596
- Park, S. R., Y. K. Kim, S. H. Kim, and K.-S. Lee. 2013. Nitrogen budget of the eelgrass, *Zostera marina* in a bay system on the south coast of Korea. *Ocean Science Journal* **48**: 301–310. doi:10.1007/s12601-013-0028-8
- Park, S. R., W. T. Li, S. H. Kim, J. . Kim, and K. . Lee. 2010. A comparison of methods for estimating the productivity of *Zostera marina*. *Journal of Ecology and Field Biology* **33**: 59–65. doi:10.5141/JEFB.2010.33.1.059
- Pedersen, M. F., and J. Borum. 1993. An annual nitrogen budget for a seagrass *Zostera marina* population. *Marine Ecology-Progress Series* **101**: 169–169.

- Pedersen, M. F., S. L. Nielsen, and G. T. Banta. 2004. Interactions between vegetation and nutrient dynamics in coastal marine ecosystems: An introduction, p. 1–16. *In* Estuarine nutrient cycling: the influence of primary producers. Kluwer Academic Publishers.
- Risgaard-Petersen, N., T. Dalsgaard, S. Rysgaard, P. B. Christensen, J. Borum, K. McGlathery, and L. P. Nielsen. 1998. Nitrogen balance of a temperate eelgrass *Zostera marina* bed. *Marine Ecology-Progress Series* **174**: 281–291.
- Romero, J., K. Lee, M. Pérez, M. A. Mateo, and T. Alcoverro. 2006. Nutrient dynamics in seagrass ecosystems, p. 227–254. *In* Seagrasses: Biology, ecology, and conservation. Springer.
- Romero, J., M. Pérez, and M. A. Mateo. 1994. The belowground organs of the Mediterranean seagrass *Posidonia oceanica* as a biogeochemical sink. *Aquatic Botany* **47**: 13–19.
- Russell, D. G., F. Y. Warry, and P. L. M. Cook. 2016. The balance between nitrogen fixation and denitrification on vegetated and non-vegetated intertidal sediments. *Limnology and Oceanography* **61**: 2058–2075. doi:10.1002/lno.10353
- Seitzinger, S. P., and J. H. Garber. 1987. Nitrogen fixation and $^{15}\text{N}_2$ calibration of the acetylene reduction assay in coastal marine sediments. *Marine Ecology Progress Series* **37**: 65–73.
- Short, F. T., and C. M. Duarte. 2001. Methods for the measurement of seagrass growth and production, p. 155–182. *In* Global Seagrass Research Methods. Elsevier.
- Smyth, A. R., S. P. Thompson, K. N. Siporin, W. S. Gardner, M. J. McCarthy, and M. F. Piehler. 2013. Assessing nitrogen dynamics throughout the estuarine landscape. *Estuaries and Coasts* **36**: 44–55. doi:10.1007/s12237-012-9554-3
- Unsworth, R. K. F., L. J. McKenzie, L. M. Nordlund, and L. C. Cullen-Unsworth. 2018. A changing climate for seagrass conservation? *Current Biology* **28**: R1229–R1232. doi:10.1016/j.cub.2018.09.027
- Waycott, M., C. M. Duarte, T. J. Carruthers, and others. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proceedings of the National Academy of Sciences* **106**: 12377–12381. doi:10.1073/pnas.0905620106
- Welsh, D. T. 2000. Nitrogen fixation in seagrass meadows: Regulation, plant–bacteria interactions and significance to primary productivity. *Ecology Letters* **3**: 58–71.
- Welsh, D. T., M. Bartoli, D. Nizzoli, G. Castaldelli, S. A. Riou, and P. Viaroli. 2000. Denitrification, nitrogen fixation, community primary productivity and inorganic-N and oxygen fluxes in an intertidal *Zostera noltii* meadow. *Marine Ecology Progress Series* **208**: 65–77.
- Welsh, D. T., S. Bourgues, R. De Wit, and R. A. Herbert. 1996. Seasonal variations in nitrogen-fixation (acetylene reduction) and sulphate-reduction rates in the rhizosphere of *Zostera noltii*: Nitrogen fixation by sulphate-reducing bacteria. *Marine Biology* **125**: 619–628.
- Zarnoch, C. B., T. J. Hoellein, B. T. Furman, and B. J. Peterson. 2017. Eelgrass meadows, *Zostera marina* (L.), facilitate the ecosystem service of nitrogen removal during simulated nutrient pulses in Shinnecock Bay, New York, USA. *Marine Pollution Bulletin* **124**: 376–387. doi:10.1016/j.marpolbul.2017.07.061

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626

Figure Legends

Figure 1. Map of the South Bay meadow showing the original seeded plots and meadow expansion over time.

Figure 2. Daily rates of denitrification (negative values, filled bars) and N fixation (positive values, patterned bars) scaled from hourly measurements reported previously by Aoki and McGlathery (2018 and 2019). Measurements in seagrass sediments are light grey; measurements in bare sediments are dark grey. Error bars are SE, n=18-22 for summer values, n=6-9 for other seasons.

Figure 3. Accretion rates (from Greiner et al. 2013) and sediment N content increased with meadow age (time since restoration). Error bars are SE, n=3 (bare), 8 (1-5 years), 4 (5-9 years), 10 (>9 years).

Figure 4 The restored meadow expanded rapidly through 2015, leading to spatial variability in the age (time since restoration) of the meadow.

Figure 5. Shoot-specific productivity (A), seagrass tissue N content (B), shoot-specific elongation (C), and seagrass shoot density (D) were measured seasonally from June 2016-August 2017. Error bars are standard error, n=3 for Oct, April, August, n=6 for June measurements.

Figure 6. Areal N assimilation rates were calculated from the shoot-specific productivity, N content, and shoot density (shown in Figure 5). Error bars are standard error, n=6 in summer, n=3 in fall and spring.

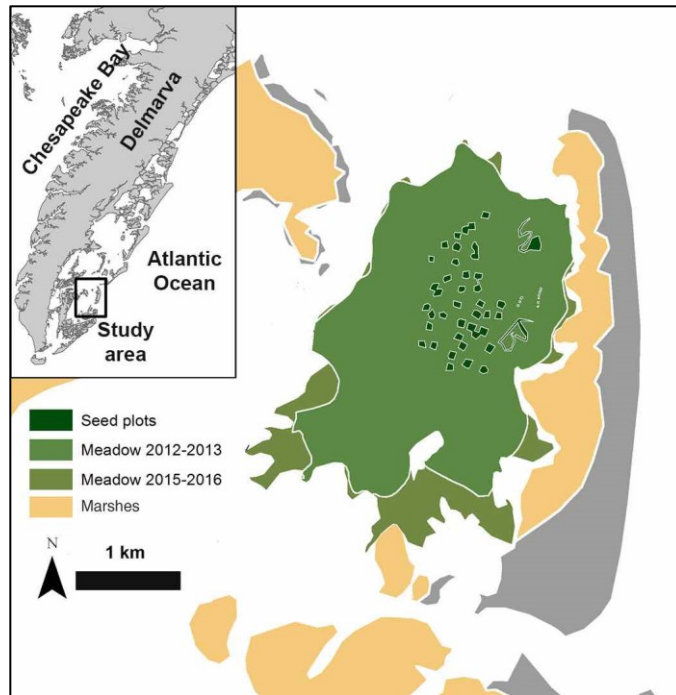


Figure 1. Map of the South Bay meadow showing the original seeded plots and meadow expansion over time.

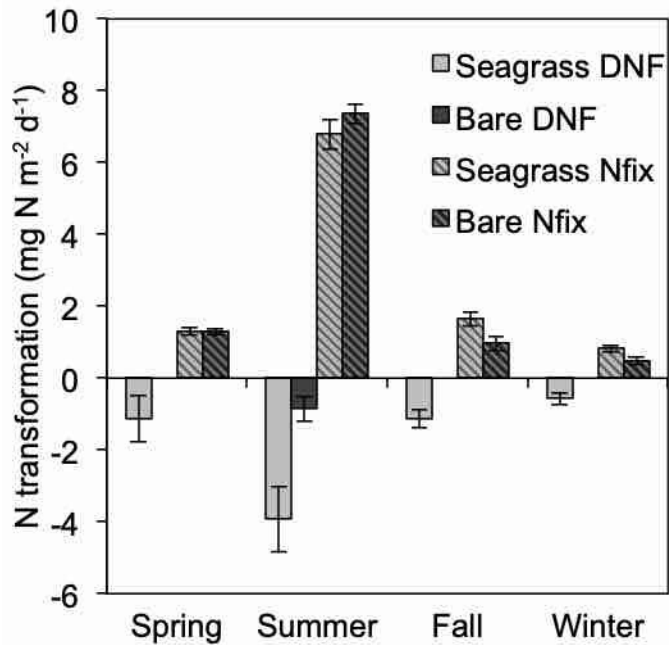


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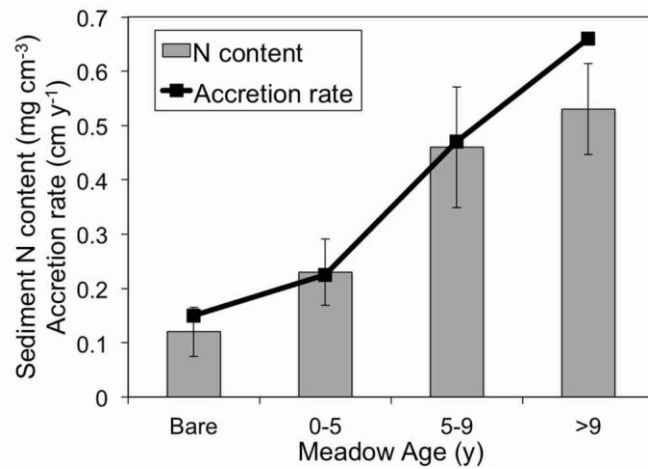


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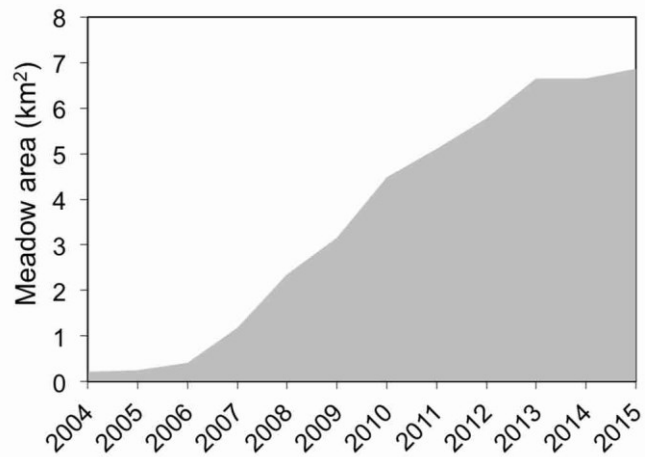


Figure 4 The restored meadow expanded rapidly through 2015, leading to spatial variability in the age (time since restoration) of the meadow.

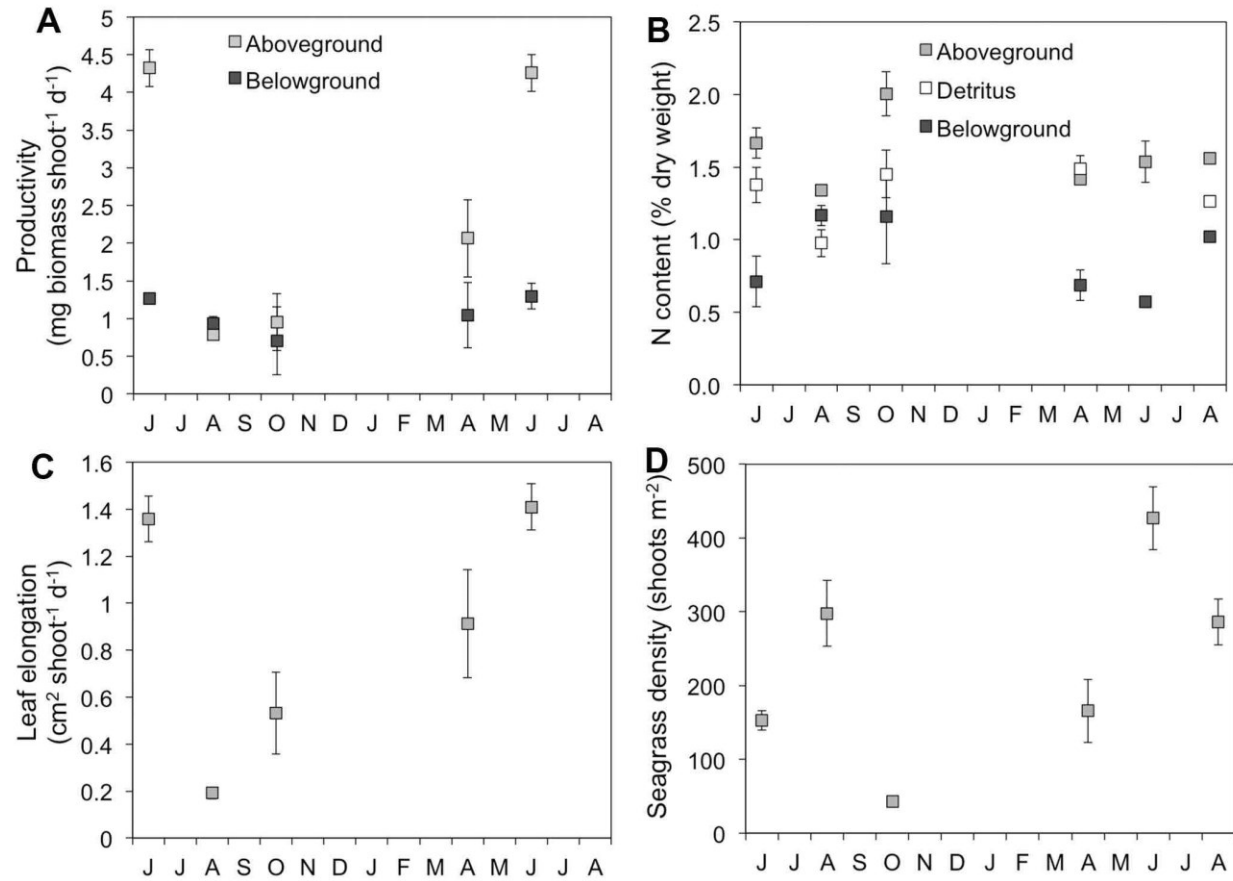


Figure 5. Shoot-specific productivity (A), seagrass tissue N content (B), shoot-specific elongation (C), and seagrass shoot density (D) were measured seasonally from June 2016-August 2017. Error bars are standard error, n=3 for Oct, April, August, n=6 for June measurements.

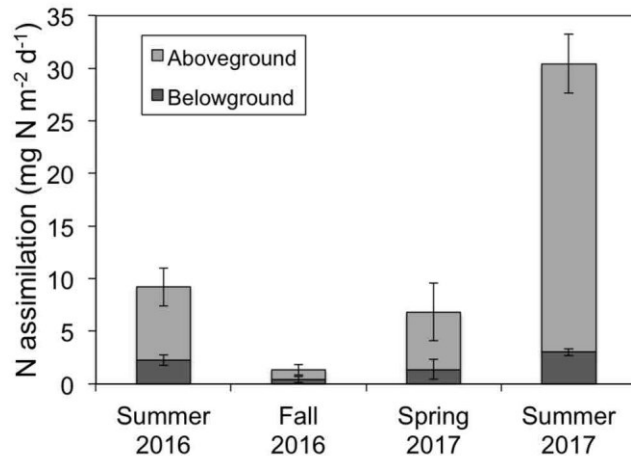


Figure 6. Areal N assimilation rates were calculated from the shoot-specific productivity, N content, and shoot density (shown in Figure 5). Error bars are standard error, $n=6$ in summer, $n=3$ in fall and spring.

676 **Tables**

677 **Table 1.** Rates of N burial in seagrass meadow sediments have so far been reported mainly for

678 *Posidonia* meadows and mainly for oligotrophic ecosystems

Location	Species	N burial (g N m ⁻² y ⁻¹)	Source
SE Australia [†]	<i>Zostera</i> sp.	2.7-3.9	Eyre et al. 2016
	<i>Halophila</i> sp.	1.3-8	
	<i>Ruppia</i> sp.	1.8-8.2	
	<i>Posidonia</i> sp.	5.8	
NW Mediterranean	<i>Posidonia oceanica</i>	13.4	Gacia et al. 2002
NW Mediterranean	<i>Posidonia oceanica</i> *	0.5-1.1	Mateo and Romero 1997
NW Mediterranean	<i>Cymodocea nodosa</i>	3.57	Pedersen et al. 1997
Central Mediterranean	<i>Posidonia oceanica</i> *	0.2-2.3	Romero et al. 1994
Virginia coastal bays, USA [†]	<i>Zostera marina</i>	3.52	This study

679 *Burial rate of seagrass detritus only, excluding particulate N in sediment

680 †Measurements made in oligotrophic systems

681 **Table 2.** N Loading values were adopted from Anderson et al. 2010, and loads were calculated
 682 based on catchment and lagoon areas; loading rates are shown as mean (95% CI min-max).

		N loading	Area	N load
	N Source	(kg N ha ⁻¹ y ⁻¹)	(ha)	(kg N y ⁻¹)
<i>Catchment</i>			6037	47000
	Base Flow	4.59 (0.86-8.76)		
	Surface water runoff	1.84 (0.34-3.50)		
	Groundwater discharge	1.51 (0.28-2.89)		
<i>Lagoon</i>			16946	161000
	Atmospheric deposition	9.49 (7.97-11.01)		
<i>Total</i>				208000

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684

685 **Table 3.** Burial rates were calculated from the accretion rate and N content for different ages of
686 restored meadow; values are mean (SE), n=3 (bare), 8 (1-5 years), 4 (5-9 years), 10 (>9 years).

	Bare sediment	1-5 years old	5-9 years old	>9 years old
Burial (g m ⁻² y ⁻¹)	0.17 (0.07)	0.53 (0.14)	2.16 (0.52)	3.52 (0.55)
Proportion of meadow area in 2015	--	26%	57%	17%

687

688

689 **Table 4.** Comparison of annual N inputs, N retention, and N removal in the meadow and bare
690 sediments*

	N flux	Mean rate (g N m ⁻² y ⁻¹)	95% confidence limit	Source
<i>Seagrass</i>				
Loading		1.23	0.39	Adapted from Anderson et al. 2010
Fixation		0.93	0.17	Aoki & McGlathery 2019
Denitrification		-0.62	0.21	Aoki & McGlathery 2018
Burial		-3.52	0.69	This study
Assimilation		-2.62	0.78	This study
<i>Bare</i>				
Loading		1.23	0.39	Adapted from Anderson et al. 2010
Fixation		0.74	0.16	Aoki & McGlathery 2019
Denitrification		-0.16	0.05	Aoki & McGlathery 2018
Burial		-0.17	0.14	This study

691 *Positive rates indicate N inputs, negative rates indicate N removal and retention.
692
693