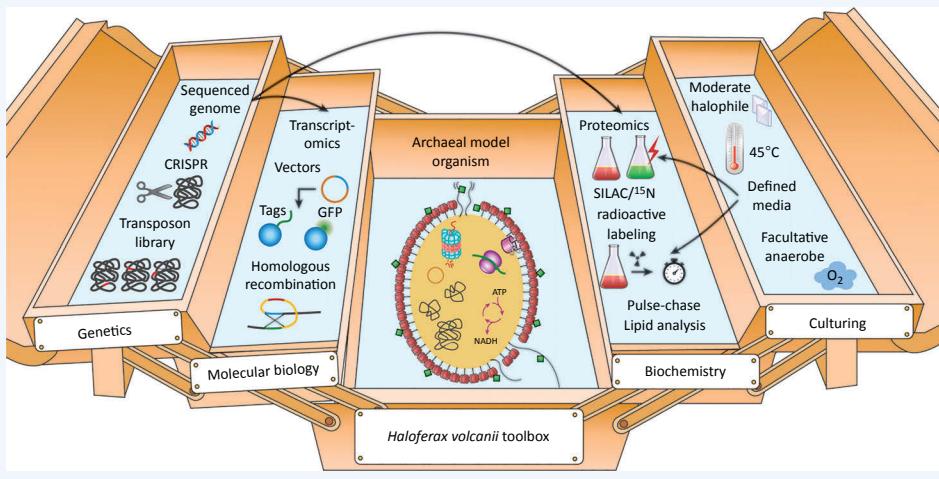


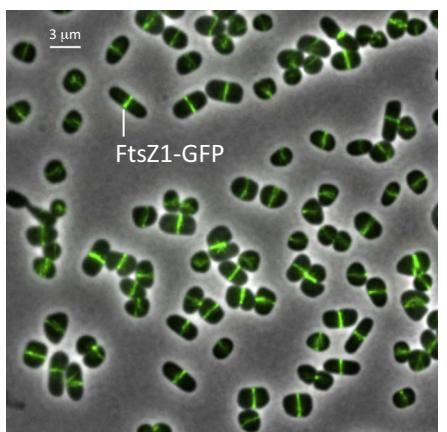
*Haloferax volcanii*Mechthild Pohlschroder^{1,*} and Stefan Schulze¹¹University of Pennsylvania, Philadelphia, PA 19104, USA

Trends in Microbiology

Isolated from the Dead Sea in 1975, *Haloferax volcanii* thrives in high salt environments and has emerged as an important archaeal model system. An extensive repertoire of genetic, molecular biological, and biochemical tools has been developed for this fast-growing, easily cultivated haloarchaeon, including expression vectors and gene-deletion strategies, including CRISPR. Its low mutation rate and ability to grow on defined media allow straightforward application of methods such as metabolic labeling, and the sequenced genome laid the foundation for transcriptomics and proteomics studies. These tools have allowed examination of key pathways such as transcription, noncoding RNAs, protein synthesis and degradation, protein glycosylation, motility, and biofilm formation. With the collaborative spirit of the *H. volcanii* community, this model system has become invaluable not only for enhancing our understanding of archaea but also for improving the development of biotech applications.

TAXONOMY AND CLASSIFICATION:

DOMAIN: Archaea
PHYLUM: Euryarchaeota
CLASS: Halobacteria
ORDER: Halobacterales
FAMILY: Halobacteriaceae
GENUS: *Haloferax*
SELECTED SPECIES: *Haloferax volcanii* DS2 – first sequenced strain



Trends in Microbiology

KEY FACTS:

First isolated by Benjamin Elazari Volcani.

Chemoorganotroph, uses polysaccharides, sugars, and amino acids as carbon sources.

Pleomorphic disk-shaped, or, when motile, rod-shaped cells.

First archaeon shown to transfer genes by a natural mating system.

Cells have one main chromosome (2.9 Mb), three secondary ones, (85–690 kb) and a plasmid (6.4 kb). They contain six origins of replication and 3996 genes encoding proteins.

Polypliod cells can contain up to 30 chromosome copies.

Archaea-specific features include, among others, archaea that drive motility, ether-linked isoprenoid membrane and cell wall composed of S-layer glycoprotein, the best-studied archaeal glycoprotein (N- and O-glycosylation), which is C-terminally lipid-anchored.

Bacteria-like CRISPR, transcription regulators, chemotaxis machinery, and extracytoplasmic proteases.

Eukaryote-like basal transcription factors, proteasome substrate targeting and degradation.

Several pathways, for example, thiamine biosynthesis, performed by chimera of eukaryote- and bacteria-like pathways.

About 50% of exported proteins are transported across the membrane via the Tat pathway; mostly predicted lipoproteins.

Survival in high salt depends on salt-in strategy maintaining homeostasis and low protein pI preventing misfolding.

Nonpathogen with simple growth conditions and low risk of contamination due to high salt media; great tool for K-16 experiments demonstrating principles of microbiology, genetics, and evolution.

*Correspondence:
 pohlschr@sas.upenn.edu (M. Pohlschroder).

Acknowledgments

Iain Duggin (University of Technology, Sydney) provided Figure 2, and Mike Dyall-Smith (University of Melbourne) and Friedhelm Pfeiffer (Max Planck Institute of Biochemistry, Martinsried) provided helpful comments.

Resources

www.haloarchaea.com/resources/halohandbook/

Literature

1. Mullakhanbhai, M.F. and Larsen, H. (1975) *Halobacterium volcanii* sp. nov., a Dead Sea halobacterium with a moderate salt requirement. *Arch. Microbiol.* 104, 207–214
2. Lam, W.L. and Doolittle, W.F. (1989) Shuttle vectors for the archaebacterium *Halobacterium volcanii*. *Proc. Natl. Acad. Sci. U. S. A.* 86, 5478–5482
3. Gophna, U. *et al.* (2017) Finally archaea get their CRISPR-Cas Toolbox. *Trends Microbiol.* 25, 430–432
4. Duggin, I.G. *et al.* (2014) CetZ tubulin-like proteins control archaeal cell shape. *Nature* 519, 362–365
5. Hartman, A.L. *et al.* (2010) The complete genome sequence of *Haloflexax volcanii* DS2, a model archaeon. *PLoS One* 5, e9605
6. Mevarech, M. and Werczberger, R. (1985) Genetic transfer in *Haloflexax volcanii*. *J. Bacteriol.* 162, 461–462
7. Zerulla, K. *et al.* (2014) DNA as a phosphate storage polymer and the alternative advantages of polyploidy for growth or survival. *PLoS One* 9, 94819
8. Pohlschroder, M. *et al.* (2018) Archaeal cell surface biogenesis. *FEMS Microbiol. Rev.* 42, 694–717
9. Kouassi, J.E. *et al.* (2017) Laboratory activity to promote student understanding of UV mutagenesis and DNA repair. *J. Microbiol. Biol. Educ.* 18, 18.1.16
10. McMillan, L.J. *et al.* (2018) Multiplex quantitative SILAC for analysis of archaeal proteomes: a case study of oxidative stress responses. *Environ. Microbiol.* 20, 385–401