

Predicting rates of consumer-mediated nutrient cycling by a diverse herbivore assemblage

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Abstract

Herbivores mediate the abundances of primary producers both from the top-down, by consuming them, and from the bottom-up, by recycling nutrients. Whereas the top-down effects of herbivores on algae in marine ecosystems are well-documented, less is known about their roles as mediators of local-scale nutrient availability. We conducted a series of surveys and measurements of tide pools and the grazers in those pools between October of 2016 and June of 2017 at an intertidal site on the coast of Southern California, USA (33° 35' 16.3" N, 117° 52' 1.5" W). We surveyed grazer abundances in the field, measured biomass of representatives from four different grazer groups (littorine snails, limpets, chitons, and turban snails), measured ammonium excretion rates, and quantified ammonium accumulation rates in tide pools at our study site. We found that different grazer groups were characterized by different per-biomass ammonium excretion rates. Some grazer groups – turban snails and chitons – contributed more ammonium than predicted by their biomass, whereas other grazer groups – littorine snails and limpets – contributed less ammonium than predicted by biomass. Because of these differences between grazer groups, ammonium accumulation rates in tide pools at our study site were effectively predicted based on the ammonium excretion rates of the different grazer groups. However, ammonium accumulation rates were not related to total herbivore biomass. Our results highlight the importance of grazer identity – and particularly the role of species such as turban snails that contribute disproportionately to nutrient recycling – in understanding the contributions of grazers as mediators of bottom-up processes in marine systems.

Key words: ammonium; bottom-up; diversity; grazer; herbivore; nitrogen; rocky intertidal

Introduction

Despite decades of work documenting the roles of marine herbivores in mediating the diversity and abundance of primary producers (e.g., Kitching and Ebling 1961; Lubchenco 1978; Nielsen 2001; Williams et al. 2013), previous work in benthic marine systems has often ignored the fact that herbivores not only consume algae, they also affect nutrient availability. Herbivores therefore affect primary producers not only from the top-down, via consumption, but also from the bottom-up, by excreting inorganic nutrients as waste products. For example, marine invertebrate herbivores excrete ammonium (Carpenter 1986; Taylor and Rees 1998; Bracken et al. 2014), thereby enhancing nutrient availability, algal growth (Bracken et al. 2014) and productivity (Carpenter 1986). Nitrogen is an important limiting nutrient in coastal marine systems (Ryther and Dunstan 1971; Corwith and Wheeler 2002), so predicting rates of consumer-mediated nitrogen recycling is essential to understanding nutrient availability and limitation in marine ecosystems.

However, most marine systems are characterized by diverse consumer assemblages, which makes predictions of those consumers' roles as mediators of nutrient availability potentially difficult (Burkepile et al. 2013; Layman et al. 2013). For example, Taylor and Rees (1998) found that ammonium excretion rates of a diverse assemblage of mobile epifauna in seaweed beds are nonlinearly related to invertebrate body mass, suggesting that mass- or taxon-specific ammonium excretion rates are necessary in order to predict the contribution of a diverse invertebrate assemblage to nutrient availability. Furthermore, McIntyre et al. (2008) quantified ammonium excretion rates of 47 co-occurring species of freshwater fish and found that even after accounting for body mass, there were significant differences between species' ammonium excretion rates.

We took a similar approach to understanding the role of herbivores as mediators of local-scale nutrient availability in a rocky intertidal ecosystem. Recent work has highlighted the importance of local-scale consumer-mediated nutrient loading in mediating the diversity (e.g., Bracken and Nielsen 2004) and growth (e.g., Bracken 2004; Pfister 2007; Aquilino et al. 2009) of algae on rocky shores, but much of that work has focused either on a single consumer species (e.g., the snail *Littorina littorea*; Bracken et al. 2014) or on consumers that do not actually eat the algae that benefit from the nutrients that they excrete (e.g., the mussel *Mytilus californianus*; Bracken 2004; Bracken and Nielsen 2004; Pfister 2007; Aquilino et al. 2009). Given the high diversity of herbivores in many rocky shore systems (Nielsen 2001; O'Connor et al. 2015), there is a clear need to evaluate the collective role of these consumers as not only top-down consumers of algae but also as bottom-up facilitators.

We addressed this knowledge gap by using a combination of field surveys, measurements of biomass and ammonium excretion of different herbivore species, and measurements of ammonium accumulation rates in tide pools to evaluate the role of a diverse herbivore assemblage in mediating nutrient availability on a Southern California rocky shore. Local-scale nutrient inputs are likely to be especially important in this system, as ambient inorganic nitrogen concentrations in the adjacent nearshore ocean are generally low ($< 5 \mu\text{mol L}^{-1}$) and have been declining for the past several years (Martiny et al. 2016). Based on previous work, we hypothesized that different grazer groups would be characterized by different ammonium excretion rates. We therefore predicted that the rate of ammonium accumulation in the field would be better predicted based on the ammonium excretion rates of the component species and not by the total biomass of herbivores in the tide pools.

Materials and Methods

Study location and species

Surveys, field measurements, and collections were conducted in a set of natural tide pools in a rocky reef at Corona del Mar State Beach on the coast of Southern California, USA (33° 35' 16.3" N, 117° 52' 1.5" W). All research was conducted between October of 2016 and June of 2017 under California Department of Fish and Wildlife Scientific Collecting Permit SCP-13405. The rocky substratum at the site is composed of sedimentary Monterey formation intertidal reefs. Volumes of the tide pools in this study averaged $X \pm SE = 22.6 \pm 3.6$ L, $n = 18$, and bottom surface areas averaged 0.46 ± 0.05 m².

These tide pools were very high on the shore; tidal elevations, determined using a self-leveling laser level (CST/berger, Watseka, Illinois, USA), averaged $X \pm SE = 1.58 \pm 0.04$ m above the local tidal datum at mean lower-low water. Based on predicted tide heights at the entrance to Newport Bay (Flater 1998), 1.1 km from our study site, these tide pools were only submerged for 5.8% of the time over the year immediately preceding our measurements. In the absence of waves, based on these predictions, tide pools were submerged only during the highest tides of each month, and they were often isolated for several days at a time during neap tides. The median isolation time for these pools in the absence of wave splash was 23.0 h, but waves reduced the typical duration of pool emersion (M. Bracken, *personal observation*). Thus, given a wave swell height of 0.5 m, which is not unusual at our study location (O'Reilly et al. 2016), tide pools would be either washed over or covered for 32.6% of the time, with a median isolation time of 8.3 h.

Despite their elevation on the shore, these pools were characterized by a diverse invertebrate assemblage primarily composed of herbivorous gastropods. These included littorine

snails (*Littorina scutulata* [Gould 1849] and *Littorina plena* [Gould 1849]), limpets (*Lottia limatula* [Carpenter 1864], *L. scabra* [Gould 1846], and *L. strigatella* [Carpenter 1864]), chitons (*Cyanoplax hartwegii* [Carpenter 1855] and *Nuttalina californica* [Reeve 1847]), and turban snails (*Tegula funebris* [A. Adams 1854] and *T. gallina* [Forbes 1850]). Collectively, these grazing mollusks represented the vast majority of invertebrates in the tide pools, though pools also contained occasional mussels (e.g., *Brachidontes adamsianus* [Dunker 1856] and *Mytilus californianus* [Conrad 1837]), hermit crabs (e.g., *Pagurus samuelis* [Stimpson 1857]), and sea anemones (*Anthopleura* spp.). None of these invertebrates were abundant enough, relative to the grazers, to appreciably modify nutrient availability. Other, smaller invertebrates, such as copepods and amphipods, were rare to absent in the pools. Macroalgae were also virtually absent from the tide pools, and the grazers primarily consumed periphyton.

Grazer abundances and attributes

We surveyed grazer abundances in 18 tide pools by spreading a flexible mesh net across the bottom of each pool (Foulweather Trawl Supply, Newport, Oregon, USA; Bracken and Nielsen 2004). The net was composed of 10 cm × 10 cm squares and facilitated both counting of grazers and measurement of tide pool surface area. Grazer abundances were divided by the volume of each tide pool (i.e., m^3), as we were interested in the potential for grazers to mediate the concentration ($\mu\text{mol L}^{-1}$) of ammonium in the pools.

We collected 10 representative individuals each of *Littorina scutulata/plena* (these species are not differentiable in the field), *Lottia limatula*, *Lottia scabra*, *Lottia strigatella*, *Cyanoplax hartwegii*, *Nuttalina californica*, *Tegula funebris*, and *Tegula gallina* from the tide pools. Samples were representative of the size range of each taxon present in the tide pools. We

dried individuals to constant mass at 60 °C, weighed them, combusted them for 4 h in a muffle furnace at 450 °C, and weighed them again to calculate mean ash-free dry mass values for each species. These were then averaged to calculate mean values (mg ind⁻¹) for each grazer group: littorine snails, limpets, chitons, and turban snails.

Ammonium excretion rates of members of each grazer group were evaluated in microcosms containing 200 mL of saltwater (salinity of 35; Instant Ocean® Sea Salt, Blacksburg, Virginia, USA). Each microcosm contained a travertine tile that approximated the sedimentary Monterey formation rocks at our study site. Water in the microcosms was not stirred or aerated in order to simulate a still-water tide pool environment. Temperatures were maintained at 20 °C to ensure constant conditions across experimental trials that were representative of field conditions. Grazers were collected from the field immediately prior to experimental measurements of ammonium accumulation. We made sure that the individuals collected were representative of the size range present in the tide pools. A consistent biomass of grazers ($X \pm SE = 0.91 \pm 0.01$ g) was added to each microcosm at the beginning of each experimental trial. Because members of the different grazer groups were characterized by different individual masses (Table 1), maintaining a constant mass across grazer groups necessitated different

Table 1. Biomass (ash-free dry mass) and ammonium (NH₄⁺) excretion rates of tide pool herbivores. Values are $X \pm SE$.

Grazer group	Biomass	NH ₄ ⁺ excretion	
	(mg ind ⁻¹)	(μmol h ⁻¹ ind ⁻¹)	(μmol h ⁻¹ g ⁻¹)
Littorine snails	6.8 ± 0.5	0.021 ± 0.004	3.1 ± 0.6
Limpets	20.6 ± 1.8	0.046 ± 0.015	2.2 ± 0.7
Chitons	58.3 ± 11.1	0.357 ± 0.359	6.1 ± 1.0
Turban snails	183.8 ± 11.6	1.751 ± 0.353	9.5 ± 1.9

145 numbers of individuals for each group in each microcosm: 135 littorine snails, 45 limpets, 15
146 chitons, or 5 turban snails. During trials, microcosms were covered with flexible window screen
147 mesh secured by a rubber band to prevent escapes and limit external sources of potential
148 ammonium contamination.

149 Microcosm trials were run for ~19 h, which was between the median isolation time of
150 field tide pools in the absence of wave splash (23.0 h) and after accounting for a 0.5 m swell
151 height (8.3 h). Initial water samples ($n = 2$) were taken from each microcosm prior to adding the
152 grazers, and a second set of samples was taken at the end of the trial. The ammonium
153 concentration in the water samples was analyzed using the phenolhypochlorite method
154 (Solórzano 1969) on a UV-1800 benchtop spectrophotometer (Shimadzu, Carlsbad, California,
155 USA). Ammonium accumulation rates were calculated on both a per-individual ($\mu\text{mol h}^{-1} \text{ ind}^{-1}$)
156 and per-biomass ($\mu\text{mol h}^{-1} \text{ g}^{-1}$) basis based on the change in ammonium concentrations over
157 time. Ammonium accumulation rates were measured in $n = 8$ microcosms for each grazer group,
158 split into two trials of $n = 4$ replicate microcosms each. Changes in ammonium concentrations in
159 an equivalent number of control microcosms without grazers were minimal and were accounted
160 for when calculating ammonium accumulation rates. Initial ammonium concentrations averaged
161 $\bar{X} \pm \text{SE} = 0.9 \pm 0.2 \mu\text{mol L}^{-1}$.

162 Assessing changes in ammonium concentrations using only two points assumes a linear
163 relationship between ammonium accumulation and time. To verify this assumption, we
164 conducted a second set of trials where we collected water samples over time (0.00, 0.25, 0.50,
165 1.00, 2.00, 4.00, 6.00, and 22.75 h) instead of only at the beginning and end of trials. We
166 conducted these trials for littorine snails and turban snails because they are the most abundant
167 grazer groups in the tide pools, collectively composing > 90% of grazer biomass in the field. We

compared linear and saturating (Michaelis-Menten) fits to the relationship between ammonium concentration ($\mu\text{mol L}^{-1}$) and time (h) using the corrected Akaike Information Criterion (AIC_c; Burnham and Anderson 2002) and found that a linear relationship always provided a better fit to the data.

Relative contributions to ammonium accumulation and biomass

Relative contributions of different grazer groups to ammonium accumulation rates and biomass were calculated based on the per-individual rates of ammonium excretion ($\mu\text{mol h}^{-1} \text{ ind}^{-1}$, Table 1), the average biomass of each individual (mg ind^{-1} , Table 1), and the total number of individuals of each grazer group in each of tide pools ($n = 18$) at Corona del Mar State Beach. We estimated the total ammonium accumulation rate in each tide pool by multiplying the abundance of each grazer in that pool by the measured ammonium excretion rate for that grazer group. These values were then summed across the four grazer groups. The relative contribution of each grazer group to the total ammonium accumulation rate was then calculated as a percentage of the total. Similarly, we estimated the total biomass of grazers in each tide pool by multiplying the average biomass of the members of each grazer group by the number of individuals of that grazer group quantified in our field surveys. The relative contribution of each grazer group to total biomass was then calculated as a percentage of that total. We calculated the difference between each grazer group's contribution to excretion and its contribution to biomass by subtracting, for each tide pool, the percentage contribution to biomass from the contribution to excretion.

Predicting the contribution of a diverse grazer assemblage to ammonium accumulation

We quantified ammonium accumulation rates in tide pools ($n = 5$) at Corona del Mar State Beach over a single day-time low tide. We deliberately chose a subset of pools that were fully submerged during high tide and then isolated by the receding tide in the morning. Waves were relatively large that day, so pools were only completely isolated from the ocean for ~6 h before they were splashed again by the combination of waves and the incoming tide. We collected water samples from each pool every hour while the pools were isolated and used the slope of the relationship between ammonium concentration ($\mu\text{mol L}^{-1}$) and time (h) to calculate the observed rate of ammonium accumulation ($\mu\text{mol L}^{-1} \text{ h}^{-1}$). We also counted and identified all grazers in those pools on that day.

We compared ammonium accumulation rates measured in the field with predictions based on (1) the total estimated biomass of grazers in each pool or (2) the ammonium accumulation rate in the pool predicted based on the measured ammonium excretion rates of each grazer group. Total estimated biomass was calculated by multiplying the average biomass of the members of each grazer group (Table 1) by the number of individuals of that grazer group quantified in our field surveys on that day. The predicted ammonium accumulation rate was estimated by multiplying the average ammonium excretion rate of the members of each grazer group ($\mu\text{mol h}^{-1} \text{ ind}^{-1}$, Table 1) by the number of individuals of that grazer group quantified in our field surveys.

Statistical analyses

Data were primarily analyzed using general linear models (PROC GLM) and t tests in SAS v. 9.4 (SAS Institute 2012), including regression and ANOVA, after verifying that the data met the assumptions of normality and homogeneity of variances. Analyses included evaluations

of mean individual mass as a function of grazer group (littorine snails, limpets, chitons, and turban snails) and ammonium excretion rates (both per-individual and per-biomass) as a function of trial (as a blocking factor) and grazer group. Relative contributions of each grazer group to biomass versus ammonium accumulation were assessed for each grazer group by subtracting, for each tide pool, the calculated % contribution to biomass from the % contribution to excretion. Averages for the $n = 18$ tide pools were then compared to zero using one-sample t tests. The metabolic scaling relationship between biomass (mg ind^{-1}) and ammonium excretion rate ($\mu\text{mol h}^{-1} \text{ind}^{-1}$) was evaluated by taking the logarithm (\log_{10}) of the mean biomass and excretion rate of each herbivore group, then quantifying the relationship between them using a general linear model (i.e., $\log_{10}[\text{excretion}] = a \cdot \log_{10}[\text{biomass}] + b$; Glazier 2005). Of particular interest was the slope of this relationship (a), which corresponds to the scaling exponent. Observed rates of ammonium accumulation in the field were evaluated as either a function of (1) predicted ammonium accumulation rates based on measured excretion rates of the different grazer groups or (2) estimated total grazer biomass. These relationships were evaluated using general linear models.

Results

Grazer abundances and attributes

Littorine snails were by far the most numerically abundant grazers in tide pools at Corona del Mar State Beach (Fig. 1A); the number of *Littorina scutulata/plena* individuals per volume was two orders of magnitude higher than the number of any other grazer group, and they were the most abundant grazer in 17 of the 18 tide pools. However, littorines were also the smallest of the grazers, with average masses 1-2 orders of magnitude lower than the other grazer groups

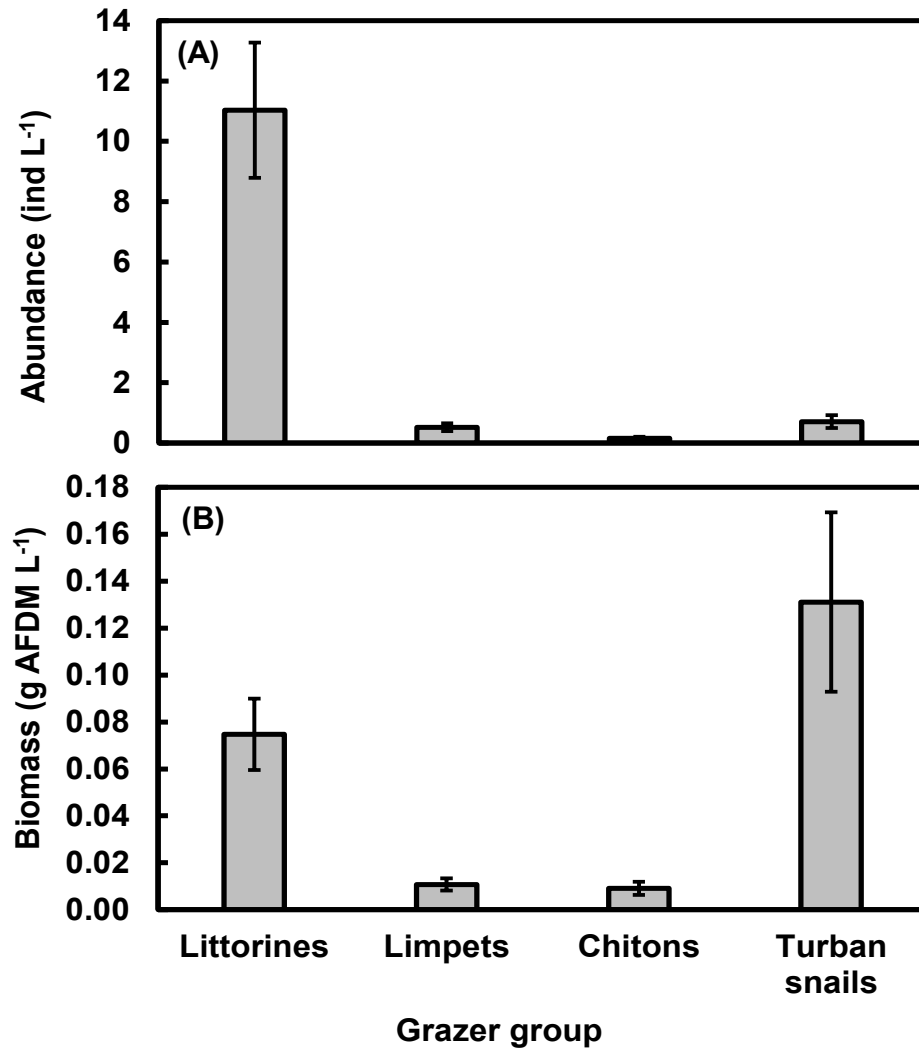


Figure 1. Abundance and biomass estimates of tide pool herbivores. (A) Littorine snails (*Littorina scutulata/plena*) were by far the most numerically abundant herbivores in the tide pools (ind L⁻¹). (B) Turban snails (*Tegula* spp.) were the herbivores with the highest total biomass in the tide pools (g ash-free dry mass [AFDM] L⁻¹), followed by littorine snails, limpets (*Lottia* spp.), and chitons (*Nuttalina fluxa* and *Cyanoplax hartwegii*). Note that counts of different herbivore species in tide pools were non-independent of each other, so statistical comparisons were not made for abundance or biomass. In both panels, values are $X \pm SE$.

(Table 1). Grazer groups, thus, differed substantially in mass (ANOVA, $F(3,56) = 149.97$, $P < 0.001$). Turban snails were the heaviest grazers, followed by chitons, limpets, and littorines, though littorine and limpet masses were statistically indistinguishable (Tukey test, $P > 0.05$). Thus, despite their relatively low abundances, the majority of the biomass in tide pools was composed of turban snails, followed by littorines, limpets, and chitons (Fig. 1B).

The different grazer groups also differed substantially with respect to their ammonium excretion rates on both a per-individual basis (ANOVA, $F(3,27) = 21.3$, $P < 0.001$; Table 1) and a per-biomass basis (ANOVA, $F(3,27) = 7.6$, $P < 0.001$; Table 1, Fig. 2). Turban snails excreted the most ammonium, both per-individual and per-biomass, excreting at rates 3-4 times higher than those of littorine snails and limpets (Table 1).

Relative contributions to ammonium accumulation and biomass

Relative contributions of different grazer groups to ammonium accumulation rates and biomass were expressed as average percentage contributions of each group to the total biomass and the total ammonium accumulation in the tide pools. Turban snails made the greatest contribution to total ammonium excretion rates, followed by littorine snails, chitons, and limpets (Fig. 3A). In contrast, littorines made the greatest contribution to biomass, followed by turban snails, limpets, and chitons (Fig. 3A).

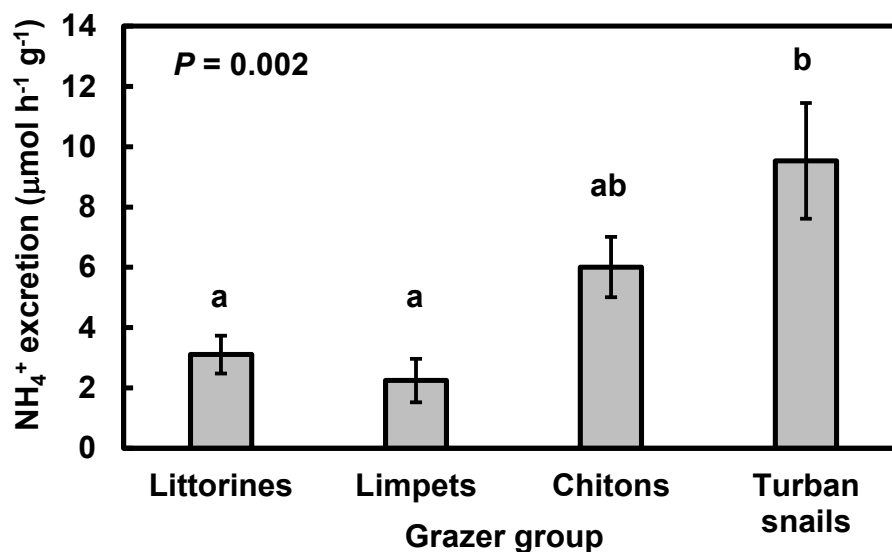


Figure 2. Ammonium (NH_4^+) excretion rates of tide pool herbivores. Per-biomass ammonium excretion rates ($\mu\text{mol h}^{-1} \text{g}^{-1}$) differed substantially between different herbivore groups ($P = 0.002$). Letters indicate statistically significant differences ($P < 0.05$) after Tukey correction for multiple comparisons. Values are $\bar{X} \pm \text{SE}$.

Two grazer groups, turban snails (one-sample t test, $t_{17} = 5.2$, $P < 0.001$) and chitons (one-sample t test, $t_{17} = 3.0$, $P = 0.008$), were predicted – based on laboratory excretion measurements and abundances in tide pools – to make greater contributions to ammonium

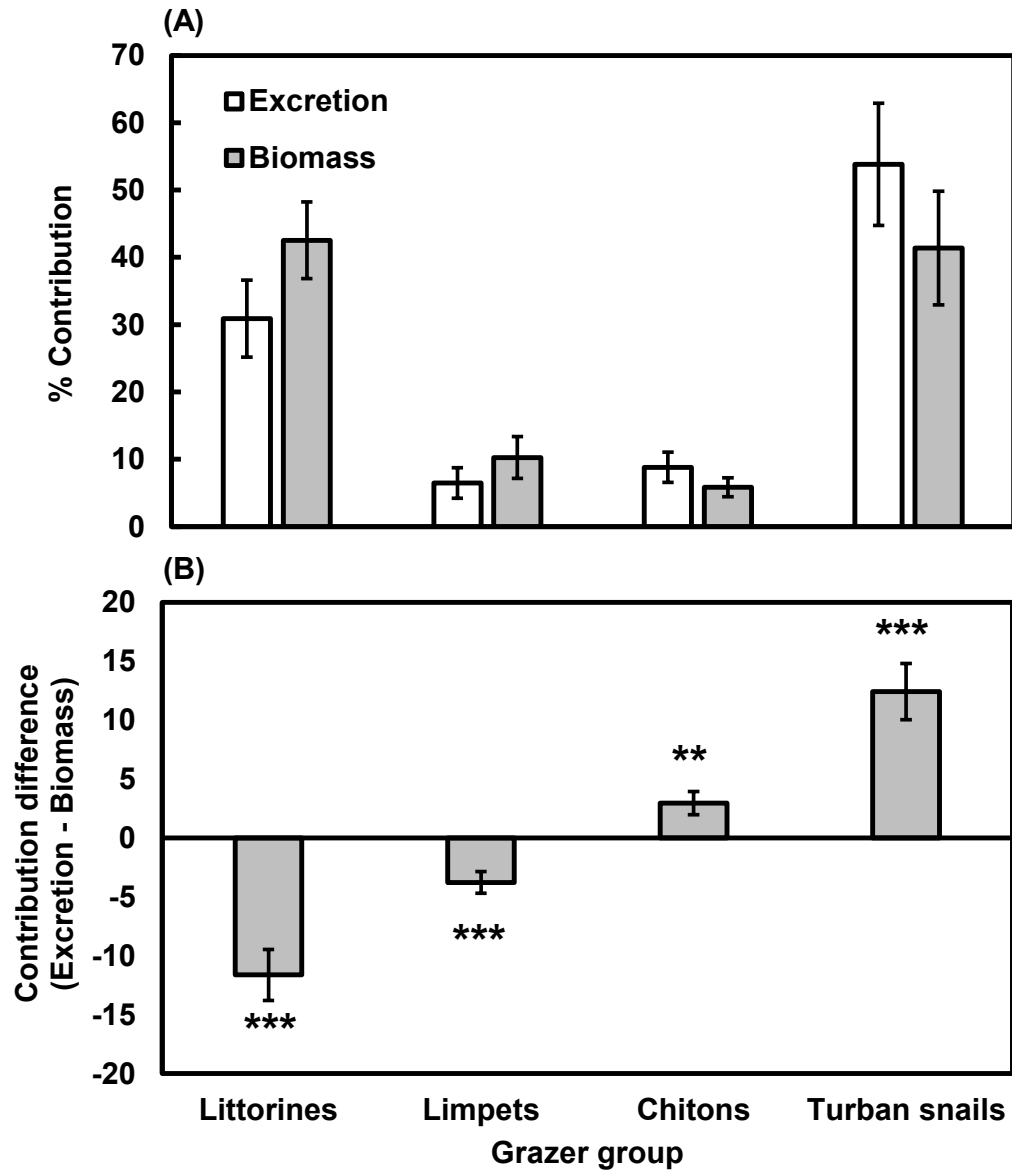


Figure 3. Estimated contributions of different herbivores to ammonium excretion rates and total herbivore biomass. (A) Percent contributions of each grazer group to the estimated total excretion rate and biomass in 18 tide pools on a California rocky shore. (B) Differences (excretion *minus* biomass) between grazer group contributions to excretion and biomass. A positive value (> 0) indicates that a group's predicted contribution to excretion exceeds its contribution to biomass, whereas a negative value (< 0) indicates that a group's predicted contribution to excretion is less than its contribution to biomass. Asterisks indicate statistically significant differences from zero: $P < 0.001$ (***) and $P < 0.01$ (**). In both panels, values are $X \pm SE$.

accumulation rates in tide pools than predicted based on their biomass (Fig. 3B). In contrast, the other two groups, littorines (one-sample t test, $t_{17} = 5.4$, $P < 0.001$) and limpets (one-sample t test, $t_{17} = 4.1$, $P < 0.001$), were predicted to make lesser contributions to ammonium accumulation than predicted based on biomass (Fig. 3B).

We evaluated the relationship between individual grazers' excretion rates ($\mu\text{mol h}^{-1} \text{ ind}^{-1}$) and biomasses (mg ind^{-1}) on a log-log plot (Fig. 4). This transformation linearized the curvilinear relationship between the variables and provided insights into the nature of the scaling relationship. Excretion increased with biomass ($r^2 = 0.97$) with a scaling exponent (the slope of the relationship) of 1.40.

Predicting the contribution of a diverse grazer assemblage to ammonium accumulation

We quantified rates of ammonium accumulation over time in $n = 5$ tide pools at Corona del Mar State Beach (Fig. 5). Ammonium accumulated at a rate of $X \pm \text{SE} = 0.16 \pm 0.06 \mu\text{mol}$

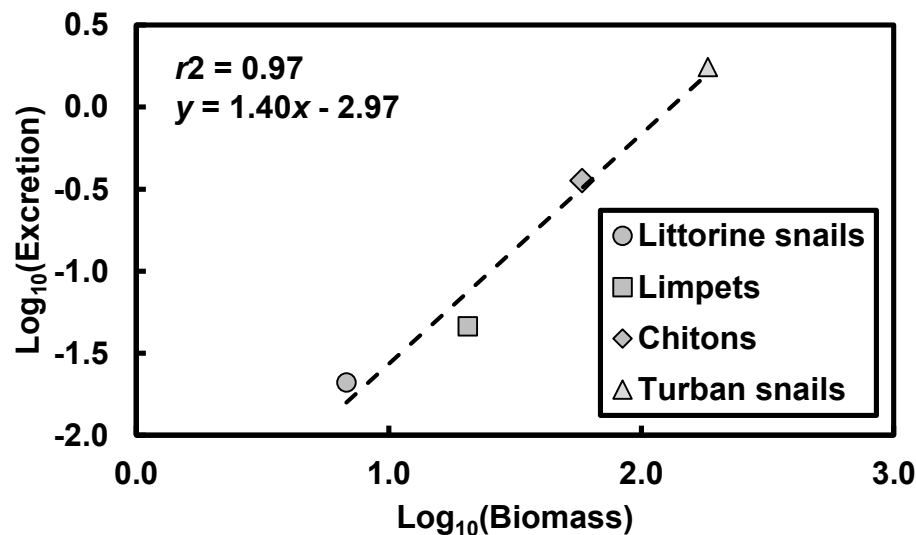


Figure 4. Log-log plot of biomass vs. excretion rates of individual herbivores. Excretion rates (originally measured in $\mu\text{mol h}^{-1} \text{ ind}^{-1}$) were linearly related to biomass (originally measured in $\text{mg ash-free dry mass ind}^{-1}$) when both variables are plotted on logarithmic scales. The slope of this relationship (1.40) represents the scaling relationship in the power curve.

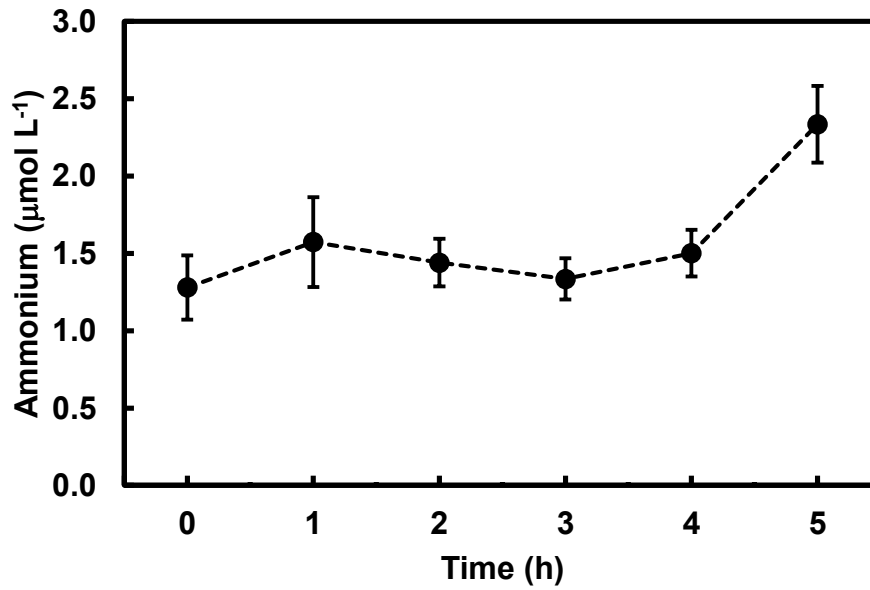


Figure 5. Ammonium concentrations in tide pools. Ammonium ($\mu\text{mol L}^{-1}$) accumulated in pools after they were isolated by the receding tide. Values are $X \pm \text{SE}$ of samples taken from $n = 5$ tide pools at Corona del Mar State Beach.

271 $\text{L}^{-1} \text{h}^{-1}$ (one-sample t test, $t_4 = 14.6$, $P < 0.001$). Five hours after pools were isolated, average
 272 ammonium concentrations in the pools were higher than the concentration in the adjacent ocean
 273 (one-sample t test, $t_4 = 5.4$, $P = 0.006$). The rate of ammonium accumulation in those tide pools
 274 was predicted more effectively by incorporating the excretion rates of the different grazer groups
 275 (Fig. 6A; Linear regression, $r^2 = 0.81$, $F_{1,3} = 13.1$, $P = 0.036$) than by the total estimated
 276 biomass of the grazers present in the tide pools (Fig. 6B; Linear regression, $r^2 = 0.42$, $F_{1,3} = 2.2$,
 277 $P = 0.236$). Note, however, that even when rates of ammonium accumulation were linearly
 278 related to rates predicted based on ammonium excretion by the component species (i.e., Fig. 6A),
 279 accumulation rates were substantially lower than predicted based on excretion rates. If
 280 ammonium accumulation rates matched predicted rates, they would fall on the dashed line in Fig.
 281 6A. Instead, measured accumulation rates were $< 30\%$ of predicted rates.

282 In these tide pools, littorines composed $X \pm SE = 68 \pm 14\%$ of the biomass but
 283 contributed only $55 \pm 19\%$ of the ammonium accumulation. In contrast, turban snails composed
 284 only $31 \pm 14\%$ of the biomass but contributed $45 \pm 19\%$ of the ammonium.

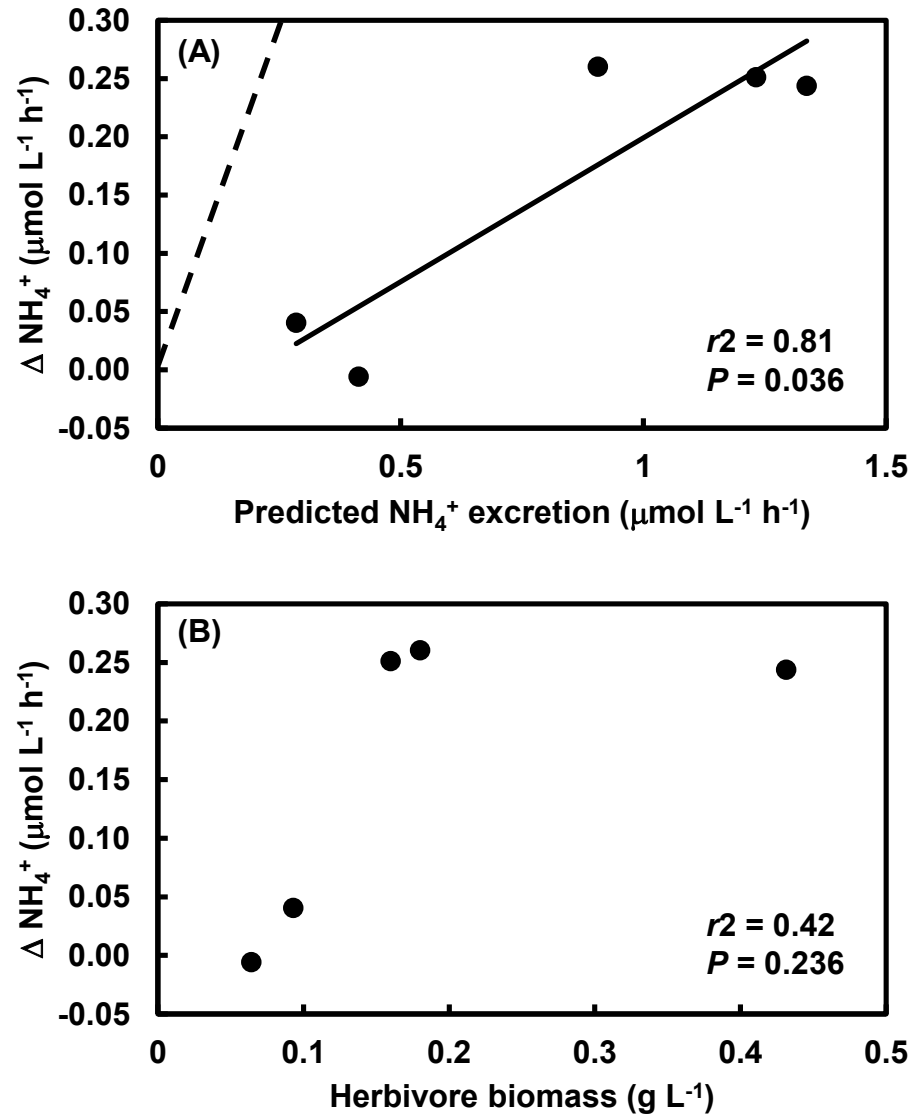


Figure 6. Measured rates of ammonium (NH_4^+) accumulation as functions of predicted ammonium excretion rates and estimated total herbivore biomass. (A) Rates of NH_4^+ accumulation ($\mu\text{mol L}^{-1} \text{h}^{-1}$) in $n = 5$ tide pools were strongly correlated to excretion rates estimated using measured rates of the grazer groups in each tide pool ($r^2 = 0.81$, $P = 0.036$). The dashed line indicates the rate of ammonium accumulation predicted by laboratory excretion rates. (B) Measured NH_4^+ accumulation rates were not correlated to estimates of total herbivore biomass in the pools ($r^2 = 0.42$, $P = 0.236$).

Discussion

Different grazer groups were characterized by different per-biomass ammonium excretion rates. Some groups (i.e., turban snails, chitons) contributed more to ammonium accumulation than predicted based on biomass, whereas others (i.e., littorine snails, limpets) contributed less to ammonium excretion than predicted by their biomass. These differences are supported by the scaling relationship between ammonium excretion ($\log_{10}[\text{excretion}]$) and biomass ($\log_{10}[\text{biomass}]$). The scaling exponent of 1.40 is higher than the typical value of ~ 0.75 (i.e., the $3/4$ power law; Glazier 2005). In general, mass-specific metabolic rates tend to decline as individual body mass increases (i.e., scaling exponents < 1). However, values > 1 are not uncommon, and scaling exponents > 2 have been reported for invertebrates (Glazier 2005). These “positively allometric” relationships occur when larger organisms have higher mass-specific metabolic rates. This is the pattern we observed here, where the largest grazers (*Tegula* spp.) were also characterized by the highest excretion rates. One grazer group, the limpets, deviated from the regression line on the log-log plot, with lower values than the other three groups. This reflects limpets’ lower contribution to ammonium accumulation rates – relative to the other grazer groups – than expected based on biomass. Note also that our comparisons were made across species, whereas most comparisons are based on scaling relationships calculated within species (Glazier 2005). For example, Carey et al. (2013) suggested that differences in the scaling exponents of six chiton species were related to differences in activity, metabolism, and habitat. Temperature can also modify scaling exponents (Glazier 2005), though our measurements of excretion were all measured at a constant, field-relevant temperature.

Measured rates of ammonium accumulation in tide pools on the shore were therefore better predicted based on the ammonium excretion rates of the component grazer groups than by their estimated total biomass. These results support our hypothesis that different grazer groups would be characterized by different ammonium excretion rates (e.g., Bray et al. 1988) and align well with previous findings, especially from freshwater systems, that taxon-specific ammonium excretion rates are necessary in order to predict spatial variation in nutrient cycling (McIntyre et al. 2008).

However, observed rates of ammonium accumulation were < 30% of predicted rates. This discrepancy between observed and predicted rates of ammonium accumulation may be explained by uptake of ammonium by periphyton in the tide pools. Despite the apparent lack of primary producers in these tide pools – there are few to no macroalgae in them – the pools are highly productive; rates of net primary production ($\text{mg O}_2 \text{ L}^{-1} \text{ h}^{-1}$) are equivalent to those that we have measured in macroalgae-dominated pools (M. Bracken and G. Bernatchez, *unpublished data*). The periphyton in the pools are likely taking up substantial quantities of excreted ammonium, as has been observed in other intertidal systems (Longphuir et al. 2009). We have observed appreciable differences between ammonium fluxes in the dark (accumulation) and light (uptake) in these tide pools, further supporting the role of periphyton in mediating ammonium availability (M. Bracken and G. Bernatchez, *unpublished data*). Subsequent work could include experimental ammonium additions to tide pools with and without grazers to evaluate whether uptake by periphyton can account for the difference between observed and predicted rates of ammonium accumulation.

What mechanisms could potentially underlie the observed differences between species with respect to ammonium excretion rates? One possibility is dietary specificity. However, little

is known about the diets of these co-occurring herbivores. All of the grazer groups consume diatoms (Castenholz 1961; Best 1964; Nicotri 1977; LaScala-Gruenewald et al. 2016), but the identities of the resources available (likely a diverse mixture of benthic microalgae, cyanobacteria, and macroalgal sporelings [LaScala-Gruenewald et al. 2016]) remain unknown at our study site. Furthermore, there is little evidence for resource partitioning among co-occurring molluscan grazers (Nicotri 1977; Hawkins et al. 1989; LaScala-Gruenewald et al. 2016). Thus, underlying mechanisms for differences in per-biomass nutrient recycling rates, which include not only diet but also organismal physiology, remain unknown.

Regardless of the underlying mechanism, it is clear that some species contribute more than others to ammonium accumulation rates in tide pools, and the loss of these species may have disproportionate effects on nutrient availability. McIntyre et al. (2007) describe how consumer extinctions can influence nutrient cycling and highlight the fact that the loss of certain vulnerable species (e.g., those targeted by humans) may have particularly large effects on nutrient availability. Populations of large, conspicuous gastropods such as turban snails have declined in Southern California due to human impacts (Murray et al. 1999). We demonstrated that turban snails contributed substantially more ammonium than predicted based on their biomass. Thus, whereas turban snails represented less than a third of the total herbivore biomass in the tide pools where we measured ammonium accumulation rates, they contributed nearly half of the ammonium.

More generally, we found that different functional groups of grazers differ with respect to their effects on an important biogeochemical processes. Understanding the roles of species in ecosystems (Lawton 1994) is essential for predicting rates of nutrient cycling and other biogeochemical rates (Naeem 2002). Intertidal grazers play an essential role in marine

ecosystems by converting organic nitrogen in the algae that they eat into inorganic nitrogen that can be readily taken up and assimilated by primary producers (Giannotti and McGlathery 2001; Bracken et al. 2014). And – given differences between grazer species in their ammonium excretion rates – a diverse grazer assemblage (e.g., one that contains groups such as turban snails and chitons characterized by higher rates of per-biomass nitrogen excretion) may be more effective at recycling nutrients.

Our study also adds another dimension to the body of research that links trophic complexity, biodiversity, and ecosystem functioning. Many studies in marine systems have demonstrated that more diverse grazer assemblages are more effective at controlling algal biomass (Duffy et al. 2003, 2015; Matthiessen et al. 2007; Eklöf et al. 2012). If those grazers also contribute nutrients – and especially if grazer diversity affects not only top-down control but bottom-up facilitation by grazers – then a mechanistic understanding of the effects of grazer diversity on primary producers requires partitioning grazers' consumptive and facilitative effects (Bracken et al. 2014).

One important caveat regarding our work is that our measurements and surveys were conducted in tide pools, which are isolated at low tide, allowing ammonium to accumulate (Bracken and Nielsen 2004). Tide pools are functionally field mesocosms – they contain most species present on local rocky shores and are amenable to measuring nutrient excretion and uptake rates and conducting experimental manipulations (Nielsen 2001; Bracken and Nielsen 2004; Pfister 2007) – but they are also hydrodynamically different from wave-swept shores and nearshore systems, where excreted nitrogen is likely to be advected away. Macroalgae were also virtually absent from these tide pools due to a combination of grazing activity and environmental stress. The microalgal biofilms in the pools likely assimilated much of the ammonium from the

water column – observed rates of ammonium accumulation were < 30% of the predicted rates – but the simplicity of the system probably enhanced our ability to link observed and predicted rates of ammonium accumulation. However, consumer-mediated nutrient inputs are important even in subtidal and wave-exposed intertidal habitats (Taylor and Rees 1998; Aquilino et al. 2009), suggesting that our findings are relevant to a broader suite of marine systems.

In conclusion, we have shown that grazers are important local-scale contributors of nitrogen to intertidal habitats. Thus, in addition to their traditional top-down role, grazers play potentially important roles in nutrient cycling. Because different groups in diverse grazer assemblages are characterized by different rates of per-biomass ammonium excretion, predicting rates of grazer-mediated ammonium accumulation requires measurement of the ammonium excretion rates of each grazer group. However, once these data are incorporated, ammonium accumulation rates in the field can be effectively predicted. Understanding the roles of consumers in ecosystems – including not only consumption but also facilitation – is essential for understanding marine biodiversity and ecosystem functioning.

Compliance with Ethical Standards

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References

- Adams A (1854) Descriptions of twenty-seven new species of shells from the collection of Hugh Cuming, Esq. Proc Zool Soc Lond 22: 311-317. doi: 10.1111/j.1469-7998.1854.tb07283.x
- Aquilino KM, Bracken MES, Faubel MN, Stachowicz JJ (2009) Local-scale nutrient regeneration facilitates seaweed growth on wave-exposed rocky shores in an upwelling system. Limnol Oceanogr 54: 309–317. doi: 10.4319/lo.2009.54.1.0309.
- Best B (1964) Feeding activities of *Tegula funebris*. Veliger 6 (suppl): 42-45.
- Bracken MES (2004) Invertebrate-mediated nutrient loading increases growth of an intertidal macroalga. J Phycol 40: 1032-1041. doi: 10.1111/j.1529-8817.2004.03106.x.
- Bracken MES, Dolecal RE, Long JD (2014) Community context mediates the top-down vs. bottom-up effects of grazers on rocky shores. Ecology 95: 1458-1463. doi 10.1890/13-2094.1d.

- 421 Bracken MES, Nielsen KJ (2004) Diversity of intertidal macroalgae increases with nutrient
422 loading by invertebrates. *Ecology* 85: 2828-2836. doi: 10.1890/03-0651.
- 423 Bray RN, Miller AC, Johnson S, Krause PR, Robertson DL, Westcott AM (1988) Ammonium
424 excretion by macroinvertebrates and fishes on a subtidal rocky reef in southern
425 California. *Mar Biol* 100: 21-30. doi: 10.1007/bf00392951.
- 426 Burkepile DE, Allgeier JE, Shantz AA, Pritchard CE, Lemoine NP, Bhatti LH, Layman CA
427 (2013) Nutrient supply from fishes facilitates macroalgae and suppresses corals in a
428 Caribbean coral reef ecosystem. *Sci Rep* 3: 1493. doi: 10.1038/srep01493
- 429 Burnham KP, Anderson DR (2002) Model selection and inference: a practical information-
430 theoretic approach. 2nd ed. Springer-Verlag, New York.
- 431 Carey N, Sigwart JD, Richards JG (2013) Economies of scaling: more evidence that allometry of
432 metabolism is linked to activity, metabolic rate and habitat. *J Exp Mar Biol Ecol* 439: 7-
433 14. doi: 10.1016/j.jembe.2012.10.013.
- 434 Carpenter PP (1855) Descriptions of (supposed) new species and varieties of shells, from the
435 Californian and west Mexican coasts, principally in the collection of Hugh Cuming, Esq.
436 *Proc Zool Soc Lond* 23: 228-235. doi: 10.1111/j.1469-7998.1855.tb00330.x.
- 437 Carpenter PP (1864) Supplementary report on the present state of our knowledge with regard to
438 the Mollusca of the west coast of North America. *Rep Br Assoc Adv Sci* 33: 517-686.
- 439 Carpenter RC (1986) Partitioning herbivory and its effect on coral reef algal communities. *Ecol*
440 *Monogr* 56: 345-363. doi: 10.2307/1942551.
- 441 Castenholz RW (1961) The effect of grazing on marine littoral diatom populations. *Ecology* 42:
442 783-794. Doi: 10.2307/1933507.

- 443 Conrad TA (1837) Description of new marine shells, from Upper California. Collected by
 444 Thomas Nuttall, Esq. J Acad Nat Sci Phila 7: 227-268.
- 445 Corwith HL, Wheeler PA (2002) El Niño related variations in nutrient and chlorophyll
 446 distributions off Oregon. Prog Oceanogr 54: 361-380. doi: 10.1016/S0079-
 447 6611(02)00058-7.
- 448 Duffy JE, Reynolds PL, Boström C, Coyer JA, Cusson M, Donadi S, Douglass JG, Eklöf JS,
 449 Engelen AH, Eriksson BK (2015) Biodiversity mediates top-down control in eelgrass
 450 ecosystems: a global comparative-experimental approach. Ecol Lett 18: 696-705. doi:
 451 10.1111/ele.12448.
- 452 Duffy JE, Richardson JP, Canuel EA (2003) Grazer diversity effects on ecosystem functioning in
 453 seagrass beds. Ecol Lett 6: 637-645. doi: 10.1046/j.1461-0248.2003.00474.x.
- 454 Dunker G (1856) Mytilacea nova collectionis Cumingianae. Proc Zool Soc Lond 24: 358-366.
 455 doi: 10.1111/j.1469-7998.1856.tb00343.x.
- 456 Eklöf JS, Alsterberg C, Havenhand JN, Sundbäck K, Wood HL, Gamfeldt L (2012)
 457 Experimental climate change weakens the insurance effect of biodiversity. Ecol Lett 15:
 458 864-872. doi: 10.1111/j.1461-0248.2012.01810.x.
- 459 Flater D (1998) XTide v. 2.13. FlaterCo, Germantown, Maryland, USA.
- 460 Giannotti AL, McGlathery KJ (2001) Consumption of *Ulva lactuca* (Chlorophyta) by the
 461 omnivorous mud snail *Ilyanassa obsoleta* (Say). J Phycol 37: 209-215. doi:
 462 10.1046/j.1529-8817.2001.037002209.x.
- 463 Glazier, DS (2005) Beyond the ‘3/4-power law’: variation in the intra-and interspecific scaling of
 464 metabolic rate in animals. Biol Rev 80: 611-662. doi: 10.1111/j.1469-
 465 185X.2009.00095.x.

- 466 Gould AA (1846) Description of new shells, collected by the United States Exploring
467 Expedition, and belong to the genus *Patella*. Proc Boston Soc Nat Hist 2: 148-152.
- 468 Gould AA (1849) Descriptions of the following new species of shells, brought home by the U.S.
469 Exploring Expedition. Proc Boston Soc Nat Hist 3: 83-85.
- 470 Hawkins SJ, Watson DC, Hill AS, Harding SP, Kyriakides MA, Hutchinson S, Norton TA
471 (1989) A comparison of feeding mechanisms in microphagous, herbivorous, intertidal
472 prosobranchs in relation to resource partitioning. J Mollusc Stud 55: 151-165. doi:
473 10.1093/mollus/55.2.151.
- 474 Kitching JA, Ebling FJ (1961) The ecology of Lough Ine: XI. the control of algae by
475 *Paracentrotus lividus* (Echinoidea). J Anim Ecol 30: 373-383. doi: 10.2307/2304.
- 476 LaScala-Gruenewald DE, Miller LP, Bracken MES, Allen BJ, Denny MW (2016) Quantifying
477 the top-down effects of grazers on a rocky shore: selective grazing and the potential for
478 competition. Mar Ecol Prog Ser 553: 49-66. doi: 10.3354/meps11774.
- 479 Lawton JH (1994) What do species do in ecosystems? Oikos 71: 367-374. doi: 10.2307/3545824.
- 480 Layman CA, Allgeier JE, Yeager LA, Stoner EW (2013) Thresholds of ecosystem response to
481 nutrient enrichment from fish aggregations. Ecology 94: 530-536. doi: 10.1890/12-
482 0705.1.
- 483 Longphuiert SN, Lim J-H, Leynaert A, Claquin P, Choy E-J, Kang C-K, An S (2009) Dissolved
484 inorganic nitrogen uptake by intertidal microphytobenthos: nutrient concentrations, light
485 availability and migration. Mar Ecol Prog Ser 379: 33-44. doi:
486 10.1016/j.ecss.2007.04.025.

- 487 Lubchenco J (1978) Plant species diversity in a marine intertidal community: importance of
488 herbivore food preference and algal competitive abilities. *Am Nat* 112: 23-39. doi:
489 10.1086/283250.
- 490 Martiny AC, Talarmin A, Mouginot C, Lee JA, Huang JS, Gellene AG, Caron DA (2016)
491 Biogeochemical interactions control a temporal succession in the elemental composition
492 of marine communities. *Limnol Oceanogr* 61: 531-542. doi: 10.1002/lno.10233.
- 493 Matthiessen B, Gamfeldt L, Jonsson PR, Hillebrand H (2007) Effects of grazer richness and
494 composition on algal biomass in a closed and open marine system. *Ecology* 88: 178-187.
495 doi: 10.1890/0012-9658(2007)88[178:EOGRAC]2.0.CO;2.
- 496 McIntyre PB, Flecker AS, Vanni MJ, Hood JM, Taylor BW, Thomas SA (2008) Fish
497 distributions and nutrient cycling in streams: can fish create biogeochemical hotspots?
498 *Ecology* 89: 2335-2346. doi: 10.1890/07-1552.1.
- 499 McIntyre PB, Jones LE, Flecker AS, Vanni MJ (2007) Fish extinctions alter nutrient recycling in
500 tropical freshwaters. *Proc Natl Acad Sci USA* 104: 4461-4466. doi:
501 10.1073/pnas.0608148104.
- 502 Murray SN, Denis TG, Kido JS, Smith JR (1999) Human visitation and the frequency and
503 potential effects of collecting on rocky intertidal populations in southern California
504 marine reserves. *Calif Coop Ocean Fish Invest Rep* 40: 100-106.
- 505 Naeem S (2002) Ecosystem consequences of biodiversity loss: the evolution of a paradigm.
506 *Ecology* 83: 1537-1552. doi: 10.1890/0012-9658(2002)083[1537:ECOBLT]2.0.CO;2.
- 507 Nicotri ME (1977) Grazing effects of four marine intertidal herbivores on the microflora.
508 *Ecology* 58: 1020-1032. doi: 10.2307/1936922.

- 509 Nielsen KJ (2001) Bottom-up and top-down forces in tide pools: test of a food chain model in an
510 intertidal community. *Ecol Monogr* 71: 187-217. doi: 10.1890/0012-
511 9615(2001)071[0187:BUATDF]2.0.CO;2.
- 512 O'Connor NE, Bracken MES, Crowe TP, Donohue I (2015) Nutrient enrichment alters the
513 consequences of species loss. *J Ecol* 103: 862-870. doi: 10.1111/1365-2745.12415.
- 514 O'Reilly WC, Olfe CB, Thomas J, Seymour RJ, Guza RT (2016) The California coastal wave
515 monitoring and prediction system. *Coast Eng* 116: 118-132. doi:
516 10.1016/j.coastaleng.2016.06.005.
- 517 Pfister CA (2007) Intertidal invertebrates locally enhance primary production. *Ecology* 88: 1647-
518 1653. doi: 10.1890/06-1913.1.
- 519 Reeve LA (1847) Monograph of the genus *Chiton*. In: *Conchologia iconica*, or, illustrations of
520 the shells of molluscos animals. Reeve, Benham, and Reeve, London
- 521 Ryther JH, Dunstan WM (1971) Nitrogen, phosphorus, and eutrophication in the coastal marine
522 environment. *Science* 171: 1008-1013. doi: 10.1126/science.171.3975.1008.
- 523 SAS Institute (2012) SAS version 9.4. SAS Institute, Cary, North Carolina, USA.
- 524 Solórzano L (1969) Determination of ammonia in natural waters by the phenolhypochlorite
525 method. *Limnol Oceanogr* 14: 799-801. doi: 10.4319/lo.1969.14.5.0799.
- 526 Stimpson W (1857) Notices of new species of Crustacea of western North America; being an
527 abstract from a paper to be published in the journal of the Society. *Proc Bost Soc Nat Hist*
528 6: 84-89.
- 529 Taylor RB, Rees TAV (1998) Excretory products of mobile epifauna as a nitrogen source for
530 seaweeds. *Limnol Oceanogr* 43: 600-606. doi: 10.4319/lo.1998.43.4.0600.

531 Williams SL, Bracken MES, Jones E (2013) Additive effects of physical stress and herbivores on
532 intertidal seaweed biodiversity. *Ecology* 94: 1089-1101. doi: 10.1890/12-0401.1.