

ORIGINAL RESEARCH

WILEY



Environmental factors function as constraints on soil nitrous oxide fluxes in bioenergy feedstock cropping systems

David S. Duncan¹ | Lawrence G. Oates¹ | Ilya Gelfand^{2,3} | Neville Millar^{2,3} | G. Philip Robertson^{2,3} | Randall D. Jackson¹

¹Department of Agronomy, DOE-Great Lakes Bioenergy Research Center, University of Wisconsin-Madison, Madison, Wisconsin

²W. K. Kellogg Biological Station, Michigan State University, Hickory Corners, Michigan

³Department of Plant, Soil, and Microbial Sciences, DOE-Great Lakes Bioenergy Research Center, Michigan State University, East Lansing, Michigan

Correspondence

Lawrence G. Oates, Department of Agronomy, DOE-Great Lakes Bioenergy Research Center, University of Wisconsin-Madison, Madison, WI.
Email: oates@wisc.edu

Present Address

Ilya Gelfand, The French Associates Institute for Agriculture and Biotechnology of Drylands, The Jacob Blaustein Institutes for Desert Research, Ben-Gurion University of the Negev, Beer-Sheva, Israel

Funding information

Michigan AgBioResearch; National Science Foundation LTER, Grant/Award Number: DEB 1027253; DOE BER Office of Science, Grant/Award Number: DE-FC02-07ER64494; DOE OBP Office of Energy Efficiency and Renewable Energy, Grant/Award Number: DE-AC05-76RL01830

Abstract

Nitrous oxide (N₂O) is a potent greenhouse gas and major component of the net global warming potential of bioenergy feedstock cropping systems. Numerous environmental factors influence soil N₂O production, making direct correlation difficult to any one factor of N₂O fluxes under field conditions. We instead employed quantile regression to evaluate whether soil temperature, water-filled pore space (WFPS), and concentrations of soil nitrate (NO₃⁻) and ammonium (NH₄⁺) determined upper bounds for soil N₂O flux magnitudes. We collected data over 6 years from a range of bioenergy feedstock cropping systems including no-till grain crops, perennial warm-season grasses, hybrid poplar, and polycultures of tallgrass prairie species each with and without nitrogen (N) addition grown at two sites. The upper bounds for soil N₂O fluxes had a significant and positive correlation with all four environmental factors, although relatively large fluxes were still possible at minimal values for nearly all factors. The correlation with NH₄⁺ was generally weaker, suggesting it is less important than NO₃⁻ in driving large fluxes. Quantile regression slopes were generally lower for unfertilized perennials than for other systems, but this may have resulted from a perpetual state of nitrogen limitation, which prevented other factors from being clear constraints. This framework suggests efforts to reduce concentrations of NO₃⁻ in the soil may be effective at reducing high-intensity periods—"hot moments"—of N₂O production.

KEYWORDS

bioenergy, biogeochemical cycling, cropping systems, greenhouse gas, hot moments, nitrous oxide, quantile regression

1 | INTRODUCTION

Nitrous oxide (N₂O) is a major contributor to global radiative forcing (Forster et al., 2007) and is currently the single most important ozone-depleting substance in the atmosphere

(Portmann, Daniel, & Ravishankara, 2012). Agricultural soils are responsible for 77% of N₂O emissions in the United States (U.S. Environmental Protection Agency, 2018) and 55% of global emissions (Hu, Chen, & He, 2015). N₂O emissions can reduce, or even completely negate, the benefits of

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2018 The Authors. *GCB Bioenergy* Published by John Wiley & Sons Ltd.

fossil fuel displacement for many bioenergy feedstock cropping systems (Crutzen, Mosier, Smith, & Winiwarter, 2008; Robertson, Paul, & Harwood, 2000), making their management and mitigation of N_2O production a major part of assessing net long-term environmental impact (Gelfand & Robertson, 2015). This is especially important for dedicated biomass production cropping systems such as miscanthus plantations or harvested native grass fields (Gelfand et al.,), whose appeal rests heavily on their anticipated positive environmental benefits (Landis et al., 2017; Werling et al., 2014). Many studies report markedly different N_2O emissions levels among cropping systems, with lower emissions frequently observed in perennial, species-rich, or minimally fertilized systems (Gelfand, Shcherbak, Millar, Kravchenko, & Robertson, 2016; Niklaus, Wardle, & Tate, 2006; Oates et al., 2016; Stehfest & Bouwman, 2006). It is less clear whether these differences primarily derive from environmental conditions (e.g., higher soil N) or how agroecosystems respond to those conditions, with major implications for predicting their behavior under novel conditions.

The microbial processes of nitrification (ammonia [NH_3] or ammonium [NH_4^+] oxidation) and denitrification (nitrate [NO_3^-] reduction), respectively, produce N_2O as a by-product/intermediate product, which can be emitted from the soil if not fully consumed (Butterbach-Bahl, Baggs, Dannenmann, Kiese, & Zechmeister-Boltenstern, 2013; Robertson & Tiedje, 1987). In soils that are regularly oxygen-limited, denitrification produces the majority of N_2O (Mathieu et al., 2006). Nitrification and denitrification rates are usually limited by environmental factors, increasing with substrate concentrations (NH_3 , NO_3^- , and labile carbon), oxygen availability, water-filled pore space (WFPS), temperature, and pH (Wallenstein, Myrold, Firestone, & Voytek, 2006). Some factors, including substrates and temperature, monotonically increase rates of N_2O -generating processes while others such as WFPS and pH have optimal values for different processes (Hénault et al., ; Wallenstein et al., 2006).

Studies linking environmental factors directly to N_2O production have found these factors vary substantially among regions (Dechow & Freibauer, 2011) and cropping systems (Gelfand et al., 2016). Similarly, studies such as a Bayesian recalibration of the nitrous oxide emission (NOE) module of the agroecosystem model CERES by Lehuger et al. (2009) and a meta-analysis of 14 published models by Surendran Nair et al. (2012) show responses to environmental factors differing among sites or cropping systems. Thus, while the biology of N_2O production is well understood, predicting and modeling this process in a particular agroecological context remains challenging.

Much of the difficulty in monitoring, modeling, and managing soil N_2O arises from “hot moments,” high-intensity short-duration bursts of activity occurring when multiple environmental factors align to create near-ideal conditions for

denitrification (Groffman et al., 2009). For example, fertilizer application followed by heavy rain and high temperatures creates a substrate-rich, oxygen-limited environment with high microbial activity. These events produce flux orders of magnitude greater than is typical of the system and can contribute 25%–50% of cumulative annual N_2O emissions (Flesch et al., 2018; Saha et al., 2017; Zenone et al., 2016). This dynamic likely contributes to the high interannual variability of cumulative annual N_2O emissions (Oates et al., 2016) and makes it challenging to accurately characterize the probable range of annual emissions without multiple years of measurement.

The prevalence of hot moments suggests N_2O -generating processes may be subject to the ecological law of the minimum (Hiddink & Kaiser, 2005), wherein the rate of a process separately constrained by multiple factors is determined by the single most limiting factor. Conventional regression, which evaluates the central tendency of the process rate for a given value of a predictor (the conditional mean), would perform poorly under such conditions, as the response of each observation would be independent of all but the single most limiting factor. In contrast, quantile regression can be used to evaluate the maximum observed value for the process rate for a given value of the predictor (the conditional upper limit; Cade & Noon, 2003). If the upper limit for N_2O flux magnitudes changes with the value of an environmental factor, it would suggest that factor serves as a constraint on N_2O production.

We correlated individual N_2O flux measurements with paired environmental measurements collected from seven bioenergy feedstock cropping systems at two sites over a 6 year period (Oates et al., 2016). These cropping systems covered multiple dimensions of agroecological intensity including perennality, plant species richness, and nitrogen addition, allowing us to explore relationships with environmental drivers across a wide range of relevant field conditions. Using quantile regression, we evaluated two hypotheses: (a) that soil temperature, water-filled pore space, and concentrations of NO_3^- and NH_4^+ all constrained the upper limits of N_2O flux measurements, and (b) that the nature of these constraints differed among the cropping systems in our study. This work provides a framework for an alternative approach to relating N_2O fluxes to environmental measurements in field studies.

2 | MATERIALS AND METHODS

2.1 | Experimental design and study sites

We conducted this study on the DOE Great Lakes Bioenergy Research Center's Bioenergy Cropping Systems Experiment (BCSE), an agronomic trial situated at the W. K. Kellogg Biological Station Long-term Ecological Research site (KBS, 42°23'47"N, 85°22'26"W, 288 m a.s.l.) in Michigan and the Arlington Agricultural Research Station (ARL,

43°17'45"N, 89°22'48"W, 315 m a.s.l.) in Wisconsin. The BCSE consisted of 10 systems. Three systems were phases of a corn (*Zea mays* L.)–soybean (*Glycine max* L.)–canola (*Brassica napus* L.) rotation between 2009 and 2011; between 2012 and 2014, these systems consisted of continuous corn and phases of a corn–soybean rotation, all with a rye (*Secale cereale* L.) and Austrian winter pea (*Pisum sativum* L.) cover crop (Supporting Information Table S1). The remaining treatments were in place throughout the measurement period and consisted of continuous no-till corn without a cover crop, monocultures of switchgrass (*Panicum virgatum* L.), miscanthus (*Miscanthus × giganteus*), and hybrid poplar (*Populus nigra* × *P. maximowiczii* “NM6”), a five species native grass mix, an early successional field recruited from the pre-existing seedbank, and an 18 species restored tallgrass prairie. Species and variety information is presented in Supporting Information Table S1 of Oates et al. (2016).

All treatments were planted in 27 × 43 m plots in a five-replicate randomized complete block design ($n = 5$ blocks) and managed with field-scale equipment. Annual grain systems were managed following recommendations from local university extension programs. The poplar system was fertilized in 2010 (210 kg N/ha as 34-0-0 granular ammonium nitrate) and harvested by coppicing during the 2013–2014 winter. Microplots (10 × 43 m) were established in all other systems to test effects of N addition. The restored prairie microplot and main plots of all other systems received annual spring N addition (56 kg N/ha as 34-0-0 granular ammonium nitrate). The main restored prairie plot and microplots of all other systems received no added N. Aboveground biomass in these systems was harvested to 10 cm residual stubble height following the first frost event in the fall. N-addition dates for all systems are given in Supporting Information Table S2, and full details on agronomic management were presented in Sanford et al. (2016).

Soils at KBS are primarily Kalamazoo loam (USDA soil classification: Fine-Loamy, Mixed, Semiactive, Mesic Typic Hapludalfs). Mean annual temperature from 1981 to 2010 was 9.9°C, and mean annual precipitation was 1,027 mm (MSCO, 2013). Prior to BCSE establishment, the field was planted to alfalfa (*Medicago sativa* L.) and a corn–soybean rotation. The switchgrass, native grass mix, and restored prairie treatments at KBS suffered seed loss following flooding in 2008 and were reseeded in 2009. Soils at ARL are predominantly Plano silt loam (USDA classification: Fine-Silty, Mixed, Superactive, Mesic Typic Arguidolls). Mean annual temperature from 1981 to 2010 was 6.8°C, and mean annual precipitation was 869 mm (NWS, 2013). Pre-BCSE land use differed among blocks: the corn phase of a corn–soybean rotation (blocks A1–A3) or an alfalfa–orchardgrass (*Dactylis glomerata* L.) hay mixture (blocks A4–A5). We replanted miscanthus at ARL in 2010 following stand loss during the 2008/2009 winter.

2.2 | Data generation

Nitrous oxide was measured biweekly during the growing season, with additional sampling following fertilization and major precipitation events. Sampling frequency was reduced during the winter, particularly at KBS and in the early years of the study. Static chambers were used to estimate trace gas emissions, with one chamber per plot/microplot. Chambers were cylindrical (28.5 cm diameter, ~17 cm effective height, ~10 L volume) and inserted to a soil depth of ~5 cm. We ensured adequate headspace mixing by selecting an appropriately low volume:surface area ratio and by keeping vegetation trimmed within chambers (Livingston, Hutchinson, & Spartalian, 2006). Chamber lids were fitted with a septum for gas extraction and a 2 mm diameter vent tube for pressure equilibration. For each chamber, four headspace gas samples of 30 ml were collected: immediately upon chamber closure then subsequently at three ~20 min intervals. Samples were placed in glass 5.9 ml Exetainer vials (Labco Limited, Buckinghamshire, UK), using 20 ml of sample to flush the vial before overpressurizing with another 10 ml. Following gas chromatography, CO₂ concentration was detected using an infrared gas analyzer (IRGA, LiCor 820, Lincoln, NE, USA) and N₂O concentration was detected using an electron capture detector (micro-ECD, Agilent 7890A GC System, Santa Clara, CA, USA). We avoided ECD cross-sensitivity issues by using an argon–methane carrier gas and setting the detector temperature to 350°C (Wang, Wang, & Ling, 2010).

Prior to estimation of N₂O fluxes, CO₂ accumulation curves were visually inspected for outlier samples indicating compromised vial integrity or other mechanical errors, removing these outliers. In time series with four valid measurements, nonlinearity of fluxes was evaluated using the *HMR* package (v0.3.1, Pedersen, 2015) in the R statistical environment (v3.5.0, R Core Team, 2018). Following this classification, time series received a secondary visual inspection focused on identifying outlier samples in N₂O concentrations, particularly those that might drive a nonlinear fit. Nonlinear flux estimates from the *HMR()* function were used for samples that passed this secondary inspection without any data removal and whose nonlinear estimate was outside the 95% confidence interval for the linear flux estimate. All other samples used linear flux estimates. All flux observations were used for analysis.

From 2010 onward, soil cores (3.7 cm diameter, 15 cm depth at ARL, 25 cm depth at KBS) were collected concurrently with trace gas sampling. Inorganic soil N was extracted from a 10 g field-moist subsample using 2 M KCl following Robertson, Sollins, Ellis, and Lajtha (1999). Ammonium (NH₄⁺) and nitrate (NO₃[−]) concentrations were determined using a Flow Solution 3100 segmented flow injection analyzer (OI Analytical, College Station, TX, USA), using USEPA methods 27200110 and 27190110, respectively. The

instrument has a detection limit of 0.05 $\mu\text{g N/g soil}$, which we used as a floor for concentrations.

Soil temperature was measured at the time of trace gas sampling using a 15 cm temperature probe (Checktemp 1C, Hanna Instruments, Smithfield, RI, USA). Soil moisture was measured at KBS by determining gravimetric water content (GWC) for the soil N samples. At ARL, moisture was measured as volumetric water content (VWC) within 1 m of the static chamber using a time domain reflectometer with 20 cm rods (FieldScout 300, Spectrum Technologies, Plainfield, IL, USA). Bulk density was measured for all plots in 2008 and 2013. We estimated annual changes in bulk density by linear

interpolation, calculating mean values for groups of cropping systems (three groups: annual grain crops, poplar, and all other systems) and sets of blocks (four sets: A1 and A3, A2, A4 and A5, and all KBS blocks) with similar distributions of measurements. Water-filled pore space (WFPS) was estimated from bulk density (Bd) and soil particle density (Pd, assumed to be a constant 2.65 g/cm^3):

$$\text{WFPS} = \text{VWC} \times \left(1 - \frac{\text{Bd}}{\text{Pd}}\right) = \text{GWC} \times \text{Bd} \times \left(1 - \frac{\text{Bd}}{\text{Pd}}\right).$$

For our analyses, we constrained WFPS values to be $\leq 100\%$, as deviations between local bulk density and the

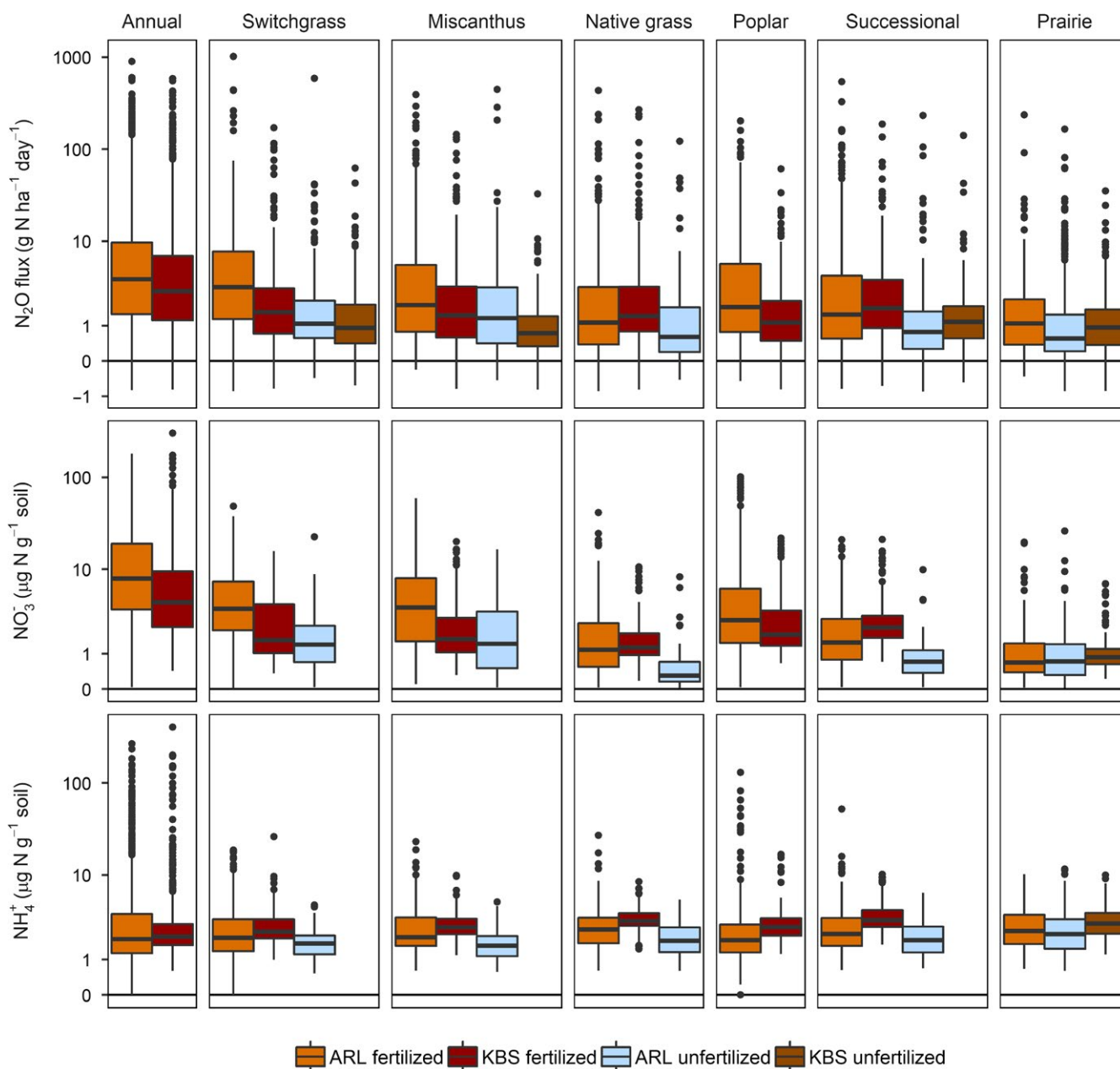


FIGURE 1 Effects of cropping system, site, and N addition (fertilized) on distributions of N_2O fluxes and concentrations of soil inorganic N. Values are plotted on an inverse hyperbolic sine scale. Median indicated by thick black line, box indicates the 75th percentile, and whiskers indicate the 25th percentile

average values we used could result in estimated WFPS values >100%.

2.3 | Data analysis

Nitrous oxide fluxes, NH_4^+ , and NO_3^- values all varied over orders of magnitude and exhibited a strong right skew; to mitigate this, observations were inverse hyperbolic sine (IHS) transformed prior to analysis. This transformation commonly used in the social sciences to handle overdispersed variables (Burbidge, Magee, & Robb, 1988) but can also be used in the natural sciences when values near or below 0 are relevant (Sekhon et al.,). This allowed us to include negative N_2O fluxes, which are periodically observed (Molodovskaya et al., 2012) and to avoid amplifying measurement errors for values close to the detection limit of our instruments. Following this transformation, median values were centered between the 25th and 75th percentiles (Figure 1 and Supporting Information Figure S1).

All analyses were conducted in the R statistical environment. Graphics were generated using the *ggplot2* package (Wickham, 2009). Boxplot quantiles (Figure 1, Supporting Information Figures S1 and S2) and LOESS fits (Supporting Information Figure S3) were generated using the default settings of *geom_boxplot()* and *stat_smooth()*. Quantile regression was carried out using the *rq()* function in the *quantreg* package (Koenker, 2018) using $\tau = 0.95$ and estimating standard errors using the kernel method. Briefly, quantile regression operates similarly to regular regression but with asymmetrically weighted errors with the parameter tau indicating the quantile used to calculate weights. In our case, $\tau = 0.95$ meant 5% of observations would be above the regression line and their errors would be weighted to match the 95% of observations below the regression. Analogous to regular regression, a slope of 0 signifies the τ th quantile of the response is the same for all values of the predictor, while

a significant slope indicates the maximum values observed in the response will depend on the predictor. The statistical significance of differences between nested model structures was evaluated with *anova.rq()*, using a Wald test. Statistical significances of differences among cropping systems slopes were evaluated by setting the annual fertilized systems (described in Results) as the baseline and assessing the significance of interaction terms.

3 | RESULTS

3.1 | Evaluating the range and depth of N_2O flux and environmental factor measurements

Our dataset consisted of 9,542 individual N_2O flux measurements with at least one paired measurement of NH_4^+ , NO_3^- , WFPS, or soil temperature, with 4,273 observations having all four factors (Table 1). Soil inorganic N data were most limited, with no data in 2009 and reduced data collection frequency from 2013 onward. We analyzed the annual cropping systems as a group (rotations and rotational phases described in Supporting Information Table S1), as they had nearly identical distributions within a site for all five measured variables (Supporting Information Figure S1).

Within cropping systems, N_2O flux measurements varied over multiple orders of magnitude (Figure 1). Fertilized perennial systems at both sites overlapped considerably with the annual systems, although annual systems had a much higher prevalence of high fluxes. Similarly, unfertilized perennial systems tended to have lower and less variable distributions of fluxes than their fertilized counterparts. Similar patterns were visible in NO_3^- concentrations, although with a smaller range. The effects of fertilization at ARL were more pronounced, as were differences between annuals and perennials at both sites. In contrast, NH_4^+ concentrations were less dynamic across cropping systems and fertilization levels,

Year	Site	N_2O flux	Soil inorganic N	Soil temperature	Water-filled pore space
2009	ARL	692	—	684	663
	KBS	495	—	483	447
2010	ARL	887	640	834	864
	KBS	499	301	316	351
2011	ARL	925	820	922	875
	KBS	634	456	486	483
2012	ARL	1,388	1,254	1,388	1,320
	KBS	594	431	460	456
2013	ARL	1,387	311	1,386	1,383
	KBS	733	40	460	565
2014	ARL	1,334	130	1,229	1,194
	KBS	1,004	118	659	581

TABLE 1 Number of N_2O flux and environmental parameter samples collected for each site and year

although values tended to be higher and less variable at KBS than at ARL. Cropping system differences in soil temperature and WFPS were minimal relative to within-system variability, although WFPS was much higher at ARL than at KBS (Supporting Information Figure S2). At low concentrations, NO_3^- and NH_4^+ were minimally correlated, although at high levels they were somewhat correlated in annual systems (Supporting Information Figure S3).

3.2 | Environmental factors correlate to upper bounds of N_2O fluxes

We used quantile regression to independently evaluate the correlation between the four environmental factors and the upper limit of N_2O fluxes (Figure 2). Modeling separate slope and intercept terms for each site significantly improved fits for NH_4^+ ($F_{2,4497} = 10.9$, $p < 0.05$), WFPS ($\text{Wald } F_{2,9178} = 321.6$, $p < 0.05$), and soil temperature ($\text{Wald } F_{2,9417} = 22.1$, $p < 0.05$) but not NO_3^- ($\text{Wald } F_{2,4497} = 1.2$, $p = 0.30$). In all cases, regression slopes were positive and significant at $p < 0.05$, signifying that the maximum observed N_2O flux magnitude increased at higher levels of all four environmental factors. Minimal fluxes were observed at high levels of all factors, suggesting individual factors were not sufficient to drive high fluxes. There may have been insufficient observations at the upper range of NH_4^+ and NO_3^- concentrations to appropriately support quantile regression. Even after IHS transformation, NH_4^+ measurements had a

long rightward tail. Observations with these extremely high NH_4^+ values had fluxes below the upper limit predicted by quantile regression, suggesting a factor other than NH_4^+ limited N_2O production. NO_3^- was more evenly distributed across its range, providing greater support for its role as a limiting factor. The clearest difference between sites was observed with WFPS, with the coarse texture of the soil at KBS restricting the range of values that could be observed. The high concentration of soil temperature values near 0°C resulted from higher sampling intensity outside of the growing season at that site (Table 1).

3.3 | Environmental factor constraints differ broadly among cropping systems

Fitting separate quantile regression slopes and intercepts for each cropping system-fertilization combination significantly improved model performance (all Wald test $p < 0.05$). Based on the slope and intercept parameters by cropping system and site (Supporting Information Figures S4 and S5), we aggregated cropping systems into three groups: annual crops, fertilized perennials, and unfertilized perennials. The unfertilized systems were the only ones that did not have significantly positive slopes for all environmental factors, with a neutral slope for soil temperature at both sites and a negative one for NH_4^+ at ARL (Figure 3). At ARL, perennial systems had lower slopes than annual systems in their response to NH_4^+ and soil temperature. There was more inconsistency at

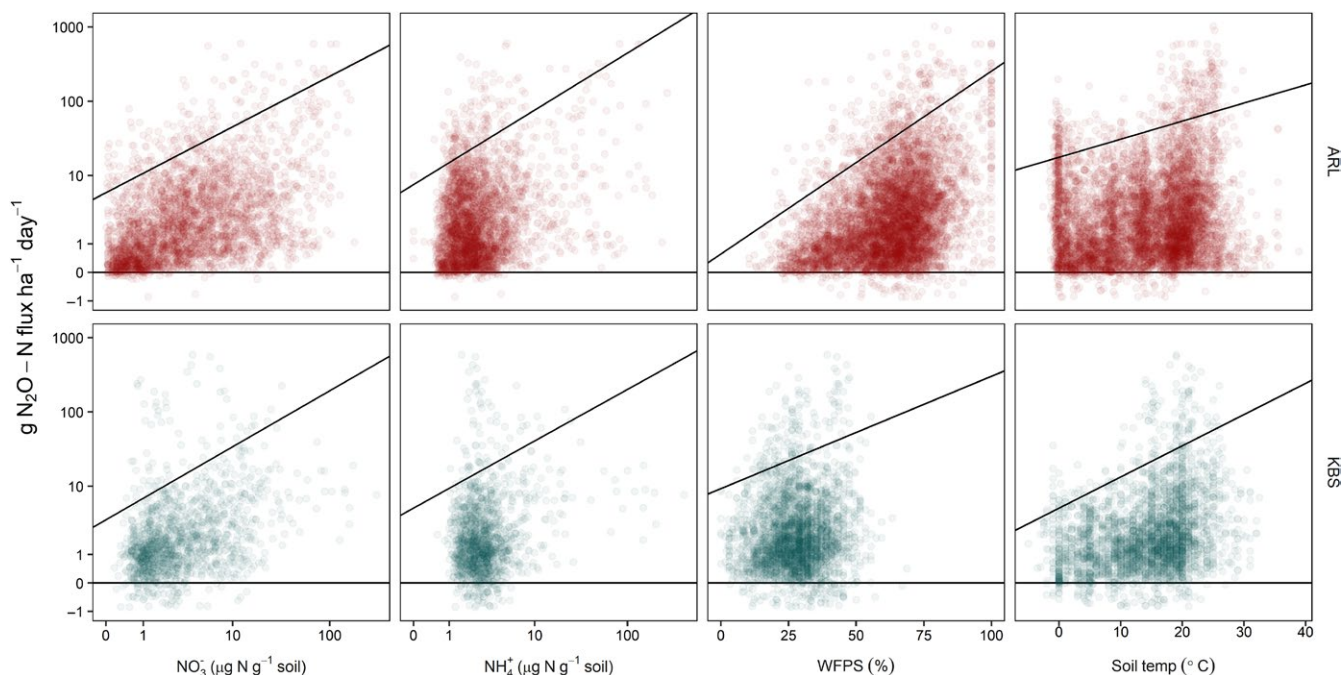


FIGURE 2 Quantile regression between soil N_2O fluxes and environmental factors. Lines indicate the quantile regression at $\tau = 0.95$, representing the 95th percentile of fluxes conditional on each environmental factor. Regressions were calculated independently for each environmental parameter. Flux data and soil inorganic N concentration data are presented on an inverse hyperbolic sine scale. All slope and intercept terms were significant at $p < 0.05$

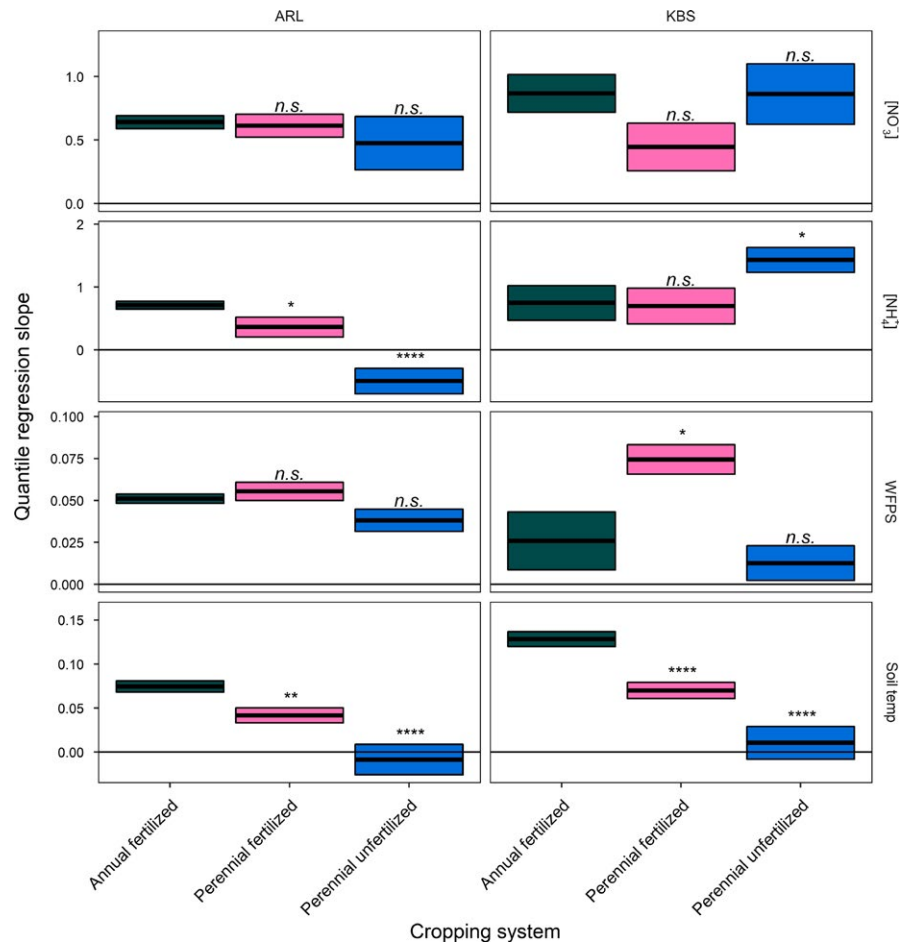


FIGURE 3 Cropping system-specific slopes for quantile regression of soil N_2O fluxes and environmental parameters. Quantile regression was conducted at $\tau = 0.95$. Crossbars indicate slope ± 1 SE (based on a kernel estimate). Slope denominators are unit changes in IHS-transformed NO_3^- or NH_4^+ concentrations, percentage points of WFPS, or $^\circ\text{C}$ of soil temperature. Symbols indicate significance of difference to annual fertilized slope ($p < 0.05^*$, $p < 0.01^{**}$, $p < 0.001^{***}$, $p < 0.0001^{****}$). Corresponding intercepts are given in Supporting Information Figure S6

KBS, with unfertilized perennials responding more strongly to NH_4^+ , fertilized perennials responding more strongly to WFPS, and both responding less strongly to soil temperature. Intercepts were all positive except for the response of unfertilized perennials at KBS to NH_4^+ (Supporting Information Figure S6).

4 | DISCUSSION

4.1 | Environmental factors correlate to upper bound of soil N_2O fluxes

Using quantile regression (Cade & Noon, 2003), we found that NO_3^- , NH_4^+ , WFPS, and soil temperature were each correlated to the upper quantiles of soil N_2O flux measurements collected from a broad range of potential bioenergy cropping systems. This supports our hypothesis that these environmental factors function as constraints on soil N_2O flux magnitudes. None of these factors drove high fluxes by itself. The largest measured fluxes occurring when WFPS,

soil temperature, and NO_3^- concentrations were all high, but minimal fluxes were observed at the highest levels of all factors. The importance of these factors fits with the consensus understanding of their importance to rates of N_2O -generating processes (Robertson & Groffman, 2015).

Quantile regression requires a large amount of data across the dynamic range (Cade & Noon, 2003). We had the greatest number of observations and relatively even distributions of observations for soil temperature and WFPS. Most models evaluating the impacts of WFPS use more complex relationships than linear regression (Heinen, 2006); although as shown in Figure 1, this seems to be a reasonable approximation at ARL. The low WFPS values observed at KBS are due to the low water holding capacity of the site's sandy soils. Castellano et al. (2010) found that matric potential was a better predictor of N_2O production in soils, explaining why a sandy soil could generate high N_2O fluxes at low WFPS. The limited effect of temperature at ARL likely results from the large number of observations taken from near-frozen soils, many of which had substantial fluxes. Near-frozen soils can

generate substantive N_2O fluxes, particularly in association with freeze–thaw events (Teepe, Brumme, & Beese, 2000; Wagner-Riddle et al., 2017). The different distribution at KBS almost certainly resulted from reduced sampling outside of the growing season, as previous work at that site found sizeable wintertime N_2O fluxes (Ruan & Robertson, 2017). Interestingly, these findings suggest the impact of soil temperature may differ during the winter, as restricting the analysis to soil temperatures $>0^\circ\text{C}$ increases the slope of the soil temperature constraint by 16% at KBS and 71% at ARL.

We observed a much stronger effect for NO_3^- than NH_4^+ at both sites, although large N_2O fluxes could occur at low levels of both compounds. Separate processes use the two compounds, and while denitrification of NO_3^- drives the largest N_2O fluxes, nitrification of $\text{NH}_3/\text{NH}_4^+$ is still relevant under certain conditions (Gelfand & Yakir, 2008; Mathieu et al., 2006). We observed a correlation between NO_3^- and NH_4^+ only when both were at high concentrations, shortly after fertilizer application, while at lower concentrations, the two were uncorrelated. Thus, in a system in which denitrification-derived N_2O production was limited by NO_3^- , nitrification-derived N_2O might not be similarly constrained (and vice versa), weakening the apparent importance of both substances at lower concentrations. While statistically significant, the effect of NH_4^+ is certainly the weakest we observed, which is consistent with its role as a N_2O -generating process of secondary importance.

4.2 | Limited evidence that system properties, rather than environmental conditions, drive differences in N_2O production

We hypothesized that cropping systems would differ in their response to environmental conditions, driving the observed differences in N_2O production. The literature provides ample reasons to expect this to be the case. Oxygen sensitivities for key denitrification enzymes differ among locations (Cavigelli & Robertson, 2001), which could shift the response to soil moisture conditions. The response to soil inorganic N concentrations also differs among systems (Lehuger et al., 2009), with some work suggesting nonagricultural systems may respond less strongly to N additions (Lu et al., 2011). Soil carbon availability and microbial biomass influence denitrification potential (Heinen, 2006), which in turn would determine how much N_2O flux would increase with the lifting of an environmental constraint. In our study, ARL had substantially higher microbial biomass and soil carbon than KBS (Liang et al., 2012), suggesting it should have higher denitrification potential and be more responsive to reductions in constraints. Despite these potential mechanisms, our dataset provides limited support for this hypothesis.

Quantile regression depends on individual measurements limited primarily by the factor being tested across its dynamic

range, requiring far more data than more conventional regression approaches. Quantile regression coefficients were highly variable across individual cropping systems, with extremely large standard errors in some cases, and some systems had highly implausible negative coefficients. Aggregating systems into annual, perennial fertilized, and perennial unfertilized groups reined in much of that variability, suggesting there were insufficient data at the level of individual cropping systems for an accurate analysis.

More problematically, unfertilized systems almost always had low concentrations of NO_3^- and NH_4^+ . In this light, the low regression slopes calculated for WFPS and soil temperature in the group of unfertilized systems seems to reflect that these factors were rarely limiting, rather than a different response. This illustrates to a key challenge of this approach in a field setting; it is highly dependent on the coincidence of permissive levels of multiple environmental factors, which may not happen very frequently. The specificity of cropping system responses to environmental factors remains a highly relevant question that is extremely important for designing management approaches aimed at minimizing N_2O fluxes.

4.3 | Constraint framework suggests steps to avoid hot moments

Conceptualizing N_2O production in terms of hot moments and environmental constraints provides a useful framework for identifying areas for reducing both the magnitude and variability of cumulative annual N_2O production. Managing NO_3^- concentrations in the soil may be particularly effective at minimizing the impact of hot moments when other factors are highly conducive to denitrification and is more easily accomplished than managing WFPS or soil temperature. Land managers are unlikely to eschew nitrogen fertilization altogether, but there may be alternative means to reduce NO_3^- concentrations. The use of enhanced efficiency fertilizers (EEFs) such as those containing nitrification inhibitors can promote a more gradual release of NO_3^- over time (e.g., Akiyama, Yan, & Yagi, 2010). Also, there is considerable evidence that increasing the diversity and perenniality of cropping systems reduces the amount of available NO_3^- and NH_4^+ in the soil (Duran, Duncan, Oates, Kucharik, & Jackson, 2016; Lu et al., 2011; Stehfest & Bouwman, 2006), likely constraining the magnitude of fluxes that could occur even with optimal temperature and moisture. Diversity and perenniality extend the range of spatial, temporal, and functional niche that can be exploited, reducing high resource concentrations (Oelmann et al., 2007; Palmberg et al., 2005). Further reductions may be possible by addressing processes that increase mineralization of organic nitrogen, such as the increase in freeze–thaw cycles caused by snow removal (Ruan & Robertson, 2017). While these recommendations are largely sound on their own merit, thinking of how they interact with other constraining

environmental factors could lead to a more nuanced set of management approaches.

ACKNOWLEDGEMENTS

We thank SK Hamilton, GR Sanford, K Kahmark, JT Simmons, SL VanderWulp, AP Dean, JR Tesmer, AP Butz, JO Sustachek, CE Menick, CC McMinn, NE Tautges, A Miller, Z Andersen, B Faust, and numerous others for providing help on this project. S Sippel and S Bohm contributed to database management and the data quality control framework. Funding was provided by the DOE Great Lakes Bioenergy Research Center (DOE BER Office of Science DE-FC02-07ER64494) and the DOE OBP Office of Energy Efficiency and Renewable Energy (DE-AC05-76RL01830), the NSF LTER Program (DEB 1027253), and Michigan AgBioResearch.

ORCID

David S. Duncan  <https://orcid.org/0000-0002-2867-0378>

Lawrence G. Oates  <http://orcid.org/0000-0003-4829-7600>

Ilya Gelfand  <http://orcid.org/0000-0002-8576-0978>

Neville Millar  <https://orcid.org/0000-0003-0297-0804>

G. Philip Robertson  <https://orcid.org/0000-0001-9771-9895>

Randall D. Jackson  <https://orcid.org/0000-0002-8033-1250>

REFERENCES

- Akiyama, H., Yan, X., & Yagi, K. (2010). Evaluation of effectiveness of enhanced-efficiency fertilizers as mitigation options for N₂O and NO emissions from agricultural soils: Meta-analysis. *Global Change Biology*, 16, 1837–1846.
- Burbidge, J. B., Magee, L., & Robb, A. L. (1988). Alternative transformations to handle extreme values of the dependent variable. *Journal of the American Statistical Association*, 83, 123–127. <https://doi.org/10.1080/01621459.1988.10478575>
- Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., & Zechmeister-Boltenstern, S. (2013). Nitrous oxide emissions from soils: How well do we understand the processes and their controls? *Philosophical Transactions of the Royal Society B: Biological Sciences*, 368, 1–13.
- Cade, B. S., & Noon, B. R. (2003). A gentle introduction to quantile regression for ecologists. *Frontiers in Ecology and the Environment*, 1, 412–420. [https://doi.org/10.1890/1540-9295\(2003\)001\[0412:AGITQR\]2.0.CO;2](https://doi.org/10.1890/1540-9295(2003)001[0412:AGITQR]2.0.CO;2)
- Castellano, M. J., Schmidt, J. P., Kaye, J. P., Walker, C., Graham, C. B., Lin, H., & Dell, C. J. (2010). Hydrological and biogeochemical controls on the timing and magnitude of nitrous oxide flux across an agricultural landscape. *Global Change Biology*, 16, 2711–2720. <https://doi.org/10.1111/j.1365-2486.2009.02116.x>
- Cavigelli, M. A., & Robertson, G. P. (2001). Role of denitrifier diversity in rates of nitrous oxide consumption in a terrestrial ecosystem. *Soil Biology and Biochemistry*, 33, 297–310. [https://doi.org/10.1016/S0038-0717\(00\)00141-3](https://doi.org/10.1016/S0038-0717(00)00141-3)
- Crutzen, P. J., Mosier, A. R., Smith, K. A., & Winiwarter, W. (2008). N₂O release from agro-biofuel production negates global warming reduction by replacing fossil fuels. *Atmospheric Chemistry and Physics*, 8, 389–395. <https://doi.org/10.5194/acp-8-389-2008>
- Dechow, R., & Freibauer, A. (2011). Assessment of German nitrous oxide emissions using empirical modelling approaches. *Nutrient Cycling in Agroecosystems*, 91, 235–254. <https://doi.org/10.1007/s10705-011-9458-9>
- Duran, B. E. L., Duncan, D. S., Oates, L. G., Kucharik, C. J., & Jackson, R. D. (2016). Nitrogen fertilization effects on productivity and nitrogen loss in three grass-based perennial bioenergy cropping systems. *PLoS One*, 11, e0151919. <https://doi.org/10.1371/journal.pone.0151919>
- Flesch, T., Baron, V., Wilson, J., Basarab, J., Desjardins, R., Worth, D., & Lemke, R. (2018). Micrometeorological measurements reveal large nitrous oxide losses during spring thaw in alberta. *Atmosphere*, 9, 128. <https://doi.org/10.3390/atmos9040128>
- Forster, P., Ramaswamy, V., Artaxo, P., Bernsten, T., Betts, R., Fahey, D., ... Van Dorland, R. (2007). Changes in atmospheric constituents and in radiative forcing. In S. Solomon, D. Qin, M. Manning, Z. Chen, M. Marquis, K. B. Averyt, M. Tignor & H. L. Miller (Eds.), *Climate change 2007: The physical science basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* (pp. 129–234). Cambridge, UK and New York, NY: Cambridge University Press.
- Gelfand, I., & Robertson, G. P. (2015). A reassessment of the contribution of soybean biological nitrogen fixation to reactive N in the environment. *Biogeochemistry*, 123, 175–184. <https://doi.org/10.1007/s10533-014-0061-4>
- Gelfand, I., Sahajpal, R., Zhang, X., Izaurralde, R. C., Gross, K. L., & Robertson, G. P. (2013). Sustainable bioenergy production from marginal lands in the US Midwest. *Nature*, 493, 514–517.
- Gelfand, I., Shcherbak, I., Millar, N., Kravchenko, A. N., & Robertson, G. P. (2016). Long-term nitrous oxide fluxes in annual and perennial agricultural and unmanaged ecosystems in the upper Midwest USA. *Global Change Biology*, 22, 3594–3607. <https://doi.org/10.1111/gcb.13426>
- Gelfand, I., & Yakir, D. (2008). Influence of nitrite accumulation in association with seasonal patterns and mineralization of soil nitrogen in a semi-arid pine forest. *Soil Biology and Biochemistry*, 40, 415–424. <https://doi.org/10.1016/j.soilbio.2007.09.005>
- Groffman, P. M., Butterbach-Bahl, K., Fulweiler, R. W., Gold, A. J., Morse, J. L., Stander, E. K., ... Vidon, P. (2009). Challenges to incorporating spatially and temporally explicit phenomena (hotspots and hot moments) in denitrification models. *Biogeochemistry*, 93, 49–77. <https://doi.org/10.1007/s10533-008-9277-5>
- Heinen, M. (2006). Simplified denitrification models: Overview and properties. *Geoderma*, 133, 444–463. <https://doi.org/10.1016/j.geoderma.2005.06.010>
- Hénault, C., Bizouard, F., Laville, P., Gabrielle, B., Nicoulaud, B., Germon, J. C., & Cellier, P. (2005). Predicting in situ soil N₂O emission using NOE algorithm and soil database. *Global Change Biology*, 11, 115–127. <https://doi.org/10.1111/j.1365-2486.2004.00879.x>
- Hiddink, J. G., & Kaiser, M. J. (2005). Implications of Liebig's law of the minimum for the use of ecological indicators based on abundance. *Ecography*, 28, 264–271. <https://doi.org/10.1111/j.0906-7590.2005.04063.x>

- Hu, H.-W., Chen, D., & He, J.-Z. (2015). Microbial regulation of terrestrial nitrous oxide formation: Understanding the biological pathways for prediction of emission rates. *FEMS Microbiology Reviews*, 39, 729–749. <https://doi.org/10.1093/femsre/fuv021>
- Koenker, R. (2018). quantreg: Quantile regression. Version 5.36.
- Landis, D. A., Gratton, C., Jackson, R. D., Gross, K. L., Duncan, D. S., Liang, C., ... Werling, B. P. (2017). Biomass and biofuel crop effects on biodiversity and ecosystem services in the North Central US. *Biomass and Bioenergy*, 114, 18–29.
- Lehuger, S., Gabrielle, B., van Oijen, M., Makowski, D., Germon, J. C., Morvan, T., & Hénault, C. (2009). Bayesian calibration of the nitrous oxide emission module of an agro-ecosystem model. *Agriculture, Ecosystems & Environment*, 133, 208–222. <https://doi.org/10.1016/j.agee.2009.04.022>
- Liang, C., Jesus, E. da. C., Duncan, D. S., Jackson, R. D., Tiedje, J. M., & Balser, T. C. (2012). Soil microbial communities under model biofuel cropping systems in southern Wisconsin, USA: Impact of crop species and soil properties. *Applied Soil Ecology*, 54, 24–31. <https://doi.org/10.1016/j.apsoil.2011.11.015>
- Livingston, G. P., Hutchinson, G. L., & Spartalian, K. (2006). Trace gas emission in chambers. *Soil Science Society of America Journal*, 70, 1459. <https://doi.org/10.2136/sssaj2005.0322>
- Lu, M., Yang, Y., Luo, Y., Fang, C., Zhou, X., Chen, J., ... Li, B. (2011). Responses of ecosystem nitrogen cycle to nitrogen addition: A meta-analysis. *The New Phytologist*, 189, 1040–1050. <https://doi.org/10.1111/j.1469-8137.2010.03563.x>
- Mathieu, O., Hénault, C., Lévêque, J., Baujard, E., Milloux, M.-J., & Andreux, F. (2006). Quantifying the contribution of nitrification and denitrification to the nitrous oxide flux using ^{15}N tracers. *Environmental Pollution*, 144, 933–940. <https://doi.org/10.1016/j.envpol.2006.02.005>
- Molodovskaya, M., Singurindy, O., Richards, B. K., Warland, J., Johnson, M. S., & Steenhuis, T. S. (2012). Temporal variability of nitrous oxide from fertilized croplands: Hot moment analysis. *Soil Science Society of America Journal*, 76, 1728–1740. <https://doi.org/10.2136/sssaj2012.0039>
- MSCO (2013). Michigan State Climatologist's Office: 27 year summary of annual values for Gull Lake (3504) 1981–2010.
- Niklaus, P. A., Wardle, D. A., & Tate, K. R. (2006). Effects of plant species diversity and composition on nitrogen cycling and the trace gas balance of soils. *Plant and Soil*, 282, 83–98. <https://doi.org/10.1007/s11104-005-5230-8>
- NWS (2013). National Weather Service: Wisconsin 30 year average temperature and precipitation 1981–2010.
- Oates, L. G., Duncan, D. S., Gelfand, I., Millar, N., Robertson, G. P., & Jackson, R. D. (2016). Nitrous oxide emissions during establishment of eight alternative cellulosic bioenergy cropping systems in the North Central United States. *GCB Bioenergy*, 8, 539–549. <https://doi.org/10.1111/gcbb.12268>
- Oelmann, Y., Wilcke, W., Temperton, V. M., Buchmann, N., Roscher, C., Schumacher, J., ... Weisser, W. W. (2007). Soil and plant nitrogen pools as related to plant diversity in an experimental grassland. *Soil Science Society of America Journal*, 71, 720–729. <https://doi.org/10.2136/sssaj2006.0205>
- Palmborg, C., Scherer-Lorenzen, M., Jumpponen, A., Carlsson, G., Huss-Danell, K., & Höglberg, P. (2005). Inorganic soil nitrogen under grassland plant communities of different species composition and diversity. *Oikos*, 110, 271–282. <https://doi.org/10.1111/j.0030-1299.2005.13673.x>
- Pedersen, A. R. (2015). HMR: Flux estimation with static chamber data. Version 0.4.1.
- Portmann, R. W., Daniel, J. S., & Ravishankara, A. R. (2012). Stratospheric ozone depletion due to nitrous oxide: Influences of other gases. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 367, 1256–1264. <https://doi.org/10.1098/rstb.2011.0377>
- R Core Team (2018). R: A language and environment for statistical computing. Version 3.5.0.
- Robertson, G. P., & Groffman, P. M. (2015). Nitrogen transformations. In E. A. Paul (Ed.), *Soil microbiology, ecology and biochemistry* (4th ed., pp. 421–446). Burlington, MA: Academic Press. <https://doi.org/10.1016/B978-0-12-415955-6.00014-1>
- Robertson, G. P., Paul, E. A., & Harwood, R. R. (2000). Greenhouse gases in intensive agriculture: Contributions of individual gases to the radiative forcing of the atmosphere. *Science*, 289, 1922–1925. <https://doi.org/10.1126/science.289.5486.1922>
- Robertson, G. P., Sollins, P., Ellis, B. G., & Lajtha, K. (1999). Exchangeable ions, pH, and cation exchange capacity. In G. P. Robertson, D. C. Coleman, C. S. Bledsoe, & P. Sollins (Eds.), *Standard soil methods for long term ecological research* (pp. 106–114). Oxford, UK: Oxford University Press.
- Robertson, G. P., & Tiedje, J. M. (1987). Nitrous oxide sources in aerobic soils: Nitrification, denitrification and other biological processes. *Soil Biology and Biochemistry*, 19, 187–193. [https://doi.org/10.1016/0038-0717\(87\)90080-0](https://doi.org/10.1016/0038-0717(87)90080-0)
- Ruan, L., & Robertson, G. P. (2017). Reduced snow cover increases wintertime nitrous oxide (N_2O) emissions from an agricultural soil in the upper U.S. Midwest. *Ecosystems*, 20, 917–927. <https://doi.org/10.1007/s10021-016-0077-9>
- Saha, D., Rau, B. M., Kaye, J. P., Montes, F., Adler, P. R., & Kemanian, A. R. (2017). Landscape control of nitrous oxide emissions during the transition from conservation reserve program to perennial grasses for bioenergy. *GCB Bioenergy*, 9, 783–795. <https://doi.org/10.1111/gcbb.12395>
- Sanford, G. R., Oates, L. G., Jasrotia, P., Thelen, K. D., Robertson, G. P., & Jackson, R. D. (2016). Comparative productivity of alternative cellulosic bioenergy cropping systems in the North Central USA. *Agriculture, Ecosystems & Environment*, 216, 344–355. <https://doi.org/10.1016/j.agee.2015.10.018>
- Sekhon, R. S., Briskine, R., Hirsch, C. N., Myers, C. L., Springer, N. M., Buell, C. R., ... Kaeppler, S., (2013). Maize gene atlas developed by RNA sequencing and comparative evaluation of transcriptomes based on RNA sequencing and microarrays. *PLoS ONE*, 8, e61005. <https://doi.org/10.1371/journal.pone.0061005>
- Stehfest, E., & Bouwman, L. (2006). N_2O and NO emission from agricultural fields and soils under natural vegetation: Summarizing available measurement data and modeling of global annual emissions. *Nutrient Cycling in Agroecosystems*, 74, 207–228. <https://doi.org/10.1007/s10705-006-9000-7>
- Surendran Nair, S., Kang, S., Zhang, X., Miguez, F. E., Izaurralde, R. C., Post, W. M., ... Wullschlegel, S. D. (2012). Bioenergy crop models: Descriptions, data requirements, and future challenges. *GCB Bioenergy*, 4, 620–633. <https://doi.org/10.1111/j.1757-1707.2012.01166.x>
- Teepe, R., Brumme, R., & Beese, F. (2000). Nitrous oxide emissions from frozen soils under agricultural, fallow and forest land. *Soil Biology and Biochemistry*, 32, 1807–1810. [https://doi.org/10.1016/S0038-0717\(00\)00078-X](https://doi.org/10.1016/S0038-0717(00)00078-X)

- U.S. Environmental Protection Agency (2018). Inventory of US greenhouse gas emissions and sinks: 1990–2016. Washington, DC.
- Wagner-Riddle, C., Congreves, K. A., Abalos, D., Berg, A. A., Brown, S. E., Ambadan, J. T., ... Tenuta, M. (2017). Globally important nitrous oxide emissions from croplands induced by freeze-thaw cycles. *Nature Geoscience*, 10, 279–283. <https://doi.org/10.1038/ngeo2907>
- Wallenstein, M. D., Myrold, D. D., Firestone, M. K., & Voytek, M. (2006). Environmental controls on denitrifying communities and denitrification rates: Insights from molecular methods. *Ecological Applications*, 16, 2143–2152. [https://doi.org/10.1890/1051-0761\(2006\)016\[2143:ECODCA\]2.0.CO;2](https://doi.org/10.1890/1051-0761(2006)016[2143:ECODCA]2.0.CO;2)
- Wang, Y., Wang, Y., & Ling, H. (2010). A new carrier gas type for accurate measurement of N₂O by GC-ECD. *Advances in Atmospheric Sciences*, 27, 1322–1330. <https://doi.org/10.1007/s00376-010-9212-2>
- Werling, B. P., Dickson, T. L., Isaacs, R., Gaines, H., Gratton, C., Gross, K. L., ... Landis, D. A. (2014). Perennial grasslands enhance biodiversity and multiple ecosystem services in bioenergy landscapes. *Proceedings of the National Academy of Sciences*, 111, 1652–1657. <https://doi.org/10.1073/pnas.1309492111>
- Wickham, H. (2009). *ggplot2: Elegant graphics for data analysis*. New York, NY: Springer-Verlag. <https://doi.org/10.1007/978-0-387-98141-3>
- Zenone, T., Zona, D., Gelfand, I., Gielen, B., Camino-Serrano, M., & Ceulemans, R. (2016). CO₂ uptake is offset by CH₄ and N₂O emissions in a poplar short-rotation coppice. *GCB Bioenergy*, 8, 524–538. <https://doi.org/10.1111/gcbb.12269>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

How to cite this article: Duncan DS, Oates LG, Gelfand I, Millar N, Robertson GP, Jackson RD. Environmental factors function as constraints on soil nitrous oxide fluxes in bioenergy feedstock cropping systems. *GCB Bioenergy*. 2019;11:416–426. <https://doi.org/10.1111/gcbb.12572>