



Discontinuity Analysis Reveals Alternative Community Regimes During Phytoplankton Succession

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It is well-recognized in plankton ecology that phytoplankton development can lead to distinct peaks (i.e., blooms) during spring and summer. We used a 5-year (2007–2011) phytoplankton data set and utilized discontinuity analysis to assess resilience attributes of spring and summer blooms based on the cross-scale resilience model. Using the size structure (i.e., cross-scale structure as an indicator of resilience) in the sampled plankton data, we assessed whether spring and summer blooms differ substantially between but not within blooms; that is, whether they comprise alternative community regimes. Our exploratory study supported this expectation and more broadly resilience theory, which posits that ecological systems can manifest in and change between alternative regimes. The dynamics of regimes receives increased attention because rapid environmental change potentially irreversibly alters ecosystems. Model organisms are needed that allow revealing patterns and processes of various aspects of regime dynamics at tractable time scales. Our preliminary findings suggest that phytoplankton can be suitable models for assessing the intricacies of regimes and regime changes.

Keywords: Baltic Sea, resilience, cross-scale structure, phytoplankton, blooms, alternative regimes, discontinuities, community succession

INTRODUCTION

Patterns of discrete size structures of organisms in communities are related to a number of abiotic and biotic factors that operate across distinct scales of space and time; that is, they reflect the hierarchical organization of ecosystems (Nash et al., 2014; Sundstrom et al., 2018). This is due to competitive interactions and behavioral, life-history, and morphological adaptations to resources (e.g., food and shelter) that prevail at each scale (Holling, 1992; Segura et al., 2013). For instance, an elephant and an ant in a savannah interact with and exploit very different scales in the system. The discrete size structure within communities, also referred to as cross-scale structure (Sundstrom et al., 2018), has been suggested to serve as a surrogate of resilience in ecosystems (Angeler et al., 2016), and other complex systems (Sundstrom et al., 2014). This is due to the ability of one scale (discrete size structure) in the system being able to buffer against disturbances at other scales as a function of their functional trait characteristics and diversity (Angeler and Allen, 2016).

Resilience theory posits that ecological systems undergo substantial reorganization when they shift between alternative regimes (Angeler and Allen, 2016). Spanbauer et al. (2016) showed that the discrete size structure of benthic diatoms in Foy lake (USA) substantially

changed as a result of a regime shift. This is consistent with predictions that cross-scale structure as an indicator of resilience can identify regimes and regime changes (Baho et al., 2017). Phytoplankton communities are both discontinuously structured (Segura et al., 2013; Downing et al., 2014) and they can undergo substantial seasonal reorganization (e.g., Sommer et al., 2012; Behrenfeld and Boss, 2014). For instance, in the Baltic Sea phytoplankton communities show multiple seasonal peaks (“blooms”). These blooms have been shown to undergo repeated cycles of reorganization and collapse, which from a resilience perspective suggest community-level regime shifts (Angeler et al., 2015). However, it is still unclear if these dynamics reflect the manifestation of and change between alternative regimes posited by resilience theory. Therefore, research is needed to address this question, especially to shed light on potential similarities and differences between resilience-based approaches and taxonomic studies that have dominated research agendas in ecology in the past.

Phytoplankton is potentially a useful model for studying alternative community regimes because they have fast turnover, communities have high species diversity, and they show complex system dynamics (Leibold and Norberg, 2004). This behavior is influenced by feedbacks that emanate from interacting biotic (trophic cascades and competition) and abiotic (stratification, nutrients, temperature) variables (Sommer et al., 1986; Leibold and Norberg, 2004). These dynamics are well-documented in ecology, particularly ecological succession, and plankton ecology (Clements, 1916; Sommer et al., 2012). These paradigms are therefore valuable for assessing alternative community regimes, based on the cross-scale structure present in the study system. Phytoplankton communities develop into spring and summer blooms and in relation to a spring clear-water phase (Sommer et al., 1986; Hajdu, 2002), although these patterns can vary among ecosystems (Jaanus et al., 2011). In our study system, the Baltic Sea, spring blooms develop from March to May and summer blooms occur between June and August (Angeler et al., 2015). These blooming periods are stable and recur annually, although there is annual variation in bloom timing. This timing provides an opportunity to assess whether spring and summer blooms comprise alternative phytoplankton community regimes.

In the Baltic Sea, spring blooms are the result of several factors. Among those are seasonal increases in temperature and solar radiation. Temperature and saline gradients that lead to stratification after winter mixing can also be important. At the beginning, spring blooms, which are characterized by high biomass, are dominated by diatoms that grow fast and later replaced by dinoflagellates with slower growth. With increasing temperature increase and nutrient depletion, spring blooms collapse, and reorganize in species-rich summer blooms with many inedible flagellates and cyanobacteria (Angeler et al., 2015). This reorganization is partly the result of changing food webs that alter biotic feedbacks; specifically, top-down effects, including zooplankton grazing that shapes the dynamics of phytoplankton (Sommer et al., 1986, 2012). In addition, Baltic Sea phytoplankton shows variability in taxonomic structure and the phenology of blooms (Jaanus et al., 2011; Klais et al., 2011; Suikkanen et al., 2013), which results from the interaction of anthropogenic (eutrophication, overfishing), climatic, and biotic

factors (Elmgren, 1989; Wasmund and Uhlig, 2003; Österblom et al., 2007). Complex interactions between winter temperature and nutrient dynamics further affect community responses during spring and summer (Janssen et al., 2004).

The seasonally recurring spring and summer blooms that reflect phytoplankton dynamics allow for assessing assumptions regarding community dynamics during different successional stages. In this exploratory study, we assess cross-scale structure in spring and summer phytoplankton blooms to test the following expectations:

Phytoplankton blooms across seasons represent alternative regimes, reflected in different cross-scale structure, because of differing abiotic and biotic conditions between both bloom seasons.

During the duration of individual blooms, cross-scale structure between sampling events should not be significantly different as a result of the intrinsic regime properties that organize community dynamics within each bloom.

MATERIALS AND METHODS

Ethics Statement

All field and laboratory work is approved by the Swedish Agency for Marine and Water Management (HaV) and are part of the Baltic Sea Monitoring Program. Data used in this study are available through the Swedish Meteorological and Hydrological Institute (SMHI) and the Dryad Digital Repository (<http://dx.doi.org/10.5061/dryad.8hj8t>). No endangered or protected species were involved in this study.

Sites and Sampling

Phytoplankton communities were assessed at the 40 m deep B1 station near the coast of Askö (58°48' N, 17°38' E) and at the 459 m deep BY31 offshore station at Landsort Deep (58°35.90' N, 18°14.21' E). These sites are located in the southern area of the Baltic Sea, specifically in the NW Baltic Proper. Data were collected from both stations in weekly to fortnightly intervals in spring (March–May) and summer (June–August) between 2007 and 2011. Sampling and analysis adhered to standardized protocols. Phytoplankton samples were taken as integrated samples with a sampling hose (inner diameter 19 mm) from 0 to 20 m and preserved with acid Lugol's solution (Willén, 1962). In this way we could integrate abiotic and biotic heterogeneity in the water column and more accurately capture bloom aspects over the spring and summer. Taxonomic experts carried out the evaluation of phytoplankton adhering to standard protocols. Briefly, an inverted microscope with phase contrast was used to count phytoplankton (>2 µm) after sedimentation in 10- or 25-mL chambers. Cells were measured and their sizes classified following HELCOM (2008). Carbon content was calculated from the biovolume of all individuals of a species, including colonial taxa. These evaluations were carried out following Olenina et al. (2006) and standardized volumes (http://www.ices.dk/marine-data/vocabularies/Documents/PEG_BVOL.zip).

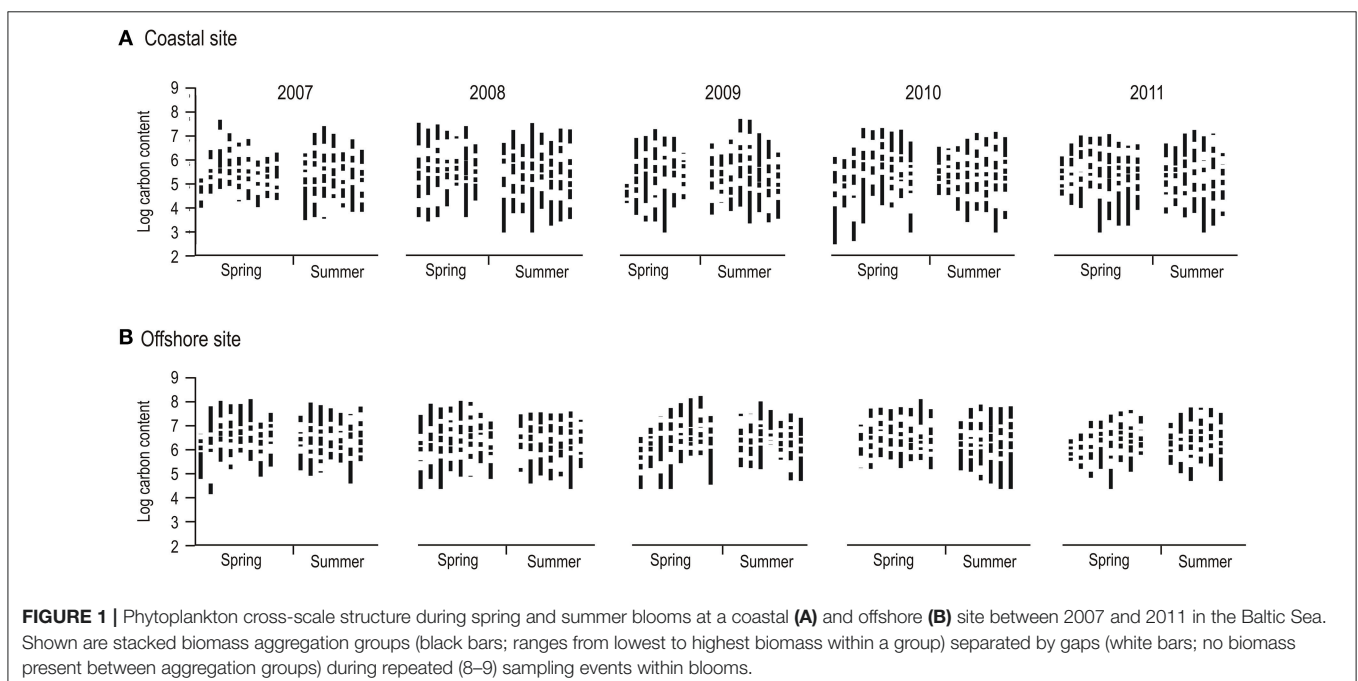
Statistical Analyses

We used discontinuity analysis for assessing the cross-scale structure present in the phytoplankton communities (Barichievy

et al., 2018). The evaluation of cross-scale structure has been originally based on animal body sizes (Holling, 1992). More recently it has been extended to a broader discontinuity framework that accounts for abundances in ecological studies (Angeler et al., 2014; Sundstrom et al., 2018) and that accommodates metrics from non-ecological systems (e.g., city size: Garmestani et al., 2008). There is also evidence that plankton studies based on body mass of single species and biovolume of populations give similar results (Baho et al., 2015). In this study we used volumetric data and assessed cross-scale structure in the carbon content of phytoplankton using Bayesian Classification and Regression Trees (BCART) (Chipman et al., 1998). Phytoplankton carbon content was rank ordered based on ascending log-transformed measures of individual populations in the communities. Rank-ordered matrices were created for each phytoplankton community at each sampling date during the study period. BCART was conducted individually on these matrices to identify biomass groups in the phytoplankton community data for each sampling date by assessing within-group homogeneity (Stow et al., 2007). The analysis was based on a million iterations repeated 25 times. From this universe of trees the best 20 were displayed and the tree with the best (highest) log-likelihood ratio was selected for further analysis. The trees branch into distal nodes that comprise groups with highest homogeneity. In our study, these homogeneity groups are composed of phytoplankton populations that differ between groups in terms of their homogeneities in carbon content, and they likely emerge from the patterns-process relationships present within each bloom regime (Holling, 1992; Allen et al., 2005; Angeler et al., 2012). That is, the homogeneity groups identified by the BCART trees were used for classifying the phytoplankton populations into aggregation groups, thereby determining the cross-scale structure in the community. These

homogeneity or aggregation groups were used to calculate nine diagnostics of phytoplankton cross-scale structure in carbon content for further analysis. These diagnostics are rooted in and therefore represent individually and collectively the cross-scale resilience model (Peterson et al., 1998), a commonly applied tool for quantifying resilience (Angeler and Allen, 2016). These diagnostics are: (1) total number of aggregation groups for the phytoplankton community at each sampling date, (2) the average of their aggregation group lengths (each aggregation length was measured as the difference between the lowest and highest log-transformed carbon content of a specific aggregation group), (3) averages of gap lengths (each gap length was measured as the difference between the log-biomass value of edges between adjacent aggregation groups), (4) standard deviation of aggregation lengths within a community as a variability measure of aggregation lengths in the community, (5) standard deviations of gap lengths, (6) average number of phytoplankton species composing carbon content aggregation groups of each analysis, (7) standard deviation of species composing aggregation groups, (8) lowest carbon content, and (9) highest carbon content values for each community were used for bounding the other diagnostics according to carbon content ranges of the phytoplankton communities.

Permutational multivariate ANOVA (PERMANOVA) (Anderson, 2005) was used to contrast the diagnostics of phytoplankton biomass cross-scale structure following the design of Angeler et al. (2015). The PERMANOVA model had three main terms. Factor 1 (Blooms) was fixed and contrasted blooms between spring and summer based on average cross-scale structure of phytoplankton. Factor 2 (Years) was random and categorical and comprised the study years from 2007 to 2011. Factor three (Months nested in years) was also random and comprised within-bloom dynamics during spring (March,



April, May) and summer (June, July, August). Interaction terms between these factors were also tested. Three terms allowed assessing whether phytoplankton communities organize in distinct spring and summer community regimes in the Baltic Sea under our study period: (1) The term “Blooms” tests if phytoplankton cross-scale structure differs between spring and summer. This term is expected to be significant if spring and summer blooms comprise alternative community regimes. (2) The interaction term “Blooms \times Months (Years)” assesses dynamic change of cross-scale structure within blooms. Because

individual blooms are considered alternative community regimes, cross-scale structure between sampling events should not be significantly different due to regime-inherent properties that steer community dynamics within each bloom (Angeler et al., 2015). (3) The interaction term “Blooms \times Years” tests if diagnostics of cross-scale structure are stable during spring and summer over the study years. If phytoplankton organizes in different spring and summer alternative community regimes consistently over time this term is not expected to be significant. PERMANOVA was calculated on Euclidean distance matrix of

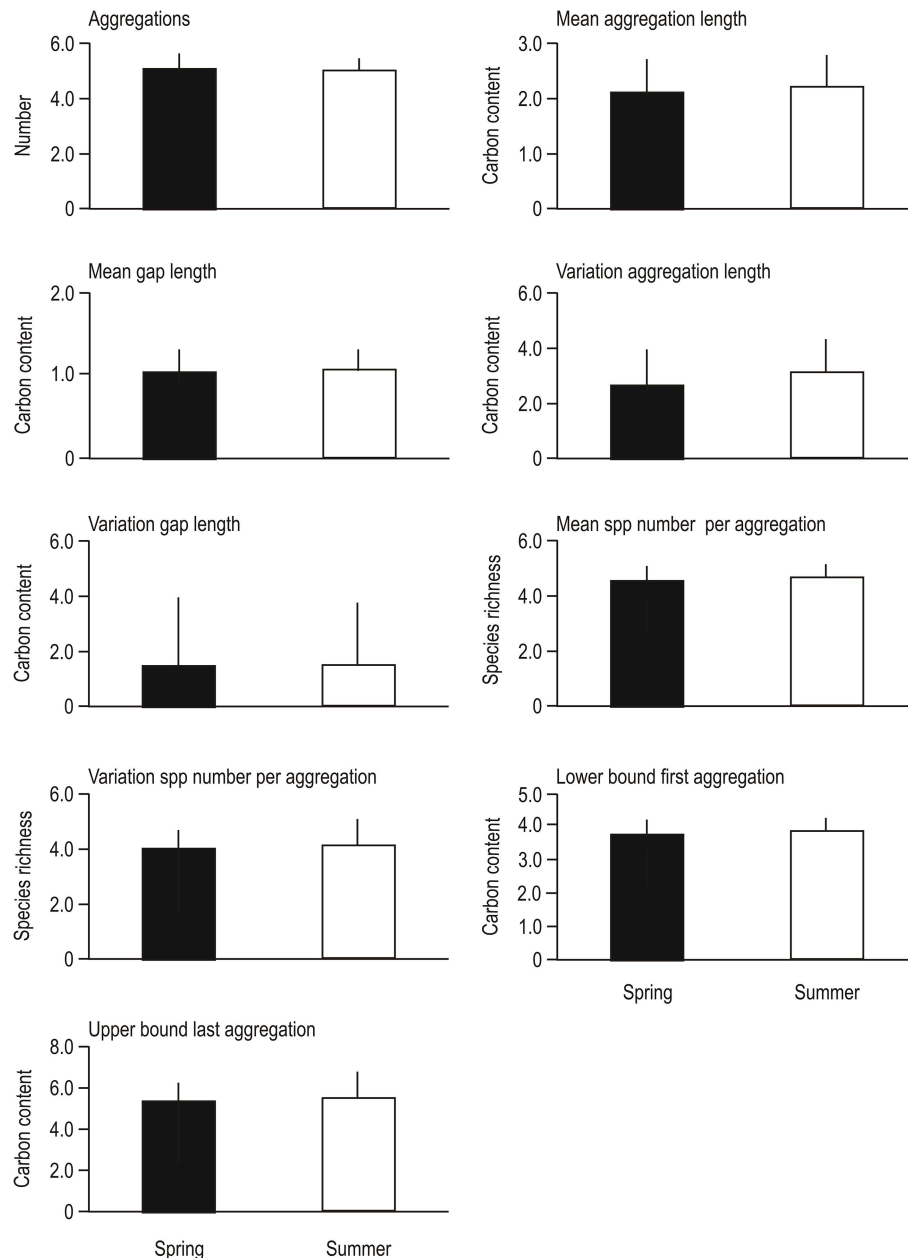


FIGURE 2 | Comparison of resilience metrics based on the cross-scale resilience model used for analysis. Shown are the means and standard deviations from 3 yearly measurements for each site (coastal and offshore) for the study period 2007–2011 ($n = 36$). See section Materials and Methods for descriptions and calculations of these metrics.

standardized diagnostics of phytoplankton biomass cross-scale structure using the coastal and offshore sites as replicates. Nine thousand nine hundred and ninety-nine unrestricted permutations of raw data were used and calculations were carried out with PERMANOVA v1.6 (Anderson, 2005). Significance testing was based on Monte Carlo asymptotic *P*-values.

RESULTS

Phytoplankton community cross-scale structure showed substantial variation between and within blooms between 2007 and 2011 at the coastal and offshore site in the Baltic Sea (Figure 1). Examining visually individual resilience metrics used for analysis revealed subtle differences between spring and summer blooms, with the exception of the variation (standard deviation) in the length of aggregations (i.e., the difference between the lowest and highest log-transformed biomass of a specific aggregation group), which was higher in summer compared to spring blooms (Figure 2). Despite the apparent similarities observed in individual metric comparisons and the variability in the data set, PERMANOVA detected a significant “Blooms” effect (Table 1). This suggests that phytoplankton cross-scale structure may reorganize in alternative community regimes between spring and summer. The effect of “Years” (study period between 2007 and 2011) was not significant, highlighting that the phytoplankton cross-scale structure of blooms is not changing during the study period. The insignificant interaction term “Blooms × Years” highlights that phytoplankton cross-scale structure present during spring and summer blooms remains differentiated over the study period. Finally, the term “Blooms × Months (Years)” was not significant. This suggests similar cross-scale structure between sampling events within each regime.

DISCUSSION

The results of our exploratory study show that phytoplankton seasonal development reflects the organization into alternative regimes, consistent with ecological resilience theory. This finding aligns with a previous study that assessed patterns of bloom

collapse and reorganization following the adaptive cycle (Angeler et al., 2015), a heuristic of complex system change (Holling, 1986; Gunderson and Holling, 2002). The results are also in agreement with a plethora of taxonomic studies in marine (Edwards and Richardson, 2004; Lindemann and St John, 2014; Vidal et al., 2017) and freshwater (Sommer, 1985; Munawar and Munawar, 1986; Reynolds, 2006) environments that have documented substantial community changes in terms of phytoplankton species composition and biomass as a result of abiotic variability.

Although our results need to be interpreted with caution due to low sample size and heterogeneity of sites, the significant “blooms” term in the PERMANOVA model preliminarily supports the interpretation that spring and summer blooms generally comprise alternative phytoplankton community regimes. Alternative regimes are usually associated with ecosystem dynamics (e.g., Beisner et al., 2003); however, alternative regimes at the community level, which are often transient, have also been documented (Fukami and Nakajima, 2011; Jiang et al., 2011). Such changes can occur when variability at the ecosystem level creates abiotic and biotic conditions that entail a substantial restructuring at lower hierarchical levels in the system; i.e., at the scale of ecological communities (Allen et al., 2014). Such reorganization was the case in our study. Although the Baltic Sea is considered to operate in a stable eutrophic regime (Yletyinen et al., 2017), the substantial seasonal changes in the abiotic and biotic environment can lead to a transitioning of phytoplankton communities between alternative regimes. In the Baltic Sea, abiotic changes are related to temperature, nutrients, stratification, and salinity (Angeler et al., 2015). Biotic changes are manifested in alterations of food webs; i.e., trophic cascades including zooplankton grazing affect the dynamics of phytoplankton assemblages (Winder and Cloern, 2010; Sommer et al., 2012; Behrenfeld and Boss, 2014).

Most phytoplankton successional studies are based on taxonomic analyses. These studies show high community turnover between spring and summer blooms and also a high replacement of species and major taxonomic groups from the onset to the end of blooms (Reynolds, 2006). For instance, in the Baltic Sea dinoflagellates replace initially dominant diatoms toward the end of spring blooms. Similarly, summer blooms are characterized by a dynamic replacement between cyanobacteria, cryptophytes, and dinoflagellates (Angeler et al., 2015). These changes are due to functional attributes of phytoplankton species with some species (diatoms) being *r*-strategists; that is, they grow fast when nutrient availability is high (Sommer, 1981). On the other hand there are some species that are *K*-strategists, which characterize slow growing species (dinoflagellates) that are competitively weaker compared to diatoms, and that become abundant during periods of low nutrient availability (Sommer, 1981; Lembi and Waaland, 2007).

Using cross-scale structure in the carbon content of phytoplankton populations in the discontinuity analysis rather than taxonomic information, our results show an important difference between both approaches. A previous taxonomy based study using the same data set, which thus allows for direct comparisons, found significant within-bloom variability in phytoplankton community composition (Angeler et al., 2015).

TABLE 1 | Results of PERMANOVA analysis contrasting multivariate cross-scale structure across blooms (averaged spring and summer blooms of phytoplankton), years (2007–2011), months nested in years (3 months comprising each bloom), and their interactions.

Factor	df	SS	MS	F	<i>P</i>
Blooms	1	0.06	0.06	7.40	0.004
Years	4	0.16	0.04	1.42	0.193
Months (Years)	10	0.28	0.03	1.69	0.023
Blooms × Years	4	0.03	0.01	0.44	0.951
Blooms × Months (Years)	10	0.19	0.02	1.19	0.258
Residual	30	0.49	0.02		
Total	59	1.21			

Shown are degrees of freedom (df), sums of squares (SS), mean squares (MS), *F*-ratios (*F*), and the Monte Carlo asymptotic *P*-values (*P*). Significant *P*-values are highlighted in bold.

However, such an effect was not detected using the cross-scale structure of carbon content in this study. Both findings are not mutually exclusive. Taxonomic studies capture the pronounced abiotic and biotic variability within blooms while cross-scale structure as a measure of system resilience provides a more conservative, systemic measure of relative stability, and persistence of a regime. In this context stability and persistence are defined as the regime dynamics that are bound within a basin of attraction (Angeler and Allen, 2016). Specifically, alternative regimes are characterized by patterns-process relationships and feedbacks that are relatively stable (Beisner et al., 2003), although variability occurs within a regime when they adapt and cope with disturbances (Gunderson, 2000; Angeler et al., 2019). Such variability is clearly evident in the dynamics of cross-scale patterns within spring and summer regimes (Figure 1), which may reflect adaptive community dynamics within the basins of attraction of spring and summer blooms.

We conclude with acknowledging that we could only use two sites for this study, which prevents us from drawing firm conclusions about phytoplankton dynamics in the Baltic Sea. However, the exploratory results allow us to highlight the potential to study plankton seasonality from a resilience perspective. Accounting for the complexity inherent in resilience might potentially contribute to a better understanding of ecosystem dynamics and potentially management (Angeler et al., 2014). Specifically, our preliminary findings broadly supported resilience theory, which posits that ecological systems can manifest and change between alternative regimes. The dynamics of regimes receive increased attention by scientists and managers because rapid environmental change potentially irreversibly alters ecosystems. Model organisms are needed that allow revealing patterns and processes of various aspects of regime dynamics at tractable time scales. Our findings suggest that phytoplankton can be suitable models for assessing the intricacies of regimes

and regime changes repeatedly over relatively short time spans.

AUTHOR CONTRIBUTIONS

DA conceived and designed study, carried out analysis, and wrote the paper. CA, DT, and MW contributed to idea development and the writing. All authors provided funding and approved submission.

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