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Role of carbonate burial in Blue Carbon budgets

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Calcium carbonates (CaCO_3) often accumulate in mangrove and seagrass sediments. As CaCO_3 production emits CO_2 , there is concern that this may partially offset the role of Blue Carbon ecosystems as CO_2 sinks through the burial of organic carbon (C_{org}). A global collection of data on inorganic carbon burial rates (C_{inorg} , 12% of CaCO_3 mass) revealed global rates of $0.8 \text{ TgC}_{\text{inorg}} \text{ yr}^{-1}$ and $15\text{--}62 \text{ TgC}_{\text{inorg}} \text{ yr}^{-1}$ in mangrove and seagrass ecosystems, respectively. In seagrass, CaCO_3 burial may correspond to an offset of 30% of the net CO_2 sequestration. However, a mass balance assessment highlights that the C_{inorg} burial is mainly supported by inputs from adjacent ecosystems rather than by local calcification, and that Blue Carbon ecosystems are sites of net CaCO_3 dissolution. Hence, CaCO_3 burial in Blue Carbon ecosystems contribute to seabed elevation and therefore buffers sea-level rise, without undermining their role as CO_2 sinks.

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Mangrove forests and seagrass meadows have the capacity to elevate the seabed through the accretion of inorganic and organic particles¹ at global rates of ~ 0.5 and ~ 0.2 cm yr^{-1} , respectively¹. Sediment accretion in mangrove forests and seagrass meadows leads to the sequestration of organic carbon (C_{org})^{2,3} originating from within and outside of the vegetated ecosystem⁴. Although mangroves and seagrass ecosystems occupy only a small fraction of the total coastal area ($< 2\%$), they contribute 10% and 25% to the yearly C_{org} sequestration in the coastal zone^{1,5}, respectively. Recognition of mangrove and seagrass meadows, together with saltmarshes, as sites of intense C_{org} burial led to the formulation of Blue Carbon strategies to mitigate and adapt to climate change, through conservation and restoration of these ecosystems^{1,6–8}. The focus on Blue Carbon has provided substantial impetus to assess sediment C_{org} concentrations and burial rates in vegetated coastal ecosystems, which recently have been widely reviewed⁹.

C_{org} generally represents a minor fraction (2–3%) of buried material within mangrove and seagrass sediments^{10,11} (although this is highly variable¹²), the rest being siliciclastic and carbonate particles. A global assessment of the concentration of inorganic carbon concluded that C_{inorg} can exceed C_{org} concentration in seagrass sediments¹³. Seagrass and mangrove plants do not calcify per se; however, they provide habitats for an abundant associated calcifying fauna and flora (e.g., crabs, sea stars, snails, bivalves, calcified algae, foraminifera), whose shells and skeletons may be deposited and buried in the sediment along with the plant litter, and the organic and inorganic particles imported from adjacent ecosystems.

Counterintuitively, CaCO_3 production represents a source of CO_2 to the atmosphere, as calcification produces CO_2 with a ratio of ~ 0.6 mol of CO_2 emitted per mol of CaCO_3 precipitated¹⁴. This has led to the argument that high CaCO_3 burial may partially offset CO_2 sequestration associated with C_{org} burial in some seagrass meadows and mangrove forests¹⁵. However, there are several caveats that affect these arguments and render inferences on the role of Blue Carbon ecosystems as net CO_2 sinks or sources inconclusive^{13,16}, based on the comparison of C_{org} and C_{inorg} sediment burial rates. To date, very few articles report the burial rates of CaCO_3 in mangrove and seagrass ecosystems^{15–17}, and the role of CaCO_3 burial in sediments and CO_2 emissions depends on the balance between dissolution and production. If CaCO_3 dissolution equals local calcification, then the burial of CaCO_3 is supported exclusively by allochthonous inputs and is neutral in terms of CO_2 emissions or sequestration. If dissolution exceeds local calcification, then CaCO_3 dynamics add to the CO_2 sink capacity of Blue Carbon ecosystems, even if CaCO_3 , which must be subsidized from allochthonous sources, is buried in the sediments. Only if CaCO_3 dissolution is lower than local calcification does CaCO_3 burial result in CO_2 emissions.

Here we address the current gap in global estimates of C_{inorg} burial in seagrass and mangrove ecosystems by providing first estimates of contemporary (last century) C_{inorg} burial rates. We rely on a compilation and analysis of data on sediment chronologies (i.e., including radiometric dating of sediment cores with ^{210}Pb) and C_{inorg} concentrations from around the world (Fig. 1). We compare burial, calcification and dissolution rates in three locations where most of the carbon mass balance terms were available. We then address the role of CaCO_3 burial in CO_2 emissions by resolving the source of the CaCO_3 buried in seagrass meadows as either allochthonous or autochthonous (i.e., from associated flora and fauna). We conclude that the high amounts of CaCO_3 found in Blue Carbon sediments can not be converted into CO_2 emissions.

Results

Global disparities in Blue Carbon sediments. CaCO_3 supports an important part of sediment accretion rates (SARs) in seagrass ecosystems, although with large geographical disparities and a non-normal distribution (Shapiro–Wilks test, $p < 0.001$). Indeed, in 40% of global locations, the CaCO_3 concentration was under 10% dry weight (DW), whereas in 28% of locations the CaCO_3 content exceeded 80% DW (see Supplementary Figure 1a). Overall, the median (interquartile range: IQR) global concentration of CaCO_3 in seagrass meadow sediments was 61 (56) %DW (mean \pm SE of 54 ± 7).

In mangrove forests, we observe a large difference between the mean (\pm SE) and the median (IQR) CaCO_3 concentration: $21 \pm 11\%$ and 3 (31)%, respectively. This is explained by the strong non-normal distribution between the eight study locations examined, including a group of five locations with $< 5\%$ DW CaCO_3 in their sediments and three locations with CaCO_3 contents between 20 and 75 %DW (Shapiro–Wilks test, $p < 0.001$, see Supplementary Fig. 1b). Converted into C_{inorg} concentrations (after normalization for the sediment bulk density), we obtain median (IQR) C_{inorg} concentrations in seagrass and mangrove sediments of 59 (66) and 1 (21) $\text{mgC}_{\text{inorg}} \text{cm}^{-3}$, respectively (means \pm SE of 63 ± 11 and $35 \pm 17 \text{ mgC}_{\text{inorg}} \text{cm}^{-3}$) (Fig. 2a).

Using the median SARs in seagrass and mangrove ecosystems compiled in this study (0.22 and 0.23 cm yr^{-1} , respectively; Fig. 2b), we estimate median (IQR) C_{inorg} burial rates in seagrass and mangrove ecosystems of 87 (154) and 6 (207) $\text{gC}_{\text{inorg}} \text{m}^{-2} \text{yr}^{-1}$, respectively (means \pm SE of 182 ± 94 and $90 \pm 43 \text{ gC}_{\text{inorg}} \text{m}^{-2} \text{yr}^{-1}$) (Fig. 2c, Fig. 3). These values correspond to vertical accretion rates of CaCO_3 of the order of 0.1 and 0.001 cm yr^{-1} in seagrass and mangrove ecosystems, respectively. Our mean SAR values agree with previously reported global values^{1,3}. However, our new estimates of burial rates are lower than the previous, indirect median estimate of C_{inorg} burial rate of $108 \text{ gC}_{\text{inorg}} \text{m}^{-2} \text{yr}^{-1}$ (mean \pm SE of $126 \pm 31 \text{ gC}_{\text{inorg}} \text{m}^{-2} \text{yr}^{-1}$) reported by Mazarrasa et al.¹³.

Global annual burial rates of C_{inorg} . Scaling up to the global seagrass coverage ($150,000$ – $600,000 \text{ km}^2$)⁹, the annual burial rate of C_{inorg} ranged from 13 to 52 $\text{TgC}_{\text{inorg}} \text{yr}^{-1}$ for the twentieth century (Table 1). Partitioning between tropical and non-tropical seagrass meadows as in Mazarrasa et al.¹³ showed that 90% of the global C_{inorg} burial occurs in the tropics (Table 1). In seagrass meadows, our estimates of global burial of C_{inorg} are equivalent to 31–55% of the available estimates of contemporary C_{org} burial rates (48 – $112 \text{ TgC}_{\text{org}} \text{yr}^{-1}$)^{1,7}, depending on the estimated global seagrass coverage considered. If all buried CaCO_3 is locally produced (i.e., of autochthonous origin), the burial rates of C_{inorg} in seagrass meadows would represent emissions of 8 – 37 TgC yr^{-1} to the atmosphere and thus would offset their role as CO_2 sinks through the sequestration of C_{org} by ~ 17 – 33% .

The extent of global mangrove coverage yields a median burial rate of $0.8 \text{ TgC}_{\text{inorg}} \text{yr}^{-1}$ (mean of $13 \text{ TgC}_{\text{inorg}} \text{yr}^{-1}$) (Table 1). This value should be considered as a first-order estimate because of the scarcity of data available on C_{inorg} burial rates in mangroves and because of the non-normal distribution between CaCO_3 -rich and CaCO_3 -poor mangrove sediments (Supplementary Figure 1b). When comparing with the global C_{org} burial rates estimate of $31 \text{ TgC}_{\text{org}} \text{yr}^{-1}$ ³, the median C_{inorg} burial rates would correspond to a negligible reduction of net atmospheric CO_2 uptake. However, assuming that all sedimentary CaCO_3 was produced in situ, the C_{inorg} burial rates can largely outweigh C_{org} burial in CaCO_3 -rich mangroves. For example, the C_{inorg} burial rate corresponds to 10–20 times the C_{org} burial rate in the Arabian Peninsula^{17–19}.

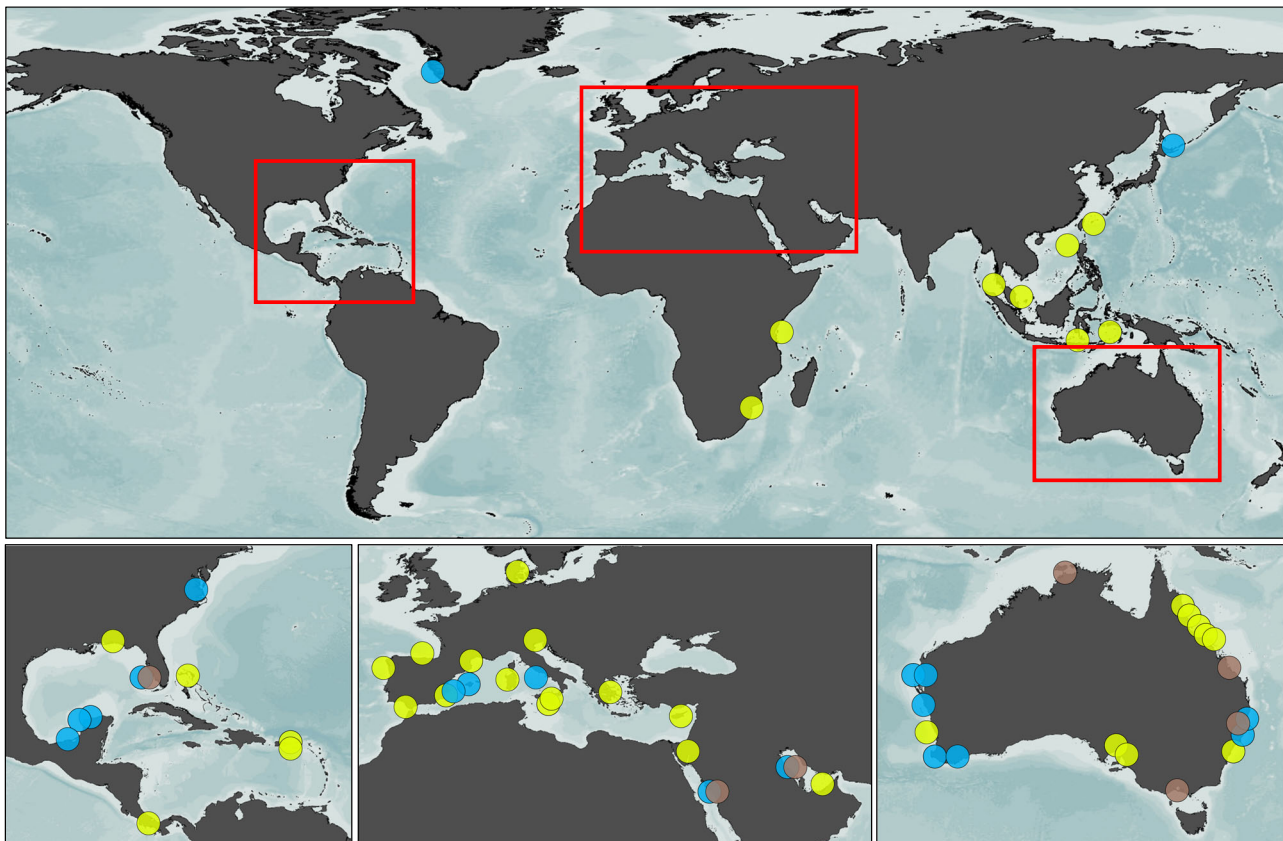


Fig. 1 World map of sediment cores locations. Brown circles: mangrove cores locations; blue: seagrass cores locations; yellow: seagrass cores non-dated but with inorganic carbon content measured¹³

Discussion

CaCO_3 burial in Blue Carbon ecosystems is the balance between inputs (autochthonous and allochthonous) and losses (dissolution and export). Assessments of the mass balance of CaCO_3 in seagrass meadows are few and none have been reported, to our knowledge, in mangrove forests. For seagrass ecosystems, we assessed the balance between calcification, dissolution and burial of CaCO_3 in three locations: the Balearic Islands, Spain^{20,21}, Shark Bay in Western Australia²² and Florida Bay, USA^{23,24} (Table 2).

The most comprehensive assessment of seagrass carbon budgets is that reported for a Mediterranean *Posidonia oceanica* meadow at Magalluf (Mallorca Island, Spain)^{20,21,25,26}. In this meadow, Barrón et al.²¹ estimated a net CO_2 uptake by primary production of $8.4 \text{ gC m}^{-2} \text{ yr}^{-1}$. This estimate was corroborated by the C_{org} burial rate, estimated independently, at $9 \pm 2 \text{ gC}_{\text{org}} \text{ m}^{-2} \text{ yr}^{-1}$ ²⁷. Barrón et al.²¹ also estimated net calcification rates of $51 \text{ gCaCO}_3 \text{ m}^{-2} \text{ yr}^{-1}$, corresponding to $6 \text{ gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$. This amount of calcification would result in a CO_2 emission of $3.6 \text{ gC m}^{-2} \text{ yr}^{-1}$ (0.6-fold the net calcification¹⁴). The CO_2 emission by calcification therefore represents an offset of 40% of the CO_2 uptake from net primary production (thereby yielding a total CO_2 sequestration of $4.8 \text{ gC m}^{-2} \text{ yr}^{-1}$ ²¹). However, the C_{inorg} burial rate in this meadow is estimated here at $226 \text{ gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$. This is two orders of magnitude greater than the net calcification rate of $6 \text{ gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$ ²¹ (Table 2). This implies that about 90% of the CaCO_3 burial in this seagrass meadow must be supported by allochthonous inputs. Therefore, calculation of the CO_2 sequestration by comparing C_{org} and C_{inorg} burial rates or stocks would have concluded that this meadow is a strong source of CO_2 , whereas estimates of calcification rates and net primary production concludes that it is a sink (as confirmed independently through air–sea flux measurements²⁶).

Similarly, in Shark Bay, the burial of C_{inorg} is four times higher than the independently reported net calcification rate²² (Table 2). This again could require large allochthonous carbonate inputs.

In Florida Bay, the low ratio between C_{org} and C_{inorg} concentration in the sediment (g cm^{-3}) implied that seagrass meadows may be a net source of CO_2 to the atmosphere¹⁵. However, such assessment requires consideration of the full carbon mass balance, including accounting for allochthonous inorganic carbon inputs and the balance between calcification and dissolution in the meadows. The contemporary C_{inorg} burial rates in Florida Bay are approximately ninefold higher than the global median, whereas median SAR is about fourfold higher than estimated globally, in an area where 80% of the sediment dry mass is composed of CaCO_3 . However, attempts to assess the gross or net calcification rates in the area yielded values one and two orders of magnitude lower than the estimated CaCO_3 burial rates (Table 2)^{23,24}. In contrast, past geological work in the Bay has suggested that it is a net producer of CaCO_3 ²⁸. It is likely to be that some areas within this large Bay act as sources of CaCO_3 and some others as sinks, helping explain the discrepancy between reported production and burial estimates. Hence, internal redistribution of CaCO_3 production within Florida Bay needs to be considered when drawing inferences on the role of seagrass meadows from sediment composition.

These three example locations are in areas close to coral reefs and/or terrestrial lithogenic sources of CaCO_3 . We could not find estimates of calcification rates (net or gross) in areas without external sources of CaCO_3 . The discrepancies between calcification rates and burial rates in the three example locations could indicate that an important fraction of CaCO_3 burial (> 90%) is supported by CaCO_3 produced elsewhere and trapped in the seagrass sediments. This conclusion is consistent with

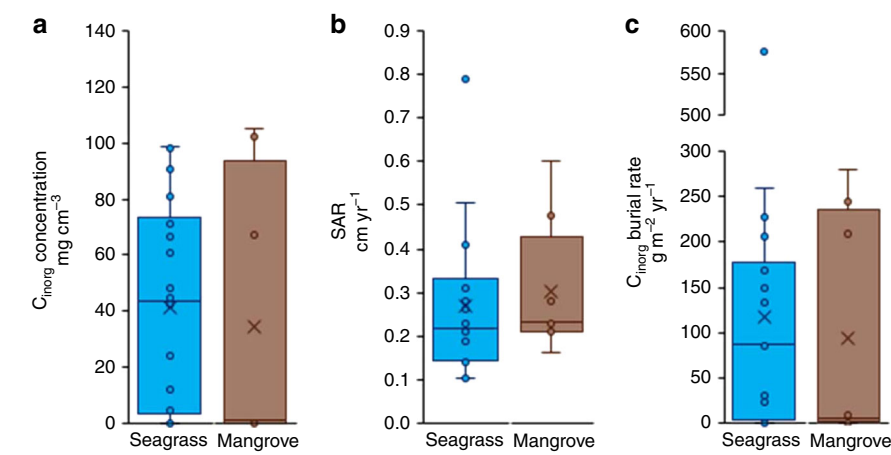


Fig. 2 Sediment cores data. **a** Inorganic carbon (C_{inorg}) concentration, **b** sediment accumulation rates (SAR), and **c** C_{inorg} burial rates. The x represents the mean. Bars are the first and last quartile

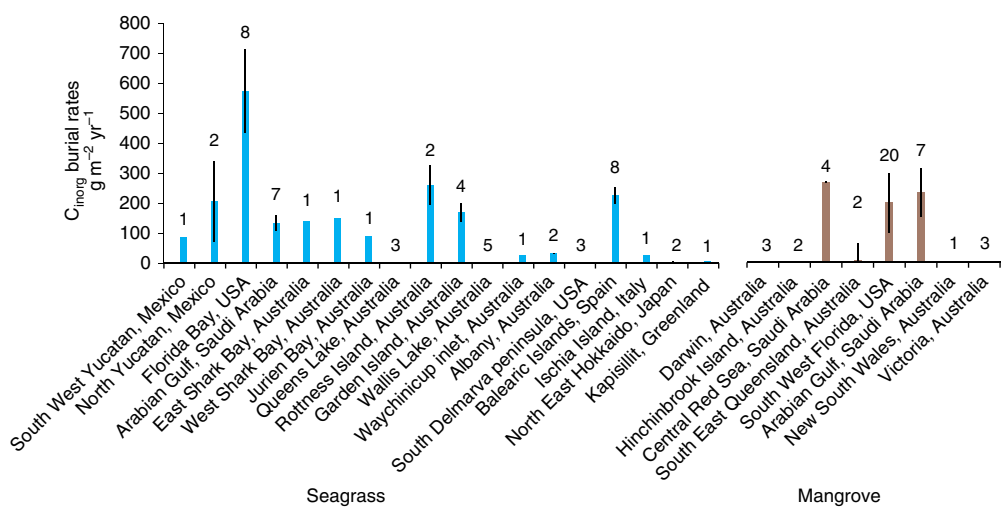


Fig. 3 Inorganic carbon burial rates in all locations. Mean C_{inorg} burial rates in all locations in sediment cores for seagrass meadows and mangrove forests, organized from low to high latitudes. Bars are the SE. Labels are the number of cores per location

Table 1 Median (mean) global C_{inorg} burial rates for seagrass meadows and mangrove forests considering one, and, for seagrass, two world regions (tropical and higher latitudes)				
		Burial rate, ($TgC_{inorg} yr^{-1}$)		
		Global	Tropical	Higher lat.
Seagrass	This study	13(27)–52(109)	14(41)–57(163)	1(3)–5(14)
	Mazarassa et al. ¹³	19(28)–65(79)		
Mangrove	This study	0.8(12)		

comparable C_{inorg} concentrations within and outside seagrass meadows, whereas, in contrast, C_{org} concentrations are higher in seagrass sediments¹³. A large role of C_{inorg} import is also consistent with the large $CaCO_3$ export from coral reefs to adjacent waters, equivalent to 25–50% of the $CaCO_3$ production, predominantly to reef lagoons²⁷, where seagrass meadows and mangroves often grow. Mangroves, seagrass and saltmarsh ecosystems are likely to be sites of net carbonate dissolution. Roots of

marine plants release organic compounds and oxygenate the sediments during the day, promoting microbial aerobic remineralization of organic matter, thereby increasing sedimentary respiratory CO_2 ^{29,30} and/or stimulating the re-oxidation of reduced metabolites. These processes result in the release of strong acids (e.g., H_2SO_4 , HNO_3)^{31–33}, which leads to $CaCO_3$ dissolution in the sediment^{34,35} (although re-precipitation can occur³⁴).

Table 2 Burial rates of CaCO_3 compared to calcification rates in seagrass ecosystems

	Community production rate of CaCO_3		Community net calcification rate		Sediment		
	$\text{gCaCO}_3 \text{ m}^{-2} \text{ yr}^{-1}$	$\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$	$\text{gCaCO}_3 \text{ m}^{-2} \text{ yr}^{-1}$	$\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$	% CaCO_3	$\text{gCaCO}_3 \text{ m}^{-2} \text{ yr}^{-1}$	$\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$
Florida Bay, USA	626 ^{23,24}	75	182 ²⁵	2.2	82 ± 2	4792 ± 756	756 ± 91
Balearic Islands, Spain	68 ²⁰	8	51 ²¹	6	81 ± 3	1886 ± 214	226 ± 30
West Shark Bay, Australia	375 ± 62 ²²	45 ± 7	295 ²²	35	60 ± 5	1240 ± 232	149 ± 30

Comparison between seagrass-associated community production rate of carbonate (obtained from standing stock assessments and leaves or calcifiers turnover rates) and community net calcification rates (balance between calcification and dissolution, calculated from variations of total alkalinity) from the literature, and carbonate burial rate in three locations with carbonate-rich sediments

Dissolution of CaCO_3 might also be influenced by the CO_2 system in the water column of Blue Carbon ecosystems. Respiration and photosynthesis of the flora and fauna, together with sediment redox processes in seagrass and mangrove ecosystems, strongly influence the chemistry of the water column, generating large diel amplitudes of the saturation state for CaCO_3 (Ω) with a tendency towards dissolution or the reduction of calcification at nighttime, amplified at low tide^{36–40}. The dissolution of allochthonous CaCO_3 in carbonate platform areas, caused directly or indirectly by metabolism of the marine vegetation and associated biota, leads to a reduction in pCO_2 through the release of fossilized total alkalinity. This sink of atmospheric CO_2 should be incorporated into the Blue Carbon framework. A recent assessment considers alkalinity addition through the dissolution of allochthonous carbonate as a very effective geo-engineering approach to remove atmospheric CO_2 and mitigate climate change^{41,42}.

Similarly, saltmarshes are not known to host high levels of calcifying organisms but can accumulate CaCO_3 from allochthonous sources. In arid tropical saltmarshes of the Western Arabian Gulf, dominated by succulent shrubs, a concentration of CaCO_3 of 57 ± 8% in sediments and a contemporary burial rate of C_{inorg} of 100 ± 15 $\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$ (mean ± SE) were found¹⁷. In a temperate saltmarsh of the Western Scheldt estuary in the Netherlands, a concentration of CaCO_3 of 14 ± 1% and a high contemporary burial rate of 467 ± 99 $\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$, mostly due to high SAR (1.1 ± 0.3 cm yr^{-1}), were measured⁴³. Yet, this does not imply that CaCO_3 dynamics have a negligible role in saltmarsh carbon budgets, as they may still act as sites of net dissolution of CaCO_3 , adding to the removal of CO_2 associated with C_{org} burial. A dissolution rate of 24–96 $\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$ was estimated in the sediment of the saltmarshes of the Eastern Scheldt estuary, corresponding to ~85% of the C_{inorg} burial rate⁴⁴.

To further examine the conclusion that Blue Carbon ecosystems are sites of substantial allochthonous CaCO_3 deposition, based on existing mass balances for Blue Carbon sediments, we examined (qualitatively) the relationship between the CaCO_3 % DW in sediments and the presence/absence of sources of CaCO_3 adjacent to the coring locations, including coral reefs and terrestrial lithogenic sources of CaCO_3 (Fig. 4, see dataset in Supplementary Information). Seagrass and mangrove ecosystems without potentially large adjacent allochthonous CaCO_3 sources have a remarkably lower median (IQR) sediment CaCO_3 content of 4 (15) and 1 (1) %DW (means ± SE of 11 ± 4 and 1.7 ± 0.8 % DW), respectively, compared with 59 (51) and 61 (27) %DW (means ± SE of 56 ± 5 and 53 ± 16 %DW) when at least one allochthonous CaCO_3 source was present (Fig. 3). For sediments in seagrass meadows, the presence of coral reefs (t -value = 4.68, df = 48.5, p < 0.0001) and lithogenic sources (t -value = 4.76, df = 57.3, p < 0.0001) increased the amount of CaCO_3 in the sediment. However, there was a significant interaction between these factors (t -value = -3.29, df = 53.2, p = 0.0018), because the CaCO_3 %

DW in the presence of both allochthonous sources was less than would be expected if these variables were additive. The presence/absence of coral reefs and lithogenic sources accounted for 36% of the variation in CaCO_3 , whereas the random variables (study, lithology grouping and marine province) accounted for 54% of the variation in CaCO_3 (see Methods for model description). Mangrove sediment samples showed a similar pattern to the seagrass meadows and the presence of allochthonous sources had a marginally significant positive effect on the amount of CaCO_3 in the sediment (t -value = 4.29, df = 1.81, p = 0.0596). The presence/absence of a CaCO_3 source accounted for 71% of the variation of in CaCO_3 within mangrove sediments, whereas the random variables accounted for 20% of the variation in CaCO_3 . In testing for biases of outlying cores and studies, we found that one study from Western Florida had an outlying data point that disproportionality skewed the results. The study from Western Florida had relatively low CaCO_3 but did have an allochthonous source of CaCO_3 . When this study was removed from the analysis, the presence of an allochthonous source became significant (t -value = 7.92, df = 4.16, p = 0.0012). This highlights the need for more studies in mangrove sediments to determine the global influence of allochthonous sources on CaCO_3 content.

In seagrass meadows, the median (IQR) C_{inorg} burial rate found in areas where no allochthonous sources were identified was 1 (13) $\text{gC}_{\text{inorg}} \text{ m}^{-2} \text{ yr}^{-1}$ (mean ± SE of 8 ± 4), only 1.1% of the global median. This contrast is consistent with our hypothesis that much of the C_{inorg} buried in seagrass and mangrove sediments is allochthonous. It explains the non-normal distribution of CaCO_3 concentrations observed in sediments of seagrass and mangroves (Supplementary Figure 1), and indicates that the import of CaCO_3 from carbonate-forming ecosystems or adjacent karstic areas is the norm²⁷. The global burial rate of C_{inorg} in seagrass meadows is between a third to a half of their C_{org} burial rate^{1,7}, whereas for mangroves our first estimate of global C_{inorg} burial rates is only 3% of the C_{org} burial rate³. If the buried CaCO_3 and C_{org} in seagrass sediments were produced entirely in situ, C_{inorg} burial would offset up to a third of CO_2 sequestration through C_{org} burial, particularly in tropical seagrass ecosystems where ~90% of the global C_{inorg} burial occurs. However, imbalances between production, dissolution and burial, and the observation of much higher CaCO_3 concentrations in sediments near lithogenic formations and coral reefs, suggest that, where present, allochthonous CaCO_3 inputs are substantial and support most of the net CaCO_3 burial.

Locally, despite supporting significant CaCO_3 burial, Blue Carbon ecosystems may be sites where imported CaCO_3 dissolves, strengthening rather than weakening the capacity of these ecosystems to sequester CO_2 . Whereas there is emphasis on apportioning the sources of autochthonous and allochthonous C_{org} in Blue Carbon sediments (up to 50% of the buried C_{org})^{4,9}, determining the sources of CaCO_3 in Blue Carbon sediments is just as important, to resolve the role of vegetated coastal

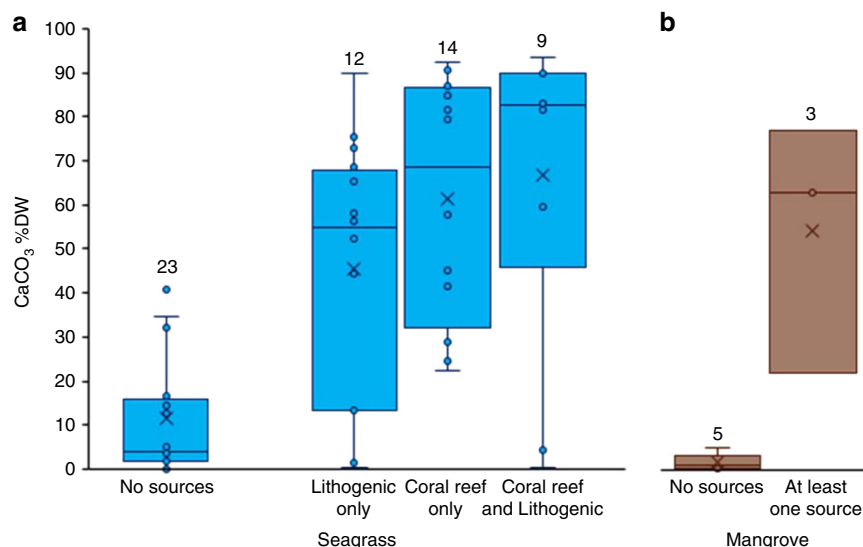


Fig. 4 Allochthonous sources and inorganic carbon in sediments. **a** %DW of carbonates (CaCO_3) in seagrass sediments depending on the presence of potential allochthonous sources (lithogenic and/or coral reefs). All data distributions for locations with allochthonous sources are significantly different to the distributions for locations without allochthonous sources (Mann-Whitney U -tests, all $p < 0.001$). **b** %DW CaCO_3 in mangrove sediments with and without allochthonous sources (coral reef and lithogenic source). Number on top of the box plots indicate the number of locations. The x represents the mean. Bars are the first and last quartile

ecosystems as CO_2 sinks and, hence, their potential to support climate change mitigation. The current focus on C_{org} budgets in vegetated coastal ecosystems needs to be augmented with integrative assessments of organic and inorganic carbon fluxes and budgets, including both allochthonous and autochthonous inputs. Moreover, these assessments must consider the sources and fate of carbon exchanged between Blue Carbon and adjacent ecosystems, as Blue Carbon ecosystems export important amounts of their organic production^{45–47} but also import significant amounts of CaCO_3 and organic matter from adjacent sources. A comparison of paired vegetated and unvegetated sediment CaCO_3 % DW showed that vegetated and adjacent unvegetated sediments have similar carbonate concentrations, both using standard parametric statistics (general linear model (GLM), t -value = 1.32, $df = 83.1$, $p = 0.191$) and meta-analysis (z -value = 0.88, $p = 0.379$; Supplementary Fig. 2A,B), which also showed no evidence for reporting bias (all points within the 95% confidence lines of the funnel plot, Supplementary Fig. 2C). This provides further support to the hypothesis that much of the carbonate buried in vegetated coastal sediments derives from allochthonous sources rather than being produced within the habitat.

Inorganic carbon burial in Blue Carbon ecosystems has been overlooked, with the rates compiled here representing the first direct estimates reported in the literature. These estimates confirm that seagrass ecosystems, and to lesser extent mangrove ecosystems, are intense sites of CaCO_3 burial, supporting sediment accretion. CaCO_3 burial is a fundamental process supporting the role of Blue Carbon ecosystems in climate change adaptation, which is underpinned by their capacity to rapidly accrete sediments, reducing relative SLR by raising the seafloor^{1,17}.

Methods

Calculation of the C_{inorg} accretion rate. We searched the peer-reviewed literature for data on sediment cores dated with ^{210}Pb , including CaCO_3 or C_{inorg} concentration in seagrass and mangrove sediments. Search terms on Google Scholar were seagrass OR mangrove AND ^{210}Pb OR SAR OR sediment accretion rate. We then searched returned articles that contained data on SAR and CaCO_3 or C_{inorg} data. We found only one study presenting CaCO_3 content in a dated sediment core. However, we found 15 and 22 studies with SAR for seagrass and mangrove

sediments, respectively. To obtain the CaCO_3 or C_{inorg} concentrations needed to calculate C_{inorg} burial rates, we used the database of Mazarrasa et al.¹³, which was the most recent exhaustive compilation of sediment cores from Blue Carbon habitats, for data on CaCO_3 in seagrass sediments. We also contacted experts in Blue Carbon studies (published studies using cores from Blue Carbon habitats) for unpublished CaCO_3 sediment concentration data (see data and references in Supplementary Data 1). In total, we compiled 42 and 53 ^{210}Pb dated cores with CaCO_3 content in mangrove and seagrass ecosystems, respectively (see PRISMA checklist and flow diagram⁴⁸ in Supplementary Note 1).

The SARs (cm yr^{-1}) from the literature were re-calculated according to the constant flux–constant sedimentation model⁴⁹, to have a coherent and comparable dating system between all cores. The CaCO_3 concentration (% sediment DW) was calculated as the mean between all slices younger than 1900, for cores with the contemporary ^{210}Pb chronologies. The C_{inorg} concentration in sediment ($\text{gC}_{\text{inorg}} \text{m}^{-3}$) was calculated from the dry bulk density (g m^{-3}) and the percentage of CaCO_3 content (using sediment DW), considering a mass ratio of 12% carbon in CaCO_3 . The C_{inorg} burial rate ($\text{gC}_{\text{inorg}} \text{m}^{-2} \text{yr}^{-1}$) was then calculated as the product of the SAR and the C_{inorg} concentration for each sediment core. Cores with negligible content of CaCO_3 were also included in the calculation (see Supplementary Figure 1).

All cores from the same site or area and with similar presence or absence of allochthonous sources of CaCO_3 (see below) were treated as replicates for a global location and averaged for the analysis (geologic grouping). For seagrass, the 51 cores dated with ^{210}Pb were grouped into 17 locations (Figs. 2, 3). For mangroves, we compiled a total of 42 cores dated with ^{210}Pb in 8 locations (Figs. 2, 3). Seagrass locations ranged from tropical to sub-arctic locations, with 50% of estimates derived from tropical and subtropical locations and 50% from higher latitudes. Mangrove sediment derived mostly from subtropical locations (seven out of eight locations), particularly in Australia and the Arabian Peninsula (Supplementary Figure 2).

Determination of the influence of allochthonous sources of CaCO_3 . We analysed the influence of the presence/absence of proximity of coral reefs and continental surface lithology (qualitative data), as potential allochthonous sources of CaCO_3 in seagrass and mangrove sediments (in %DW) (see dataset in Supplementary Data 1). For seagrass, we expanded our dataset by including CaCO_3 concentrations from 264 cores compiled by Mazarrasa et al.¹³, reaching a total of 341 cores with measured CaCO_3 %DW.

We estimated the presence/absence of coral reefs using the map of the global distribution of warm-water coral reefs compiled by the UNEP-WCMC⁵⁰ and the presence/absence of nearby lithogenic sources using the global lithology map of Hartmann and Moosdorf⁵¹ and the world soil map of the FAO/UNESCO (<http://www.fao.org/soils-portal/en/>). The coring locations were associated with climate regions following the Köppen–Geiger classification system⁵².

Statistical analysis. All data distributions were tested for normality to determine the most reliable central tendency measured with Shapiro–Wilks normality test (Statistica, Dell Software). None of the datasets of SAR, C_{inorg} concentration, C_{inorg}

burial rate or CaCO_3 %DW were normally distributed (all $p < 0.05$). We therefore chose to use the median (IQR) as the most appropriate description of central tendency. Traditional meta-analysis tools, which calculate effect sizes to standardize the difference between control and experimental treatments, thereby allowing comparison among disparate response variables and weighting to account for unequal variance among studies, could not be used for this analysis for multiple reasons. These reasons include that the question posed and the studies available did not include experimental designs with paired control and experimental plots required for effect size calculations, that there was a single response variable facilitating direct comparison and data integration, and, most importantly, that we used the raw data for each core. Instead, we ran a statistical test using a mixed effect GLM to determine the effect of coral reefs and lithogenic sources on the CaCO_3 % DW of the sediment. For sediments within seagrass meadows, the GLM included two fixed factors (presence/absence of coral reefs and of lithogenic sources), as well as the interaction between the two factors. For sediment within mangrove forests, the GLM included one fixed factor (presence/absence of allochthonous sources), because replication did not exist for all combinations of the two factors. The data had unequal samples among studies and studies were not evenly distributed around the globe (Fig. 1), which could result in pseudo-replication and biased results. To account for the data structure and minimize non-independence, we included three separate random variables, which included study, lithology grouping and marine province. The marine province was determined for each sample location using the marine provinces of the world as defined by Spalding et al.⁵³. Separate models were run for seagrass and mangrove sites. The statistical model was produced using the lmer function within the lme4 package⁵⁴ and p -values were calculated with the lmerTest package⁵⁵. The R^2 was calculated for the fixed and random effects using the r.squared GLMM function in the MuMIn package⁵⁶. The response variable was log transformed, which improved the model fit compared with raw data. The model fit was assessed by plotting the Q-Q plot (linear relationship) and the fitted values compared with the residuals (random distribution). To test whether individual cores or studies were biased and having a disproportionate influence on findings, we systematically removed any studies that contained outlying samples as determined from being outside the 95% confidence interval for the fitted values vs. residuals comparison using the plot model function from sjPlot package⁵⁷. This analysis was conducted in R version 3.4.2.

Reporting bias and its effect on findings is an important consideration for meta-analyses⁵⁸ and when the result from a meta-analysis is not the same as it would have been if data from all correctly conducted studies were included in the analysis⁵⁹. A main cause of reporting bias is not publishing research because of a lack of merit as determined by the researcher, reviewer or editor⁶⁰. As indicated by the data inclusion flow diagram (Supplementary Fig. 3), researchers often measured but did not publish data on soil CaCO_3 content and authors needed to be directly contacted for these results. In addition, the researchers not only provided information from published studies but also unpublished data on CaCO_3 content (10 of 51 seagrass studies included in the analysis were not published). For these reasons, it is unlikely to be that our findings were affected by reporting bias. A subset of data collected for this study included the appropriate information to run both a GLM and a traditional meta-analysis (effect size could be calculated between paired data). The data included information from nine studies that measured the CaCO_3 content of sediment from both vegetated and unvegetated habitats. There were 92 core samples with 32 from unvegetated and 60 from vegetated habitats (Supplementary Fig. 2A). The GLM followed the same procedures as detailed in the main text, except it had only two random factors, study and marine province, because study and lithology grouping differed in only one instance. For the meta-analysis, the data were paired for each study and the mean CaCO_3 %DW, number of samples and SD were calculated for vegetated and unvegetated cores for each study. Two studies included in the GLM were removed for the meta-analysis, because they only had one core for an unvegetated habitat and SD could not be calculated, leaving seven comparisons for this analysis. Hedges' g was calculated for the effect size following Borenstein et al.⁶⁰ (Eqs. 1–3) and a variance for each effect size was also calculated (V_g)⁵⁹ (Eq. 4), as indicated by equations:

$$\text{Hedges' } g = \frac{(X_E - X_C)J}{\text{SD}_{\text{pooled}}} \quad (1)$$

$$\text{SD}_{\text{pooled}} = \sqrt{\frac{(n_E - 1)(\text{SD}_E)^2 + (n_C - 1)(\text{SD}_C)^2}{n_E + n_C - 2}} \quad (2)$$

$$J = 1 - \frac{3}{4(n_E + n_C - 2) - 1} \quad (3)$$

$$V_g = \frac{n_E + n_C}{n_E \times n_C} + \frac{g^2}{2(n_E + n_C)} \quad (4)$$

X_E and X_C are the mean (n is sample size) of vegetated and unvegetated sediments, along with $\text{SD}_{\text{pooled}}$ and J , which accounts for biases associated with different sample sizes. The meta-analysis included the same two random variables as the GLM and was conducted using the rma.mv function from the metafor package⁶¹.

Calculation of global yearly burial rates of C_{inorg} . The global annual burial of inorganic carbon ($\text{TgC}_{\text{inorg}} \text{ yr}^{-1}$) in seagrass meadows was calculated as the product of the global median C_{inorg} burial rates and the estimated global seagrass area, which ranges from 150,000 to 600,000 km^2 ²⁹. We also calculated the global annual burial of C_{inorg} as the sum of separate calculations for tropical and arid climates and meadows at higher latitude climates. Median C_{inorg} burial rates were calculated for tropical (core locations with tropical and hot desert climates) and non-tropical areas (temperate, continental and polar climates) and multiplied by the global seagrass cover range under the assumption that 2/3 of the seagrass area is in the tropical and subtropical zone¹³. The global annual burial of inorganic carbon ($\text{TgC}_{\text{inorg}} \text{ yr}^{-1}$) in mangroves was calculated as the product of the global median C_{inorg} burial rates and the estimated global mangrove cover of 137,760 km^2 ⁶².

Data availability

The dataset is available as Supplementary Data 1.

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References

- Duarte, C. M., Losada, I. J., Hendriks, I. E., Mazarrasa, I. & Marbà, N. The role of coastal plant communities for climate change mitigation and adaptation. *Nat. Clim. Chang.* **3**, 961–968 (2013).
- Duarte, C. M., Middelburg, J. & Caraco, N. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* **1**, 1–8 (2005).
- Rosentreter, J. A., Maher, D. T., Erler, D. V., Murray, R., & Eyre, B. D. CH_4 emissions partially offset 'blue carbon' burial in mangroves. *Sci. Adv.* **4**, ea04985 (2018).
- Kennedy, H. et al. Seagrass sediments as a global carbon sink: Isotopic constraints. *Glob. Biogeochem. Cycles* **24**, 1–8 (2010).
- Alongi, D. M. Carbon cycling and storage in mangrove forests. *Ann. Rev. Mar. Sci.* **6**, 195–219 (2014).
- Nellemann, C. et al. (eds.) *Blue Carbon: A rapid response assessment*. (United Nations Environment Programme, GRID-Arendal, 2009).
- McLeod, E. et al. A blueprint for blue carbon: toward an improved understanding of the role of vegetated coastal habitats in sequestering CO_2 . *Front. Ecol. Environ.* **9**, 552–560 (2011).
- Chmura, G. L., Anisfeld, S. C., Cahoon, D. R. & Lynch, J. C. Global carbon sequestration in tidal, saline wetland soils. *Glob. Biogeochem. Cycles* **17**, 1111 (2003).
- Duarte, C. M. Reviews and syntheses: hidden forests, the role of vegetated coastal habitats in the ocean carbon budget. *Biogeosciences* **14**, 301–310 (2017).
- Kristensen, E., Bouillon, S., Dittmar, T. & Marchand, C. Organic carbon dynamics in mangrove ecosystems: a review. *Aquat. Bot.* **89**, 201–219 (2008).
- Fourqurean, J. W. et al. Seagrass ecosystems as a globally significant carbon stock. *Nat. Geosci.* **5**, 505–509 (2012).
- Sanderman, J. et al. A global map of mangrove forest soil carbon at 30 m spatial resolution. *Environ. Res. Lett.* **13**, 055002 (2018).
- Mazarrasa, I. et al. Seagrass meadows as a globally significant carbonate reservoir. *Biogeosciences* **12**, 4993–5003 (2015).
- Ware, J. R., Smith, S. V. & Reaka-Kudla, M. L. Coral reefs: sources or sinks of atmospheric CO_2 ? *Coral Reefs* **11**, 127–130 (1992).
- Howard, J. L., Creed, J. C., Aguiar, M. V. P. & Fouquerean, J. W. CO_2 released by carbonate sediment production in some coastal areas may offset the benefits of seagrass 'Blue Carbon' storage. *Limnol. Oceanogr.* **63**, 160–172 (2017).
- Macreadie, P. I., Serrano, O., Maher, D. T., Duarte, C. M. & Beardall, J. Addressing calcium carbonate cycling in blue carbon accounting. *Limnol. Oceanogr. Lett.* **2**, 195–201 (2017).
- Saderne, V. et al. Accumulation of carbonates contributes to coastal vegetated ecosystems keeping pace with sea level rise in an arid region (Arabian Peninsula). *J. Geophys. Res. Biogeosci.* **2050**, 1–13 (2018).
- Almahasheer, H. et al. Low carbon sink capacity of Red Sea mangroves. *Sci. Rep.* **7**, 1–10 (2017).
- Cusack, M. et al. Organic carbon sequestration and storage in vegetated coastal habitats along the western coast of the Arabian Gulf. *Environ. Res. Lett.* **13**, 074007 (2018).
- Canals, M. & Ballesteros, E. Production of carbonate by phytobenthic communities on the Mallorca -Menorca shelf, northwestern Mediterranean sea. *Deep Sea Res. Part II* **44**, 611–629 (1997).
- Barrón, C., Duarte, C. M., Frankignoulle, M. & Borges, A. V. Organic carbon metabolism and carbonate dynamics in a Mediterranean seagrass (*Posidonia oceanica*), meadow. *Estuaries Coasts* **29**, 417–426 (2006).

22. Walker, D. & Woelkerling, W. Quantitative study of sediment contribution by epiphytic coralline red algae in seagrass meadows in Shark Bay, Western Australia. *Mar. Ecol. Prog. Ser.* **43**, 71–77 (1988).
23. Bosence, D. W. J. Biogenic carbonate production in Florida Bay. *Bull. Mar. Sci.* **44**, 419–433 (1989).
24. Yates, K. K. & Halley, R. B. Diurnal variation in rates of calcification and carbonate sediment dissolution in Florida Bay. *Estuaries Coasts* **29**, 24–39 (2006).
25. Mazarrasa, I. et al. Effect of environmental factors (wave exposure and depth) and anthropogenic pressure in the C sink capacity of *Posidonia oceanica* meadows. *Limnol. Oceanogr.* **62**, 1436–1450 (2017).
26. Gazeau, F. et al. Whole-system metabolism and CO₂ fluxes in a Mediterranean Bay dominated by seagrass beds (Palma Bay, NW Mediterranean). *Biogeosciences* **2**, 43–60 (2005).
27. Milliman, J. D. Production and accumulation of calcium carbonate in the ocean: Budget of a nonsteady state. *Glob. Biogeochem. Cycles* **7**, 927–957 (1993).
28. Stockman, K. W., Ginsburg, R. N. & Shinn, E. A. The production of lime mud by algae in South Florida. *J. Sed. Petrol.* **37**, 633–648 (1967).
29. Mucha, A. P., Almeida, C. M. R., Bordalo, A. A. & Vasconcelos, M. T. S. D. Exudation of organic acids by a marsh plant and implications on trace metal availability in the rhizosphere of estuarine sediments. *Estuar. Coast. Shelf Sci.* **65**, 191–198 (2005).
30. Haoliang, L., Chongling, Y. & Jingchun, L. Low-molecular-weight organic acids exuded by Mangrove (*Kandelia candel* (L.) Druce) roots and their effect on cadmium species change in the rhizosphere. *Environ. Exp. Bot.* **61**, 159–166 (2007).
31. Middelburg, J., Nieuwenhuize, J., Slim, F. & Ohowa, B. Sediment biogeochemistry in an East African mangrove forest (Gazi Bay, Kenya). *Biogeochemistry* **34**, 133–155 (1996).
32. Ku, T. C. W., Walter, L. M., Coleman, M. L., Blake, R. E. & Martini, A. M. Coupling between sulfur recycling and syndepositional carbonate dissolution: evidence from oxygen and sulfur isotope composition of pore water sulfate, South Florida Platform, U.S.A. *Geochim. Cosmochim. Acta* **63**, 2529–2546 (1999).
33. Burdige, D. J. & Zimmerman, R. C. Impact of sea grass density on carbonate dissolution in Bahamian sediments. *Limnol. Oceanogr.* **47**, 1751–1763 (2002).
34. Hu, X. & Burdige, D. J. Enriched stable carbon isotopes in the pore waters of carbonate sediments dominated by seagrasses: evidence for coupled carbonate dissolution and reprecipitation. *Geochim. Cosmochim. Acta* **71**, 129–144 (2007).
35. Linto, N. et al. Carbon dioxide and methane emissions from mangrove-associated waters of the Andaman Islands, Bay of Bengal. *Estuaries Coasts* **37**, 381–398 (2014).
36. Saderne, V., Fietzek, P. & Herman, P. M. J. Extreme variations of pCO₂ and pH in a Macrophyte Meadow of the Baltic Sea in summer: evidence of the effect of photosynthesis and local upwelling. *PLoS ONE* **8**, e62689 (2013).
37. Sippo, J. Z., Maher, D. T., Tait, D. R., Holloway, C. & Isaac, R. Are mangroves drivers or buffers of coastal acidification? Insights from alkalinity and dissolved inorganic carbon export estimates across a latitudinal transect. *Glob. Biogeochem. Cycles* **30**, 753–766 (2016).
38. Challenger, R. C., Robbins, L. L. & McClintock, J. B. Variability of the carbonate chemistry in a shallow, seagrass-dominated ecosystem: implications for ocean acidification experiments. *Mar. Freshw. Res.* **67**, 163–172 (2016).
39. Ray, R., Baum, A., Rixen, T., Gleixner, G. & Jana, T. K. Exportation of dissolved (inorganic and organic) and particulate carbon from mangroves and its implication to the carbon budget in the Indian Sundarbans. *Sci. Total Environ.* **621**, 535–547 (2018).
40. Maher, D. T., Santos, I. R., Golsby-Smith, L., Gleeson, J. & Eyre, B. D. Groundwater-derived dissolved inorganic and organic carbon exports from a mangrove tidal creek: The missing mangrove carbon sink? *Limnol. Oceanogr.* **58**, 475–488 (2013).
41. Gattuso, J.-P. et al. Ocean solutions to address climate change and its effects on marine ecosystems. *Front. Mar. Sci.* **5**, 337 (2018).
42. Maher, D. T., Call, M., Santos, I. R., & Sanders, C. J. Beyond burial: Lateral exchange is a significant atmospheric carbon sink in mangrove forests. *Biol. Lett.* **14**, 20180200 (2018).
43. Zwolsman, J. J. G., Berger, G. W. & Van Eck, G. T. M. Sediment accumulation rates, historical input, postdepositional mobility and retention of major elements and trace metals in salt marsh sediments of the Scheldt estuary, SW Netherlands. *Mar. Chem.* **44**, 73–94 (1993).
44. Vranken, M., Oenema, O. & Mulder, J. Effects of tide range alterations on salt marsh sediments in the Eastern Scheldt, S. W. Netherlands. *Hydrobiologia* **195**, 13–20 (1990).
45. Duarte, C. M. & Cebrián, J. The fate of marine autotrophic production. *Limnol. Oceanogr.* **41**, 1758–1766 (1996).
46. Krause-Jensen, D. & Duarte, C. M. Substantial role of macroalgae in marine carbon sequestration. *Nat. Geosci.* **9**, 737–742 (2016).
47. Duarte, C. M. & Krause-Jensen, D. Export from seagrass meadows contributes to marine carbon sequestration. *Front. Mar. Sci.* **4**, 13 (2017).
48. Moher, D., Liberati, A., Tetzlaff, J., Altman, D. G. & The PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA Statement. *PLoS Med.* **6**, e1000097 (2009).
49. Krishnaswamy, S., Lal, D., Martin, J. M. & Meybeck, M. Geochronology of lake sediments. *Earth Planet. Sci. Lett.* **11**, 407–414 (1971).
50. UNEP-WCMC. *Global distribution of warm-water coral reefs, compiled from multiple sources including the Millennium Coral Reef Mapping Project*, Version 1.3. (UNEP World Conservation Monitoring Centre, 2010).
51. Hartmann, J. & Moosdorf, N. The new global lithological map database GLiM: A representation of rock properties at the Earth surface. *Geochim. Geophys. Geosyst.* **13**, Q12004 (2012).
52. Rubel, F., Brugger, K., Haslinger, K. & Auer, I. The climate of the European Alps: Shift of very high resolution Köppen-Geiger climate zones 1800–2100. *Meteorol. Zeitschrift* **26**, 115–125 (2016).
53. Spalding, M. D. et al. Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. *Bioscience* **57**, 573–583 (2007).
54. Bates, D., Maechler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**, 1–48 (2015).
55. Kuznetsova, A., Brockhoff, P. B. & Christensen, R. H. B. “lmerTest Package: tests in Linear Mixed Effects Models.”. *J. Stat. Softw.* **82**, 1–26 (2017).
56. Bartoň, K. *MuMIn: Multi-Model Inference*. R package version 1.42.1. <https://CRAN.R-project.org/package=MuMIn> (2018).
57. Lüdtke, D. *sjPlot: Data Visualization for Statistics in Social Science*. R package version 2.6.0. <https://CRAN.R-project.org/package=sjPlot> (2018).
58. Gurevitch, J., Koricheva, J., Nakagawa, S. & Stewart, G. Meta-analysis and the science of research synthesis. *Nature* **555**, 175–182 (2018).
59. Koricheva, J., Gurevitch, J. & Mengersen, K. *Handbook of Meta-analysis in Ecology and Evolution* (Princeton Univ. Press, 2013).
60. Borenstein, M., Hedges, L. V., Higgins, J. P. & Rothstein, H. R. *Introduction to meta-analysis*. (John Wiley & Sons, 2011).
61. Viechtbauer, W. Conducting meta-analyses in R with the metafor package. *J. Stat. Softw.* **36**, 1–48 (2010).
62. Giri, C. et al. Status and distribution of mangrove forests of the world using earth observation satellite data. *Glob. Ecol. Biogeogr.* **20**, 154–159 (2011).

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Author contributions

C.M.D. and V.S. conceived and designed this work. V.S. wrote the manuscript with support from N.R.G., P.I.M., D.T.M., J.J.M., O.S. and C.M.D. V.S., N.R.G., P.I.M., D.T.M., J.J.M., O.S., H.A., A.A.-O., M.C., B.D.E., J.W.F., H.K., D.K.-J., T.K., P.S.L., C.E.L., N.M., P.M., M.A.M., I.M., K.J.M.G., M.P.J.O., C.J.S., I.R.S., J.M.S., T.T., K.W. and C.M.D. contributed data and quality check was done by A.A.-O. and P.M. Analysis of data was performed by V.S. and N.R.G. V.S., N.R.G., P.I.M., D.T.M., J.J.M., O.S., H.A., A.A.-O., M.C., B.D.E., J.W.F., H.K., D.K.-J., T.K., P.S.L., C.E.L., N.M., P.M., M.A.M., I.M., K.J.M.G., M.P.J.O., C.J.S., I.R.S., J.M.S., T.T., K.W. and C.M.D. discussed and commented on the manuscript.

Additional information

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