Tracking the assembly of nested parasite communities: using  $\beta$ -diversity to understand

## variation in parasite richness and composition over time and scale

Running header: Parasite community assembly

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# ABSTRACT

- Community composition is driven by a few key assembly processes: ecological selection, drift, and dispersal. Nested parasite communities represent a powerful study system for understanding the relative importance of these processes and their relationship with biological scale. Quantifying β-diversity across scales and over time additionally offer mechanistic insights into the ecological processes shaping the distributions of parasites and therefore infectious disease.
- 7 2. To examine factors driving parasite community composition, we quantified the parasite communities of 959 amphibian hosts representing two species (the Pacific chorus frog. 8 Pseudacris regilla, and the California newt, Taricha torosa) sampled over three months 9 from 10 ponds in California. Using additive partitioning, we estimated how much of 10 regional parasite richness ( $\gamma$ -diversity) was composed of within-host parasite richness ( $\alpha$ -11 diversity) and turnover (β-diversity) at three biological scales: across host individuals, 12 across species, and across communities (ponds). We also examined how β-diversity varied 13 across time at each biological scale. 14
- 15 3. Differences among ponds comprised the majority (40%) of regional parasite diversity, followed by differences among host species (23%) and among host individuals (12%). Host 16 species supported parasite communities that were less similar than expected by null models. 17 consistent with ecological selection, although these differences lessened through time, 18 likely due to high dispersal rates of infectious stages. Host individuals within the same 19 population supported more similar parasite communities than expected, suggesting that host 20 heterogeneity did not strongly impact parasite community composition and that dispersal 21 was high at the individual-host level. Despite the small population sizes of within-host 22 parasite communities, drift appeared to play a minimal role in structuring community 23 composition. 24
- Dispersal and ecological selection appear to jointly drive parasite community assembly,
   particularly at larger biological scales. The dispersal ability of aquatic parasites with
   complex life cycles differs strongly across scales, meaning that parasite communities may

- 28 predictably converge at small scales where dispersal is high, but may be more stochastic
- and unpredictable at larger scales. Insights into assembly mechanisms within multi-host,
- 30 multi-parasite systems provide opportunities for understanding how to mitigate the spread
- 31 of infectious diseases within human and wildlife hosts.
- 32 Key Words: community assembly,  $\beta$ -diversity, trematode, multi-scale, amphibian disease,
- 33 coinfection, disease ecology, infectious disease

#### 34 INTRODUCTION

35 Parasites comprise more than one-third of known species on Earth and can be an 36 important driving force in ecosystem dynamics, the maintenance of biodiversity, and 37 evolutionary change (Hatcher et al. 2006, Hudson et al. 2006, Dobson et al. 2008, Paterson et al. 38 2010). Although epidemiology has traditionally focused on one-host, one-parasite interactions, parasites typically interact with rich assemblages of hosts and other symbionts (Rigaud et al. 39 2010). Moreover, interactions between host and parasite communities can combine to shape 40 41 ecological processes across multiple biological scales (Telfer et al. 2010, Seabloom et al. 2015). For instance, co-infecting parasites can dramatically alter disease outcomes for individual hosts 42 43 (Griffiths et al. 2011, Johnson and Hoverman 2012) as well as transmission rates within 44 populations (Jolles et al. 2008, Susi et al. 2015). Scaling up even further, parasite diversity within ecosystems can functionally dictate food web linkages and patterns of energy flow (Lafferty et 45 al. 2006), underscoring the importance of characterizing parasite community composition at 46 multiple scales – from across individual hosts to entire regions (Pedersen and Fenton 2007). 47

A key challenge in the ongoing study of parasite community ecology is to understand the 48 49 ecological processes underlying community structure and how these processes vary with scale (Holmes 1987, Sousa 1990). Historically, many studies have employed comparative approaches 50 to quantify compositional differences in parasite communities (e.g. Sousa 1990, Vidal-Martínez 51 52 and Poulin 2003, Krasnov et al. 2011, Altman and Byers 2014). This work has accumulated valuable information as to how parasite communities are structured at different biological scales: 53 54 e.g. how parasite communities differ across host individuals (Guégan and Hugueny 1994), across 55 host populations (Holmes 1990), across host species (Locke et al. 2013, Fecchio et al. 2017), and 56 across environmental or spatial gradients (Poulin 2003, Thieltges et al. 2010). However, fewer

studies have integrated multiple scales into the same study (but see Krasnov et al. 2010, Dallas 57 and Presley 2014), for example, to compare whether host species identity or habitat represent a 58 stronger structuring force. Comparisons of community composition across scales can reveal 59 where and by what mechanisms species distributions are most strongly limited and identify 60 whether assembly is non-random (Crist et al. 2003a). Identifying these core structuring processes 61 62 and their scale-dependence in natural host-parasite systems offers essential opportunities to test hypotheses about the drivers of parasite distributions and better inform strategies for disease 63 mitigation (Rynkiewicz et al. 2015). For instance, if parasite communities are strongly structured 64 65 across environmental gradients, managing habitat features might be an important disease control strategy, whereas high among-individual or among-species variation might require a targeted 66 vaccination program (Paull et al. 2012). More broadly, due to their hierarchically nested structure 67 (Guégan et al. 2005), parasite communities offer potential for understanding how assembly 68 processes vary over scale (Mihaljevic 2012) and therefore represent a powerful study system for 69 70 addressing one of the major challenges in community ecology (Levin 1992, Chase and Myers 2011). 71

72 Community ecology theory advances that variation among ecological communities is driven by four key processes: dispersal, ecological selection, drift, and speciation (Vellend 73 2010). Dispersal is the process by which parasites colonize a site from a given species pool and 74 75 operates at multiple biological scales; for instance, dispersal structures which parasites arrive to a particular habitat patch from the regional species pool, as well as the parasites reaching a given 76 host within that habitat patch (Guégan et al. 2005). Ecological selection is a niche-based process, 77 78 whereby abiotic or biotic conditions filter which parasites can persist following dispersal. Selection acts at the habitat-level (if habitat features influence the species able to persist), at the 79

host species-level (if species vary in their susceptibility to different parasites), and at the
individual host-level (if hosts differ in susceptibility or if parasite interactions occur). Finally,
drift is the change in community composition resulting from stochastic fluctuations; this can
occur across all biological scales but is expected to occur most strongly for small communities
(Chase and Myers 2011), such as those at the within-host level.

To better understand at how the relative importance of these mechanisms varies with 85 86 scale, metrics of community dissimilarity ( $\beta$ -diversity) can be compared through approaches such as additive partitioning (Veech et al. 2002, Johnson et al. 2016). For instance, if  $\beta$ -diversity 87 is higher across host species than across habitats, ecological selection at the host-species level 88 could be more important than dispersal among habitats or selection due to environmental 89 90 variables. Moreover, because assembly processes are often temporally dynamic (Penczykowski et al. 2016, Fitzgerald et al. 2017), quantifying  $\beta$ -diversity across a temporal window can further 91 inform how composition shifts with host development or environmental change (Cohen et al. 92 2015), offering further insight into process. Decreasing  $\beta$ -diversity over time indicates 93 94 convergence among communities, due to high dispersal rates or similar selection regimes 95 (Leibold et al. 2004), whereas increasing  $\beta$ -diversity between similar sampling units often indicates an important role of drift (Chase 2007). Studies examining  $\beta$ -diversity over time and 96 scale have revealed that assembly processes and community compositions are temporally and 97 spatially dynamic (Soininen et al. 2007, McIntire and Fajardo 2009) and have reinforced the idea 98 that temporal studies yield a more process-based understanding of community assembly. For 99 example, by partitioning variance in community composition across scale and over time, 100 101 Costello and others (2009) showed that temporal variation in gut microbiomes within the same 102 individual was smaller than among-individual variation, even on the same day, suggesting that

different hosts selected particular microbial communities that remained distinct from other hosts'
microbiota over time. Temporal analyses of parasite community composition are still relatively
rare (but see Vidal-Martínez and Poulin 2003, Fallon et al. 2004, Cohen et al. 2015, Budischak et
al. 2016), and even fewer studies have longitudinally tracked parasite community composition
across multiple distinct scales to infer the general processes driving assembly.

Here, we compared variation in parasite communities across multiple, discrete biological 108 scales and over time to better characterize the processes driving community composition. We 109 110 quantified parasite communities (trematodes, nematodes, and protists) within two species of amphibian hosts (Pacific chorus frogs *Pseudacris regilla* and California newts *Taricha torosa*), 111 sampled across 10 different ponds within the California Bay Area. By re-visiting each pond 112 throughout host development, we generated a unique dataset on parasite accumulation and 113 primary succession. Using an additive partitioning approach (Figure 1), we tested the extent to 114 which parasite community composition was driven by differences among host individuals within 115 the same population ( $\beta_{\text{host}}$ ), among host species within the same community ( $\beta_{\text{species}}$ ), or among 116 117 host communities ( $\beta_{pond}$ ). We next examined how  $\beta$ -diversity at each of these scales varied across time to gain further insight into how structuring processes changed over the course of 118 primary succession. We also modelled temporal variation in richness ( $\alpha$ -diversity) at each 119 120 biological scale to better infer whether changes in community composition were operating through species losses (consistent with ecological selection or drift) or through species gain 121 (consistent with dispersal). We expected that, if ecological selection at the community or species 122 123 scale is a major structuring force, we would observe large differences among host species (B<sub>species</sub>) and host ponds (B<sub>ponds</sub>) relative to null models (Müller and Goßner 2010). If ecological 124 125 drift is an important structuring processes, among-host variation (Bhost) would be consistent with

or larger than null models, and would increase through time (Püttker et al. 2015). Finally, we expected that if dispersal was not limiting at a particular scale,  $\alpha$ -diversity would increase while  $\beta$ -diversity would decrease through time at that scale (Leibold et al. 2004). Overall, we intend that the application of a consistent framework and terminology can help to identify the drivers of parasite assembly across a range of multi-host, multi-parasite systems, thereby facilitating future comparisons focused on scale-dependent (or invariant) assembly processes related to disease.

### **132 MATERIALS AND METHODS**

#### 133 Field surveys and dissection

Over the summer of 2017 (May – August), we characterized parasite communities within 134 the larvae of two amphibian species (the Pacific chorus frog, P. regilla, and the California newt, 135 T. torosa). Hosts were sampled from 10 ponds in California Bay Area (Appendix S1: Figure S1). 136 All ponds were small habitats containing both species of focal amphibian host and the snail 137 138 species *Helisoma trivovlis* and *Physa* spp. (P. acuta or P. gyrina), which are intermediate hosts for a diversity of trematode species found in amphibians (McCaffrey and Johnson 2017). Each 139 pond was visited four to five times across the summer, during which we sampled 10 to 20 140 141 individuals of each host species, representing juvenile life stages from post-hatching to perimetamorphosis (Appendix S1: Figure S2). Previous research in the same system has shown that 142 143 this sample size is sufficient for estimating parasite richness within a site (Johnson et al. 2012; 2013). After euthanizing hosts with MS-222, we examined their organs and tissues under an 144 Olympus SZX10 dissection microscope following standard methods (Johnson et al. 2018) to 145 characterize the presence and identity of parasites (Schell 1985, Sleigh 1991, Gibson et al. 2002, 146 Duszynski et al. 2007, Anderson et al. 2009). For trematode and nematode infections, we 147

quantified both parasite identity and load (number of parasites per host), whereas for protozoans
we noted only presence or absence. We also measured host size (snout-vent length) and
developmental stage (Gosner 1960, Wong and Liversage 2004).

## 151 *Richness and composition across biological scale and time*

152 To examine how parasite richness varied with biological scale and sampling date, we quantified host-level richness ( $\alpha_{host}$ ) as the number of parasite taxa within an individual host, 153 population-level richness ( $\alpha_{population}$ ) as the number of parasite taxa across all individuals of a 154 given host species at a given pond, and community-level richness ( $\alpha_{pond}$ ) as the number of 155 156 parasite taxa across both host species at a given pond. Regional richness ( $\gamma$ ) was estimated as the 157 total number of parasite taxa across all ponds. We quantified  $\alpha_{host}$ ,  $\alpha_{population}$ ,  $\alpha_{pond}$  and  $\gamma$ separately for each time point and modelled richness as a function of time separately for each 158 159 biological scale, using generalized linear mixed models (GLMM). We included polynomial terms for sample date to test whether richness accumulation was linear or unimodal and selected 160 the model that minimized AIC (Appendix S1). We used indicator species analysis in the package 161 indicspecies for R (De Caceres and Legendre 2009), to determine whether certain parasite 162 species were associated with particular time points ("indicator species analysis"), including early 163 or late summer. The code for this analysis and all subsequent statistical analyses is included in 164 Appendix S2. 165

### 166 $\beta$ -diversity across biological scale and time

To quantify how differences among parasite communities (i.e., turnover or β-diversity)
changed with time, we estimated pairwise dissimilarity metrics using Jaccard's distance (Jaccard
169 1912), which is based on presence-absence data (Appendix S1). We estimated between-host

turnover ( $\overline{\beta}_{J, host}$ ) as the average pairwise dissimilarity between individuals of the same species collected in the same pond on the same visit. Between-species turnover ( $\overline{\beta}_{J, species}$ ) was the average difference between *P. regilla* and *T. torosa* parasite communities at a given site-visit, and between-community turnover ( $\overline{\beta}_{J, pond}$ ) was the average difference in parasite communities among ponds. We calculated all three metrics separately at each time point. Using a separate linear mixed model for each biological scale, we quantified the relationship between Jaccard's βdiversity and sample date (Appendix S1).

Because Jaccard's  $\beta$ -diversity metric is sensitive to total taxonomic richness and thus should not be used to compare turnover across scales, we used an additive partitioning approach to compare  $\beta$ -diversity among hosts, species, and communities (Lande 1996, Crist et al. 2003b, Gering et al. 2003, Veech and Crist 2010, Johnson et al. 2016). This approach partitions regional parasite diversity ( $\gamma$ ) into scale-specific values of turnover ( $\beta$  diversity) and richness ( $\alpha$  diversity) using the formula:

 $\gamma = \alpha host$  (within host) +  $\beta host$  (between hosts) +  $\beta species$  (between species) +  $\beta pond$  (between 183 communities (Figure 1). We used the function *adipart* in the R package vegan (Oksanen et al. 184 2018), which implements random permutations of species occurrence data to produce estimates 185 186 of significance relative to a null model. The null distribution was generated using 1000 random permutations which hold site-level richness constant and randomly distribute parasite species 187 across sites relative to their abundance (Crist et al. 2003). We present the difference between 188 observed values ( $\bar{\alpha}_{host, obs}$  and  $\bar{\beta}_{host, obs}$ ) and mean null simulation values ( $\bar{\alpha}_{host, sim}$  and  $\bar{\beta}_{host, sim}$ ) to 189 evaluate whether observed diversity differed from random chance and then assessed how the 190

proportional contribution of each component to total γ diversity varied with time as hostsprogressively developed and infections accumulated.

We evaluated the sensitivity of the above analyses to the inclusion of rare species by rerunning the analyses without rare species, as well as using the Bray-Curtis abundance-based
dissimilarity metric (Appendix S1).

196 **RESULTS** 

197 Field surveys and parasite community

We quantified the richness and community composition of parasites from 959 individual 198 199 hosts (445 T. torosa, 514 P. regilla) from 10 ponds across three months of sampling. The majority of hosts (n = 795; 82%) were infected by at least one parasite. On average, ponds 200 contained 6.2 parasite taxa per visit. We identified 18 distinct taxa of parasites, 11 of which were 201 202 present in >1% of hosts (six digenetic trematodes, two nematodes, and three protists; Appendix S1: Table S1). The most common parasite was the trematode *Ribeiroia ondatrae* (present in 44% 203 204 of *P. regilla* and 48% of *T. torosa*), followed by the trematode *Echinostoma* spp. (59% of *P. regilla* and 16% of *T. torosa*). Some parasites were specialists (e.g., the trophically-transmitted 205 nematode Chabaudgolvania sp. was only found in T. torosa, whereas two protists [Opalina 206 207 ranarum and Nyctotherus cordiformus] and the nematode Gyrinicola batrachiensis were only found in P. regilla). Other parasites (e.g. the trematodes R. ondatrae and Manodistomum 208 syntomentera) infected both host species at roughly equal proportions. 209

## 210 Richness and composition across biological scale and time

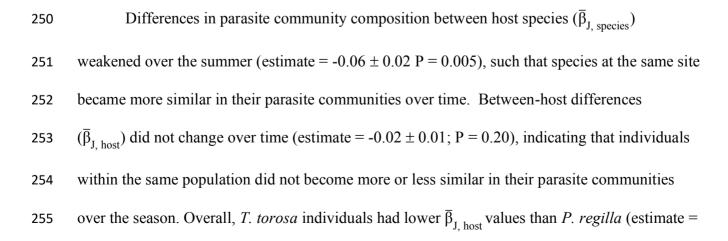
Richness showed a non-linear relationship with time at both the host- and populationlevels (Appendix S1: Figure S3). The top model for predicting host-level richness (αhost)

213	included significant terms for host species, an interaction between visit and species, and a					
214	quadratic term for visit. This model showed improved fit over the next best model, which used					
215	visit as a linear term ( $\Delta$ AIC = -10.47). Therefore, hosts accumulated parasites non-linearly with					
216	respect to time, with richness peaking in late June to mid-July. The median richness within P.					
217	<i>regilla</i> hosts was three parasites, which was significantly higher than <i>T. torosa</i> (median = 1;					
218	estimate = -1.65 $\pm$ 0.15; P < 0.0001; Appendix S1: Figure S3). The best performing model for					
219	predicting population-level richness ( $\alpha_{population}$ ) included significant terms for host species, an					
220	interaction between visit and species, and a second-order polynomial term for visit. Similarly,					
221	within-population parasite richness was higher in <i>P. regilla</i> populations (median = 5) than in <i>T.</i>					
222	<i>torosa</i> populations (median = 2, estimate = $-1.42 \pm 0.34$ ; P < 0.0001). While community-level					
223	richness ( $\alpha_{pond}$ ) exhibited a similarly unimodal relationship, the best model was a null model,					
224	indicating that visit was a poor predictor of parasite richness within ponds. Regional richness ( $\gamma$ ;					
225	aggregated over all ponds) did not change over time (estimate [visit] = $0.009 \pm 0.09$ ; P = $0.92$ )					
226	Of 11 parasite taxa, four were strongly associated with a particular time point or group of					
227	time points based on indicator species analysis (Appendix S1: Figure S4). The protozoan					
228	<i>Tritrichomonas augustus</i> was associated with the first and second visits ( $P = 0.003$ ), and the					
229	nematode <i>Gyrinicola batrachiensis</i> was associated with the first through third visits ( $P = 0.001$ ).					
230	Two trematode parasites, Echinostoma spp. and Gorgoderina sp., were "late season" parasites,					
231	associated with the third through fifth visits ( $P = 0.001$ and $P = 0.02$ , respectively).					

# $\beta$ -diversity across biological scale and time

Additive partitioning of diversity (Figure 1) revealed that differences among communities
(βponds) had the strongest influence on regional parasite diversity (γ), and were considerably

greater than the effects of differences between host species ( $\beta_{species}$ ) or among individual hosts 235 ( $\beta$ host; Figure 2). Among-community turnover averaged over all visits ( $\overline{\beta}_{pond, obs}$ ) accounted for 236 40% of regional parasite diversity, and ponds were significantly less similar than expected from 237 a null model ( $\overline{\beta}_{pond, obs} - \overline{\beta}_{pond, sim} = 0.16$ ; P < 0.001). When rare species were removed, the same 238 overall pattern remained, but  $\bar{\beta}_{pond, obs}$  declined to 0.29. Within-host parasite richness ( $\bar{\alpha}_{host}$ ) was 239 the second largest contributor to regional diversity, accounting for 25% of regional parasite 240 diversity, which was slightly higher than expected ( $\bar{\alpha}_{host, obs} - \bar{\alpha}_{host, sim} = 0.040$ ; P< 0.001). 241 Differences among host species ( $\overline{\beta}_{species}$ ) accounted for 23% of regional parasite diversity and 242 were only slightly less similar ( $\overline{\beta}_{\text{species, obs}} - \overline{\beta}_{\text{species, sim}} = 0.08$ ; P<0.001) than predicted by null 243 models. Finally, differences among host individuals within the same population ( $\overline{\beta}_{host}$ ) accounted 244 for just 12% of regional diversity, and individuals were much more similar in their parasite 245 communities than predicted by null models ( $\bar{\beta}_{host obs} - \bar{\beta}_{host sim} = -0.29$ ; P=0.001). Therefore, 246 turnover between communities (ponds) accounted for the highest percentage of regional 247 diversity, followed by within-host richness, between-species at the same site, and between-hosts 248 of the same population (Figure 2). 249



-0.16 ± 0.03; P < 0.0001); therefore, *T. torosa* individuals shared more parasites with other members of their population than did *P. regilla*. Differences among communities ( $\overline{\beta}_{J, \text{ pond}}$ ) did not change appreciably over time (estimate= -0.01 ± 0.02; P = 0.64). These patterns were robust to the dissimilarity metric used, as well as the inclusion of rare species (Appendix S1).

260 **DISCUSSION** 

Despite increasing interest in describing how parasite community composition changes 261 262 across habitats, host species, and individuals (Pedersen and Fenton 2007, Johnson et al. 2015, Seabloom et al. 2015), comparatively few studies have investigated the drivers of compositional 263 differences (i.e. "parasite community assembly") especially across both biological scale and 264 265 time. In this study, we compared patterns of parasite composition across nested biological scales and used repeated sampling over a temporal window of host development to derive insight into 266 the potential roles of drift, dispersal, and ecological selection. This integration of scales indicated 267 that parasite communities were more different across habitat patches than across host species, 268 and that individuals within the same population contained highly similar parasite communities. 269 Differences among communities (i.e., ponds) were consistent with dispersal limitation, whereas 270 within communities, high rates of dispersal appeared to erode individual and species-level 271 differences. Our study illustrates that, as a whole, dispersal processes can play a strong role in 272 273 structuring parasite communities, but that rates of dispersal strongly differed across scales.

Based on an additive partitioning of regional diversity, hierarchical differences in parasite community between communities ( $\beta_{ponds}$ ) comprised the majority (40%) of regional level diversity, more than differences between host species ( $\beta_{species}$ ; 23%) or individuals ( $\beta_{host}$ ; 12%). Community-level differences ( $\beta_{ponds}$ ) were significantly greater at each time point than expected

by chance (Figure 2), suggesting that parasites were distributed non-randomly across ponds. 278 These differences were, in part, driven by the occurrence of rare parasites; when they were 279 280 removed, the contribution of  $\beta_{\text{ponds}}$  was lower (28%). Differences among ponds could be the result of ecological selection if sites filter out certain parasite taxa based on abiotic or biotic 281 282 conditions. For example, the only two ponds that lacked a common trematode parasite 283 (Cephalogonimus americanus) were also the only ponds with American bullfrog (Rana *catesbeiana*) larvae, which are a less competent host that could reduce C. *americanus* abundance 284 (Johnson et al. 2013). Consequently, the host community could represent one possible selection 285 286 mechanism driving parasite community composition, as previously reported in similar studies (Krasnov et al. 2005, Johnson et al. 2013, Maestri et al. 2017, Mihaljevic et al. 2017). More 287 broadly, our results are consistent with a large body of work indicating that ecological selection 288 can strongly structure parasite communities across localities (e.g. Dallas and Presley 2014, 289 Krasnov et al. 2015, Warburton et al. 2016). To further elucidate the role of ecological selection, 290 291 interannual variation could be assessed; differences among ponds that are stable across years and correlate strongly to environmental factors could point to important selection mechanisms 292 shaping parasite community composition. 293

Differences among ponds could also be generated by dispersal limitation; for instance, ponds lacking certain parasites may have not been colonized, even if suitable conditions for that parasite's survival exist. Differences among communities were consistent over time, using both Jaccard's distance and additive partitioning approaches (Figure 2), and community-level richness remained stable over time (Figure S3), suggesting that dispersal did not occur during the sampling window at levels high enough to attenuate differences in pond parasite communities. As such, lack of dispersal during the sampling season created stable differences among sites.

Parasites are spread from site-to-site through the movement of hosts (Price 1980). Despite the 301 large dispersal potential of parasites within vagile hosts, host activity may be seasonal or 302 303 infrequent, limiting colonization opportunities. Indeed, variation in definitive host activity is known to limit dispersal (Kuris and Lafferty 1994), and parasite communities are often strongly 304 structured by this dispersal limitation at landscape scales (Smith 2001, Thieltges et al. 2009; 305 306 Hartson et al. 2011). Moreover, habitat-level factors can strongly influence the activity of hosts and therefore the probability of dispersal (Sousa and Grosholz 1991, Hartson et al. 2011) making 307 parasite dispersal across sites a non-neutral process. Consequently, relationships between 308 309 geographic distance and community dissimilarity, which are commonly used to infer dispersal limitation in free-living communities (Cottenie 2005, Soininen et al. 2007), may not be expected 310 for parasite communities structured by dispersal (Poulin 2003, Thieltges et al. 2010). On the 311 other hand, a relationship between environmental attributes and community composition, 312 commonly interpreted as evidence of *selection* in free-living community ecology (Smith et al. 313 314 2015, Vellend 2016), may actually be evidence of a system driven by non-neutral dispersal limitation. Therefore tracking parasite dispersal more directly, for instance by monitoring or 315 manipulating host movement (Wood et al. 2019), could be an important tool for further 316 317 evaluating the role of dispersal limitation in driving among-site variation in parasite communities. 318

In addition to high variation across sites, host species also demonstrated significant and
 non-random differences in parasite community composition. Differences between the parasite
 communities of *P. regilla* and *T. torosa* (β<sub>species</sub>) were greater than expected by chance (Figure 2).
 Therefore, ecological selection appeared to play a role in structuring the parasite communities
 among different host species, consistent with other systems (e.g. Friggens and Brown 2005,

MacColl 2009, Dallas and Presley 2014). Indeed, almost half (5/11) of the parasite taxa in this 324 study were strict specialists on one host (Appendix S1: Table S1) and previous studies (Johnson 325 326 et al. 2016) have demonstrated that host diversity is an important driver of regional parasite diversity in this same system. Intriguingly, differences among host species ( $\beta_{J, \text{ species}}$ ) decreased 327 over time, revealing that *P. regilla* and *T. torosa* within the same site became more similar in 328 329 their parasite communities. This pattern appeared driven by a combination of decreasing selection and increasing dispersal pressure over time. Specialist parasites (protozoans and 330 nematodes) tended to peak earlier in the season (Appendix S1: Figure S4); this observation was 331 332 supported by indicator species analysis showing that both early season parasites were specialists with direct lifecycles. On the other hand, generalist parasites in this study were all digenetic 333 trematodes requiring a snail intermediate host, meaning that infections occurred later in 334 development, after snails began shedding infectious trematode stages around June (Paull and 335 Johnson 2014). Host species became more similar due to the accumulation of infections by 336 337 generalist parasites, which reached high prevalences later in summer. Therefore, high dispersal of generalist parasites homogenized host species over time, and parasite specialization, a form of 338 selection, attenuated through time. This differs from free-living communities, wherein specialist 339 340 taxa often appear to colonize later in succession (Piechnik et al. 2008, Raevel et al. 2012, Helsen et al. 2016). 341

We found little evidence of heterogeneity among individual hosts; hosts within the same
population shared significantly more parasites than expected by chance (Figure 2). Low βdiversity at this scale can result from high dispersal rates ("propagule pressure"), which act to
homogenize communities (Leibold et al. 2004, Qian 2009), and it is well-established that
dispersal to individual hosts can be high in pond ecosystems given the high biomass of infectious

forms produced by trematodes (Preston et al. 2013; Lambden and Johnson 2013). Many parasites 347 reached high prevalences within host populations, including numerous taxa that reached 100% 348 349 prevalence (Appendix S1: Figure S4), indicating a lack of dispersal limitation to individual hosts. Our evidence differs from previous studies documenting high heterogeneities among hosts (Paull 350 et al. 2012), driven by selection (e.g. host traits influencing susceptibility or exposure risk; 351 352 Johnson et al. 2005), dispersal (e.g. spatial variation in exposure risk; Calabrese et al. 2011) or interactions between dispersal and drift (e.g. priority effects; Budischak et al. 2016). Overall, 353 parasite interactions and variation in host competency or exposure risk did not appear to be 354 355 important processes in this system. We note, however, that by sampling similarly-aged, nonreproductive hosts within similar habitats, our approach effectively limited several of the sources 356 of intra-host variability in parasite communities. In natural populations containing a mixture of 357 different larval cohorts, differences among individuals would likely be larger, with younger 358 individuals supporting more protozoan infections, and older individuals supporting more 359 360 trematode infections. Hosts at intermediate stages of development supported the most diverse parasite communities, as evidenced by the unimodal relationship between richness and time 361 (Figure S3). Finally, the low divergence among host individuals indicated that drift was not a 362 363 strong structuring force, since drift is expected to increase divergence over time (Gilbert and Levine 2017). Drift can represent a powerful stochastic force driving community dissimilarity 364 (Hubbell 2001) and is expected to operate more strongly when community sizes are small, such 365 as the within-host scale (Vellend 2010). The most prevalent parasites in this system, digenetic 366 trematodes, do not reproduce within our focal amphibian hosts, requiring a definitive host to 367 complete reproduction; thus demographic fluctuations and thus the influence of drift were likely 368 dampened at the within-host scale. Therefore, unlike free-living species, the effect of drift may 369

be more important at larger scales which encompass the entire parasite life cycle. The role of
drift has not received as much attention in parasite community ecology (Seabloom et al. 2015)
but its scale dependency may represent an important future avenue of research.

Empirical data from hierarchically nested parasite communities provide opportunities to 373 374 understand how ecological processes vary over biological scale and enable syntheses across community ecology. However, key differences among these study systems present challenges for 375 integrating free-living and parasite community ecology. For example, within parasite community 376 377 ecology, the use of multiple, alternative sets of terminologies to describe assembly have led to both confusion and a disconnect from community ecology generally. For instance, parasitologists 378 may refer to parasites as being limited by "encounter filters" or "compatibility filters" (Combes 379 380 2001) or evaluate the role of "host-level" processes such as host age, sex, or body size in driving community composition (Blaylock et al. 2011). Yet mechanistically such traits can affect 381 community composition through dispersal (e.g., if larger hosts experience greater colonization 382 Kuris et al. 1980), selection (e.g., if body size correlates with immune function; Venesky et al. 383 2012), or drift (e.g., if larger hosts contain larger parasite communities more resistant to 384 stochastic extinctions; Guégan and Hugueny 1994). Additionally, the same process may be 385 referred to by multiple names depending on the scale of study – for instance, dispersal may be 386 called "transmission" when parasites disperse to hosts, but may be referred to as "colonization" 387 388 when that host or population is previously uninfected (Bush et al. 1997). The concept of "transmission" further tends to combine both selection and dispersal-based processes (McCallum 389 et al. 2017), making it challenging to compare with assembly processes in free-living ecology. 390 391 As empirical data on parasite communities accumulates, using a common language about the

underlying processes will further facilitate comparisons across studies and biological scales,improving links with free-living community ecology.

394 Studies of parasite community ecology have generated numerous fundamental insights 395 into how parasite community composition alters disease dynamics (Johnson et al. 2015), but 396 understanding how those communities arise is still difficult with observational studies in natural populations. Owing to the inherently nested nature of parasite communities, implementing cross-397 scale comparisons can more accurately capture the drivers of parasite community composition 398 399 (Esch 2002, Penczykowski et al. 2016). Using additive partitioning is one way to make direct, cross-scale comparisons in nested communities, and we have used this approach herein to 400 401 compare processes acting at the within-host, within-population, and within-pond level. We 402 additionally demonstrate that comparing communities over a temporal window, especially over primary succession, can aid in disentangling process and pattern. Community assembly 403 mechanisms influence important facets of infectious disease-causing agents; where they are, 404 when they arrive, and at what scales they are most limited. A process-based approach therefore 405 improves our ability to predict where parasites will spread and at what scales they may be 406 407 appropriately managed.

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# 418 AUTHOR CONTRIBUTIONS

- 419 WEM and PTJ designed the study; WEM, TMG, and DMC collected the data; WEM
- 420 analysed the data; WEM led the manuscript writing. All authors contributed to editing and
- 421 reviewing the manuscript and gave final approval for publication.

# 422 DATA ACCESSIBILITY

423 Upon acceptance, individual dissection data and R code necessary to reproduce analyses424 will be archived in the Dryad Data Repository.

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#### **FIGURE LEGENDS**

**Figure 1**. Contribution of different biological scales to regional parasite diversity. At the largest scale, regional richness ( $\gamma$ ) is the total number of parasite species within all ponds at any given point in time. Regional richness can be partitioned additively into within-host richness ( $\alpha$ -diversity) and turnover at each biological scale ( $\beta$ -diversity) using the equation:

 $\gamma = \alpha_{host}$  (within host) +  $\beta_{host}$  (between hosts) +  $\beta_{species}$  (between species) +

 $\beta_{pond}$  (between pond communities). We compared the relative contributions of each component across time.

**Figure 2.** Additive partitioning of regional parasite richness ( $\gamma$ ). Each panel shows the observed contribution of each level to overall regional parasite richness (solid line), compared to null simulations where parasites are distributed randomly (illustrated by a shaded ribbon showing the 95 % CI of simulations). Differences among communities or ponds ( $\beta_{pond}$ ) represented the largest contribution to regional parasite richness and variation among individual hosts in the same population ( $\beta_{host}$ ) represented the smallest contribution to regional parasite richness.

