

Contaminants in Aquatic and Terrestrial Environments

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Photochemical Transformations of Carbon Dots in Aqueous Environments

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Abstract

The unique physicochemical and luminescent properties of carbon dots (CDs) have motivated research efforts towards their incorporation into commercial products. Increased use of CDs will inevitably lead to their release into the environment where their fate and persistence will be influenced by photochemical transformations, the nature of which is poorly understood. This knowledge gap motivated the present investigation of the effects of direct and indirect photolysis on citric and malic acid-based CDs. Our results indicate that natural sunlight will rapidly and non-destructively photobleach CDs into optically inactive carbon nanoparticles. We demonstrate that after photobleaching, $\cdot\text{OH}$ exposure degrades CDs in a two-step process that will span several decades in natural waters. The first step, occurring over several years of $\cdot\text{OH}$ exposure, involves depolymerization of the CD structure, characterized by volatilization of over 60% of nascent carbon atoms and the oxidation of nitrogen atoms into nitro groups. This is followed by a slower oxidation of residual carbon atoms first into carboxylic acids and then volatile carbon species, while nitrogen atoms are oxidized into nitrate ions. Considered alongside related CD studies, our findings suggest that the environmental behavior of CDs will be strongly influenced by the molecular precursors used in their synthesis.

Introduction

Carbon dots (CDs) are an emerging class of colloiddally stable carbon-based nanomaterials with high fluorescence quantum yields and tunable emission properties.¹ As an environmentally benign alternative to inorganic quantum dots that contain toxic metals, CDs have attracted great interest for their potential in sensing,²⁻³ fuel cells,⁴⁻⁵ drug delivery,⁶⁻⁷ and bioimaging.⁸⁻⁹ As a result, the quantity of CDs entering aquatic environments will inevitably increase during the production, use, and disposal of CD-enabled products. While CDs are of similar size to other carbon-based nanomaterials such as fullerenes, they are likely to exhibit different environmental behavior due to the high surface charge imparted by the functional groups (e.g., carboxylic acid, amine, and amide groups) embedded within their structure.¹⁰

To date, research on the environmental behavior of CDs has largely focused on their colloidal stability,¹¹⁻¹⁴ transport,¹⁵ and toxicity¹⁶⁻¹⁷ in environmental media. Bayati et al. investigated the effect of ionic strength, pH, and natural organic matter (NOM) on the aggregation of glycerol-based CDs and aminated CDs, finding both species largely stable at the upper bound of environmental ionic strength (i.e., 10 mmol/L NaCl and CaCl₂) and across a pH range of 3-11.¹² Li et al. showed that graphene CDs were colloiddally stable in solutions of NaCl, KCl, and MgCl₂ with an ionic strength of >1000 mM, but were prone to aggregation in solutions containing 1 mM CaCl₂.¹³ Liu et al. determined that hydrothermally synthesized sodium citrate-based CDs were stable to homoaggregation in environmentally relevant pH and ionic strength conditions, but this stability could be perturbed by low pH (pH < 3) and high ionic strength conditions (> 30 mM NaCl).¹¹ The high degree of colloidal stability exhibited by CDs translates into high transport capacity, as exhibited by Kamrani et al., in which citric acid-based CDs readily eluted through quartz media at environmentally relevant conditions and were only partially retained in the column

even at extremely high ionic strength and low pH.¹⁵ As a result of their colloidal stability and transport properties, CDs are likely to be present in the water column for significant periods of time. Consequently, their interactions with sunlight are likely to play an important role in determining their fate and persistence in aquatic environments.

The primary photolytic processes acting on CDs in the environment will be direct irradiation by natural sunlight and indirect photolysis via exposure to reactive oxygen species (ROS) of which hydroxyl radicals ($\cdot\text{OH}$) produced by the interaction of sunlight with naturally occurring species (e.g., natural organic matter (NOM) or nitrate/nitrite)¹⁸⁻²⁰ are typically the most reactive, and as such are often used experimentally to simulate the indirect photolysis of environmental contaminants.²¹⁻²⁵ While the effects of direct and indirect photolysis on other carbon nanomaterials (e.g., CNTs,²⁶⁻²⁷ GO,^{24, 28} fullerenes²⁹⁻³⁰) have been the subject of significant research, the unique chemical and fluorescent properties of CDs are likely to impart different photochemical behavior. A recent study by Chen et al. identified that CDs synthesized via a hydrothermal route using citric acid and urea rapidly degraded into low molecular weight compounds and could be mineralized to carbon dioxide under both UVB irradiation and natural sunlight exposure.³¹ These transformations occurred over the course of 2 h UVB or 8 h natural sunlight exposure and were attributed to processes involving ROS formed by the irradiation of the CDs themselves, with $\cdot\text{OH}$ being the dominant radical species. The rapid photodegradation and mineralization of these CDs suggest they will not persist once they are released into the environment. However, these results cannot necessarily be generalized to all CDs, as an enormous array of precursors and methods (e.g. hydrothermal, isolation from organic waste, thermal decomposition, microwave)²⁻³ are currently being used in their synthesis, the effect of which on the chemical, physical, and photochemical properties of CDs remains unknown.

CDs produced from small organic acid molecules, via microwave irradiation, are some of the most widely studied for consumer and research applications due to the low cost and ease of “bottom-up” synthesis.³² Therefore it is important to develop a mechanistic understanding of their behavior in aquatic environments. In this study, the effects of direct and indirect photolysis on microwave synthesized citric acid-based CDs (CACDs) and malic acid-based CDs (MACDs) were investigated. CDs were exposed to natural sunlight for up to 6 weeks, while the effects of indirect photolysis were simulated by exposing CD solutions to $\cdot\text{OH}$ radicals generated by the photolysis of H_2O_2 under UV light. Total carbon analysis (TC), total nitrogen analysis (TN), and ion chromatography (IC) were used to identify the changes in carbon content, nitrogen content, and nitrogen speciation over time. Changes to optical properties were examined using photoluminescence (PL) spectroscopy. The physicochemical transformations of CDs were characterized by a combination of attenuated total reflectance Fourier transform infrared spectroscopy (ATR-FTIR), ^1H - and ^{13}C -nuclear magnetic resonance spectroscopy (NMR), and X-ray photoelectron spectroscopy (XPS). Using this suite of analytical techniques, we have identified both the photochemical transformations and the kinetics of such transformations that these CDs will undergo upon release into natural waters.

Materials and Methods

Additional details of synthetic procedures, characterization methods, photochemical exposures, and $\cdot\text{OH}$ radical quantification are included in the SI. For all suspensions prepared, the initial concentration of CDs was first determined via the recorded mass of dry CD powder added to a known volume of water.

Materials

All chemicals were purchased from Sigma-Aldrich and used without further purification. Milli-Q[®] water (18.2 Ω , Millipore, USA) was used to prepare all aqueous solutions.

Synthesis of CDs

CDs were prepared using a bottom-up microwave-assisted pyrolysis method as described by Zhi et al.³³ Two types of CDs were synthesized, using ethylene diamine (EDA) and either citric or malic acid as precursors to form CACDs and MACDs, respectively.

Characterization of CDs

CDs were characterized initially and after exposure to the effects of direct and indirect photolysis using a combination of TC, TN, IC, ATR-FTIR, XPS, ¹H- and ¹³C-NMR, and PL. Transmission electron microscopy (TEM) and UV-Vis spectroscopy were used to further characterize the as-synthesized CDs.

Exposure to Natural Sunlight

Solar irradiation experiments were conducted by adding suspensions of CDs in Milli-Q water to borosilicate glass test-tubes (16 mm outer diameter x 125 mm, Corning, USA). Sample test-tubes were sealed with PTFE-lined caps, and placed outside on a rooftop in Baltimore, MD (39° 19' W, 76° 37' N). Equivalent dark controls were prepared by wrapping glass test-tubes in aluminum foil, and CD-free controls were prepared identically without CDs. At time points ranging from 0 to 6 weeks, samples and controls were retrieved sacrificially for analysis. Unless otherwise specified, natural sunlight exposures were performed under ambient conditions with no sparging of the samples. Therefore, dissolved oxygen levels should be representative of those encountered in natural waters.

Exposure to $\cdot\text{OH}$ Radicals

Suspensions of MACDs and CACDs were prepared and added to quartz test tubes, sealed with an aluminum foil cap, and vented via a syringe tip. Samples were exposed to $\cdot\text{OH}$ radicals generated by the photolysis of H_2O_2 in a photochemical reactor equipped with 16 low pressure mercury lamps emitting 300 nm light (RPR 100, Southern New England Ultraviolet Company, Branford, CT, approx. 1.5×10^{17} photons/s). $\cdot\text{OH}$ exposure studies performed for TC, TN, ATR-FTIR, and XPS were all carried out at concentrations at or below 350 mg/L CD. ^{13}C -NMR experiments required a higher CD concentration (1.7×10^4 mg/L) to achieve reasonable signal-to-noise ratio.

Determination of $\cdot\text{OH}$ Radical Dose

The total dose of $\cdot\text{OH}$ generated by the photodegradation of H_2O_2 as well as the equivalent environmental exposure time in natural surface waters (Table S1) were determined using methods described by Lankone et al.³⁴ Briefly, the steady-state $\cdot\text{OH}$ concentration in solution generated during the photolysis of H_2O_2 was determined via monitoring the rate of salicylic acid (i.e., a probe molecule) decomposition from an initial concentration of 0.07 mM. With this methodology, the steady-state concentration of $\cdot\text{OH}$ with CDs present in solution could be determined with a standard error of less than 10%. Duration of $\cdot\text{OH}$ exposure was also recorded, enabling a molar*time $\cdot\text{OH}$ dose to be determined. This dose ($\text{M} \cdot \text{min}$) was then related to an equivalent duration of exposure to $\cdot\text{OH}$ in the natural environment, wherein the steady-state concentration ranges between 10^{-15} - 10^{-17} M.^{19, 24, 35} For example, a $\cdot\text{OH}$ concentration of 10^{-15} M, results in a monthly $\cdot\text{OH}$ dose of $10^{-15} \text{ M} \cdot 2.2 \times 10^4 \text{ min/month} = 2.2 \times 10^{-11} \frac{\text{M} \cdot \text{min}}{\text{month}}$. Importantly, control studies found that CD concentrations used in this study (≤ 319 mg/L) had no effect on the steady-state $\cdot\text{OH}$ concentration (Figure S1).

CD Settling Tests

Suspensions of CACDs (15 mg/L) were allowed to settle for 1 week in either MilliQ water or a mixture of NaCl, KCl, and CaCl₂ (4 mM Ca²⁺, 13 mM Cl⁻, 0.3 mM Na⁺, 5 mM K⁺) at pH 7 or 8. Supernatant was analyzed for suspended CDs using PL.

Results and Discussion

Initial CD Characterization

The as-synthesized CACDs were characterized by an emission peak at 480 nm and an absorbance below 400 nm, with a peak centered at 350 nm (Figure S2a). As-synthesized MACDs emitted most strongly at 470 nm with a broad absorbance below 400 nm and a weak peak around 350 nm (Figure S2b). TEM images (Figure S2c and S2d) of the as-synthesized CACDs and MACDs reveal spherical nanoparticles with sub-10 nm diameters. ATR-FTIR of CACDs and MACDs (Figure S3a and S3b) both exhibited broad IR features at 3280 cm⁻¹, 3085 cm⁻¹, and 2927 cm⁻¹ corresponding to N-H, O-H, and C-H stretching modes, respectively.³⁶⁻³⁷ For CACDs, features at 1700 cm⁻¹ (carboxyl) and 1646 cm⁻¹ (amide) are attributed to carbonyl (C=O) stretches,³⁸⁻³⁹ while the feature at 1547 cm⁻¹ is indicative of an N-H bend in an amide group (O=C-NH);^{37, 40} MACDs share the same carbonyl bands, however the N-H bend occurs at 1527 cm⁻¹.^{37, 40-41} The C (1s) envelope of the native CDs, as measured by XPS, contained contributions from C-C and C-H species centered at 285 eV, with a higher binding energy shoulder at ~ 288 eV due to the presence of more oxidized carbon atoms (e.g. amide and carboxyl groups; Figure S4 and S5). It should be noted that due to their small size (i.e., diameters < 10 nm), XPS spectra reflect the entire CD nanoparticle with respect to composition and chemical bonding environment.

Effects of Direct Photolysis

Upon entering aquatic environments, CDs will immediately experience the effects of direct photolysis due to their absorbance within the solar power distribution reaching the earth's surface (Figure S6a). Photoluminescence (PL) spectroscopy was used to monitor the fluorescent properties of CDs after exposure to sunlight and artificial indoor light, while changes in composition and chemical bonding were evaluated using a combination of TC/TN, XPS, ATR-FTIR, and ^1H -NMR.

CACDs exposed to natural sunlight experienced rapid photobleaching, as their fluorescence disappeared after 12 h of irradiation, observed both spectroscopically (Figure 1a, S7) and visually (Figure S8). This low photostability is in stark contrast to the persistence of fluorescence (Figure S9a) observed under artificial laboratory lighting. This difference in photostability is attributed to the differences in overlap between the absorbance spectrum of the CDs and the respective emission spectra from natural sunlight (solar irradiance, Figure S6a) and fluorescent bulbs (Figure S9b). Thus, it is likely that the < 400 nm component of natural sunlight drives photobleaching of CDs in the environment. The loss of fluorescence induced by exposure to sunlight was also found to be more rapid in solutions initially saturated with O_2 , but unaffected in solutions sparged with N_2 (Figure 1b). This behavior indicates that photobleaching in natural sunlight likely involves reactions with ROS formed via the presence of dissolved oxygen in solution.

Despite the rapid loss of fluorescence in CD solutions (i.e., < 12 h), exposure of up to 6 weeks of sunlight had no observable effect on the carbon or nitrogen content as exhibited by TC and TN (Figure 1c). Similarly, XPS data revealed that the composition within the CACDs (Figure 2a) and MACDs (Figure S10) as well as the chemical bonding environment within both CACDs (Figures 2b, S4) and MACDs (Figures S5) also remained constant over the course of the 6-week

exposure. ATR-FTIR spectra of CACDs and MACDs (Figure 2c) were largely unchanged after 6 weeks of exposure to sunlight, though for CACDs, loss of intensity at 1547 cm^{-1} was observed during the first two weeks of outdoor exposure, suggesting photobleaching could be driven through proton abstraction from the amide groups initially present in the CD structure, though not extensively enough to meaningfully change the CD composition.

^1H -NMR spectra of CACDs acquired before and after photobleaching also indicated only minor chemical changes to proton bonding environments (Figure 2d). As-synthesized CACDs featured clusters of ^1H -NMR peaks in several diagnostic regions: 1.8 ppm to 2.5 ppm, 2.5 ppm to 4.5 ppm, and at 8.4 ppm, indicative of protons in the α - or β -position to carbonyl or amide groups,⁴²⁻⁴⁴ protons bound to or in the α -position to oxidized carbon species (i.e., alcohol, ether, ester, carbonyl),^{42, 45} and non-exchangeable amide protons⁴⁶⁻⁴⁸ respectively. In contrast, no carboxylic acid protons were observed due to exchange of these acidic protons in the (D_2O) solvent. The photobleached CACDs exhibited a largely similar ^1H -NMR spectrum to the parent CACDs. The only changes observed were the loss of peak intensity at 4.2 ppm and the increase in intensity of the peak at 8.4 ppm, likely indicating some degradation of the amide groups initially present in the CDs.

Collectively, these findings indicate that the principle effect of natural sunlight on organic acid-based CDs immediately after entering an aquatic environment is rapid (i.e., $< 12\text{ h}$) photobleaching (Figure 1a) in a process modulated by the relative level of dissolved oxygen therein (Figure 1b). While the initial rate of photobleaching may vary to some degree dependent on the concentration of dissolved oxygen available to generate ROS, the complete photobleaching of CDs is expected to occur in all aqueous environments, irrespective of dissolved oxygen content (Figure 1b). Findings also indicate that the CD structure and composition will remain largely unchanged

during photobleaching, supported by mass-recovery studies which demonstrated that 101.4 % \pm 2.1 % of CD mass is recovered after 21 d of natural sunlight exposure (see SI for details).

Interestingly, these findings contrast with a study published by Chen et al., in which it was reported that citric acid/urea CDs were rapidly volatilized in totality after 8 h of natural sunlight exposure due to extensive structural decomposition driven by CD-produced $\cdot\text{OH}$.³¹ We posit one possibility for the contrasting photochemical behavior observed between these studies is that the different structural features present in the two different types of CDs, arising by virtue of the different precursors used (e.g., urea features a carbonyl group which EDA lacks), strongly regulate the persistence of CD photoactivity, the overall phototransformation pathway, and resultant kinetics of sunlight driven degradation. While we did not observe degradation or volatilization for CDs exposed to sunlight in the present study, the photobleaching and comparatively small changes that did occur to the CDs indeed could arise from similar reactions between CD-generated ROS and the CDs themselves³¹ (e.g., $\cdot\text{OH}$ -driven proton abstraction)⁴⁹ as suggested by the increase in photobleaching rate with increased dissolved oxygen content.

Importantly, because as-synthesized CDs remained colloidally stable in the presence of divalent cations known to destabilize other nanomaterials (Figure S11), the lack of significant changes to the physicochemical properties of CDs after photobleaching suggests that they will remain colloidally stable in natural environments after irradiation by natural sunlight. Specifically, spectroscopic measurements (i.e., XPS, ATR-FTIR) of the photobleached CDs indicate the retention of negatively charged oxygen-containing functional groups (i.e., carboxyl, hydroxyl) which are responsible for the electrostatic repulsion between CDs, an effect which is responsible for the colloidal stability of carbon dots⁵⁰⁻⁵⁴ as well as other carbon nanomaterials (CNTs,⁵⁵⁻⁵⁷ graphene⁵⁸⁻⁶⁰) in aqueous environments. Consequently, photobleached CDs can be expected to

remain stable for long periods in the water column where they will be subject to reactions with ROS (primarily $\cdot\text{OH}$) generated by the irradiation of common constituents of natural water (NOM, NO_3^-).¹⁸⁻²⁰

Effects of Indirect Photolysis.

To assess the effects of indirect photolysis on CDs, CACD and MACD suspensions were exposed to $\cdot\text{OH}$ produced via 300 nm irradiation of H_2O_2 . 300 nm irradiation of CDs in the absence of H_2O_2 photobleached CDs without any measurable structural changes. A previous analytical study enabled us to determine the Molar \times time dose of $\cdot\text{OH}$ generated during H_2O_2 photolysis (Table S1), permitting comparison to the equivalent dose of $\cdot\text{OH}$ CDs would experience in the natural environment. The kinetics of CD photodegradation by $\cdot\text{OH}$ was assessed using TC and TN, while reaction products were identified through a combination of IC, ATR-FTIR, ^{13}C -NMR, and XPS.

TC data indicated that during $\cdot\text{OH}$ exposure, both CACDs and MACDs reacted in two well defined temporal stages. First, $\sim 60\%$ of carbon atoms initially present in both CD types were lost after a $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$ exposure to $\cdot\text{OH}$ (Figure 3a). Following this rapid and extensive loss of carbon atoms, the residual carbon-containing species were significantly less susceptible to further $\cdot\text{OH}$ -mediated degradation during the second stage of the reaction as the $\cdot\text{OH}$ dose increased beyond $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$ $\cdot\text{OH}$. At an $\cdot\text{OH}$ dose of $2.8 \times 10^{-8} \text{ M} \cdot \text{min}$, the carbon content only decreased by an additional 10% for MACDs, while a dose of $4.0 \times 10^{-8} \text{ M} \cdot \text{min}$ $\cdot\text{OH}$ only decreased the carbon content by 15% for CACDs.

Complementary TN analysis (Figure 3b) revealed that the total nitrogen content was relatively insensitive to $\cdot\text{OH}$ exposure ($\cdot\text{OH}$ doses of $4.0 \times 10^{-8} \text{ M} \cdot \text{min}$), in marked contrast to the

considerable loss of carbon (compare Figures 3a and 3b). Although the total nitrogen content remained essentially constant during $\cdot\text{OH}$ exposure, nitrogen speciation evolved. Specifically, the production of nitrate was observed for $\cdot\text{OH}$ doses in excess of $5.5 \times 10^{-9} \text{ M} \cdot \text{min}$ (Figure 3b) and increased steadily until all CD nitrogen atoms were converted into nitrate ions. The constant TN content coupled with the steady increase in nitrate concentration in solution indicates that over time the $\cdot\text{OH}$ -driven degradation of CDs will produce nitrate ions.

ATR-FTIR spectra shown in Figure 4a provide spectroscopic evidence of two distinct phases in the reaction of MACDs with $\cdot\text{OH}$. The first phase of degradation was initiated upon exposure to an $\cdot\text{OH}$ dose of $6.9 \times 10^{-10} \text{ M} \cdot \text{min}$, which resulted in a decrease in intensity of the amide N-H bending mode at 1527 cm^{-1} , and a red-shift of the (carboxylic) C=O stretch to 1680 cm^{-1} indicative of a change in the chemical bonding environment in the MACDs. The simultaneous diminishment and transformation of these features indicates that $\cdot\text{OH}$ -mediated decomposition of CDs involves a cleavage of the amide moieties initially present in the CD backbone. After $\cdot\text{OH}$ exposure increased from $6.9 \times 10^{-10} \text{ M} \cdot \text{min}$ to $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$, the amide groups initially present in the CDs had been oxidized into nitro groups (i.e., NO_2), as evidenced by the simultaneous loss of the O=C-NH bend at 1527 cm^{-1} and the appearance of the ν_4/ν_1 stretches of nitro groups at 1590 cm^{-1} and 1414 cm^{-1} , respectively.⁶¹ Additionally, the appearance of peaks at $\sim 3400 \text{ cm}^{-1}$ and 1100 cm^{-1} indicate the formation of hydroxyl groups in the photoproducts^{38, 62} whose sharpness and peak positions are very similar to the hydroxyl features present in malic acid (Figure 4b). By a $\cdot\text{OH}$ dose of $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$, the previous carbonyl feature at 1700 cm^{-1} had shifted and is now centered on 1680 cm^{-1} , indicating that the carbonyl species are now in a more highly hydrogen-bound chemical environment (i.e., small carboxylic acids, such as malic acid). As the $\cdot\text{OH}$ dose increases, new bands at $1313 \text{ cm}^{-1}/831 \text{ cm}^{-1}$ also emerge which can be assigned to and the ν_3 stretch

(E')/ ν_2 bend (A_2'') of nitrate ions.⁶³ Collectively, the formation of these oxidized nitrogen groups and the similarity of degraded CD spectra to malic acid strongly suggests that CDs were depolymerized in the presence of $\cdot\text{OH}$ through the cleavage of their initial amide bonds to form nitro groups and malic acid photofragments.

The second phase of degradation shown by ATR-FTIR was observed for $\cdot\text{OH}$ doses beyond $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$, and is defined as being the period of exposure during which the spectra remained relatively unchanged with increasing $\cdot\text{OH}$ dose, aside from an increase in nitrate and nitro peak intensities. ATR-FTIR spectra observed for the solid phase (i.e., lyophilized) species produced by MACDs exposed to $\cdot\text{OH}$ doses in excess of $5.5 \times 10^{-9} \text{ M} \cdot \text{min}$ continued to exhibit peaks similar to those of sodium nitrate and the solid malic acid precursor (Figure 4b, common peaks: free O-H at 3400 cm^{-1} , carboxy C=O at 1680 cm^{-1} , and O-H deformation at 1100 cm^{-1}). Furthermore, Figure 4b demonstrates sustained exposure of both MACDs and CACDs to $\cdot\text{OH}$ in excess of $5.5 \times 10^{-10} \text{ M} \cdot \text{min}$ resulted in the formation of near identical photoproducts as suggested by the similarity of their spectra. It should be noted that XPS and ATR-FTIR analysis was not possible for $\cdot\text{OH}$ doses above $1.1 \times 10^{-8} \text{ M} \cdot \text{min}$, as the mass loss due to volatilization of carbon (as shown by TC) precluded sufficient sample recovery.

XPS spectra of CACDs exposed to $\cdot\text{OH}$ also indicate a two-phase degradation process. During the first period of degradation, $\cdot\text{OH}$ exposure up to $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$ $\cdot\text{OH}$ resulted in a loss of higher binding energy features (i.e., carbonyl carbon), and the C (1s) region was dominated by the C-C/C-H species at 285 eV with only a small shoulder at 288 eV (Figure 5a). The CDs then entered the second phase of $\cdot\text{OH}$ exposure from doses of $5.5 \times 10^{-9} \text{ M} \cdot \text{min}$ to $1.1 \times 10^{-8} \text{ M} \cdot \text{min}$, at which point the carbonyl feature reemerged in the C (1s) region at 288 eV. XPS analysis also revealed that exposure to $\cdot\text{OH}$ produced an overall decrease in the carbon content and a small

increase in the nitrogen content of the non-volatile photoproducts (Figure S12). Specifically, CACDs were initially composed of 55% carbon and 15% nitrogen, but exposure to $1.1 \times 10^{-8} \text{ M} \cdot \text{min}$ $\cdot\text{OH}$ led to products that contained 35% carbon and 20% nitrogen. This enrichment of nitrogen with increasing $\cdot\text{OH}$ exposure as measured by XPS is qualitatively consistent with the trends observed by TC/TN in that it further indicates that carbon was lost and volatilized from the CDs, while nitrogen atoms remain associated with the CD photoproduct.

The ^{13}C -NMR of CACDs exposed to $\cdot\text{OH}$ also indicated extensive degradation to the CDs. Photobleached CACDs (Figure 5b) prior to $\cdot\text{OH}$ exposure contained various carbon functional groups including carboxylic acids/amides (160-180 ppm)⁶⁴, alcohols/ethers (70-90 ppm),⁶⁵⁻⁶⁶ and both substituted (i.e., branched) and primary alkane carbons centered around 30-50 ppm and 10-30 ppm, respectively.⁶⁷⁻⁶⁸ After a $\cdot\text{OH}$ dose sufficient to cause a loss of 60% TC, ^{13}C -NMR was performed again to assess the CDs nearing the end of the first phase of degradation (see Figure 3). The resultant ^{13}C -NMR indicates that the CDs degraded through the loss of substituted/branched alkane groups (30 ppm to 50 ppm). This suggests that the crosslinked network initially present in the CDs was degraded during $\cdot\text{OH}$ exposure. Interestingly, the growth of one peak around 165 ppm, likely a carboxyl species, agrees with the increase in such species as shown with ATR-FTIR and XPS spectra.

To contextualize these results in terms of the equivalent environmental $\cdot\text{OH}$ exposure, we applied a methodology developed by Lankone et al. which allowed the integrated $\cdot\text{OH}$ dose to be benchmarked to the maximum $\cdot\text{OH}$ dose a sample would experience during one month of exposure in sunlit waters ($10^{-15} \text{ M} \cdot 2.2 \times 10^4 \text{ min/month} = 2.2 \times 10^{-11} \frac{\text{M} \cdot \text{min}}{\text{month}}$). For example, exposure to a solution of H_2O_2 at an initial concentration of 100 mM, as it is photolyzed for 4 hours results in CD exposure of $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$; this dose corresponds to an equivalent environmental exposure

of approximately 64 months ($\frac{1.4 \times 10^{-9} \text{ M} \cdot \text{min}}{2.2 \times 10^{-11} \frac{\text{M} \cdot \text{min}}{\text{month}}}$). In this study, relations to environmental timescale were done under the assumption that CDs consistently experience the maximum $\cdot\text{OH}$ dose (i.e., 10^{-15} M) in the environment, though calculations based on the full range of $[\cdot\text{OH}]_{\text{ss}}$ are reported in the SI (Table S1). Using these calculations and assuming an environmental steady state $\cdot\text{OH}$ concentration of 10^{-15} M , we can discuss CD photodegradation by indirect photolysis in terms of the equivalent environmental timescale.

The two-step degradation process first observed with TC and TN data (Figure 3a and 3b) can now be interpreted to indicate that the period of initial CD degradation in the presence of $\cdot\text{OH}$ occurs over approximately 64 +/- 5 months of indirect photolysis in sunlit waters. During this time, as indicated by ATR-FTIR and XPS, the CDs are depolymerized via the cleavage of amide groups, resulting in the formation of nitro groups and carboxylic acid species. Additionally, TC indicates that 60% of carbon atoms are lost from solution, most likely as volatile species, similar to the reactions of $\cdot\text{OH}$ with GO which evolve CO_2 as an end product.²⁴ We note the rate of loss of the amide feature at 1527 cm^{-1} closely corresponds to the rate at which the initial carbon volatilization occurs (compare TC and ATR-FTIR data in Figures 3a and 4a, respectively), suggesting that the first step in the degradation of CDs by $\cdot\text{OH}$ is hydrogen abstraction from N-H and/or C-H groups,^{49, 69-70} leading to the degradation of amide linkages. Additional evidence of this depolymerization is seen in ^{13}C -NMR data via the loss of C=C (120 ppm) and substituted/branched alkane character (30-50 ppm), representing the degradation of crosslinked character within the CDs, resulting in the primary alkane signature which persists in the resultant photoproducts (10-30 ppm) which are also carboxylated as seen in both ATR-FTIR (1680 cm^{-1} ; Figure 4a) and ^{13}C -NMR (165 ppm; Figure 5b). The redshift of the carboxyl C=O stretch coupled with the persistence of primary alkanes seen in ^{13}C -NMR provides evidence for the evolution of low molecular weight carboxylic acids after

depolymerization which are less sterically hindered than in the parent CD, thus able to participate in more extensive hydrogen bonding.³⁸

In contrast to the significant loss/volatilization of carbon atoms, the TN and ATR-FTIR data reveal that during the first 64 +/- 5 months of environmental exposure, there is little to no change in the total nitrogen content, although the nitrogen species initially present in the CDs have been converted/oxidized into nitro groups by $\cdot\text{OH}$. This oxidation occurs over the same $\cdot\text{OH}$ dose as the loss in amide character in ATR-FTIR (1646 cm^{-1}), suggesting that nitrogen atoms involved in amide linkages in the parent CDs are oxidized into nitro groups after cleavage during indirect photolysis.

Following the initial $\cdot\text{OH}$ -mediated depolymerization of the CD structure, the second phase of indirect photolysis is observed for cumulative $\cdot\text{OH}$ doses in excess of approximately $1.4 \times 10^{-9}\text{ M}\cdot\text{min}$, or beyond the first 64 +/- 5 months of environmental exposure, and is characterized by a much slower decrease in carbon content. During this phase, residual carbon atoms are further oxidized by $\cdot\text{OH}$, leading to a significant increase in the concentration of highly oxidized photoproducts structurally similar to the dicarboxylic acids used in the original CD synthesis (e.g. malic acid as shown with ATR-FTIR in Figure 4b), and related carboxylic acids (e.g. glutaric acid, Figure S13). The formation of carboxylated carbon species is consistent with the increase in concentration of the C(1s) shoulder above 286 eV in XPS. Because both CACDs and MACDs generated photoproducts with similar spectroscopic signatures, this degradation pathway is likely generalizable to CDs synthesized from EDA and carboxylic acid precursors.

When the changes in composition and structure are considered together, our results indicate that during this second stage of indirect photolysis, residual carbon atoms are oxidized by $\cdot\text{OH}$ to carboxyl groups which then serve as precursors for the formation of volatile species⁷⁰ such as

CO₂²⁴ or acetone.⁷¹ Continued •OH-mediated oxidation of previously formed nitro groups leads to the evolution of nitrate ions⁷² as indicated by the IC and ATR-FTIR data (Figures 3b and 4a). Thus, extrapolation of these trends to sustained (maximum) •OH exposure in the natural environment, in excess of the 43 years of equivalent exposure simulated in laboratory studies, indicates CDs would eventually degrade such that all carbon atoms will be volatilized while nitrogen atoms are stoichiometrically converted into nitrate ions.

The reason for the two distinct temporal regimes of carbon loss, however, remains somewhat unclear. One possible explanation is that the more recalcitrant carbon atoms seen in the second phase of the process are associated with fragments containing carbon-nitrogen bonds residual from reactions of •OH with the parent CDs. This interpretation is supported by IC data which clearly shows that the formation of nitrate ions from the CDs requires a dose of •OH in excess of 5.5×10^{-9} M•min. The evolution of nitrate ions from nitro groups requires C-N bond cleavage, a limiting prerequisite step in the volatilization of these carbon atoms compared to those in other bonding environments.

Environmental Implications

Due to their highly charged surface and small size, CDs are colloidally stable in the water column.¹¹⁻¹⁴ Thus, the effects of both direct and indirect photolysis will play an important role in determining the fate and persistence of CDs in aquatic environments. In contrast to their photostability under illumination in laboratory settings (i.e., under fluorescent lighting), the citric and malic acid-based CDs investigated in this study will rapidly photobleach after only a few hours of direct photolysis by natural sunlight, likely precluding the use of these particular CDs as sensors in outdoor settings. Once the CACDs and MACDs have photobleached, the resultant carbon

nanoparticles exhibit similar physicochemical properties to the parent CDs, suggesting a high degree of colloidal stability. Consequently, these photobleached CDs will persist for extended periods of time in the environment while being difficult to monitor or track due their poor light scattering properties and lack of fluorescent signature.

Ultimately, photobleached CDs will be degraded by indirect photolysis via reactions with ROS, primarily $\cdot\text{OH}$ produced by exposure of NOM and other constituents of water (e.g., metal ions, nitrate, dissolved oxygen) to natural sunlight.²⁰ Based on the relation to environmental timescale which was used (assuming exposure to the maximum environmental $[\cdot\text{OH}]_{\text{ss}}$),³⁴ the first stage of this indirect photolysis will occur over the course of approximately 5-6 years of environmental exposure in sunlit waters, resulting in depolymerization of the CD via cleavage of its amide bonds and the oxidation of a majority of the carbon atoms in the CDs into volatile carbon species (e.g., CO_2 and acetone). Over the same time span, nitrogen atoms will be oxidized into nitro groups bound to fragments of the parent CDs. Environmental exposure of the residual photoproducts will lead to the formation of carboxylic acid species similar to the organic acids used in the CD synthesis (e.g. malic acid). These carboxylic acids will serve as precursors to the production of volatile carbon species, a process which will continue until all carbon atoms in the CDs are ultimately mineralized (timespan of several decades based on TC trend). In contrast, nitrogen atoms in the CDs will be stoichiometrically oxidized from nitro groups into nitrate ions as indicated by IC data (half-life roughly 75 years). Given the projected lifetime and colloidal stability of photobleached CDs and their photoproducts throughout this prolonged photodegradation process, future studies identifying the environmental impact of these various intermediate species will be prudent.

A comparison of the results from this study with previous studies highlights the diversity of photochemical transformations experienced by carbon-based nanomaterials in the environment. For example, GO rapidly photofragments in natural sunlight as a result of the reactivity of the highly strained epoxide groups that exist on its surface, leading to the formation of reduced graphene oxide species (rGO) and the evolution of CO₂.²⁸ Conversely, oxidized carbon nanotubes (CNTs) do not fragment upon exposure to sunlight or ROS, but rather undergo photodecarboxylation, causing them to lose surface charge which leads to sedimentation.²⁶ Aggregates of fullerenes are prone to ROS-driven degradation by sunlight, leading to disaggregation, the formation of dissolved organic species, and eventual mineralization.²⁹ However, under simulated sunlight³⁰ and UVA light conditions,⁷³ fullerenes showed a higher level of resistance to mineralization, only exhibiting surface oxidation. These observations clearly indicate that the structure and physicochemical properties of carbon nanomaterials play a determinant role in regulating their photochemical reactions in the environment. With this in mind, it is not unreasonable that the photochemical transformations of CDs will be sensitive to their chemical structure and composition.

Although the phototransformations of the microwave-synthesized CACDs and MACDs using ethylene diamine outlined in this study parallel one another and were structurally stable for 6 weeks in natural sunlight, it has been reported that CACDs synthesized via a hydrothermal route using urea are completely mineralized in sunlight after less than one day at concentrations similar to the ones used in the present study (50 mg/L).³¹ Thus, the intrinsic chemical and physical characteristics of CDs imparted via their precursors appear to dictate their photostability. As such, it will be important to identify the different phototransformations of CDs synthesized with a wide variety of precursors and synthetic pathways. Additionally, a detailed understanding of the

mechanisms and influential factors which dictate CD photobleaching will be needed to improve photostability if CDs are to be used as environmental sensors and tracers in outdoor settings.

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Supporting Information

Detailed calculation of $\cdot\text{OH}$ dose and relation to environmental timescale (Table S1, Figure S1) and experimental procedures for CD synthesis and characterization (PL, UV-Vis, TEM, ATR-FTIR, XPS, TC, TN, IC, ^1H - and ^{13}C -NMR; Figures S2, S3, S4, S5). CD absorption compared to solar emission spectrum (Figure S6) and emission spectrum of fluorescent bulbs (Figure S9), effects of photobleaching after exposure to natural sunlight (Figure S7, S8, S10) along with stability studies of CDs in the presence of divalent cations (Figure S11). Additional figures illustrating change in XPS composition of CDs after $\cdot\text{OH}$ exposure (Figure S12) are included and ATR-FTIR comparing degraded MACDs to glutaric acid (Figure S13).

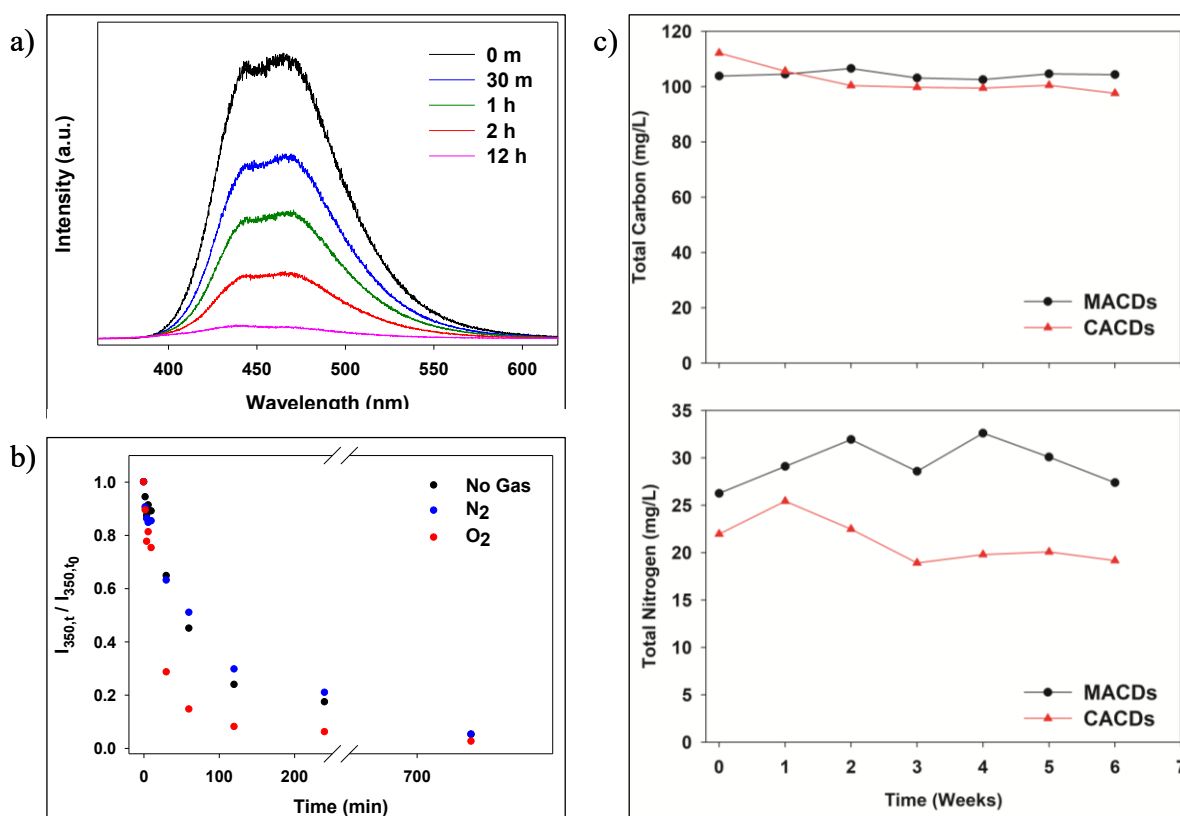


Figure 1. Effect of natural sunlight on CDs. a) Emission profiles of CACDs after exposure to 0 min (black), 30 min. (blue), 1 hr. (green), 2 hr. (red), and 12 hr. (pink) of natural sunlight. b) Emission of CACDs at 350 nm (black) compared to emission of CACDs after sparging with N₂ (blue) or O₂ (red) gas as a function of exposure to natural sunlight. c) Total carbon (top) and total nitrogen (bottom) concentration of CACDs (red triangles) and MACDs (black circles) after exposure to 0-6 weeks of natural sunlight in non-sparged water.

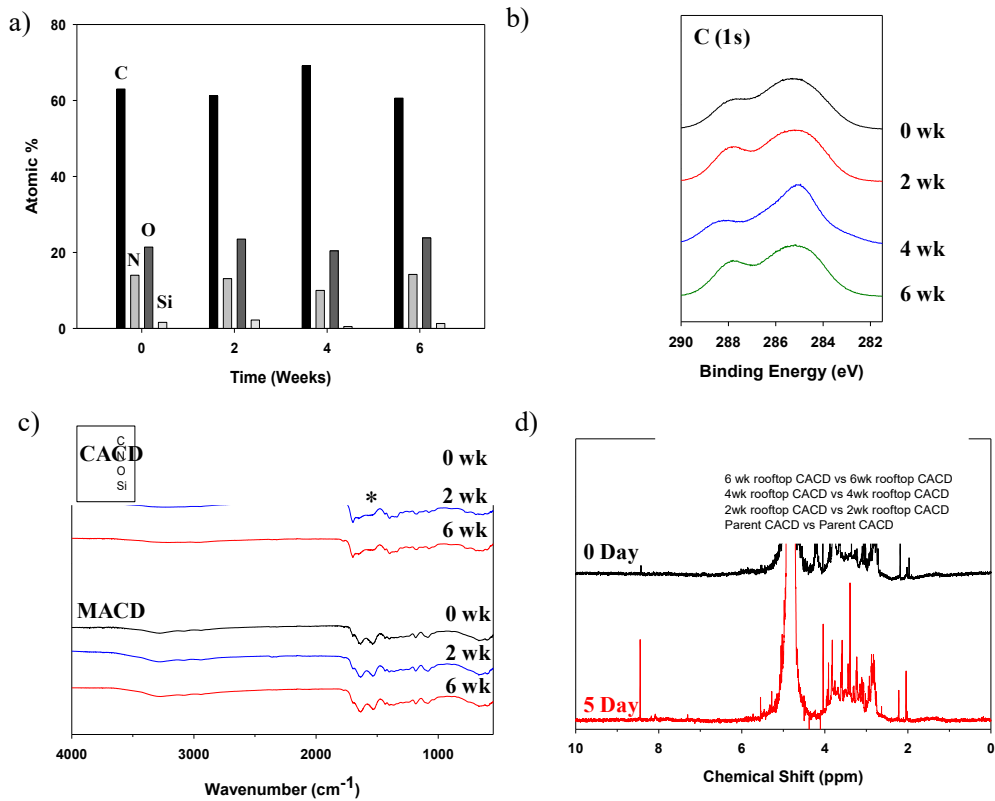


Figure 2. Effect of natural sunlight on CD structure. a) Atomic composition of lyophilized CACDs after natural sunlight exposure for 0-6 wk as determined with XPS. b) XPS C (1s) regions of lyophilized parent CACDs after 0 (black), 2 (red), 4 (blue), and 6 (green) weeks of natural sunlight exposure. (c) ATR-FTIR spectra of CACDs (top) and MACDs (bottom) exposed to natural sunlight for 0, 2, and 6 weeks, with (*) indicating the N-H bend at 1547 cm⁻¹. (d) ¹H-NMR spectra of CACDs (black) and CACDs exposed to 5 days of natural sunlight (red).

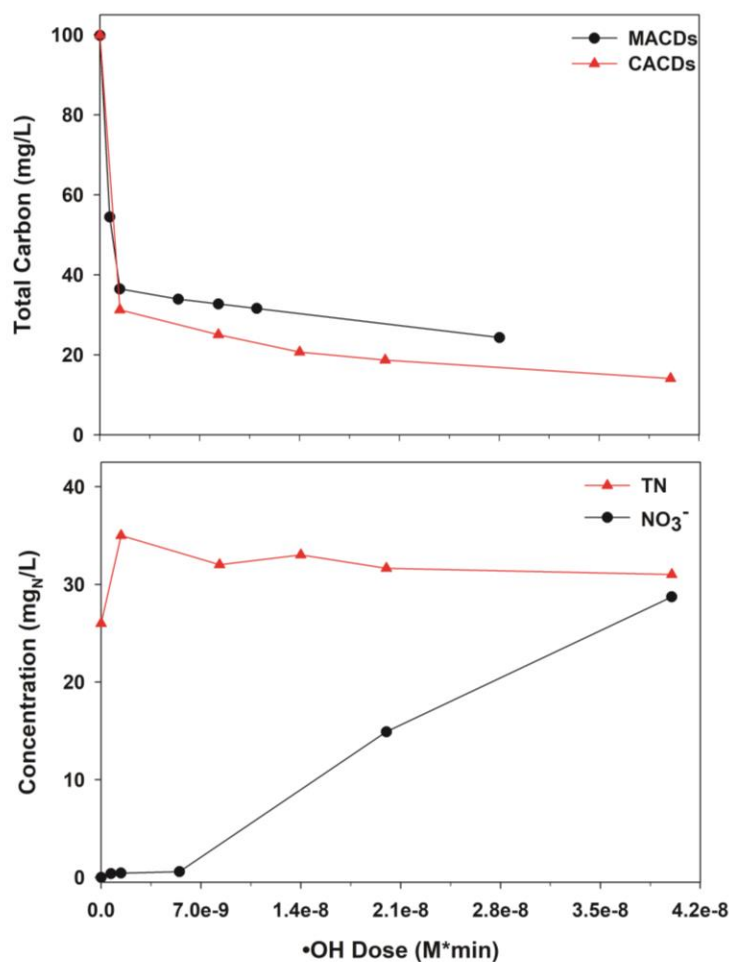


Figure 3. Degradation of CDs by $\bullet\text{OH}$. (a) Total carbon concentration of MACD (black) and CACD (red) as a function of $\bullet\text{OH}$ dose. (b) Total nitrogen (red) and nitrate concentration (black) of CACD solution in terms of mg/L nitrogen as a function of $\bullet\text{OH}$ dose.

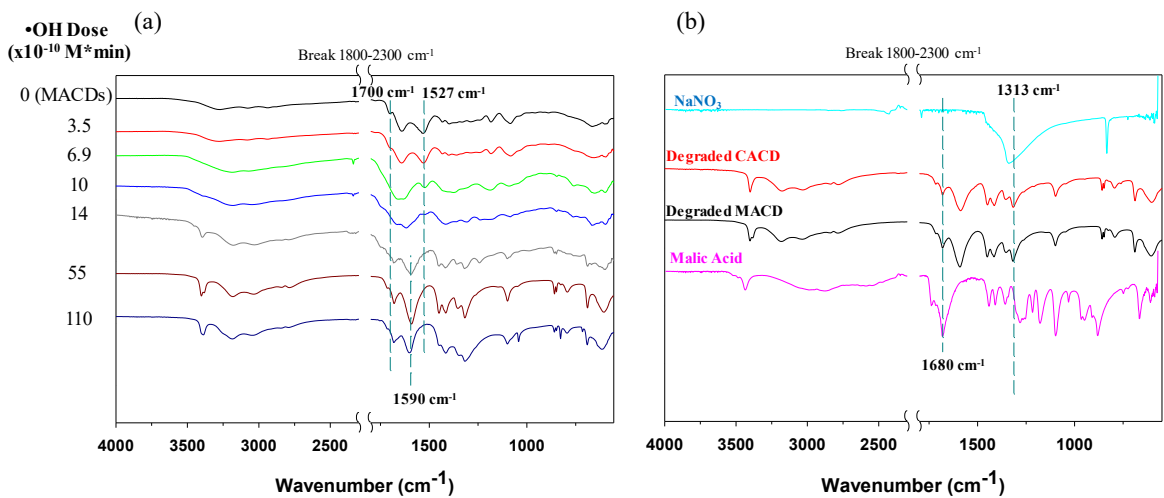


Figure 4. ATR-FTIR of lyophilized MACDs after exposure to different $\cdot\text{OH}$ doses. (a) MACDs (black) after exposure to $3.5 \times 10^{-10} \text{ M} \cdot \text{min}$ (red), $6.9 \times 10^{-10} \text{ M} \cdot \text{min}$ (light green), $1.0 \times 10^{-9} \text{ M} \cdot \text{min}$ (blue), $1.4 \times 10^{-9} \text{ M} \cdot \text{min}$ (grey), $5.5 \times 10^{-9} \text{ M} \cdot \text{min}$ (dark red), and $1.1 \times 10^{-8} \text{ M} \cdot \text{min}$ (dark blue) $\cdot\text{OH}$. Insufficient masses of photoproducts were recovered for CDs reacted with $\cdot\text{OH}$ doses $> 1.1 \times 10^{-8} \text{ M} \cdot \text{min}$, precluding ATR-FTIR analysis. Dotted lines mark carboxyl C=O (stretch, 1700 cm^{-1}), N-H (bend, 1527 cm^{-1}), and nitro (stretch, 1590 cm^{-1}) modes. (b) Degradation products of CACDs (red) and MACDs (black) after exposure to $5.5 \times 10^{-9} \text{ M} \cdot \text{min}$ $\cdot\text{OH}$ shown in comparison to malic acid (MA, pink) and sodium nitrate (blue). Dotted lines mark carboxyl C=O (stretch, 1680 cm^{-1}) and nitrate ν_3 (stretch, 1313 cm^{-1}) modes.

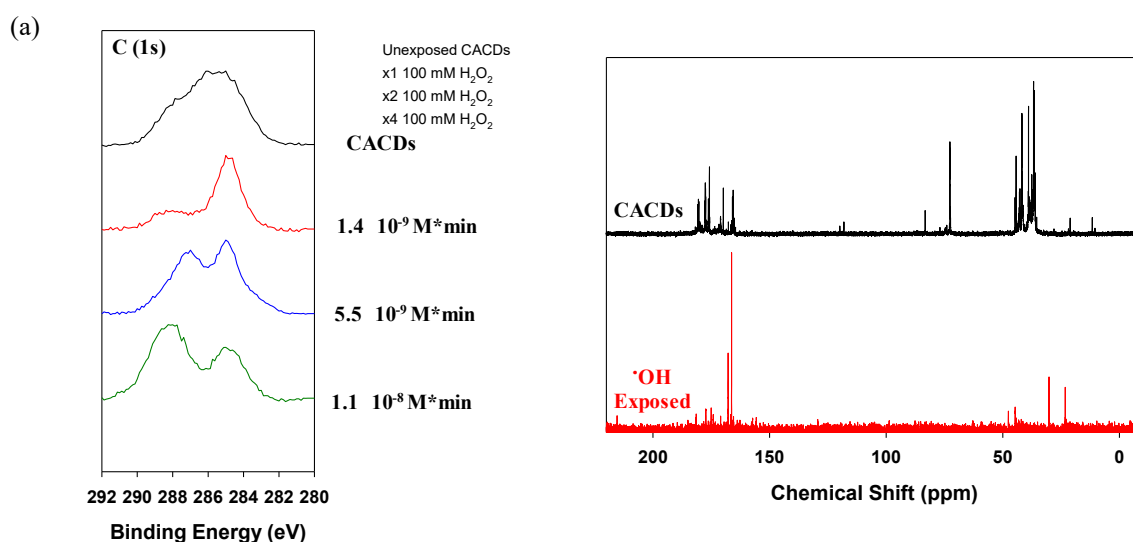


Figure 5. a) C 1s) region of lyophilized CACDs (black) after exposure to a dose of 1.4×10^{-9} M*min (red), 5.5×10^{-9} M*min (blue), and 1.1×10^{-8} M*min (green) [•]OH. Insufficient masses of photoproducts were recovered for CDs reacted with [•]OH doses $> 1.1 \times 10^{-8}$ M*min, precluding XPS analysis. b) ¹³C-NMR of photobleached CACDs in D₂O before (black) and after (red) exposure to 5.7×10^{-8} M*min [•]OH. At the significantly higher CD concentration required for NMR (1.7×10^4 mg/L), this [•]OH dose leads to the loss of 60% of carbon from solution as determined with TC.

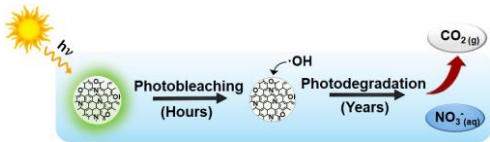
References

1. Yan, F.; Sun, Z.; Zhang, H.; Sun, X.; Jiang, Y.; Bai, Z., The fluorescence mechanism of carbon dots, and methods for tuning their emission color: a review. *Microchimica Acta* **2019**, *186* (8), 583.
2. Liu, M. L.; Chen, B. B.; Li, C. M.; Huang, C. Z., Carbon dots: synthesis, formation mechanism, fluorescence origin and sensing applications. *Green Chem.* **2019**, *21* (3), 449-471.
3. Huang, S.; Li, W.; Han, P.; Zhou, X.; Cheng, J.; Wen, H.; Xue, W., Carbon quantum dots: synthesis, properties, and sensing applications as a potential clinical analytical method. *Anal. Methods* **2019**, *11* (17), 2240-2258.
4. Zhang, Z.; Zhang, J.; Chen, N.; Qu, L., Graphene quantum dots: an emerging material for energy-related applications and beyond. *Energ. Environ. Sci.* **2012**, *5* (10), 8869-8890.
5. Fei, H.; Ye, R.; Ye, G.; Gong, Y.; Peng, Z.; Fan, X.; Samuel, E. L. G.; Ajayan, P. M.; Tour, J. M., Boron- and nitrogen-doped graphene quantum dots/graphene hybrid nanoplatelets as efficient electrocatalysts for oxygen reduction. *ACS Nano* **2014**, *8* (10), 10837-10843.
6. Hettiarachchi, S. D.; Graham, R. M.; Mintz, K. J.; Zhou, Y.; Vanni, S.; Peng, Z.; Leblanc, R. M., Triple conjugated carbon dots as a nano-drug delivery model for glioblastoma brain tumors. *Nanoscale* **2019**, *11* (13), 6192-6205.
7. Zhang, Z.; Lei, Y.; Yang, X.; Shi, N.; Geng, L.; Wang, S.; Zhang, J.; Shi, S., High drug-loading system of hollow carbon dots–doxorubicin: preparation, in vitro release and pH-targeted research. *J. Mater. Chem. B* **2019**, *7* (13), 2130-2137.
8. Zhi, B.; Cui, Y.; Wang, S.; Frank, B. P.; Williams, D. N.; Brown, R. P.; Melby, E. S.; Hamers, R. J.; Rosenzweig, Z.; Fairbrother, D. H.; Orr, G.; Haynes, C. L., Malic acid carbon dots: from super-resolution live-cell imaging to highly efficient separation. *ACS Nano* **2018**, *12* (6), 5741-5752.
9. Boakye-Yiadom, K. O.; Kesse, S.; Opoku-Damoah, Y.; Filli, M. S.; Aquib, M.; Joelle, M. M. B.; Farooq, M. A.; Mavlyanova, R.; Raza, F.; Bavi, R.; Wang, B., Carbon dots: Applications in bioimaging and theranostics. *Int. J. Pharm.* **2019**, *564*, 308-317.
10. Sciortino, A.; Cannizzo, A.; Messina, F., Carbon nanodots: a review—from the current understanding of the fundamental photophysics to the full control of the optical response. *C — Journal of Carbon Research* **2018**, *4* (4), 67.
11. Liu, X.; Li, J.; Huang, Y.; Wang, X.; Zhang, X.; Wang, X., Adsorption, aggregation, and deposition behaviors of carbon dots on minerals. *Environ. Sci. Technol.* **2017**, *51* (11), 6156-6164.
12. Bayati, M.; Dai, J.; Zambrana, A.; Rees, C.; Fidalgo de Cortalezzi, M., Effect of water chemistry on the aggregation and photoluminescence behavior of carbon dots. *J. Environ. Sci.* **2018**, *65*, 223-235.
13. Li, Q.; Chen, B.; Xing, B., Aggregation kinetics and self-assembly mechanisms of graphene quantum dots in aqueous solutions: cooperative effects of pH and electrolytes. *Environ. Sci. Technol.* **2017**, *51* (3), 1364-1376.
14. Dager, A.; Uchida, T.; Maekawa, T.; Tachibana, M., Synthesis and characterization of mono-disperse carbon quantum dots from fennel seeds: photoluminescence analysis using machine learning. *Sci. Rep.* **2019**, *9* (1), 14004.
15. Kamrani, S.; Rezaei, M.; Kord, M.; Baalousha, M., Transport and retention of carbon dots (CDs) in saturated and unsaturated porous media: role of ionic strength, pH, and collector grain size. *Water Res.* **2018**, *133*, 338-347.
16. Havrdova, M.; Hola, K.; Skopalik, J.; Tomankova, K.; Petr, M.; Cepe, K.; Polakova, K.; Tucek, J.; Bourlinos, A. B.; Zboril, R., Toxicity of carbon dots – effect of surface functionalization on the cell viability, reactive oxygen species generation and cell cycle. *Carbon* **2016**, *99*, 238-248.

17. Tabish, T. A.; Scotton, C. J.; Ferguson, D. C. J.; Lin, L.; Veen, A. v. d.; Lowry, S.; Ali, M.; Jabeen, F.; Ali, M.; Winyard, P. G.; Zhang, S., Biocompatibility and toxicity of graphene quantum dots for potential application in photodynamic therapy. *Nanomedicine* **2018**, *13* (15), 1923-1937.
18. Zepp, R. G.; Hoigne, J.; Bader, H., Nitrate-induced photooxidation of trace organic chemicals in water. *Environ. Sci. Technol.* **1987**, *21* (5), 443-450.
19. Page, S. E.; Logan, J. R.; Cory, R. M.; McNeill, K., Evidence for dissolved organic matter as the primary source and sink of photochemically produced hydroxyl radical in arctic surface waters. *Environ. Sci. Proc. Imp.* **2014**, *16* (4), 807-822.
20. Gligorovski, S.; Streckowski, R.; Barbati, S.; Vione, D., Environmental implications of hydroxyl radicals ($\bullet\text{OH}$). *Chem. Rev.* **2015**, *115* (24), 13051-13092.
21. Miller, P. L.; Chin, Y.-P., Indirect Photolysis Promoted by Natural and Engineered Wetland Water Constituents: Processes Leading to Alachlor Degradation. *Environ. Sci. Technol.* **2005**, *39* (12), 4454-4462.
22. Lam, M. W.; Mabury, S. A., Photodegradation of the pharmaceuticals atorvastatin, carbamazepine, levofloxacin, and sulfamethoxazole in natural waters. *Aquatic Sciences* **2005**, *67* (2), 177-188.
23. Vione, D.; Maddigapu, P. R.; De Laurentiis, E.; Minella, M.; Pazzi, M.; Maurino, V.; Minero, C.; Kouras, S.; Richard, C., Modelling the photochemical fate of ibuprofen in surface waters. *Water Res.* **2011**, *45* (20), 6725-6736.
24. Hou, W.-C.; Henderson, W. M.; Chowdhury, I.; Goodwin, D. G.; Chang, X.; Martin, S.; Fairbrother, D. H.; Bouchard, D.; Zepp, R. G., The contribution of indirect photolysis to the degradation of graphene oxide in sunlight. *Carbon* **2016**, *110*, 426-437.
25. Zepp, R. G.; Faust, B. C.; Hoigne, J., Hydroxyl radical formation in aqueous reactions (pH 3-8) of iron(II) with hydrogen peroxide: the photo-Fenton reaction. *Environ. Sci. Technol.* **1992**, *26* (2), 313-319.
26. Chen, C.-Y.; Jafvert, C. T., Photoreactivity of carboxylated single-walled carbon nanotubes in sunlight: reactive oxygen species production in water. *Environ. Sci. Technol.* **2010**, *44* (17), 6674-6679.
27. Hou, W.-C.; Beigzadeh Milani, S.; Jafvert, C. T.; Zepp, R. G., Photoreactivity of unfunctionalized single-wall carbon nanotubes involving hydroxyl radical: chiral dependency and surface coating effect. *Environ. Sci. Technol.* **2014**, *48* (7), 3875-3882.
28. Hou, W.-C.; Chowdhury, I.; Goodwin, D. G.; Henderson, W. M.; Fairbrother, D. H.; Bouchard, D.; Zepp, R. G., Photochemical transformation of graphene oxide in sunlight. *Environ. Sci. Technol.* **2015**, *49* (6), 3435-3443.
29. Hou, W.-C.; Jafvert, C. T., Photochemical transformation of aqueous C60 clusters in sunlight. *Environ. Sci. Technol.* **2009**, *43* (2), 362-367.
30. Sanchis, J.; Aminot, Y.; Abad, E.; Jha, A. N.; Readman, J. W.; Farré, M., Transformation of C60 fullerene aggregates suspended and weathered under realistic environmental conditions. *Carbon* **2018**, *128*, 54-62.
31. Chen, X.; Fang, G.; Liu, C.; Dionysiou, D. D.; Wang, X.; Zhu, C.; Wang, Y.; Gao, J.; Zhou, D., Cotransformation of carbon dots and contaminant under light in aqueous solutions: a mechanistic study. *Environ. Sci. Technol.* **2019**, *53* (11), 6235-6244.
32. de Medeiros, T. V.; Manioudakis, J.; Noun, F.; Macairan, J.-R.; Victoria, F.; Naccache, R., Microwave-assisted synthesis of carbon dots and their applications. *J. Mater. Chem. C* **2019**, *7* (24), 7175-7195.
33. Zhi, B.; Gallagher, M. J.; Frank, B. P.; Lyons, T. Y.; Qiu, T. A.; Da, J.; Mensch, A. C.; Hamers, R. J.; Rosenzweig, Z.; Fairbrother, D. H.; Haynes, C. L., Investigation of phosphorous doping effects on polymeric carbon dots: fluorescence, photostability, and environmental impact. *Carbon* **2018**, *129*, 438-449.
34. Lankone, R. S.; Barclay, M.; Deline, A. R.; Fairbrother, D. H., Quantifying hydroxyl radical concentrations and total dose via principle component analysis of UV-Vis Spectroscopy. *Anal. Methods* **Submitted**.

- 581 35. Haag, W. R.; Hoigné, J., Photo-sensitized oxidation in natural water via .OH radicals.
582 *Chemosphere* **1985**, *14* (11), 1659-1671.
- 583 36. Schneider, J.; Reckmeier, C. J.; Xiong, Y.; von Seckendorff, M.; Sussha, A. S.; Kasák, P.; Rogach,
584 A. L., Molecular fluorescence in citric acid-based carbon dots. *J. Phys. Chem. C* **2017**, *121* (3), 2014-
585 2022.
- 586 37. Yuan, H.; Yu, J.; Feng, S.; Gong, Y., Highly photoluminescent pH-independent nitrogen-doped
587 carbon dots for sensitive and selective sensing of p-nitrophenol. *RSC Adv.* **2016**, *6* (18), 15192-15200.
- 588 38. Max, J.-J.; Chapados, C., Infrared spectroscopy of aqueous carboxylic acids: malic acid. *J. Phys.*
589 *Chem. A* **2002**, *106* (27), 6452-6461.
- 590 39. Dousseau, F.; Pezolet, M., Determination of the secondary structure content of proteins in
591 aqueous solutions from their amide I and amide II infrared bands. Comparison between classical and
592 partial least-squares methods. *Biochemistry* **1990**, *29* (37), 8771-8779.
- 593 40. Liu, W.; Li, C.; Ren, Y.; Sun, X.; Pan, W.; Li, Y.; Wang, J.; Wang, W., Carbon dots: surface
594 engineering and applications. *J. Mater. Chem. B* **2016**, *4* (35), 5772-5788.
- 595 41. Colthup, N. B., Spectra-structure correlations in the infra-red region. *J. Opt. Soc. Am.* **1950**, *40*
596 (6), 397-400.
- 597 42. Wishart, D. S.; Case, D. A., [1] - Use of Chemical Shifts in Macromolecular Structure
598 Determination. In *Methods in Enzymology*, James, T. L.; Dötsch, V.; Schmitz, U., Eds. Academic Press:
599 2002; Vol. 338, pp 3-34.
- 600 43. Fouda, A. S.; Elmorsi, M. A.; Shaban, S. M.; Fayed, T.; Azazy, O., Evaluation of N-(3-(dimethyl
601 hexadecyl ammonio)propyl) palmitamide bromide as cationic surfactant corrosion inhibitor for API N80
602 steel in acidic environment. *Egypt. J. Pet.* **2018**, *27* (4), 683-694.
- 603 44. Shaban, S. M., N-(3-(Dimethyl benzyl ammonio)propyl)alkanamide chloride derivatives as
604 corrosion inhibitors for mild steel in 1 M HCl solution: experimental and theoretical investigation. *RSC*
605 *Adv.* **2016**, *6* (46), 39784-39800.
- 606 45. Szilágyi, L.; Jardetzky, O., α -Proton chemical shifts and secondary structure in proteins. *Journal*
607 *of Magnetic Resonance (1969)* **1989**, *83* (3), 441-449.
- 608 46. Blundell, C. D.; DeAngelis, P. L.; Day, A. J.; Almond, A., Use of ^{15}N -NMR to resolve
609 molecular details in isotopically-enriched carbohydrates: sequence-specific observations in hyaluronan
610 oligomers up to decasaccharides. *Glycobiology* **2004**, *14* (11), 999-1009.
- 611 47. Blundell, Charles D.; Deangelis, Paul L.; Almond, A., Hyaluronan: the absence of amide-
612 carboxylate hydrogen bonds and the chain conformation in aqueous solution are incompatible with stable
613 secondary and tertiary structure models. *Biochem J.* **2006**, *396* (3), 487-498.
- 614 48. Hill, S. A.; Benito-Alifonso, D.; Davis, S. A.; Morgan, D. J.; Berry, M.; Galan, M. C., Practical
615 three-minute synthesis of acid-coated fluorescent carbon dots with tuneable core structure. *Sci. Rep.* **2018**,
616 *8* (1), 12234.
- 617 49. Stadtman, E. R., Oxidation of free amino acid and amino acid residues in proteins by radiolysis
618 and by metal-catalyzed reactions. *Annu. Rev. Biochem.* **1993**, *62* (1), 797-821.
- 619 50. Zhao, P.; Zhu, L., Dispersibility of carbon dots in aqueous and/or organic solvents. *Chemical*
620 *Communications* **2018**, *54* (43), 5401-5406.
- 621 51. Zhang, B.; Liu, C.-y.; Liu, Y., A Novel One-Step Approach to Synthesize Fluorescent Carbon
622 Nanoparticles. *European Journal of Inorganic Chemistry* **2010**, *2010* (28), 4411-4414.
- 623 52. Xu, M.; Xu, S.; Yang, Z.; Shu, M.; He, G.; Huang, D.; Zhang, L.; Li, L.; Cui, D.; Zhang, Y.,
624 Hydrophilic and blue fluorescent N-doped carbon dots from tartaric acid and various alkylol amines under
625 microwave irradiation. *Nanoscale* **2015**, *7* (38), 15915-15923.
- 626 53. Wang, W.; Damm, C.; Walter, J.; Nacken, T. J.; Peukert, W., Photobleaching and stabilization of
627 carbon nanodots produced by solvothermal synthesis. *Physical Chemistry Chemical Physics* **2016**, *18* (1),
628 466-475.
- 629 54. Fiuza, T.; Gomide, G.; Campos, A. F. C.; Messina, F.; Depeyrot, J., On the Colloidal Stability of
630 Nitrogen-Rich Carbon Nanodots Aqueous Dispersions. *C — Journal of Carbon Research* **2019**, *5* (4), 74.

55. Smith, B.; Wepasnick, K.; Schrote, K. E.; Cho, H.-H.; Ball, W. P.; Fairbrother, D. H., Influence of Surface Oxides on the Colloidal Stability of Multi-Walled Carbon Nanotubes: A Structure–Property Relationship. *Langmuir* **2009**, *25* (17), 9767-9776.
56. Zhang, L.; Wang, M.; Fang, J.; Yang, K.; Lin, D., The effect of oxidation on physicochemical properties and aqueous stabilization of multiwalled carbon nanotubes: comparison of multiple analysis methods. *Science China Chemistry* **2016**, *59* (11), 1498-1507.
57. Bai, Y.; Wu, F.; Lin, D.; Xing, B., Aqueous stabilization of carbon nanotubes: effects of surface oxidization and solution chemistry. *Environmental Science and Pollution Research* **2014**, *21* (6), 4358-4365.
58. Hadadian, M.; Goharshadi, E. K.; Youssefi, A., Electrical conductivity, thermal conductivity, and rheological properties of graphene oxide-based nanofluids. *Journal of Nanoparticle Research* **2014**, *16* (12), 2788.
59. Nuncira, J.; Seara, L. M.; Sinisterra, R. D.; Caliman, V.; Silva, G. G., Long-term colloidal stability of graphene oxide aqueous nanofluids. *Fullerenes, Nanotubes and Carbon Nanostructures* **2019**, 1-11.
60. Qi, Y.; Xia, T.; Li, Y.; Duan, L.; Chen, W., Colloidal stability of reduced graphene oxide materials prepared using different reducing agents. *Environ. Sci.: Nano* **2016**, *3* (5), 1062-1071.
61. Iogansen, A. V.; Litovchenko, G. D., The characteristic bands of the stretching vibrations of the nitro group in infrared absorption. *J. Appl. Spectrosc.* **1965**, *3* (6), 404-411.
62. Barańska, H.; Kuduk-Jaworska, J.; Szostak, R.; Romaniewska, A., Vibrational spectra of racemic and enantiomeric malic acids. *J. Raman Spectrosc.* **2003**, *34* (1), 68-76.
63. Brooker, M. H.; Irish, D. E., Vibrational frequency assignments of isotopically different forms of nitrate ion in ionic nitrate crystals. *Can. J. Chem.* **1970**, *48* (8), 1198-1201.
64. Howarth, O. W.; Lilley, D. M. J., Carbon-13-NMR of peptides and proteins. *Prog. Nucl. Mag. Res. Sp.* **1978**, *12* (1), 1-40.
65. Katsuraya, K.; Hatanaka, K.; Matsuzaki, K.; Amiya, S., Assignment of finely resolved ¹³C NMR spectra of poly(vinyl alcohol). *Polymer* **2001**, *42* (24), 9855-9858.
66. Ding, S.; Hong, Y.-W.; Chen, C.-Y.; Chang, N.-C., One and two dimensional ¹H and ¹³C high resolution NMR investigation of lariat ethers and their alkali metal ionic complexes: A more tangible evidence for the presence of less common C–H···O hydrogen bonds. *Biophys. Chem.* **2006**, *121* (2), 75-83.
67. Gilbert, A.; Yamada, K.; Yoshida, N., Exploration of intramolecular ¹³C isotope distribution in long chain n-alkanes (C11–C31) using isotopic ¹³C NMR. *Org. Geochem.* **2013**, *62*, 56-61.
68. Ritter, W.; Hull, W.; Cantow, H. J., Determination of the most stable conformers of branched alkanes by ¹³C-NMR spectroscopy at very low temperatures. *Tetrahedron Lett.* **1978**, *19* (34), 3093-3096.
69. Doan, H. Q.; Davis, A. C.; Francisco, J. S., Primary steps in the reaction of OH radicals with peptide systems: perspective from a study of model amides. *J. Phys. Chem. A* **2010**, *114* (16), 5342-5357.
70. Vel Leitner, N. K.; Berger, P.; Legube, B., Oxidation of amino groups by hydroxyl radicals in relation to the oxidation degree of the α-carbon. *Environ. Sci. Technol.* **2002**, *36* (14), 3083-3089.
71. Quici, N.; Morgada, M. E.; Gettar, R. T.; Bolte, M.; Litter, M. I., Photocatalytic degradation of citric acid under different conditions: TiO₂ heterogeneous photocatalysis against homogeneous photolytic processes promoted by Fe(III) and H₂O₂. *Appl. Catal. B-Environ.* **2007**, *71* (3), 117-124.
72. Pelizzetti, E.; Minero, C.; Piccinini, P.; Vincenti, M., Phototransformations of nitrogen containing organic compounds over irradiated semiconductor metal oxides: Nitrobenzene and Atrazine over TiO₂ and ZnO. *Coord. Chem. Rev.* **1993**, *125* (1), 183-193.
73. Hwang, Y. S.; Li, Q., Characterizing photochemical transformation of aqueous nC₆₀ under environmentally relevant conditions. *Environ. Sci. Technol.* **2010**, *44* (8), 3008-3013.



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