

Practical Speedup of Bayesian Inference of Species Phylogenies by Restricting the Space of Gene Trees

Yaxuan Wang,¹ Huw A. Ogilvie,¹ and Luay Nakhleh^{*,1}

¹Computer Science Department, Rice University, Houston, TX

*Corresponding author: E-mail: nakhleh@rice.edu.

Associate editor: Kelley Harris

Abstract

Species tree inference from multilocus data has emerged as a powerful paradigm in the postgenomic era, both in terms of the accuracy of the species tree it produces as well as in terms of elucidating the processes that shaped the evolutionary history. Bayesian methods for species tree inference are desirable in this area as they have been shown not only to yield accurate estimates, but also to naturally provide measures of confidence in those estimates. However, the heavy computational requirements of Bayesian inference have limited the applicability of such methods to very small data sets. In this article, we show that the computational efficiency of Bayesian inference under the multispecies coalescent can be improved in practice by restricting the space of the gene trees explored during the random walk, without sacrificing accuracy as measured by various metrics. The idea is to first infer constraints on the trees of the individual loci in the form of unresolved gene trees, and then to restrict the sampler to consider only resolutions of the constrained trees. We demonstrate the improvements gained by such an approach on both simulated and biological data.

Key words: species tree, multispecies coalescent, Bayesian MCMC, efficiency.

Introduction

Species tree inference under the multispecies coalescent (MSC) model accounts for gene tree heterogeneity that arises due to incomplete lineage sorting (Tajima 1983). This model has gained much attention in the years since the first inferential methods to implement it were developed (Takahata et al. 1995; Yang 2002; Degnan and Rosenberg 2009). A wide array of methods that assume or are inspired by the MSC have been devised (Liu et al. 2010; Liu and Yu 2011; Chifman and Kubatko 2014; Mirarab et al. 2014; Wang and Nakhleh 2018), including the Bayesian methods of Ogilvie et al. (2017), Flouri et al. (2018), and Zhu et al. (2018). The MSC was recently extended to the multispecies network coalescent to account for reticulation (in addition to incomplete lineage sorting; see Yu et al. 2014) and Bayesian methods for inference under this model have been devised (Wen et al. 2016; Wen and Nakhleh 2018; Wen et al. 2018; Zhang et al. 2018; Zhu et al. 2018).

The power of Bayesian methods lies in their ability to incorporate prior knowledge, infer values of parameters beyond the tree topology, and provide measures of confidence in the inference based on the posterior that they sample (Huelsenbeck et al. 2001). However, for these Bayesian methods to approximate the true posterior distribution, they demand significant computational resources, an issue that has thus far limited their applicability in terms of both the number of taxa and number of loci in the data set (Ogilvie et al. 2016). This is why incomplete lineage sorting aware methods that have been proven to be statistically consistent under the MSC and, at the same time very efficient computationally, are

used to infer large-scale species trees (Liu et al. 2010; Liu and Yu 2011; Chifman and Kubatko 2014; Mirarab et al. 2014). However, these methods focus almost exclusively on the species tree topology and provide neither accurate information on other parameters, such as divergence times and population sizes, nor confidence intervals for their inferences. The question we address in this article is: Can the convergence of Bayesian methods be improved in practice without sacrificing the accuracy of the information they provide?

A rich body of literature exists on the development of methods for statistical inference outside phylogenetics, much of which has been adopted by Bayesian phylogenetic methods. The ubiquitous Markov chain Monte Carlo (MCMC) arose from nuclear weapons research (Robert and Casella 2011), and is the basis for tree and network inference in MrBayes, *BEAST, PhyloNet, and other software tools. The efficiency of MCMC for phylogenetics has been improved with the development of new MCMC proposals (e.g., Höhna and Drummond 2012; Yang and Rodríguez 2013; Zhang et al. 2020), including proposals designed to improve the mixing of MSC models (e.g., Yang and Rannala 2014; Jones 2017; Rannala and Yang 2017).

Metropolis coupling to accelerate MCMC was developed for the inference of spatial statistics (Geyer 1991), and has been implemented in various phylogenetics software (Ronquist et al. 2012; Wen et al. 2018; Bouckaert et al. 2019). Variational Bayes is a radically different approach which fits parametric distributions to model parameters, unlike MCMC which is nonparametric. Variational Bayes was originally developed for graphical models (Attias 1999), and has recently been applied to compute posterior distributions and

marginal likelihoods of phylogenetic trees (Fourment and Darling 2019; Zhang and Matsen 2019).

All of these methods were developed decades before their adoption for phylogenetic inference because tree and network space is far more complex than the typical multidimensional parameter space. The number of unrooted or rooted trees grows superexponentially with the number of taxa (Felsenstein 1978), and is for all practical purposes infinite when the number of taxa is large. Multilocus MSC inference embeds gene trees within a species tree, with the constraint that between-species coalescent events must take place earlier in time than the most recent common ancestor (MRCA) time of the involved species. This multiplies the complexity of the inference problem by increasing the number of trees to infer, and because the probability distributions of node heights for different trees are not independent.

Rather than trying to adapt an algorithm developed for other fields of natural sciences or mathematics, we have developed a heuristic method that specifically applies to the problem of multi-locus MSC inference. The heuristic method constrains the space of gene tree topologies to allow for faster convergence and, consequently, analyses of larger data sets. The idea behind our approach is very simple: A set of constraints in the form of a tree which is usually less than fully resolved is estimated independently for each individual locus, and then MCMC walks in the portion of the tree space that is consistent with these constraints.

In other words, the MCMC sampler considers only gene trees that are consistent with the constraints on the individual loci. Using simulated data under a variety of conditions and employing several metrics for assessing performance, we demonstrate that this simple approach results in computational improvements relative to unconstrained Bayesian MCMC without sacrificing accuracy. We then analyze a biological data set and show that the new approach enables analyses that had before necessitated dividing the data set into smaller ones.

Our work presents an approach for improving the computational requirements of Bayesian inference of species phylogenies. The constraints on the individual loci can be obtained in various ways and the proposals that satisfy these constraints can be derived in multiple ways as well. In this work, we implemented one specific method for obtaining the constraints and a standard set of proposals that satisfy them. As this approach can be adopted by any Bayesian species phylogeny inference method, both of these components can be further modified to achieve even further improvement to the computational requirements of Bayesian inference.

New Approach

A Bayesian formulation of the multi-locus species tree inference problem is to estimate the posterior distribution over species tree topologies, population sizes and species divergence times from multiple sequence alignments given a model which includes at least some demographic function (e.g., constant population sizes) and substitution process (e.g., Jukes and Cantor 1969).

Here, we use S to represent the species tree, Θ to represent the population sizes and divergence times, and X to represent the multiple sequence alignments. Inspired by Rannala and Yang (2017), we can formulate the model as

$$f(S, \Theta|X) = \frac{1}{f(X)} f(S, \Theta) f(X|S, \Theta). \quad (1)$$

In this formulation, $f(S, \Theta|X)$ is the posterior probability of the species tree topology and associated parameters, $f(X)$ is the marginal likelihood, $f(S, \Theta)$ is the prior on the species tree topology and associated parameters, and $f(X|S, \Theta)$ is the likelihood. To calculate the likelihood we must integrate over gene trees G and substitution model parameters ψ :

$$f(X|S, \Theta) = \int_{\psi} \int_G f(X|G, \psi) f(G|S, \Theta) f(\psi) dG d\psi. \quad (2)$$

In the above formulation, $f(X|G, \psi)$ is the phylogenetic likelihood and substitution model parameters, $f(G|S, \Theta)$ is the coalescent likelihood, and $f(\psi)$ is the prior for the same parameters. Note that the coalescent and phylogenetic likelihoods are functions of a gene tree, and are calculated over the space of all possible gene trees. When using MCMC, we only need a density proportional to the posterior probability, so the marginal likelihood can be omitted.

Furthermore, under the common assumption of recombination-free, unlinked loci, the likelihood can be derived from the product of integrations for each locus:

$$f(X|S, \Theta) = \prod_i \int_{\psi} \int_G f(X_i|G_i, \psi) f(G_i|S, \Theta) f(\psi) dG_i d\psi. \quad (3)$$

where X_i is the multiple sequence alignment of the i th locus in the data set and G_i is the gene tree sampled at locus i .

As the integration in equation (3) cannot be derived analytically, MCMC sampling algorithms are often employed to obtain samples from the posterior distribution and approximate it based on those samples. Due to the scaling problems of MCMC inference (Ogilvie et al. 2016), current algorithms to approximate equation (3) become computationally infeasible for larger data sets, getting stuck in the peaks and troughs of the posterior distribution and requiring extremely large numbers of iterations to converge.

Our approach to tackle the computational challenge works as follows. For each sequence alignment X_i , maximum likelihood (ML) with bootstrapping is run to obtain a set of gene trees from which a majority-rule consensus tree with a prespecified support threshold x is built. For example, for $x = 90$, a majority-rule consensus tree is built where only clades that appear in at least 90% of the bootstrap trees are included. We denote this majority-rule consensus tree by C_i (we use the value of x explicitly in the naming only when it is not clear from the context) and call it a constraint gene tree (CGT). Our approach now samples according to equation (3) with one difference: The integration is taken over gene trees that are consistent with, that is, refinements of, their respective C_i constraints. For example, if $C_i = (((A, B), C), (D, E, F))$, the sampler considers gene

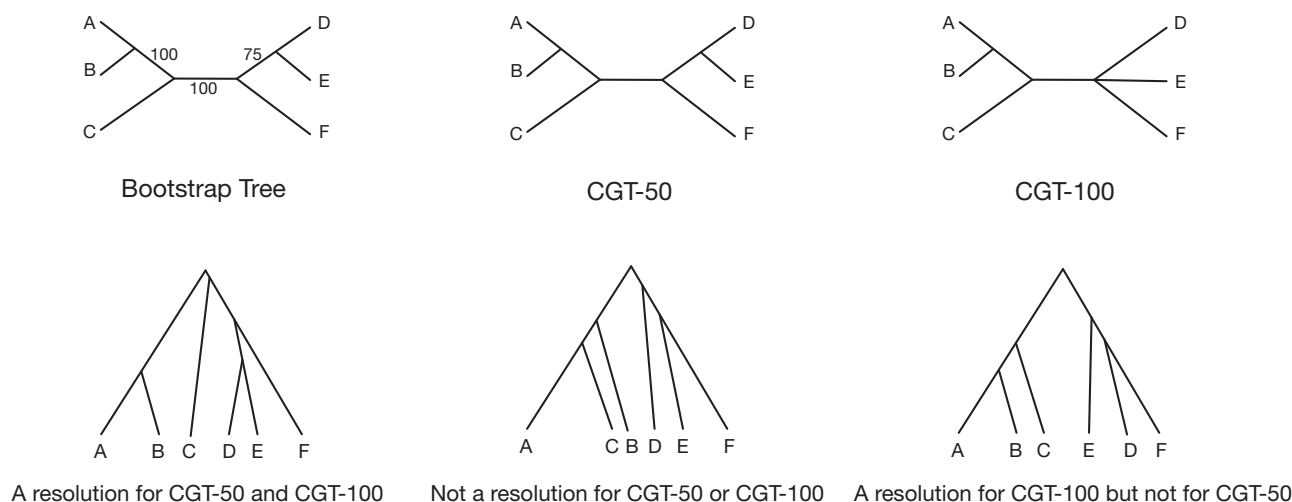


Fig. 1. Constraint trees, their resolutions, and acceptable moves. Bootstrap tree and CGTs under the consensus threshold of 50 (CGT-50) and 100 (CGT-100) are shown in the first row. In the second row, three possible proposed gene trees are provided. The left tree is acceptable given the constraints CGT-50 and CGT-100. The middle tree is not acceptable given the constraints CGT-50 or CGT-100. The right tree is acceptable given the constraint CGT-100 but not CGT-50.

tree $G_i = (((A, B), C), ((D, E), F))$ as it is a refinement of C_i , but does not consider gene tree $G_i = (((A, B), (C, D)), (E, F))$ as it is not a refinement of C_i . This concept is illustrated in figure 1.

We implemented this restricted sampling of gene tree topologies by comparing proposed topologies with the constraint trees, and rejecting incompatible proposals. Future implementations may be made even more efficient by only proposing compatible topologies.

Observe that if C_i is a star phylogeny (a tree that has no internal branches), then the method is effectively sampling according to equation (3), whereas if C_i is fully resolved (a binary tree), then the sampler fixes the gene tree topology for locus i and only samples its parameters. It is important to note that C_i imposes only topological constraints; that is, C_i has no branch lengths. Furthermore, we take C_i to be unrooted, so that the sampler is allowed to sample the roots of the gene trees.

The posterior of species trees includes a “long tail” region of model trees where the likelihood of each tree is very small but the number of trees within such region is very large. Unfortunately, as the scale of the data set increases, the autocorrelation time of the MCMC chain increases dramatically (Ogilvie et al. 2016). The motivation of our approach is that by constraining the gene trees, the sampler can avoid the long tail and have better mixing. While utilizing these constraints necessarily means that the sampler is not sampling from the same posterior distribution as an unconstrained version of the sampler (except for the case where the constraints are star phylogenies), we demonstrate below that this has very little impact on the accuracy of the sampler in practice.

Hereafter, we write CMCMC to denote constrained MCMC according to our new approach and UCMCMC to denote the unconstrained version of MCMC. We also write CMCMC- x , where x is a value between 50 and 100, to denote the use of CMCMC with support threshold x in the majority-

rule consensus tree, or a value of 0 to denote use with the ML tree.

Parameter values must be initialized somehow at the beginning of each MCMC chain. For both CMCMC and UCMCMC we initialized gene trees by inferring the ML tree for each locus in RAXML (Stamatakis 2014). We used the topology of the ML tree, and for each internal node used the maximum distance from that node to any descendant leaf as the node height. Although there is a chance that the ML tree is incompatible with the constraint tree due to the stochastic nature of bootstrapping, this is probably very uncommon as we did not encounter this in any of our analyses.

Results and Discussion

A simulation study was carried out to comprehensively analyze the performance of CMCMC and UCMCMC. We varied the simulation data set along three dimensions: evolutionary scenarios of species, the number of loci and the number of taxa. When we focused on one dimension, the other two dimensions were fixed. More details are provided in the Materials and Methods section. To simulate different scenarios of complexity and signal in the data, we varied evolutionary time scales and population sizes in four categories:

- “OH”: old divergence times and high population size;
- “OL”: old divergence times and low population size;
- “YH”: young divergence times and high population size; and
- “YL”: young divergence times and low population size.

To further examine the performance of each method, we varied the number of loci (10, 20, 40) for the YH condition whereas fixing the number of taxa as 16. We also varied the numbers of taxa (16, 32, 48) for the YH condition and fixed the number of loci as 10. Unless otherwise stated, there are 10 replicates for each condition.

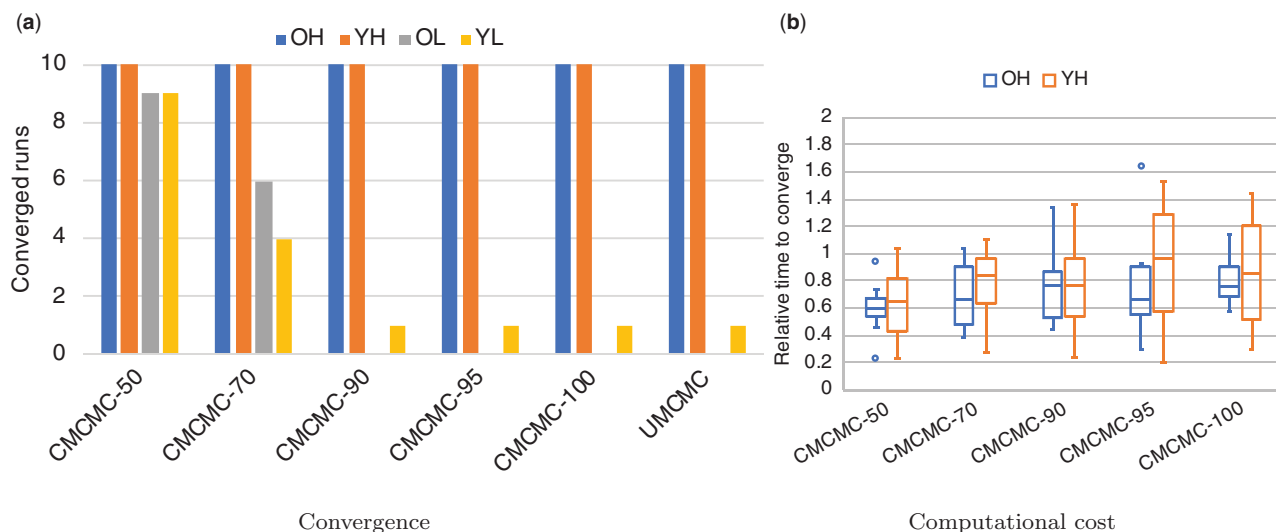


Fig. 2. Convergence and efficiency of CCMCMC and UCMCMC. (a) Convergence of samplers with or without constraint gene trees. Different samplers are shown on the x-axis and the y-axis shows the number of data sets (out of 10) on which the sampler converged. (b) Ratios of iterations required for convergence using CCMCMC compared with UCMCMC. Values above 1 are faster using UCMCMC, below 1 are faster using CCMCMC.

CGTs Improve the Convergence of MCMC

The ability to converge within a reasonable time is a key metric to evaluate the performance of an MCMC sampler. An effective sample size (ESS) of at least 200 is used as a threshold for convergence in the popular MCMC diagnostic and analysis tool Tracer (Rambaut et al. 2018). In this work, we target the same convergence standard for continuous parameters including the posterior probability, likelihood, prior probability, coalescent likelihood, tree height and population size. We terminated any chain still running after 72 h.

Figure 2(a) shows that decreasing the consensus threshold enables convergence for low population size conditions, which is impossible for UCMCMC within 72 h. We also show the improved convergence of CCMCMC as the number of loci increases in [supplementary figure S1, Supplementary Material online](#) and as the number of taxa increases in [supplementary figure S2, Supplementary Material online](#).

When MCMC chains are able to converge, CCMCMC reduces the number of iterations required for convergence into less than half that of UCMCMC under various evolutionary parameters as shown in [figure 2b](#). Furthermore, CCMCMC took fewer iterations than UCMCMC to converge for different numbers of loci and different numbers of taxa ([supplementary figs. S3 and S4, Supplementary Material online](#)).

CCMCMC and UCMCMC Derive Similar Posterior Distributions

The ultimate goal of phylogenetic inference problem with Bayesian sampling is to approximate the posterior distribution of the true species tree. One way to verify the posterior distribution is to compare the average standard deviation of split frequencies (ASDSF; Lakner et al. 2008) of the 95% credible set. Note that the 95% credible set or interval of a well calibrated Bayesian method covers the true value in 95% of cases.

The smaller the ASDSF is, the more similar the species tree distributions are. A threshold of 0.01 on the ASDSF is commonly used to assess the convergence of two chains. An ASDSF value below 0.01 is taken to indicate that the chains are likely to be sampling from the same underlying distribution (for examples, see [Stunženas et al. 2011](#); [Stensvold et al. 2012](#); [Mazza et al. 2016](#)).

Figure 3 shows the ASDSF between the CCMCMC and UCMCMC chains for different evolutionary scenarios, different numbers of loci, and different numbers of taxa. To compare the ASDSF, the MCMC chains must be converged. However, although CCMCMC has better convergence ability than UCMCMC, we only compare the ASDSF when all MCMC chains can converge. For all simulated scenarios in [figure 3a](#), the ASDSF values of CCMCMC and UCMCMC in most replicates are below 0.01. As we increased the number of loci and taxa, all ASDSF interquartile ranges (IQRs) fell below 0.01, except for CCMCMC-50 for which the top of the range could be slightly above 0.01, as shown in [figure 3b and c](#).

As the number of loci was increased, ASDSF decreased. This was expected as with more data more nodes in the species tree become fully resolved, and the difference in support between CCMCMC and UCMCMC for those nodes will be zero. Conversely, ASDSF increased as the number of taxa was increased. Again this was expected, as denser taxon sampling will reduce the proportion of fully resolved nodes, and hence the proportion of nodes with zero difference between methods. In summary, although the underlying distributions sampled by CCMCMC and UCMCMC are different by construction, our results show that in practice they are almost the same.

If an internal node in one CGT is binary, we consider such node as resolved. For young divergence time scenarios, there are fewer substitutions and hence less information available with which to reconstruct the phylogeny. For a given threshold, CGTs in young divergence time scenarios were less

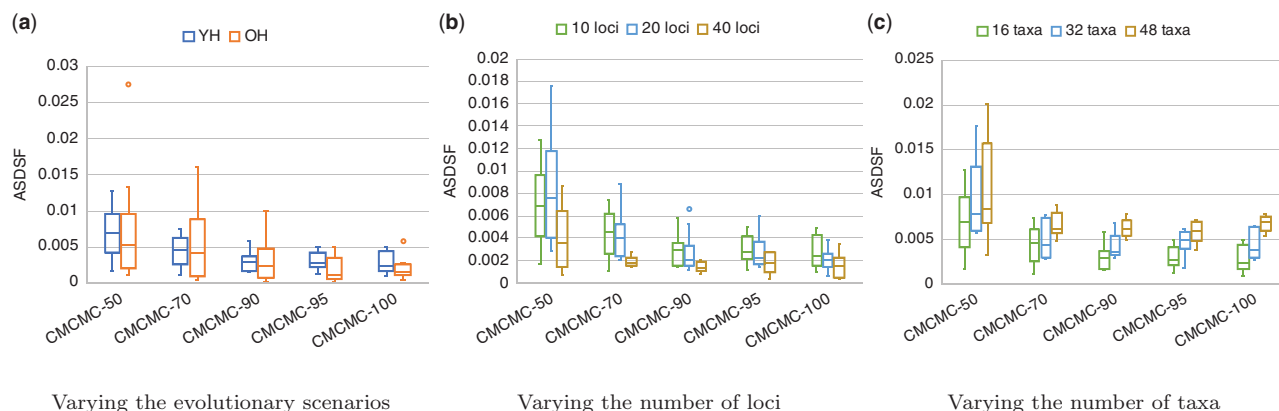


FIG. 3. ASDSF between the CMC and UMC chains. The x-axis lists CMC samplers with different support thresholds and the y-axis shows the ASDSF between each CMC method and UMC. (a) ASDSF values when varying the divergence times fixing the number of taxa and loci as 16 and 10. Only “YH” and “OH” are shown because UMC cannot converge in “YL” and “OL” scenarios. (b) ASDSF values when varying the number of loci, restricted to the “YH” scenario. There are 10 replicates for 10- and 20-locus data sets. For 40-locus data sets, results are shown for the 6 out of 10 replicates where all methods converged within 20 days. (c) ASDSF values when varying the number of taxa, restricted to the “YH” scenario. There are 10 replicates for 16-, 32-, and 48-taxon data sets.

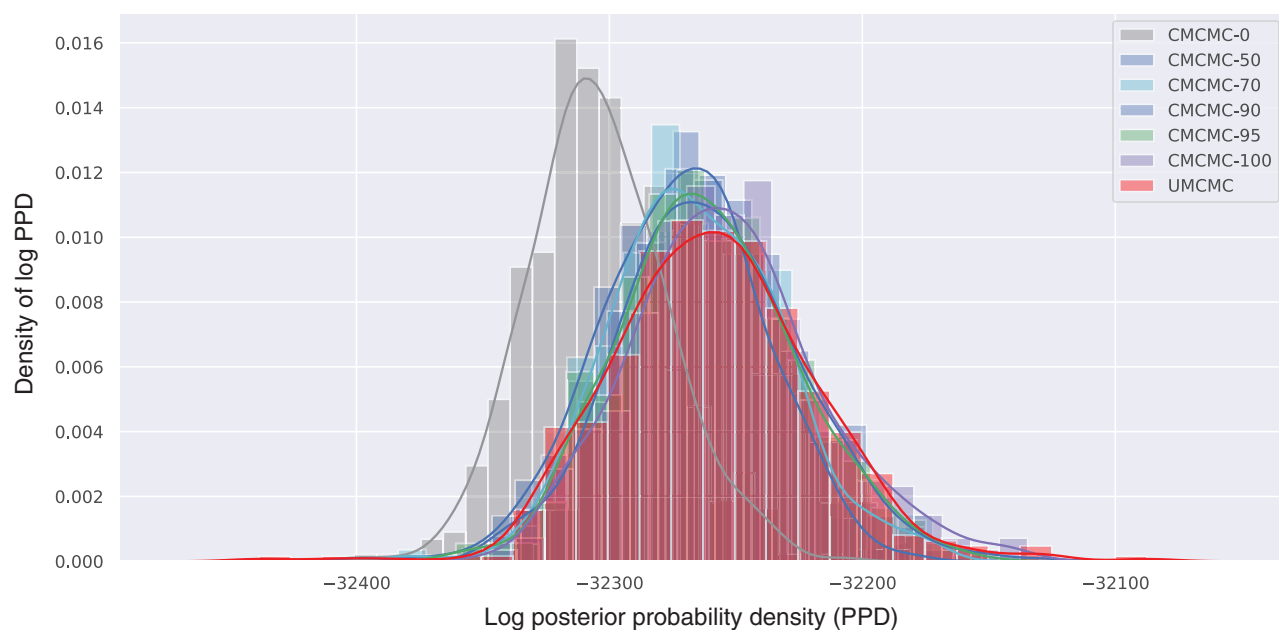


FIG. 4. Kernel Density Estimation of the posterior probability distribution of CMC and UMC samples. Different methods are shown in different colors. All methods are run on the example sequence data which contains 16 taxa and 10 independent loci for the young divergence times and high population size scenario.

resolved than CGTs in old divergence time scenarios in [supplementary figure S5, Supplementary Material](#) online. But for all conditions the proportion of resolved nodes steadily decreased as the threshold was raised. A similar trend was observed when increasing the number of taxa in [supplementary figure S6, Supplementary Material](#) online. This shows the role that the support threshold plays as a useful tuning parameter for our heuristic.

Although the ASDSF provides a numeric measure reflecting the similarity between the distributions being sampled, we visualize in [figure 4](#) the distributions sampled by the various samplers as an illustration of this similarity. Although decreasing the support threshold increases the difference in the

posterior distribution, all CMC methods approximate posterior density distributions similar to UMC, except for CMCMC-0.

Our aim for CMC is to closely approximate the unconstrained posterior distribution of species trees, and to do so faster than UMC. The posterior density distribution of CMCMC-0 is very divergent from UMC and from CMC when using other thresholds, implying that it is no longer closely approximating the unconstrained posterior distribution. For this reason, we do not recommend using CMCMC-0 (i.e., gene trees with fixed, ML estimated unrooted topologies) for Bayesian inference of species trees from sequences.

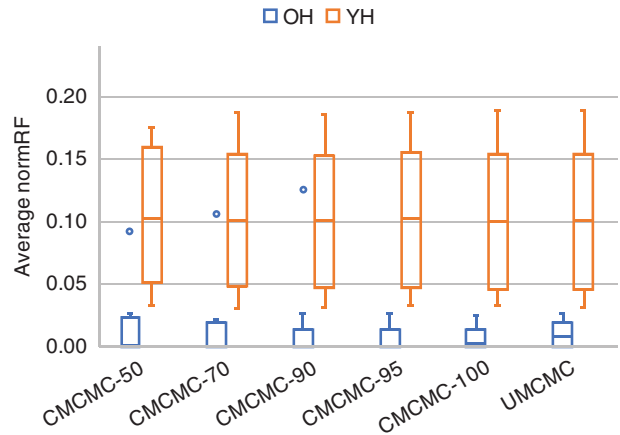


Fig. 5. Topological accuracy of the species trees inferred by CMC and UMC. The data pertained to simulations of 10 loci from 16 taxa under the OH and YH scenarios.

CMC and UMC Predictions Are Essentially Identical in Accuracy

Although Bayesian MCMC provides an approximation of the posterior distribution over parameters, the topology of the species tree is often the main quantity of interest. We compared the average Robinson–Foulds (RF) distance (Robinson and Foulds 1981) between the true species tree and the inferred species tree topology in the 95% credible set to assess the topological accuracy. For each topology in the 95% credible set, we calculated the RF distance and divided it by the maximum possible RF distance (twice the number of internal branches in the species tree) to derive the normalized RF distance (normRF; Kupczok et al. 2010). Then we averaged all normRF distances, weighted by the frequency of each topology in the 95% credible set. More details about how to calculate average normRF distance are provided in the Evaluation Metrics section.

Figure 5 shows the topological accuracy of the species trees inferred by the various methods. As the figure shows, under both evolutionary scenarios, all samplers infer almost the same species trees. For the OH scenario the outlier corresponds to a single-species tree which is difficult to accurately infer because of its short internal branches (supplementary fig. S7, Supplementary Material online). The same lack of variation is also observed when varying the numbers of loci and taxa (supplementary figs. S8 and S9, Supplementary Material online).

Within a condition, there was no observed trend in average normRF distance (fig. 5, supplementary figs. S8 and S9, Supplementary Material online). In addition to topological error, we also calculated “branch length error” as the average branch score of trees in the credible set. The calculation is described in detail in the Evaluation Metrics subsection. As expected, branch length error was higher for the “old” case because branch score is not scale invariant, and average normRF distance was higher for the “young” case due to the lower rate of informative mutations. But within a condition, there was no difference in branch length error between CMC and UMC (supplementary figs. S10–S12,

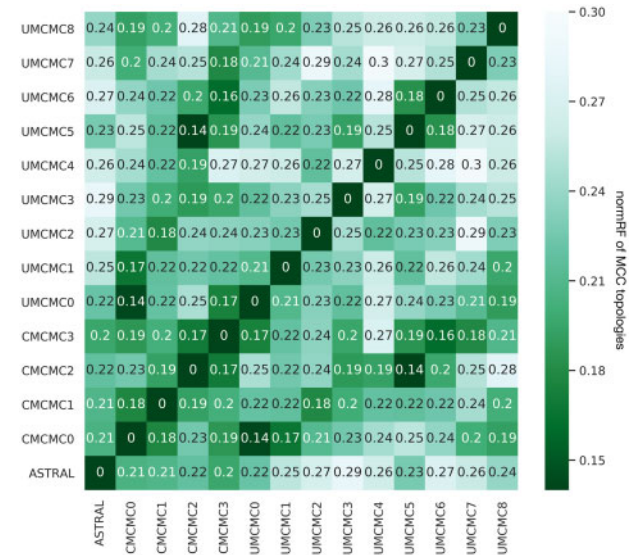


Fig. 6. Discordance among phylogenies estimated by ASTRAL, CMC, and UMC. CMC was applied to four nonoverlapping subsets of 64 loci each, UMC was applied to nine nonoverlapping subsets of 32 loci each, and ASTRAL was applied to gene trees inferred from all 304 loci. The color and the number in each entry of the matrix indicate the normalized Robinson–Foulds distance between maximum clade credibility (MCC) species trees estimated from each subset.

Supplementary Material online). These results further demonstrate the proximity of the posterior distribution of species trees inferred by CMC to UMC.

Analysis of a Biological Data Set

Recently, a study that applied exon capture sequencing to Australian rainbow skinks (Bragg et al. 2018) compared the phylogenies inferred using summary MSC methods, a full Bayesian MSC method and concatenation. However, due to the computational time required, the full Bayesian species tree method was only applied to nonoverlapping 32 locus subsets of the data, despite 304 highly informative loci being available. This data set contains 46 taxa from 43 recognized species.

CMC enabled us to double the number of loci, so inspired by Bragg et al. we compared the species trees inferred using UMC from nine nonoverlapping 32 locus subsets with those inferred using CMC from four nonoverlapping 64 locus subsets. All analyses were run until satisfactory convergence was observed. The samples for each analysis were summarized as maximum clade credibility (MCC) trees. To quantify the variation between species trees inferred from different subsets, we calculated the normRF distance between the MCC tree from one MCMC chain or the inferred tree from ASTRAL (Mirarab et al. 2014). As shown in figure 6, CMC derived more consistent results compared with UMC, as the highest normRF distance between CMC subsets was 0.23, but the highest pairwise distance between the smaller UMC subsets was 0.3.

The precision of the 64 locus CMC posterior distributions was higher, as expected given the larger number of loci

employed. For both normRF and branch score, the average distances between the MCC tree and individual samples in the posterior distribution were smaller for CMCMC (supplementary figs. S13 and S14, Supplementary Material online).

The increased precision of the CMCMC analyses enables taxonomic refinement of rainbow skinks. When 32 loci are used with UCMCMC, the trio *Carlia inconnexa*, *Carlia pectoralis*, and *Carlia rubigo* form a clade but the relationships within that clade are unclear, as the best supported topology for this trio has *C. rubigo* as sister with an average posterior probability of 69% across subsets. When 64 loci are used with CMCMC, this average rises to 98% (supplementary figs. S15 and S16, Supplementary Material online).

Conclusions

In this article, we reported on a simple heuristic method for speeding the convergence of Bayesian MCMC under the MSC. The heuristic works by restricting the space of gene trees that can be sampled. The constraints can be obtained in various ways including bootstrap trees contracted according to some support threshold, majority-rule consensus trees of posterior samples, or even constraints provided based on biological knowledge. As the approach restricts the explored space by design, we evaluated the method's performance in terms of convergence and, when converged, the distribution it samples. The evaluation was done on simulated data sets as well as a biological data set, and for evaluation metrics, we focused mainly on the time to convergence, the ASDSF between the constrained version of the sampler (CMCMC) and the unconstrained one (UCMCMC), as well as the topological accuracy of the inferred species trees.

We have demonstrated that CGTs are advantageous in two distinct ways. For data sets where it is challenging to achieve convergence with UCMCMC, for example, those which did not converge even after 20 days in our study, CMCMC can converge within a reasonable time. For data sets which did readily converge using UCMCMC, applying CGTs reduced the required number of iterations.

Following from our results, we recommend that CMCMC can be applied in two ways. The first is to accelerate preliminary analyses, where CMCMC-50 can be used to infer posterior distributions of species trees which are very close to the unconstrained posterior distributions. This has the additional benefit of reducing the amount of resources required for a given study, which range from grant money to pay for computer time, to CPU hours which may be in high demand at a given institution, to the electricity and natural resources needed to manufacture and operate computer nodes. As many preliminary analyses may have to be run for a given study, this reduction can be substantial.

The second is for final published analyses, where researchers may wish to be more conservative and avoid approximations and heuristics wherever possible. For example, MCMC chains of finite lengths and variational Bayes are both approximate and heuristic methods, but are unavoidable for Bayesian inference of phylogenetic trees from sequences. CGTs are an additional heuristic which are avoidable for data sets where

convergence is possible within a reasonable time using UCMCMC, but unavoidable otherwise. So our final recommendation is to employ CMCMC for final published analyses of data sets which fail to converge using UCMCMC.

Materials and Methods

MCMC Implementation and Settings

CMCMC can be easily implemented in commonly used Bayesian based phylogenetics software packages, such as PhyloNet (Wen et al. 2016), BEAST (Drummond and Rambaut 2007), and BEAST2.5 (Bouckaert et al. 2019). In this article, we implemented CMCMC in PhyloNet. In all our experiments, we generated 50 bootstrap trees for each locus and obtained the majority-rule consensus trees from those. Firstly, we generated bootstrap trees given an alignment using RAxML (Stamatakis 2014). Then, we estimated the constraint tree given a specific support threshold.

We applied a uniform prior over the species tree topologies, a uniform prior $U(0, \infty)$ on species tree node heights, and a $1/X$ prior on the mean population size. More implementation details are provided in supplementary data, Supplementary Material online.

Executing MCMC Chains

For the simulation study, we first ran each chain for three days on the DAVinCI computing cluster. All jobs executed on this cluster ran on 2.83 GHz Intel Westmere CPUs. Jobs were restarted each day, and so the total running time of the MCMC chain was <72 h, as for each job some time was spent queuing and postprocessing.

For any chain that did not converge within three days, we restarted it from the beginning on the NOTS computing cluster at Rice University. Jobs executed on this cluster were randomly assigned to one of the following CPUs: Intel Xeon E5-2650 v2 at 2.6 GHz, Xeon E5-2650 v4 at 2.2 GHz, Xeon Gold 6126 at 2.6 GHz, or Intel Xeon Gold 6230 at 2.1 GHz. With the exception of the 48 taxon analyses, all chains were run for 20 days. The 48 taxon chains were run for only 10 days because we noticed they had all converged by that time. As with the shorter chains, jobs were restarted each day so the total runtime was <480 h, and <240 h for the 48 taxon chains.

For the empirical study, 10 independent chains with different random seeds but otherwise identical data and settings were run for each locus subset and method. This was necessary to achieve convergence on these relatively large data sets. CMCMC chains on 64 loci were run for 160 million iterations, taking ~ 25 days. UCMCMC chains on 32 loci were run for 120 million iterations, taking ~ 10 days. After all 10 chains had finished running for a given subset and method, the remaining samples were concatenated after removing the 10% burnin.

Simulating Data

For all simulated data sets, we used DendroPy (Sukumaran and Holder 2010) to generate random species trees and ms (Hudson 2002) to generate gene trees on these species trees under the MSC. Sequence data were generated by Seq-Gen (Rambaut and Grass 1997) under the Jukes–Cantor model

Table 1. The Evolutionary Parameters Varied to Control the Complexity and Signal in the Data.

	Low Population Sizes	High Population Sizes
Old divergence times	OL: 50 Ma, 100 k individuals	OH: 50 Ma, 500 k individuals
Young divergence times	YL: 10 Ma, 100 k individuals	YH: 10 Ma, 500 k individuals

(Jukes and Cantor 1969). We derived the CGT for each locus by bootstrapping from the sequences by RAxML (Stamatakis 2014).

Because species tree inference methods are employed over a range of evolutionary time scales and to clades with different population sizes, we varied both parameters for our simulation study which are shown in table 1. For each simulated species tree we scaled its root to different heights; an “old” height of 50 Ma akin to the rice-Pooideae split (Sandve et al. 2008), and a “young” height of 10 Ma akin to the split of gorillas with humans and chimpanzees (Langergraber et al. 2012). Gene trees were scaled before simulating sequences so that the branch lengths in substitutions per site corresponded to a substitution rate of 2.5×10^{-3} per million years. This rate is slightly faster than the rates observed for the RAG1 nuclear gene in animals (Hugall et al. 2007), but within the ranges observed for plant nuclear genes (Huang et al. 2003).

For both the old and young species tree scales, we simulated gene trees under large and small population sizes of 500,000 and 100,000, respectively, with annual generation times. Chimpanzees, gorillas and ancient humans all have effective population sizes (N_e) of around 20,000 individuals (Huff et al. 2010). Assuming a great ape generation time of 25 years, this population will have the same distribution of coalescent times as a clade of species with annual generation times and an N_e of 500,000, the same as our large population size condition. Given that human effective population sizes are often considered low, the small population size condition therefore corresponds to species with very low effective population sizes.

The evolutionary parameters also affect the proportion of resolved internal nodes of the CGT as shown in supplementary figure S5, Supplementary Material online. The “OH” and “OL” scenarios have higher proportion of resolved internal nodes than the “YH” and “YL” which means that the substitution rate more effectively restricts the gene tree search space. The proportion of resolved internal nodes in consensus trees decreases as the number of taxa increases as shown in supplementary figure S6, Supplementary Material online. In contrast, the population size does not have such obvious effect as the substitution rate or number of taxa.

More details on the simulations are provided in supplementary data, Supplementary Material online.

Biological Data

We analyzed the Australian skinks data set which is provided in Bragg et al. (2018). We randomly selected one sample from each species. Note that the species names in the data set and in the article are not consistent. More details about how to

map the species in the data set and in the article are shown in supplementary table S1, Supplementary Material online.

The Australian skinks data set contains three ingroup genera: *Carlia*, *Lygisaurus*, and *Liburnascincus*. There are 46 taxa from 43 recognized species. All details of the biological data including genus, species, tissue, collection, sample library, and focal clade are provided in supplementary table S2, Supplementary Material online.

To obtain informative gene trees, we included 304 complete informative loci whose length ranges from 240 to 6,534 sites. Supplementary figure S17, Supplementary Material online shows the proportion of resolved internal nodes of CGTs for different ranges of sequence length. In general, as the length of sequence increases the number of resolved internal nodes gets larger. This is because longer sequences are likely to contain more substitutions to inform the resolution of nodes.

Evaluation Metrics

Effective Sample Size

The ESS is the number of effectively independent draws from some distributions sampled by the MCMC chain. Adequate ESS is a sign of good mixing of the MCMC chain and it has been argued that the ESS should be >200 (Kuhner 2009), a value that has been adopted in the Bayesian phylogenetics community. Therefore, an MCMC chain where the ESS of all selected probability densities and parameter values were >200 was considered to have converged. The probability densities were of the posterior, phylogenetic likelihood, prior and coalescent likelihood, all of which are dependent on the tree topologies and continuous parameters. The parameter values were tree height and population size.

Average RF Distance

When calculating the total RF distance, we only considered posterior samples where the species tree topology was within the 95% credible set. We call this credible set of posterior samples T^* to distinguish it from the full set of posterior samples T . To quantify differences between true tree t and the 95% credible set T^* , we calculate the average normRF distance (Kupczok et al. 2010) as

$$\frac{1}{|T^*|} \sum_{t^* \in T^*} \text{normRF}(t, t^*). \quad (4)$$

Average Branch Length Error

To evaluate the accuracy of branch length estimates, we calculated the average error between the true tree t and the 95% credible set T^* using a measure based on Euclidean branch score distances (Kuhner and Felsenstein 1994; St. John 2017). For every tree t^* in the credible set, we take the union B of all branches in t^* and t , where a branch $b \in B$ is defined by the taxa associated with its tipward node (i.e., the corresponding clade). We define $\delta(b)$ as the difference between the length of b in t and t^* . If a branch is missing in one of t and t^* , its length in that tree is defined to be zero. The sum of square differences $\sum_{b \in B} \delta(b)^2$ is known as the branch score distance,

which we will treat as a function $BSD(t, t^*)$. The square root of the branch score is a Euclidean distance, and we define branch length error as

$$\frac{1}{|T^*|} \sum_{t^* \in T^*} \sqrt{BSD(t, t^*)}. \quad (5)$$

Average Standard Deviation of Split Frequencies

ASDSF is a measure of convergence that has been used in tools, such as ExaBayes (Aberer et al. 2014) and MrBayes (Ronquist et al. 2012). ASDSF can be calculated by comparing split or clades frequencies between two MCMC chains. Given two posterior distributions T_1 and T_2 from two MCMC chains and their corresponding 95% credible sets T_1^* and T_2^* , C is all unique, nontrivial clades in $T_1^* \cup T_2^*$. Set C^* is defined as

$$C^* = \{c \in C | \max(f(c, T_1), f(c, T_2)) \geq \epsilon\}, \quad (6)$$

where $f(c, T)$ is the frequency of clade c in distribution T , and ϵ is a threshold. We used $\epsilon = 0.1$, the same as the default setting in MrBayes 3.2 (Ronquist et al. 2012). Finally, the ASDSF between T_1 and T_2 is defined as

$$ASDSF(T_1, T_2) = \frac{1}{|C^*|} \sum_{c \in C^*} \frac{|f(c, T_1) - f(c, T_2)|}{2}, \quad (7)$$

because the standard deviation of two numbers is half of the absolute difference.

Supplementary Material

Supplementary data are available at *Molecular Biology and Evolution* online.

Acknowledgments

This study was supported in part by National Science Foundation (NSF) grants DBI-1355998, CCF-1514177, CCF-1800723, DMS-1547433, and by the Data Analysis and Visualization Cyberinfrastructure funded by NSF under grant OCI-0959097 and Rice University. Ana C. Afonso Silva assisted us in interpreting and mapping the species names of the biological data set.

References

- Aberer AJ, Kobert K, Stamatakis A. 2014. ExaBayes: massively parallel Bayesian tree inference for the whole-genome era. *Mol Biol Evol.* 31(10):2553–2556.
- Attias H. 1999. Inferring parameters and structure of latent variable models by variational Bayes. In: *Proceedings of the Fifteenth Conference on Uncertainty in Artificial Intelligence, UAI'99*. San Francisco (CA): Morgan Kaufmann Publishers Inc. p. 21–30.
- Bouckaert R, Vaughan TG, Barido-Sottani J, Duchêne S, Fourment M, Gavryushkina A, Heled J, Jones G, Kühnert D, De Maio N, et al. 2019. BEAST 2.5: an advanced software platform for Bayesian evolutionary analysis. *PLOS Comput Biol.* 15(4):e1006650.
- Bragg JG, Potter S, Silva ACA, Hoskin CJ, Bai BY, Moritz C. 2018. Phylogenomics of a rapid radiation: the Australian rainbow skinks. *BMC Evol Biol.* 18(1):15.
- Chifman J, Kubatko L. 2014. Quartet inference from SNP data under the coalescent model. *Bioinformatics* 30(23):3317–3324.
- Degnan JH, Rosenberg NA. 2009. Gene tree discordance, phylogenetic inference and the multispecies coalescent. *Trends Ecol Evol.* 24(6):332–340.
- Drummond AJ, Rambaut A. 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol Biol.* 7(1):214.
- Felsenstein J. 1978. The number of evolutionary trees. *Syst Biol.* 27(1):27–33.
- Flouri T, Jiao X, Rannala B, Yang Z. 2018. Species tree inference with BPP using genomic sequences and the multispecies coalescent. *Mol Biol Evol.* 35(10):2585–2593.
- Fourment M, Darling AE. 2019. Evaluating probabilistic programming and fast variational Bayesian inference in phylogenetics. *PeerJ.* 7:e8272.
- Geyer CJ. 1991. Markov Chain Monte Carlo maximum likelihood. In: Keramidas EM, editor. *Computing Science and Statistics: Proceedings of the 23rd Symposium on the Interface*. Fairfax Station, Virginia: Interface. p. 156–163.
- Höhna S, Drummond AJ. 2012. Guided tree topology proposals for Bayesian phylogenetic inference. *Syst Biol.* 61(1):1–11.
- Huang S, Su X, Haselkorn R, Gornicki P. 2003. Evolution of switchgrass (*Panicum virgatum* L.) based on sequences of the nuclear gene encoding plastid acetyl-CoA carboxylase. *Plant Sci.* 164(1):43–49.
- Hudson RR. 2002. Generating samples under a Wright–Fisher neutral model of genetic variation. *Bioinformatics* 18(2):337–338.
- Huelsenbeck JP, Ronquist F, Nielsen R, Bollback JP. 2001. Bayesian inference of phylogeny and its impact on evolutionary biology. *Science* 294(5550):2310–2314.
- Huff CD, Xing J, Rogers AR, Witherspoon D, Jorde LB. 2010. Mobile elements reveal small population size in the ancient ancestors of *Homo sapiens*. *Proc Natl Acad Sci USA.* 107(5):2147–2152.
- Hugall AF, Foster R, Lee MSY. 2007. Calibration choice, rate smoothing, and the pattern of tetrapod diversification according to the long nuclear gene RAG-1. *Syst Biol.* 56(4):543–563.
- Jones G. 2017. Algorithmic improvements to species delimitation and phylogeny estimation under the multispecies coalescent. *J Math Biol.* 74(1–2):447–467.
- Jukes TH, Cantor CR. 1969. Evolution of protein molecules. In: *Mammalian protein metabolism*. New York: Academic Press. p. 21–132.
- Kuhner MK. 2009. Coalescent genealogy samplers: windows into population history. *Trends Ecol Evol.* 24(2):86–93.
- Kuhner MK, Felsenstein J. 1994. A simulation comparison of phylogeny algorithms under equal and unequal evolutionary rates. *Mol Biol Evol.* 11(3):459–468.
- Kupczok A, Schmidt HA, von Haeseler A. 2010. Accuracy of phylogeny reconstruction methods combining overlapping gene data sets. *Algorithms Mol Biol.* 5(1):37.
- Lakner C, Van Der Mark P, Huelsenbeck JP, Larget B, Ronquist F. 2008. Efficiency of Markov chain Monte Carlo tree proposals in Bayesian phylogenetics. *Syst Biol.* 57(1):86–103.
- Langergraber KE, Prüfer K, Rowney C, Boesch C, Crockford C, Fawcett K, Inoue E, Inoue-Muruyama M, Mitani JC, Muller MN, et al. 2012. Generation times in wild chimpanzees and gorillas suggest earlier divergence times in great ape and human evolution. *Proc Natl Acad Sci USA.* 109(39):15716–15721.
- Liu L, Yu L. 2011. Estimating species trees from unrooted gene trees. *Syst Biol.* 60(5):661–667.
- Liu L, Yu L, Edwards SV. 2010. A maximum pseudo-likelihood approach for estimating species trees under the coalescent model. *BMC Evol Biol.* 10(1):302.
- Mazza G, Menchetti M, Sluys R, Sola E, Riutort M, Tricarico E, Justine J-L, Caviglioli L, Mori E. 2016. First report of the land planarian *Diversibipalium multilineatum* (Makino & Shirasawa, 1983) (Platyhelminthes, Tricladida, Continenticola) in Europe. *Zootaxa* 4067(5):577–580.
- Mirarab S, Reaz R, Bayzid MS, Zimmermann T, Swenson MS, Warnow T. 2014. ASTRAL: genome-scale coalescent-based species tree estimation. *Bioinformatics* 30(17):i541–i548.

- Ogilvie HA, Bouckaert RR, Drummond AJ. 2017. StarBEAST2 brings faster species tree inference and accurate estimates of substitution rates. *Mol Biol Evol.* 34(8):2101–2114.
- Ogilvie HA, Heled J, Xie D, Drummond AJ. 2016. Computational performance and statistical accuracy of *BEAST and comparisons with other methods. *Syst Biol.* 65(3):381–396.
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018. Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Syst Biol.* 67(5):901–904.
- Rambaut A, Grass NC. 1997. Seq-Gen: an application for the Monte Carlo simulation of DNA sequence evolution along phylogenetic trees. *Bioinformatics* 13(3):235–238.
- Rannala B, Yang Z. 2017. Efficient Bayesian species tree inference under the multispecies coalescent. *Syst Biol.* 66(5):823–842.
- Robert C, Casella G. 2011. A short history of Markov Chain Monte Carlo: subjective recollections from incomplete data. *Stat Sci.* 26(1):102–115.
- Robinson DF, Foulds LR. 1981. Comparison of phylogenetic trees. *Math Biosci.* 53(1–2):131–147.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol.* 61(3):539–542.
- Sandve SR, Rudi H, Asp T, Rognli OA. 2008. Tracking the evolution of a cold stress associated gene family in cold tolerant grasses. *BMC Evol Biol.* 8(1):245.
- St. John K. 2017. Review paper: the shape of phylogenetic treespace. *Syst Biol.* 66(1):e83–e94.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30(9):1312–1313.
- Stensvold CR, Lebbad M, Clark CG. 2012. Last of the human protists: the phylogeny and genetic diversity of *Iodamoeba*. *Mol Biol Evol.* 29(1):39–42.
- Stunžėnas V, Petkevičiūtė R, Stanevičiūtė G. 2011. Phylogeny of *Sphaerium solidum* (Bivalvia) based on karyotype and sequences of 16S and ITS1 rDNA. *Central Eur J Biol.* 6(1):105–117.
- Sukumaran J, Holder MT. 2010. DendroPy: a Python library for phylogenetic computing. *Bioinformatics* 26(12):1569–1571.
- Tajima F. 1983. Evolutionary relationship of DNA sequences in finite populations. *Genetics* 105(2):437–460.
- Takahata N, Satta Y, Klein J. 1995. Divergence time and population size in the lineage leading to modern humans. *Theor Popul Biol.* 48(2):198–221.
- Wang Y, Nakhleh LK. 2018. Towards an accurate and efficient heuristic for species/gene tree co-estimation. *Bioinformatics* 34(17):i697–i705.
- Wen D, Nakhleh L. 2018. Coestimating reticulate phylogenies and gene trees from multilocus sequence data. *Syst Biol.* 67(3):439–457.
- Wen D, Yu Y, Nakhleh L. 2016. Bayesian inference of reticulate phylogenies under the multispecies network coalescent. *PLOS Genet.* 12(5):e1006006.
- Wen D, Yu Y, Zhu J, Nakhleh L. 2018. Inferring phylogenetic networks using PhyloNet. *Syst Biol.* 67(4):735–740.
- Yang Z. 2002. Likelihood and Bayes estimation of ancestral population sizes in hominoids using data from multiple loci. *Genetics* 162(4):1811–1823.
- Yang Z, Rannala B. 2014. Unguided species delimitation using DNA sequence data from multiple loci. *Mol Biol Evol.* 31(12):3125–3135.
- Yang Z, Rodríguez CE. 2013. Searching for efficient Markov chain Monte Carlo proposal kernels. *Proc Natl Acad Sci USA.* 110(48):19307–19312.
- Yu Y, Dong J, Liu KJ, Nakhleh L. 2014. Maximum likelihood inference of reticulate evolutionary histories. *Proc Natl Acad Sci USA.* 111(46):16448–16453.
- Zhang C, Huelsenbeck JP, Ronquist F. 2020. Using parsimony-guided tree proposals to accelerate convergence in Bayesian phylogenetic inference. *Syst Biol.* Advance Access published January 27, 2020, doi: 10.1093/sysbio/syaa002.
- Zhang C, Matsen FA. 2019. Variational Bayesian phylogenetic inference. In: *International Conference on Learning Representations*. [accessed 2019 Sep 15]. Available from: <https://openreview.net/forum?id=SJVmjjR9FX>.
- Zhang C, Ogilvie HA, Drummond AJ, Stadler T. 2018. Bayesian inference of species networks from multilocus sequence data. *Mol Biol Evol.* 35(2):504–517.
- Zhu J, Wen D, Yu Y, Meudt HM, Nakhleh L. 2018. Bayesian inference of phylogenetic networks from bi-allelic genetic markers. *PLOS Comput Biol.* 14(1):e1005932.