

Highlights

- *Cavilignum pratchettii* is a new Neogene angiosperm from eastern Tennessee, U.S.A.
- *Cavilignum* is based on four-chambered fossil endocarps with open germination pores.
- *Cavilignum* cannot be definitively classified within angiosperms.
- *Cavilignum* is the first extinct plant macrofossil genus from Gray Fossil Site.

A suite of fossil endocarps representing a new taxon from the early Pliocene Gray Fossil Site, Tennessee, U.S.A., is described as *Cavilignum pratchettii* gen. et sp. nov.

Cavilignum is represented by circular to oblong endocarps that have a truncate apex, a mucronate base, and a smooth outer surface. The endocarp wall has three layers. The outermost layer is of uncertain composition. The inner wall structure is fibrous, with an outer layer of radially-oriented fibers and an inner layer of circumferentially-oriented fibers. The endocarps typically have four chambers (locules) divided by thick septa; two narrow canals penetrate the septa from the apex to the base of each endocarp. The locules are open apically. Because no evidence of structures that may have sealed the chambers was found, *Cavilignum* is interpreted as having open germination pores. Apically-oriented, open germination pores most obviously suggest an affinity with the angiosperm family Symplocaceae, but endocarps of Symplocaceae differ from *Cavilignum* in several critical characteristics (e.g., wall histology, presence of a basal pit). While several other groups (e.g., Anacardiaceae, Cornales) produce drupaceous fruits with endocarps comparable in some characteristics to those of *Cavilignum*, none is structurally congruent enough to suggest an affinity with the new fossil genus. Thus, *Cavilignum* cannot be conclusively assigned to a known extinct or extant angiosperm genus or family. *Cavilignum* is the first extinct plant genus described from the Gray Fossil Site macroflora.

Keywords: angiosperm, fossil, endocarp, Gray Fossil Site, Neogene, Pliocene

1 *Cavilignum pratchettii* gen. et sp. nov., a novel type of fossil endocarp with open locules
2 from the Neogene Gray Fossil Site, Tennessee, USA

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4 Caroline Siegert^a and Elizabeth J. Hermsen^{a,b,c,*}

5

6 ^a Department of Environmental and Plant Biology, Ohio University, Athens, Ohio 45701.

7

8 ^b Paleontological Research Institution, 1259 Trumansburg Road, Ithaca, New York
9 14850.

10

11 ^c Plant Biology Section, School of Integrative Plant Science, Cornell University, Ithaca,
12 New York 14853.

13

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15 *Corresponding author: Elizabeth J. Hermsen, Paleontological Research Institution,
16 1259 Trumansburg Road, Ithaca, NY 14850. hermsen@priweb.org.

18 **1. Introduction**

19

20 The Neogene is a time of transition, when the relatively warm world of the early
21 to middle Miocene gave way to a world shaped by global cooling in the late Miocene to
22 Pliocene, eventually culminating in the widespread northern hemisphere glaciations of
23 the Pleistocene (e.g., Tiffney, 1985; Tiffney and Manchester, 2001; Zachos et al., 2001;
24 Milne, 2006). These changes helped to shape the flora of eastern North America, with
25 certain genera identified from Neogene macrofossil floras—such as *Paliurus* Mill. (Lott
26 et al., 2019), *Pterocarya* Kunth (McCartan et al., 1990), *Sargentodoxa* Rehder &
27 E.H.Wilson (Tiffney, 1993; McNair et al., 2019), and *Sinomenium* Diels (Liu and
28 Jacques, 2010)—disappearing from the region prior to the present day. Our
29 understanding of plant taxonomic richness and extinction in eastern North America from
30 the Neogene to the present is hampered by the sparseness of the known plant
31 macrofossil record (Fig. 1; Tiffney and Manchester, 2001; Corbett, 2004; Stults et al.,
32 2010; Stults and Axsmith, 2011a, 2015; Lott et al., 2019). The distribution of Neogene
33 floras in this region is also highly uneven, with the greatest concentration of material
34 across space and time occurring in the Gulf Coast states, in a band of the Neogene
35 formations extending from eastern Texas to the Florida panhandle. Among these floras,
36 Alum Bluff, Citronelle, and Hattiesburg are the best documented (Fig. 1; Berry, 1916a,
37 b, c; Corbett, 2004; Stults et al., 2010; Stults and Axsmith 2011a, b, 2015; Dockery and
38 Thompson, 2016; Lott et al., 2019; McNair et al., 2019). Outside of the Gulf Coast
39 region, the early Miocene Brandon Lignite flora of Vermont is the most thoroughly
40 studied (see Tiffney, 1994, and references therein; Haggard and Tiffney 1997; Tiffney et

41 al., 2018), whereas formal systematic treatments are lacking for many macrofossil taxa
42 (e.g., *Potentilla* L., *Pterocarya*, *Trapa* L.) preliminarily identified from some other
43 important floras (e.g., Brandywine, Pipe Creek Sinkhole: McCartan et al., 1990; Farlow
44 et al., 2001).

45 The Pliocene Gray Fossil Site (GFS), eastern Tennessee, U.S.A., is an isolated
46 Neogene paleobiota from the Appalachian region (Fig. 1) that provides data critical for
47 understanding the evolution of eastern North American vegetation during this time of
48 climatic and environmental transition. GFS is interpreted as an ancient sinkhole deposit
49 preserving a paleobiota that is especially rich in fossil vertebrates, but that also includes
50 abundant and diverse plant microfossil and macrofossil remains (e.g., Parmalee et al.,
51 2002; Wallace and Wang, 2004; Mead et al., 2012; Bourque and Schubert 2015; Ochoa
52 et al., 2016). The GFS macrofossil flora includes wood, fruits, seeds, and leaves. Over
53 thirty genera of plants are thought to be represented in the GFS macroflora (Mead et al.,
54 2012; Ochoa et al., 2016), although only a subset have been described (Gong et al.,
55 2010; Liu and Jacques, 2010; Brandon, 2013; Noll, 2013; Huang et al., 2014, 2015).
56 The described taxa have all been assigned to extant plant genera and show strongest
57 affinities to taxa from Laurasian continents, especially those found in the modern and
58 Cenozoic fossil floras of North America and Asia, as well as the Cenozoic flora of
59 Europe (Gong et al., 2010; Liu and Jacques, 2010; Brandon, 2013; Noll, 2013; Huang et
60 al., 2014, 2015). Angiosperms described from GFS fruit and seed fossils include extinct
61 species of Chinese moonseed (*Sinomenium macrocarpum* Liu & Jacques: Liu and
62 Jacques, 2010), bladdernut (*Staphylea levisemia* Huang et al.: Huang et al., 2015),

63 hickory (*Carya tennesseensis* Huang et al.: Huang et al., 2014), tupelo (*Nyssa* L., sp.
64 nov.: Noll, 2013), and grapes (*Vitis* L., 3 species: Gong et al., 2010).

65 In this study, we describe a new genus and species of angiosperm from GFS
66 represented by fossil endocarps. These endocarps are unusual in having four, or less
67 commonly three, open locules. While the endocarps share features with endocarps of
68 some modern flowering plant groups, most notably Symplocaceae Desf. (Ericales) and
69 certain Anacardiaceae R.Br. (Sapindales), they also differ in critical ways that exclude
70 them from these families. Thus, their affinities are unclear, and we interpret them as
71 representing an extinct Pliocene element of the flora.

72

73 **2. Methods**

74

75 *2.1. Context of the flora*

76

77 Gray Fossil Site was discovered during road construction in the year 2000 and
78 covers up to about 3 ha of land (Parmalee et al., 2002). Although initially macrofossil-
79 bearing sediments at the site were estimated to be late Miocene to early Pliocene in age
80 (Hemphillian, 4.5–7 million years old) (Paramlee et al. 2002; Wallace and Wang, 2004),
81 a more recent assessment by Samuels et al. (2018) has narrowed the age range of the
82 sediments to the early Pliocene (Blancan, 4.5–4.9 million years old). Many vertebrate
83 fossils have been uncovered at GFS, including a mixture of extinct animals, groups
84 extirpated from eastern North America but still present elsewhere, and genera still
85 native to the region; the fauna includes fish, amphibians, reptiles, birds, and mammals

86 such as bats (*Eptesicus* Rafinesque), tapirs (*Tapirus* Brünnich), extinct North American
87 camels (cf. *Megatylopus* Matthew & Cook), red pandas (*Pristinailurus bristoli* Wallace &
88 Wang), wolverines (*Gulo sudorus* Samuels et al.), and mastodons (e.g., Parmalee et al.,
89 2002; Wallace and Wang, 2004; Hulbert et al., 2009; Boardman and Schubert 2011;
90 Mead et al., 2012; Bourque and Schubert, 2015; Czaplewski, 2017; Jasinski and
91 Moscato, 2017; Doughty et al., 2018; Samuels et al., 2018, 2019).

92 The Pliocene climate at GFS is interpreted as somewhat warmer with less
93 seasonal temperature variation than the modern climate of Gray, Tennessee (Shunk et
94 al., 2006; Mead et al., 2012; Noll, 2013; Baumgartner, 2014; Simpson and Mickle,
95 2019). The site was inhabited by cold-intolerant vertebrates like *Alligator* Cuvier and
96 beaded lizards (Shunk et al., 2006; Mead et al., 2012), as well as tropical to warm
97 temperate plant genera such as *Taxodium* Rich. (Brandon, 2013; Noll, 2013),
98 *Corylopsis* Siebold & Zucc. (Ochoa et al., 2016; Quirk and Hermsen, unpublished data),
99 *Nyssa* (Noll, 2013), and *Sinomenium* (Liu and Jacques, 2010). Evidence from isotopes,
100 palynomorphs, wood, the carpopossil flora, and the fauna suggests that the environment
101 surrounding the GFS sinkhole was wooded (e.g., Wallace and Wang, 2004; Shunk et
102 al., 2006, 2009; DeSantis and Wallace, 2008; Ochoa et al., 2012), although
103 interpretations of tree density vary. DeSantis and Wallace (2008) interpreted the local
104 habitat as a “moderately dense” forest based on carbon isotope data from grazing
105 animals, whereas Ochoa et al. (2012, 2016) interpreted the environment as more open
106 (woodland to savanna) and subject to disturbance based on the composition of the
107 flora, the presence of large herbivores, and indicators of drought (wood with false
108 growth rings, sedimentology) and fire (occurrence of charcoal, perylene, and the

109 bamboo genus *Arundinaria* Michx.). The taxonomic composition and frequency
110 distribution of the palynomorphs recovered from the GFS sediments indicates that the
111 dominant tree types were oak (*Quercus* L.) and hickory (*Carya* Nutt.), with pine (*Pinus*
112 L.) also common (Wallace and Wang, 2004; Ochoa et al., 2012; Worobiec et al., 2013).

113

114 2.2. *Fossil specimens*

115

116 Excavations at the site are ongoing. Fruits and seeds are collected from GFS as
117 part of bulk sediment samples. To separate the fossils from the surrounding sediments,
118 GFS staff and volunteers wet-screen the sediments, after which the screened material
119 is dried; screened and dried fossils are then picked under dissecting microscopes.

120 Many specimens of the type of endocarp described for this study have been
121 recovered from GFS. Of these, many exhibit various degrees of compression; some
122 were also incomplete, brittle, cracked, or deteriorated at the time of study. The
123 description herein is based on 35 specimens that were complete or nearly complete,
124 although they may have experienced compression or cracking; three specimens that
125 were sectioned transversely by previous investigators; and two additional specimens
126 that appear to have broken longitudinally on their own (specimen numbers are given
127 below in the “Systematic Paleontology” section). All fossil material examined for this
128 study is permanently held at the East Tennessee State University Museum of Natural
129 History (ETMNH) collections, Gray Fossil Site, Gray, Tennessee, U.S.A.

130

131 2.3. *Imaging & measurements*

132

133 Fossil endocarps were studied using a Nikon SMZ1500 stereomicroscope with a
134 Nikon Digital Sight DS-Fi2 camera head and a Nikon SMZ18 stereomicroscope with a
135 Nikon DS-Ri2 camera. Images were captured and measurements taken using NIS-
136 Elements software (1991–2018, Laboratory Imaging); ImageJ was also used for
137 measurements (Rasband 1997–2018). A micro-computed tomography (micro-CT) scan
138 of three specimens (ETMNH 18139, 18148, and 19561) was performed at Ohio
139 University using a TriFoil Imaging eXplore CT 120 Small Animal X-Ray CT Scanner
140 (see associated Research Data). The specimens were placed in a row end-to-end and
141 scanned in a single session. Micro-CT scans were examined using MicroView software
142 (2011–2018, Parallax Innovations), and selected slices were saved as TIFF files to
143 illustrate the internal structure of the endocarps. Adobe Photoshop CC (1990–2018,
144 Adobe) was used to reorient (rotate or flip) saved slices and construct plates. The
145 brightness or levels on some photomicrographs were adjusted in Photoshop.

146

147 **3. Systematic Paleontology**

148

149 Class: Magnoliopsida (angiosperms)

150 Order: Unknown

151 Family: Unknown

152 Genus: *Cavilignum* Siegert & Hermsen, gen. nov.

153

154 Generic diagnosis: Endocarp lignitic, circular to oblong in lateral view, apex
155 truncate, base mucronate, outer surface smooth. Locules three to four, divided by septa
156 oriented perpendicularly to one another; locules open at the endocarp apex and closed
157 at the base. Two small, unbranched canals traversing the full length of the endocarp;
158 canals open apically on the longer septum to either side of the intersection between the
159 septa; canals open basally to either side of the basal mucro. Endocarp wall with an
160 outer layer of unknown composition and two inner fibrous layers; outer layer of fibers
161 radially oriented and inner layer of fibers circumferentially oriented.

162

163 Type: *Cavilignum pratchettii* Siegert & Hermsen, sp. nov. (Plates I, II)

164 Etymology: *Cavilignum* is a compound of the Latin *cavus* and *ignum* meaning
165 “hollow stone” (or hollow fruit pit), referring to the four hollow locules of the endocarps.

166

167 Species: *Cavilignum pratchettii* Siegert & Hermsen, sp. nov.

168 Species diagnosis: As for the genus.

169 Holotype: ETMNH 18149 (Plate I, 1, 9). Held at the East Tennessee State
170 University Museum of Natural History fossil collections, Gray Fossil Site, Gray,
171 Tennessee, U.S.A.

172 Paratypes: ETMNH 18128–18132, 18134–18146, 18148, 18150 (broken
173 longitudinally), 18151 (sectioned specimen), 18153 (sectioned specimen), 19566
174 (broken longitudinally), 19557 (sectioned specimen), 19560–19563, 19565, 19569,
175 19570, 19576, 19578, 19580, 22637–22640, 24514.

176 Type locality & stratigraphy: All specimens come from the Gray Fossil Site, Gray,
177 Tennessee, USA (36.386° N, 82.498° W). Sediments at GFS are considered Blancan
178 (early Pliocene), 4.5–4.9 Ma (Samuels et al., 2018).

179 Etymology: The species epithet is named after deceased author Sir Terry
180 Pratchett, who wrote numerous books and plays throughout his career that inspired
181 millions. He taught us how to understand the world and our place in it, which is really
182 what science is about. He told us that, in order to really see the world, you have to
183 “[o]pen your eyes and then open your eyes again” (Pratchett, 2003).

184 Description: The specimens represent lignitic endocarps that are circular to
185 oblong in lateral view, with a truncate apex, a mucronate base, and a smooth outer
186 surface (Plate I, 1–8). They are 9.4–15.5 mm in length and 8.0–14.2 mm in width,
187 length/width ratio 0.9–1.5. The endocarps are somewhat flattened in cross-section
188 (Plate I, 9–13; Plate II, 1, 2). We interpret the cross-sectional shape as naturally
189 elliptical and greater in width than depth; the elliptical shape is exaggerated in some
190 specimens that show evidence of compression (Plate II, 1). Specimens showing little or
191 no distortion from compression (Plate I, 9–13) measure 5.7–7.6 mm in depth; the ratio
192 of the longer radius (width) to the shorter radius (depth) is 1.5–1.9.

193 The endocarps typically have four locules (Plate I, 9–12), although sometimes
194 only three fully develop (Plate I, 11, 13; Plate II, 2). The apex of each locule is open,
195 and the openings of the locules are oval in shape (Plate I, 9–11). The locules measure
196 about up to about 2.1 mm x 3.9 mm along the radii. In longitudinal view, the locules are
197 oblong, rounded at the base, truncate apically, and extend nearly the full length of the
198 endocarp (Plate II, 3–6, 11).

199 The locules are partitioned by two septa oriented perpendicularly to each other
200 (Plate I, 9–13). The longer septum is 0.4–0.8 mm thick, whereas the shorter septum is
201 0.9–2.8 mm thick. Two narrow canals occur to either side of the intersection between
202 the septa, measuring about 0.2–0.9 mm wide (Plate I, 9–13; Plate II, 2, 7). They extend
203 the full length of the endocarp (Plate II, 10, 11). The canals are open at the apex of the
204 endocarp on the longer septal wall, to either side of the junction between the septa
205 (Plate I, 9–11). At the base, the openings of the canals can be observed on the outer
206 surface of the endocarp on either side of the basal mucro, where they sometimes
207 appear to be slit-like openings (Plate II, 8, 9). The canals may represent the former
208 positions of vascular bundles, although no vascular tissue was observed within them.

209 The endocarp wall is 1.3–2.7 mm thick. The outermost layer of the endocarp wall
210 has an indistinct composition (Plate II, 12, 14). The inner wall is fibrous, with two distinct
211 fiber orientations in cross section. The outer fibers are radially oriented (Plate I, 12, 13;
212 Plate II, 1, 2, 12–14). The inner fibers are oriented in a direction perpendicular to the
213 outer fibers, roughly circumferential or tangential (Plate II, 12, 14). Amber-colored
214 deposits are present in cracks in the wall of one specimen (Plate II, 14), which may
215 indicate the presence of resin.

216

217 **4. Discussion**

218

219 *4.1. Structural interpretation of Cavilignum*

220

221 *Cavilignum pratchettii* is represented by stony, typically four-chambered
222 structures, each chamber presumably representing a carpel. The chambers are open at
223 one end, here interpreted as the apical end (Plate I, 9–11). The apical openings of the
224 chambers appear to be intrinsic structural features of *Cavilignum* rather than, for
225 example, holes created by foraging animals, because they are consistently present and
226 uniform in appearance. No valves, opercula, or plugs that may once have sealed the
227 chambers have been discovered attached to or in association with the *Cavilignum*
228 structures; there are also no discernable dehiscence scars. Thus, the chambers were
229 likely persistently open rather than pores revealed upon dehiscence, as in a capsule.
230 Because the *Cavilignum pratchettii* structures are dry, stony, four-chambered, and open
231 but lacking a dehiscence mechanism, we are interpreting them as endocarps with open
232 germination pores; it is possible that the pores were once covered by additional layers
233 of the pericarp (e.g., a fleshy mesocarp) that are not preserved.

234

235 4.2. *Similar endocarps and potential affinities*

236

237 The presence of three (rare) to four locules, lidless germination pores, and a
238 fibrous endocarp wall are the most obvious characters that distinguish *Cavilignum* from
239 endocarps of other angiosperm taxa and that may provide clues to the affinities of the
240 genus. Specimens here assigned to *Cavilignum pratchettii* are likely the GFS plant
241 macrofossils previously reported to have affinities to Symplocaceae (Noll, 2011; Y. Liu,
242 pers. comm. in table 1 of Ochoa et al., 2016). Symplocaceae include two genera and
243 about 320 species of trees and shrubs present in tropical to warm temperate climates;

244 extant species are distributed in eastern North America to the Caribbean, Central
245 America, and northern South America in the Western Hemisphere and southern and
246 eastern Asia to eastern Australia in the Eastern Hemisphere (Nooteboom, 1975; Fritsch
247 et al., 2008, 2015; Liu and Qin, 2012). The vast majority of species in the family are
248 included within *Symplocos* Jacq., whereas only two species are assigned to the East
249 Asian genus *Cordyloblaste* Hensch. ex Moritzi (Fritsch et al., 2008). Plants in the family
250 are trees or shrubs that produce drupes with hard or sometimes papery endocarps (see,
251 e.g., Liu and Qin, 2012; Manchester and Fritsch, 2014; Tiffney et al., 2018). Endocarps
252 may have one to five chambers and characteristically have open apical germination
253 pores (Nooteboom, 1975; Mai and Martinetto, 2006; Fritsch et al., 2008; Liu and Qin,
254 2012). Fossil symplocaceous endocarps are widespread in Cenozoic deposits of the
255 Northern Hemisphere (e.g., Chandler, 1961a, b; Nooteboom, 1975; Mai & Martinetto,
256 2006; Manchester and Fritsch, 2014; Fritsch et al., 2015; Tiffney et al., 2018).

257 While endocarps of *Cavilignum* are similar to endocarps of members of the family
258 Symplocaceae in that they have locules that are open apically, they differ in other
259 characteristics. *Symplocos* endocarps are often characterized by the presence of an
260 apical depression in which the germination pores occur; the lip of this depression
261 sometimes exhibits a thickened apical collar or "bulge" (Mai and Martinetto, 2006).
262 *Cavilignum* endocarps show neither depression nor collar (Plate I, 1–11). The base of
263 *Symplocos* endocarps is rounded to tapered with a small central pit (Mai and Martinetto,
264 2006), whereas *Cavilignum* endocarps are mucronate, i.e., have a short, extended
265 basal point (Plate I, 1–8). *Symplocos* endocarps often have a central canal at the
266 junction of the septa separating the locules (Mai and Martinetto, 2006); *Cavilignum*, in

267 contrast, has dual canals that are offset from the intersection of the septa and that open
268 to either side of the basal point on the outer surface of the endocarp (Plate I, 9–13;
269 Plate II, 7, 8, 9). While the structure of the endocarp or drupe wall of *Symplocos* is
270 somewhat varied (Liu and Qin, 2012; Manchester and Fritsch, 2014; fig. S2 of Fritsch et
271 al., 2015; Tiffney et al., 2018), no illustrated sections of *Symplocos* show a fibrous wall
272 structure similar to that observed in *Cavilignum* (Plate II, 12–14).

273 *Cordyloblaste*, with the species *C. henschelii* Mortizi and *C. pendula* (Wight)
274 Alston (Fritsch et al. 2008), is sometimes included in *Symplocos* (Nooteboom, 1975; Wu
275 and Nooteboom, 1996). *Cordyloblaste* endocarps are similar to those of *Cavilignum* in
276 that they lack an apical bulge (Mai and Martinetto, 2006) and have two to five locules
277 (Tiffney et al., 2018). However, *Cordyloblaste* endocarps are circular in cross section
278 (see fig. S2b in Fritsch et al., 2015); the individual locules are small compared to the
279 overall diameter of the endocarp (see fig. S2b in Fritsch et al., 2015) and circular to oval
280 in transverse section (supporting information in Fritsch et al., 2015; Tiffney et al., 2018).
281 Furthermore, the endocarp wall of *Cordyloblaste* is “mottled” and penetrated by a
282 reticulum of “narrow tubes” (Manchester and Fritsch, 2014, p. 72; Fritsch et al., 2015,
283 appendix S3, p. 15; Tiffney et al., 2018, p. 189), quite distinct from the fibrous wall of
284 *Cavilignum* endocarps (Plate II, 12–14).

285 Another family known for drupaceous fruits with endocarps sharing some
286 similarities with *Cavilignum* is Anacardiaceae R.Br. Anacardiaceae are a diverse group
287 of woody plants found in tropical to temperate climates worldwide (Pell et al., 2011;
288 Weeks et al., 2014); they are well represented in the fossil record, including on the basis
289 of their endocarps (e.g., Chandler, 1961a, b; Tiffney et al., 1994; Herrera et al., 2012;

290 Rozefelds et al. 2015; Fu et al., 2017). Endocarps of Anacardiaceae can be
291 multiloculate (Pell et al., 2011; Herrera et al., 2018, 2019), like *Cavilignum*. As in
292 *Cavilignum*, anacardiaceous endocarps have fibrous walls, although the orientation of
293 the fibers in anacardiaceous endocarps has been described as “tortuous” (Herrera et
294 al., 2018, 2019). Germination pores in anacardiaceous endocarps, when present, are
295 typically sealed by a plug, valve, or other structure (Herrera et al., 2018); no such
296 structures have been found in the germination pores of *Cavilignum*. Furthermore, even
297 when the germination pores of anacardiaceous endocarps are located near the apical
298 end of the endocarp (as in, e.g., *Sclerocarya* Hochst. and *Choerospondias* B.L.Burtt &
299 A.W.Hill), their apertures are still somewhat lateral in orientation (see Herrera et al.,
300 2018). The germination pores of *Cavilignum* are truly apical with no lateral deflection
301 (Plate I, 9–11). Finally, endocarps of Anacardiaceae often have pitted, textured, or
302 sculptured surfaces and/or internal lacunae (Herrera et al., 2018, 2019), none of which
303 are observed in *Cavilignum* (Plate I, 1–8, 12, 13; Plate II, 1–5, 6, 10, 11).

304 Other endocarps comparable to those of *Cavilignum* are found in Cornales,
305 Humiriaceae A.Juss., and *Vitex* L. Cornalean endocarps are similar to *Cavilignum*
306 endocarps in that they can have several locules and a smooth surface; however,
307 cornalean endocarps open by germination valves (Atkinson et al., 2016, 2017;
308 Manchester and Collinson, 2019). Endocarps of Humiriaceae, which may have apical
309 foramina (depressions), have germination valves; some also have a lacunose wall that
310 appears spongiform in transverse section (Herrera et al. 2010, 2014). *Vitex*
311 (Verbenaceae J. St.-Hil.) has a four-chambered endocarp, although the endocarp is
312 strongly ovoid, has operculate germination pores, and has a single, central channel

313 for the vascular tissue (Godley, 1971). No other families of angiosperms produce
314 endocarps that are particularly comparable to those assigned to *Cavilignum*.

315

316 *4.3. Conclusions*

317

318 The combination of smooth outer surface, four (sometimes three) locules,
319 truncate apex, basal mucro, open and apical germination pores, two small canals
320 penetrating the septa that are open apically and basally, and fibrous wall histology
321 distinguish the new genus *Cavilignum* from endocarps of other angiosperm taxa, both
322 living and extinct. While the locule number and open, apical germination pores are
323 similar to those of members of Symplocaceae, the fibrous wall structure, two small
324 canals that open to the outside of the endocarp basally, and the presence of a basal
325 mucro rather than a basal pit preclude assignment to that family. Endocarps from other
326 families investigated also show critical differences from *Cavilignum*. Thus, the affinities
327 of the endocarps remain unresolved. Angiosperm taxa previously described from Gray
328 Fossil Site have all been assigned to extant genera and support a biogeographic
329 interpretation of a Pliocene flora with strong connections to fossil and modern Eurasian
330 and North American floras. *Cavilignum* is the first report of a new genus of angiosperms
331 from the Gray Fossil Site macroflora, and it suggests the presence of an extinct
332 Pliocene element in the flora.

333

334 **5. Acknowledgments**

335

336 We thank L. Witmer and R. Ridgley (Ohio University) for micro-CT scanning of
337 specimens, Z. Quirk for assistance in arranging the micro-CT scanning, S. Welker for
338 assistance in initial characterization of the fossils, and M.A. Gandolfo (Cornell
339 University) for access to the SMZ18 stereomicroscope and attached camera. Thank you
340 also to B.H. Tiffney for providing information about endocarps of Symplocaceae from
341 the Brandon Lignite flora, and to M. DeVore and other colleagues who provided helpful
342 discussion and insights related to this project. Thank you to Gray Fossil Site for
343 providing the loan of specimens used in this study. Funding for this project was provided
344 by Ohio University and the National Science Foundation [DEB-1829376 to EJH].

345

346 Declarations of interest: none.

347

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659 **Plate captions**

660

661 Figure 1. Locations of major Neogene macrofossil floras in eastern North America
662 discussed in the text. For a review of Miocene localities, see Lott et al. (2019). **Miocene**:
663 1. Brandon Lignite, early Miocene, Vermont (Tiffney, 1994); 2. Alum Bluff, middle
664 Miocene, Florida (Berry, 1916a; Corbett, 2004; Jarzen et al., 2010; Lott et al., 2019); 3.
665 Hattiesburg, middle Miocene, Mississippi (McNair et al., 2019); 4. Brandywine, late
666 Miocene, Maryland (McCartan et al. 1990). **Pliocene**: 5. Gray Fossil Site, early
667 Pliocene, Tennessee; 6. Pipe Creek Sinkhole, early Pliocene, Indiana (Farlow et al.,
668 2001); 7. Citronelle, late Pliocene, Alabama and Florida (Berry, 1916b; Stults et al.,
669 2010; Stults and Axsmith, 2011a, b; 2015). Point map created with SimpleMapp
670 (Shorthouse, 2010).

671

672 Plate I. *Cavilignum pratchettii* C. Siegert & Hermsen, gen. et sp. nov. All scale bars = 2
673 mm.

674 1–8. Endocarps in lateral view, showing variation in size, shape, and prominence of the
675 basal mucro. Longitudinal cracks or splits visible on some specimens are interpreted as
676 artifacts rather than intrinsic structural features. 1. ETMNH 18149 (holotype). 2. ETMNH
677 18132. 3. ETMNH 18139 4. ETMNH 18128. 5. ETMNH 19562. 6. ETMNH 18143. 7.
678 ETMNH 18142. 8. ETMNH 18148.

679 9–11. Apical view of endocarps, showing open locules. 9. ETMNH 18149 (holotype). 10.
680 ETMNH 18151. 11. ETMNH 19561.

681 12–13. Transverse sections of endocarps, showing locules, fibrous wall structure, and
682 central canals. 12. ETMNH 18151. 13. ETMNH 18153.

683

684 Plate II. *Cavilignum pratchettii* C. Siegert & Hermsen, gen. et sp. nov. Structural details
685 of the endocarps. Figures 1–5, 11, are virtual slices from micro-CT scans. Scale bars: 6,
686 7, 12–14 = 1 mm; 8–10 = 2 mm.

687 1. Transverse section of endocarp showing collapsed locules. ETMNH 18148.

688 2. Transverse section of endocarp showing three well-developed locules (A, B, C), and
689 a central canal (Ca, arrow); second canal obscured by crack in septum between locules
690 A and B. ETMNH 19561.

691 3. Longitudinal section of endocarp showing two oblong locules with portions of the
692 collapsed septa near their apices (upper two locules in 1). ETMNH 18148.

693 4. Longitudinal section of endocarp showing two locules corresponding to A and B in 2.
694 ETMNH 19561.

695 5. Longitudinal section of endocarp perpendicular to section in 4, showing two locules
696 corresponding to B and C in 2. ETMNH 19561.

697 6. Endocarp, broken longitudinally, showing two locules. ETMNH 19566.

698 7. Transverse section of an endocarp at the junction of the septa, showing two canals.
699 ETMNH 18151.

700 8. Base of sectioned endocarp, showing two canals opening to the outside. ETMNH
701 18151.

702 9. Base of sectioned endocarp, showing two canals opening to the outside. ETMNH
703 19557.

704 10. Endocarp, broken longitudinally, showing the length of one of the central canals.

705 ETMNH 18150.

706 11. Longitudinal section of endocarp (perpendicular to section in 4, parallel to section in

707 5) showing one locule (corresponding to B in 2) and one central canal (corresponding to

708 canal in 2). ETMNH 19561.

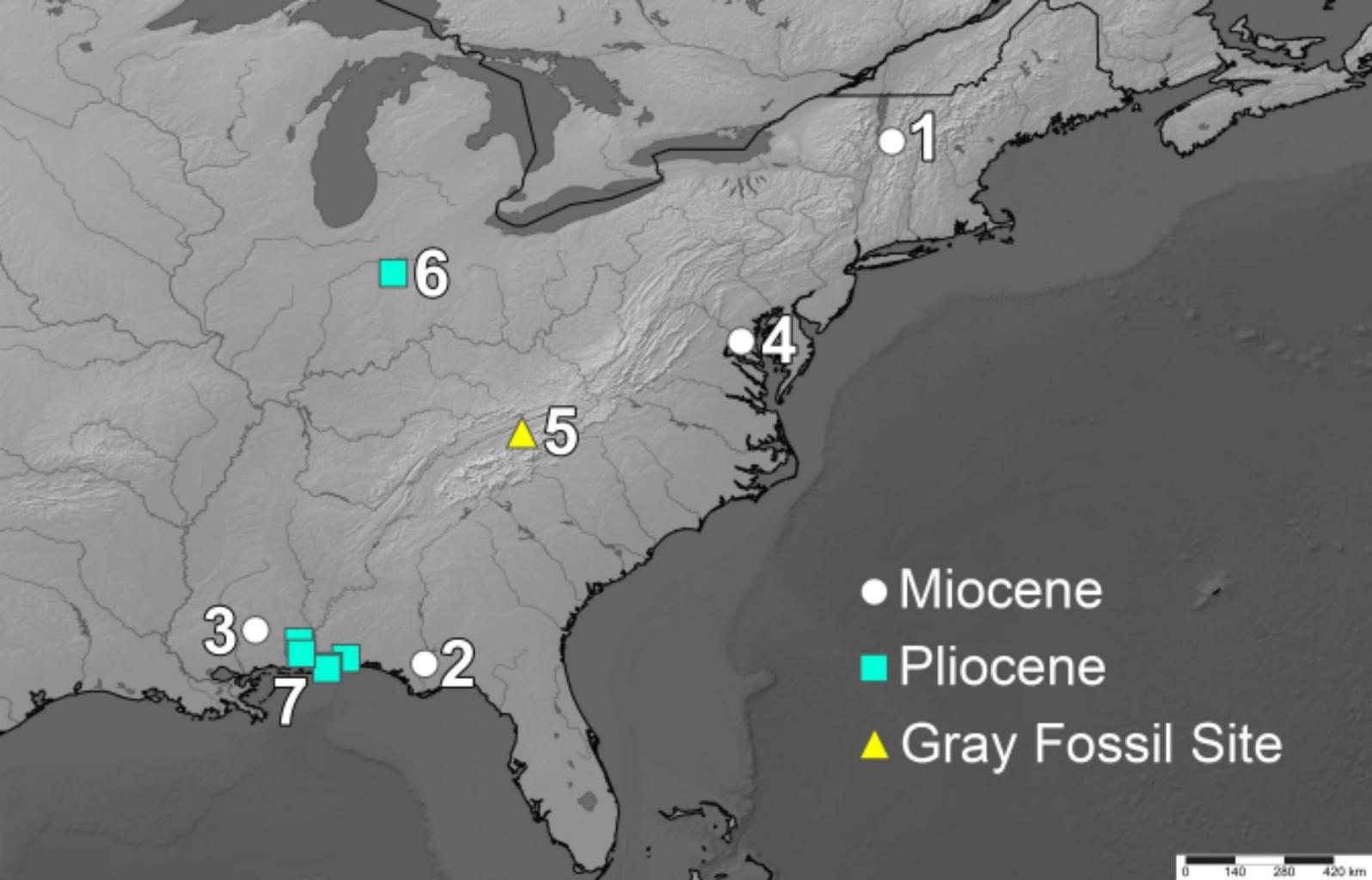
709 12. Transverse section of an endocarp showing the fibrous endocarp wall. ETMNH

710 18151.

711 13. Endocarp, broken longitudinally, showing fibrous wall. ETMNH 18150.

712 14. Transverse section showing cracks in the endocarp wall with amber-colored

713 substance. ETMNH 18151.



● Miocene

■ Pliocene

▲ Gray Fossil Site



