

# Predicting Rearrangement-Competent Terpenoid Oxidation Levels

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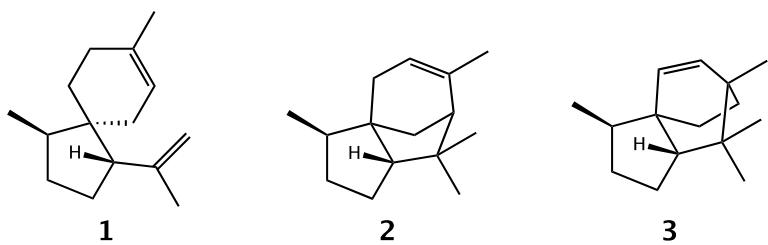
## Abstract

Results of density functional calculations on rearrangements of potential biosynthetic precursors to the sesquiterpenoid illisimonin A reveal that only some possible precursors, those with certain specific oxidation patterns, are rearrangement-competent.

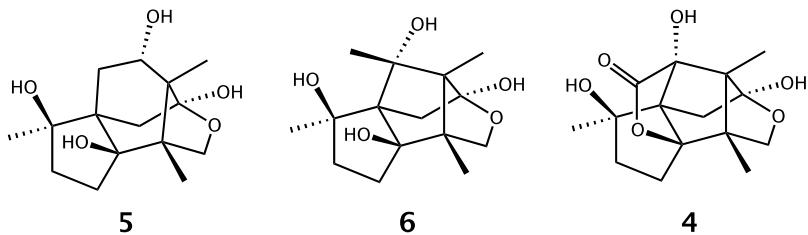
## Introduction

The diversity of Nature's library of terpenoid natural products arises, for the most part, from two sources. First, acyclic, achiral metabolic intermediates—oligoisoprenyl diphosphates—are converted into terpenes, which are often polycyclic and rich with stereogenic centers (Figure 1a), by terpene synthase-mediated carbocation cyclization/rearrangement reactions.<sup>1</sup> Hundreds of possible hydrocarbon skeletons are known for sesquiterpenes, for example. Second, heteroatoms are added—most often oxygen atoms—by enzymes such as cytochrome P450s, to produce terpenoids (Figure 1b).<sup>2</sup> In some cases, oxidation is accompanied by further rearrangement.<sup>3</sup> An example of a skeletal rearrangement that may occur after oxidation is discussed here.

(a) representative terpenes



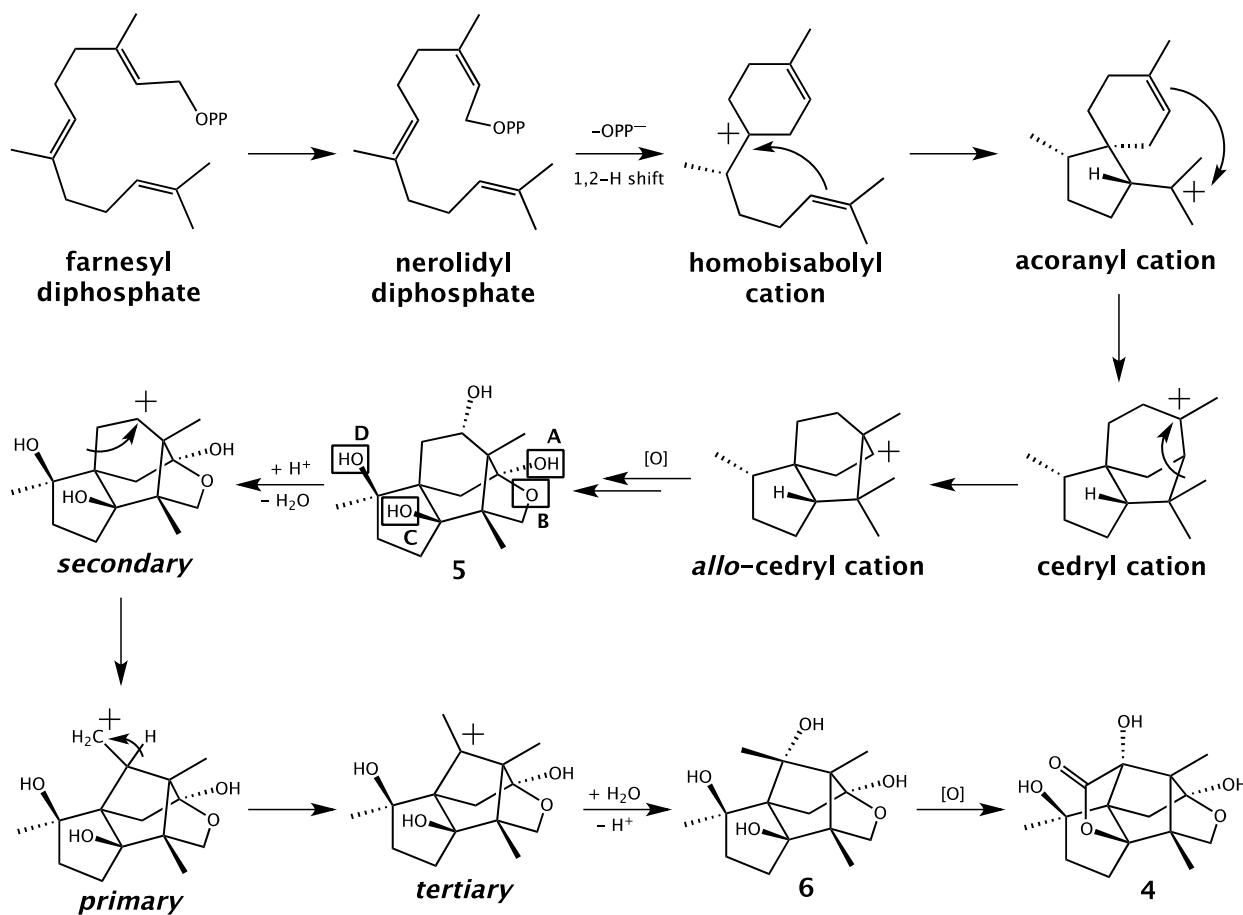
(b) representative terpenoids



**Figure 1.** Representative terpene and terpenoid structures, here sesquiterpenes and sesquiterpenoids: acorene (**1**), cedrene (**2**) and *allo*-cedrane (**3**, also known as khusiene).<sup>5</sup>

The authors of the paper describing the isolation of illisimonin A (**4**, Figure 1) proposed that a skeletal rearrangement occurs from a highly oxidized form of *allo*-cedrane, **5**→**6** (Scheme 1).<sup>4</sup> We report the results of density functional theory (DFT) computations used to probe the feasibility of rearrangement at this oxidation level and others.<sup>6</sup> These results indicate that rearrangement from a less (but still) oxidized structure is accompanied by a lower overall barrier. We hope that our approach will find utility in predicting rearrangement-competent oxidation levels for other terpenoids, which could facilitate both biosynthetic experiments and total synthesis efforts.<sup>7</sup>

**Scheme 1.** Rearrangements proposed in ref. 4 (stereochemistry and nerolidyl diphosphate added).



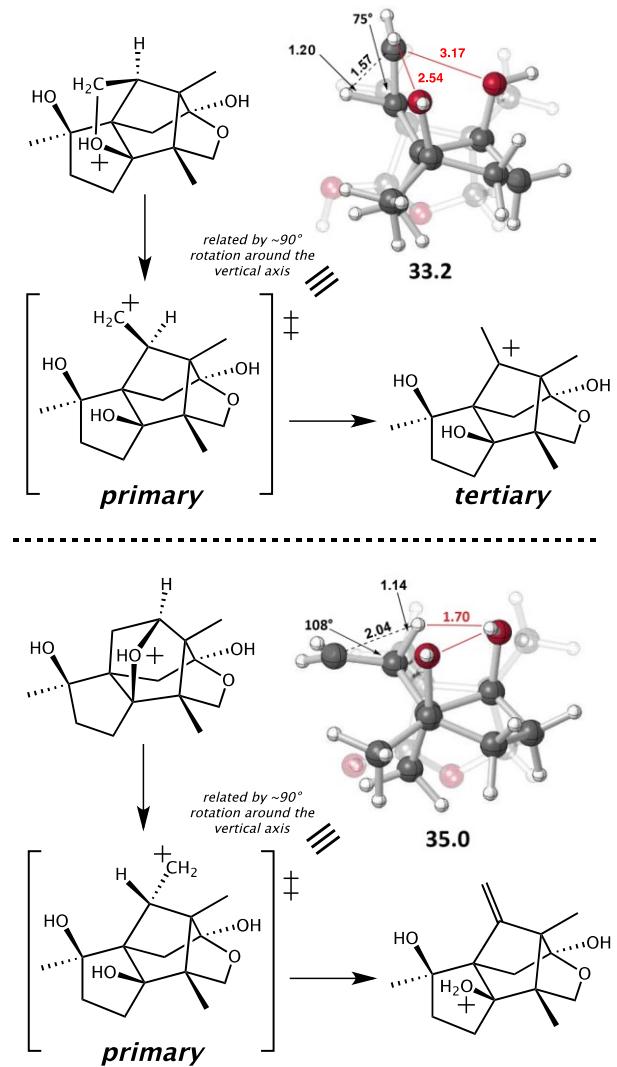
## Methods

Computations were performed at the mPW1PW91/6-31+G(d,p) level of theory, a level of theory used previously to model many carbocation rearrangements,<sup>6,8</sup> using *Gaussian09*.<sup>9</sup> All minima and transition state structures (TSSs) were confirmed by frequency analysis, and intrinsic reaction coordinate (IRC) calculations were performed on unconstrained transition state structures to confirm the minima to which they are connected.<sup>10</sup> All energies shown are free energies at room temperature, except for those in IRC plots, which are electronic energies. Three-dimensional molecular images were generated with *CylView*.<sup>11</sup>

## Results and Discussion

### *The previously proposed rearrangement*

The secondary→primary→tertiary carbocation rearrangement mechanism in Scheme 1 was the focus of our attention, as it is exceedingly unlikely that a primary carbocation is a minimum.<sup>12</sup> For the originally proposed system, two epimers of the primary carbocation are possible. Both were found to be transition state structures (TSSs), not minima (Figure 2). In one case (Figure 2, top), the core H–C–C substructure (top left in each ball-and-stick image) is partly bridged, while in the other (Figure 2, bottom) an intramolecular CH–O interaction<sup>13</sup> provides an alternative to bridging. These modes of delocalization are representative of the products connected to each TSS: bridging leads to a 1,2-hydride shift to form the tertiary carbocation originally proposed as an intermediate (Figure 2, top), while the CH–O interaction presages an intramolecular proton transfer to form a C=C double bond. In neither case, however, is a simple secondary carbocation (Scheme 1) found to connect to the TSS in the reactant direction.<sup>6,14</sup> Instead, oxygen-bridged structures are found – in one case (Figure 2, top) bridging to the site of the putative primary carbocation and in the other case (Figure 2, bottom) bridging to the site of the putative secondary carbocation connected to the TSS by a 1,2-alkyl shift (see the Supporting Information for IRC plots).<sup>15</sup> Thus, for neither configuration is the primary carbocation a minimum, nor is it connected to the previously proposed secondary and tertiary carbocations, i.e., the two proposed steps are not simply merged into a concerted process as has been observed for many other carbocation rearrangements.<sup>6,12,14</sup> In any case, the barriers for rearrangement associated with the TSSs in Figure 2 are predicted to be too high to be relevant to biology without considerable intervention (on the order of 10 kcal/mol barrier lowering),<sup>16</sup> so other oxidation levels were examined.

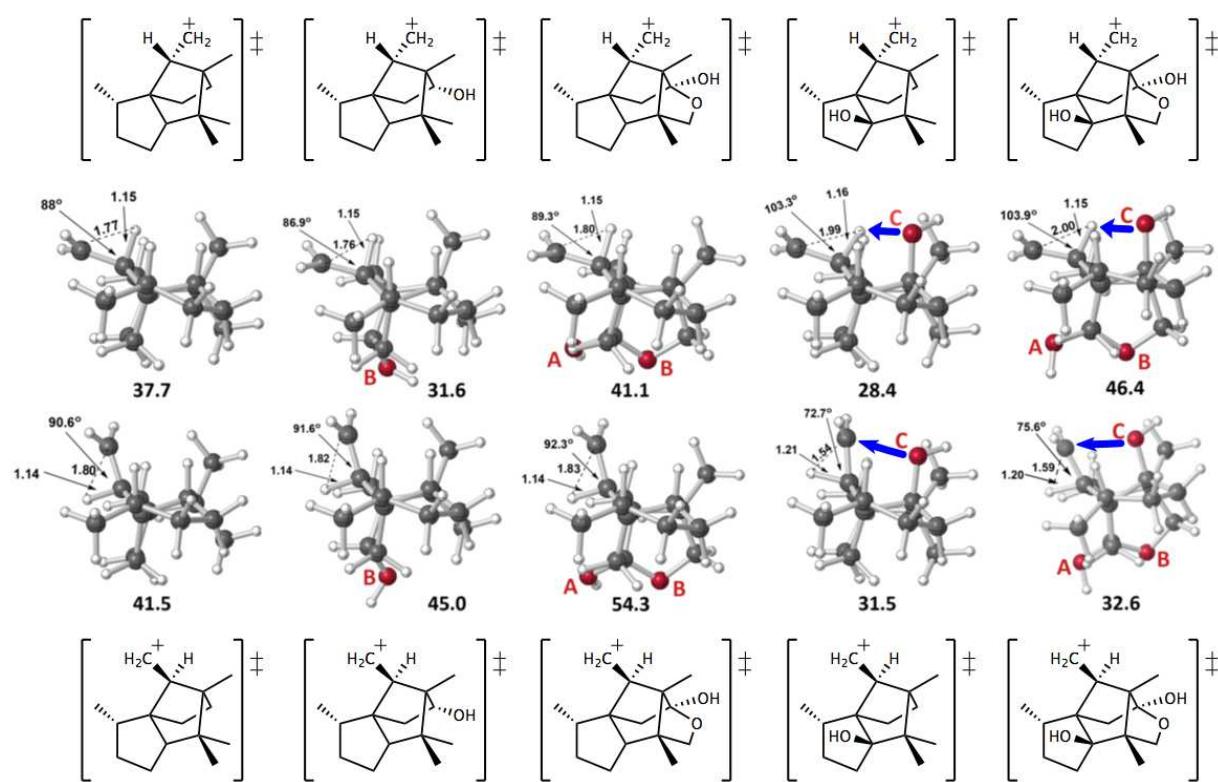


**Figure 2.** Rearrangements found for the fully substituted system (oxygens at positions A, B, C and D are shown), i.e., cations derived from **5**. Ball-and-stick images of the computed transition state structures (mPW1PW91/6-31+G(d,p)) resembling primary carbocations for the two epimers. Selected distances are shown in Å. Predicted rearrangement barriers ( $\Delta G^\ddagger$ ) in kcal/mol are listed below each transition state structure.

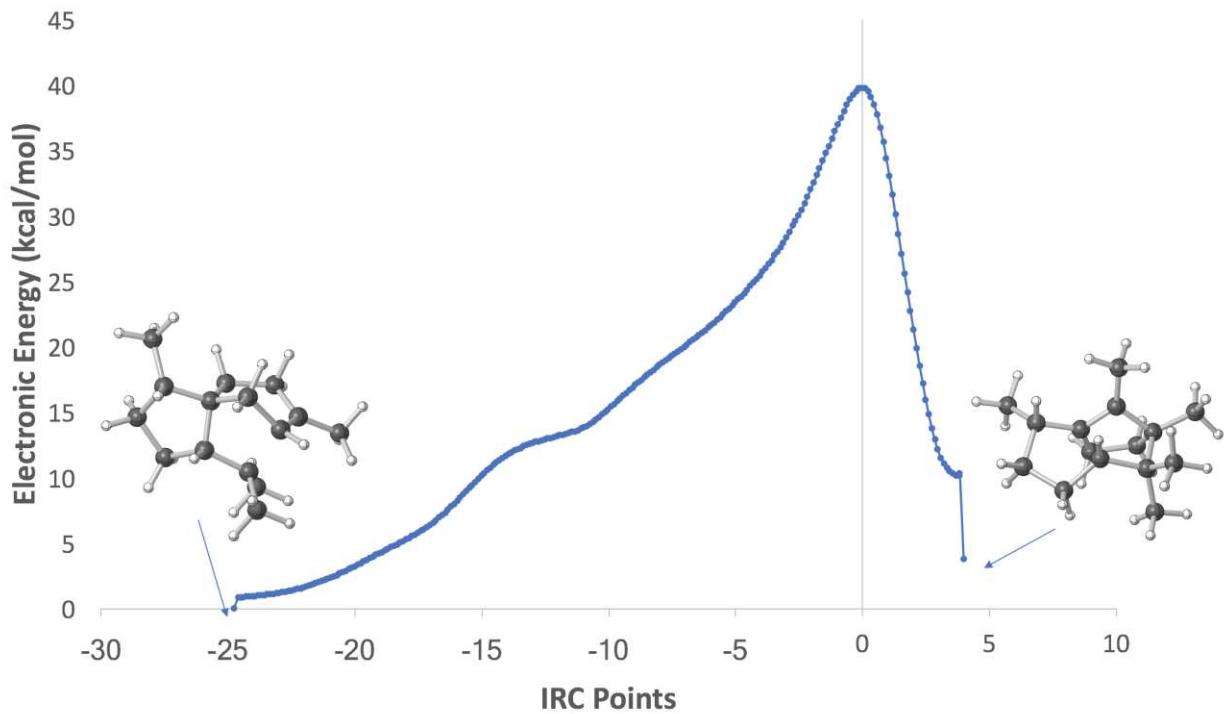
#### *The hydrocarbon case*

Carbon skeletons of terpenoids are usually constructed at the hydrocarbon stage, i.e., through the action of synthases/cyclases that produce terpenes,<sup>1,3</sup> thus we looked for an energetically viable pathway that involved non-oxygenated carbocations preceding **5** in Scheme 1. Again, we began by locating primary carbocation-like structures for two epimers at the

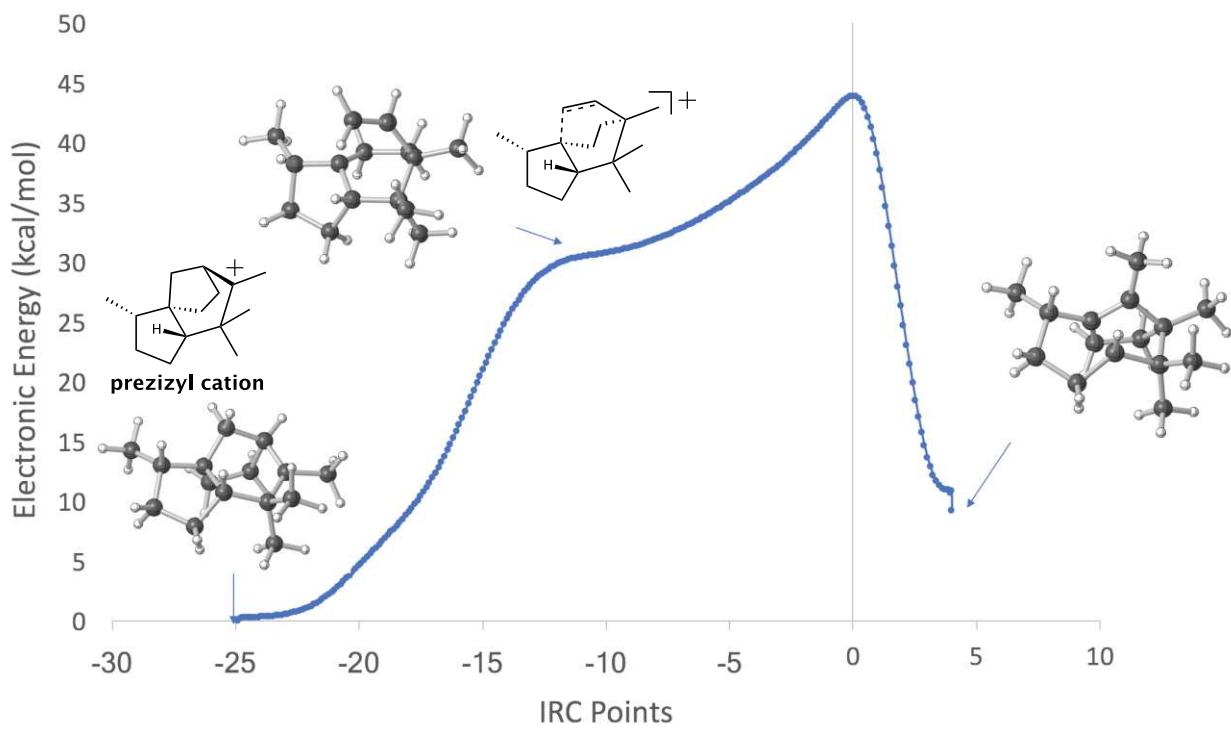
stereogenic center next to the site of the putative primary carbocation. In both cases, primary carbocation-like TSSs were again found (Figure 3, first column) and again, these were associated with high rearrangement barriers (here, approximately 40 kcal/mol). Note that these TSSs lack the possibility of intramolecular CH–O or O–C<sup>+</sup> interactions. One of these TSSs is connected to a cedryl cation (an epimer of that shown in Scheme 1) and the sans-oxygen version of the originally proposed tertiary carbocation (Figure 4). The other TSS is connected to a prezizyl cation<sup>17</sup> and the sans-oxygen version of the originally proposed tertiary carbocation (Figure 5).<sup>18</sup>



**Figure 3.** Representative transition state structures with various carbons oxidized (see SI for others), oriented to show <sup>13</sup>C–C–H substructures (top left of each ball-and-stick image) side-on to facilitate comparisons. Predicted activation barriers ( $\Delta G^\ddagger$ ) are shown below each in kcal/mol. Selected distances are shown in Å. Key CH–O and O–C<sup>+</sup> interactions are highlighted in blue.



**Figure 4.** IRC plot for rearrangement of cedryl cation epimer.



**Figure 5.** IRC plot for rearrangement of prezizyl cation.

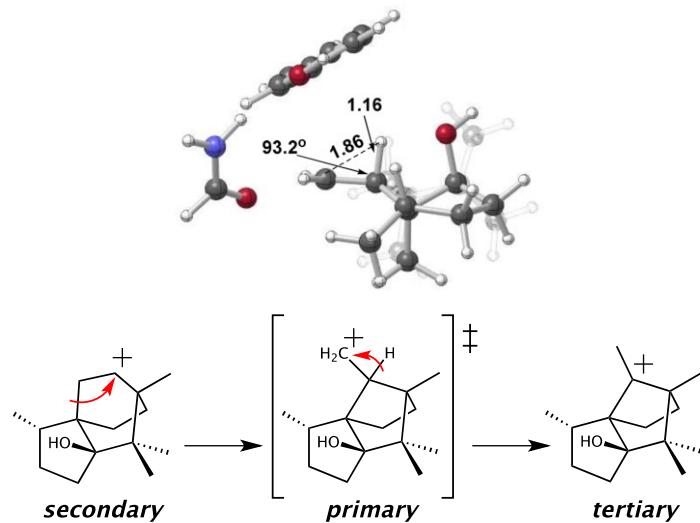
### *Varying the oxidation level*

Carbocations with a variety of alternative oxidation levels were explored in hopes that one or more would allow for an energetically viable rearrangement of the carbon skeleton. Oxygens were added at positions A, B and C (as labeled in Scheme 1), in various combinations. All TSSs for the rearrangements examined (Figure 3) contain a hyperconjugated primary carbocation center (some on the verge of H-bridging, e.g., two bottom right structures in Figure 3). However, these TSSs connect minima with a variety of polycyclic skeletons (see SI for details). For systems without an oxygen at position C, much higher barriers are predicted for migration of hydrogen to the bottom face (as drawn in Figure 3) of the carbocation center,<sup>19</sup> and the difference in predicted barriers increases as more oxygens are added. While it is difficult to pin down the origins of these barrier differences, since different types of reactant minima are connected to very similar TSSs, it is observed that in the preferred TSSs, the C–H bond  $\alpha$  to the primary carbocation center is slightly longer and its hydrogen is slightly closer to the  $\text{CH}_2$  group, i.e., it is closer to bridging. Having an OH group at position C (Figure 3, column 4) leads to the lowest barrier,  $\sim 28$  kcal/mol, likely due in part to the presence of a favorable internal CH–O interaction between the hyperconjugated H and the OH group ( $\sim 1.7$  Å; note that the epimeric TSS also has a favorable interaction, in that case between the carbocation center and an oxygen lone pair; see SI for additional details on these interactions). This TSS leads to an epimer (at the spiro center) of an acorenyl cation as reactant and an alkene resulting from intramolecular proton transfer (analogous to the reaction shown at the bottom of Figure 2) as product. Structures with an OH at position C and additional OH groups did not lead to lower barriers, likely the result of differences in strain and additional avenues for selective reactant stabilization (e.g., Figure 2).

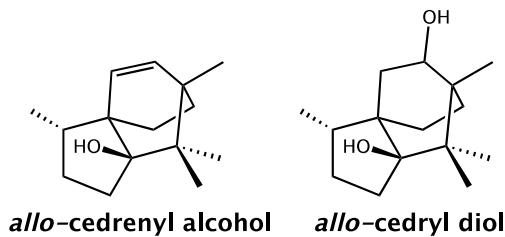
### *Theozymes*

In hopes that noncovalent interactions with an enzyme active site might lower the rearrangement barrier into a reasonable range ( $\sim 20$  kcal/mol),<sup>16</sup> we undertook theozyme calculations.<sup>20</sup> In these, structures were reoptimized in the presence of various combinations of benzene, indole, phenol and formamide – mimics of sidechains of phenylalanine, tryptophan, tyrosine and asparagine/glutamine (or backbone amides), respectively (see SI for details), on the

system from Figure 3 with the lowest barrier (Figure 6, bottom). In many cases, alkylation of the residue models by the carbocations occurred, so constraints were added during optimization to mimic the restrictions that could be imposed by an enzyme active site.<sup>21</sup> The system for which the lowest barrier was found is shown at the top of Figure 6: formamide/phenol with the primary carbocation center constrained to be 2.92 Å from one *ortho* carbon of the phenol (corresponding to a reasonable carbocation–π distance).<sup>22</sup> The predicted rearrangement barrier for this system is only 12.6 kcal/mol (and the reaction is predicted to be exergonic by ~9 kcal/mol) and the reactant in this case is a hydroxyl-bearing *allo*-cedryl cation (Figure 6). Carbocation–π interactions<sup>22</sup> between the phenol and both the primary carbocation center and the hyperconjugated hydrogen, as well as a CH–O interaction<sup>13</sup> between one CH at the primary carbocation center and the amide oxygen (~2 Å) conspire to provide selective stabilization to the TSS, for which these groups bear the largest positive charge (NBO charges on H: +0.43 in TSS, +0.27 in reactant; see SI for additional details). While this barrier is derived from a theozyme with constraints, and while no enzyme for this reaction has actually been isolated, the arrangement shown is not unusual.<sup>23</sup> Theozymes may also lower the rearrangement barrier into the biologically reasonable range for other systems in Figure 3, but the system described here has the advantage of starting with the lowest inherent rearrangement barrier (by at least 3 kcal/mol; Figure 3). *Our proof-of-principle results show that rearrangement is at least possible and lead us to propose that one of the compounds shown in Figure 7 is likely the biosynthetic precursor to ilisimonin A.* Related oxidized *allo*-cedrane have been described previously.<sup>4b,24</sup>



**Figure 6.** Transition state structure for the migration shown in the presence of a formamide/phenol theozyme. Selected distances are shown in Å.



**Figure 7.** Proposed, rearrangement-competent, biosynthetic precursors to illisimonin A.

## Conclusions

Our computational study has shed light on several aspects of illisimonin A biosynthesis: (1) Primary carbocations are almost certainly not involved as minima. (2) Rearrangement at the oxidation level of **5** would require an unreasonably large degree of enzymatic intervention. (3) Rearrangement at the hydrocarbon level would as well. (4) Some structures with intermediate oxidation levels have lower rearrangement barriers. (5) Specifically oriented noncovalent interactions can lower some barrier for these structures into the biologically reasonable range. We hope that an illisimonin A-producing enzyme will be isolated and characterized so that these predictions can be tested. We also hope that the approach described herein will find utility in predicting rearrangement-capable oxidation levels for structures involved in the biosynthesis (or synthesis) of other natural products.

## ASSOCIATED CONTENT

### Supporting Information

Additional details on computations, coordinates and energies for computed structures (PDF)

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### Notes

The authors declare no competing financial interest.

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## TOC graphic

