

**Adhesion and Running Speed of a Tropical Arboreal Ant
(*Cephalotes atratus*) on Rough, Narrow, and Inclined
Substrates**

Journal:	<i>Integrative and Comparative Biology</i>
Manuscript ID	ICB-2020-0055.R1
Manuscript Type:	Symposium article
Date Submitted by the Author:	03-Jun-2020
Complete List of Authors:	Stark, Alyssa; Villanova University, Biology Yanoviak, Stephen; University of Louisville, Department of Biology
Keywords:	performance, Formicidae, Panama, locomotion, behavior

SCHOLARONE™
Manuscripts

Adhesion and Running Speed of a Tropical Arboreal Ant (*Cephalotes atratus*) on Rough, Narrow, and Inclined Substrates

Alyssa Y. Stark^{1,*} and Stephen P. Yanoviak^{2,3}

¹Department of Biology, Villanova University, 800 E. Lancaster Ave., Villanova, PA 19085 USA

²Department of Biology, University of Louisville, 139 Life Sciences Building, Louisville, KY 40292 USA

³ Smithsonian Tropical Research Institute, Balboa, Republic of Panama

*Author for correspondence: alyssa.stark@villanova.edu; (610) 519-4838

Keywords: performance, Formicidae, Panama, locomotion, behavior

Running Title: Arboreal Ant Adhesion and Locomotion

Word Count: 4,765

21 Abstract

22 Arboreal ants must navigate variably sized and inclined linear structures across a range of
23 substrate roughness when foraging tens of meters above the ground. To achieve this, arboreal
24 ants use specialized adhesive pads and claws to maintain effective attachment to canopy
25 substrates. Here, we explored the effect of substrate structure, including small and large-scale
26 substrate roughness, substrate diameter, and substrate orientation (inclination), on adhesion and
27 running speed of workers of one common, intermediately-sized, arboreal ant species. Normal
28 (orthogonal) and shear (parallel) adhesive performance varied on sandpaper and natural leaf
29 substrates, particularly at small size scales, but running speed on these substrates remained
30 relatively constant. Running speed also varied minimally when running up and down inclined
31 substrates, except when the substrate was positioned completely vertical. On vertical surfaces,
32 ants ran significantly faster down than up. Ant running speed was slower on relatively narrow
33 substrates. The results of this study show that variation in the physical properties of tree surfaces
34 differentially affects arboreal ant adhesive and locomotor performance. Specifically, locomotor
35 performance was much more robust to surface roughness than was adhesive performance. The
36 results provide a basis for understanding how performance correlates of functional morphology
37 contribute to determining local ant distributions and foraging decisions in the tropical rainforest
38 canopy.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Introduction

An animal's ability to locomote over the range of conditions that occur within its local environment ultimately affects its survival (Arnold 1983; Vogel 1988; Bennett and Huey 1990; Garland Jr and Losos 1994; Ricklefs and Miles 1994; Alexander 2003; Irschick and Higham 2016; Biewener and Patek 2018). For cursorial animals, substrate characteristics are strong selective pressures; camels and mountain goats have limb and biomechanical modifications that promote efficient locomotion on sand and rock faces (respectively), but not vice versa (Dagg 1974; Lewinson and Stefanyshyn 2016). Such traits facilitate foraging and escape from predation, but are only effective across a range of substrate characteristics. Thus, selection should favor biased use of pathways that minimize costs relative to benefits (e.g., Schoener 1979; Stephens and Krebs 1986; Ydenberg et al. 1994; Yunker et al. 2002). Potentially important costs include metabolic energy expenditure, frequency of injury, and duration of exposure to hazards (Stephens and Krebs 1986).

The biased use of pathways to minimize costs is particularly relevant for small organisms like ants, where local habitat complexity rapidly increases as body size decreases (Kaspari and Weiser 1999; Yanoviak and Kaspari 2000; Grevé et al. 2019). The duration of foraging trips and the frequency of worker loss are likely to increase in more complex habitats (Fewell et al. 1996). Longer foraging trips result in reduced food intake rate (Carroll and Janzen 1973; Fewell 1988) and lost workers are costly to ant colonies as lost biomass (Wilson 1968). Thus, ants and other small foragers commonly choose paths of least resistance (i.e., shorter, less complex routes; Fewell 1988; Torres-Contreras and Vasquez 2004; Devigne and Detrain 2006; Farji-Brener et al. 2007; Clay et al. 2010).

Worker ants that forage primarily in tree crowns must maintain contact with three-dimensional substrates to avoid falling. Indeed, unique behaviors among small wingless arboreal organisms such as gliding and swimming suggest that falling is costly and climbing should be optimized (Cartmill 1985; Yanoviak et al. 2005; Yanoviak et al. 2009; Yanoviak and Frederick 2014; Yanoviak et al. 2015). Arboreal ants generally use compliant tarsal pads coated with an adhesion-mediating fluid and curved claws to adhere to, and mechanically interlock with canopy substrates, respectively (Federle et al. 2002; Endlein and Federle 2008). Although adhesion and locomotion are functionally linked (Federle and Endlein 2004; Endlein and Federle 2008), direct comparisons between adhesive performance and locomotor performance are few, particularly under variable environmental conditions (but see Federle et al. 2000; Stark and Yanoviak 2018).

Given that the energetic costs of locomotion in ants are relatively low (e.g., Fewell 1988), and endurance constraints (e.g., oxygen debt during bursts of activity; Bennett and Huey 1990) are trivial at this small scale, the maintenance of secure attachment and consistent motion during foraging bouts in a variety of environmental conditions presumably are key limiting factors. Locomotor speed likely is constrained by the robustness and versatility of the adhesive system in complex environments, especially for broad-ranging foragers. Thus, when attachment is compromised, we expect a compensatory reduction in locomotor performance (measured as running speed); however, this expectation is not always supported, suggesting that the functional morphology of locomotion in ants is very robust to environmental variation (Federle et al. 2000; Clay et al. 2010; Yanoviak et al. 2012; Stark and Yanoviak 2018).

Although the forest canopy is a linear network of stems from the perspective of small cursorial organisms like ants, rough, narrow, and steeply inclined surfaces are difficult to avoid in this setting (Schoener 1968; Mattingly and Jayne 2004). Stem roughness is likely to be

1
2
3 85 especially problematic for ants because adhesive performance is dependent on the relationship
4
5 86 between surface asperity size and adhesive unit size (e.g., tarsal pad, claw tip). In instances
6
7
8 87 where the asperity size and adhesive unit size match, adhesion and mechanical interlocking can
9
10 88 fail due to a decrease in contact area (Figure 1; Scholz et al. 2010; Zhou et al. 2014; Song et al.
11
12 89 2016). Conversely, as surface asperity size increases, adhesive performance should increase,
13
14 90 although this is dependent on body size (Figure 1; Yanoviak et al. 2017). In addition to surface
15
16 91 roughness, natural vegetative substrates have diverse features that potentially impact ant
17
18 92 performance (Federle et al. 2000; Koch et al. 2008; Barthlott et al. 2017). Finally, arboreal ants
19
20 93 commonly locomote on the stems of climbing plants (e.g., vines) that vary in inclination and
21
22 94 diameter, and these factors potentially influence ant foraging behavior (Clay et al. 2010;
23
24 95 Yanoviak et al. 2012).

25
26
27
28 96 The primary goal of this study was to measure the effect of local substrate characteristics
29
30 97 (roughness, diameter, and inclination) on arboreal ant adhesive and locomotor performance. We
31
32 98 hypothesized that adhesive performance and running speed vary according to the match between
33
34 99 substrate asperity size and attachment structure size. Specifically, we predicted lower adhesive
35
36 100 performance on substrates with fine asperity size and higher adhesive performance on substrates
37
38 101 with large asperities, relative to smooth surfaces (Figure 1). We expected locomotor
39
40 102 performance, measured as running speed, to slow as a consequence of reduced adhesion on fine
41
42 103 rough substrates and on very rough, structurally complex substrates like tree bark. Secondly,
43
44 104 we predicted that ant running speed would also slow with declining stem diameter and increasing
45
46 105 angle of stem inclination, due to the biomechanical limitations of appendages and adhesive
47
48 106 anatomy. We explored these predictions with laboratory and field experiments on artificial and
49
50 107 natural substrates with one common tropical arboreal ant species.
51
52
53
54
55
56
57
58
59
60

108

109 **Materials and Methods**

110 Data were collected on Barro Colorado Island (BCI), Panama (9.15°N, 79.85°W) during
111 the 2015 wet season (May - July) and in June 2019. More information about this site is available
112 elsewhere (Croat 1978; Leigh 1996). We measured adhesive and locomotor performance of
113 workers of a common arboreal ant species (*Cephalotes atratus* L., Hymenoptera, Formicidae,
114 Myrmicinae). Workers were hand collected from at least two colonies daily, and housed (alive)
115 with their nestmates in small plastic containers until needed (generally < 12 hr). Only intact
116 individuals that could articulate all legs were used in experiments. Prior to experimental testing
117 in the laboratory or the field, focal ants were acclimated to the appropriate conditions
118 (laboratory: ca. 22°C and 80% RH; field: ca. 30°C and 80% RH) for at least 1 hr. All ants were
119 weighed to the nearest 0.1 mg following experiments.

120

121 *Ant Adhesive Morphology*

122 We used tarsal pad area and tarsal claw diameter as basic measures of ant adhesive
123 morphology. Tarsal pad area values were obtained from prior studies (Stark et al. 2018) in which
124 digital images of tarsal pads engaged with a glass surface were measured using NIH ImageJ
125 (version 1.46r; see Stark et al. 2018 for details). Tarsal claw diameter was measured using
126 scanning electron microscopy (SEM; Zeiss Supra 35 SEM, Jena, Germany). Ant tarsi were
127 separated from three frozen individuals and mounted so that the claw tips were in clear view
128 (i.e., facing up or laterally). We used NIH ImageJ to measure the diameter of the claw tips on
129 digital SEM images. Specifically, we used the *oval measurement* tool (locking it into a circle) to

outline the perimeter of the tarsal claw tip, then converted the area of the circular overlay to diameter.

Surface Characterization

We used plate glass and seven different grades of sandpaper (grit size = P2500, P2000, P1500, P320, P120, P80, and P36; Table 1) to test ant adhesion and locomotor performance on smooth and variably rough substrates. For the purposes of this project, we assumed glass roughness is effectively zero. The surface asperities of plate glass exist at the nanometer scale, thus they are at least three orders of magnitude smaller than the micrometer-scale size of the attachment structures of ants.

We quantified the asperity size of each sandpaper grit size as the average maximum length of 5 particles chosen in three different locations on a sandpaper sample (i.e., 15 particles per sandpaper type). Specifically, a digital grid was overlaid on images of each of the three sample locations on a sheet of sandpaper. Due to the broad range in asperity size, grid areas were set at 10 mm² for P36 grit sandpaper, 2 mm² for P80 grit sandpaper, 0.75 mm² for P120 grit sandpaper, and 0.05 mm² for the remaining grit sizes (P2500, P2000, P1500, P320). These grid sizes provided six complete squares per sample area; we selected the largest exposed particle within each of the 5 clearest grid squares for measurement. Digital images of the sample areas were taken with a Zeiss Axio Zoom V16 microscope fitted with an AxioCam 305 color camera (Jena, Germany). We used the *grid plug-in* and *line tool* in NIH ImageJ to draw grid lines and measure the largest dimension of selected particles, respectively.

We measured ant adhesion on leaves from five different tree species: *Anacardium excelsum*, *Cavanillesia platanifolia*, *Dipteryx oleifera*, *Ochroma pyramidale*, and *Pseudobombax*

1
2
3 153 *septenatum*. We chose these species because they are common trees near the BCI lab clearing,
4
5 154 and superficially appear to span a broad range of surface roughness. In each case, we harvested
6
7 155 mature leaves from portions of the trees receiving full sun exposure. Only leaves free of
8
9 156 conspicuous epiphylls and damage were used for experiments. We quantified the surface
10
11 157 characteristics of each leaf species by first making sodium alginate impressions of the leaf
12
13 158 surface, and then making plaster casts of the impressions. The roughness profiles of these casts
14
15 159 were obtained with a Veeco contact profilometer (Plainview, NY, USA). On each cast, three
16
17 160 linear 6.102 mm passes using 5 mg of force were made with a 12.5 μ m stylus tip. These passes
18
19 161 did not include leaf veins. Profilometry data were uploaded to TopoBank (version 0.7.5;
20
21 162 <https://contact.engineering/>) to calculate root-mean-square (RMS) height deviation, following the
22
23 163 conventions of Jacobs et al. (2017). RMS height deviation is the average of the profile height
24
25 164 deviations from the mean height. Curvature was removed from the profile plots. The RMS height
26
27 165 deviation of each pass was used to calculate the mean RMS height deviation for each leaf
28
29 166 species. All leaves were measured and used in experiments within 24 h of collection. To prevent
30
31 167 dehydration and decay between trials, leaves were stored in a plastic bag with a wet paper towel
32
33 168 in the laboratory.

34
35
36
37
38
39
40 169 We used stems of two liana (woody vine) taxa (*Tontelea ovalifolia* and *Bauhinia* sp.) and
41
42 170 bark from the trunks of three tree species (*Anacardium excelsum*, *Alseis blackiana* and *Dipteryx*
43
44 171 *oleifera*) to characterize ant locomotor performance on non-foliar natural plant substrates.
45
46 172 Similar to the leaf substrates, we chose these focal species because they differed conspicuously
47
48 173 in surface roughness, which was quantified in a separate study (Yanoviak et al. 2017).
49
50 174 Specifically, field-based laser scanning was used to measure the mean amplitude of the bark
51
52 175 surface asperities, and a laser scanning confocal microscope was used to image surface
53
54
55
56
57
58
59
60

topography of liana stem samples (Yanoviak et al. 2017). Hereafter, all natural substrates used in experiments are referred to by genus only.

Adhesive Performance Experiments

The adhesive performance of ants was measured on sandpaper and tree leaves mounted to a flat surface. Adhesion to plate glass was used as a reference. In each case, tarsal adhesion was measured by clamping a thin nylon thread (Aurlfil, Milano, Italy) to a hand-held 10 or 20 g analog spring gauge (0.1g and 0.2g resolution respectively; Pesola, Schindellegi, Switzerland), and tying the free end around the ant petiole (Stark et al. 2018; Stark and Yanoviak 2018; Stark et al. 2019). We measured the adhesion of live ants clinging to experimental substrates in two orientations. First, ants were pulled approximately orthogonal to the horizontally positioned substrate surface (i.e., normal adhesion), then the substrate was mounted vertically using a stage with a clamp and ants were slid approximately parallel to the vertical substrate surface (i.e., shear adhesion; see Stark and Yanoviak 2018 for a schematic of the experimental set-up). In both orientations, each ant was induced to securely attach its adhesive pads by applying slight pressure to the gaster. Normal adhesion was defined as the maximum load (g) an ant could maintain before detaching from the experimental substrate (i.e., losing grip and springing free of the horizontal surface). Shear adhesion was defined as the maximum load (g) an ant resisted during an approximately 3 cm slide along the vertically positioned substrate. Ants were oriented head down for these experiments, so that their tarsal pads were fully engaged during vertical sliding. Each ant was tested in both orientations three times, but only the maximum of the three measurements in each orientation was used for statistical analysis. We tested normal and shear adhesion of fifteen ants per substrate.

199

200 *Locomotor Performance Experiments*

201 Locomotor performance (running speed, cm s^{-1}) was measured on 1 cm diameter wooden
202 dowels wrapped with sandpaper, leaves attached to a flat substrate, or ca. 1 cm diameter vines.
203 Running speed on bark was measured in the field by releasing ants on tree trunks at
204 approximately 1.5m above the ground (Yanoviak et al. 2017). A 1 cm glass rod was used as a
205 baseline reference for running speed on a smooth, hard substrate. We tested the effect of
206 substrate diameter on locomotor performance by measuring ant running speed on glass rods of
207 varying diameter (2, 5, 7, 9, 12 mm). Similarly, we tested the effects of substrate inclination on
208 locomotor performance by measuring ant running speed on a 1 cm glass rod secured at different
209 angles relative to horizontal (0, 30, 60, and 90°). In all experiments, running speed was measured
210 as the time required for an ant to traverse a 10 cm section of the test substrate. Only
211 uninterrupted, straight runs were used. Each ant was tested three times ($n = 30$ ants per
212 treatment), and only the maximum speed observed for a given ant was used for statistical
213 analysis.

215 *Statistical Analysis*

216 Normal and shear adhesion data did not fit the assumptions of a linear model, so we used
217 nonparametric Kruskal-Wallis tests with post hoc Wilcoxon comparisons when significant
218 differences occurred among treatment groups. Normal and shear adhesion were analyzed
219 separately as a function of either sandpaper grit size or leaf species. Load resisted divided by
220 body mass was used as the response variable to account for differences in body size. Body mass

and body size measures (e.g., head width) are highly correlated in *C. atratus* and other ants (Corn 1980; Kaspari and Weiser 1999; Yanoviak et al. 2005).

Running speed data fit the assumptions of a linear model, therefore we used three separate analysis of covariance (ANCOVA) tests to investigate differences in running speed on sandpaper substrates and natural substrates (leaves, vines) in the laboratory, and tree trunks in the field. Running speed on tree trunks in the field was analyzed separately from leaves and vines because ambient temperature differences affect running speed (Hurlbert et al. 2008; Yanoviak et al. 2017). We used multiple regression to test for differences in running speed on dowels of different diameter, where mass and diameter were the independent variables. Finally, we used an ANCOVA to test for differences in running speed on 1 cm dowels secured at different angles of inclination. Ant mass was used as the covariate in all ANCOVA models. Differences among means in ANCOVA tests were determined with post hoc Tukey HSD tests. Data were transformed when necessary to meet normality and homogeneity of variance assumptions. Analyses were conducted with JMP 10.0.0 software (SAS Institute, Inc. 2012) and all means are reported \pm s.e.m.

Results

Ant Morphology

The average mass of individual *C. atratus* workers used in this study was 40.3 ± 0.32 mg. The range of worker mass (12.4 - 79.0 mg) used in this study matches the range of worker sizes commonly encountered in the field (Corn 1980; Yanoviak et al. 2005). Their average tarsal pad area was 0.0140 ± 0.00078 mm² (Stark et al. 2018) and average tarsal claw diameter was 0.014 ± 0.0005 mm.

244

245 *Surface Characterization*

246 Sandpaper asperity sizes ranged from 0.02-1.19 mm in average maximum length (Table
247 1). These values are higher than reported by Federation of European Producers of Abrasives
248 (FEPA) because we used the largest linear dimension of maximum particle size, not average
249 particle size. The RMS height deviation of fresh leaves ranged ca. 0.01-0.02 mm (Table 2). Vine
250 and tree bark asperity height was reported elsewhere (Yanoviak et al. 2017), and ranged ca. 0.09-
251 2.15 mm (Table 2).

252

253 *Adhesive and Locomotor Performance on Artificially Rough Substrates*

254 Normal and shear adhesion of *C. atratus* ant workers on sandpaper substrates varied as a
255 function of the asperity size (i.e., sandpaper grit size; Normal Adhesion: $\chi^2 = 111.32$, $df = 7$, $P <$
256 0.0001 ; Shear Adhesion: $\chi^2 = 98.67$, $df = 7$, $P < 0.0001$; Figure 2A and 2B). In general, shear
257 adhesion was higher than normal adhesion, and substrates with larger asperity size produced
258 higher forces. Specifically, normal adhesion was significantly different among asperity sizes
259 except the largest sandpaper grits (P120, P80, P36). Normal adhesion to smooth glass produced
260 the same load resistance as adhesion to sandpaper grit P320, but not smaller asperity sizes (grit $>$
261 P320). Indeed, adhesion was lower (effectively zero) on these substrates than on smooth glass
262 (0.5 ± 0.07 g load resisted). Likewise, shear adhesion differed among asperity sizes except the
263 roughest sandpaper grits (P120, P80, P36). Adhesion consistently was higher on these rough
264 substrates than on substrates with small asperity sizes. Shear adhesion loads did not differ for
265 ants tested on smooth glass and on fine-grit sandpaper (P2000, P1500).

1
2
3 266 The running speed of *C. atratus* ant workers varied as a function of substrate asperity size
4
5 267 ($F_{15,224} = 3.45$, $P < 0.0001$; Figure 2C). Specifically, running speed was consistently slower on
6
7 268 smooth glass than on sandpaper, and tended to be marginally slower on the roughest sandpaper
8
9 269 grits (P80 and P36).

10 270
11
12
13
14 271 *Adhesive and Locomotor Performance on Natural Substrates*

15
16
17 272 Normal and shear adhesive performance of *C. atratus* workers differed among natural
18
19 273 substrate types (i.e., leaves; Normal Adhesion: $\chi^2 = 12.84$, $df = 4$, $P = 0.0121$; Shear Adhesion:
20
21 274 $\chi^2 = 20.74$, $df = 4$, $P = 0.0004$; Figure 3). Similar to adhesion results on sandpaper, normal
22
23 275 adhesion was much lower than shear adhesion on leaves. Normal adhesion was higher on
24
25 276 *Ochroma* leaves than on the *Cavanillesia* and *Dipteryx* leaves, but was similar among all other
26
27 277 leaf comparisons. Shear adhesion was higher on *Ochroma* and *Anacardium* leaves than on
28
29 278 *Dipteryx* and *Pseudobombax* leaves. All other paired comparisons were statistically similar.

30
31
32
33 279 The running speed of *C. atratus* ant workers differed as a function of leaf and vine type in
34
35 280 the laboratory ($F_{13,196} = 2.82$, $P = 0.0010$) and bark type in the field ($F_{5,85} = 8.05$, $P < 0.0001$).
36
37 281 Specifically, ant running speed was faster on *Dipteryx* and *Anacardium* leaves and *Bauhinia* vine
38
39 282 stems than on *Ochroma* leaves in the laboratory (Figure 4). In the field, ant running speed was
40
41 283 lower on *Anacardium* bark than on *Dipteryx* and *Alseis* bark (Figure 4).

42
43 284
44
45
46 285 *Locomotion on Narrow and Inclined Substrates*

47
48
49 286 The average running speed of *C. atratus* ant workers increased as substrate diameter
50
51 287 increased ($F_{2,147} = 69.90$, $P < 0.0001$, $R^2 = 0.49$; Figure 5). Running speed also differed as a
52
53 288 function of running orientation (running up or down an incline; $F_{15,96} = 2.35$, $P = 0.0065$; Figure

6) and substrate angle had a significant effect on running speed (interaction: $F = 3.69$, $df = 3$, $P = 0.0146$). The significant interaction was driven by orientation (single effect: $F = 12.55$, $df = 1$, $P = 0.0006$), such that ants running upward on a vertical dowel ran slower than those running downward.

Discussion

The relatively linear, reticulate substrates that characterize the tropical rainforest canopy present a variety of physical challenges for wingless arboreal ants. Reduction in worker ant adhesion and running speed on highly variable substrates may increase the likelihood of falling, extend the time needed to discover food, elevate energy requirements, and prolong exposure to dangerous abiotic and biotic factors (i.e., temperature, rain, wind, predators, competitors; Denny et al. 2001; Gissel Nielsen 2001; Sarty et al. 2006; Farji-Brener et al. 2018; Radnan et al. 2018; Stark et al. 2018). The results of this study show that the adhesive performance and locomotor performance of workers of one common arboreal ant species are resilient to variation in substrate roughness, diameter, and orientation. However, in some conditions the adhesive mechanism completely failed or running speed significantly declined. Thus, it is likely that *C. atratus* workers establish foraging routes that minimize the inclusion of substrates effecting reduced adhesion and running speed, as observed in other cursorial animals (Pounds 1988; Irschick and Losos 1999; Jones and Jayne 2012).

Surface roughness is a ubiquitous and unpredictable environmental variable that can impact the adhesive performance of many organisms, especially when asperity size matches adhesive unit size (Huber et al. 2007; Gorb and Gorb 2009; Scholz et al. 2010; Wolff and Gorb 2012; Gillies et al. 2014; Salerno et al. 2017; Pillai et al. 2020). The results of this study show

1
2
3 312 that when comparing asperity size, tarsal pad area, and claw tip size of *C. atratus* workers, the
4
5 313 finest polishing paper is similarly scaled (i.e., asperity size of P2500 is ca. 0.02 mm and tarsal
6
7 314 pad area and claw tip diameter are ca. 0.01 mm² and ca. 0.01 mm wide, respectively). This
8
9
10 315 similarity likely explains the low adhesive performance of *C. atratus* workers on P2500
11
12 316 sandpaper, even in shear sliding. Presumably this outcome is due to additional friction between
13
14
15 317 the tarsal pads and claws when sliding (Dai et al. 2002; Federle et al. 2004).
16

17 318 By contrast, the increase in adhesive performance with asperity size is counter to
18
19 319 theoretical predictions and other experimental results (Fuller and Tabor 1975). However, these
20
21 320 models do not take into account mechanical interlocking and friction from claws, which only
22
23 321 occurs at high surface asperity sizes (Dai et al. 2002). The results of this study suggest that
24
25 322 significant mechanical interlocking and friction from claws of *C. atratus* workers begins to occur
26
27 323 when surface asperities are ca. 0.13 mm wide (P320 sandpaper grit substrate) and dominate when
28
29 324 surface asperities are ca. 0.4 mm wide (P120 sandpaper grit substrate). The lack of difference in
30
31 325 adhesive and locomotor performance on grits at the larger end of our experimental range of
32
33 326 asperity (> 0.5 mm grain size) suggests that such asperities are the equivalent of boulders for
34
35 327 ants. Consequently, it appears that differences in size among grains averaging 0.5-1.2 mm in
36
37 328 maximum dimension are irrelevant to locomotion in *C. atratus* workers. The results also suggest
38
39 329 that shear sliding helps claws interlock at smaller surface asperity sizes better than when pulled
40
41 330 orthogonally from a substrate (i.e., asperity size \geq 0.3 mm vs. asperity size \geq 0.4 mm,
42
43 331 respectively). Evaluating these possibilities was beyond the scope of this study, but could be
44
45 332 accomplished via microscopic examination of tarsal-substrate interactions on diverse surfaces
46
47 333 (e.g., Stark et al. 2018).
48
49
50
51
52
53
54
55
56
57
58
59
60

Surface roughness is just one parameter that potentially influences ant performance on natural substrates; variables like wettability, trichome density, and the presence of wax crystals are also important (Davidson et al. 1989; Federle et al. 2000; Gorb and Gorb 2011; Stark and Yanoviak 2018). The plaster casting process used in this study generates a structurally imperfect replica of leaf surface properties, which likely contributed to the lack of strong RMS height deviation differences among leaf species that were otherwise superficially distinct. Indeed, the range of RMS height deviation observed among leaf casts was small, and matched the attachment structure size of ants (tarsal pads and claws). Regardless, when qualitatively comparing adhesion to glass and rough sandpaper with leaf adhesion, adhesion of *C. atratus* workers to natural leaf substrates was much lower overall. However, the normal and shear loads ants resisted on leaves were much higher than their body mass, suggesting that the risk of falling from a leaf surface is low under most natural circumstances. While the surface roughness of sandpaper and natural substrates (leaves, bark) used in this study are not easily comparable due to differences in characterization (i.e., asperity length vs. RMS height deviation vs. peak-valley asperity height), it was clear that surface roughness is significantly smaller on leaves than sandpaper and bark surfaces. Improving surface roughness measurement and quantification of substrates in the field would be a potentially useful future direction for this work.

The results of this study highlight the robustness of running speed across a variety of natural and artificial surface asperity sizes. Specifically, the running speed of *C. atratus* workers is very consistent across a range of leaf, vine, bark, and sandpaper substrates, as observed in related studies (Yanoviak et al. 2017). However, the slower running speeds on *Anacardium* bark (and our observations in the field) suggest that roughness amplitudes matching larger anatomical structures of worker ants, such as tarsi and legs, impose significant obstacles to locomotion

1
2
3 357 (Figure 1; Yanoviak et al. 2017). Very rough or unstable substrates like bark or gravel potentially
4
5 358 reduce foraging efficiency (e.g., causing dropping of heavy loads when returning to the nest;
6
7 359 Bernadou et al. 2011), making navigation on rough substrates even more costly. Thus, we would
8
9
10 360 expect *C. atratus* workers to avoid foraging on very rough bark and very smooth substrates (at
11
12 361 the roughness scale of glass), which also reduced running speed.

14
15 362 A key result of this work is the comparison of adhesive performance and running speed.
16
17 363 In particular, while running speed across artificial and natural substrates did not show strong
18
19 364 differences, adhesion varied significantly. This countered our prediction that adhesive
20
21 365 performance dictates running speed (Figure 1). The mechanism responsible for this discontinuity
22
23 366 has not been resolved (Stark et al. 2015; Stark and Yanoviak 2018). Many biological materials
24
25 367 like ant adhesive pads are viscoelastic, and thus dependent on the rate of material extension
26
27 368 (Federle et al. 2004; Vincent 2012). Therefore, it is possible that speed is maintained on poorly
28
29 369 adhesive substrates by increasing shear sliding rate (i.e., strain rate; Stark et al. 2015). It is also
30
31 370 possible that ants do not need their adhesive structures while running. For example, ants running
32
33 371 horizontally and vertically can make contact mainly with their proximal tarsal segments (i.e.,
34
35 372 running on their "heels"), and use tarsal hairs for additional friction as needed (Reinhardt et al.
36
37 373 2009; Wöhrle et al. 2017).

38
39 374 Running up or down inclined substrates often reduces running speed (Birn-Jeffery and
40
41 375 Higham 2014), and running on narrow substrates likely makes correct tarsal placement more
42
43 376 difficult, thereby slowing forward progress (Lammers and Biknevicius 2004; Frantsevich and
44
45 377 Cruse 2005). Given the prevalence of narrow and inclined substrates in arboreal habitats, (e.g.,
46
47 378 50% of the surfaces were < 0.8 cm in diameter and angled at ca. 60° in a Bahamian forest;
48
49 379 Mattingly and Jayne 2004), arboreal organisms likely routinely encounter physical challenges
50
51
52
53
54
55
56
57
58
59
60

that impact running speed. The results of this work show that substrate inclination and diameter affect running speed of *C. atratus* workers, although such effects are limited to extreme circumstances (i.e., narrow or vertical substrates). Similar inclination-related results were reported in the field and laboratory for ants running up and down slopes (Seidl and Wehner 2008; Ravindra 2014), but other studies found limited effects of substrate inclination (Seidl and Wehner 2008; Weihmann and Blickhan 2009; Holt and Askew 2012; Khuong et al. 2013; Norton et al. 2014). Inclination generally slows ascent in cursorial animals (Jayne and Byrnes 2015), and different experimental outcomes likely reflect differences among species (Grevé et al. 2019) and slope (i.e., most studies were not conducted on vertical substrates). Likewise, only very narrow vine substrates seem to constrain foraging substrate choice in arboreal ants (Clay et al. 2010). This pattern supports the habitat constraint hypothesis - i.e., that foraging worker ants should avoid substrates and conditions that negatively impact performance (Irschick and Losos 1999).

Conclusion

The results of this study show that canopy substrates can affect adhesive performance of foraging worker ants, but that locomotor performance is maintained in most contexts. Given this outcome, we would expect ants like *C. atratus* (with relatively larger workers) to avoid foraging in locations of the canopy where adhesion is compromised (i.e., smooth substrates). Conversely, we do not expect foraging paths to be influenced by substrate structure except where substrates are vertical or narrow. Indeed, running down a vertical substrate (verses a lower angle of inclination) may maximize time and energy benefits. Future work should focus on field observations of foraging arboreal ants in the canopy and more detailed analyses of the kinematics of adhesion and locomotion on variable substrates.

1

2

3

403

4

5

404

Acknowledgements

6

7

405

We thank Oris Acevedo, Belkys Jimenez, Melissa Cano, and the staff of the Smithsonian

8

9

406

Tropical Research Institute for logistical support in Panama, and the faculty and staff at the

10

11

407

Micro/Nano Technology Center and the Huson Imaging & Characterization Laboratory at the

12

13

408

University of Louisville. We also thank Keegan Thompson for help with sample preparation and

14

15

409

Tevis Jacobs for help with leaf surface profilometry analysis. This research was supported by

16

17

410

National Science Foundation grant DEB-1252614 to SPY.

18

19

20

21

22

23

24

25

26

27

411

412

Data Availability Statement

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

The data underlying this article will be shared on request to the corresponding author.

Figure Captions

Figure 1. Predicted performance (adhesion and running speed) of adhesive animals on rough substrates. On smooth to rough substrates performance is expected to be lower on fine-scale rough substrates that match the asperity size of the attachment structures (pads, claws) due to low contact area. Within this regime the attachment structure size dominates performance. On very rough substrates, body size effects dominate and performance is lowest on very rough substrates that match the size scale of feet and legs. Under this circumstance, surface asperities become significant obstacles. As substrate roughness increases, performance increases because the asperities essentially become hills at the scale of the whole animal while being relatively smooth at the adhesive interface. Ultimately, larger and larger hills increasingly have their own surface asperities (i.e., at the micrometer scale), which interact with adhesive structures at the smooth to rough end of the scale while also resisting the gravitational forces of inclination. This latter complex interaction is not illustrated here, but was partially explored experimentally.

Figure 2. Normal adhesion (A), shear adhesion (B), and running speed (C) of *Cephalotes atratus* ant workers on sandpaper substrates that vary in asperity size (i.e., grit size) and plate glass (reference substrate). Treatment groups denoted with the same letter are not significantly different. Boxes in A and B represent the 25th and 75th percentiles, the line marks the median, bars are the 90th and 10th percentiles. Filled circles in C are means and error bars are \pm s.e.m.

Figure 3. Normal adhesion (A) and shear adhesion (B) of *Cephalotes atratus* ant workers on leaves. Substrates are abbreviated by genus name (ANA: *Anacardium*; CAV: *Cavanillesia*; DIP:

1
2
3 436 *Dipteryx*; OCH: *Ochroma*; PSE: *Pseudobombax*). Treatment groups denoted with the same letter
4
5
6 437 are not significantly different. Plotted values are described in Figure 2A and B.
7
8 438
9
10 439 **Figure 4.** Average running speed of *Cephalotes atratus* ant workers on natural substrates (A,
11
12 440 leaves and vine stems; B, tree bark). Substrates in A are abbreviated by genus name (ANA:
13
14 441 *Anacardium*; BAU: *Bauhinia* (vine); CAV: *Cavanillesia*; DIP: *Dipteryx*; OCH: *Ochroma*; PSE:
15
16 442 *Pseudobombax*; TON: *Tontelea* (vine)). Treatment groups denoted with the same letter are not
17
18
19 443 significantly different. Error bars are \pm s.e.m.
20
21
22 444
23
24 445 **Figure 5.** Running speed of *Cephalotes atratus* ant workers on horizontal glass rods that vary in
25
26 446 diameter. Error bars are \pm s.e.m.
27
28
29 447
30
31 448 **Figure 6.** Running speed of *Cephalotes atratus* ant workers on a glass rod (1 cm) positioned at
32
33 449 various angles relative to horizontal (0, 30, 60, and 90°). Ants were positioned on the rod and
34
35 450 induced to either run up or down. Means that differ are indicated with an asterisk. Error bars are
36
37
38 451 \pm s.e.m.
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

References

- Alexander RM. 2003. Principles of animal locomotion. Princeton University Press.
- Arnold SJ. 1983. Morphology, performance and fitness. *American Zoologist* 23: 347-361.
- Barthlott W, Mail M, Bhushan B, Koch K. 2017. Plant surfaces: structures and functions for biomimetic innovations. *Nano-Micro Letters* 9: 23.
- Bennett AF, Huey R. 1990. Studying the evolution of physiological performance. *Oxford Surveys in Evolutionary Biology* 7: 251-284.
- Bernadou A, Espadaler X, Dos-Reis V, Fourcassié V. 2011. Effect of substrate roughness on load selection in the seed-harvester ant *Messor barbarus* L.(Hymenoptera, Formicidae). *Behavioral Ecology and Sociobiology* 65: 1763.
- Biewener A, Patek S. 2018. Animal locomotion. Oxford University Press.
- Birn-Jeffery AV, Higham TE. 2014. The scaling of uphill and downhill locomotion in legged animals. *Integrative and Comparative Biology* 54: 1159-1172.
- Carroll CR, Janzen DH. 1973. Ecology of foraging by ants. *Annual Review of Ecology and Systematics* 4: 231-257.
- Cartmill M. 1985. Climbing. Functional vertebrate morphology. Harvard University Press.
- Clay NA, Bauer M, Solis M, Yanoviak SP. 2010. Arboreal substrates influence foraging in tropical ants. *Ecological Entomology* 35: 417-423.
- Corn M. 1980. Polymorphism and polyethism in the neotropical ant *Cephalotes atratus* (L.). *Insectes Sociaux* 27: 29-42.
- Croat TB. 1978. Flora of Barro Colorado Island. Stanford University Press.
- Dagg AI. 1974. The locomotion of the camel (*Camelus dromedarius*). *Journal of Zoology* 174: 67-78.

- 475 Dai Z, Gorb SN, Schwarz U. 2002. Roughness-dependent friction force of the tarsal claw system
476 in the beetle *Pachnoda marginata* (Coleoptera, Scarabaeidae). Journal of Experimental
477 Biology 205: 2479-2488.
- 478 Davidson DW, Snelling RR, Longino JT. 1989. Competition among ants for myrmecophytes and
479 the significance of plant trichomes. Biotropica 21: 64-73.
- 480 Denny AJ, Wright J, Grief B. 2001. Foraging efficiency in the wood ant, *Formica rufa*: is time of
481 the essence in trail following? Animal Behaviour 62: 139-146.
- 482 Devigne C, Detrain C. 2006. How does food distance influence foraging in the ant *Lasius niger*:
483 the importance of home-range marking. Insectes Sociaux 53: 46-55.
- 484 Endlein T, Federle W. 2008. Walking on smooth or rough ground: passive control of pretarsal
485 attachment in ants. Journal of Comparative Physiology A 194: 49-60.
- 486 Farji-Brener AG, Dalton MC, Balza U, Curtis A, Lemus-Domínguez I, Fernández-Hilario R,
487 Cáceres-Levi D. 2018. Working in the rain? Why leaf-cutting ants stop foraging when
488 it's raining. Insectes Sociaux 65: 233-239.
- 489 Farji-Brener AG, Barrantes G, Laverde O, Fierro-Calderón K, Bascopé F, López A. 2007. Fallen
490 branches as part of leaf-cutting ant trails: their role in resource discovery and leaf
491 transport rates in *Atta cephalotes*. Biotropica 39: 211-215.
- 492 Federle W, Baumgartner W, Hölldobler B. 2004. Biomechanics of ant adhesive pads: frictional
493 forces are rate-and temperature-dependent. Journal of Experimental Biology 207: 67-74.
- 494 Federle W, Endlein T. 2004. Locomotion and adhesion: dynamic control of adhesive surface
495 contact in ants. Arthropod Structure & Development 33: 67-75.

- 496 Federle W, Riehle M, Curtis AS, Full RJ. 2002. An integrative study of insect adhesion:
497 mechanics and wet adhesion of pretarsal pads in ants. Integrative and Comparative
498 Biology 42: 1100-1106.
- 499 Federle W, Rohrseitz K, Holldobler B. 2000. Attachment forces of ants measured with a
500 centrifuge: better 'wax-runners' have a poorer attachment to a smooth surface. Journal of
501 Experimental Biology 203: 505-512.
- 502 Fewell JH. 1988. Energetic and time costs of foraging in harvester ants, *Pogonomyrmex*
503 *occidentalis*. Behavioral Ecology and Sociobiology 22: 401-408.
- 504 Fewell JH, Harrison JF, Lighton JR, Breed MD. 1996. Foraging energetics of the ant,
505 *Paraponera clavata*. Oecologia 105: 419-427.
- 506 Frantsevich LI, Cruse H. 2005. Leg coordination during turning on an extremely narrow
507 substrate in a bug, *Mesocerus marginatus* (Heteroptera, Coreidae). Journal of Insect
508 Physiology 51:1092-1104.
- 509 Fuller K, Tabor D. 1975. The effect of surface roughness on the adhesion of elastic solids.
510 Proceedings of the Royal Society of London A Mathematical and Physical Sciences 345:
511 327-342.
- 512 Garland Jr T, Losos JB. 1994. Ecological morphology of locomotor performance in squamate
513 reptiles. Ecological Morphology: Integrative Organismal Biology. p. 240-302.
- 514 Gillies AG, Henry A, Lin H, Ren A, Shiuan K, Fearing RS, Full RJ. 2014. Gecko toe and
515 lamellar shear adhesion on macroscopic, engineered rough surfaces. Journal of
516 Experimental Biology 217: 283-289.
- 517 Gissel Nielsen M. 2001. Energetic cost of foraging in the ant *Rhytidoponera aurata* in tropical
518 Australia. Physiological Entomology 26: 248-253.

- 519 Gorb E, Gorb S. 2009. Effects of surface topography and chemistry of *Rumex obtusifolius* leaves
520 on the attachment of the beetle *Gastrophysa viridula*. *Entomologia Experimentalis et*
521 *Applicata* 130: 222-228.
- 522 Gorb E, Gorb S. 2011. How a lack of choice can force ants to climb up waxy plant stems.
523 *Arthropod-Plant Interactions* 5: 297-306.
- 524 Grevé ME, Bláha S, Teuber J, Rothmaier M, Feldhaar H. 2019. The effect of ground surface
525 rugosity on ant running speed is species-specific rather than size dependent. *Insectes*
526 *Sociaux* 66: 355-364.
- 527 Holt NC, Askew GN. 2012. Locomotion on a slope in leaf-cutter ants: metabolic energy use,
528 behavioural adaptations and the implications for route selection on hilly terrain. *Journal*
529 *of Experimental Biology* 215: 2545-2550.
- 530 Huber G, Gorb SN, Hosoda N, Spolenak R, Arzt E. 2007. Influence of surface roughness on
531 gecko adhesion. *Acta Biomaterialia* 3: 607-610.
- 532 Hurlbert AH, Ballantyne F, Powell S. 2008. Shaking a leg and hot to trot: the effects of body size
533 and temperature on running speed in ants. *Ecological Entomology* 33: 144-154.
- 534 Irschick DJ, Higham TE. 2016. *Animal athletes: An ecological and evolutionary approach*.
535 Oxford University Press.
- 536 Irschick DJ, Losos JB. 1999. Do lizards avoid habitats in which performance is submaximal?
537 The relationship between sprinting capabilities and structural habitat use in Caribbean
538 anoles. *American Naturalist* 154: 293-305.
- 539 Jacobs TD, Junge T, Pastewka L. 2017. Quantitative characterization of surface topography
540 using spectral analysis. *Surface Topography: Metrology and Properties* 5: 013001.

- 541 Jayne BC, Byrnes G. 2015. The effects of slope and branch structure on the locomotion of a
542 specialized arboreal colubrid snake (*Boiga irregularis*). Journal of Experimental Zoology
543 Part A: Ecological Genetics and Physiology 323: 309-321.
- 544 Jones ZM, Jayne BC. 2012. Perch diameter and branching patterns have interactive effects on the
545 locomotion and path choice of anole lizards. Journal of Experimental Biology 215: 2096-
546 2107.
- 547 Kaspari M, Weiser M. 1999. The size–grain hypothesis and interspecific scaling in ants.
548 Functional Ecology 13: 530-538.
- 549 Khuong A, Lecheval V, Fournier R, Blanco S, Weitz S, Bezia J-J, Gautrais J. 2013. How do
550 ants make sense of gravity? A Boltzmann Walker analysis of *Lasius niger* trajectories on
551 various inclines. PLOS One 8: e76531.
- 552 Koch K, Bhushan B, Barthlott W. 2008. Diversity of structure, morphology and wetting of plant
553 surfaces. Soft Matter 4: 1943-1963.
- 554 Lammers AR, Biknevicius AR. 2004. The biodynamics of arboreal locomotion: the effects of
555 substrate diameter on locomotor kinetics in the gray short-tailed opossum (*Monodelphis*
556 *domestica*). Journal of Experimental Biology 207: 4325-4336.
- 557 Leigh EG Jr, Rand AS, Windsor DM. 1996. The ecology of a tropical forest, second ed.
558 Smithsonian Institution, Washington, DC.
- 559 Lewinson RT, Stefanyshyn DJ. 2016. A descriptive analysis of the climbing mechanics of a
560 mountain goat (*Oreamnos americanus*). Zoology 119: 541-546.
- 561 Mattingly WB, Jayne BC. 2004. Resource use in arboreal habitats: structure affects locomotion
562 of four ecomorphs of *Anolis* lizards. Ecology 85: 1111-1124.

1
2
3 563 Norton V, Stevens-Wood B, Harris WE. 2014. Flexibility of individual load-mass selection in
4
5 564 relation to foraging trail gradient in the leaf-cutter ant *Acromyrmex octospinosus*. Journal
6
7 565 of Insect Behavior 27: 370-384.
8
9
10 566 Pillai R, Nordberg E, Riedel J, Schwarzkopf L. 2020. Nonlinear variation in clinging
11
12 567 performance with surface roughness in geckos. Ecology and Evolution 10: 2597-2607.
13
14 568 Pounds JA. 1988. Ecomorphology, locomotion, and microhabitat structure: patterns in a tropical
15
16 569 mainland *Anolis* community. Ecological Monographs 58: 299-320.
17
18 570 Radnan GN, Gibb H, Eldridge DJ. 2018. Soil surface complexity has a larger effect on food
19
20 571 exploitation by ants than a change from grassland to shrubland. Ecological Entomology
21
22 572 43: 379-388.
23
24 573 Ravindra P. 2014. Ant runners: an analysis of running speed of *Leptogenys processionalis*
25
26 574 (Hymenoptera: Formicidae: Ponerinae). Current Science 106: 1187-1189.
27
28 575 Reinhardt L, Weihmann T, Blickhan R. 2009. Dynamics and kinematics of ant locomotion: do
29
30 576 wood ants climb on level surfaces? Journal of Experimental Biology 212: 2426-2435.
31
32 577 Ricklefs RE, Miles DB. 1994. Ecological and evolutionary inferences from morphology: an
33
34 578 ecological perspective. Ecological Morphology: Integrative Organismal Biology. p. 13-
35
36 579 41.
37
38 580 Salerno G, Rebora M, Gorb E, Kovalev A, Gorb S. 2017. Attachment ability of the southern
39
40 581 green stink bug *Nezara viridula* (Heteroptera: Pentatomidae). Journal of Comparative
41
42 582 Physiology A 203: 601-611.
43
44 583 Sarty M, Abbott KL, Lester PJ. 2006. Habitat complexity facilitates coexistence in a tropical ant
45
46 584 community. Oecologia 149: 465-473.
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 585 Schoener TW. 1968. The *Anolis* lizards of Bimini: resource partitioning in a complex fauna.
4
5
6 586 Ecology 49: 704-726.
7
8 587 Schoener TW. 1979. Generality of the size-distance relation in models of optimal feeding.
9
10 588 American Naturalist 114: 902-914.
11
12 589 Scholz I, Bückins M, Dolge L, Erlinghagen T, Weth A, Hischen F, Mayer J, Hoffmann S,
13
14
15 590 Riederer M, Riedel M. 2010. Slippery surfaces of pitcher plants: *Nepenthes* wax crystals
16
17 591 minimize insect attachment via microscopic surface roughness. Journal of Experimental
18
19 592 Biology 213: 1115-1125.
20
21 593 Seidl T, Wehner R. 2008. Walking on inclines: how do desert ants monitor slope and step length.
22
23
24 594 Frontiers in Zoology 5: 8.
25
26 595 Song Y, Dai Z, Wang Z, Ji A, Gorb SN. 2016. The synergy between the insect-inspired claws
27
28 596 and adhesive pads increases the attachment ability on various rough surfaces. Scientific
29
30
31 597 Reports 6: 26219.
32
33 598 Stark AY, Arstingstall K, Yanoviak SP. 2018. Adhesive performance of tropical arboreal ants
34
35 599 varies with substrate temperature. Journal of Experimental Biology 221: jeb171843.
36
37
38 600 Stark AY, Davis HR, Harrison WK. 2019. Shear adhesive performance of leaf-cutting ant
39
40 601 workers (*Atta cephalotes*). Biotropica 51: 572-580.
41
42 602 Stark AY, Ohlemacher J, Knight A, Niewiarowski PH. 2015. Run don't walk: locomotor
43
44 603 performance of geckos on wet substrates. Journal of Experimental Biology 218: 2435-
45
46 604 2441.
47
48
49 605 Stark AY, Yanoviak SP. 2018. Adhesion and running speed of a tropical arboreal ant
50
51 606 (*Cephalotes atratus*) on wet substrates. Journal of the Royal Society Open Science 5:
52
53 607 181540.
54
55
56
57
58
59
60

- Stephens DW, Krebs JR. 1986. Foraging theory. Princeton University Press.
- Torres-Contreras H, Vasquez R. 2004. A field experiment on the influence of load transportation and patch distance on the locomotion velocity of *Dorymyrmex goetschi* (Hymenoptera, Formicidae). *Insectes Sociaux* 51: 265-270.
- Vincent J. 2012. Structural biomaterials. Princeton University Press.
- Vogel S. 1988. Life's devices: The physical world of animals and plants. Princeton University Press.
- Weihmann T, Blickhan R. 2009. Comparing inclined locomotion in a ground-living and a climbing ant species: sagittal plane kinematics. *Journal of Comparative Physiology A* 195: 1011.
- Wilson EO. 1968. The ergonomics of caste in the social insects. *American Naturalist* 102: 41-66.
- Wöhrle T, Reinhardt L, Blickhan R. 2017. Propulsion in hexapod locomotion: how do desert ants traverse slopes? *Journal of Experimental Biology* 220: 1618-1625.
- Wolff JO, Gorb SN. 2012. Surface roughness effects on attachment ability of the spider *Philodromus dispar* (Araneae, Philodromidae). *Journal of Experimental Biology* 215: 179-184.
- Yanoviak S, Frederick D. 2014. Water surface locomotion in tropical canopy ants. *Journal of Experimental Biology* 217: 2163-2170.
- Yanoviak S, Kaspari M. 2000. Community structure and the habitat templet: ants in the tropical forest canopy and litter. *Oikos* 89: 259-266.
- Yanoviak SP, Dudley R, Kaspari M. 2005. Directed aerial descent in canopy ants. *Nature* 433: 624-626.

- 630 Yanoviak SP, Kaspari M, Dudley R. 2009. Gliding hexapods and the origins of insect aerial
631 behaviour. *Biology Letters* 5: 510-512.
- 632 Yanoviak SP, Munk Y, Dudley R. 2015. Arachnid aloft: directed aerial descent in neotropical
633 canopy spiders. *Journal of the Royal Society Interface* 12: 20150534.
- 634 Yanoviak SP, Silveri C, Hamm CA, Solis M. 2012. Stem characteristics and ant body size in a
635 Costa Rican rain forest. *Journal of Tropical Ecology* 28: 199-204.
- 636 Yanoviak SP, Silveri C, Stark AY, Van Stan JT, Levia DF. 2017. Surface roughness affects the
637 running speed of tropical canopy ants. *Biotropica* 49: 92-100.
- 638 Ydenberg RC, Welham CVJ, Schmid-Hempel R, Schmid-Hempel P, Beauchamp G. 1994. Time
639 and energy constraints and the relationships between currencies in foraging theory.
640 *Behavioral Ecology* 5: 28-34.
- 641 Yunker JA, Meserve PL, Gutiérrez JR. 2002. Small-mammal foraging behavior: mechanisms for
642 coexistence and implication for population dynamics. *Ecological Monographs* 72: 561-
643 577.
- 644 Zhou Y, Robinson A, Steiner U, Federle W. 2014. Insect adhesion on rough surfaces: analysis of
645 adhesive contact of smooth and hairy pads on transparent microstructured substrates.
646 *Journal of the Royal Society Interface* 11: 20140499.

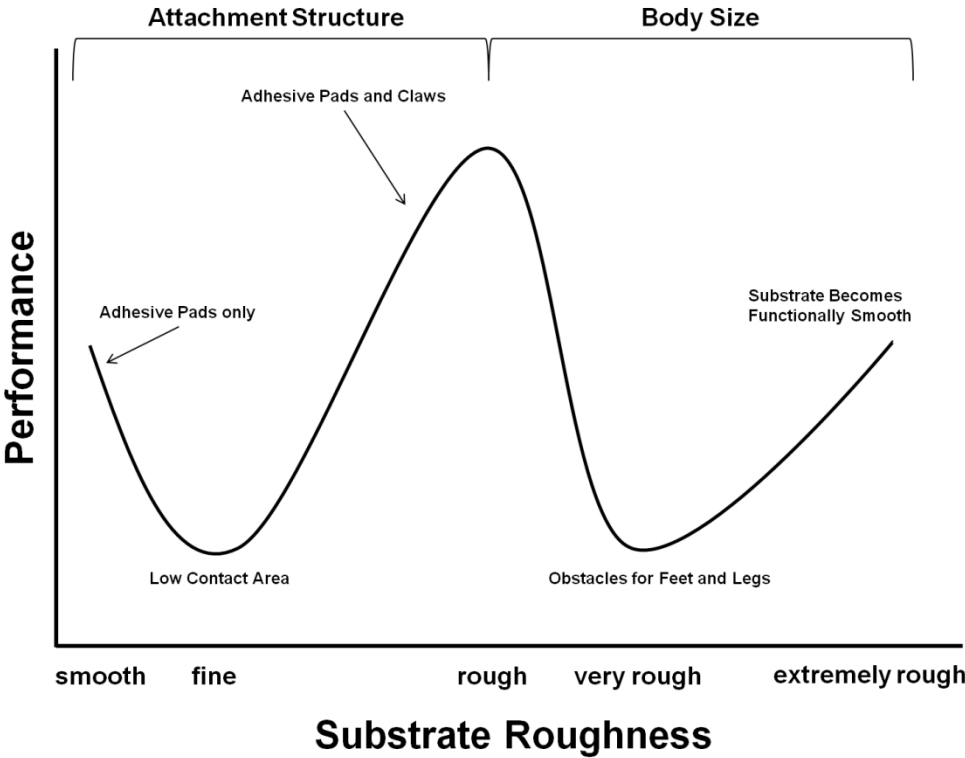


Figure 1. Predicted performance (adhesion and running speed) of adhesive animals on rough substrates. On smooth to rough substrates performance is expected to be lower on fine-scale rough substrates that match the asperity size of the attachment structures (pads, claws) due to low contact area. Within this regime the attachment structure size dominates performance. On very rough substrates, body size effects dominate and performance is lowest on very rough substrates that match the size scale of feet and legs. Under this circumstance, surface asperities become significant obstacles. As substrate roughness increases, performance increases because the asperities essentially become hills at the scale of the whole animal while being relatively smooth at the adhesive interface. Ultimately, larger and larger hills increasingly have their own surface asperities (i.e., at the micrometer scale), which interact with adhesive structures at the smooth to rough end of the scale while also resisting the gravitational forces of inclination. This latter complex interaction is not illustrated here, but was partially explored experimentally.

63x48mm (600 x 600 DPI)

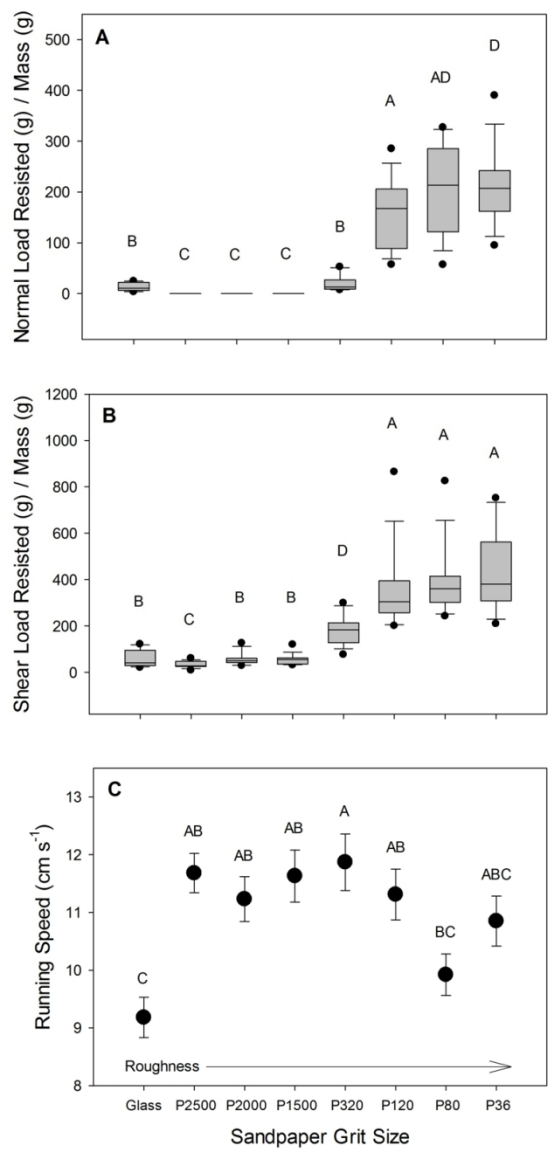


Figure 2. Normal adhesion (A), shear adhesion (B), and running speed (C) of *Cephalotes atratus* ant workers on sandpaper substrates that vary in asperity size (i.e., grit size) and plate glass (reference substrate). Treatment groups denoted with the same letter are not significantly different. Boxes in A and B represent the 25th and 75th percentiles, the line marks the median, bars are the 90th and 10th percentiles. Filled circles in C are means and error bars are \pm s.e.m.

34x71mm (600 x 600 DPI)

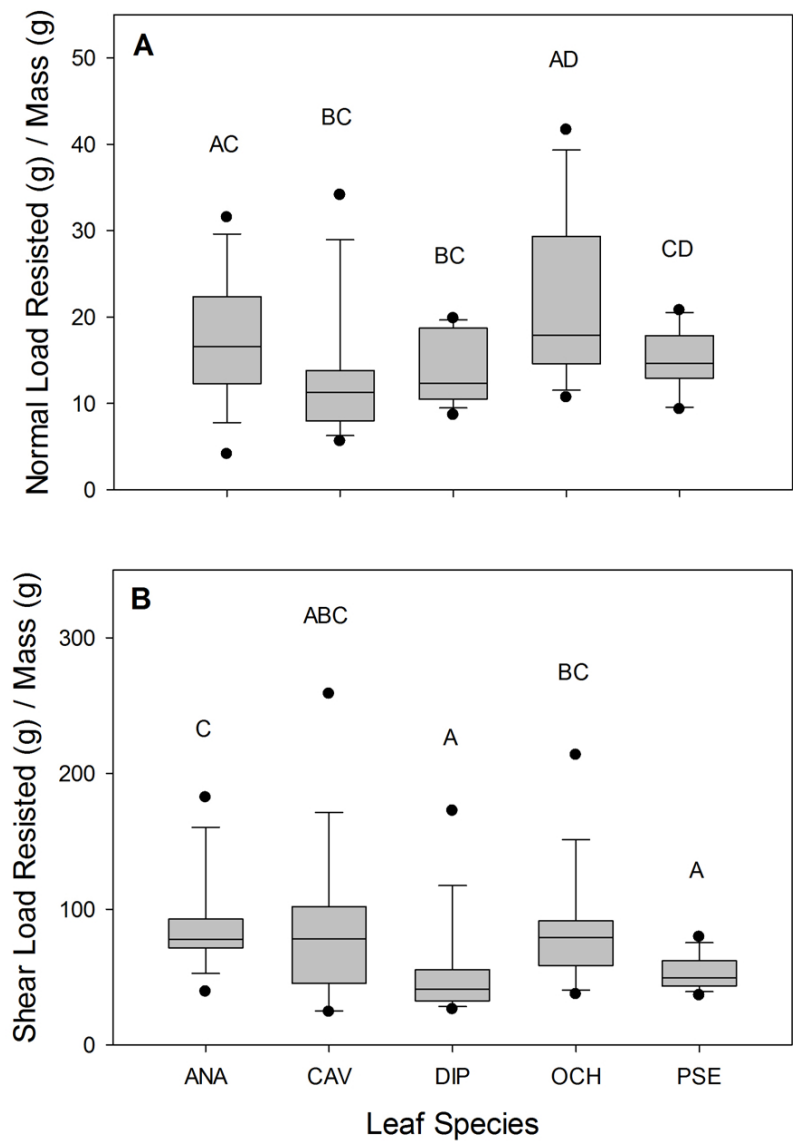


Figure 3. Normal adhesion (A) and shear adhesion (B) of *Cephalotes atratus* ant workers on leaves. Substrates in are abbreviated by genus name (ANA: *Anacardium*; CAV: *Cavanillesia*; DIP: *Dipteryx*; OCH: *Ochroma*; PSE: *Pseudobombax*). Treatment groups denoted with the same letter are not significantly different. Plotted values are described in Figure 2A and B.

40x56mm (600 x 600 DPI)

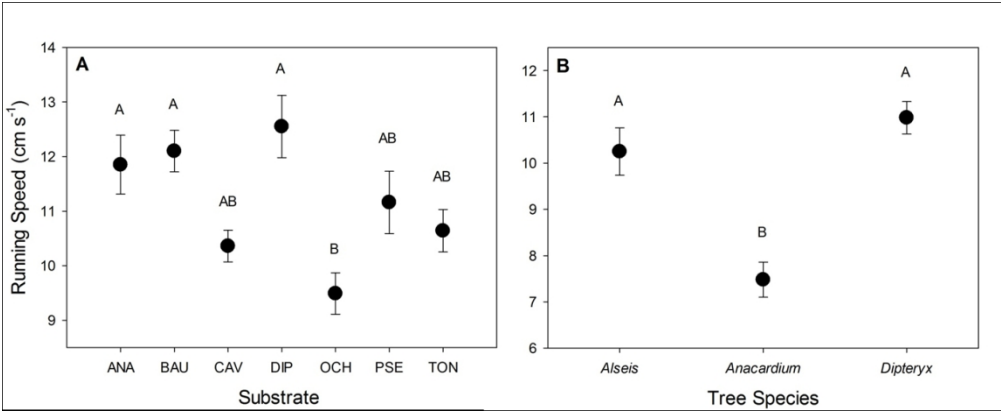


Figure 4. Average running speed of *Cephalotes atratus* ant workers on natural substrates (A, leaves and vine stems; B, tree bark). Substrates in A are abbreviated by genus name (ANA: *Anacardium*; BAU: *Bauhinia* (vine); CAV: *Cavanillesia*; DIP: *Dipteryx*; OCH: *Ochroma*; PSE: *Pseudobombax*; TON: *Tontelea* (vine)). Treatment groups denoted with the same letter are not significantly different. Error bars are \pm s.e.m.

59x24mm (600 x 600 DPI)

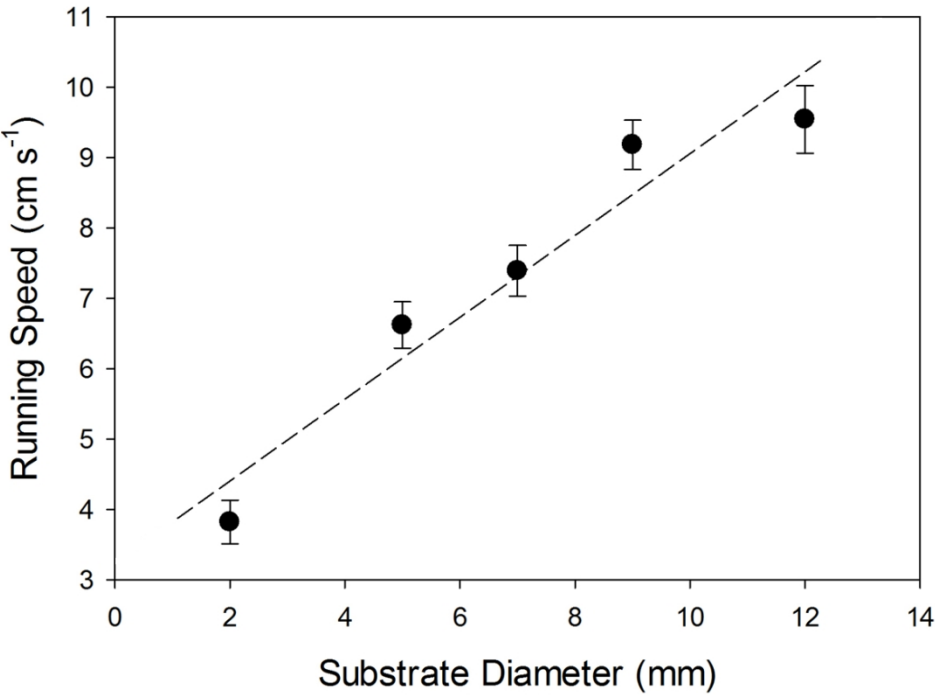


Figure 5. Running speed of *Cephalotes atratus* ant workers on horizontal glass rods that vary in diameter. Error bars are ± s.e.m.

53x40mm (600 x 600 DPI)

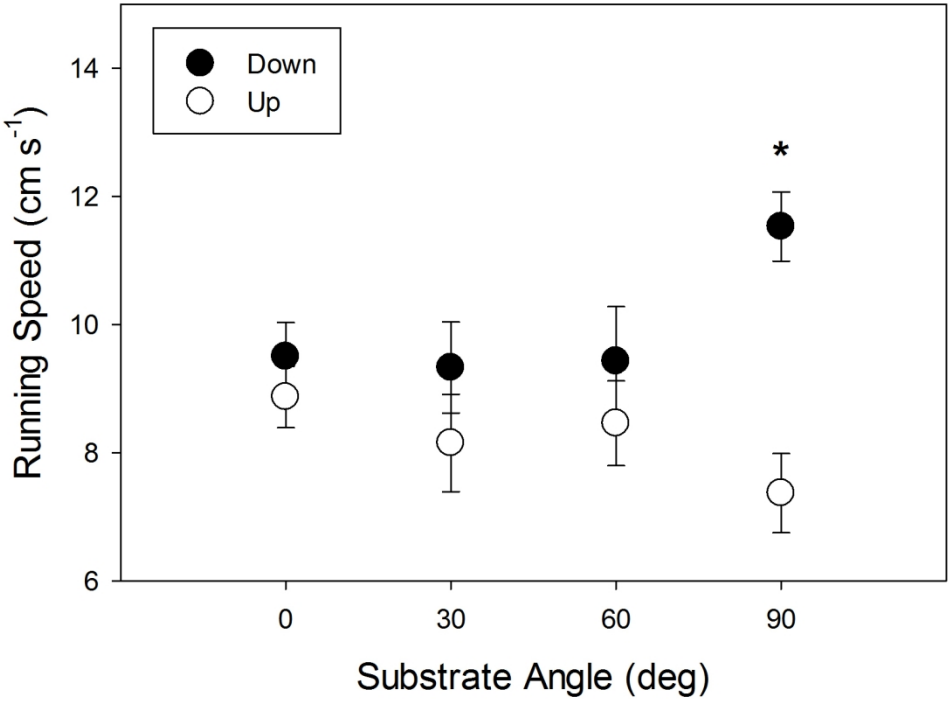


Figure 6. Running speed of *Cephalotes atratus* ant workers on a glass rod (1 cm) positioned at various angles relative to horizontal (0, 30, 60, and 90°). Ants were positioned on the rod and induced to either run up or down. Means that differ are indicated with an asterisk. Error bars are \pm s.e.m.

76x60mm (600 x 600 DPI)

Table 1. Average maximum asperity size (measured as particle length) of sandpaper substrates characterized by commercially available grit size. Federation of European Producers of Abrasives (FEPA) asperity size values for the same grit size sandpaper substrates are provided as reference. Means are reported \pm s.e.m.

Sandpaper Grit Size (FEPA)	Average Maximum Asperity Size (mm)	FEPA Asperity Size Reference Values (mm)
P36	1.186 \pm 0.0891	0.538
P80	0.521 \pm 0.0232	0.201
P120	0.354 \pm 0.0190	0.125
P320	0.130 \pm 0.0066	0.0462
P1500	0.035 \pm 0.0008	0.0126
P2000	0.031 \pm 0.0012	0.0103
P2500	0.022 \pm 0.0008	0.0084

Table 2. Average asperity size of natural leaf substrates. Vine and bark substrate asperity sizes are reported elsewhere (Yanoviak et al. 2017). Surface asperity size is measured as root mean square (RMS) height deviation. Means are reported \pm s.e.m.

Species	Surface Asperity Size (mm)
<i>Anacardium excelsum</i>	0.016 ± 0.0023
<i>Cavanillesia platanifolia</i>	0.018 ± 0.0009
<i>Dipteryx oleifera</i>	0.010 ± 0.0009
<i>Ochroma pyramidale</i>	0.012 ± 0.0018
<i>Pseudobombax septenatum</i>	0.016 ± 0.0031