



Short-Term Coral Bleaching Is Not Recorded by Skeletal Boron Isotopes

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Abstract

Coral skeletal boron isotopes have been established as a proxy for seawater pH, yet it remains unclear if and how this proxy is affected by seawater temperature. Specifically, it has never been directly tested whether coral bleaching caused by high water temperatures influences coral boron isotopes. Here we report the results from a controlled bleaching experiment conducted on the Caribbean corals *Porites divaricata*, *Porites astreoides*, and *Orbicella faveolata*. Stable boron ($\delta^{11}\text{B}$), carbon ($\delta^{13}\text{C}$), oxygen ($\delta^{18}\text{O}$) isotopes, Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca ratios, as well as chlorophyll *a* concentrations and calcification rates were measured on coral skeletal material corresponding to the period during and immediately after the elevated temperature treatment and again after 6 weeks of recovery on the reef. We show that under these conditions, coral bleaching did not affect the boron isotopic signature in any coral species tested, despite significant changes in coral physiology. This contradicts published findings from coral cores, where significant decreases in boron isotopes were interpreted as corresponding to times of known mass bleaching events. In contrast, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ exhibited major enrichment corresponding to decreases in calcification rates associated with bleaching. Sr/Ca of bleached corals did not consistently record the 1.2°C difference in seawater temperature during the bleaching treatment, or alternatively show a consistent increase due to impaired photosynthesis and calcification. Mg/Ca, U/Ca, and Ba/Ca were affected by coral bleaching in some of the coral species, but the observed patterns could not be satisfactorily explained by temperature dependence or changes in coral physiology. This demonstrates that coral boron isotopes do not record short-term bleaching events, and therefore cannot be used as a proxy for past bleaching events. The robustness of coral boron isotopes to changes in coral physiology, however, suggests that reconstruction of seawater pH using boron isotopes should be uncompromised by short-term bleaching events.

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Introduction

The world's oceans are simultaneously warming and acidifying at unprecedented pace due to rising atmospheric CO₂ concentrations [1], thereby severely threatening marine ecosystems [2,3]. Coral reefs are particularly vulnerable because they are highly sensitive to changes in both seawater temperature and pH [4–6].

Coral reefs are increasingly subject to periods with unusually warm seawater temperatures which can cause coral bleaching, defined as a significant loss of photosynthetic pigments and/or algal endosymbionts from the coral tissue [7,8–12]. Although corals can sometimes recover from bleaching events [13–18], other corals are significantly impacted by bleaching events, resulting in widespread mortality for more sensitive species [10,11,19]. As surface ocean temperatures have already increased by 0.6°C since preindustrial times and are projected to increase by at least

another 2.0°C under a business as usual scenario by the year 2100 [1], coral bleaching events are expected to increase in frequency and intensity [11,20–22], thus contributing to the worldwide decline of corals reefs [23,24].

In addition to greenhouse-induced warming, rising levels of atmospheric CO₂ concentrations have already caused a drop in surface seawater pH by approximately 0.1 unit compared to preindustrial times [25], and a further decrease of 0.3 pH units by the end of this century has been predicted [1,26]. This is of particular concern for marine calcifying organisms including corals because ocean acidification (OA) typically compromises calcification [27–33]. Although it is now recognised that scleractinian corals have a capacity to internally up-regulate pH [34–40], it has been estimated that coral calcification will decrease by up to 22% by the year 2100 [41]. Therefore, predicting how

global climate change will affect ocean chemistry and marine ecosystems in the coming decades is of imminent concern and requires understanding of how factors such as ocean temperature and pH have varied naturally in the past. Since direct measurements of ocean chemistry are only available for the most recent decades, proxy records are of crucial importance.

Coral skeletal boron isotopes ($\delta^{11}\text{B}$) are promising proxies of seawater pH [31,34,40,42–45] that have been used to reconstruct paleo-pH [46–48]. Their use as a pH-proxy is based on the fact that the relative abundance of the two major aqueous boron species in seawater, boric acid ($\text{B}(\text{OH})_3$) and borate ion ($\text{B}(\text{OH})_4^-$), are dependent on seawater pH. Further, their isotopic composition is also pH dependent and differs by about 27‰. It is generally thought that corals mainly incorporate the charged borate ion into the skeleton as their isotopic composition reflects that of borate [34,42,49], although this has been questioned by some studies [50,51]. However, it is now established that coral skeletal $\delta^{11}\text{B}$ does not directly record seawater pH but rather reflects the pH at the site of calcification [34,35,40,42], which is typically elevated by up to 1 unit [35,37–39] or more [36] relative to ambient seawater. Therefore, species-specific calibrations have to be performed to quantify the amount of pH-upregulation in order to reconstruct seawater pH [34].

One key requirement for any proxy is that it faithfully records the environmental variable of interest without being influenced by other factors. Several experimental studies have attempted to identify potentially confounding effects of light, depth, temperature, and feeding on coral $\delta^{11}\text{B}$ but, taken together, their results are contradictory. For example, Hönisch et al. [45] showed that various light, depth, and feeding regimes did not influence $\delta^{11}\text{B}$ of *Porites compressa* and *Montipora compressa*. However, another study using branching *Acropora* sp. found that $\delta^{11}\text{B}$ decreased as light intensity increased, introducing a bias of about 0.05 pH units [43].

Regarding temperature, Reynaud et al. [44] observed no temperature effect on $\delta^{11}\text{B}$ of *Acropora* sp. cultured at 25 and 28°C and two pCO₂ levels. Interestingly, Dissard et al. [43] cultured *Acropora* sp. using the same protocol as in Reynaud et al. [44] at 22, 25 and 28°C, and found a significant temperature effect only at the lower temperatures of 22 and 25°C, corresponding to an increase in $\delta^{11}\text{B}$ of about 0.02 pH units. A significant temperature effect was not observed between 25 and 28°C, consistent with the findings of Reynaud et al. [44]. It therefore appears that other factors may have confounded this work or, at least in *Acropora*, that temperature effects on coral $\delta^{11}\text{B}$ are non-linear and only exist for lower temperature ranges. However, given that both studies only used nubbins prepared from a single parent colony of one species, it is not clear whether these results could be reproduced if nubbins were used from multiple parent colonies (thus representing genetically different individuals), and/or different species of coral such as massive *Porites* sp., which is frequently used for paleoclimate reconstruction.

Although the experimental evidence available to date suggests that temperature may only affect coral $\delta^{11}\text{B}$ at lower temperatures (i.e., <25°C), no studies have examined stressfully high temperatures that can result in coral bleaching. Evidence from a massive *Porites* core from the Great Barrier Reef suggests that significant short-term decreases in $\delta^{11}\text{B}$ could be the result of coral bleaching events [47]. They proposed that this was due to a major reduction in pH at the site of calcification associated with impaired calcification, and suggest that coral $\delta^{11}\text{B}$ may be used as a proxy for past bleaching events [47]. Although coral $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ as well as Sr/Ca have been suggested as potential bleaching proxies, they were unreliable in this regard or in the case of Sr/Ca lack

sufficient detail [52–55]. Therefore, establishing coral $\delta^{11}\text{B}$ as a bleaching proxy could present a major step forward in understanding how often bleaching events occurred in the past. Further, it is essential that temperature effects on coral $\delta^{11}\text{B}$ be better understood over a large range of temperatures and using a variety of coral species to improve the reconstruction of past seawater pH.

To address these questions, we conducted a controlled, replicated bleaching experiment using the Caribbean coral species *Porites divaricata*, *Porites astreoides* and *Orbicella faveolata*. Coral skeletal $\delta^{11}\text{B}$, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ as well as Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca were analysed in samples collected immediately after the bleaching experiment and again after 6 weeks of recovery on the reef. Several physiological measurements were concurrently performed on these corals [13,56–59], providing a rich background of physiological information within which to interpret our findings. Based on the evidence presented by Wei et al. [47], we hypothesized that $\delta^{11}\text{B}$ of bleached corals decreases compared to non-bleached controls, and approaches (i.e., increases to) the values of non-bleached controls during recovery.

Methods

Coral Bleaching Experiment

The corals used for this study were taken from an experiment where three species of Caribbean corals were experimentally bleached in two consecutive summers [13]. These corals were physiologically fully recovered (i.e., there were no significant differences between treatment and control corals in any of the measured variables) after a year on the reef prior to exposure to the elevated temperature stress for a second time [57,59]. A detailed description of the experimental design can be found in Grottoli et al. [13].

Briefly, coral fragments were collected from 9 healthy colonies of *Porites divaricata* (branching morphology), *Porites astreoides* (mounding/encrusting morphology), and mounding *Orbicella faveolata* (formerly *Montastraea faveolata*) [60] (large mounding morphology) in July 2009 from reefs near Puerto Morelos, Yucatan Peninsula, Mexico (20°50'N, 86°52'W). All collections and experiments were conducted following the rules and regulations of Mexico and imported to the USA under CITES permits held by UNAM-ICML and the Ohio State University. Half of the coral fragments from each parent colony were randomly assigned to each treatment: (1) ambient control fragments were maintained in tanks with ambient seawater temperature ($30.66 \pm 0.24^\circ\text{C}$), and (2) treatment fragments were placed in tanks with elevated seawater temperature ($31.48 \pm 0.20^\circ\text{C}$). Seawater temperature in the treatment tanks was gradually elevated over the course of a week. Corals were not fed but had access to unfiltered seawater. High-precision seawater pH measurements were not made. After a total of 15 days, temperature in all tanks was returned to ambient levels, and all coral fragments were placed back *in situ* on the reef for one year.

In July 2010, the experiment was repeated. All corals that had served as ambient control fragments the previous summer were placed in tanks with ambient seawater ($30.40 \pm 0.23^\circ\text{C}$), whereas all corals that had been used as treatment fragments were maintained in tanks with elevated temperature ($31.60 \pm 0.24^\circ\text{C}$). After 17 days, all tanks were returned to ambient temperature levels and one control and one treatment fragment per colony of each species were then frozen for geochemical analyses (= 0 weeks on the reef). All remaining fragments were placed on the back reef. All remaining corals (i.e., one control and one treatment fragment

per colony of each species) were recollected from the reef after 6 weeks and frozen for geochemical analyses.

Physiological Analyses

Chlorophyll *a*. Coral tissue was removed from the skeleton of a portion of each fragment with a WaterPik, homogenized, and centrifuged to separate the algal endosymbiont from the animal tissue. A subsample of the resuspended algal pellet was broken using glass beads in 100% acetone and subsequently stored at -20°C overnight. Chlorophyll *a* was then determined using a Shimadzu UV-VIS spectrophotometer and the equations of Jeffrey and Humphrey [61]. Chlorophyll *a* content was standardized to surface area, which was determined using the aluminium foil method [62].

Calcification. Published calcification rates determined using the buoyant weight technique [63] were reproduced from Grottoli et al. [13] with permission of the publisher (license 3424840151859).

Isotopic Analyses

Coral tissue was removed from the skeleton using a dental hygiene tool. The uppermost layer (approx. 0.25–0.5 mm) of the dried skeleton was then gently shaved with a diamond-tipped Dremel tool and ground to fine powder using agate mortar and pestles [52,64]. Approximately 2–11 mg of the skeletal powder was weighed in pre-cleaned, pre-weighed Teflon tubes. An aliquot of this powder was used for $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ analyses (see below). When there was <2 mg of powder available from individual fragments, samples from multiple fragments of the same species and time point were combined for $\delta^{11}\text{B}$ analyses.

Boron isotopes. The following procedures were carried out in a clean lab. To remove organic matter, samples were rinsed with H_2O , incubated with 2 ml of 6.25% NaClO for 15 min including 5 min of sonication, then centrifuged for 2 min at highest speed, and the supernatant removed. Powders were then rinsed $3\times$ with H_2O , with each rinse including 5 min sonication and 2 min centrifugation at highest speed. After cleaning, tubes were lightly capped and samples dried to constant dry weight by placing them on a 60°C hotplate with a heat-lamp next to it. Cleaned samples were dissolved in 0.58 N HNO_3 . An aliquot of this solution was then diluted for trace element measurements (see below), and the remaining solution extracted for boron using ion exchange resins [35,65].

The extracted boron was analysed at the University of Western Australia using either a Neptune Plus Multi-Collector Inductively Coupled Plasma Mass Spectrometer (MC-ICP-MS; Thermo Fisher Scientific) fitted with a PFA nebulizer and a cyclonic quartz spray chamber or a NU Plasma II MC-ICP-MS (NU Instruments). The boron isotopic composition of the skeleton ($\delta^{11}\text{B}$) was reported as the per mil deviation of the stable isotopes ^{11}B : ^{10}B relative to SRM-951. Sample measurements were bracketed by SRM-951 enriched to 24.7‰ and blank measurements [66], and all samples were analysed in duplicate. Repeated measurements of SRM-951 had a standard deviation (1σ) of $\pm 0.37\text{‰}$ ($n = 124$) for concentrations ranging from 100–200 ppb B.

Carbon and oxygen isotopes. A 80–100 μg aliquot of the dried and ground skeletal powder was analysed for $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ using an automated Kiel Carbonate Device coupled to a Stable Isotope Ratio Mass Spectrometer (SIRMS; Finnigan Delta IV) at The Ohio State University. Samples were not pre-treated [67]. Samples were acidified under vacuum with 100% ortho-phosphoric acid. The carbon isotopic composition of the skeleton ($\delta^{13}\text{C}$) was reported as the per mil deviation of the stable isotopes ^{13}C : ^{12}C relative to Vienna-Peedee Belemnite Limestone standard (v-PDB).

Skeletal oxygen isotopes ($\delta^{18}\text{O}$) were reported as the per mil deviation of the stable isotopes ^{18}O : ^{16}O relative to v-PDB. Approximately 10% of all samples were run in duplicate. The standard deviation (1σ) of repeated measurements of internal standards was $\pm 0.03\text{‰}$ for $\delta^{13}\text{C}$ and $\pm 0.07\text{‰}$ for $\delta^{18}\text{O}$ ($n = 55$).

Trace Element Analyses

From the solution used for $\delta^{11}\text{B}$ analysis, a 2–7 μL aliquot was diluted to a final concentration of 10 ppm Ca in 2% HNO_3 spiked with ~ 19 ppb Sc, 19 ppb Y, 0.19 ppb Pr, 0.095 ppb Bi, and 19 ppb V. Samples were then analysed for Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca on an X-Series 2 Quadrupole Inductively Coupled Plasma Mass Spectrometer (Q-ICPMS; Thermo Fisher Scientific) at the University of Western Australia using the standard Xt interface and the plasma screen fitted. Between-run reproducibility (RSD) of a coral standard subjected to the same chemistry as the coral samples was 0.4, 0.4, 0.8 and 0.4% for Sr/Ca, Mg/Ca, U/Ca and Ba/Ca, respectively ($n = 6$).

Statistical Analyses

Two-way analysis of variance (ANOVA) was used to test for the effects of temperature and time on coral chlorophyll *a*, $\delta^{11}\text{B}$, $\delta^{13}\text{C}$, $\delta^{18}\text{O}$, Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca. Temperature was fixed and fully crossed with two levels (treatment, control), and time was fixed and fully crossed with two levels (0, 6 weeks on the reef). Residual values for each variable and species were calculated and tested for normality using a Shapiro-Wilk's test and homogeneity of variance was assessed with plots of expected versus residual values. Non-normal data sets were transformed. Post hoc Tukey tests were performed when main effects – but no interaction terms – were significant. A posteriori slice tests (e.g., tests of simple effects) [68] determined if the treatment and control averages significantly differed at each time interval. Calcification data was previously analysed in Grottoli et al. [13].

Corals were considered to be fully recovered once average chlorophyll *a*, $\delta^{11}\text{B}$, $\delta^{13}\text{C}$, $\delta^{18}\text{O}$, Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca values of treatment corals no longer significantly differed from the average values of controls. Since all fragments were exposed to identical conditions except temperature during the tank portion of the experiment, any differences in the observed responses were due to temperature effects alone and independent of seasonal variation. Bonferroni corrections were not applied because they increase the risk of false negatives [69,70]. *P*-values ≤ 0.05 were considered significant. Statistical analyses were performed using SAS software, version 9.3 of the SAS System for Windows.

Results

Physiology

A significant interaction of temperature and time was observed for area-normalized chlorophyll *a* concentrations of *P. divaricata*, with treatment corals having 42% lower and 46% higher chlorophyll *a* concentrations than controls after 0 and 6 weeks on the reef, respectively (Fig. 1A, Table 1). Calcification rates were the same for both treatment and control *P. divaricata* throughout the study (Fig. 1B) [13].

In *P. astreoides*, area-normalized chlorophyll *a* concentrations were influenced by temperature, with treatment corals having 75% and 76% lower chlorophyll *a* rates than the controls at both 0 and 6 weeks on the reef, respectively (Fig. 1C, Table 1). Calcification rates of treatment corals were the same as in controls initially, but were 69% lower after 6 weeks on the reef (Fig. 1D) [13].

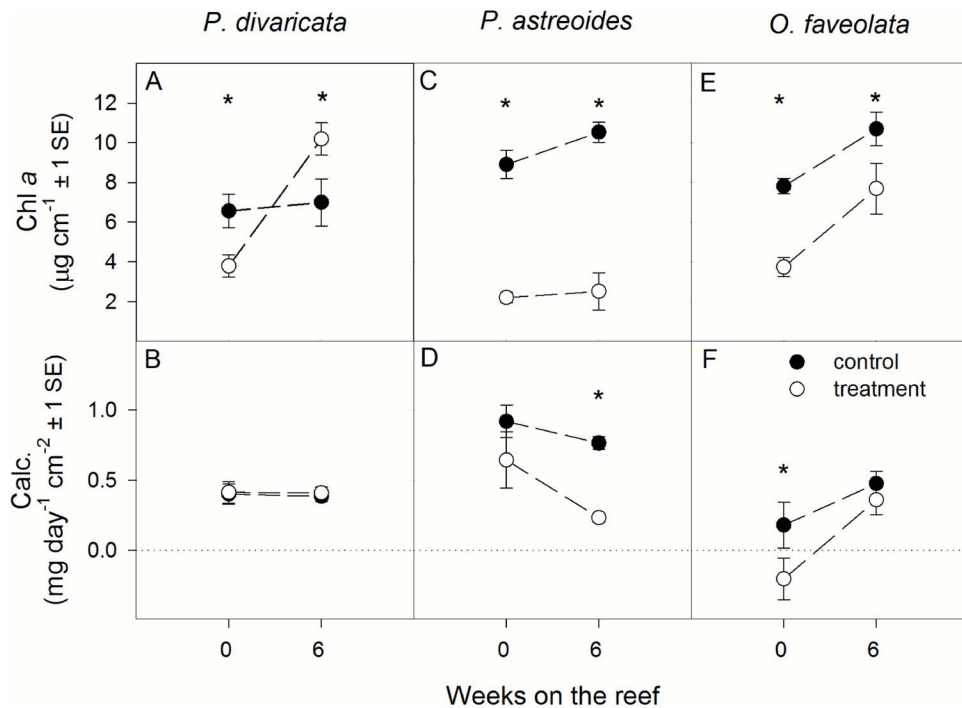


Figure 1. Average area-normalized chlorophyll *a* (=chl *a*) concentration and calcification (=calc.) rate of (A, B) *Porites divaricata*, (C, D) *Porites astreoides*, and (E, F) *Orbicella faveolata* after 0 and 6 weeks on the reef. Asterisks indicate significant differences between control and treatment corals at a specific time interval. Sample size ranges from 6–9. Calcification data reproduced from Grottoli et al. [13] with permission of the publisher.
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In *O. faveolata*, area-normalized chlorophyll *a* concentrations were influenced by both temperature and time (Table 1). They were 52% and 28% lower in treatment corals compared to controls at 0 and 6 weeks on the reef, respectively (Fig. 1E,

Table 1). Further, they were higher at 6 weeks on the reef compared to 0 weeks (Fig. 1E, Table 1). Calcification rates of treatment corals were significantly lower (–212%) than in controls initially but had fully recovered 6 weeks later (Fig. 1F) [13].

Table 1. Results of two-way ANOVA for area-normalized chlorophyll *a* content of *P. divaricata*, *P. astreoides*, and *O. faveolata*.

Variable	Effect	df	SS	F-statistic	p-value	Tukey
<i>P. divaricata</i>						
Chlorophyll <i>a</i>	Model	3, 28	164.4113	9.82	0.0002	
	Temp.	1	0.3307	0.06	0.8097	
	Time	1	82.8258	14.84	0.0007	
	Temp. x Time	1	63.1744	11.32	0.0025	
<i>P. astreoides</i>						
Chlorophyll <i>a</i>	Model	3, 33	469.1181	43.44	<0.0001	
	Temp.	1	456.8590	126.91	<0.0001	CO>TR
	Time	1	7.9244	2.20	0.1483	
	Temp. x Time	1	3.6588	1.02	0.3214	
<i>O. faveolata</i>						
Chlorophyll <i>a</i>	Model	3, 33	206.4635	12.44	<0.0001	
	Temp.	1	106.3292	19.22	0.0001	CO>TR
	Time	1	99.1921	17.93	0.0002	6>0
	Temp. x Time	1	2.4176	0.44	0.5136	

The effect of temperature (Temp.) was fixed with two levels (CO = control 30.4°C, TR = treatment 31.6°C), and time was fixed with 2 levels (0, 6 weeks). When main effects (but no interaction terms) were significant, Tukey post hoc results are shown. Significant *p*-values (*p*≤0.05) are highlighted in bold. df = degrees of freedom, SS = sum of squares of the effect.

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Isotopes

$\delta^{11}\text{B}$ was not influenced by temperature or time in any of the three species (Table 2, Fig. 2A, D, G). Similarly, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ of *P. divaricata* were not affected by either temperature or time (Table 2, Fig. 2B, C). However, a significant interaction of temperature and time was observed for $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ of *P. astreoides* (Table 2, Fig. 2E, F). Initially, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ did not differ between treatment and control corals, but after 6 weeks on the reef, both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ were significantly higher in treatment compared to control corals (Fig. 2E, F). In *O. faveolata*, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ were not influenced by temperature or time, but post hoc slice tests revealed that treatment corals had higher values than the controls at 0 weeks on the reef (Fig. 2H, I, Table 2).

Trace Elements

In *P. divaricata*, Sr/Ca, Mg/Ca, and U/Ca were not affected by either temperature or time (Table 3, Fig. 3A–C). However, Ba/Ca was affected by both temperature and time, with higher ratios found in controls than in treatment corals and increases in Ba/Ca after 6 weeks on the reef compared to 0 weeks (Table 3, Fig. 3D).

In *P. astreoides*, Sr/Ca, Mg/Ca and Ba/Ca were not affected by either temperature or time (Table 3, Fig. 3E, F, H). However, post hoc slice tests revealed that Mg/Ca ratios were higher

(+16.5%) in treatment than in control corals at 0 weeks (Table 3, Fig. 3F). A significant interaction between temperature and time was observed for U/Ca, with ratios being significantly higher (+20.3%) in treatment than in control corals after 6 weeks on the reef (Table 3, Fig. 3G).

In *O. faveolata*, Sr/Ca, Mg/Ca and U/Ca were not influenced by either temperature or time (Table 3, Fig. 3I–K). However, post hoc slice tests revealed that treatment Mg/Ca ratios were lower (−26.3%) than the controls at 0 weeks on the reef (Fig. 3J). A significant interaction of temperature and time was observed for Ba/Ca, with treatment values being higher (+5.8%) than the control initially, but no longer different after 6 weeks on the reef (Fig. 3L, Table 3).

Discussion

Isotopes

Coral skeletal boron isotopes have been established as a proxy for seawater pH, yet it remains unclear if and how this proxy is affected by seawater temperature. Specifically, it has never been tested whether coral bleaching due to elevated temperature affects coral $\delta^{11}\text{B}$. Here, we show for the first time that experimental coral bleaching does not affect $\delta^{11}\text{B}$ of three Caribbean coral species

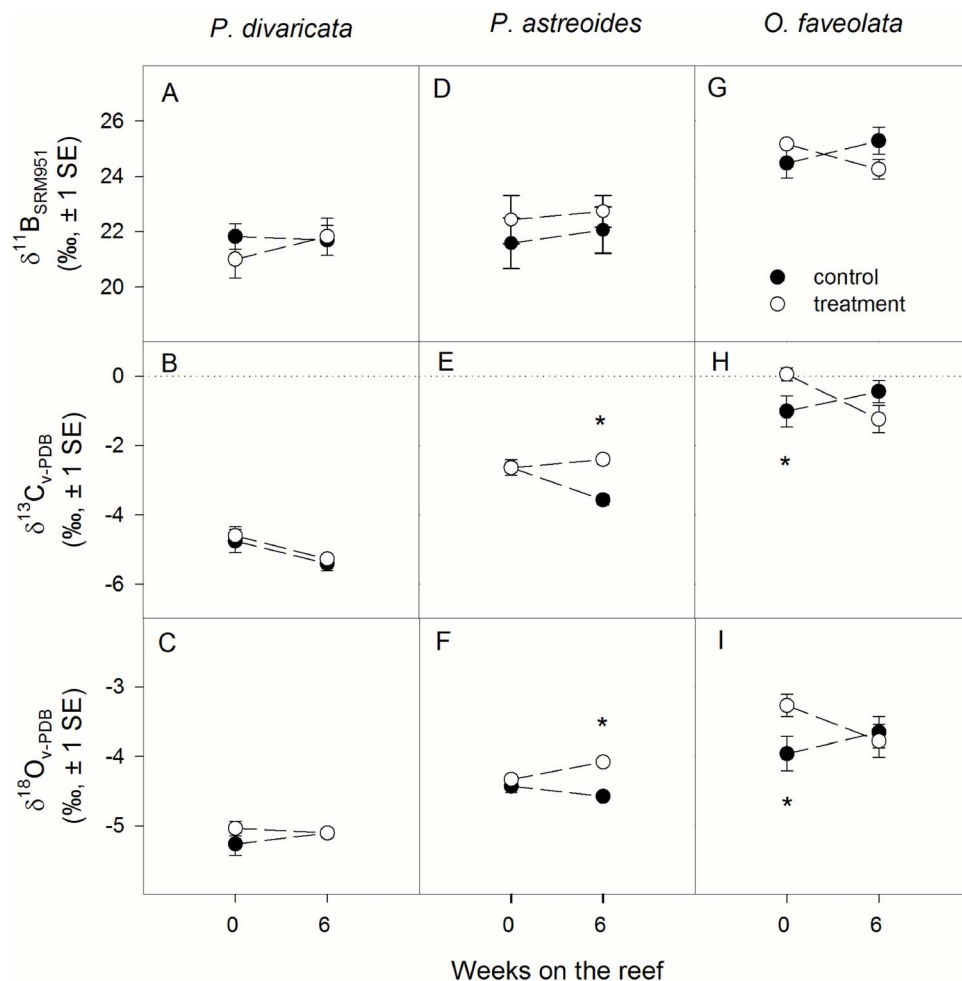


Figure 2. Average $\delta^{11}\text{B}$, $\delta^{13}\text{C}$, and $\delta^{18}\text{O}$ of (A–C) *Porites divaricata*, (D–F) *Porites astreoides*, and (G–I) *Orbicella faveolata* after 0 and 6 weeks on the reef. Asterisks indicate significant differences between control and treatment corals at a specific time interval. Sample size ranges from 3–9.

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Table 2. Results of two-way ANOVAs for $\delta^{11}\text{B}$, $\delta^{13}\text{C}$, and $\delta^{18}\text{O}$ of *P. divaricata*, *P. astreoides*, and *O. faveolata*.

Variable	Effect	df	SS	F-statistic	p-value
<i>P. divaricata</i>					
$\delta^{11}\text{B}$	Model	3, 19	2.7948	0.46	0.7148
	Temp.	1	0.5602	0.28	0.6066
	Time	1	0.5602	0.28	0.6066
	Temp. x Time	1	1.0643	0.52	0.4795
$\delta^{13}\text{C}$	Model	3, 23	2.8226	2.76	0.0689
	Temp.	1	0.1219	0.36	0.5565
	Time	1	2.5603	7.51	0.0126
	Temp. x Time	1	0.0008	0.00	0.9610
$\delta^{18}\text{O}$	Model	3, 23	0.1570	0.89	0.4619
	Temp.	1	0.0778	1.33	0.2628
	Time	1	0.0111	0.19	0.6683
	Temp. x Time	1	0.0778	1.33	0.2628
<i>P. astreoides</i>					
$\delta^{11}\text{B}$	Model	3, 22	3.8106	0.32	0.8094
	Temp.	1	3.2908	0.83	0.3725
	Time	1	0.8574	0.22	0.6464
	Temp. x Time	1	0.0420	0.01	0.9189
$\delta^{13}\text{C}$	Model	3, 34	7.2196	10.56	<0.0001
	Temp.	1	2.9506	12.95	0.0011
	Time	1	1.0442	4.58	0.0403
	Temp. x Time	1	3.0344	13.32	0.0010
$\delta^{18}\text{O}$	Model	3, 34	1.1779	10.85	<0.0001
	Temp.	1	0.7728	21.36	<0.0001
	Time	1	0.0240	0.66	0.4217
	Temp. x Time	1	0.3460	9.56	0.0042
<i>O. faveolata</i>					
$\delta^{11}\text{B}$	Model	3, 13	2.6525	1.25	0.3430
	Temp.	1	0.0981	0.14	0.7173
	Time	1	0.0086	0.01	0.9145
	Temp. x Time	1	2.5358	3.59	0.0876
$\delta^{13}\text{C}$	Model	3, 33	8.3114	2.56	0.0739
	Temp.	1	0.1528	0.14	0.7100
	Time	1	1.0742	0.99	0.3275
	Temp. x Time	1	7.3876	6.82	0.0140
$\delta^{18}\text{O}$	Model	3, 33	2.1365	1.69	0.1903
	Temp.	1	0.6800	1.61	0.2138
	Time	1	0.0845	0.20	0.6576
	Temp. x Time	1	1.4076	3.34	0.0776

The effect of temperature (Temp.) was fixed with two levels (control 30.4°C, treatment 31.6°C), and time was fixed with 2 levels (0, 6 weeks). Significant *p*-values ($p \leq 0.05$) are highlighted in bold. df = degrees of freedom, SS = sum of squares of the effect.

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(Fig. 2A, D, G). This finding does not support our initial hypothesis based on evidence presented by Wei et al. [47] that coral $\delta^{11}\text{B}$ may decrease significantly during coral bleaching events. However, it is in agreement with experimental studies showing that $\delta^{11}\text{B}$ is not affected by moderate increases in temperatures from 25°C to 28°C [43,44], which do not induce coral bleaching.

Coral bleaching is caused by a significant loss of algal endosymbionts and/or photosynthetic pigments from the coral

tissue [7,9,12]. In the current study, treatment corals of all species had significantly lower area-normalized chlorophyll *a* levels than controls throughout the study (Fig. 1A, C, E), primarily driven by lower endosymbiont densities [13]. Further, treatment corals of all species had a significantly lower maximum quantum yield of photosystem II (F_v/F_m) than controls for up to 6 weeks on the reef [59]. Thus, treatment corals of all three species were bleached. This demonstrates that even though corals experienced significant

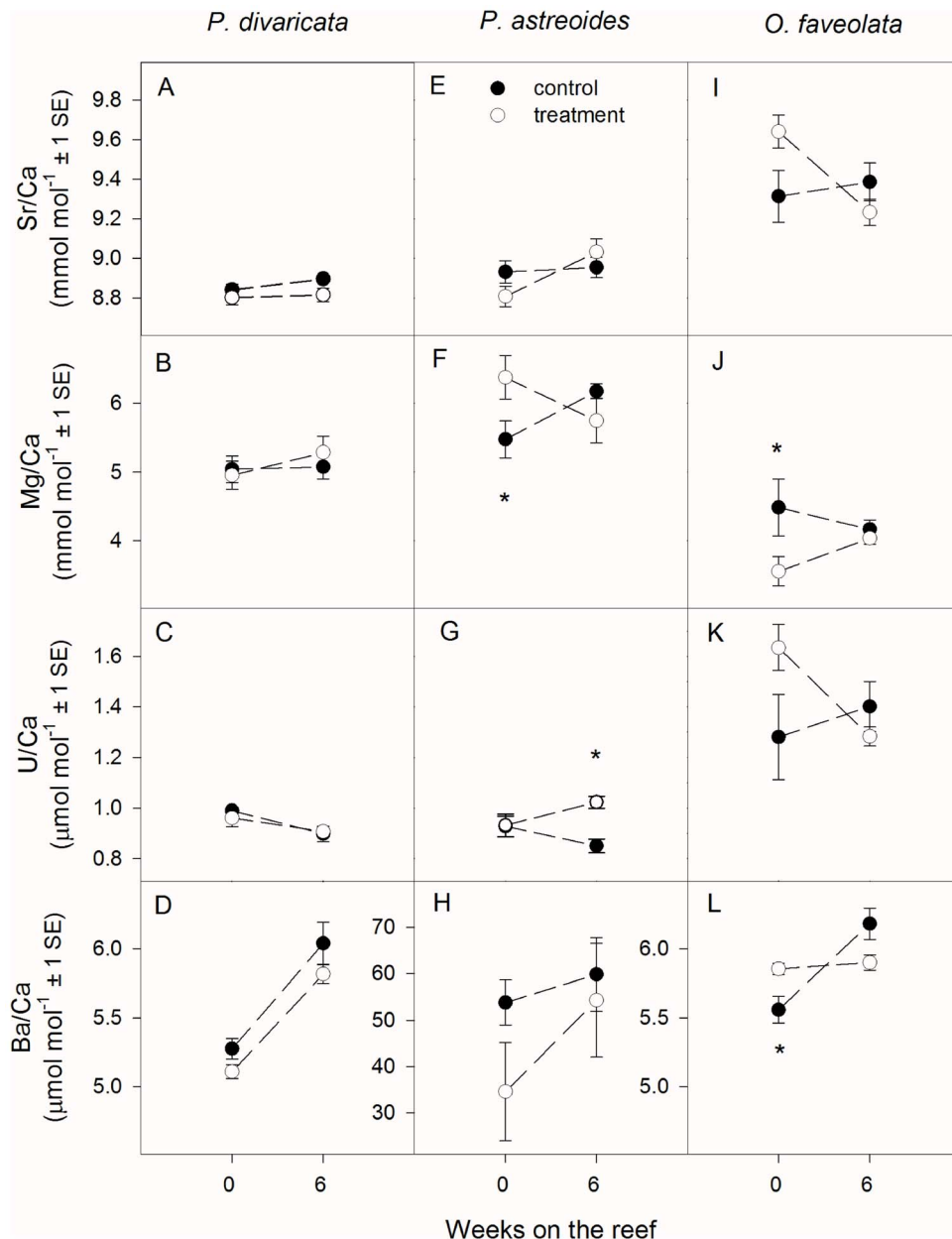


Figure 3. Average Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca of (A–D) *Porites divaricata*, (E–H) *Porites astreoides*, and (I–L) *Orbicella faveolata* after 0 and 6 weeks on the reef. Asterisks indicate significant differences between control and treatment corals at a specific time interval. Sample size ranges from 3–7. Note that the Y-axis for Ba/Ca differs for *P. astreoides* and the other two species.
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temperature stress and bleaching, coral skeletal $\delta^{11}\text{B}$ remained unaffected.

Calcification rates are often compromised in bleached corals [17,52,54,71], and may therefore have affected the response of skeletal isotopes and trace elements [72,73]. Indeed, calcification rates of treatment *O. faveolata* were significantly lower than in the controls at 0 weeks on the reef (Fig. 1D). In *P. astreoides*, calcification was not significantly lower than the controls until after 6 weeks on the reef (Fig. 1F) because the bleaching response of the animal host showed a lag of several weeks in this coral species [13]. Such lags in the bleaching response have been previously observed in three species of Hawaiian corals as well [17,52]. However, none of these declines in calcification resulted in significant differences

in $\delta^{11}\text{B}$ between treatment and control corals (Fig. 2D, G) despite the fact that treatment corals were significantly bleached at this time (Fig. 1C, E). Similarly, when calcification rates were maintained in bleached corals, as was the case in *P. divaricata* and in *P. astreoides* initially at the end of the heating period (Fig. 1B, D), $\delta^{11}\text{B}$ of treatment corals did not differ from the controls (Fig. 2A, D). Based on these results, coral $\delta^{11}\text{B}$ is unaffected by calcification rate.

The insensitivity of $\delta^{11}\text{B}$ to calcification rates is further corroborated by the fact that both skeletal $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ changed in response to decreases in calcification rates in two species, yet $\delta^{11}\text{B}$ did not (Figs. 1, 2). In treatment *P. astreoides* and *O. faveolata*, both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ increased as calcification

Table 3. Results of two-way ANOVAs for Mg/Ca, Sr/Ca, Ba/Ca, and U/Ca of *P. divaricata*, *P. astreoides*, and *O. faveolata*.

Variable	Effect	df	SS	F-statistic	p-value	Tukey
<i>P. divaricata</i>						
Sr/Ca	Model	3, 19	0.0241	1.49	0.2557	
	Temp.	1	0.0170	3.15	0.0950	
	Time	1	0.0055	1.02	0.3273	
	Temp. x Time	1	0.0019	0.35	0.5622	
Mg/Ca	Model	3, 19	0.2820	0.42	0.7427	
	Temp.	1	0.0170	0.08	0.7868	
	Time	1	0.1575	0.70	0.4152	
	Temp. x Time	1	0.1045	0.46	0.5054	
U/Ca	Model	3, 19	0.0264	2.02	0.1520	
	Temp.	1	0.0005	0.10	0.7504	
	Time	1	0.0252	5.78	0.0287	
	Temp. x Time	1	0.0017	0.39	0.5435	
Ba/Ca	Model	3, 19	2.8756	27.77	<0.0001	
	Temp.	1	0.1797	5.21	0.0365	CO>TR
	Time	1	2.5673	74.38	<0.0001	6>0
	Temp. x Time	1	0.0038	0.11	0.7456	
<i>P. astreoides</i>						
Sr/Ca	Model	3, 22	0.1453	2.69	0.0751	
	Temp.	1	0.0029	0.16	0.6926	
	Time	1	0.0860	4.78	0.0415	
	Temp. x Time	1	0.0570	3.17	0.0911	
Mg/Ca	Model	3, 22	2.7689	2.47	0.0929	
	Temp.	1	0.3173	0.85	0.3681	
	Time	1	0.0069	0.02	0.8934	
	Temp. x Time	1	2.5033	6.71	0.0180	
U/Ca	Model	3, 22	0.0872	4.11	0.0210	
	Temp.	1	0.0442	6.24	0.0218	
	Time	1	0.0003	0.04	0.8529	
	Temp. x Time	1	0.0398	5.61	0.0286	
Ba/Ca	Model	3, 21	2242.8826	1.63	0.2181	
	Temp.	1	808.5285	1.76	0.2012	
	Time	1	871.9859	1.90	0.1851	
	Temp. x Time	1	246.0976	0.54	0.4736	
<i>O. faveolata</i>						
Sr/Ca	Model	3, 13	0.2863	2.48	0.1206	
	Temp.	1	0.0259	0.67	0.4308	
	Time	1	0.0962	2.50	0.1447	
	Temp. x Time	1	0.1983	5.16	0.0464	
Mg/Ca	Model	3, 13	1.5265	1.94	0.1879	
	Temp.	1	0.9719	3.70	0.0835	
	Time	1	0.0238	0.09	0.7696	
	Temp. x Time	1	0.5522	2.10	0.1779	
U/Ca	Model	3, 13	0.2635	1.69	0.2309	
	Temp.	1	0.0479	0.92	0.3591	
	Time	1	0.0458	0.88	0.3696	
	Temp. x Time	1	0.1922	3.71	0.0831	
Ba/Ca	Model	3, 13	0.7797	8.77	0.0038	
	Temp.	1	0.0001	0.00	0.9463	
	Time	1	0.3844	12.97	0.0048	

Table 3. Cont.

Variable	Effect	df	SS	F-statistic	p-value	Tukey
	Temp. x Time	1	0.2847	9.61	0.0113	

The effect of temperature (Temp.) was fixed with two levels (CO = control 30.4°C, TR = treatment 31.6°C), and time was fixed with 2 levels (0, 6 weeks). When main effects (but no interaction terms) were significant, Tukey post hoc results are shown. Significant *p*-values (*p*≤0.05) are highlighted in bold. df = degrees of freedom, SS = sum of squares of the effect.

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decreased. This is unexpected given that both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ are assumed to decrease when photosynthesis decreases and seawater temperature increases (provided there were no changes in salinity), respectively [74–79]. Instead, the enrichment in both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ indicates that skeletal carbonate started approaching isotopic equilibrium with seawater as calcification rates slowed down [76] and that calcification rate was the primary control on $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$. Similar relationships between skeletal $\delta^{13}\text{C}$, $\delta^{18}\text{O}$, and calcification were also noted in other corals when bleached [52,80]. For *O. faveolata*, this strongly suggests that some skeletal material was deposited during the bleaching treatment, even though on average net calcification rates were slightly negative. Further, in *P. divaricata*, $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ were unaffected by bleaching at any point, which is consistent with no changes in calcification rates.

Clearly, the carbon and oxygen isotopic signal was dominated by kinetic effects associated with calcification rates [76] in all three species. Kinetic effects likely overpowered any metabolic effects in $\delta^{13}\text{C}$ due to reduced chlorophyll *a* concentrations (implying reduced photosynthesis rates) as well as the expected temperature-dependent effect on $\delta^{18}\text{O}$ [74]. Although a simple data correction has been proposed to correct for kinetic isotope effects in $\delta^{13}\text{C}$ [81], it does not reliably remove kinetic isotope effects in bleached corals [82]. Unfortunately, these findings confirm that coral $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ cannot be used as a proxy for past bleaching events [17,52,54,55], and that coral $\delta^{11}\text{B}$ does not record bleaching events as suggested by Wei et al. [47], at least not in the coral species studied here.

The observed lack of a bleaching effect on coral $\delta^{11}\text{B}$ suggests that coral bleaching – and therefore the physiological status of the algal endosymbiont – does not affect pH-upregulation at the site of calcification. This is surprising given that coral calcification is enhanced in the light via photosynthesis [83–85] due to a variety of mechanisms including removal of protons from the site of calcification, supply of ATP for ion transport and/or organic matrix synthesis, production of oxygen, supply of precursors for organic matrix production, and removal of phosphates [86,87]. Thus, if the symbiotic relationship is disrupted during bleaching, any of the mechanisms listed above would likely be interrupted as well, including the impaired removal of protons from the site of calcification that affects $\delta^{11}\text{B}$ values. Clearly, this was not the case in the coral species studied here, which further suggests that decreased calcification rates of bleached corals may not be due to impaired control over the calcifying environment.

Heterotrophy and energy reserves can promote many physiological aspects including calcification in healthy corals [88], and also enhances resistance to and recovery from coral bleaching [13,18,89,90]. It is therefore possible that heterotrophic energy input may have helped bleached corals to maintain pH-upregulation at the site of calcification. However, feeding rates and total carbon acquired by feeding relative to metabolic demand (CHAR) did not differ between treatment and control corals immediately after the bleaching treatment in any of the species

[13]. It is possible that other heterotrophic sources of carbon such as dissolved and particulate organic carbon (DOC and POC, respectively) could be providing the necessary supplemental energy for calcification in bleached corals. For example, Levas [57] showed that DOC can provide 15% of daily metabolic carbon in bleached *O. faveolata* corals.

Collectively, these results demonstrate that pH-upregulation at the site of calcification is robust to significant physiological changes caused by short bleaching events. This suggests either that this process has high priority during energy allocation even under scenarios of resource limitation such as coral bleaching, or that pH-upregulation does not require that much energy and can thus be maintained during coral bleaching. However, it is possible that when coral bleaching events are prolonged, a certain threshold of resource limitation may be reached at which corals are no longer able to maintain pH-upregulation at the site of calcification. Resource limitation in this scenario would not only be related to a decrease in transferred carbon by the algal endosymbiont [18,91], but also to oxygen limitation [92,93] and potentially other mechanisms involved in light-enhanced calcification that would be interrupted due to bleaching. This would then result in decreased $\delta^{11}\text{B}$ values of bleached corals, consistent with observations by Wei et al. [47] for a *Porites* coral core, where significant decreases in boron isotopes were interpreted as corresponding to the 1998 mass bleaching event on the Great Barrier Reef. This threshold, if existent, would likely be on the order of at least several months because the physiology of *P. astreoides* in this study was affected for at least 6 weeks [13], yet showed no decrease in boron isotopes. Further, several studies have shown that coral growth rates can take several years to fully recover from bleaching [71,80,94,95].

Trace Elements

Coral bleaching did not affect trace element composition (Sr/Ca, Mg/Ca, U/Ca, and Ba/Ca) of *P. divaricata*, but influenced Mg/Ca and U/Ca ratios of *P. astreoides* as well as Mg/Ca and Ba/Ca in *O. faveolata* (Fig. 3). The lack of any temperature effect on Sr/Ca is particularly surprising, because it is generally considered to be one of the most reliable sea surface temperature (SST) proxies available [96,97,98], although it is not free of complications [99–105]. Sr/Ca sensitivity is about 0.04–0.08 mmol mol⁻¹ per °C [100,101,106–109]. Thus the treatment corals in this study were expected to have Sr/Ca ratios that were 0.05–0.1 mmol mol⁻¹ lower than the control corals given the inverse correlation between temperature and Sr/Ca. At 0 weeks on the reef, treatment *P. divaricata* and *P. astreoides* had indeed lower Sr/Ca ratios than the controls (0.04 and 0.12 mmol mol⁻¹, respectively), whereas treatment *O. faveolata* had higher Sr/Ca values (0.33 mmol mol⁻¹). However, none of these differences were statistically significant (Fig. 3A, E, I).

During cold and hot temperature stress, sharp increases in Sr/Ca have been observed in *Porites* coral from the Great Barrier Reef [53,96]. Marshall and McCulloch [53] proposed that

temperature stress disrupts the Sr/Ca – SST relationship by inhibiting Ca transport enzymes, whereas Sr transport is unaffected. Thus, the Sr/Ca ratio increases because relatively less Ca is made available to the calcifying fluid. Similarly, Cohen et al. [99] suggested that endosymbiont photosynthesis enhances Ca but not Sr transport to the calcifying fluid, thus resulting in lower Sr/Ca ratios during times of high photosynthesis. Following this reasoning, Sr/Ca should therefore increase in bleached corals when photosynthesis is impaired and calcification slows down. However, the findings from this study do not generally support this hypothesis. Even though Sr/Ca ratios were higher in treatment than in control *O. faveolata* initially (Fig. 3I) when chlorophyll *a* concentrations and calcification rates were significantly lower (Fig. 1F), this difference was not statistically significant ($p = 0.0536$). Marshall and McCulloch [53] suggested that a particular temperature threshold needs to be reached before Sr/Ca ratios are affected by temperature stress. Potentially, this threshold differs for different coral species and therefore resulted in equivocal results here.

Changes in calcification rates due to coral bleaching may have influenced Sr/Ca [100,101,110]. Slower growing corals typically have higher Sr/Ca values than faster growing corals [100,101,110]. However, this was not generally observed, and many other studies also failed to detect growth rate effects on Sr/Ca [102,106,111–113]. It therefore seems that overall coral skeletal Sr/Ca is not influenced by the physiological changes or temperature changes occurring during short coral bleaching events.

Mg/Ca ratios were affected by coral bleaching in *P. astreoides* and *O. faveolata*, but not *P. divaricata* (Fig. 3B, F, J). Sensitivity of Mg/Ca is about $0.09\text{--}0.16\text{ mmol mol}^{-1}\text{ per }^{\circ}\text{C}$ [108,109,114], and therefore treatment corals were expected to have Mg/Ca ratios that were higher by $0.1\text{--}0.2\text{ mmol mol}^{-1}$ compared to control corals at 0 weeks on the reef. Although treatment *P. astreoides* had significantly higher Mg/Ca ratios than control corals at this time point (Fig. 3F), the difference was 0.9 mmol mol^{-1} , which is much higher than expected based on temperature dependence alone. Further, Mg/Ca decreased with bleaching in *O. faveolata* instead of the expected increase (Fig. 3J). However, the reliability of Mg/Ca as a SST proxy has been questioned by many studies [98,108,112]. Mg/Ca ratios showed similar patterns as calcification rates in both *P. divaricata* and *O. faveolata* and could therefore be mainly controlled by calcification rate. However, they were not significantly correlated in either species (Spearman's $r = 0.40$, $p = 0.10$ and $r = 0.63$, $p = 0.13$, respectively). They were also not significantly correlated in *P. astreoides* ($r = -0.06$, $p = 0.84$). It is therefore unlikely that Mg/Ca ratios are primarily controlled by calcification rate.

Sensitivity of U/Ca is about $0.03\text{--}0.05\text{ }\mu\text{mol mol}^{-1}\text{ per }^{\circ}\text{C}$ [115] and treatment corals were therefore expected to have U/Ca ratios that are $0.04\text{--}0.06\text{ }\mu\text{mol mol}^{-1}$ lower than in control corals at 0 weeks on the reef. However, either no change or the opposite (i.e., increases in U/Ca) was observed. It is therefore likely that seawater temperature is not the primary control on coral U/Ca. Potential salinity [115] and pH effects [115,116] can be excluded as treatment and control corals were kept in the same seawater (except for temperature). Coral $\delta^{11}\text{B}$ further suggests that the pH of the calcifying fluid did not significantly differ between treatment and control corals (Fig. 2).

Although Ba/Ca ratios are typically used as a proxy for upwelling of nutrient-rich water [108,117,118] and river input and flood events [119,120], in some species Ba/Ca ratios can also be influenced by SST [117,121]. If that were the case in the present study, treatment corals would be expected to have lower Ba/Ca

ratios than control corals at 0 weeks on the reef. However, this was not generally the case. As all corals were kept in the same seawater and location on the reef, both river input and temperature effects are unlikely to have caused the observed pattern.

Interestingly, Ba/Ca ratios of *P. astreoides* were an order of magnitude higher than in either *P. divaricata* or *O. faveolata*, showed high within-treatment variability, and were also much higher than typical flood or upwelling related Ba signals in corals (Fig. 3H) [118,119,120]. Considering that all three coral species were exposed to identical environmental conditions except for temperature during the bleaching treatment, this finding is difficult to explain and cannot be related to seawater Ba concentrations. Even in exceptionally high Ba seawater, coral skeletal Ba/Ca concentrations do not exceed $6.5\text{ }\mu\text{mol mol}^{-1}$ [122]. However, similar differences in Ba/Ca between different coral genera have also been observed in another study [123].

One possibility is that particulates and/or organic phases rich in Ba may have become trapped within the skeleton [124,125]. Due to its encrusting to mounding morphology, sediment settles more easily on *P. astreoides* compared to the other two species (V. Schoepf, pers. observation) and may somehow result in enriched Ba concentrations in the surrounding seawater and/or skeleton. Spawning has also been hypothesized to influence Ba/Ca ratios [125], but the seasonal peak in reproduction occurs in April for *P. astreoides* [126], whereas the corals here were collected in August and September, respectively. Overall, these findings add to the existing literature of anomalously high Ba concentrations in corals, which at present cannot be satisfactorily explained [118,124,125,127].

Overall, trace element data often showed trends that were inconsistent with either changes in seawater temperature or the observed physiological effects. This was in stark contrast to the isotopic data, which showed consistent trends across species, with calcification rate being the main influence on $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ but not $\delta^{11}\text{B}$ in bleached corals. Given that it is difficult to explain these inconsistencies based on our current understanding of trace elements, these findings demonstrate that the mechanisms of trace element incorporation in bleached corals are poorly understood and that they are likely species-specific. Species-specific differences in the bleaching response of the three coral species studied here [13] may in part be responsible for the observed inconsistencies. For example, *P. divaricata* was least affected by bleaching whereas *P. astreoides* suffered the largest declines of all three species in both symbiont and host performance (Fig. 1A–D) [13]. In contrast, *O. faveolata* showed large health declines initially but was able to recover calcification rates, energy reserves, and endosymbiont densities within 6 weeks (Fig. 1E, F) [13]. Similarly, trace elements showed no significant temperature effects and little within-treatment variability in *P. divaricata* whereas temperature effects were often evident in the other two species (Fig. 3). Although the magnitude and direction of these effects were typically inconsistent with our current knowledge about trace element incorporation and often differed between *P. astreoides* and *O. faveolata* (see above), this could nevertheless indicate that more severely bleached corals may show larger excursions in their trace elements. This study therefore highlights the need to study trace element incorporation under stressful temperatures in a variety of coral species, especially in the context of accompanying physiological information.

Implications for Paleo-Climate Reconstruction

The present study provides the first experimental evidence that short coral bleaching events do not affect boron isotopes in three Caribbean coral species. This is generally good news because it

suggests that past seawater pH conditions can be reconstructed without any conflicting effects introduced by significant physiological changes due to short bleaching events. This is particularly important for massive coral species such as *O. faveolata* or *Porites* sp., which are frequently used for paleo-pH reconstruction. However, at the same time, this study indicates that coral $\delta^{11}\text{B}$ cannot be used as a novel proxy for past bleaching events as suggested by Wei et al. [47]. Similarly, our findings confirm that $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ cannot be used as bleaching proxies when calcification rates are compromised, demonstrating that there is currently no reliable proxy available to identify past bleaching events. However, future studies are needed to confirm these findings for prolonged coral bleaching events.

Regarding trace elements, Sr/Ca of bleached corals did not consistently record the 1.2°C difference in seawater temperature during the bleaching treatment, or alternatively show a consistent increase due to impaired photosynthesis and calcification. These findings suggest that the mechanisms of Sr/Ca incorporation are not well understood at temperatures that are stressful to corals. Further, the observed changes in Mg/Ca, U/Ca and Ba/Ca due to coral bleaching could not be satisfactorily explained using either temperature dependence or changes in coral physiology. There-

fore, it is likely that additional factors influence these geochemical proxies and that these factors are species-specific. It is therefore not recommended to use corals with a history of bleaching for paleo-climate reconstruction using trace elements.

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Author Contributions

Conceived and designed the experiments: AGG MEW VS. Performed the experiments: VS SJL MDA YM AGG MEW. Analyzed the data: VS. Contributed reagents/materials/analysis tools: VS MTM MEW SJL YM MDA AGG. Wrote the paper: VS MTM MEW SJL YM MDA AGG.

References

- IPCC (2013) Climate Change 2013: The physical science basis. Summary for Policy Makers. Available: <http://www.ipcc.ch>. Accessed 2014 Oct 26.
- Hoegh-Guldberg O, Bruno JF (2010) The impact of climate change on the world's marine ecosystems. *Science* 328: 1523–1528.
- Harvey BP, Gwynn-Jones D, Moore PJ (2013) Meta-analysis reveals complex marine biological responses to the interactive effects of ocean acidification and warming. *Ecol Evol* 3: 1016–1030.
- Wild C, Hoegh-Guldberg O, Naumann MS, Colombo-Pallotta MF, Ateweberhan M, et al. (2011) Climate change impedes scleractinian corals as primary reef ecosystem engineers. *Mar Freshwater Res* 62: 205–215.
- Kleypas J, Buddemeier RW, Archer D, Gattuso J-P, Langdon C, et al. (1999) Geochemical consequences of increased atmospheric carbon dioxide on coral reefs. *Science* 284: 118–120.
- Hoegh-Guldberg O, Mumby PJ, Hooten AJ, Steneck R, Greenfield P, et al. (2007) Coral reefs under rapid climate change and ocean acidification. *Science* 318: 1737–1742.
- Hoegh-Guldberg O, Smith GJ (1989) The effect of sudden changes in temperature, light, and salinity on the population density and export of zooxanthellae from the reef coral *Stylophora pistillata* Esper and *Seriatopora hystrix* Dana. *J Exp Mar Biol Ecol* 129: 279–303.
- Fitt WK, Brown BE, Warner ME, Dunne RP (2001) Coral bleaching: interpretation of thermal tolerance limits and thermal thresholds in tropical corals. *Coral Reefs* 20: 51–65.
- Jokiel PL, Coles SL (1990) Response of Hawaiian and other Indo-Pacific reef corals to elevated temperature. *Coral Reefs* 8: 155–162.
- Baker AC, Glynn PW, Riegl B (2008) Climate change and coral reef bleaching: An ecological assessment of long-term impacts, recovery trends and future outlook. *Estuar Coast Shelf Sci* 80: 435–471.
- Hoegh-Guldberg O (1999) Climate change, coral bleaching and the future of the world's coral reefs. *Mar Freshwater Res* 50: 839–866.
- Glynn PW (1996) Coral reef bleaching: facts, hypotheses and implications. *Global Change Biol* 2: 495–509.
- Grottoli AG, Warner M, Levas SJ, Aschaffenburg M, Schoepf V, et al. (2014) The cumulative impact of annual coral bleaching can turn some coral species winners into losers. *Global Change Biol*. doi: 10.1111/gcb.12658.
- Nakamura T, Yamasaki H, van Woesik R (2003) Water flow facilitates recovery from bleaching in the coral *Stylophora pistillata*. *Mar Ecol Prog Ser* 256: 287–291.
- Fitt WK, Spero HJ, Halas J, White MW, Porter JW (1993) Recovery of the coral *Montastraea annularis* in the Florida Keys after the 1987 Caribbean "bleaching event". *Coral Reefs* 12: 57–64.
- Rodrigues IJ, Grottoli AG (2007) Energy reserves and metabolism as indicators of coral recovery from bleaching. *Limnol Oceanogr* 52: 1874–1882.
- Levas SJ, Grottoli AG, Hughes AD, Osburn CL, Matsui Y (2013) Physiological and biogeochemical traits of bleaching and recovery in the mounding species of coral *Porites lobata*: Implications for resilience in mounding corals. *PLoS One* 8: e63267. doi:10.1371/journal.pone.0063267.
- Grottoli AG, Rodrigues IJ, Palardy JE (2006) Heterotrophic plasticity and resilience in bleached corals. *Nature* 440: 1186–1189.
- Loya Y, Sakai K, Yamazato K, Nakan Y, Sambali H, et al. (2001) Coral bleaching: the winners and the losers. *Ecol Lett* 4: 122–131.
- Donner SD (2009) Coping with commitment: Projected thermal stress on coral reefs under different future scenarios. *PLoS One* 4: e5712. doi:10.1371/journal.pone.0005712.
- Teneva L, Karnauskas M, Logan C, Bianucci L, Currie J, et al. (2012) Predicting coral bleaching hotspots: the role of regional variability in thermal stress and potential adaptation rates. *Coral Reefs* 31: 1–12.
- Frieler K, Meinshausen M, Golly A, Mengel M, Lebek K, et al. (2013) Limiting global warming to 2°C is unlikely to save most coral reefs. *Nat Clim Change* 3: 165–170.
- Wilkinson C (2008) Status of coral reefs of the world: 2008. Global Coral Reef Monitoring Network and Reef and Rainforest Research Centre. 296 p.
- Carpenter KE, Abrar M, Aeby G, Aronson RB, Banks S, et al. (2008) One-third of reef-building corals face elevated extinction risk from climate change and local impacts. *Science* 321: 560–563.
- Orr JC, Fabry VJ, Aumont O, Bopp L, Doney SC, et al. (2005) Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature* 437: 681–686.
- Caldeira K, Wickett ME (2003) Anthropogenic carbon and ocean pH. *Nature* 425: 365.
- Langdon C, Atkinson MJ (2005) Effect of elevated pCO₂ on photosynthesis and calcification on corals and interactions with seasonal change in temperature/irradiance and nutrient enrichment. *J Geophys Res* 110: C09S07. doi:10.1029/2004JC002576.
- Holcomb M, McCorkle DC, Cohen AL (2010) Long-term effects of nutrient and CO₂ enrichment on the temperate coral *Astrangia poculata* (Ellis and Solander, 1786). *J Exp Mar Biol Ecol* 386: 27–33.
- Schoepf V, Grottoli AG, Warner M, Cai W-J, Melman TF, et al. (2013) Coral energy reserves and calcification in a high-CO₂ world at two temperatures. *PLoS One* 8: e75049. doi:10.1371/journal.pone.0075049.
- Marubini F, Ferrier-Pages C, Cuif J-P (2003) Suppression of skeletal growth in scleractinian corals by decreasing ambient carbonate-ion concentration: a cross-family comparison. *Proc R Soc B* 270: 179–184.
- Krief S, Hendy EJ, Fine M, Yam R, Meibom A, et al. (2010) Physiological and isotopic responses of scleractinian corals to ocean acidification. *Geochim Cosmochim Acta* 74: 4988–5001.
- Edmunds PJ, Brown D, Moriarty V (2012) Interactive effects of ocean acidification and temperature on two scleractinian corals from Moorea, French Polynesia. *Global Change Biol* 18: 2173–2183.
- Comeau S, Edmunds PJ, Spindel NB, Carpenter RC (2013) The responses of eight coral reef calcifiers to increasing partial pressure of CO₂ do not exhibit a tipping point. *Limnol Oceanogr* 58: 388–398.
- Trotter J, Montagna P, McCulloch MT, Silenzi S, Reynaud S, et al. (2011) Quantifying the pH 'vital effect' in the temperate zooxanthellate coral *Cladocora caespitosa*: Validation of the boron seawater pH proxy. *Earth Planet Sci Lett* 303: 163–173.
- Holcomb M, Venn AA, Tambutte E, Tambutte S, Allemand D, et al. (2014) Coral calcifying fluid pH dictates response to ocean acidification. *Sci Rep* 4. doi:10.1038/srep05207.
- Ries JB (2011) A physicochemical framework for interpreting the biological calcification response to CO₂-induced ocean acidification. *Geochim Cosmochim Acta* 75: 4053–4064.

37. Venn AA, Tambutte E, Holcomb M, Laurent J, Allemand D, et al. (2013) Impact of seawater acidification on pH at the tissue-skeleton interface and calcification in reef corals. *Proc Natl Acad Sci USA* 110: 1634–1639.
38. Venn A, Tambutte E, Holcomb M, Allemand D, Tambutte S (2011) Live tissue imaging shows reef corals elevate pH under their calcifying tissue relative to seawater. *PLoS One* 6: e20013. doi: 10.1371/journal.pone.0020013.
39. Al-Horani FA, Al-Moghrabi SM, de Beer D (2003) The mechanism of calcification and its relation to photosynthesis and respiration in the scleractinian coral *Galaxea fascicularis*. *Mar Biol* 142: 419–426.
40. McCulloch MT, Falter J, Trotter J, Montagna P (2012) Coral resilience to ocean acidification and global warming through pH up-regulation. *Nat Clim Change* 2: 623–627.
41. Chan NCS, Connolly SR (2013) Sensitivity of coral calcification to ocean acidification: a meta-analysis. *Global Change Biol* 19: 282–290.
42. McCulloch MT, Trotter J, Montagna P, Falter J, Dunbar R, et al. (2012) Resilience of cold-water scleractinian corals to ocean acidification: Boron isotopic systematics of pH and saturation state up-regulation. *Geochim Cosmochim Acta* 87: 21–34.
43. Dissard D, Douville E, Reynaud S, Juillet-Leclerc A, Montagna P, et al. (2012) Light and temperature effect on $\delta^{11}\text{B}$ and B/Ca ratios of the zooxanthellate coral *Acropora* sp.: results from culturing experiments. *Biogeosciences* 9: 4589–4605.
44. Reynaud S, Hemming NG, Juillet-Leclerc A, Gattuso J-P (2004) Effect of $p\text{CO}_2$ and temperature on the boron isotopic composition of the zooxanthellate coral *Acropora* sp. *Coral Reefs* 23: 539–546.
45. Hönisch B, Hemming NG, Grottoli AG, Amat A, Hanson GN, et al. (2004) Assessing scleractinian corals as recorders for paleo-pH: Empirical calibration and vital effects. *Geochim Cosmochim Acta* 68: 3675–3685.
46. Douville E, Paterne M, Cabioch G, Louvat P, Gaillardet J, et al. (2010) Abrupt sea surface pH change at the end of the Younger Dryas in the central sub-equatorial Pacific inferred from boron isotope abundance in corals (*Porites*). *Biogeosciences* 7: 2445–2459.
47. Wei G, McCulloch MT, Mortimer G, Deng W, Xie L (2009) Evidence for ocean acidification in the Great Barrier Reef of Australia. *Geochim Cosmochim Acta* 73: 2332–2346.
48. Pelejero C, Calvo E, McCulloch MT, Marshall JF, Gagan MK, et al. (2005) Preindustrial to modern interdecadal variability in coral reef pH. *Science* 309: 2204–2207.
49. Hemming NG, Hanson GN (1992) Boron isotopic composition and concentration in modern marine carbonates. *Geochim Cosmochim Acta* 56: 537–543.
50. Klockho K, Cody GD, Tossell JA, Dera P, Kaufman AJ (2009) Re-evaluating boron speciation in biogenic calcite and aragonite using ^{11}B MAS NMR. *Geochim Cosmochim Acta* 73: 1890–1900.
51. Rollion-Bard C, Blamart D, Trebosc J, Tricot G, Mussi A, et al. (2011) Boron isotopes as pH proxy: A new look at boron speciation in deep-sea corals using ^{11}B MAS NMR and EELS. *Geochim Cosmochim Acta* 75: 1003–1012.
52. Rodrigues LJ, Grottoli AG (2006) Calcification rate and the stable carbon, oxygen, and nitrogen isotopes in the skeleton, host tissue, and zooxanthellae of bleached and recovering Hawaiian corals. *Geochim Cosmochim Acta* 70: 2781–2789.
53. Marshall JF, McCulloch MT (2002) An assessment of the Sr/Ca ratio in shallow water hermatypic corals as a proxy for sea surface temperature. *Geochim Cosmochim Acta* 66: 3263–3280.
54. Leder JJ, Szamant AM, Swart P (1991) The effect of prolonged "bleaching" on skeletal banding and stable isotopic composition in *Montastraea annularis*. *Coral Reefs* 10: 19–27.
55. Hartmann AC, Carilli JE, Norris RD, Charles CD, Deheyn DD (2010) Stable isotopic records of bleaching and endolithic algae blooms in the skeleton of the boulder forming coral *Montastraea faveolata*. *Coral Reefs* 29: 1079–1089.
56. Schoepf V (2013) Physiology and Biogeochemistry of Corals Subjected to Repeat Bleaching and Combined Ocean Acidification and Warming. The Ohio State University. pp. 295.
57. Levas SJ (2012) Biogeochemistry and Physiology of Bleached and Recovering Hawaiian and Caribbean Corals. The Ohio State University. pp. 238.
58. McGinley M (2012) The impact of environmental stress on *Symbiodinium* spp.: A molecular and community-scale investigation. University of Delaware. pp. 173.
59. Aschaffenburg M (2012) The physiological response of *Symbiodinium* spp. to thermal and light stress: A comparison of different phylotypes and implications for coral reef bleaching. University of Delaware. pp. 155.
60. Budd AF, Fukami H, Smith ND, Knowlton N (2012) Taxonomic classification of the reef coral family Mussidae (Cnidaria: Anthozoa: Scleractinia). *Zool J Linn Soc* 166: 465–529.
61. Jeffrey SW, Humphrey GF (1975) New spectrophotometric equations for determining chlorophylls *a*, *b*, *c1*, and *c2* in higher plants, algae, and natural phytoplankton. *Biochem Physiol Pflanzen* 167: 191–194.
62. Marsh JA (1970) Primary productivity of reef-building calcareous red algae. *Ecology* 51: 255–263.
63. Jokiel PL, Maragos JE, Franzisket L (1978) Coral growth: buoyant weight technique. In: D. R. Stoddart and R. E. Johannes, editors. *Coral Reefs: Research Methods*. Paris: UNESCO. pp. 529–541.
64. Grottoli AG, Rodrigues LJ, Juarez C (2004) Lipids and stable carbon isotopes in two species of Hawaiian corals, *Porites compressa* and *Montipora verrucosa*, following a bleaching event. *Mar Biol* 145: 621–631.
65. Holcomb M, Rankenburg K, McCulloch M (2014) High-precision MC-ICP-MS measurements of $\delta^{11}\text{B}$: Matrix effects in direct injection and spray chamber sample introduction systems. In: K. Grice, editor. *Principles and Practice of Analytical Techniques in Geosciences*. The Royal Society of Chemistry, pp. 254–270. In press.
66. Guerrot C, Millot R, Robert M, Négrel P (2011) Accurate and high-precision determination of boron isotopic ratios at low concentration by MC-ICP-MS (Neptune). *Geostandard Newslett* 35: 275–284.
67. Grottoli AG, Rodrigues LJ, Matthews KA, Palardy JE, Gibb OT (2005) Pre-treatment effects on coral skeletal $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$. *Chem Geol* 221: 225–242.
68. Winer BJ (1971) *Statistical Principles in Experimental Design*. New York: McGraw-Hill.
69. Quinn GP, Keough MJ (2002) *Experimental Design and Data Analysis for Biologists*. New York: Cambridge University Press.
70. Moran MD (2003) Arguments for rejecting the sequential Bonferroni in ecological studies. *Oikos* 100: 403–405.
71. D'Olivo JP, McCulloch MT, Judd K (2013) Long-term records of coral calcification across the central Great Barrier Reef: assessing the impacts of river runoff and climate change. *Coral Reefs* 32: 999–1012.
72. Felis T, Patzelt J, Loya Y (2003) Mean oxygen-isotope signatures in *Porites* spp. corals: inter-colony variability and correction for extension-rate effects. *Coral Reefs* 22: 328–336.
73. Land LS, Lang JC, Barnes DJ (1975) Extension rate: A primary control on the isotopic composition of West Indian (Jamaican) scleractinian reef coral skeletons. *Mar Biol* 33: 221–233.
74. Weber JN, Woodhead PMJ (1972) Temperature dependence of oxygen-18 concentration in reef coral carbonates. *J Geophys Res* 77: 463–473.
75. Swart PK (1983) Carbon and oxygen isotope fractionation in scleractinian corals: A review. *Earth Sci Rev* 19: 51–80.
76. McConnaughey T (1989) ^{13}C and ^{18}O isotopic disequilibrium in biological carbonates: I. Patterns. *Geochim Cosmochim Acta* 53: 151–162.
77. McConnaughey TA, Burdett J, Whelan JF, Paull CK (1997) Carbon isotopes in biological carbonates: Respiration and photosynthesis. *Geochim Cosmochim Acta* 61: 611–622.
78. Grottoli AG (2002) Effect of light and brine shrimp levels on skeletal $\delta^{13}\text{C}$ values in the Hawaiian coral *Porites compressa*: A tank experiment. *Geochim Cosmochim Acta* 66: 1955–1967.
79. Grottoli AG, Wellington GM (1999) Effect of light and zooplankton on skeletal $\delta^{13}\text{C}$ values in the Eastern Pacific corals *Pavona clavus* and *Pavona gigantea*. *Coral Reefs* 18: 29–41.
80. Suzuki A, Gagan MK, Fabricius K, Isdale PJ, Yukino I, et al. (2003) Skeletal isotope microprofiles of growth perturbations in *Porites* corals during the 1997–1998 mass bleaching event. *Coral Reefs* 22: 357–369.
81. Heikoop JM, Dunn JJ, Risk MJ, Schwarcz HP, McConnaughey TA, et al. (2000) Separation of kinetic and metabolic isotope effects in carbon-13 records preserved in reef coral skeletons. *Geochim Cosmochim Acta* 64: 975–987.
82. Schoepf V, Levas SJ, Rodrigues LJ, McBride MO, Aschaffenburg M, et al. (in press) Kinetic and metabolic isotope effects in coral skeletal carbon isotopes: A re-evaluation using experimental coral bleaching as a case study. *Geochim Cosmochim Acta*.
83. Gattuso J-P, Allemand D, Frankignoulle M (1999) Photosynthesis and calcification at cellular, organismal, and community levels in coral reefs: a review on interactions and control by carbonate chemistry. *Am Zool* 39: 160–183.
84. Moya A, Tambutte S, Tambutte E, Zoccola D, Caminiti N, et al. (2006) Study of calcification during a daily cycle of the coral *Stylophora pistillata*: implications for light-enhanced calcification. *J Exp Biol* 17: 3413–3419.
85. Goreau TF, Goreau NI (1959) The physiology of skeleton formation in corals. II. Calcium deposition by hermatypic corals under different conditions. *Biol Bull* 117: 239–250.
86. Allemand D, Tambutte E, Zoccola D, Tambutte S (2011) Coral calcification, cells to reefs. In: Z. Dubinsky and N. Stambler, editors. *Coral Reefs: An Ecosystem in Transition*. Springer. pp.119–150.
87. Tambutte S, Holcomb M, Ferrier-Pages C, Reynaud S, Tambutte E, et al. (2011) Coral biomineralization: from the gene to the environment. *J Exp Mar Biol Ecol*. doi:10.1016/j.jembe.2011.1007.1026.
88. Houlbreque F, Ferrier-Pages C (2009) Heterotrophy in tropical scleractinian corals. *Biol Rev* 84: 1–17.
89. Anthony KRN, Hoogenboom MO, Maynard JF, Grottoli AG, Middlebrook R (2009) Energetics approach to predicting mortality risk from environmental stress: a case study of coral bleaching. *Funct Ecol* 23: 539–550.
90. Connolly SR, Lopez-Yglesias MA, Anthony KRN (2012) Food availability promotes rapid recovery from thermal stress in a scleractinian coral. *Coral Reefs* 31: 951–960.
91. Porter JW, Fitt WK, Spero HJ, Rogers AD, White MW (1989) Bleaching in reef corals: physiological and stable isotopic responses. *Proc Natl Acad Sci USA* 86: 9342–9346.
92. Holcomb M, Tambutte E, Allemand D, Tambutte S (2014) Light enhanced calcification in *Stylophora pistillata*: effects of glucose, glycerol and oxygen. *PeerJ* 2: e375.

93. Colombo-Pallotta MF, Rodriguez-Roman A, Iglesias-Prieto R (2010) Calcification in bleached and unbleached *Montastraea faveolata*: evaluating the role of oxygen and glycerol. *Coral Reefs*: doi:10.1007/s00338-00010-00638-x.
94. Cantin NE, Lough JM (2014) Surviving coral bleaching events: *Porites* growth anomalies on the Great Barrier Reef. *PLoS One* 9: e88720.
95. Carilli JE, Norris RD, Black BA, Walsh SM, McField M (2009) Local stressors reduce coral resilience to bleaching. *PLoS One* 4: e6324. doi: 6310.1371/journal.pone.0006324.
96. McCulloch MT, Gagan MK, Mortimer GE, Chivas AR, Isdale PJ (1994) A high-resolution Sr/Ca and $\delta^{18}\text{O}$ coral record from the Great Barrier Reef, Australia, and the 1982–1983 El Niño. *Geochim Cosmochim Acta* 58: 2747–2754.
97. McCulloch M, Mortimer G, Esat T, Xianhua L, Pillans B, et al. (1996) High resolution windows into early Holocene climate: Sr/Ca coral records from the Huon Peninsula. *Earth Planet Sci Lett* 138: 169–178.
98. Fallon S, McCulloch M, Alibert C (2003) Examining water temperature proxies in *Porites* corals from the Great Barrier Reef: a cross-shelf comparison. *Coral Reefs* 22: 389–404.
99. Cohen AL, Owens KE, Layne GD, Shimizu N (2002) The effect of algal symbionts on the accuracy of Sr/Ca paleotemperatures from coral. *Science* 296: 331–333.
100. de Villiers S, Shen GT, Nelson BK (1994) The Sr/Ca-temperature relationship in coralline aragonite: Influence of variability in (Sr/Ca)/seawater and skeletal growth parameters. *Geochim Cosmochim Acta* 58: 197–208.
101. de Villiers S, Nelson BK, Chivas AR (1995) Biological controls on coral Sr/Ca and $\delta^{18}\text{O}$ reconstructions of sea surface temperatures. *Science* 269: 1247–1249.
102. Allison N, Finch AA (2004) High-resolution Sr/Ca records in modern *Porites lobata* corals: Effects of skeletal extension rate and architecture. *G-cubed* 5: Q05001. doi:05010.01029/02004GC000696.
103. Sun Y, Sun M, Lee T, Nie B (2005) Influence of seawater Sr content on coral Sr/Ca and Sr thermometry. *Coral Reefs* 24: 23–29.
104. Meibom A, Stage M, Wooden J, Constantz BR, Dunbar RB, et al. (2003) Monthly Strontium/Calcium oscillations in symbiotic coral aragonite: Biological effects limiting the precision of the paleotemperature proxy. *Geophys Res Lett* 30: 1418.
105. Cohen AL, Layne GD, Hart SR, Lobel PS (2001) Kinetic control of skeletal Sr/Ca in a symbiotic coral: Implications for the paleotemperature proxy. *Paleoceanography* 16: 20–26.
106. Alibert C, McCulloch MT (1997) Strontium/calcium ratios in modern *Porites* corals from the Great Barrier Reef as a proxy for sea surface temperature: Calibration of the thermometer and monitoring of ENSO. *Paleoceanography* 12: 345–363.
107. Beck JW, Edwards RL, Ito E, Taylor FW, Recy J, et al. (1992) Sea-surface temperature from coral skeletal strontium/calcium ratios. *Science* 257: 644–647.
108. Fallon SJ, McCulloch MT, van Woessik R, Sinclair DJ (1999) Corals at their latitudinal limits: laser ablation trace element systematics in *Porites* from Shirigai Bay, Japan. *Earth Planet Sci Lett* 172: 221–238.
109. Sinclair DJ, Kinsley LPJ, McCulloch MT (1998) High resolution analysis of trace elements in corals by laser ablation ICP-MS. *Geochim Cosmochim Acta* 62: 1889–1901.
110. Ferrier-Pages C, Boisson F, Allemand D, Tambutté E (2002) Kinetics of strontium uptake in the scleractinian coral *Stylophora pistillata*. *Mar Ecol Prog Ser* 245: 93–100.
111. Reynaud S, Ferrier-Pagès C, Meibom A, Mostefaoui S, Mortlock R, et al. (2007) Light and temperature effects on Sr/Ca and Mg/Ca ratios in the scleractinian coral *Acropora* sp. *Geochim Cosmochim Acta* 71: 354–362.
112. Mitsuguchi T, Matsumoto E, Uchida T (2003) Mg/Ca and Sr/Ca ratios of *Porites* coral skeleton: Evaluation of the effect of skeletal growth rate. *Coral Reefs* 22: 381–388.
113. Hayashi E, Suzuki A, Nakamura T, Iwase A, Ishimura T, et al. (2013) Growth-rate influences on coral climate proxies tested by a multiple colony culture experiment. *Earth Planet Sci Lett* 362: 198–206.
114. Mitsuguchi T, Matsumoto E, Abe O, Uchida T, Isdale PJ (1996) Mg/Ca thermometry in coral skeletons. *Science* 274: 961–963.
115. Shen GT, Dunbar RB (1995) Environmental controls on uranium in reef corals. *Geochim Cosmochim Acta* 59: 2009–2024.
116. Rong Min G, Lawrence Edwards R, Taylor FW, Recy J, Gallup CD, et al. (1995) Annual cycles of U/Ca in coral skeletons and U/Ca thermometry. *Geochim Cosmochim Acta* 59: 2025–2042.
117. Lea DW, Shen GT, Boyle EA (1989) Coralline barium records temporal variability in equatorial Pacific upwelling. *Nature* 340: 373–376.
118. Tudhope AW, Lea DW, Shimmield GB, Chilcott CP, Head S (1996) Monsoon climate and Arabian Sea coastal upwelling recorded in massive corals from Southern Oman. *Paleaia* 11: 347–361.
119. McCulloch M, Fallon S, Wyndham T, Hendy E, Lough J, et al. (2003) Coral record of increased sediment flux to the inner Great Barrier Reef since European settlement. *Nature* 421: 727–730.
120. Alibert C, Kinsley L, Fallon SJ, McCulloch MT, Berkemans R, et al. (2003) Source of trace element variability in Great Barrier Reef corals affected by the Burdekin flood plumes. *Geochim Cosmochim Acta* 67: 231–246.
121. Allison N, Finch AA (2007) High temporal resolution Mg/Ca and Ba/Ca records in modern *Porites lobata* corals. *G-cubed* 8: Q05001. doi:05010.01029/02006gc001477.
122. Horta-Puga G, Carriquiry JD (2012) Coral Ba/Ca molar ratios as a proxy of precipitation in the northern Yucatan Peninsula, Mexico. *Appl Geochem* 27: 1579–1586.
123. Pretet C, Reynaud S, Ferrier-Pagès C, Gattuso J-P, Kamber B, et al. (2014) Effect of salinity on the skeletal chemistry of cultured scleractinian zooxanthellate corals: Cd/Ca ratio as a potential proxy for salinity reconstruction. *Coral Reefs* 33: 169–180.
124. Hart SR, Cohen AL (1996) An ion probe study of annual cycles of Sr/Ca and other trace elements in corals. *Geochim Cosmochim Acta* 60: 3075–3084.
125. Sinclair DJ (2005) Non-river flood barium signals in the skeletons of corals from coastal Queensland, Australia. *Earth Planet Sci Lett* 237: 354–369.
126. Chornesky EA, Peters EC (1987) Sexual reproduction and colony growth in the scleractinian coral *Porites astreoides*. *Biol Bull* 172: 161–177.
127. Chen T, Yu K, Li S, Chen T, Shi Q (2011) Anomalous Ba/Ca signals associated with low temperature stresses in *Porites* corals from Daya Bay, northern South China Sea. *Journal of Environmental Sciences* 23: 1452–1459.