

RESEARCH ARTICLE

HYBRID GENOMICS

Natural hybridization reveals incompatible alleles that cause melanoma in swordtail fish

Daniel L. Powell^{1,2,3*}, Mateo García-Olázabal^{2,3}, Mackenzie Keegan⁴, Patrick Reilly⁵, Kang Du⁶, Alejandra P. Díaz-Loyo⁷, Shreya Banerjee¹, Danielle Blakkan¹, David Reich^{8,9}, Peter Andolfatto¹⁰, Gil G. Rosenthal^{2,3}, Manfred Schartl^{2,3,6,11,12}, Molly Schumer^{1,2*}

The establishment of reproductive barriers between populations can fuel the evolution of new species. A genetic framework for this process posits that “incompatible” interactions between genes can evolve that result in reduced survival or reproduction in hybrids. However, progress has been slow in identifying individual genes that underlie hybrid incompatibilities. We used a combination of approaches to map the genes that drive the development of an incompatibility that causes melanoma in swordtail fish hybrids. One of the genes involved in this incompatibility also causes melanoma in hybrids between distantly related species. Moreover, this melanoma reduces survival in the wild, likely because of progressive degradation of the fin. This work identifies genes underlying a vertebrate hybrid incompatibility and provides a glimpse into the action of these genes in natural hybrid populations.

The emergence of reproductive barriers between populations is the first step in the process of speciation and drives Earth’s biological diversity, yet surprisingly little is known about how it occurs at the genetic level. The Dobzhansky-Muller model of hybrid incompatibility (1–3) posits that new mutations arising in diverging species can interact negatively in hybrids, generating lower hybrid viability or causing hybrid sterility. Although empirical work provides support for the general predictions of this model (4), progress in this area has been limited by a lack of knowledge about which genes interact to generate hybrid incompatibilities. Despite the effort devoted to this problem, only a dozen incompatible interactions have been mapped to the single-gene level [reviewed in (5)]. With so few known cases, it has been difficult to evaluate whether common genetic and evolutionary mechanisms underlie the emergence of incompatibilities (6–8).

With an increasing appreciation that hybridization is common across the tree of life (9–13), there has been renewed interest in identifying hybrid incompatibilities and understanding how these genes act as barriers in nature. Of hybrid incompatibilities that have been mapped to the single-gene level, most have been identified with crosses between model species that no longer naturally hybridize (4, 5). As a result, it is unclear whether these mapped incompatibilities were important in the initial divergence between species or arose after these lineages had stopped exchanging genes.

One such example is the melanoma receptor tyrosine-protein kinase (*xmrk*) gene in swordtail fish (genus *Xiphophorus*). *xmrk* is one of two identified genes in vertebrates that drive hybrid incompatibility (the other being the regulator of mammalian recombination hotspots, *prdm9*) (14) and was one of the earliest described hybrid incompatibilities (15). In crosses between *Xiphophorus maculatus* and *Xiphophorus hellerii*, a malignant melanoma develops in a subset of F₂ hybrids, emanating from natural pigmentation spots on the body and fins. This hybrid incompatibility is the result of an interaction between the *xmrk* gene derived from *X. maculatus* and an unknown locus derived from *X. hellerii* (16).

Despite work that demonstrated the role of *xmrk* in the development of hybrid melanomas, its importance as a barrier between species has been debated (17). This is because *X. maculatus* and *X. hellerii* diverged ~3 million years ago and do not naturally hybridize (18). Moreover, because melanoma in *X. maculatus* x *X. hellerii* laboratory hybrids develops later in life, it is unclear whether it affects survival and reproduction (17).

Melanoma occurs in hybrids between recently diverged swordtail species

We identified a phenotypically similar melanoma in natural hybrids formed between the swordtail fish species *Xiphophorus birchmanni* and *Xiphophorus malinche*. *X. birchmanni* and *X. malinche* are closely related and hybridize in the wild (~0.5% differences per base pair and ~250,000 generations diverged) (19). Although a subset of hybrids are viable and fertile, there is evidence of selection against hybrid incompatibilities (20–22). In some populations, hybrids develop melanoma early in life (13 ± 4% of males develop melanoma before sexual maturity) (fig. S1).

Melanoma in *X. birchmanni* x *X. malinche* hybrids develops from a phenotype derived from *X. birchmanni* called the “spotted caudal,” which is a dark blotch on the caudal fin generated by clusters of macromelanocyte cells (Fig. 1A and fig. S2) (23). The spotted caudal trait occurs at intermediate frequencies in *X. birchmanni* but is absent from *X. malinche* populations (Fig. 1B). Some hybrid populations have a high frequency of the trait and exhibit phenotypes that extend beyond the range of those observed in *X. birchmanni* (Fig. 1, B and C, and figs. S3 and S4), including invasion of macromelanocyte cells into the body, where they are normally absent. Tracking of hybrids in the laboratory documents the progression of the trait from a phenotype typical of the *X. birchmanni* spot to the expanded trait found in some hybrids (Fig. 1D and fig. S1) (24). Histological sections from hybrid individuals revealed penetration of melanocytes into the musculature and invasion of surrounding tissues, which is indicative of a malignant melanoma (Fig. 1E and fig. S5).

We performed mRNA-sequencing of hybrid individuals that varied in the degree of expansion of their spot (figs. S2 and S6). Functional enrichment analysis indicated changes in the regulation of a number of melanoma-associated gene categories, such as pigment cell differentiation and regulation of cytoskeletal organization, including several implicated in melanoma in other fish species (Fig. 1F and table S1) (24, 25).

Melanoma is extremely rare in nonhybrid individuals (Fig. 1C) (6, 26), and we have not identified a single wild-caught *X. birchmanni* male with melanoma (1296 males collected from 2017 to 2019, 0 with melanoma). Laboratory-reared individuals indicate that environmental levels of ultraviolet irradiance or other natural carcinogens do not underlie differences in the frequency of melanoma between hybrid and parental populations (24). The presence of melanoma in hybrids, but not the parental species, suggests that this melanoma is a hybrid incompatibility generated by interactions between alleles in the *X. birchmanni* and *X. malinche* genomes.

¹Department of Biology, Stanford University and Howard Hughes Medical Institute, Stanford, CA, USA. ²Centro de Investigaciones Científicas de las Huastecas “Aguazarca”, A.C., Calnali, Hidalgo, Mexico. ³Department of Biology, Texas A&M University, College Station, TX, USA. ⁴Department of Biology, Northeastern University, Boston, MA, USA. ⁵Lewis Sigler Institute for Integrative Genomics, Princeton University, Princeton, NJ, USA. ⁶Developmental Biochemistry, Biocenter, University of Würzburg, Würzburg, Bavaria, Germany. ⁷Laboratorio de Ecología de la Conducta, Instituto de Fisiología, Benemérita Universidad Autónoma de Puebla, Puebla, Mexico. ⁸Department of Genetics, Harvard Medical School, Howard Hughes Medical Institute, and the Broad Institute of Harvard and MIT, Cambridge, MA, USA. ⁹Department of Human Evolutionary Biology, Harvard University, Cambridge, MA 02138, USA. ¹⁰Department of Biological Sciences, Columbia University, New York, NY, USA. ¹¹Hagler Institute for Advanced Study, Texas A&M University, College Station, TX, USA. ¹²*Xiphophorus* Genetic Stock Center, Texas State University San Marcos, San Marcos, TX, USA. *Corresponding author. Email: dpowell8@stanford.edu (D.L.P.); schumer@stanford.edu (M.Schu.)

Loci associated with the spotting phenotype in *X. birchmanni*

We mapped the genetic basis of the spotted caudal in *X. birchmanni* and the genetic ba-

sis of melanoma in interspecific hybrids. We generated de novo assemblies for both species using a 10X-based linked read approach followed by assembly into chromosomes with Hi-C

data and annotated the resulting assemblies with RNA-sequencing (RNA-seq) data (24).

We collected low-coverage whole-genome sequence data for 392 adult male *X. birchmanni*

Fig. 1. Hybridization generates a high incidence of melanoma. (A) Naturally hybridizing species *X. malinche* (top) and *X. birchmanni* (middle) differ in morphological traits, including the presence of a melanin pigment spot that is polymorphic in *X. birchmanni*. In hybrids, this spotting phenotype can transform into a melanoma (bottom). (B) Whereas *X. birchmanni* populations segregate for the presence of this spot, the trait is absent in *X. malinche* populations; hybrid populations have high frequencies of this trait. (C) The trait is at higher frequencies in hybrid populations and covers more of the body. Shown here is invasion area, or the melanized body surface area outside of the caudal fin (normalized for body size). Hybrid phenotypes are shown from three populations on the Río Calnali (fig. S3): AGCZ, Aguazarca; CALL, Calnali low; CHAF, Chahuaco falls. (D) Spots expand more over a 6-month period in hybrids than in *X. birchmanni* individuals. (E) A cross section of the caudal peduncle from a Chahuaco falls hybrid (10 \times magnification). Melanoma cells invading the body and muscle bundles are visually evident (indicated with blue stars). (F) Gene ontology categories enriched in melanoma tissue compared with normal caudal tissue (24, 45). The size of the dots reflects the number of genes identified, and the color corresponds to the *P* value. Categories with undefined odds ratios (not plotted) are listed in table S1. In (B) and (D), the plot shows the mean, and whiskers indicate two standard errors of the mean. Individual points show the raw data.

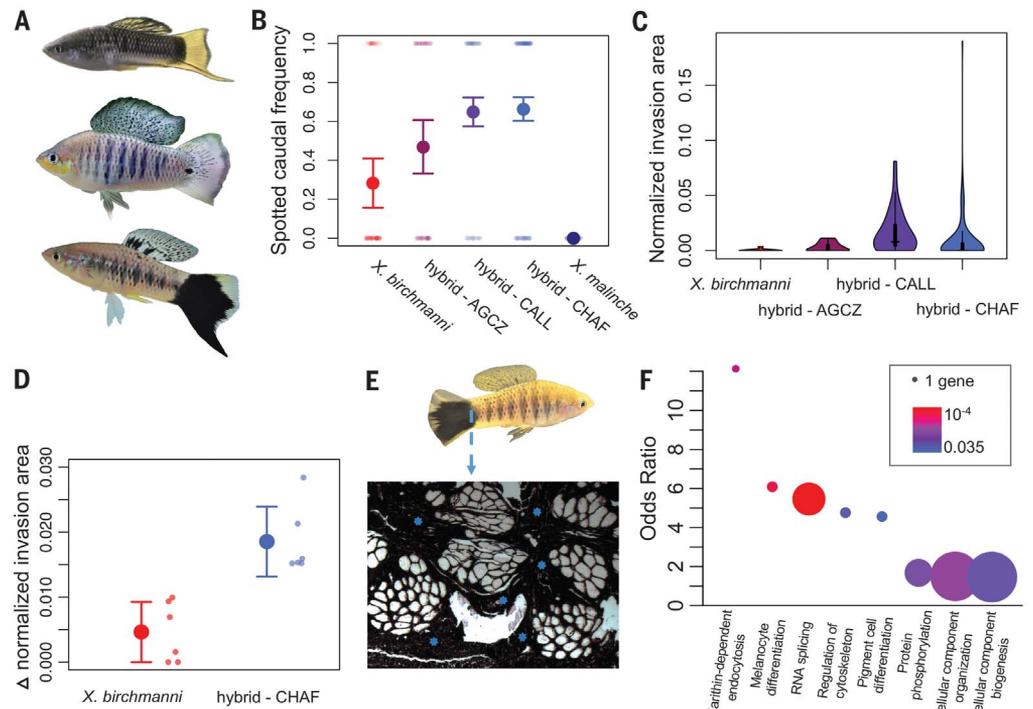
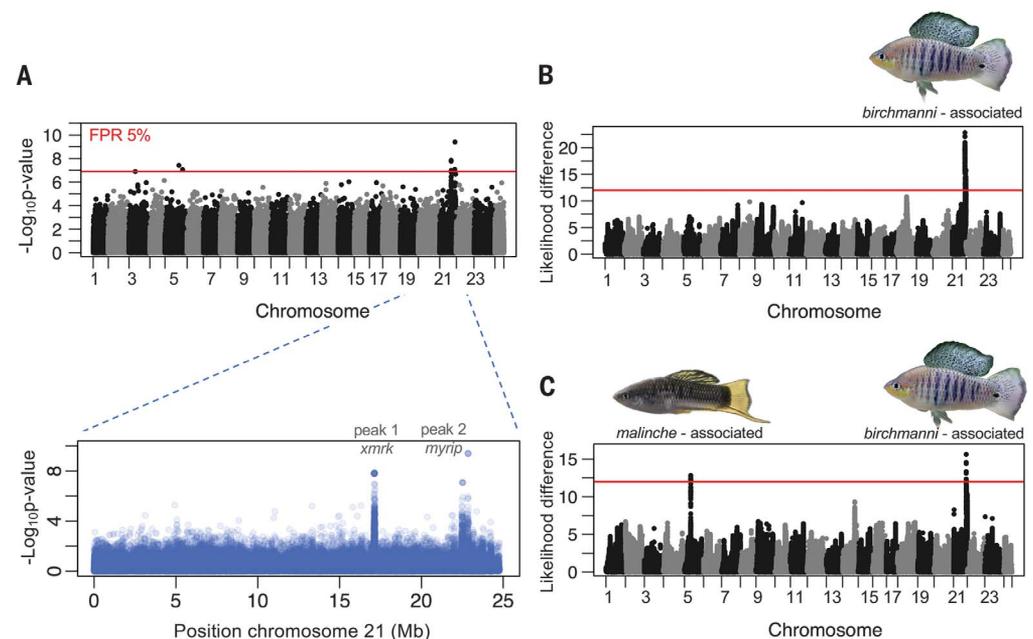


Fig. 2. Combined genome-wide association and admixture mapping approaches identify the genetic basis of the melanoma hybrid incompatibility. (A) Results of genome-wide association scan for allele frequency differences between spotted cases and unspotted controls. (Top) Results can be seen for all chromosomes, and the red line indicates the genome-wide significance threshold, determined by permutation (24). (Bottom) Results from chromosome 21, where two distinct regions are strongly associated with spotting. (B) Admixture mapping in hybrids identifies associations between *X. birchmanni* ancestry on chromosome 21 and spot presence. Plotted here are log likelihood differences between models with and without ancestry at the focal site included as a covariate. The red line indicates the genome-wide significance threshold, determined by permutation (24). (C) When we treated melanocyte invasion as the focal trait and mapped associations with ancestry, we again identified associations with *X. birchmanni* ancestry on chromosome 21 but also identified a second region on chromosome 5 associated with *X. malinche* ancestry.



individuals from a single collection site and performed a genome-wide association study (GWAS), scanning for allele frequency differences between spotted cases ($n = 159$) and unspotted controls ($n = 233$), evaluating the impact of population structure and low-coverage data (24). We identified a strong association between the spotting pattern and allele frequency differences on chromosome 21 at an estimated false positive rate of 5% (by permutation) (Fig. 2A and figs. S7 and S8) (24). Two distinct signals are evident on this chromosome, ~5 Mb apart (Fig. 2A) (24). The first peak is centered on the *xmrk* gene (fig. S9) (27), which arose through duplication of a gene homologous to the mammalian epidermal growth factor receptor ~3 million years ago (11, 28, 29). *xmrk* controls pigmentation patterns and drives hybrid melanomas in rela-

tives of *X. birchmanni* and *X. malinche* (16). The signal at the second peak on chromosome 21 contains a single gene, the melanosome transporter gene myosin VIIA and Rab interacting protein (*myrip*) (Fig. 2A) (24, 30).

Genetic architecture of the melanoma incompatibility

For the melanoma phenotype, we used an admixture mapping approach, focusing our efforts on a hybrid population with high incidence of melanoma (19 ± 3% of adult males, the Chahuaco falls population). To infer local ancestry, we generated ~1X low-coverage whole-genome sequence data for 209 adult males from this population and applied a hidden Markov model to 680,291 ancestry informative sites genome-wide (20, 22, 31) [approximately one ancestry informative site per kilobase, (24)]. Simulations and

analyses of laboratory-generated crosses indicate that we should have high accuracy in local ancestry inference (figs. S10 to S13) (24).

Using these data, we performed admixture mapping for spot presence and melanocyte invasion of the body (59% of individuals had spots, and 19% of individuals had melanoma). Admixture mapping for the presence of the spotted caudal revealed one strongly associated region on chromosome 21 (log likelihood difference of linear models, 23) (24) where spotting correlated with *X. birchmanni* ancestry (Fig. 2B). Because of the lower resolution of admixture, mapping this peak is broad, but the signal occurs in the same regions identified by our GWAS scan (24), confirming that the genetic basis of the spot is the same in *X. birchmanni* and hybrids. Simulations accounting for effect size inflations owing to the

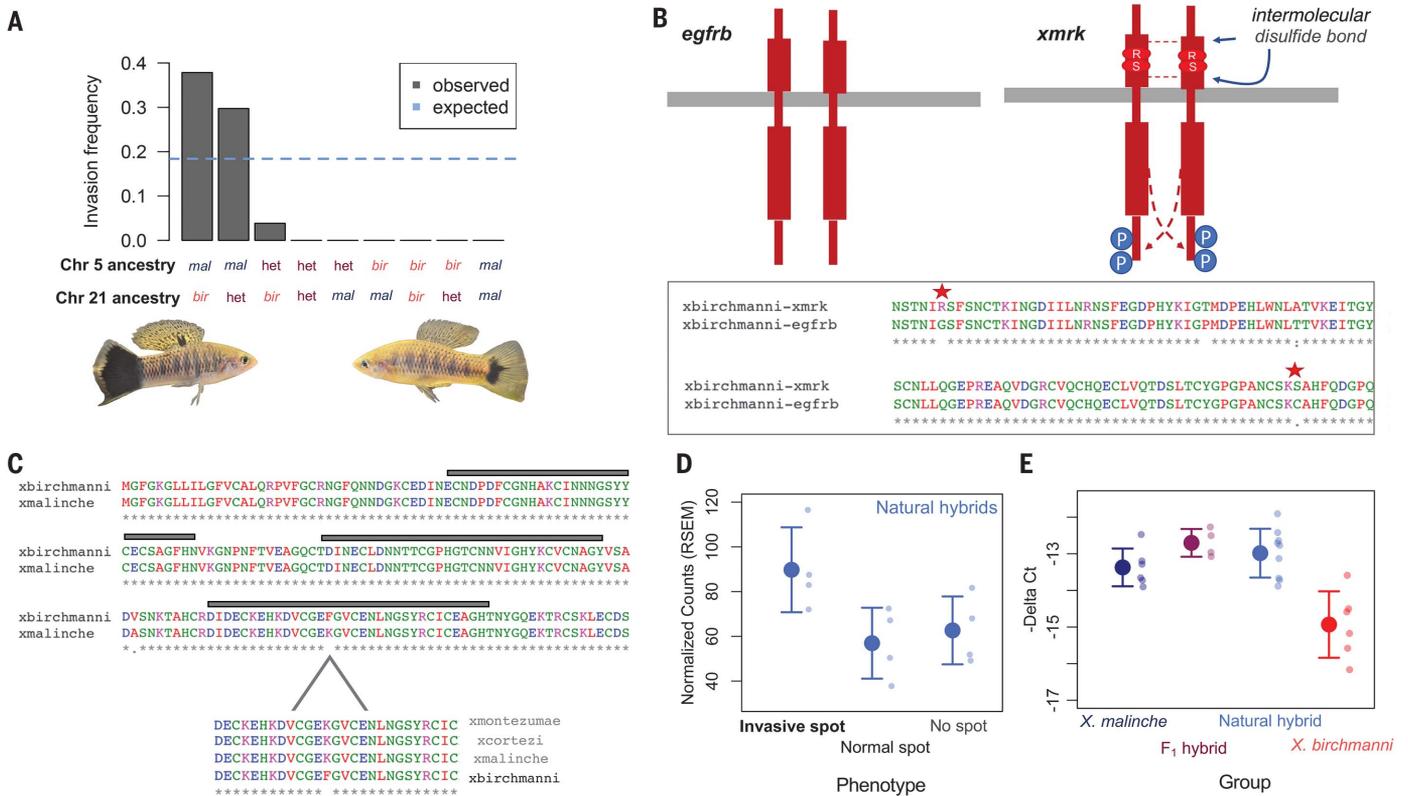


Fig. 3. Interactions between chromosomes 5 and 21 are associated with melanoma in hybrids. (A) Proportion of individuals with melanoma as a function of ancestry at the associated regions on chromosome 5 and chromosome 21. The blue dashed line indicates the expected proportion of cases if melanoma risk were equally distributed among individuals with at least one *birchmanni* allele at chromosome 21. We only had one observation for the *bir-bir* and *bir-het* genotypes. (B) The *xmrk* sequence in *X. birchmanni* harbors two mutations (G364R and C582S) that transform *xmrk* to a constitutively active state (33, 46). The schematic compares the ancestral form of the protein (*egfrb*) to the predicted structure of *xmrk* in *X. birchmanni*. Proteins are shown in red, and the cell membrane is shown in gray. In *xmrk*, residues R364 and S582 promote intramolecular disulfide bonds that cause protein dimerization and phosphorylation (blue circles) (33, 46). (Single-letter abbreviations for the amino acid residues are as follows: C, Cys; G, Gly; R, Arg; and S, Ser. In *xmrk*, amino acids were substituted at certain locations;

for example, G364R indicates that glycine at position 364 was replaced by arginine.) (Inset) A partial clustal alignment of *X. birchmanni* *egfrb* and *xmrk* with these substitutions highlighted. Colors indicate properties of the amino acid, and asterisks indicate locations where the amino acid sequences are identical. (C) Clustal alignment showing the N terminus of *cd97* in *X. birchmanni* and *X. malinche*. We observed a substitution in a conserved epidermal growth factor-binding domain (gray rectangles). (Inset) The substitution found in *X. birchmanni* is not present in closely related species. (D) Expression of *cd97* based on RNA-seq data in melanoma, spotted, and unspotted tissue from Chahuaco falls hybrids (four biological replicates per group). (E) Real-time quantitative PCR of *cd97* from caudal fin tissue from *X. malinche*, *X. birchmanni*, and natural and F₁ hybrids (four to nine biological replicates per group). In (D) and (E), large solid dots indicate the mean, and whiskers indicate two standard errors of the mean. Individual points show the raw data.

“winner’s curse” (32) suggest that *X. birchmanni* ancestry in this region explains ~75% of the variation in spot presence or absence (24).

Admixture mapping for melanoma identified an additional significant region on chromosome 5. In this case, melanocyte invasion was associated with *X. malinche* ancestry (Fig. 2C). A contingency test indicated a non-random association between *X. birchmanni* ancestry at the chromosome 21 peak and *X. malinche* ancestry at the chromosome 5 peak, with the prevalence of melanoma (Fisher’s exact test, $P = 0.0005$) (Fig. 3A). Individuals heterozygous for *X. malinche* ancestry at this region on chromosome 5 appear to have a lower risk of melanoma (Fig. 3A). Moreover, regardless of melanoma phenotype, spotted individuals that were heterozygous at the chromosome 5 peak had smaller spots than individuals that were homozygous (Student’s *t* test on log-normalized area $P = 0.007$) (fig. S14).

Linking molecular changes to hybrid melanoma

Our GWAS identified associations between the spotted caudal and both *xmrk* and *myrip* (Fig. 2A), making it initially unclear whether either or both of these genes interacts with *malinche* ancestry on chromosome 5 to produce melanoma. Although both are associated with the spotting pattern that precedes melanoma, *myrip* is not expressed in an RNA-seq dataset of adult caudal tissue, nor is it expressed in the melanoma itself (fig. S15). By contrast, *xmrk* is expressed in caudal tissue and has higher expression in spotted than unspotted tissues (fig.

S15). In addition, functional studies have linked *xmrk* to the development of melanoma. We identified two amino acid substitutions in *X. birchmanni* that fall within the extracellular domain of *xmrk* known to drive the oncogenic properties of *xmrk* in vitro (Fig. 3B) (33), and transgenic studies have demonstrated that overexpression of *xmrk* causes the formation of tumors (25, 34, 35). Although *myrip* may not be directly involved in the development of melanoma, past work in other swordtail species has suggested the presence of “patterning” loci linked to *xmrk* [reviewed in (23)]. Given *myrip*’s role in melanosome transport, we speculate that it could play a role in pigmentation patterning, which occurs in the first several weeks of life.

The region on chromosome 5 associated with *X. malinche* ancestry and melanoma contains only two genes, a gene called *cd97* and a fatty acid transporter gene (Figs. 2C and 3C). Although this is unusually high resolution given our admixture-mapping approach, subsampling the data indicated that this scale of resolution is likely the result of high recombination rates in this region (24). We therefore sought to better characterize the two genes in this region.

The ortholog of *cd97* in mammals plays a role in epithelial metastasis and is associated with tumor invasiveness in cancers (36–38). Accordingly, we found that *cd97* is up-regulated in RNA-seq data from melanotic tissue in hybrids, whereas the fatty acid transporter gene is not (Fig. 3D); nor is this pattern of up-

regulation observed in any other gene within 100 kb of this region (24). In addition, of five amino acid changes between *X. birchmanni* and *X. malinche* in *cd97*, one occurs in a conserved epidermal growth factor-like calcium-binding domain (Fig. 3C).

We further investigated differences in expression of *cd97* using a targeted quantitative polymerase chain reaction (qPCR) approach. We found that *cd97* was expressed at low levels in the caudal fin tissue of *X. birchmanni*, regardless of spotting phenotype, but at similarly high levels in *X. malinche* and in natural and artificial hybrids [analysis of variance (ANOVA) $P = 1^{-5}$, Tukey post hoc; all groups different from *X. birchmanni* at $P < 0.005$] (Fig. 3E). Higher expression of *cd97* in *X. malinche* and hybrids is not tissue-specific and surprisingly does not appear to be driven by cis-regulatory differences (fig. S16) (24). We do not know whether the link between *X. malinche* ancestry at *cd97* and melanoma is driven by coding or regulatory differences (24). However, in mammals, overexpression of *cd97* has been linked to tumor metastasis; a similar mechanism could be involved here, given that high expression of *cd97* coincides with invasion of other tissues with melanoma cells.

Independent evolution of a melanoma incompatibility

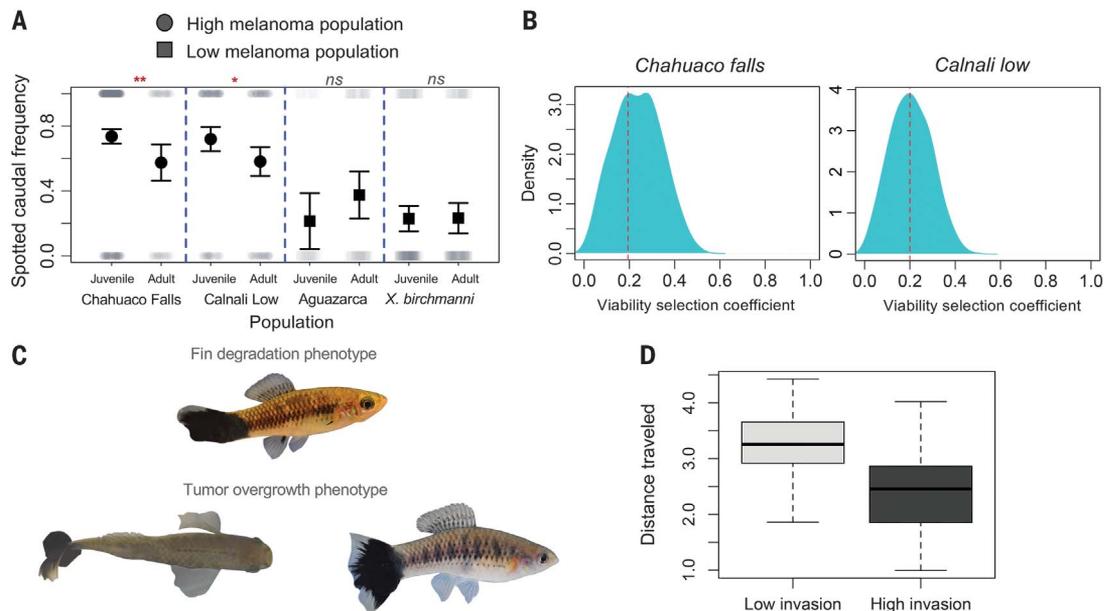
Although the role of *xmrk* in the *maculatus-hellerii* hybrid incompatibility has been known for 30 years (39), the identity of the interacting gene is not known. Laboratory crosses have

Fig. 4. Impact of the spotted caudal melanoma in natural hybrid populations.

(A) Frequency of spotting in juvenile and adult males across populations with high (circles, Calnali low and Chahuaco falls) or low (squares, Aguazarca and *X. birchmanni*) melanoma incidence. Asterisks indicate significant differences by age class ($*P < 0.05$, $**P < 0.01$; ns indicates

nonsignificant differences in a two-sample *z* test). Gray points indicate the raw data, black points indicate the mean, and error bars indicate one standard error of the mean. (B) Results of approximate Bayesian computation simulations indicate that the change in frequency of the spotting phenotype between juvenile and adult males is consistent with strong viability

selection (24). Shown here are posterior distributions of viability selection coefficients consistent with the observed frequency change data in (left) Chahuaco falls and (right) Calnali low. (C) Because of where the melanoma develops, it can cause (top) the degradation of a fin essential in swimming or (bottom) the growth of tumors on the fin (overhead and side view of the same individual). (D) Visualization of the difference in fast-start response between individuals with low and high melanoma invasion (upper and lower 25% quantiles shown here). This representation is for visualization only; the statistical analysis comes from a linear model.



narrowed to a ~5 Mb region on chromosome 5 but have not yet identified the underlying gene, although candidates have been proposed (16). We identified a distinct region on chromosome 5 (Fig. 2C), more than 7 Mb away from the region identified in the *hellerii-maculatus* cross. Alignments of chromosome 5 confirm that *cd97* is at the same location in all four species (fig. S17), and linkage disequilibrium between *cd97* and the region identified in *hellerii-maculatus* decays to background levels in hybrids (24). Using simulations, we ruled out a lack of power to detect an association between this region and melanoma, assuming a similar effect size to that seen in the *hellerii-maculatus* cross (fig. S18). These results indicate that the incompatibility has a partially distinct genetic basis in the two crosses generating hybrid melanoma. However, we may not have mapped all components of the melanoma incompatibility, particularly if other genes have subtle impacts on melanoma risk (24).

These mapping results are surprising because they suggest that a melanoma incompatibility involving *xmrk* emerged independently in two distinct lineages. Despite the evolutionary distance between these species (fig. S19), it is possible that the melanoma incompatibility arose through similar evolutionary paths in both cases. *X. hellerii* and its relatives lack *xmrk* (39), either because the lineage leading to this clade diverged before *xmrk* arose (fig. S19) or because of an ancient loss of *xmrk*. By contrast, many species in the lineage leading to *X. birchmanni* and *X. malinche* retained *xmrk* (fig. S19) (24), but we found that *xmrk* has been deleted in *X. malinche* since its divergence from *X. birchmanni* (fig. S20) (24). We speculate that the loss of *xmrk* in *X. malinche* could have changed the level of constraint on interacting genes in this lineage, and if so, similar evolutionary mechanisms could be at play in *X. hellerii*.

Selection on melanoma in natural populations

Although the melanoma that forms in *birchmanni* x *malinche* hybrids appears to be deleterious from its development early in life (fig. S1) and its malignancy (Fig. 1E and fig. S5), we wanted to evaluate its impact in natural hybrid populations. Over several years, we observed shifts in the frequency of the spotted caudal trait between juvenile and adult males (24). Specifically, in hybrid populations with high incidences of melanoma, juvenile males had a significantly higher frequency of the spotted caudal trait than that of adult males (two-sample *z* test, both $P < 0.02$) (Fig. 4A). By contrast, this pattern was not observed in the *X. birchmanni* parental population or in a hybrid population with a low incidence of melanoma (Fig. 4A). Phenotype tracking of laboratory-raised individuals shows that once

it appears, the spotted area always expands over time, indicating that we do not expect reversal of spotting due to some form of phenotypic plasticity (Fig. 1D) (24). We also did not find evidence for systematic shifts in ancestry genome-wide between the juvenile and adult male life stages that could explain this pattern (24). However, we do see a shift toward *X. birchmanni* ancestry at the melanoma risk locus (in the top 1% of changes genome-wide) (fig. S21) (24).

The observed shifts in spotting phenotype and ancestry at the melanoma risk locus, combined with an absence of substantial genome-wide shifts in ancestry, suggest that viability selection acts against spotted hybrids during maturation (24). Using an approximate Bayesian approach, we inferred that the strength of viability selection required to generate observed phenotypic shifts was extremely high and consistent across the two hybrid populations where melanoma is common (maximum a posteriori estimate of $s \sim 0.2$; 95% confidence intervals 0.05 to 0.44 and 0.04 to 0.38) (Fig. 4B).

Histology showed degradation of the muscle tissue that connects to the caudal fin in advanced melanomas (Figs. 1E and 4C, fig. S5, and movie S1). We thus measured its impact on swimming performance using two approaches. We did not find differences between phenotypes in ability to swim against a current (24). However, individuals with three-dimensional melanoma had slower escape responses when startled (linear model, $t = -2.6$, $p = 0.014$) (Fig. 4D) (24). This result is intriguing because fish with expanded spotting are likely more visible (movie S2), which could affect detection by avian and piscine predators.

Given the evidence for reduced survival of spotted individuals in populations with high rates of melanoma, it is surprising that this trait is still segregating in some hybrid populations (Fig. 4, A and B, and fig. S22) (24). Simulations suggest that high levels of gene flow from *X. birchmanni* would be required to maintain spotting at observed frequencies in hybrid populations (fig. S22) (24). However, because our inferences are based on viability rather than direct measures of fitness, we stress that there may be weaker effects of melanoma on overall fitness. Alternatively, other factors, such as mating advantages in individuals with large spots (40, 41), may explain its maintenance.

Implications

The involvement of *xmrk* in a melanoma hybrid incompatibility in two distantly related swordtail species pairs raises the question of whether certain genetic interactions are particularly prone to breakdown in hybrids. Genes that interact with many other genes or those that are involved in evolutionary arms races may be especially likely to generate hybrid incompatibilities (such as observed in *Arabidopsis*)

(42, 43). Indeed, the only other known hybrid incompatibility in vertebrates, the recombination hotspot regulator *prdm9*, causes hybrid sterility in multiple crosses in mice (44). Whether unifying molecular or evolutionary forces drive the evolution of hybrid incompatibilities will become clearer as more incompatibilities are mapped to the single-gene level.

REFERENCES AND NOTES

1. T. Dobzhansky, *Zellforsch.* **21**, 169–223 (1934).
2. N. A. Johnson, *Genetics* **161**, 939–944 (2002).
3. T. Dobzhansky, *Biol. Rev. Camb. Philos. Soc.* **11**, 364–384 (1936).
4. J. A. Coyne, H. A. Orr, *Speciation* (Sinauer Associates, 2004).
5. D. C. Presgraves, *Nat. Rev. Genet.* **11**, 175–180 (2010).
6. S. Maheshwari, D. A. Barbash, *Annu. Rev. Genet.* **45**, 331–355 (2011).
7. D. A. Barbash, P. Awadalla, A. M. Tarone, *PLoS Biol.* **2**, e142 (2004).
8. N. A. Johnson, *Trends Genet.* **26**, 317–325 (2010).
9. E. H. Stukenbrock, F. B. Christiansen, T. T. Hansen, J. Y. Dutheil, M. H. Schierup, *Proc. Natl. Acad. Sci. U.S.A.* **109**, 10954–10959 (2012).
10. Y. Brandvain, A. M. Kenney, L. Fligel, G. Coop, A. L. Sweigart, *PLoS Genet.* **10**, e1004410 (2014).
11. R. Cui et al., *Evolution* **67**, 2166–2179 (2013).
12. S. Sankararaman et al., *Nature* **507**, 354–357 (2014).
13. D. A. Turissini, D. R. Matute, *PLoS Genet.* **13**, e1006971 (2017).
14. O. Mihola, Z. Trachtulec, C. Vlcek, J. C. Schimenti, J. Forejt, *Science* **323**, 373–375 (2009).
15. M. Gordon, *J. Hered.* **28**, 223–230 (1937).
16. S. Meierjohann, M. Schartl, *Trends Genet.* **22**, 654–661 (2006).
17. M. Schartl, *BioEssays* **30**, 822–832 (2008).
18. E. Clark, L. R. Aronson, M. Gordon, *Bull. Am. Mus. Nat. Hist.* **103**, 135–226 (1954).
19. Z. W. Culumber et al., *Mol. Ecol.* **20**, 342–356 (2011).
20. M. Schumer et al., *eLife* **3**, e02535 (2014).
21. M. Schumer, Y. Brandvain, *Mol. Ecol.* **25**, 2577–2591 (2016).
22. M. Schumer et al., *Science* **360**, 656–660 (2018).
23. Z. W. Culumber, *Zebrafish* **11**, 57–70 (2014).
24. Materials and methods are available as supplementary materials.
25. B. Klotz et al., *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **208**, 20–28 (2018).
26. A. Schartl, B. Malitschek, S. Kazianis, R. Borowsky, M. Schartl, *Cancer Res.* **55**, 159–165 (1995).
27. A. Gómez, J. N. Volff, U. Hornung, M. Schartl, C. Wellbrock, *Mol. Biol. Evol.* **21**, 266–275 (2004).
28. J. N. Volff, M. Schartl, *J. Struct. Funct. Genomics* **3**, 139–150 (2003).
29. J. C. Jones, J.-A. Perez-Sato, A. Meyer, *Mol. Ecol.* **21**, 2692–2712 (2012).
30. J. S. Ramalho, V. S. Lopes, A. K. Tarafder, M. C. Seabra, A. N. Hume, *Pigment Cell Melanoma Res.* **22**, 461–473 (2009).
31. R. Corbett-DeGig, R. Nielsen, *PLoS Genet.* **13**, e1006529 (2017).
32. S. Xu, *Genetics* **165**, 2259–2268 (2003).
33. I. Gomez-Mestre, D. R. Buchholz, *Proc. Natl. Acad. Sci. U.S.A.* **103**, 19021–19026 (2006).
34. M. Schartl et al., *J. Invest. Dermatol.* **130**, 249–258 (2010).
35. J. Regneri et al., *Pigment Cell Melanoma Res.* **32**, 248–258 (2019).
36. M. Safaee et al., *Int. J. Oncol.* **43**, 1343–1350 (2013).
37. D. Liu et al., *PLoS ONE* **7**, e39989 (2012).
38. Y. Ward et al., *Cell Rep.* **23**, 808–822 (2018).
39. J. Wittbrodt et al., *Nature* **341**, 415–421 (1989).
40. A. A. Fernandez, M. R. Morris, *Proc. Natl. Acad. Sci. U.S.A.* **105**, 13503–13507 (2008).
41. Z. W. Culumber, G. G. Rosenthal, *Naturwissenschaften* **100**, 801–804 (2013).
42. E. Chae et al., *Cell* **159**, 1341–1351 (2014).
43. R. Alcázar et al., *PLoS Genet.* **10**, e1004848 (2014).
44. B. Davies et al., *Nature* **530**, 171–176 (2016).
45. F. Supek, M. Bošnjak, N. Škunca, T. Šmuc, *PLoS ONE* **6**, e21800 (2011).
46. S. Meierjohann, T. Mueller, M. Schartl, M. Buehner, *Zebrafish* **3**, 359–369 (2006).
47. D. Powell, Natural hybridization reveals incompatible alleles that cause melanoma in swordtail fish. Dryad (2020).
48. M. Schumer, schumerm/Powell_et_al_hybrid_melanoma_manuscript: Spotted caudal hybrid incompatibility scripts release. Zenodo (2020).

49. M. Schumer, Schumerlab/mixnmatch: Mixnmatch version used in Powell et al 2020. Zenodo (2020).
50. M. Schumer, Schumerlab/ncASE_pipeline: ncASE pipeline version used in Powell et al 2020. Zenodo (2020).

ACKNOWLEDGMENTS

We thank E. Calfee; H. Fraser; B. Kim; M. Lipson; P. Moorjani; M. Przeworski; and members of the Reich, Rosenthal, and Schumer laboratories for feedback on this work. We appreciate the help of A. Moyaho-Martinez in the design of swim trials, J. Lim and members of the Rosenthal laboratory in collecting samples, and O. Juárez-Mora for help conducting swim trials. We thank the federal government of Mexico for permission to collect fish. Stanford University and the Stanford Research Computing Center

provided computational support for this project. **Funding:** This work was supported by NSF LTREB 1354172 to G.G.R.; funding from the Hagler Institute for advanced study to M.Scha.; and a Hanna H. Gray fellowship, L'Oreal for Women in Science grant, and NIH 1R35GM133774 grant to M.Schu. **Author contributions:** D.L.P., M.G.-O., and M.Schu. designed the project; D.L.P., M.G.-O., M.K., A.P.D.-L., S.B., D.B., M.Schu., and M.Scha. collected data; D.L.P., M.G.-O., M.K., K.D., P.R., and M.Schu. performed analyses; D.R., P.A., M.Scha., and G.G.R. provided expertise and technical support. **Competing interests:** The authors declare no competing interests. **Data and materials availability:** Data are available through NCBI (BioProject PRJNA610049; SRA accessions: SRX7847847-SRX7847871, SRX7866838-SRX7867011, SRX7861514-SRX7861761, and SRX7860174-SRX7860180) and

Dryad (47). Code is available on github (<https://github.com/Schumerlab>) and archived at Zenodo (48–50).

SUPPLEMENTARY MATERIALS

science.sciencemag.org/content/368/6492/731/suppl/DC1
Materials and Methods
Figs. S1 to S41
Tables S1 to S3
Appendix S1
References (51–108)
Movies S1 and S2

12 December 2019; accepted 27 March 2020
10.1126/science.aba5216

Natural hybridization reveals incompatible alleles that cause melanoma in swordtail fish

Daniel L. Powell, Mateo García-Olazábal, Mackenzie Keegan, Patrick Reilly, Kang Du, Alejandra P. Díaz-Loyo, Shreya Banerjee, Danielle Blakkan, David Reich, Peter Andolfatto, Gil G. Rosenthal, Manfred Scharl and Molly Schumer

Science **368** (6492), 731-736.
DOI: 10.1126/science.aba5216

Mapping vertebrate incompatibility alleles

Deleterious gene interactions may underlie the observed hybrid incompatibilities. However, few genes underlying hybrid incompatibilities have been identified, and most of these involve species that do not hybridize in natural conditions. Powell *et al.* used genome sequencing to map genes likely responsible for incompatibilities that reduce fitness in naturally occurring hybrid swordtail fish. These gene combinations result in malignant melanoma, which is found in naturally hybridizing populations but is not present in the parental populations (see the Perspective by Dagilis and Matute). Using genome and population resequencing, the authors performed a genome-wide association study to identify potentially causative mutations. Using an admixture mapping approach that assessed introgression between multiple swordtail fish species, the authors suggest that lineages carry different genes that interact with the same candidate gene, resulting in the observed melanomas and providing insight into convergent hybrid incompatibles that arise between species.

Science, this issue p. 731; see also p. 710

ARTICLE TOOLS

<http://science.sciencemag.org/content/368/6492/731>

SUPPLEMENTARY MATERIALS

<http://science.sciencemag.org/content/suppl/2020/05/13/368.6492.731.DC1>

RELATED CONTENT

<http://science.sciencemag.org/content/sci/368/6492/710.full>

REFERENCES

This article cites 103 articles, 21 of which you can access for free
<http://science.sciencemag.org/content/368/6492/731#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)

Science (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science* is a registered trademark of AAAS.

Copyright © 2020 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works