

Not All Nitrogen Is Created Equal: Differential Effects of Nitrate and Ammonium Enrichment in Coastal Wetlands

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Excess reactive nitrogen (N) flows from agricultural, suburban, and urban systems to coasts, where it causes eutrophication. Coastal wetlands take up some of this N, thereby ameliorating the impacts on nearshore waters. Although the consequences of N on coastal wetlands have been extensively studied, the effect of the specific form of N is not often considered. Both oxidized N forms (nitrate, NO_3^-) and reduced forms (ammonium, NH_4^+) can relieve nutrient limitation and increase primary production. However, unlike NH_4^+ , NO_3^- can also be used as an electron acceptor for microbial respiration. We present results demonstrating that, in salt marshes, microbes use NO_3^- to support organic matter decomposition and primary production is less stimulated than when enriched with reduced N. Understanding how different forms of N mediate the balance between primary production and decomposition is essential for managing coastal wetlands as N enrichment and sea level rise continue to assail our coasts.

Keywords: reactive nitrogen, salt marshes, PIE LTER, nitrogen cycling, carbon cycling

The course of human history was dramatically changed when, in 1908, Fritz Haber filed a patent for ammonium (NH_4^+) production and his contemporary, Carl Bosch, industrialized the process to increase the scale of production. During the tumultuous first half of the twentieth century, this discovery facilitated arms manufacturing, which required extensive supplies of reactive nitrogen (N_r). Even more transformative was its value in fertilizer production, particularly after World War II, which enhanced food availability across the globe. Today, 40%–50% of the human population depends on food grown with fertilizers resulting from the Haber–Bosch process (Smil 2004, Stewart et al. 2005, Erisman et al. 2008), and therefore, increases in human population are tightly coupled with increases in fertilizer use (Erisman et al. 2008). Today, N_r derived via the Haber–Bosch process has more than doubled the annual supply of biologically available nitrogen entering the biosphere (Fowler et al. 2013).

The cascade of N_r from agricultural fields and other sources, through ecosystems, and into coastal waters is well documented (Galloway et al. 2003, Billen et al. 2013). Fertilizer N can be lost from agricultural land through volatilization to

the atmosphere that is later deposited as wet or dry deposition (Paelz 1995, Hinga et al. 1991), through leaching into surficial water bodies and into groundwater (Forster et al. 1982), and through harvest and subsequent movement of N_r around the planet in the form of food and feed stocks (Galloway et al. 2008) that ultimately enters wastewater streams. Much of this human-derived N_r eventually finds its way to the N-limited coastal zone, where it increases primary production and, in excess, can lead to eutrophic conditions and anoxia (Nixon 1995, Diaz and Rosenberg 2008). Unfortunately, conversion of N_r back to N_2 gas, essentially reversing the Haber–Bosch process, has limited industrial analogs, particular once N_r enters coastal waters. Instead, to prevent eutrophication of the coastal zone, a series of microbial transformations of N must occur for the excess N_r to be returned to inert N_2 gas that is then removed from the ecosystem.

Coastal emergent wetlands—principally, salt marshes in the temperate zone—sit between the land and the sea and intercept N_r before it enters open coastal waters (Valiela et al. 2002, Verhoeven et al. 2006, Sousa et al. 2008, Brin et al. 2010, Nelson and Zavaleta 2012). Coastal wetlands can

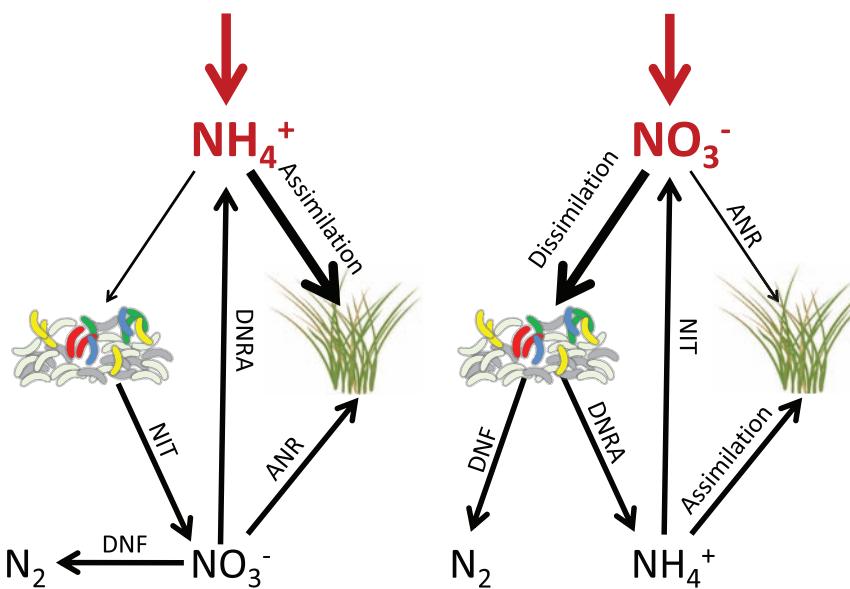


Figure 1. Conceptual diagram illustrating different possible fates of added nitrogen to salt marshes. (a) nitrogen is added in the reduced form of ammonium (NH_4^+). (b) nitrogen is added in the oxidized form of nitrate (NO_3^-). We propose that when nitrogen is added as NO_3^- , it will be predominantly used as an electron acceptor to fuel dissimilatory bacterial respiration rather than to stimulate primary production. ANR = assimilatory nitrate reduction, NIT = nitrification, DNF = denitrification, DNRA = dissimilatory nitrate reduction to ammonium.

retain or remove some fraction of anthropogenically derived N_r delivered to the coastal zone; however, the amount of N_r retained or removed varies widely, with wetlands in eutrophic systems retaining a smaller fraction of total N_r than wetlands that receive lower N_r inputs (Valiela and Cole 2002). The loss of seagrass beds is considered a sentinel indicator of eutrophic conditions (Orth et al. 2006) and estuaries with large areas of emergent wetlands have greater extents of seagrass beds than similar estuaries with no emergent wetlands because of wetlands' N_r removal capacity (Valiela and Cole 2002).

The transformations of N_r that are required to ultimately remove N from ecosystems depend on the initial form of the N and on the geochemical conditions present in the environment. There are several mechanisms by which anthropogenic N_r gets delivered to the coastal zone. N_r derived from sewage waste and fertilizers can be transported to estuaries via river (Peierls et al. 1991, Caraco and Cole 1999) and groundwater flows (Valiela et al. 1990, Caraco and Cole 1999, Bowen et al. 2007, Kroeger and Charette 2008), or can be directly discharged from treatment plants into estuarine and coastal waters. This N_r is typically delivered to estuaries as NO_3^- (Weller and Jordan 2020), with NH_4^+ often making up less than 10% of N_r as a result of oxygen-dependent nitrification, which converts reduced NH_4^+ to NO_3^- through a series of microbially mediated oxidation steps (Ward et al. 2011). Enhanced mobility of NO_3^- compared with NH_4^+ and the preferential uptake of NH_4^+

compared with NO_3^- by macrophytes, underscores the importance of separately identifying the fates of each form of N_r in coastal systems. Reactive N can also be delivered to coastal systems via atmospheric deposition both directly to the estuarine water body and via deposition on land and subsequent delivery to the estuary via river and groundwater flows. Atmospheric deposition by these mechanisms can account for 10%–40% of total N_r inputs to the coastal zone (Fisher and Oppenheimer 1991, Paerl 1995, Paerl et al. 2002). Historically, oxidized N deposition dominated this flux in eastern North America; however, in recent years, the flux of N oxides decreased as a result of emissions reduction strategies adopted by industrial processes (Lloret and Valiela 2016), whereas the flux of reduced N either increased (largely because of volatilization from animal wastes and fertilizers) or remained constant (Gilliam et al. 2019).

The retention and loss of N_r can occur via multiple pathways (figure 1), including uptake into plant biomass and storage in salt marsh sediments (Valiela and

Teal 1979, Drake et al. 2009) and through microbial denitrification, which converts N in its most oxidized form, NO_3^- , to N_2 gas through stepwise reduction (Zumft 1997). This process is largely anaerobic (requiring low oxygen conditions) and heterotrophic (requiring organic carbon as a carbon source; Burgin and Hamilton 2007). Autotrophic (fixing carbon) denitrification also exists, often coupled with reduced iron or sulfur compounds, although its quantitative importance in coastal wetlands is unclear (Rivett et al. 2008). In addition to these denitrification pathways, NO_3^- can also be converted to NH_4^+ via microbes that are capable of dissimilatory nitrate reduction to ammonium (DNRA) or anaerobic ammonia oxidation to N_2 (annamox). DNRA recycles N within the environment, rather than returning this N to the atmosphere as N_2 gas (An and Gardner 2002, Giblin et al. 2013). Organisms capable of DNRA can also be either autotrophic or heterotrophic (Burgin and Hamilton 2007). Annamox also results in loss of N_r (Dalsgaard et al. 2005); however, its importance in coastal wetlands appears limited (Koop-Jakobsen and Giblin 2009). This complex combination of processes, some autotrophic and requiring oxygen and others heterotrophic and requiring anoxic conditions, coupled with different ionic interactions with soil particles (e.g., NH_4^+ readily binds to clay particles in soils), ultimately dictates the dominant form of N_r in the environment. Understanding the specific pathways involved in the processing of N_r in the coastal zone is therefore essential for predicting the stability of coastal wetlands because shifts in

N cycle processes can have simultaneous implications for carbon cycling in these critical carbon-rich habitats (Bulseco et al. 2019).

When N_r enters coastal salt marshes as NO₃⁻, unlike its reduced forms, it can play two distinct roles (figure 1) with very different consequences at the ecosystem scale. As with NH₄⁺ and organic based fertilizers (figure 1a), NO₃⁻ can be used to relieve nutrient limitation and stimulate primary production of vascular plants, benthic algae, and phytoplankton. However, unlike these other forms of N, in the absence of oxygen, it can also be used as an electron acceptor to fuel microbial respiration through denitrification or DNRA (figure 1b). These two opposite outcomes—stimulation of primary production and enhanced decomposition—make it essential that we understand how the forms of N_r entering our coastal waters are being used. If the added NO₃⁻ relieves nutrient limitation and supports primary production, it will increase plant biomass, which will facilitate sediment trapping. This increased trapping of sediment, combined with increased peat build up through production of roots and rhizomes, increases the marsh's ability to keep pace with sea level rise (Kirwan et al. 2010, Kirwan and Megonigal 2013, Morris et al. 2013). However, if NO₃⁻ is primarily being used as an electron acceptor by the microbial community, this could stimulate microbial respiration and potentially decrease the rate of wetland carbon storage and destabilize marsh creeks (Deegan et al. 2012). By contrast, if the N_r is added in its reduced form, as NH₄⁺, the ecosystem scale outcome will depend on the balance between plant uptake and microbial nitrification. If microbes are able to use the NH₄⁺ in nitrification to produce NO₃⁻ that is coupled to denitrification, then increased NH₄⁺ should also ultimately stimulate microbial N₂ production through coupled nitrification and denitrification; however, if the plants have a higher affinity for available NH₄⁺, then we would expect increased primary production by marsh macrophytes.

Below, we synthesize multiple lines of evidence demonstrating that, when NO₃⁻ supply is abundant, primary production rates are lower than are achieved by the addition of comparable supplies of NH₄⁺ and that microbial decomposition is stimulated by NO₃⁻ additions. Therefore, it is imperative that we consider not just the quantity but also the form of N_r entering coastal systems to properly manage and mitigate its downstream consequences.

NO₃⁻ additions have a smaller effect on primary production of the foundation species *Spartina* than other forms of N_r

The salt marsh vegetation throughout the East and Gulf Coasts of the United States and in Europe is generally N limited (Valiela and Teal 1979, Kiehl et al. 1997, Tyler et al. 2003) and the addition of exogenous N, particularly when added as NH₄⁺ or organic based fertilizers, typically increases primary production of marsh grasses (figure 2). The foundation species along the East Coast of the United States, *Spartina alterniflora* and *Spartina patens*, are both

capable of taking up NO₃⁻ through their roots (Smith and McLachlan 1979, Mendelsohn 1979b, Cott et al. 2018). *S. alterniflora* can also take up NO₃⁻ through its shoots (Mozdzer et al. 2011), although we could find no studies that examined NO₃⁻ uptake via shoots in *S. patens*. In a field study in North Carolina, nitrate reductase activity was much lower than glutamate dehydrogenase activity (induced by NH₄⁺ availability) across all growth forms of *Spartina alterniflora* in both the shoots and the roots (Mendelsohn 1979b), suggesting that NH₄⁺ was the preferred nutrient for *S. alterniflora* growth. A nutrient enrichment experiment indicated that NO₃⁻ addition did increase the expression of nitrate reductase in *S. alterniflora*; however, activity under high ambient NO₃⁻ concentrations was still dramatically lower than activity of glutamate dehydrogenase (Mendelsohn 1979b). In addition, foliar uptake of *S. alterniflora* was around 70% higher for NH₄⁺ and glycine than for NO₃⁻ (Mozdzer et al. 2011). *S. patens*, similarly showed uptake rates of NH₄⁺ that were an order of magnitude higher than NO₃⁻ across a range of nutrient concentrations (Cott et al. 2018), further suggesting that, although these *Spartina* species are capable of NO₃⁻ uptake, they prefer N in the form of NH₄⁺.

We performed a meta-analysis of field studies focusing on *Spartina* marsh nutrient enrichment to assess whether the form of N_r used in an experiment affected measured outcomes. We included in our study any salt marsh nutrient enrichment experiments where the enriched habitat consisted of predominantly *S. alterniflora* or *S. patens*, both the form and quantity of N_r used in the experiment were clearly stated, N_r addition results were presented relative to an unenriched reference, and standard deviation or standard error of the mean and sample size for the response variable were reported. There were too few studies documenting the response of belowground vegetation so we focused on the response of aboveground *Spartina* biomass to N_r enrichment (figure 2).

Twenty-two studies from the Western Atlantic ($n = 17$), the Gulf of Mexico ($n = 3$) in the United States, and China ($n = 2$) clustered into four different types of added N_r: NH₄⁺ ($n = 7$), NH₄NO₃ ($n = 4$), other forms of NO₃⁻ ($n = 4$), and organic N, either in the form of pelletized sewage sludge or organic fertilizer or as urea ($n = 7$). We used the “escalc” function in the metafor v.2.1-0 R package (Viechtbauer 2010) to calculate the mean and variance in effect size using ROM, the log transformed ratio of the means (Hedges et al. 1999, Lajeunesse 2011), as the effect size measure while specifying the means, standard deviations, and sample sizes from each study.

We found that added N_r in the form of NO₃⁻ (excluding N_r that was added as NH₄NO₃) had a smaller effect size on plant biomass than other forms of N_r (figure 2a). When studies were aggregated together by the form of N_r added (figure 2b) the mean effect size was significantly less (ANOVA, $F(3,40) = 9.89$, $p < .01$) in the NO₃⁻ only additions (Tukey's posthoc test: NO₃⁻ versus NH₄⁺, $p \leq .01$;

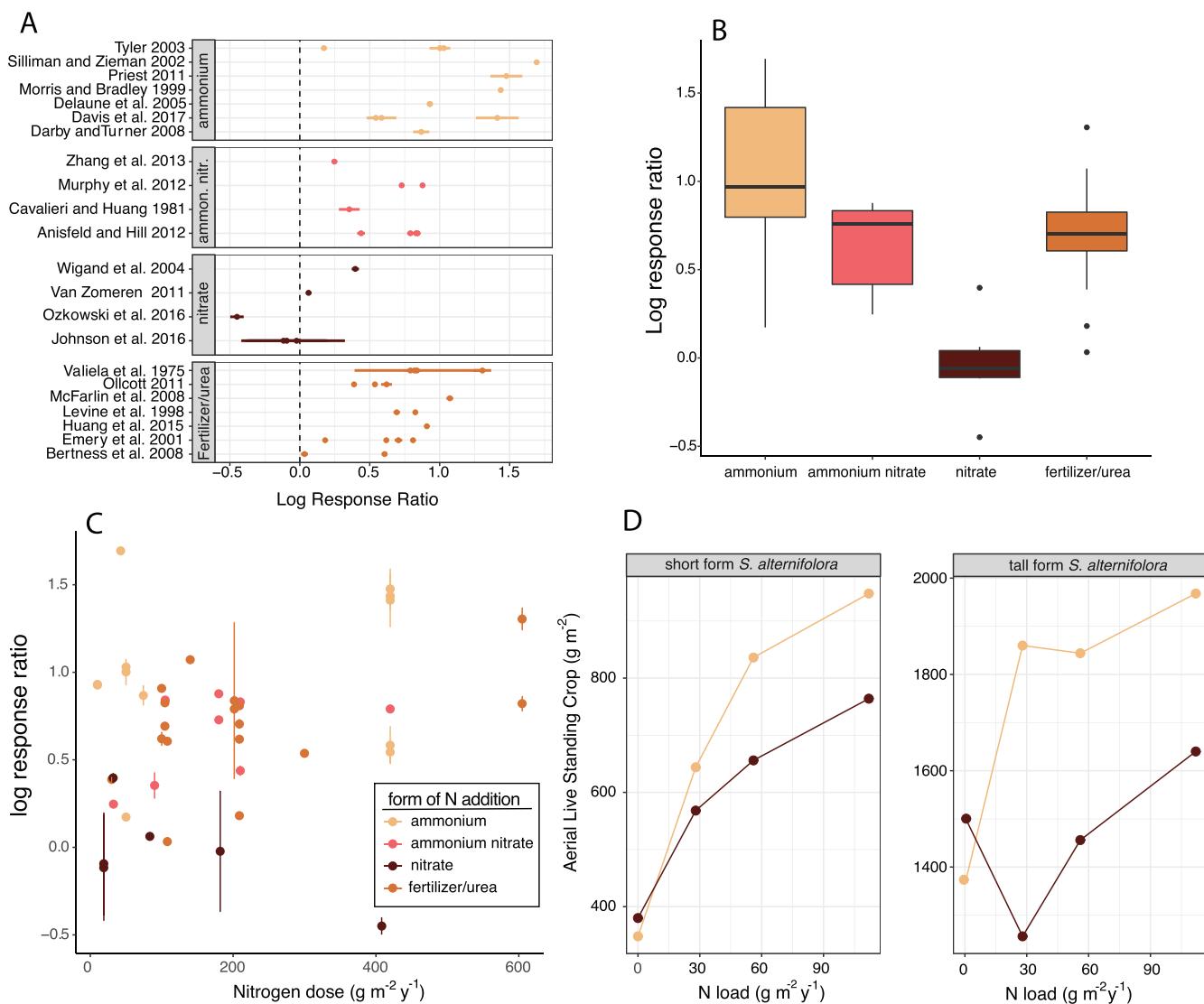


Figure 2. Results of meta-analysis that suggest response to N_r additions in the form of NO_3^- leads to a smaller positive effect on aboveground biomass of *Spartina* plants than when other forms of nitrogen enrichment are added. (a) Log response ratios of studies that report response to N_r in enriched compared with reference salt marsh samples separated by the form of N added. (b) Box and whisker plot of the log response ratios reported in panel (a). The center line represents the median response, upper and lower edges of the box represent the upper and lower quartiles, respectively, and the whiskers indicate the highest and lowest values. (c) Log response ratio plotted as a function of the amount of N added. (d) Aboveground biomass by different *Spartina* ecotypes to the direct addition of NO_3^- (brown) and NH_4^+ (yellow) under identical experimental conditions. Source: The data were plotted from table 1 in Mendelsohn (1979a).

NO_3^- versus NH_4NO_3 , $p = .044$; NO_3^- versus organic fertilizer N or urea, $p \leq .01$). There was no relationship between the quantity of N added and the effect size across all treatments (figure 2c), suggesting that the muted responses observed in NO_3^- addition experiments were not the result of a lower overall amount of N_r added in the enrichment experiments, but rather, it was the form of N that played a consequential role.

Mendelsohn (1979a) performed the only study that directly compared the change in *Spartina alterniflora* aboveground biomass when plants were grown separately

with NO_3^- or NH_4^+ as the added N_r source. This study was not included in our meta-analysis because no error was reported; however, the results are consistent with our hypothesis of a muted response by *S. alterniflora* to N_r enrichment when it is added as NO_3^- (figure 2d). Mendelsohn measured live standing crop of both the tall ecotype and the short ecotype of *S. alterniflora* from the East Coast (Walden Creek Marsh, North Carolina) under different nutrient regimes. The experiment used a $4 \times 2 \times 2$ factorial design with four N application rates ranging from 0 to 112 grams per square meter per year

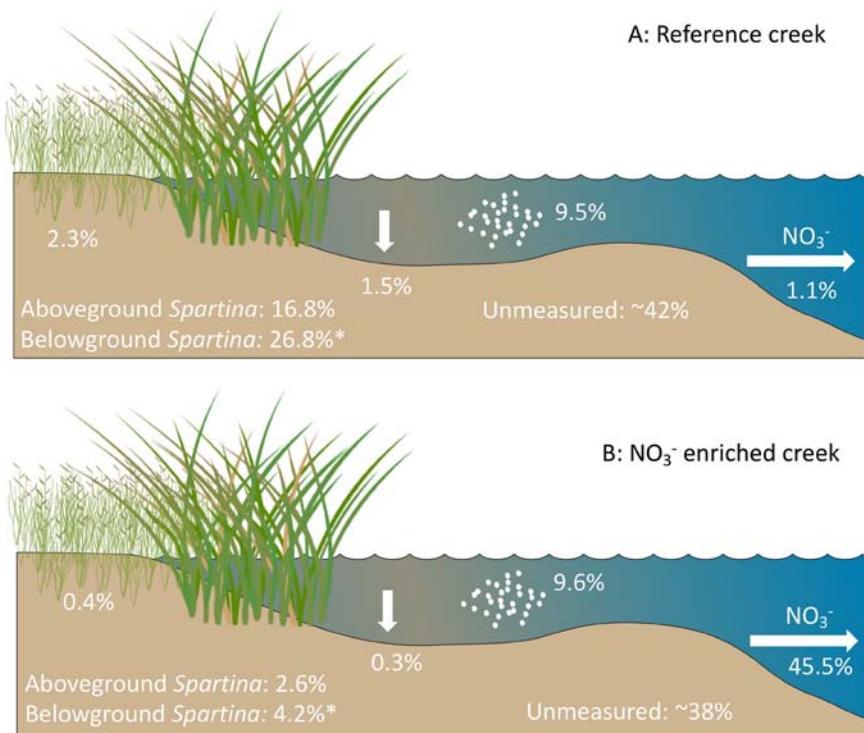


Figure 3. Mass balance of $^{15}\text{NO}_3^-$ added in low dose to an unenriched reference salt marsh creek (a) and in high dose to a NO_3^- enriched salt marsh creek (b) during a paired ^{15}N isotope enrichment experiment (Drake et al. 2009). The reference marsh retained 98.8% of added NO_3^- , compared with 54.5% in the enriched marsh. The percentages are the percentage of added NO_3^- found in each marsh pool after 5 days of dosing. NO_3^- that was unaccounted for was assumed to be lost via dissimilatory microbial processes such as denitrification. Asterisk indicates that the value was estimated and not directly measured. More details on the experimental design and measurements can be found in Drake and colleagues (2009). Source: The images of marsh vegetation were compiled from the IAN symbol repository.

of added N_p , two different application methods (band, where N_r was injected into the sediment, and broadcast, where N_r was placed on the surface sediments), and two different N forms, $(\text{NH}_4)_2\text{SO}_4$ and NaNO_3 . For both the short and tall ecotypes of *Spartina alterniflora* when N_r was added as $(\text{NH}_4)_2\text{SO}_4$ there was a greater increase in growth than when it was added as NaNO_3 (figure 2d). We show only the results of band delivery in the present article but the response to the broadcast delivery method were comparable (Mendelsohn 1979a). Our meta-analysis and Mendelsohn's direct test indicate that *Spartina* biomass accumulation is lower with NO_3^- enrichment, which could have consequences for the long-term sustainability of marshes in the face of rising sea levels.

Whole creek $^{15}\text{N-NO}_3^-$ enrichment experiments support rapid uptake by microalgae and little NO_3^- uptake by *Spartina*

In unvegetated areas of salt marshes and even in under-story areas of the marsh where there is sufficient light

penetration, benthic microalgae, along with macrophytes and denitrifying bacteria (Sundback and Miles 2000), use N_r . Benthic diatoms are capable of NO_3^- uptake and storage (Lomas and Glibert 2000); however, responses to the addition of N_r by benthic microalgae vary widely, depending on the study (Sullivan and Currin 2002). Whole ecosystem additions of enriched $^{15}\text{N-NO}_3^-$ in saltmarshes quickly and consistently result in highly labeled benthic algae (Hughes et al. 2000, Tobias et al. 2003a, Tobias et al. 2003b, Drake et al. 2009, Galván et al. 2011), suggesting uptake of NO_3^- . However, the measured uptake kinetics of benthic microalgae were consistently higher when provided ammonium, compared with NO_3^- in estuarine sediments of the Nakdong Estuary on the Korean Peninsula (Longphuirt et al. 2009). Our 13-year salt marsh NO_3^- enrichment experiment, referred to as the TIDE project, showed that the addition of NO_3^- did not appreciably increase standing stock biomass of benthic microalgae (Deegan et al. 2007), although high grazing could have masked an increase in algal productivity (Galván et al. 2011, Pascal and Fleeger 2013).

The partitioning of NO_3^- between salt marsh macrophytes, benthic microalgae, and microbial dissimilatory nitrate respiration pathways can partly be disentangled using whole marsh $^{15}\text{N-NO}_3^-$ addition experiments (Drake

et al. 2009). As a part of the TIDE project, we performed a whole-creek marsh $^{15}\text{N-NO}_3^-$ addition experiment in a creek that was amended to 70 micromolar (μM) NO_3^- dissolved into flooding tidal waters on every tide for 2 years and in reference creeks that had ambient (less than 5 μM) NO_3^- concentrations. $^{15}\text{N-NO}_3^-$ was added to both creeks for 5 days in midsummer (23–28 July 2005). In the NO_3^- enriched creek, enough $^{15}\text{N-NO}_3^-$ was added to maintain creek water at 650‰ enrichment while keeping the creek NO_3^- concentration at approximately 70 μM . In the reference creek we maintained a target enrichment of 1000‰, which resulted in a 3%–11% increase in the creek NO_3^- concentration (Drake et al. 2009). The reference creek was able to retain 98.8% of the added $^{15}\text{N-NO}_3^-$ (figure 3a). Aboveground plant biomass accumulated approximately 17% of the isotope label and belowground biomass was estimated to take up an additional approximately 25%, although belowground accumulation was not directly measured. The sediment pool, including benthic microalgae, took up less than 2% of the added NO_3^- . After all

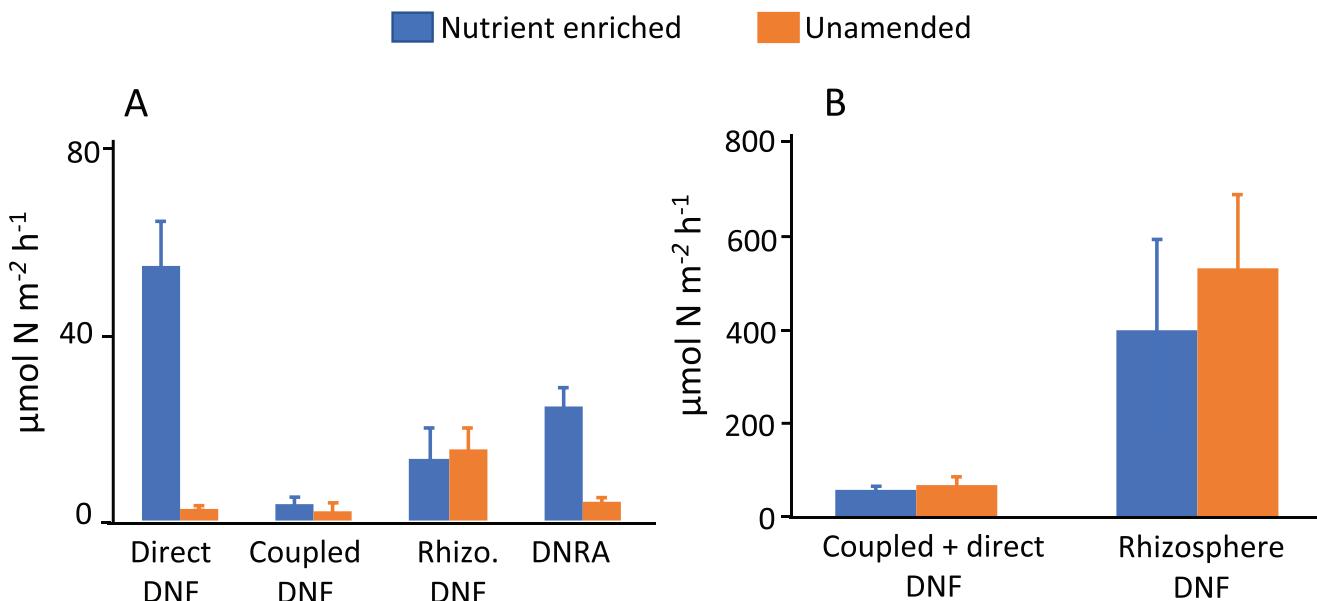


Figure 4. Field- and lab-based assessments of the fate of added NO₃⁻ in salt marsh sediments. (a) Direct denitrification, coupled denitrification, rhizosphere (rhizo.) associated denitrification and dissimilatory reduction of nitrate to ammonium (DNRA) rates measured on the marsh platform in NO₃⁻ enriched and unamended marshes (adapted from Koop-Jakobsen and Giblin 2010). (b) rates of coupled plus direct denitrification and rhizosphere denitrification in cores collected from NO₃⁻ and unamended marshes when exposed to comparable supplies of exogenous NO₃⁻.

the measured pools were accounted for, 42% of the added NO₃⁻ in the reference marsh remained unaccounted for and was assumed to be lost via microbial denitrification. In the NO₃⁻ enriched marsh, 54.5% of added NO₃⁻ was retained in the plant and sediment pools. Although the percentage retention in the enriched marsh was lower than in the reference marsh, the total mass of NO₃⁻ retained in the system was higher, because of an overall larger mass of NO₃⁻ added during the experiment. In the enriched creek, the plants and benthic microalgae retained less than 10% of the added NO₃⁻ and approximately 38% was unaccounted for and assumed to be lost via microbial denitrification. The remainder was exported from the creek in the tidal waters. These enrichment experiments occurred toward the latter part of the growing season of *Spartina*, when biomass accumulation had slowed. It remains to be seen whether similar partitioning of added NO₃⁻ would be observed during times of peak *Spartina* growth. These results further support our hypothesis that salt marsh vegetation and the benthic microalgal communities use a small amount of added NO₃⁻ compared with presumed dissimilatory NO₃⁻ respiration performed by the microbial community.

Both field and controlled laboratory experiments support microbial use of added NO₃⁻

The whole creek ¹⁵N-NO₃⁻ isotope enrichment experiment, both in enriched and reference creeks, had high proportions of ¹⁵N-NO₃⁻ that were unaccounted for and that were

assumed to be lost via microbial denitrification (Drake et al. 2009); however, denitrification was not directly measured. Additional work by Koop-Jakobsen and Giblin (2010) measured the rates of DNRA, denitrification that is coupled with nitrification, direct denitrification, and rhizosphere associated denitrification in specific subhabitats (creek sediments and high marsh platform) within nutrient enriched and unamended creeks. During flooding tides, the addition of NO₃⁻ stimulated direct denitrification, which was nearly twentyfold higher on the fertilized marsh platform than at the reference site (figure 4a). DNRA on the fertilized platform was also stimulated although it was only a fourfold stimulation. In contrast, there was no stimulation of coupled denitrification in either the surface sediments or the rhizosphere (figure 4a).

The major reason for this increase in direct denitrification appears to be a direct and rapid response to added NO₃⁻. Pore water measurements showed that the flooding NO₃⁻ did not reach the rhizosphere in the high marsh and therefore rates were similar between reference and enriched sites. When sediments collected from reference sites were amended with NO₃⁻ in whole core incubations, denitrification rates increased and did not differ from sediments in enriched sites when both treatments received comparable amounts of NO₃⁻ (figure 4b). Similarly, when NO₃⁻ was added to the rhizosphere of sediments from the reference and the enriched sites, there was a large stimulation in denitrification rates in sediments from both sites. Therefore, in all cases when NO₃⁻ was directly added to

Table 1. Total nitrate reduction, denitrification (DNF) and dissimilatory nitrate reduction to ammonium (DNRA) integrated across the duration of the flow through reactor experiment.

Depth	Nitrate reduction (mmol per cm ³)		DNF (mmol per cm ³)		DNRA (mmol per cm ³)		DNRA + DNF (mmol per cm ³)		Percentage of nitrate reduction
	Mean (M)	Standard deviation (SD)	M	SD	M	SD	M	SD	
Shallow (0–5 cm)	87.5	7.4	70.6	1.7	10.2	0.6	80.9	1.8	92
Mid (10–15 cm)	61.3	4.1	58.1	3.7	4.8	1.4	63	4.0	102
Deep (20–25 cm)	70.9	20.4	55.2	10.3	3.6	0.9	58.8	10.3	82

Note: The sum of DNF and DNRA accounted for between 82%–102% of the total amount of nitrate reduced during the experiment. The data were derived from Bulseco and colleagues (2019). Abbreviations: cm³, cubic centimeters; mmol, millimoles.

marsh sediments, both to the rhizosphere and to the overlying water in intact cores, there was a rapid increase in microbial respiration of NO₃[−] through denitrification. *In situ* measurements of ecosystem respiration (Geoghegan et al. 2018) and soil respiration (Wigand et al. 2018) were also significantly higher in the nutrient enriched creeks, providing further evidence of NO₃[−] stimulated microbial processes. This suggests that salt marsh sediments have populations of microbes capable of using NO₃[−] within hours of it being added and implies that there is a sufficient supply of biologically available salt marsh organic matter to support this NO₃[−] respiration.

To explore the capacity of salt marsh microbes to use NO₃[−] to decompose organic matter over longer time periods, we used a flow through reactor (FTR) approach (Bulseco et al. 2019). In an FTR, seawater flows through reactors filled with homogenized marsh sediment and peat, half of which were enriched for approximately 90 days with ¹⁵N-NO₃[−], and we measured total NO₃[−] reduction on the basis of the difference between the NO₃[−] concentration entering the reactor and the concentration leaving the reactor. Simultaneously, we measured production of ³⁰N₂ (denitrification) and production of ¹⁵NH₄⁺ (DNRA) in sediments collected from the top 5 centimeters (cm), 10–15 cm, and 20–25 cm depth from a *Spartina* marsh (table 1). Regardless of the depth from which the sediments were collected, NO₃[−] addition resulted in a stimulation of decomposition (Bulseco et al. 2019). When we added enough NO₃[−] that it was never limiting, denitrification was the dominant NO₃[−] loss process (table 1). The combined rates of denitrification and DNRA accounted for the total amount of NO₃[−] that was consumed by the reactors (table 1), further supporting our supposition that seemingly biologically unavailable salt marsh peat can support substantial rates of denitrification.

NO₃[−] respiration liberates more free energy for microbes than respiration via sulfate reduction or fermentation, two of the most important organic matter decomposition processes in salt marsh sediments (Howarth 1984). Therefore, we hypothesized that the addition of NO₃[−] would not only stimulate additional decomposition, but also that there would be a pool of complex organic matter that could be decomposed in the presence of NO₃[−] that

would not be decomposed by microbes using less energetically favorable electron acceptors, such as sulfate (Bulseco et al. 2019). Indeed, deeper sediments showed less stimulation of dissolved inorganic carbon (DIC) production than surface sediments, indicating that the quality of the carbon in salt marsh sediments does differ; however, at all depths, respiration was stimulated by NO₃[−] addition (Bulseco et al. 2019).

We also measured the byproducts of respiration in the flow through reactor experiment and calculated the ratio of DIC:NH₄⁺ produced over time (figure 5). We found that in surface sediments, where newly deposited biologically available carbon is abundant, there was little difference in the DIC:NH₄⁺ ratio between reactors that received NO₃[−] and those that did not. However, in the mid-depth and deep sediments, where the organic matter is likely less biologically available and more complex, there was an increase in the DIC:NH₄⁺ ratio when NO₃[−] was added compared with the unamended reactors. This suggests that the addition of NO₃[−] allowed the microbes to access a pool of organic matter that was not accessed in the unamended reactors. In the absence of added NO₃[−], this carbon would remain stable, which has implications for our understanding of carbon storage in eutrophic systems. If excess NO₃[−] in coastal systems allows for enhanced decomposition of more complex carbon it could slow the rate of carbon storage in these systems, relative to systems that do not receive excess NO₃[−] supply.

To confirm the microbial role in NO₃[−] use via enhanced denitrification or DNRA and to test our supposition that microbes exposed to high concentrations of NO₃[−] are able to decompose more complex organic matter, we used metagenomics to link our rate measurements to their underlying microbial mechanisms. Analysis of sediment metagenomics is complex because the tremendous diversity of sediment microbes makes deciphering patterns in genetic changes challenging. Previous research on the microbial community structure in the TIDE project indicated that although NO₃[−] increased microbial leucine uptake (a proxy for microbial production) in bare sediment that receives sufficient light to promote increased microalgal production, there did not appear to be a stimulation of

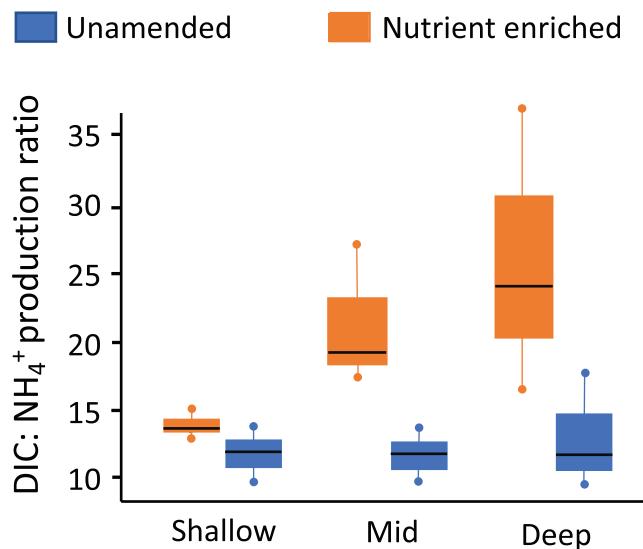


Figure 5. DIC:NH₄⁺ production ratio, as a proxy for the complexity of the organic matter being decomposed. A high ratio indicates a more complex carbon source. Horizontal black bars indicate the median values and whiskers represent the upper and lower quartiles. Source: Adapted from Bulseco and colleagues (2019).

microbial productivity in more heavily vegetated regions of the marsh (Bowen et al. 2009a). Even in regions where leucine uptake was enhanced, the increase in leucine uptake did not translate into shifts in the overall microbial community structure, which was remarkably consistent over a decade of nutrient enrichment (Bowen et al. 2009b, 2011). We did observe large shifts in the active bacterial community, including shifts in bacterial taxa known to denitrify (Kearns et al. 2016), and shifts in putative fungal denitrifiers (Kearns et al. 2019), which suggests that microbial communities using NO₃⁻ as an electron acceptor may be able to increase respiration using NO₃⁻, without leading to a whole scale shift in the community structure.

In light of these results, it is not surprising that we were unable to detect a strong signal of nutrient enrichment in the microbial metagenomes from our field experiment (figure 6a). In the FTR experiments, however, where NO₃⁻ was added at higher concentrations and in a continuous manner, we saw broad shifts in the metagenomes (figure 6b) that are consistent with the biogeochemical rate measurements (Bulseco et al. 2020). Many genes involved in central carbon metabolism were enhanced in the NO₃⁻ addition (figure 6c, supplemental table S1), including increases in the Entner–Doudoroff pathway for the generation of pyruvate from glucose, and decarboxylates that harness that pyruvate for cellular respiration, supporting the hypothesis that the addition of NO₃⁻ allows the microbial community to access otherwise inaccessible organic matter.

Surprisingly, we also observed an increase in genes associated with CO₂ fixation as a result of nutrient enrichment,

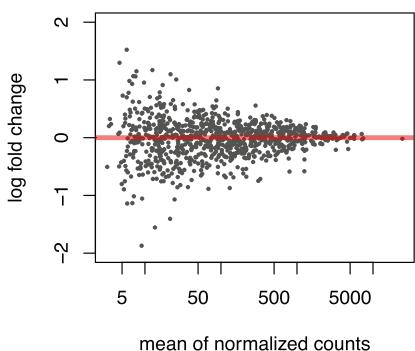
suggesting that added NO₃⁻ could also promote autotrophic denitrification pathways (Bulseco et al. 2020). Many of the genes responsible for the cycling of nitrogen, including all the genes found in the denitrification pathway, were also significantly higher in the NO₃⁻ amended FTRs, compared with the unamended reactors (figure 6d, table S1). By contrast, subsystems involved in fermentation and other low energy producing metabolisms were more abundant in the unamended treatment (figure 6c) as were genes responsible for N fixation (figure 6d). The metagenomics findings from our FTR experiment are consistent with the biogeochemical rates we measured (Bulseco et al. 2020) and provide a mechanistic link that connects microbial genetics to the NO₃⁻ loss and DIC production we observed under NO₃⁻ addition.

Conclusions

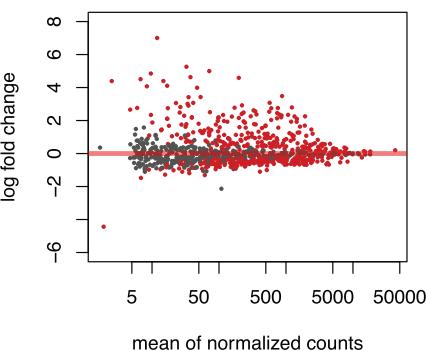
Reactive nitrogen enrichment of the coastal zone is well documented, but frequently, the form of N is overlooked. Our results suggest that understanding the form of N that is biologically available in coastal systems is critically important for managing coastal resources. Most anthropogenically derived N delivered to the coastal zone is in its oxidized form, as NO₃⁻. Our rate measurements, both in the field and in the laboratory, show that NO₃⁻ is being used as an electron acceptor to fuel decomposition, and that this overshadows its role as a nutrient to support growth of plant biomass. Our results suggest that microbes can access available NO₃⁻ and use it as an electron acceptor to respire organic matter that might have otherwise been stored. This process could potentially decrease the carbon sink capacity of marshes because of enhanced decomposition and limit effects on primary production needed to facilitate sediment trapping and offset increased decomposition.

In all likelihood, the use of NO₃⁻ in coastal marsh sediments is not a winner take all scenario, where 100% of the resource goes either to the microbes or to the plants, but rather, the ecosystem outcome is a function of the relative responses of the two groups of organisms. Several unknowns need resolution before predictive models (e.g., the Marsh Equilibrium Model, Morris and Bowden 1986, Morris et al. 2002) can accurately incorporate nutrient supply into their algorithms to predict marsh persistence relative to sea level rise. First, with a carbon to N ratio of around 30, marsh vegetation requires a fairly small amount of N to fix a fairly large amount of carbon and it remains to be seen whether losing a portion of that NO₃⁻ to the microbial community materially alters the productivity or annual storage by vascular plants. Second, we need to better parameterize whether variations in the timing, supply, and duration of use of NO₃⁻ or NH₄⁺ by plants, microalgae, and microbes has long-term consequences for marsh carbon storage. In particular, the variation in growing season length and phenology, which may drive plant nutrient uptake, varies with latitude and could affect response to increasing supply of NO₃⁻ and NH₄⁺. Third, we need a better understanding of how aboveground

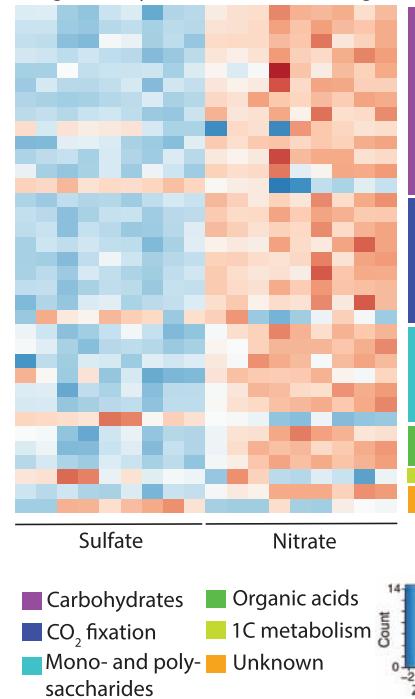
A: Metagenomic analysis of field study



B: Metagenomic analysis of FTR study



C: Significantly different metabolism genes



D: Significantly different N cycle genes

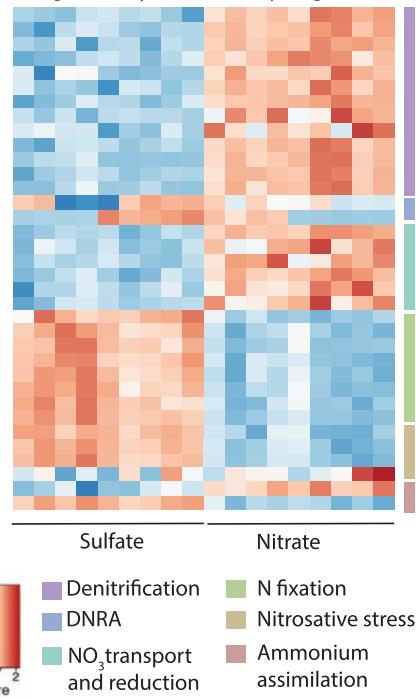


Figure 6. Metagenomic evidence for increased microbial respiration during NO_3^- addition in field and laboratory experiments. (a) Log-fold differences in gene abundance between NO_3^- enriched and unenriched salt marsh surface sediments collected from the TIDE nutrient enrichment experiment. (b) Log-fold differences in gene abundance in FTRs between NO_3^- amended reactors and reactors receiving only seawater. Red points indicate genes that are significantly different between the two treatments. (c) Heat map indicating the 35 biggest differences between amended and unamended FTRs for genes involved in carbon metabolism. (d) Heat map indicating the 35 biggest differences between amended and unamended FTRs for genes involved in the nitrogen cycle. Source: The data used in panels (c) and (d) were adapted from Bulseco and colleagues (2020).

and belowground biomass is partitioned and how these factors feedback on sediment trapping and storage when N is supplied as NO_3^- rather than as NH_4^+ . Finally, our genomic data suggest that chemoautotrophic metabolisms could play a larger than expected role in carbon fixation in nutrient enriched marshes; however, the extent of this needs to

be better parameterized. Understanding these critical unknowns will be essential for predicting marsh carbon storage in a high nutrient world.

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Supplemental material

Supplemental data are available at BIOSCI online.

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