



Main Manuscript for

Evanescence Waves with Trochoidal Polarizations Reveal a New Dichroism

Lauren A. McCarthy^{a,b}, Kyle W. Smith^{a,b,†}, Xiang Lan^{a,b}, Seyyed Ali Hosseini Jebeli^{b,c}, Luca Bursi^{d,e}, Alessandro Alabastri^{b,c,e}, Wei-Shun Chang^f, Peter Nordlander^{b,c,d,e}, Stephan Link^{a,b,c,e*}

Affiliations:

^aDepartment of Chemistry, Rice University, Houston, Texas 77005, USA.

^bLaboratory for Nanoscale Spectroscopic Imaging at Rice, Rice University, Houston, Texas 77005, USA.

^cDepartment of Electrical and Computer Engineering, Rice University, Houston, Texas 77005, USA.

^dDepartment of Physics and Astronomy, Rice University, Houston, Texas 77005, USA.

^eLaboratory for Nanophotonics, Rice University, Houston, Texas 77005, USA.

^fDepartment of Chemistry and Biochemistry, University of Massachusetts Dartmouth, Dartmouth, MA, 02747, USA.

*Corresponding Author: slink@rice.edu

†Current address: Draper Laboratory, Houston, TX, USA.

Classification

Physical Sciences: Applied Physical Sciences

Keywords

Evanescence field polarization, single-particle spectroscopy, plasmonic nanorod dimers, Born-Kuhn model for circular dichroism

Author Contributions

S.L., W.-S.C, L.A.M., and K.W.S. designed the research. L.A.M. fabricated the nanostructures. X.L. synthesized the self-assembled nanostructures. L.A.M. and K.W.S. characterized the nanostructures and analyzed the data. A.H.J. performed FDTD numerical simulations of the nanostructures. L. B. and A. A. performed finite element method COMSOL simulations of the nanostructures and were supervised by P.N. L.A.M., K. W. S., and S.L. wrote the manuscript. All authors contributed to editing the manuscript.

This PDF file includes:

Main Text

Figures 1-4

Abstract

Matter's sensitivity to light polarization is characterized by linear and circular polarization effects, corresponding to the system's anisotropy and handedness, respectively. Recent investigations into the near-field properties of evanescent waves have revealed polarization states with out-of-phase transverse and longitudinal oscillations, resulting in trochoidal, or cartwheeling, field motion. Here, we demonstrate matter's inherent sensitivity to the direction of the trochoidal field and name this new property trochoidal dichroism. We observe trochoidal dichroism in the differential excitation of bonding and antibonding plasmon modes for a system composed of two coupled dipole-scatterers. Trochoidal dichroism constitutes the observation of a geometric basis for polarization sensitivity that fundamentally differs from linear and circular dichroism. It could also be used to characterize molecular systems, such as certain light-harvesting antennas, with cartwheeling charge motion upon excitation.

Significance

The ability of certain materials to discriminate between two opposite light polarizations is the basic principle behind several technologies such as liquid crystal displays and 3D glasses. While there are numerous forms of light polarization, only linear and circular polarizations, which have wave motion in a flat sheet or helix, respectively, are typically used. Here, we utilize trochoidal

polarizations with cartwheeling wave motion. We demonstrate that single gold nanorod dimers can discriminate between trochoidal fields rotating in opposite directions, which we term trochoidal dichroism. Trochoidal dichroism is a new classification of polarized light-matter interaction and could inspire the development of optical studies uniquely sensitive to molecules with cartwheeling charge motion, potentially relevant for probing key light-harvesting antennas.

Introduction

Dichroism is the differential absorption of orthogonal light polarizations by matter and enables optical characterization of molecular geometry. Circular dichroism (CD) suffers from low sensitivity due to its reliance on far-field radiation, whose wavelength is typically orders of magnitude larger than the molecular features it is used to probe. An effective route for improving optical sensitivity is the use of highly confined, structured, light such as evanescent waves (1-5), interference fields (6-8), and focused Gaussian beams (7, 9).

Confined light has distinct polarization properties from free-space radiation. In particular, laterally confined light can support longitudinal electromagnetic oscillations, which enable rich near-field polarization phenomena (9-12). The longitudinal oscillations of an evanescent wave result in a spin angular momentum (SAM) component that is perpendicular, or transverse, to the light propagation direction. Transverse SAM components first gained interest in 2009 (13, 14) and have since been found in evanescent waves to be helicity-independent yet locked to the propagation direction of light, (10, 15) enabling spin-dependent unidirectional propagation (16-18). Kawalec *et al.* (19) and Mitsch *et al.* (20) have found that the transverse SAM of confined light can enable electronic transitions of atoms within a magnetic field (Fig. 1C), exactly analogous to the Zeeman effect observed with circularly polarized light (Fig. 1A). These studies provided some of the first experimental evidence of the existence of longitudinal oscillations and

transverse SAM in evanescent waves. Further, Junge *et al.* (21) demonstrated transverse spin-momentum locking in evanescent waves, convenient for non-destructive interference of counter-propagating light in microresonators. Finally, as predicted by Yang *et al.* (7), and experimentally demonstrated by Mathevet *et al.* (22), transverse SAM in an external magnetic field can give rise to magnetic field-induced CD signals.

The transverse SAM of confined light is also associated with cycloidal, or more generally, trochoidal field motion (10, 12). Trochoidal field motion is both planar and rotational, producing a cartwheeling motion, which results from significant phase delays between transverse and longitudinal oscillations. However, the differential extinction of orthogonal trochoidal polarizations by an analyte based on its geometry, the corresponding analog to CD (Fig. 1B), has not yet been observed (Fig. 1D). In this study, we develop a model metamaterial system composed of coupled dipole-scatterers and observe unique differential scattering with trochoidal fields of opposite rotational directions. The measurement of trochoidal differential scattering is directly analogous to circular differential scattering (23, 24), successfully employed to characterize the optical activity of single chiral nanoparticles (25-27). While CD is historically defined as the differential absorption of left- and right-handed circular polarizations, as only absorption is relevant for small molecules, a CD spectrometer's measurement is based on differential extinction (23). The scattering component of the extinction can significantly contribute to the CD of larger objects such as nanoparticles and macromolecules (23, 28). The trochoidal differential scattering we characterize here is therefore a significant component of a new effect we name trochoidal dichroism.

Result and Discussion

We use total internal reflection (TIR) of light with orthogonal linear polarizations, $E_s=\pm E_p$ ($\pm 45^\circ$) (Fig. 2A), to generate an evanescent wave with significant trochoidal character (SI Appendix, Supplementary Text, Fig. S1). Clockwise (CW) trochoidal motion results from $E_s=-E_p$ (-45°) incident polarization and anticlockwise (ACW) field motion from $E_s=E_p$ (45°) (Fig. 2B). The corresponding phase delays between the sample-plane transverse and longitudinal oscillations are -67° and 112° (Supplementary Text). In addition to the ACW and CW trochoidal fields produced in the sample plane (Fig. 2B), TIR of $E_s=\pm E_p$ polarizations also yields a trochoidal field spinning perpendicular to the sample plane (SI Appendix, Fig. S1), with a transverse SAM component parallel to the plane (S_y). However, S_y is not expected to significantly contribute to trochoidal dichroism in this work as it cannot effectively couple to the sample and its sign is determined by the propagation direction which is held constant in our measurements (10). Therefore, by using TIR of $E_s=\pm E_p$ polarizations, we characterize trochoidal dichroism that arises mainly due to the differential interaction of matter with the vertical transverse SAM component (S_x).

To demonstrate trochoidal dichroism, we probe the differential scattering of CW and ACW trochoidal fields with a model system based on the Born-Kuhn model for CD (29, 30). In this model, left- and right-handed circularly polarized light perform differential work on a chiral spring system composed of orthogonal, coupled harmonic oscillators with attached charges that are displaced along a third orthogonal axis (Fig. 1B). In the trochoidal analog (Fig. 1D), the same two oscillators are instead displaced in the plane containing the two oscillators. Importantly, the second harmonic oscillator's motion is collinear with the light propagation direction, k , to enable coupling with longitudinal oscillations. Trochoidal fields with CW and ACW rotational directions couple differently to the spring isomers, giving rise to trochoidal dichroism.

To realize such a model system, we utilize two orthogonal gold nanorods with a planar offset. The spectral response of gold nanorods is dominated by the coherent oscillation of conduction band electrons known as the localized surface plasmon resonance, accurately approximated as a damped harmonic oscillation (31). Plasmon hybridization between two adjacent nanorods dictates their collective interaction with light, similar to the principles of molecular orbital theory (32). Inspired also by Yin *et al.* who used coupled plasmonic oscillators to describe CD (33), we develop a plasmonic system for the trochoidal analog of the Born-Kuhn model. Specifically, we lithographically fabricate two nanorods with orthogonally aligned axes, aspect ratios of two, lengths of 80 nm, heights of 40 nm, and an interparticle gap of 20 nm (Fig. 2; SI Appendix, Fig. S2).

The coupling of the hybridized plasmon modes to CW or ACW trochoidal excitations depends on dimer shape and orientation relative to k . We first consider a system in which one nanorod is placed parallel to k and the other is perpendicular, forming a \perp -shaped dimer (Fig. 2C). The significant separation of the nanorods relative to the wavelength of light causes each nanorod to experience distinct phases of the electric field. Based on plasmon hybridization theory, the electric field vectors align with both nanorods under CW trochoidal excitation, resulting in the higher-energy antibonding mode (Fig. 2C). Similar considerations predict that ACW field motion excites the lower-energy bonding mode (Fig. 2C). Mirroring the \perp -dimer over the (x, z) plane containing k forms a $\perp\perp$ -dimer, and the opposite polarization-dependent mode excitation is expected (Fig. 2D).

To test these predictions, we record single-particle scattering spectra of both \perp - and $\perp\perp$ -dimers, quantitatively confirming that the bonding and anti-bonding plasmon modes are selectively excited with CW and ACW trochoidal fields. As both modes have a significant net

electric dipole, they are both readily observable in the scattering spectra (34). CW trochoidal excitation yields a single-particle scattering maximum at 740 nm (Fig. 2E) for the J -dimer, while under ACW trochoidal excitation, the resonance is redshifted to 770 nm. Electromagnetic simulations validate the assignments of antibonding and bonding modes (Fig. 2E) and suggest that changes in dimer dimensions can further enhance the spectral shift (SI Appendix, Fig. S3). In particular, charge distributions calculated at the scattering maxima visualize the antibonding and bonding plasmon modes excited with CW and ACW trochoidal fields, respectively (Fig. 2E-inset), in excellent agreement with the plasmon hybridization sketched in Fig. 2C. These results are reproduced with the mirror-image 7 -dimer, but with ACW trochoidal polarization now exciting the higher energy antibonding plasmon (Fig. 2F). Additionally, single-particle spectra and simulations verify that the resonance energy of an individual nanorod indeed lies between the two hybridized dimer modes and is independent of trochoidal excitation (SI Appendix, Fig. S4). Finally, the trochoidal polarization-dependent differential excitation of bonding and antibonding plasmon modes is well-reproduced in simulated absorption spectra (SI Appendix, Fig. S5).

Our results cannot be explained by linear dichroism, as can be demonstrated when examining geometric isomers that show opposite trochoidal and linear dichroism. Because the dimers have anisotropic geometries and our trochoidal excitation has linear components (SI Appendix, Fig. S1, S6), a minor contribution from linear dichroism is expected. While J - and 7 -dimers have trochoidal and linear dichroism of the same sign (SI Appendix, Fig. S6-S7), the L -dimer, formed by mirroring the J -dimer over the (x, y) plane, does not (Fig. 3). With k unchanged, CW trochoidal excitation resulting from TIR of $E_s = -E_p$ (-45°) has electric field vectors aligned with both nanorods, exciting an antibonding mode (Fig. 3A). The net dipole of

this antibonding mode is oriented along the 45° axis (Fig. 3B). In contrast, if the excitation did not possess trochoidal character, $E_s = -E_p$ (-45°) incident polarization would excite the bonding mode oriented along the -45° axis (Fig. 3B). Indeed, CW trochoidal excitation from TIR of $E_s = -E_p$ yields a single-particle scattering resonance that is 10 nm blue-shifted relative to ACW trochoidal excitation (Fig. 3C), well outside the experimental error in resonance shifts (SI Appendix, Fig. S4). Scattering spectra collected with identical polarizations but incident at an oblique angle (SI Appendix, Fig. S8) reveal a 15 nm red-shift for $E_s = -E_p$ (Fig. 3D). This opposite mode excitation when changing between trochoidal and purely linear polarizations demonstrates that our results are not due to linear dichroism, which is well reproduced by electromagnetic simulations (SI Appendix, Fig. S9). In total, each geometric isomer can be related to another by mirror (σ_v) and 180° rotation (C_2) symmetry operations (Fig. 3E). Performing a mirror operation over the (x, y) plane maintains trochoidal dichroism but gives opposite linear dichroism, enabling a clear distinction between the two dichroisms for both L-dimers and Γ -dimers (SI Appendix, Fig. S10).

We further investigate trochoidal differential scattering in self-assembled colloidal nanorod dimers that deviate from the ideal model geometry. We take advantage of DNA origami templates (36-38), formed into rectangles by scaffold single-stranded DNA that is pinned in place by short staple strands (SI Appendix, Fig. S13). Capture DNA strands extend from the template to place gold nanorods in an approximately orthogonal arrangement with a planar offset (Fig. 4A). L-dimers from self-assembled gold nanorods are characterized under CW and ACW trochoidal excitation at two different sample orientations: 0° and rotated 180° relative to k (Fig. 4A, 4B). The C_2 rotation maintains any linear dichroism contributions while giving opposite trochoidal dichroism, enabling their distinction (Fig. 3E, quadrant I \rightarrow III).

We find that observing trochoidal differential scattering is indeed not limited to only precisely-engineered lithographically fabricated models. Deviations from the model-dimer geometry arise in the self-assembled dimers due to particle size heterogeneity, drying of the assembly on a substrate for spectroscopy, and a lack of control over the final orientation of the dimers relative to a fixed k . Relaxing the structural requirements enables components of both CW and ACW trochoidal excitations to map onto the bonding and antibonding modes, resulting in the partial excitation of both modes with each polarization. However, the relative excitation of each mode strongly depends on the direction of trochoidal rotation. Specifically, for the L-dimers, ACW trochoidal excitation predominantly excites a bonding mode (Fig. 4C, E dashed line), directly analogous to the engineered model shown in Fig. 3C. Upon 180° sample rotation, yielding approximate $\bar{\ell}$ -dimers, ACW trochoidal polarization predominantly excites the antibonding plasmon (Fig. 4D, F dotted line), exactly analogous to Fig. 2F. These results are in excellent agreement with the mode selective excitations seen for the model-dimers presented above. Furthermore, the relative mode excitations are well-matched by theory (SI Appendix, Fig. S14) and the linear dichroism is indeed maintained upon C_2 rotation (SI Appendix, Fig. S15).

Conclusion

In conclusion, the observed dependence of the dimers' optical properties on the rotational direction of the trochoidal field constitutes a novel light-matter interaction: trochoidal dichroism. Consistent with theoretical work (7) on confined light polarization states, trochoidal polarizations may be able to probe molecular rotational symmetries to which CD is less sensitive. Relevant mutants of photosynthetic chlorosomes of green sulfur bacteria are composed of chlorophylls that self-assemble into cylinders with charge delocalization around closely stacked rings (39, 40). Trochoidal dichroism, being optimized for a planar rotation of charges, may be able to

effectively probe structure-function relationships in light-harvesting antennas, in a complementary fashion to CD. Further, any achiral molecule with perpendicular electric and magnetic dipoles experiences a planar rotation of charge upon excitation. The rotational direction of the charge is dependent on molecular orientation and could potentially be probed with trochoidal dichroism. Finally, while the longitudinal spin of surface waves can be observed through chiral structures (41), and helicity-invariant-transverse spin can be observed through spin-orbit coupling (16), here we are sensitive to the diagonal-polarization dependent transverse spin component of evanescent waves (10). Thus trochoidal dichroism, as observed through far-field scattering, completes the ability to optically probe the full spin-basis set of a confined electromagnetic wave.

Materials and Methods

Electron Beam Lithography

The lithographic nanorod dimers were fabricated with a designed width of 40 nm, a length of 80 nm, and a height of 40 nm of gold with a 2 nm Ti adhesion layer and an interparticle gap of 20 nm. Single nanorods were fabricated with the same designed dimensions. The dimer arrays each contained 32 structures, which were divided into four sub-arrays containing dimers at different interparticle orientations (SI Appendix, Fig. S2). Indium tin oxide (ITO) coated float glass (Delta Technology LTD CG-50IN-S107) was used as a transparent conductive substrate. The substrates were cleaned through sequential ten-minute sonication in a 2% V/V solution of Hellmanex® detergent, water, and ethanol. Slides were gently dried with a stream of N₂ and then spin-coated with a polymethyl methacrylate electron resist (Michrochem PMMA 950 A4) at 3000 rpm for 60 seconds. The slides were then baked for two minutes at 180° C. Patterns were written into the resist with a Nanometer Pattern Generation System on an FEI Quanta 650 SEM. The beam voltage was 30 kV, with a beam current of 40 pA, a spot size of 2.1, and a working distance of 7 mm. Samples were then developed in a 1:3 methyl isobutyl ketone (MIBK)/isopropanol solution for 70 seconds, followed by a 60-second rinse in isopropanol and gentle drying with N₂. The samples were then placed in an electron beam evaporator, where a 2 nm Ti adhesion layer was deposited, followed by a 40 nm layer of gold. Excess material was removed by soaking the sample in acetone overnight, followed by gentle sonication and drying with N₂.

DNA Origami-based Self-assembly

Rectangular DNA origami (~60 nm × 100 nm; SI Appendix, Fig. S13) was prepared by annealing a mixture of a circular single-stranded M13mp18 DNA with a set of short helper

strands, each ~32 nucleotides, as well as capture strands for positioning the nanorods. All DNA was purchased from Integrated DNA Technologies. Complete sequences of the helper and capture strands are given in Table S1 and S2. M13mp18 DNA was annealed with helper and capture strands at a 1:10 molar ratio in a TE-Mg²⁺ buffer (14 mM magnesium chloride) in a thermocycler programmed for a cooling ramp from 85 °C to 25 °C overnight. The assembled DNA structures were purified using a 30 kDa Amicon filter under centrifugation at 8000 rpm for 15 min. The concentration was estimated by measuring the absorption at 260 nm. 25 nm × 70 nm gold nanorods were purchased from Nanopartz and modified with thiolated single-stranded DNA following published protocols (42). To produce gold nanorod dimers, DNA-decorated nanorods were mixed with the purified DNA origami template with a molar ratio ≥5, followed by annealing from 45°C to 30°C, cycled 4 times over 12 hours. The assembled nanorod dimers were purified through Agarose gel electrophoresis and recovered using a Bio-Rad Freeze 'N SqueezeTM spin column under centrifugation at 13000 rpm for 5 min. Gold nanorod dimers were deposited onto indexed ITO glass and flushed three times in deionized water, then dried under N₂ flow. Finally, the deposited dimers were exposed to an O₂ plasma for 2 minutes to partially remove the DNA to allow high-resolution SEM imaging.

Single-Particle Scattering Spectroscopy

Single-particle scattering measurements were performed with an inverted dark-field microscope (Zeiss, Axio Observer m1). Light from a quartz tungsten halogen lamp (Newport 66884) was coupled by fiber optic cable to a home-built optical rail system. The light was collimated with a plano-convex lens (Thorlabs AC254-030-A) and filtered with a 304-785 nm bandpass filter (Thorlabs FGS550) to protect the film linear polarizer from undue heating. The

light was then polarized with a linear polarizer (Thorlabs LPVIS100) set to either $+-45^\circ$ from the parallel polarized (E_p) axis (Fig. 2A; SI Appendix, Fig. S1). The polarized light was focused with a plano-convex lens (Thorlabs AC254-030-A) onto an equilateral prism. TIR conditions were achieved with an input angle of 55° . Free-space, oblique incidence dark-field excitation was achieved with an input angle on the prism of $\sim 27^\circ$. The scattered light from the nanostructures was collected with either a 50x magnification air-space objective lens (Zeiss, numerical aperture of 0.8) for TIR experiments or a 40x magnification air-space objective lens (Zeiss, numerical aperture of 0.6 with an extra-long working distance) for oblique incidence experiments. The collected light was passed through the microscope and then directed to a hyperspectral detection system that has been described previously (43). Briefly, the output light was directed into a spectrograph (Acton SpectraPro 2150i) and the dispersed light was detected with a back-illuminated CCD camera (Princeton Instruments Pixis 400). The spectrograph and camera were both mounted on a scanning stage driven by a linear actuator (Newport LTA-HL). The data collection was performed in a hyperspectral fashion, with a slit aperture allowing for the collection of multiple spectrally resolved image slices as the spectrograph was scanned over the field of view. Normal incidence measurements were performed with light from a tungsten-halogen lamp (Zeiss, Axioline HAL 100) which was filtered and polarized with the same optics described above. The light was focused on the sample with an oil-immersion condenser (Zeiss, adjustable numerical aperture of 0.7-1.4) in bright-field mode (incident angle of 0°) with the numerical aperture set to 0.7. The scattered light was collected with a 74x reflecting objective (Beck Optronics Solutions, 0.65 numerical aperture) with a direct beam block aligned with the incident excitation, allowing dark-field imaging. Light scattered to a high angle was collected by the objective lens and detected with the same hyperspectral procedure described above. Spectra

with lower signal-to-noise ratios, such as the oblique incidence measurements, were subjected to a 5-point moving boxcar average.

Numerical Simulations

Finite-difference time-domain (FDTD) simulations were performed using the Lumerical FDTD package. The simulation model consisted of nanorods with a length of 85 nm, a width of 35 nm, and a height of 40 nm with a 2nm Ti adhesion layer. The simulated gap between the nanorods was 15 nm and the nanorods were arranged to form \perp , \sqcap , and L -dimers. All simulation parameters were tuned to be within 5 nm of the design parameters to achieve a reasonable resonance match with the experimental spectra. Remaining deviations from the experimentally measured spectra likely originate from the poor crystallinity of lithographically fabricated structures and increased plasmon damping from the Ti adhesion layer that is not captured in the bulk dielectric functions. The dielectric functions of gold and Ti were extracted from Johnson and Christy(44) and Palik (45), respectively, and fitted to a Drude-Lorentz model for the time-domain calculations. The substrate was glass (refractive index $n = 1.52$) with 160 nm of ITO ($n \approx 1.7$) (46). Perfectly matched layers were used to avoid any reflections from the boundaries. For TIR, the incident angle from the substrate medium was 55° from the sample normal. For oblique incidence simulations, the incident angle was 39° degrees. As in experiments, $E_s = \pm E_p$ was used as the incident polarization. The charge plots were calculated using the electric field obtained from the FDTD simulations. The charge distribution varied over the height of the dimers because the presence of a substrate changes the charge distribution compared to vacuum. To account for this variance when calculating the two-dimensional charge plots, the charge distributions were summed over the entire height of the dimers.

Data Availability

Scripts used to analyze data and all data presented in the figures are available in an online directory at <https://rice.box.com/s/rv65ydbiuxaiokxuucgkpjryxs9pelfi>

Acknowledgments

This work is funded by the Robert A. Welch Foundation (C-1664 to S.L. and C-1222 to P.N.) and the National Science Foundation (CHE1903980 to S. L.). L.A.M. and K.W.S. acknowledge that this material is based on work supported by the National Science Foundation Graduate Research Fellowship Program (1842494). We thank the Rice University Shared Equipment Authority and Electron Microscopy Center for instrument use and availability. Finally, the authors thank Mr. Rashad Baiyasi for providing analysis code used for SEM image processing.

Competing Interests

None

References

1. E. Hendry, *et al.* Ultrasensitive detection and characterization of biomolecules using superchiral fields. *Nat. Nanotechnol.* **5**, 783-787 (2010).
2. R. Tullius, *et al.* "Superchiral" Spectroscopy: Detection of Protein Higher Order Hierarchical Structure with Chiral Plasmonic Nanostructures. *J. Am. Chem. Soc.* **137**, 8380-8383 (2015).
3. L. M. Kneer, *et al.* Circular Dichroism of Chiral Molecules in DNA-Assembled Plasmonic Hotspots. *ACS Nano* **12**, 9110-9115 (2018).
4. H. Zhang & A. O. Govorov Giant circular dichroism of a molecule in a region of strong plasmon resonances between two neighboring gold nanocrystals. *Phys. Rev. B* **87**, 075410 (2013).
5. K. W. Smith, *et al.* Exploiting Evanescent Field Polarization for Giant Chiroptical Modulation from Achiral Gold Half-Rings. *ACS Nano* **12**, 11657-11663 (2018).
6. Y. Tang & A. E. Cohen Enhanced Enantioselectivity in Excitation of Chiral Molecules by Superchiral Light. *Science* **332**, 333-336 (2011).
7. N. Yang & A. E. Cohen Local Geometry of Electromagnetic Fields and Its Role in Molecular Multipole Transitions. *J. Phys. Chem. B* **115**, 5304-5311 (2011).
8. Y. Tang & A. E. Cohen Optical Chirality and Its Interaction with Matter. *Phys. Rev. Lett.* **104**, 163901 (2010).
9. A. Aiello, P. Banzer, M. Neugebauer, & G. Leuchs From transverse angular momentum to photonic wheels. *Nat. Photonics* **9**, 789-795 (2015).

10. K. Y. Bliokh, A. Y. Bekshaev, & F. Nori Extraordinary momentum and spin in evanescent waves. *Nat. Commun.* **5**, 3300 (2014).
11. K. Y. Bliokh & F. Nori Transverse spin of a surface polariton. *Phys. Rev. A* **85**, (2012).
12. P. Banzer, *et al.* The photonic wheel: demonstration of a state of light with purely transverse angular momentum. *J. Eur. Opt. Soc.-Rapid* **8**, 13032 (2013).
13. A. Aiello, N. Lindlein, C. Marquardt, & G. Leuchs Transverse Angular Momentum and Geometric Spin Hall Effect of Light. *Phys. Rev. Lett.* **103**, 100401 (2009).
14. A. Y. Bekshaev Oblique section of a paraxial light beam: criteria for azimuthal energy flow and orbital angular momentum. *J Opt a-Pure Appl Op* **11**, 094003 (2009).
15. K. Y. Bliokh, D. Smirnova, & F. Nori Quantum spin Hall effect of light. *Science* **348**, 1448-1451 (2015).
16. J. Petersen, J. Volz, & A. Rauschenbeutel Chiral nanophotonic waveguide interface based on spin-orbit interaction of light. *Science* **346**, 67-71 (2014).
17. D. O'Connor, *et al.* Spin-orbit coupling in surface plasmon scattering by nanostructures. *Nat. Commun.* **5**, 5327 (2014).
18. F. J. Rodríguez-Fortuño, *et al.* Near-Field Interference for the Unidirectional Excitation of Electromagnetic Guided Modes. *Science* **340**, 328-330 (2013).
19. T. Kawalec, *et al.* Spectroscopic measurements of the evanescent wave polarization state. *Opt. Commun.* **274**, 341-346 (2007).
20. R. Mitsch, *et al.* Exploiting the local polarization of strongly confined light for sub-micrometer-resolution internal state preparation and manipulation of cold atoms. *Phys. Rev. A* **89**, 063829 (2014).
21. C. Junge, D. O'Shea, J. Volz, & A. Rauschenbeutel Strong Coupling between Single Atoms and Nontransversal Photons. *Phys. Rev. Lett.* **110**, 213604 (2013).
22. R. Mathevet & G. L. J. A. Rikken Magnetic circular dichroism as a local probe of the polarization of a focused Gaussian beam. *Opt. Mater. Express* **4**, 2574-2585 (2014).
23. C. Bustamante, I. Tinoco, & M. F. Maestre Circular differential scattering can be an important part of the circular dichroism of macromolecules. *Proc. Natl. Acad. Sci. USA* **80**, 3568-3572 (1983).
24. I. Tinoco & A. L. Williams Differential Absorption and Differential Scattering of Circularly Polarized-Light - Applications to Biological Macromolecules. *Annu. Rev. Phys. Chem.* **35**, 329-355 (1984).
25. J. Karst, *et al.* Chiral Scatterometry on Chemically Synthesized Single Plasmonic Nanoparticles. *ACS Nano* **13**, 8659-8668 (2019).
26. Q. F. Zhang, *et al.* Unraveling the origin of chirality from plasmonic nanoparticle-protein complexes. *Science* **365**, 1475-1478 (2019).
27. K. W. Smith, *et al.* Chiral and Achiral Nanodumbbell Dimers: The Effect of Geometry on Plasmonic Properties. *ACS Nano* **10**, 6180-6188 (2016).
28. B. Auguié, J. L. Alonso-Gómez, A. s. Guerrero-Martínez, & L. M. Liz-Marzán Fingers Crossed: Optical Activity of a Chiral Dimer of Plasmonic Nanorods. *J. Phys. Chem. Lett.* **2**, 846-851 (2011).
29. M. Born The Natural Optical Activity of Liquids and Gases. *Phys. Z.* **16**, 251-258 (1915).
30. W. Kuhn The physical significance of optical rotatory power. *Transactions of the Faraday Society* **26**, 293-308 (1930).
31. B. Foerster, *et al.* Particle Plasmons as Dipole Antennas: State Representation of Relative Observables. *J. Phys. Chem. C* **122**, 19116-19123 (2018).
32. E. Prodan, C. Radloff, N. J. Halas, & P. Nordlander A Hybridization Model for the Plasmon Response of Complex Nanostructures. *Science* **302**, 419-422 (2003).
33. X. Yin, M. Schäferling, B. Metzger, & H. Giessen Interpreting Chiral Nanophotonic Spectra: The Plasmonic Born-Kuhn Model. *Nano Lett.* **13**, 6238-6243 (2013).

34. L. Shao, *et al.* Angle- and Energy-Resolved Plasmon Coupling in Gold Nanorod Dimers. *ACS Nano* **4**, 3053-3062 (2010).
35. C. A. Thibodeaux, *et al.* Impurity-induced plasmon damping in individual cobalt-doped hollow Au nanoshells. *J. Phys. Chem. B* **118**, 14056-14061 (2014).
36. P. W. K. Rothemund Folding DNA to create nanoscale shapes and patterns. *Nature* **440**, 297-302 (2006).
37. A. Kuzyk, *et al.* DNA-based self-assembly of chiral plasmonic nanostructures with tailored optical response. *Nature* **483**, 311-314 (2012).
38. A. Kuzyk, *et al.* Reconfigurable 3D plasmonic metamolecules. *Nat. Mater.* **13**, 862-866 (2014).
39. S. Ganapathy, *et al.* Alternating syn-anti bacteriochlorophylls form concentric helical nanotubes in chlorosomes. *Proc. Natl. Acad. Sci. USA* **106**, 8525-8530 (2009).
40. T. Fujita, J. C. Brookes, S. K. Saikin, & A. Aspuru-Guzik Memory-Assisted Exciton Diffusion in the Chlorosome Light-Harvesting Antenna of Green Sulfur Bacteria. *J. Phys. Chem. Lett.* **3**, 2357-2361 (2012).
41. K. W. Smith, *et al.* Chiral and Achiral Nanodumbbell Dimers: The Effect of Geometry on Plasmonic Properties. *ACS Nano* **10**, 6180-6188 (2016).
42. X. Lan, *et al.* Bifacial DNA Origami-Directed Discrete, Three-Dimensional, Anisotropic Plasmonic Nanoarchitectures with Tailored Optical Chirality. *J. Am. Chem. Soc.* **135**, 11441-11444 (2013).
43. C. P. Byers, *et al.* Single-Particle Spectroscopy Reveals Heterogeneity in Electrochemical Tuning of the Localized Surface Plasmon. *J. Phys. Chem. B* **118**, 14047-14055 (2014).
44. P. B. Johnson & R. W. Christy Optical Constants of Noble Metals. *Phys. Rev. B* **6**, 4370-4379 (1972).
45. E. D. Palik (1998) *Handbook of optical constants of solids III* (Academic Press, San Diego).
46. T. A. F. Konig, *et al.* Electrically Tunable Plasmonic Behavior of Nanocube-Polymer Nanomaterials Induced by a Redox-Active Electrochromic Polymer. *ACS Nano* **8**, 6182-6192 (2014).

Figure Legends

Fig. 1. SAM and geometry drive distinct polarization-dependent light-matter interactions. (A) Circularly polarized light has a longitudinal SAM vector (S_{CPL}) that couples to electrons with co-aligned spins (S_{el}). (B) The Born-Kuhn model for CD of two charged masses (grey spheres) attached to two coupled, orthogonal springs displaced along a third orthogonal axis. (C) Trochoidal polarizations produce a transverse SAM vector (S_{EW}) that similarly couples to electrons with co-aligned spins. (D) A modified Born-Kuhn model for trochoidal dichroism where one oscillator is aligned along the direction of light propagation.

Fig. 2. Orthogonal and offset nanorods are a model system for observing trochoidal differential scattering. (A) Experimental TIR geometry. LP: linear polarizer, E_s , E_p : incident electric field components, θ : angle of incidence. (B) Isolated in-sample-plane components of the evanescent wave for $E_s = -E_p$ and $E_s = E_p$ incident polarizations, tracing CW and ACW trochoids, respectively. (C) Plasmon hybridization for the L-dimer. The light-induced electric dipoles are shown with black arrows and are excited opposite the electric field, with the curved arrow solely indicating the trochoidal polarization. Excitation from a CW (ACW) trochoidal field gives a high (low) energy antibonding (bonding) mode. In the corresponding spring system, attached masses are positively charged and are attracted toward the trochoidal electric field drawn with dashed lines and curved arrows. (D) Plasmon hybridization for the mirror-image T-dimer. Normalized measured and simulated scattering spectra of the (E) L-dimer and (F) T-dimer under CW and ACW trochoidal excitation. Insets: correlated scanning electron microscopy (SEM) images with scale bars of 100 nm and charge distributions calculated at the scattering maxima for CW and ACW trochoidal excitation, matching the black arrows in C and D.

Fig. 3. L-dimers have trochoidal and linear dichroism of opposite sign. (A) Plasmon hybridization for the L-dimer with trochoidal excitation resulting from TIR of $E_s = E_p$ (45°) and $E_s = -E_p$ (-45°). Excitation from a CW (ACW) trochoidal field gives a high (low) energy antibonding (bonding) mode. (B) Plasmon hybridization for the L-dimer with oblique $E_s = \pm E_p$ incident polarizations. $E_s = E_p$ ($E_s = -E_p$) linearly polarized light excites the high (low) energy antibonding (bonding) mode. Note that while magenta and blue arrows indicate the $\pm 45^\circ$ linear polarization of the incident light, the polarization once projected onto the sample plane is $\pm 70^\circ$ (Supplementary Text). (C) Normalized single-particle scattering spectra of an L-dimer under CW and ACW trochoidal excitation. Inset: correlated SEM image with a scale bar of 100 nm. (D)

Normalized scattering spectra of the same L-dimer under oblique incident excitation. Consistent results are observed also with normal incidence excitation (SI Appendix, Fig. S11). The increase in noise is due to the reduced sensitivity of oblique incidence scattering spectra (35). (E)

Summary of symmetry operations relating each geometric isomer to another and the effect on trochoidal and linear dichroism (TD and LD). Each mirror operation (σ_v) gives opposite linear dichroism, but mirroring over the (x, y) plane maintains trochoidal dichroism. Therefore, the two effects are consistently distinguishable across L-dimers and Γ -dimers (quadrant I and IV) (SI Appendix, Fig. S12). These isomers have reduced resonance shifts relative to those of \perp - and Γ -dimers (quadrants II and III) (SI Appendix, Fig. S12), likely resulting from competing contributions of both dichroisms. However, trochoidal dichroism dominates (SI Appendix, Fig. S6, S12).

Fig. 4. Observation of trochoidal differential scattering in self-assembled gold nanorod dimers (A) Schematic of self-assembled dimers utilizing DNA origami, forming an L-dimer. (B) Sample rotation of 180° forms a Γ -dimer, reversing the trochoidal dichroism and promoting opposite mode excitation. (C, E) Normalized scattering spectra of the L-dimers at 0° sample orientation with CW and ACW trochoidal excitation. SEM images depict dimer orientation relative to k . Scale bar: 50 nm. (D, F) Normalized scattering spectra of the same dimers after 180° rotation. Dashed and dotted lines identify bonding and antibonding modes respectively. As for the ideal dimers in Fig. 2 and 3, trochoidal polarization of the incident field allows for mode selective excitation of hybridized nanorod dimer plasmons.