



# Effects of body size on estimation of mammalian area requirements

Michael J. Noonan <sup>1,2\*</sup> Christen H. Fleming,<sup>1,2</sup> Marlee A. Tucker,<sup>3,4,5</sup> Roland Kays <sup>6,7</sup> Autumn-Lynn Harrison,<sup>8</sup> Margaret C. Crofoot,<sup>9,10</sup> Briana Abrahms,<sup>11</sup> Susan C. Alberts,<sup>12</sup> Abdullahi H. Ali,<sup>13</sup> Jeanne Altmann,<sup>14</sup> Pamela Castro Antunes,<sup>15</sup> Nina Attias,<sup>16</sup> Jerrold L. Belant,<sup>17</sup> Dean E. Beyer Jr.,<sup>18</sup> Laura R. Bidner,<sup>9,19</sup> Niels Blaum,<sup>20</sup> Randall B. Boone,<sup>21,22</sup> Damien Caillaud,<sup>9</sup> Rogerio Cunha de Paula,<sup>23</sup> J. Antonio de la Torre,<sup>24</sup> Jasja Dekker,<sup>25</sup> Christopher S. DePerno,<sup>7</sup> Mohammad Farhadinia <sup>26,27</sup> Julian Fennessy,<sup>28</sup> Claudia Fichtel,<sup>29</sup> Christina Fischer,<sup>30</sup> Adam Ford <sup>31</sup> Jacob R. Goheen <sup>32</sup> Rasmus W. Havmøller,<sup>9</sup> Ben T. Hirsch,<sup>33</sup> Cindy Hurtado <sup>34,35</sup> Lynne A. Isbell,<sup>9,19</sup> René Janssen,<sup>36</sup> Florian Jeltsch,<sup>20</sup> Petra Kaczensky,<sup>37,38</sup> Yayoi Kaneko,<sup>39</sup> Peter Kappeler,<sup>29</sup> Anjan Katna,<sup>40,41</sup> Matthew Kauffman,<sup>42</sup> Flavia Koch,<sup>29</sup> Abhijeet Kulkarni,<sup>40</sup> Scott LaPoint,<sup>43,44</sup> Peter Leimgruber,<sup>1</sup> David W. Macdonald,<sup>26</sup> A. Catherine Markham,<sup>45</sup> Laura McMahon,<sup>46</sup> Katherine Mertes,<sup>1</sup> Christopher E. Moorman,<sup>7</sup> Ronaldo G. Morato,<sup>23,47</sup> Alexander M. Moßbrucker,<sup>48</sup> Guilherme Mourão,<sup>49</sup> David O'Connor,<sup>4,50,51</sup> Luiz Gustavo R. Oliveira-Santos,<sup>52</sup> Jennifer Pastorini,<sup>53,54</sup> Bruce D. Patterson,<sup>55</sup> Janet Rachlow,<sup>56</sup> Dustin H. Ranglack,<sup>57</sup> Neil Reid,<sup>58</sup> David M. Scantlebury,<sup>59</sup> Dawn M. Scott,<sup>60</sup> Nuria Selva,<sup>61</sup> Agnieszka Sergiel,<sup>61</sup> Melissa Songer,<sup>1</sup> Nucharin Songsasen,<sup>1</sup> Jared A. Stabach,<sup>1</sup> Jenna Stacy-Dawes,<sup>50</sup> Morgan B. Swingen,<sup>7,62</sup> Jeffrey J. Thompson,<sup>63,64</sup> Wiebke Ullmann,<sup>20</sup> Abi Tamim Vanak,<sup>40,65,66</sup> Maria Thaker,<sup>67</sup> John W. Wilson,<sup>68</sup> Koji Yamazaki,<sup>69,70</sup> Richard W. Yarnell,<sup>71</sup> Filip Zieba,<sup>72</sup> Tomasz Zwijacz-Kozica,<sup>72</sup> William F. Fagan,<sup>2</sup> Thomas Mueller,<sup>3,4</sup> and Justin M. Calabrese<sup>1,2</sup>

<sup>1</sup>Smithsonian Conservation Biology Institute, National Zoological Park, 1500 Remount Road, Front Royal, VA 22630, U.S.A.

<sup>2</sup>Department of Biology, University of Maryland, College Park, MD 20742, U.S.A.

<sup>3</sup>Senckenberg Biodiversity and Climate Research Centre, Senckenberg Gesellschaft für Naturforschung, Senckenberganlage 25, Frankfurt (Main), 60325, Germany

<sup>4</sup>Department of Biological Sciences, Goethe University, Max-von-Laue-Straße 9, Frankfurt (Main), 60438, Germany

<sup>5</sup>Department of Environmental Science, Institute for Wetland and Water Research/Radboud University, P.O. Box 9010, Nijmegen, GL NL-6500, The Netherlands

<sup>6</sup>North Carolina Museum of Natural Sciences, Biodiversity Lab, Raleigh, NC 27601, U.S.A.

<sup>7</sup>Fisheries, Wildlife, and Conservation Biology Program, College of Natural Resources Campus Box 8001, North Carolina State University, Raleigh, NC 27695, U.S.A.

<sup>8</sup>Migratory Bird Center, Smithsonian Conservation Biology Institute, Washington, D.C., 20013, U.S.A.

<sup>9</sup>Department of Anthropology, University of California, Davis, Davis, CA, 95616, U.S.A.

<sup>10</sup>Smithsonian Tropical Research Institute, Balboa Ancon, 0843-03092, Republic of Panama

<sup>11</sup>Environmental Research Division, NOAA Southwest Fisheries Science Center, Monterey, CA 93940, U.S.A.

<sup>12</sup>Departments of Biology and Evolutionary Anthropology, Duke University, Durham, NC 27708, U.S.A.

<sup>13</sup>Hirola Conservation Programme, Garissa, 1774-70100, Kenya

<sup>14</sup>Department of Ecology and Evolution, Princeton University, 106A Guyot Hall, Princeton, NJ 08544, U.S.A.

<sup>15</sup>Department of Ecology, Federal University of Mato Grosso do Sul, Campo Grande, MS 79070-900, Brazil

\*Address correspondence to Michael J. Noonan, email [noonanm@si.edu](mailto:noonanm@si.edu)

**Article impact statement:** Due to autocorrelation-induced bias, conventional methods severely underestimate the area requirements of GPS-tracked large mammals.

Paper submitted September 9, 2019; revised manuscript accepted December 24, 2019.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

<sup>16</sup>Programa de Pós-Graduação em Biologia Animal, Universidade Federal do Mato Grosso do Sul, Cidade Universitária, Av. Costa e Silva, Campo Grande, Mato Grosso do Sul, 79070-900, Brazil

<sup>17</sup>Camp Fire Program in Wildlife Conservation, State University of New York, College of Environmental Science and Forestry, Syracuse, NY 13210, U.S.A.

<sup>18</sup>Michigan Department of Natural Resources, 1990 U.S. 41 South, Marquette, MI 49855, U.S.A.

<sup>19</sup>Mpala Research Centre, Nanyuki, 555-104000, Kenya

<sup>20</sup>University of Potsdam, Plant Ecology and Nature Conservation, Am Mühlenberg 3, Potsdam, 14476, Germany

<sup>21</sup>Natural Resource Ecology Laboratory, Colorado State University, Fort Collins, CO, 80523, U.S.A.

<sup>22</sup>Department of Ecosystem Science and Sustainability, Colorado State University, Fort Collins, CO, 80523, U.S.A.

<sup>23</sup>National Research Center for Carnivores Conservation, Chico Mendes Institute for the Conservation of Biodiversity, Estrada Municipal Hisaichi Takebayashi 8600, Atibaia, SP 12952-011, Brazil

<sup>24</sup>Instituto de Ecología, Universidad Nacional Autónoma de Mexico and CONACyT, Ciudad Universitaria, Mexico, D.F. 04318, Mexico

<sup>25</sup>Jasja Dekker Dierecologie, Enkhuizenstraat 26, Arnhem, WZ 6843, The Netherlands

<sup>26</sup>Wildlife Conservation Research Unit, Department of Zoology, University of Oxford, Tubney House, Oxfordshire, Oxford, OX13 5QL, U.K.

<sup>27</sup>Future4Leopards Foundation, Tehran, Iran

<sup>28</sup>Giraffe Conservation Foundation, PO Box 86099, Windhoek, Namibia

<sup>29</sup>German Primate Center, Behavioral Ecology & Sociobiology Unit, Kellnerweg 4, Göttingen, 37077, Germany

<sup>30</sup>Restoration Ecology, Department of Ecology and Ecosystem Management, Technische Universität München, Emil-Ramann-Straße 6, Freising, 85354, Germany

<sup>31</sup>The Irving K. Barber School of Arts and Sciences, Unit 2: Biology, The University of British Columbia, Okanagan Campus, SCI 109, 1177 Research Road, Kelowna, BC V1V 1V7, Canada

<sup>32</sup>Department of Zoology and Physiology, University of Wyoming, Laramie, WY 82071, U.S.A.

<sup>33</sup>Zoology and Ecology, College of Science and Engineering, James Cook University, Townsville, QLD 4811, Australia

<sup>34</sup>Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, Lima, 15072, Peru

<sup>35</sup>Department of Forest Resources Management, The University of British Columbia, Vancouver, BC V6T 1Z4, Canada

<sup>36</sup>Bionet Natuuronderzoek, Valderstraat 39, Stein, 6171EL, The Netherlands

<sup>37</sup>Norwegian Institute for Nature Research — NINA, Sluppen, Trondheim, NO-7485, Norway

<sup>38</sup>Research Institute of Wildlife Ecology, University of Veterinary Medicine, Savoyenstraße 1, Vienna, A-1160, Austria

<sup>39</sup>Tokyo University of Agriculture and Technology, Tokyo, 183-8509, Japan

<sup>40</sup>Ashoka Trust for Research in Ecology and the Environment (ATREE), Bangalore, Karnataka 560064, India

<sup>41</sup>Manipal Academy of Higher Education, Manipal, Karnataka 576104, India

<sup>42</sup>U.S. Geological Survey, Wyoming Cooperative Fish and Wildlife Research Unit, Department of Zoology and Physiology, University of Wyoming, Laramie, WY, 82071, U.S.A.

<sup>43</sup>Max Planck Institute for Ornithology, Vogelwarte Radolfzell, Am Obstberg 1, Radolfzell, D-78315, Germany

<sup>44</sup>Black Rock Forest, 65 Reservoir Road, Cornwall, NY 12518, U.S.A.

<sup>45</sup>Department of Anthropology, Stony Brook University, Stony Brook, NY 11794, U.S.A.

<sup>46</sup>Office of Applied Science, Department of Natural Resources, Rhinelander, WI 54501, U.S.A.

<sup>47</sup>Institute for the Conservation of Neotropical Carnivores – Pró-Carnívoros, Atibaia, São Paulo 12945-010, Brazil

<sup>48</sup>Frankfurt Zoological Society, Bernhard-Grzimek-Allee 1, Frankfurt, 60316, Germany

<sup>49</sup>Embrapa Pantanal, Rua 21 de setembro 1880, Corumbá, MS 79320-900, Brazil

<sup>50</sup>San Diego Zoo Institute of Conservation Research, 15600 San Pasqual Valley Road, Escondido, CA 92027, U.S.A.

<sup>51</sup>National Geographic Partners, 1145 17th Street NW, Washington, D.C. 20036, U.S.A.

<sup>52</sup>Department of Ecology, Federal University of Mato Grosso do Sul, Campo Grande, MS 79070-900, Brazil

<sup>53</sup>Centre for Conservation and Research, 26/7 C2 Road, Kodighawewa, Julpallama, Tissamaharama, 82600, Sri Lanka

<sup>54</sup>Anthropologisches Institut, Universität Zürich, Winterthurerstrasse 190, Zurich, 8057, Switzerland

<sup>55</sup>Integrative Research Center, Field Museum of Natural History, Chicago, IL 60605, U.S.A.

<sup>56</sup>Department of Fish and Wildlife Sciences, University of Idaho, 875 Perimeter Drive MS 1136, Moscow, ID 83844-1136, U.S.A.

<sup>57</sup>Department of Biology, University of Nebraska at Kearney, Kearney, NE 68849, U.S.A.

<sup>58</sup>Institute for Global Food Security (IGFS), School of Biological Sciences, Queen's University Belfast, Belfast, BT9 5DL, U.K.

<sup>59</sup>School of Biological Sciences, Queen's University Belfast, 19 Chlorine Gardens, Belfast, Northern Ireland BT9 5DL, U.K.

<sup>60</sup>School of Life Sciences, Keele University, Keele, Staffordshire ST5 5BG, U.K.

<sup>61</sup>Institute of Nature Conservation, Polish Academy of Sciences, Mickiewicza 33, Krakow, 31-120, Poland

<sup>62</sup>1854 Treaty Authority, 4428 Haines Road, Duluth, MN 55811, U.S.A.

<sup>63</sup>Asociación Guyra Paraguay – CONACYT, Parque Ecológico Asunción Verde, Asuncion, 1101, Paraguay

<sup>64</sup>Instituto Saite, Coronel Felix Cabrera 166, Asuncion, 1101, Paraguay

<sup>65</sup>Wellcome Trust/DBT India Alliance, Hyderabad, 500034, India

<sup>66</sup>School of Life Sciences, University of KwaZulu-Natal, Westville, Durban 4041, South Africa

<sup>67</sup>Centre for Ecological Sciences, Indian Institute of Science, Bangalore, 560012, India

<sup>68</sup>Department of Zoology & Entomology, University of Pretoria, Pretoria, 0002, South Africa

<sup>69</sup>Ibaraki Nature Museum, Zoological Laboratory, 700 Osaki, Bando-city, Ibaraki 306-0622, Japan

<sup>70</sup>Forest Ecology Laboratory, Department of Forest Science, Tokyo University of Agriculture, 1-1-1 Sakuragaoka, Setagaya-Ku, Tokyo 156-8502, Japan

<sup>71</sup>School of Animal, Rural and Environmental Sciences, Nottingham Trent University, Brackenhurst Campus, Southwell, NG25 0QF, U.K.

<sup>72</sup>Tatra National Park, Kúznice 1, Zakopane, 34-500, Poland

**Abstract:** Accurately quantifying species' area requirements is a prerequisite for effective area-based conservation. This typically involves collecting tracking data on species of interest and then conducting home-range analyses. Problematically, autocorrelation in tracking data can result in space needs being severely underestimated. Based on the previous work, we hypothesized the magnitude of underestimation varies with body mass, a relationship that could have serious conservation implications. To evaluate this hypothesis for terrestrial mammals, we estimated home-range areas with global positioning system (GPS) locations from 757 individuals across 61 globally distributed mammalian species with body masses ranging from 0.4 to 4000 kg. We then applied block cross-validation to quantify bias in empirical home-range estimates. Area requirements of mammals <10 kg were underestimated by a mean approximately 15%, and species weighing approximately 100 kg were underestimated by approximately 50% on average. Thus, we found area estimation was subject to autocorrelation-induced bias that was worse for large species. Combined with the fact that extinction risk increases as body mass increases, the allometric scaling of bias we observed suggests the most threatened species are also likely to be those with the least accurate home-range estimates. As a correction, we tested whether data thinning or autocorrelation-informed home-range estimation minimized the scaling effect of autocorrelation on area estimates. Data thinning required an approximately 93% data loss to achieve statistical independence with 95% confidence and was, therefore, not a viable solution. In contrast, autocorrelation-informed home-range estimation resulted in consistently accurate estimates irrespective of mass. When relating body mass to home range size, we detected that correcting for autocorrelation resulted in a scaling exponent significantly >1, meaning the scaling of the relationship changed substantially at the upper end of the mass spectrum.

**Keywords:** allometry, animal movement, area-based conservation, autocorrelation, home range, kernel density estimation, reserve design, scaling

Efectos del Tamaño Corporal sobre la Estimación de los Requerimientos de Área de Mamíferos

**Resumen:** La cuantificación precisa de los requerimientos de área de una especie es un prerequisito para que la conservación basada en áreas sea efectiva. Esto comúnmente implica la recolección de datos de rastreo de la especie de interés para después realizar análisis de la distribución local. De manera problemática, la autocorrelación en los datos de rastreo puede resultar en una subestimación grave de las necesidades de espacio. Con base en trabajos previos, formulamos una hipótesis en la que supusimos que la magnitud de la subestimación varía con la masa corporal, una relación que podría tener implicaciones serias para la conservación. Para probar esta hipótesis en mamíferos terrestres, estimamos las áreas de distribución local con las ubicaciones en GPS de 757 individuos de 61 especies de mamíferos distribuidas mundialmente con una masa corporal entre 0.4 y 4,000 kg. Después aplicamos una validación cruzada en bloque para cuantificar el sesgo en estimaciones empíricas de la distribución local. Los requerimientos de área de los mamíferos <10 kg fueron subestimados por una media ~15% y las especies con una masa ~100 kg fueron subestimadas en ~50% en promedio. Por lo tanto, encontramos que la estimación del área estaba sujeta al sesgo inducido por la autocorrelación, el cual era peor para las especies de talla grande. En combinación con el hecho de que el riesgo de extinción incrementa conforme aumenta la masa corporal, el escalamiento alométrico del sesgo que observamos sugiere que la mayoría de las especies amenazadas también tienen la probabilidad de ser aquellas especies con las estimaciones de distribución local menos acertadas. Como corrección, probamos si la reducción de datos o la estimación de la distribución local informada por la autocorrelación minimizan el efecto de escalamiento que tiene la autocorrelación sobre las estimaciones de área. La reducción de datos requirió una pérdida de datos del ~93% para lograr la independencia estadística con un 95% de confianza y por lo tanto no fue una solución viable. Al contrario, la estimación de la distribución local informada por la autocorrelación resultó en estimaciones constantemente precisas sin importar la masa corporal. Cuando relacionamos la masa corporal con el tamaño de la distribución local, detectamos que la corrección de la autocorrelación resultó en un exponente de escalamiento significativamente >1, lo que significa que el escalamiento de la relación cambió sustancialmente en el extremo superior del espectro de la masa corporal.

**Palabras Clave:** alometría, autocorrelación, conservación basada en áreas, diseño de reserva, distribución local, escalamiento, estimación de densidad del núcleo, movimiento de mamíferos

**摘要:** 准确量化物种的栖息面积需求是开展有效区域保护的先决条件, 这一般包括收集目标物种的跟踪数据并进行家域分析。然而, 跟踪数据中的自相关性可能导致严重低估动物的空间需求。基于已有研究, 我们假设低估程度会随体重变化而变化, 这种关系可能对保护产生严重影响。为了评估陆生哺乳动物是否符合这一假设,

我们利用体重在 0.4 到 4000 千克之间的 61 种全球分布哺乳动物的 757 个个体的全球定位系统 (GPS) 定位数据, 估计了它们的家域范围。接下来, 我们应用分块交叉验证定量分析了家域经验估计的偏差。结果显示, 体重小于 10 千克的哺乳动物的面积需求平均被低估了约 15%, 体重在 100 千克左右的物种平均被低估约 50%。因此, 我们指出面积估计受到自相关引起偏差的影响, 且这种偏差对大型物种来说更为严重。考虑到物种灭绝风险随体重增加而增加这一事实, 我们观察到的偏差异速增长标度表明, 对最受威胁物种的家域估计可能也最不准确。为了修正这一偏差, 我们测试了数据抽稀和考虑自相关的家域估计是否能最大程度减少自相关对面积估计的尺度效应。数据抽稀需要丢弃约 93% 的数据才能达到 95% 置信的统计独立性, 因此该方法不可行。比较起来, 考虑自相关的家域估计始终可以获得准确估计, 而与动物体型无关。当将体重与家域大小联系起来时, 我们发现校正自相关会导致标度指数显著大于 1, 这意味着相关关系的标度在体型最大的物种中发生了显著的变化。【翻译: 胡怡思; 审校: 聂永刚】

**关键词:** 异速增长, 动物移动, 区域保护, 自相关, 家域, 核密度估计, 保护区设计, 标度

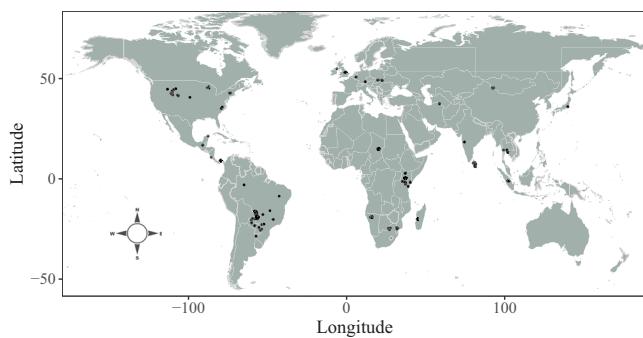
## Introduction

Globally, human-altered landscapes are restricting animal movement (Fahrig 2007; Tucker et al. 2018), and habitat loss and fragmentation are the principal threats to terrestrial biodiversity (Brooks et al. 2002; Wilson et al. 2016). A key component to conserving species in increasingly human-dominated landscapes is understanding how much space is required to maintain stable, interconnected populations (Brashares et al. 2001; Pe'er et al. 2014). Area requirements are typically quantified via home-range analysis (Burt 1943). This routinely involves collecting tracking data on species of interest (Kays et al. 2015) and then applying a home-range estimator to these data (Fleming et al. 2015; Noonan et al. 2019). These range estimates can then be used to inform recommendations on reserve sizes (Linnell et al. 2001), to advocate for specific land-tenure systems (Johansson et al. 2016; Farhadinia et al. 2018), and to make conservation policy recommendations (Barton et al. 2019). However, tracking data are often strongly autocorrelated, whereas conventional home-range estimators are based on the assumption of independent and identically distributed data (Noonan et al. 2019).

When data are autocorrelated, the total number of data points does not reflect the total amount of information in the data set (i.e., effective sample size) (Fleming & Calabrese 2017). Although the idea that autocorrelation may affect home-range estimates is not new (e.g., Swihart & Slade 1985; Fieberg 2007; Fleming et al. 2015), only recent analyses have demonstrated the seriousness of the problem. Using the largest empirical tracking data set assembled to date, Noonan et al. (2019) found conventional estimators significantly negatively biased when used on autocorrelated data. Although any form of bias is undesirable, the systematic underestimation of home-range areas is a worst-case scenario from a conservation perspective. Any policy or management decisions informed by underestimated home-range estimates could result in failed conservation initiatives (Brashares et al. 2001; Gaston et al. 2008) or exacerbate negative human-wildlife interactions at reserve boundaries (Van Eeden et al. 2018).

Noonan et al. (2019) noticed that large-bodied species tended to exhibit more negatively biased conventional home-range estimates than small-bodied species. However, the species included in their study were not selected to provide the broad range of body masses required to investigate allometric trends. We compiled an extensive empirical data set of global positioning system (GPS) locations from 757 individuals across 61 terrestrial mammalian species with body masses ranging from 0.4 to 4000 kg. We used these data to investigate whether the underestimation of home-range size scales with body mass. To see the potential for this, consider that large species have large home ranges (Jetz et al. 2004) that tend to take longer to cross than smaller home ranges (Calder 1983). In addition, range crossing time ( $\tau_p$ ) interacts with the sampling interval ( $dt$ ) in determining the amount of autocorrelation in tracking data (Fleming & Calabrese 2017; Noonan et al. 2019). When  $dt \lesssim \tau_p$ , the resulting data are autocorrelated, whereas  $dt \gg \tau_p$  results in effectively independent data. Finally, the magnitude of the negative biases in conventional home-range estimates increases in proportion to the strength of autocorrelation in the data (Noonan et al. 2019). Combining these facts, we arrived at the hypothesis that an allometry in  $\tau_p$  drives autocorrelation and negative estimation bias to scale with body size.

We examined this hypothesis in 2 ways. First, we tested whether the chain of relationships that would drive bias to scale with mass holds for empirical tracking data. Second, we explored how well 2 methods of home-range estimation for autocorrelated data eliminate the scaling of home-range estimation bias. These methods were model-informed data thinning, which removes autocorrelation from the data prior to home-range estimation, and autocorrelation-informed home-range estimation, which statistically accounts for autocorrelation in movement data. We then used model selection to determine whether significant allometry bias remains in the data for each approach and identified whether one of these corrections offers improved performance over the other. Finally, in light of our findings, we revisited



**Figure 1.** Distribution of study sites for the empirical global positioning system tracking data set spanning 757 individuals across 61 mammalian species.

the concept of home-range allometry (e.g., McNab 1963; Jetz et al. 2004; Tucker et al. 2014). Mammalian home-range area ( $H$ ) scales positively with body mass ( $M$ ) as  $H = B_0 M^b$ , where  $B_0$  is a normalization constant and  $b$  is the scaling exponent (McNab 1963). Despite decades of research, however, there has been little consensus on whether the allometry is linear (i.e.,  $M^1$ ), or super-linear (i.e.,  $M^{>1}$ ). Historically, this scaling relationship has been calculated by compiling home-range areas estimated via conventional estimators, which are subject to varying levels of autocorrelation-induced bias (Noonan et al. 2019), whereas no one has assessed this relationship directly from tracking data. Although consistent bias across the mass spectrum would lead only to a change in the normalization constant, differential bias across the mass spectrum could alter the scaling exponent, fundamentally changing the properties of the relationship. As such, we tested for any significant deviations from linear ( $M^1$ ) scaling.

## Methods

All analyses were based on precollected tracking data sets obtained under appropriate permits and that were based on Institutional Animal Care and Use Committee approved protocols.

### Data Compilation

To investigate whether biases in home-range estimation scale with body size, we compiled GPS tracking data for 61 globally distributed terrestrial mammalian species, comprising  $6.94 \times 10^6$  locations for 757 individuals collected from 2000 to 2019 (Fig. 1). Individual data sets were selected based on the criterion of range resident behavior (i.e., area-restricted space use), as evidenced by plots of the semivariance in positions as a function of the time lag separating observations (i.e., variograms) with a clear asymptote at large lags (Calabrese et al.

2016). When data do not indicate evidence of range residency, home-range estimation is not appropriate (Calabrese et al. 2016; Fleming & Calabrese 2017), so we excluded data from migratory or nonrange resident individuals. The visual verification of range residency via variogram analysis was conducted using the R package ctmm (version 0.5.3) (Calabrese et al. 2016). Further details on these data are given in Supporting Information.

For each of the species in our data set, we compiled covariate data on that species' mean adult mass in kilograms. We also identified the main food source for each species and classified them as carnivorous or omnivorous or frugivorous or herbivorous. Data from these 2 dietary classes were analyzed separately. Mass and dietary data were from the EltonTraits database (Wilman et al. 2014).

### Tracking-Data Analyses

Our conjecture that the underestimation of home-range areas increases as body size increases was based on 2 well-established biological and one methodological relationship: the positive correlation between body mass and home-range area (Jetz et al. 2004); the positive correlation between home-range area and range crossing time,  $\tau_p$  (Calder 1983); and the negative correlation between range crossing time and the effective sample size for area estimation,  $N_{\text{area}}$  (i.e., equivalent number of statistically independent locations [Noonan et al. 2019]). We hypothesized that these conspire to drive 2 previously untested relationships: a potential negative correlation between body mass and  $N_{\text{area}}$  and a potential negative correlation between body mass and home-range estimator accuracy.

Testing for these relationships first required estimating the autocorrelation structure in each of the individual tracking data sets. To accomplish this, we fitted a series of range-resident, continuous-time movement models to the data with the estimation methods developed by Fleming et al. (2019). The fitted models included the independent and identically distributed process, which features uncorrelated positions and velocities; the Ornstein-Uhlenbeck (OU) process, which features correlated positions but uncorrelated velocities (Uhlenbeck & Ornstein 1930); and an OU-foraging (OUF) process, featuring both correlated positions and velocities (Fleming et al. 2014). We used model selection to identify the best fitting model given the data (Fleming et al. 2014) from which  $\tau_p$  and  $N_{\text{area}}$  were extracted. To fit and select the movement models, we used the R package ctmm and applied the workflow described by Calabrese et al. (2016).

We estimated home-range areas for each of the 757 individuals in our tracking database via kernel density estimation (KDE) with Gaussian reference function bandwidth optimization because this is one of the most commonly applied home-range estimators in ecological research (Noonan et al. 2019). The KDE home ranges were estimated via the methods implemented in ctmm, and

the further small-sample-size bias correction that was introduced in area-corrected KDE (Fleming & Calabrese 2017).

Our primary aim was to determine the extent to which autocorrelation-induced bias in conventional home-range estimation might increase with body size. This required an objective and statistically sound measure of bias. We applied the well-established technique of block cross-validation (Noonan et al. 2019) to quantify bias in empirical home-range estimates.

By determining the extent to which the results of an analysis generalize to a statistically independent data set, cross-validation is an effective tool for quantifying bias (Pawitan 2001). For this approach, each individual data set was split in half, and a home-range area was estimated from the first half of the data only (i.e., training set). Next, the percentage of observations in the second half of the data (i.e., held-out set) that fell within the specified contour (here 50% and 95%) of the estimated home range was calculated. If the percentage of points included came out consistently higher or lower than the specified contour, then it would suggest positive or negative bias, respectively.

As a further measure of bias, we identified the contour of the home range estimated from the training set that contained the desired percentage of locations in the held-out set (i.e., 50% and 95%) and compared the area within that contour to the estimated area at the specified quantile. For example, consider that the 95% area estimated on the training data contained only 90% of the locations in the held-out set, whereas the 97% contour contained 95% of the locations. To measure bias, we would take the ratio between the 97% area and the 95% area. Cross-validating home-range estimates in this way can also be seen as providing a measure of how well a home-range estimate can be expected to capture an animal's future space use, assuming no substantial changes in movement behavior.

Block cross-validation is based on the assumption that data from the training and held-out sets are generated from the same processes. To confirm this assumption, we used the Battacharryya distance implementation in ctmm (Winner et al. 2018) as a measure of similarity (range 0– $\infty$ ) between the mean area and covariance parameters of movement models fitted to the training and held-out data sets and determined whether the confidence intervals on this distance contained 0 (details are given in Appendix S1 in Noonan et al. [2019]). Using this method, we determined that 160 of 757 individuals had movement models with significantly different parameter estimates between the first and second halves of the data, so we excluded these from our cross-validation analyses. We found no significant relationship between whether or not a data set was excluded from our analyses and which species the data were from ( $p = 0.52$ ) or between exclusion and how long an individual was tracked ( $p = 0.39$ ). This

confirmed that the subsampling required to meet the assumptions of half-sample cross-validation did not bias our sample.

### Correction Factors

We explored 2 potential solutions to the allometric scaling of autocorrelation and home-range estimation bias: thinning data to minimize autocorrelation and using autocorrelation-informed home-range estimation.

Conventional kernel methods are based on an assumption of independence; however, they can provide accurate estimates for autocorrelated processes when the sampling is coarse enough that the data appear uncorrelated over time (Hall & Hart 1990). Thus, data thinning presents a potentially straightforward solution to autocorrelation-induced bias, but requires a balance between reducing autocorrelation and retaining sample size. We, therefore, explored model-informed data thinning as a means of mitigating size-dependent home-range bias. As noted above, the parameter  $\tau_p$  relates to an individual's range-crossing time and quantifies the time scale over which positional autocorrelation decays to insignificance. More specifically, because positional autocorrelation decays exponentially at rate  $1/\tau_p$ , the time required for the percentage of the original velocity autocorrelation to decay to  $\alpha$  is  $\tau_\alpha = \tau_p \ln(1/\alpha)$ . Conventionally, data are thinned to independence with a 95% level of confidence, and approximately  $3\tau_p$  is the time it takes for 95% of the positional autocorrelation to decay. Consequently, we thinned each individual's tracking data to a sampling frequency of  $dt = 3\tau_p$ . We then used autocorrelation functions to quantify how much autocorrelation remained in the thinned data and evaluated the performance of KDEs on these thinned data.

As opposed to manipulating the data to meet the assumptions of the estimator, the second potential solution was to use an estimator that explicitly modeled the autocorrelation in the data. Autocorrelated-KDE (AKDE) is a generalization of Gaussian reference function KDE that conditions upon the autocorrelation structure of the data when optimizing the bandwidth (Fleming et al. 2015). Following the workflow described by Calabrese et al. (2016), AKDE home-range areas were estimated conditioned on the selected movement model for each data set, via the methods implemented in ctmm, with the same small-sample-size bias correction applied to the conventional KDE area estimates (Fleming & Calabrese 2017). The AKDE is available via the web-based graphical user interface at [ctmm.shinyapps.io/ctmmweb/](http://ctmm.shinyapps.io/ctmmweb/) (Dong et al. 2017).

### Correction Factor Performance

To test for body-size-dependent biases in cross-validation success, we fitted 3 regression models to the

cross-validation results as a function of  $\log_{10}$ -scaled mass. The models included an intercept-only model (i.e., no change in bias with mass), linear model, and logistic model. We then identified the best model for the data via small-sample-size corrected quasi-Akaike information criterion (Burnham et al. 2011).

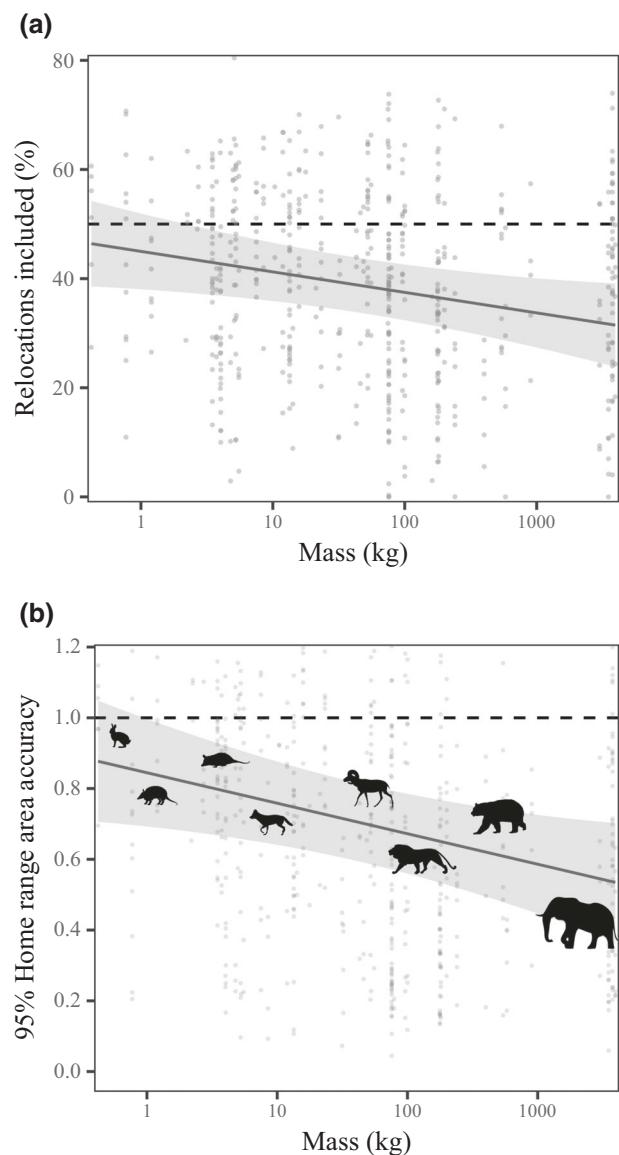
Species may exhibit similarities in traits due to phylogenetic inertia and the constraints of common ancestry; thus, controlled comparisons are required (Harvey & Pagel 1991). Accordingly, we did not treat species data records as independent; rather, we used the phylogenetic distances among species to construct a variance-covariance matrix and defined the correlation structure in our allometric regressions with the R package nlme (version 3.1-137) (Pinheiro et al. 2018). Phylogenetic relationships between eutherian mammalian orders were based on genetic differences and taken from Liu et al. (2001). Intraorder relationships were taken from more targeted studies aimed at resolving species-level relationships, including Price et al. (2005) for Artiodactyla, Matthee et al. (2004) for Lagomorpha, Steiner and Ryder (2011) for Perissodactyla, Barriel et al. (1999) for Proboscidea, Perelman et al. (2011) for Primates, and Agnarsdóttir et al. (2010) for Carnivora. For Canidae, however, we took relationships from Lindblad-Toh et al. (2005), due to better coverage of the species in our data set. The phylogenetic tree was built with the R package ape (version 5.2) (Paradis & Schliep 2019), and branch lengths were computed following Grafen (1989). Phylogenies are given in Supporting Information.

## Results

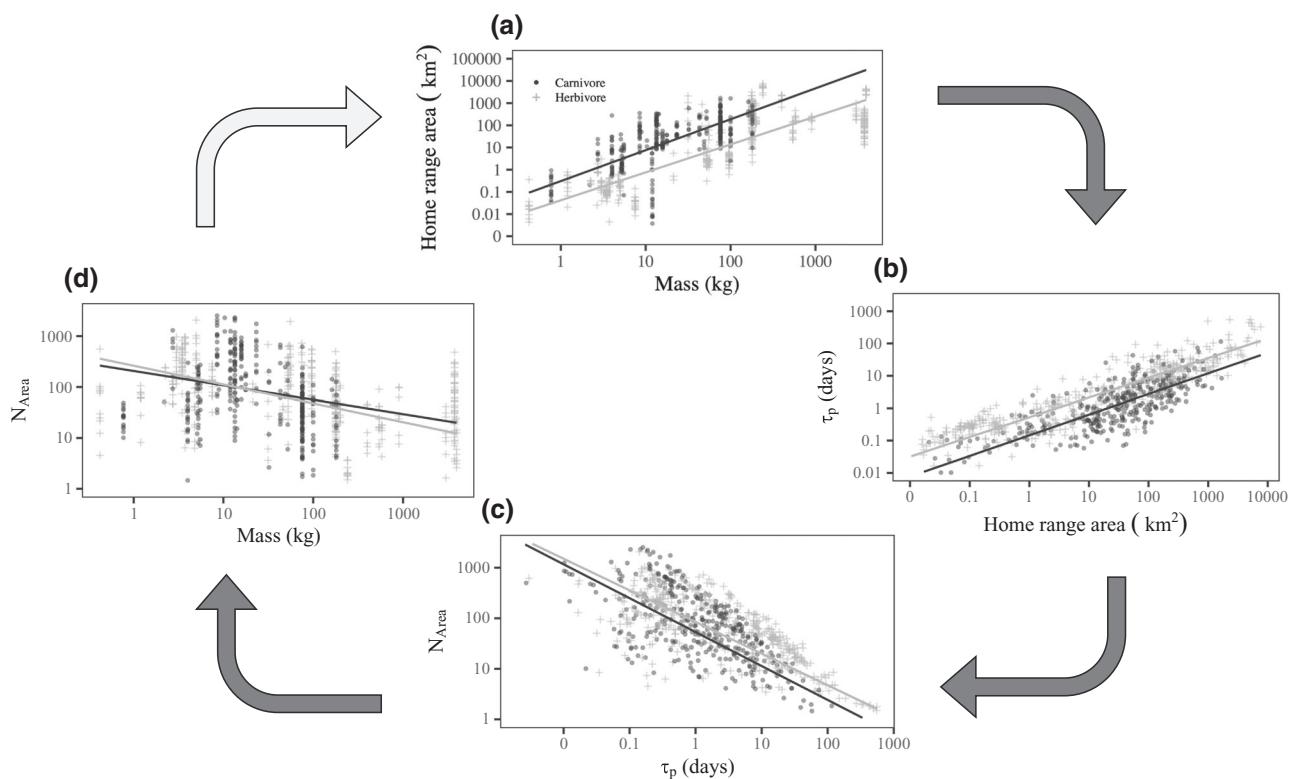
### Allometric Scaling of Bias

Out of 757 data sets, only one was independent and identically distributed and free from significant autocorrelation. Conventional KDE 95% home-range areas cross-validated at a median rate of 88.3% (95% CI 87.2–90.1), which was below the target 95% quantile and demonstrated a tendency to underestimate home-range areas on average. Similarly, KDE 50% home-range areas cross-validated at a median rate of 41.5% (95% CI 39.4–43.3), which was again below the target 50% quantile. The magnitude of KDE's underestimation worsened as body mass increased ( $t = 2.30, p = 0.02$ ) (Fig. 2a), carnivores and herbivores did not differ significantly ( $t = 0.31; p = 0.75$ ). Cross-validation success of 50% home-range areas across the mass spectrum was best described by a linear decay model with an intercept of 47.2 (95% CI 39.9–54.5) and a slope of -3.9 (95% CI -7.0 to -0.8). In other words, for every order of magnitude increase in body mass, home-range estimates captured approximately 4% less of an individual's future space use.

When comparing the 95% area estimates with the area estimates for the contours that contained 95% of



**Figure 2.** Cross-validation of conventional kernel density estimation (KDE) across the mammalian body-mass spectrum: (a) percentage of locations from the second half of the data (held-out set) included in KDE 50% home ranges estimated from the first half of the data (training set) as a function of body mass (dashed line, target 50% quantile; solid line, phylogenetically controlled regression model fit to cross-validation results; shading, 95% CI of the fit) and (b) regression model describing the accuracy of 95% KDE area estimates across the mass spectrum. Accuracy was quantified as the ratio between estimated 95% area of the training set and the area contained within the contour that encompassed 95% of locations in the held-out set. The horizontal dashed line represents an unbiased area estimate. The x-axes in are log scaled.



**Figure 3.** Mechanisms driving body-size-dependent estimation bias: (a) positive allometry of home-range areas, (b) correlation between home-range area and range-crossing time ( $\tau_p$ ), (c) negative correlation between  $\tau_p$  and effective sample size ( $N_{\text{area}}$ ) governed by duration of observation period ( $T$ ) and  $\tau_p$  such that  $N_{\text{area}} \approx T/\tau_p$ , and (d) resulting negative allometry of  $N_{\text{area}}$  (axes, log scaled; lines, phylogenetically controlled fitted regression models). From (a) to (d), 1 axis is preserved from the previous panel to demonstrate the inherent link between each of these relationships (arrows, visual aid of link; top-left arrow, end of the chain).

locations, KDE accuracy across the mass spectrum was best described by linear decay (Fig. 2b). Consequently, whereas the home-range areas of mammals weighing  $<10$  kg were underestimated by 13.6% (95% CI 6.3–18.6), those of species weighing  $>100$  kg were underestimated by 46.0% on average (95% CI 36.7–51.4).

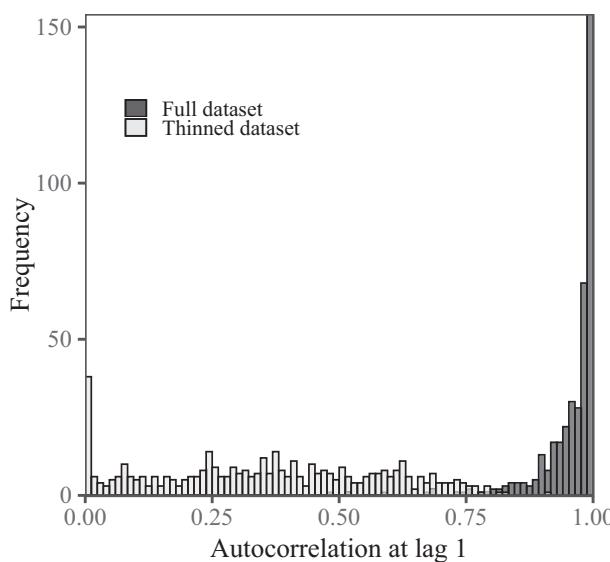
#### Mechanisms Driving Body Size-Dependent Estimation Bias

We found significant positive relationships between body mass and home-range area (regression parameter:  $\beta = 1.18$ , 95% CI 0.92–1.43,  $t = 9.09$ ,  $p < 0.0001$ ) (Fig. 3a) and between home-range area and range crossing time,  $\tau_p$  ( $\beta = 7.09$ , 95% CI 4.78–9.41,  $t = 6.00$ ,  $p < 0.0001$ ) (Fig. 3b) and a negative relationship between  $\tau_p$  and the effective sample size,  $N_{\text{area}}$  ( $\beta = -0.65$ , 95% CI -0.70 to -0.60,  $t = 25.46$ ,  $p < 0.0001$ ) (Fig. 3c). The former 2 scaling relationships differed significantly between carnivorous and herbivorous mammals ( $t = 3.08$ ,  $p < 0.005$  and  $t = 2.37$ ,  $p = 0.02$ , respectively). Carnivores tended to have larger home ranges and shorter range crossing times than comparably sized herbivores, and herbivores tended to have longer range crossing times. The relationship

between  $N_{\text{area}}$  and mass did not differ between dietary classes ( $t = 0.82$ ,  $p = 0.06$ ). The  $N_{\text{area}}$  was governed by both  $\tau_p$  and sampling duration,  $T$ , such that  $N_{\text{area}} \approx T/\tau_p$ . Although we noted a positive correlation between body mass and  $T$  in the studies we sampled ( $\beta = 0.24$ , 95% CI 0.09–0.39,  $t = 3.17$ ,  $p < 0.005$ ), this was not enough to counter the positive correlation between mass and  $\tau_p$ . Consequently, the net result was a negative relationship between body mass and  $N_{\text{area}}$  ( $\beta = -0.23$ , 95% CI -0.39 to -0.08,  $t = 2.98$ ,  $p < 0.005$ ) (Fig. 3d).

#### Correction Factors

Model-informed data thinning served to reduce the mean autocorrelation at lag 1 from 0.96 (95% CI 0.96–0.97) to 0.32 (95% CI 0.30–0.35) (Fig. 4). Hence, an independent and identically distributed model was the best fit for 167 of the 463 individuals for which sufficient data ( $>2$  locations) remained after data thinning. The remaining individuals were best described by OU and OUF processes whose autocorrelation parameters were not significant. Although thinning mitigated the correlation between bias and body mass ( $\beta = -2.41$ , 95% CI -6.08 to



**Figure 4.** Frequency of amounts of autocorrelation at lag 1 in the full tracking data sets for each of the 757 individuals used to estimate home ranges via conventional kernel density estimation (KDE), compared with the thinned data sets for individuals for which sufficient data remained after thinning to apply KDE.

1.26,  $t = 1.29$ ,  $p = 0.20$ ), the median cross-validation rate of 95% home ranges estimated using the thinned data was only 85.1% (95% CI 83.6–86.5). This approximately 3% decrease in performance, as compared with conventional KDE on the full data, was likely the result of the small sample size. Model-informed data thinning resulted in a mean data loss of 93.2% (95% CI 92.1–94.3), and the median number of approximately independent locations left in each data set after thinning was only 23 (95% CI 18–26). Furthermore, in approximately 20% of the individuals,  $\leq 2$  locations remained after thinning, making it impossible to estimate a home-range area on the thinned data.

#### Autocorrelation-Informed Home-Range Estimation

Like model-informed data thinning, autocorrelation-informed home-range estimation via AKDE also eliminated the correlation between cross-validation success and body mass ( $\beta = -0.51$ , 95% CI -1.88 to 0.86,  $t = 0.73$ ,  $p = 0.47$ ). However, without the data loss required by the thinning approach, AKDE resulted in a median cross-validation rate of 95.2% (95% CI 94.2–95.9) for 95% home ranges and 51.3% (95% CI 49.26–54.36) for 50% home ranges. In other words, AKDE exhibited consistent accuracy across species, irrespective of the allometries in autocorrelation time scales and effective sample sizes.

**Table 1.** Estimates of the scaling exponent ( $b$ ) of mass to home-range area relationship\*.

Category	KDE (95% CI)	AKDE (95% CI)
All mammals	1.20 (0.95–1.45)	1.28 (1.01–1.54)
Herbivores and frugivores	1.26 (0.99–1.52)	1.38 (1.09–1.66)
Carnivores and omnivores	1.23 (0.95–1.50)	1.27 (1.01–1.56)

\* Abbreviations: KDE, kernel density estimation; AKDE, autocorrelated-kernel density estimation.

#### Scaling of Mammalian Space Use

When regressing home-range area against mass with conventional KDE estimates, we documented no significant difference from linear scaling for either herbivores or carnivores (Table 1). For AKDE-derived area estimates, however, we detected that the scaling exponent was significantly  $>1$  for both taxonomic groups, suggesting home-range area scales with mass according to a power function.

#### Discussion

The importance of autocorrelation in animal-tracking data has been an active area of research for decades (Swihart & Slade 1985; Fieberg 2007; Fleming et al. 2015). We, however, are the first to demonstrate that mass-specific space requirements driven by autocorrelation-induced underestimation of home-range areas are worse for larger species. From a fundamental perspective, the continuous nature of animal movement means quantities, such as positions, velocities, and accelerations, are necessarily autocorrelated (Fleming et al. 2014). Autocorrelation time scales ( $\tau$ ) should, therefore, be viewed as explicit attributes of an animal's movement process (Gurarie & Ovaskainen 2015) that are revealed when the temporal resolution of measurement becomes  $\lesssim \tau$ . As technological advances continue to permit ever-finer sampling (Kays et al. 2015), persistent autocorrelation is likely to become the norm in animal-tracking data. Pairing data from inherently autocorrelated processes with statistical approaches that ignore autocorrelation not only risks biasing any derived quantities, but also effectively negates the technological advances that are improving data quality. Unless analyses that are informed by autocorrelation become adopted by movement ecologists and conservationists, the issue of autocorrelation-induced bias will only worsen. Conversely, properly harnessing the wealth of information provided by autocorrelation can dramatically improve the accuracy of tracking-data-derived measures (see also Fleming & Calabrese 2017; Winner et al. 2018; Noonan et al. 2019). Our findings, therefore, highlight the need for more statistical estimators that can handle biologically induced variance without introducing bias.

### Implications of Size-Dependent Bias

From a conservation perspective, the underestimation of home-range areas is a worst-case scenario. When reserves are too small, relative to their target species' area requirements, the probability of local populations undergoing declines or extirpations increases significantly (Brashares et al. 2001; Gaston et al. 2008). Undersized protected areas resulting from poorly estimated space needs also risk exacerbating the issue of negative human–wildlife interactions at reserve boundaries (Van Eeden et al. 2018) as animals move beyond reserve boundaries to meet their energetic requirements (Farhadinia et al. 2018). It is thus of critical importance that policy actions be well informed about species' spatial requirements. To this end, we analyzed a broad taxonomic and geographic range of data and identified a strong correlation between home-range underestimation and body size when autocorrelation was ignored; average bias was approximately 50% at the upper end of the mass spectrum. In this regard, the majority of home ranges are estimated via methods based on the assumption of statistically independent data (Noonan et al. 2019). Combined with the facts that humans are the dominant mortality source for terrestrial vertebrates globally (Hill et al. 2019), that this mortality is higher for large-bodied species (Hill et al. 2020), and that megafauna are experiencing more severe range contractions (Tucker et al. 2018) and extinction risk (Cardillo et al. 2005), the most threatened species are also likely to be those with the least accurate home-range estimates, a worrying combination.

Based on these findings, we suggest that any conservation initiatives or policy based on home-range estimates derived from estimators based on the assumption of statistically independent data be revisited, especially where large-bodied species are involved. To facilitate this, we developed HRcorrect, an open-access application that allows users to correct a home-range area estimate for their focal species' body-mass-specific-bias with a correction factor calculated from our cross-validation regression models. The current version of HRcorrect is freely available from <https://hrcorrect.shinyapps.io/HRcorrect/>. However, there are numerous factors beyond body mass that influence an individual's home-range size. For instance, mammalian home-range areas are well known to covary with the spatial distribution of resources (Litvaitis et al. 1986; Boutin 1990), social structure (Lukas & Clutton-Brock 2013), sex (Cederlund & Sand 1994; Lukas & Clutton-Brock 2013; Noonan et al. 2018), age (Cederlund & Sand 1994), population density (Adler et al. 1997), and reproductive status (Rootes & Chabreck 1993; Noonan et al. 2018). Furthermore, if an individual's space use changes over time (e.g., interseasonal and -annual variation), a home-range area estimated from a single observation period may not be representative of its long-term area requirements. As

such, the deterministic trend-based correction provided by HRcorrect is not a substitute for more rigorous data collection and home-range estimation and should only be used for cases where the underlying tracking data are not accessible.

### Allometries and Conservation Theory

The metabolic theory of ecology (West et al. 1997) suggests that body mass represents a super trait that governs a wide range of ecological processes. Prime among these is the relationship between body mass and home-range area, an allometry that has guided ecological theory for more than 50 years (McNab 1963; Calder 1983; Jetz et al. 2004). More recently, attempts have been made to integrate this allometry into conservation theory. For instance, Hilbers et al. (2016) incorporated the home-range allometry into a method for quantifying mass-specific extinction vulnerability, and Hirt et al. (2018) highlighted how allometries in movement and space use can be used to make testable predictions of movement and biodiversity patterns at the landscape scale. Similarly, Pereira et al. (2004) used allometries of space use and movement rates to predict species-level vulnerability to land-use change. If the underlying allometries are biased, however, hypothesis testing and conservation planning in this context can fail even if the logic behind the experimental design is perfectly sound. Although the earliest derivation of the home-range allometry proposed a metabolically determined  $M^{0.75}$  allometry (McNab 1963), subsequent revisions showed no support for a purely energetic basis for home-range scaling (Calder 1983; Kelt & Van Vuren 2001; Jetz et al. 2004; Tucker et al. 2014; Tamburello et al. 2015). Although all these studies concluded that home-range area should scale with an exponent greater than the 0.75 predicted by metabolic requirements alone, there has been little consensus on whether the allometry is linear ( $M^1$ ) or superlinear ( $M^{>1}$ ). Our results suggest that at least part of the confusion can be attributed to the increasing bias in underestimating home ranges with increasing body size. Ours is the first study to estimate this relationship directly from tracking data by applying a consistent estimator across all individuals and, crucially, correcting for any potential autocorrelation-induced bias (Noonan et al. 2019). In doing so, we documented a super-linear relationship between body mass and home-range area (exponent of approximately 1.25 for  $M$ ). This shift from linear to power-law scaling fundamentally changes the behavior of the relationship, particularly at the upper end of the mass spectrum. Although we did not investigate the mechanisms behind the deviation from the metabolically determined  $M^{0.75}$ , we encourage future work on this subject to be based on the assumption of a superallometry, as opposed to linear allometry. Accurately quantifying species' area requirements is a prerequisite for

successful, area-based conservation planning. Our results highlight an important yet hitherto unrecognized aspect of home-range estimation: autocorrelation-induced negative bias in home-range estimation that is systematically worse for large species. Crucially, however, our findings also outline a readily applicable solution to the problem of size-dependent bias. We demonstrated that home-range estimation that properly accounts for the autocorrelation structure of the data is currently the only consistently reliable solution for eliminating allometric biases in home-range estimation (see also Noonan et al. 2019). We emphasize that the differential scaling of autocorrelation across the mass spectrum be a key consideration for movement ecologists and conservation practitioners and suggest avoiding home-range estimators that assume statistically independent data.

## Acknowledgments

This work was supported by a Smithsonian Institution Scholarly Studies Award to M.J.N., J.M.C., and A.L.H. and by the U.S. NSF Advances in Biological Informatics program (ABI-1458748 to J.M.C., W.F.F., and C.H.F.). N.B., F.J., and W.U. were supported by Deutsche Forschungsgemeinschaft in the framework of the BioMove Research Training Group (DFG-GRK 2118/1). T.M. and M.T. were funded by the Robert Bosch Foundation. S.L. was supported by Animals on the Move (NNX15AV92A), a NASA Arctic Boreal Vulnerability Experiment-funded project. This work was supported in part by the Wellcome Trust/DBT India Alliance Fellowship to A.T.V. (grant IA/CPHI/15/1/502028) and an IISc-ISRO Space Technology Cell Grant to M.T. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

## Supporting Information

Data set summary statistics (Appendix S1), individual tracking data set summaries (Appendix S2), and mammalian phylogenetic relationships (Appendix S3) are available online. The authors are solely responsible for the content and functionality of these materials. Queries (other than absence of the material) should be directed to the corresponding author.

### Supplementary Material

## Literature Cited

Abrahms B. 2017. Data from: Suite of simple metrics reveals common movement syndromes across vertebrate taxa. MovebankData Repository. <https://doi.org/10.5441/001/1.hm5nk220>.

Adler G, Endries M, Piotter S. 1997. Spacing patterns within populations of a tropical forest rodent, *Proechimys semispinosus*, on five Panamanian islands. *Journal of Zoology* **241**:43–53.

Agnarsson I, Kuntner M, May-Collado LJ. 2010. Dogs, cats, and kin: a molecular species-level phylogeny of carnivorans. *Molecular Phylogenetics and Evolution* **54**:726–745.

Barriel V, Thuet E, Tassy P. 1999. Molecular phylogeny of Elephantidae. Extreme divergence of the extant forest African elephant. *Comptes Rendus de l'Académie des Sciences-Series III-Sciences de la Vie* **322**:447–454.

Barton KA, Zwijacz-Kozica T, Zieba F, Sergiel A, Selva N. 2019. Bears without borders: long-distance movement in human-dominated landscapes. *Global Ecology and Conservation* **17**:e00541.

Boutin S. 1990. Food supplementation experiments with terrestrial vertebrates: patterns, problems, and the future. *Canadian Journal of Zoology* **68**:203–220.

Brashares JS, Arcese P, Sam MK. 2001. Human demography and reserve size predict wildlife extinction in West Africa. *Proceedings of the Royal Society B: Biological Sciences* **268**:2473–2478.

Brooks TM, et al. 2002. Habitat loss and extinction in the hotspots of biodiversity. *Conservation Biology* **16**:909–923.

Burnham KP, Anderson DR, Huyvaert KP. 2011. AIC model selection and multimodel inference in behavioral ecology: some background, observations, and comparisons. *Behavioral Ecology and Sociobiology* **65**:23–35.

Burt WH. 1943. Territoriality and home range concepts as applied to mammals. *Journal of Mammalogy* **24**:346–352.

Calabrese JM, Fleming CH, Gurarie E. 2016. ctmm: an R package for analyzing animal relocation data as a continuous-time stochastic process. *Methods in Ecology and Evolution* **7**:1124–1132.

Calder WA, III. 1983. Ecological scaling: mammals and birds. *Annual Review of Ecology and Systematics* **14**:213–230.

Cardillo M, Mace GM, Jones KE, Bielby J, Bininda-Emonds OR, Sechrest W, Orme CDL, Purvis A. 2005. Multiple causes of high extinction risk in large mammal species. *Science* **309**:1239–1241.

Cederlund G, Sand H. 1994. Home-range size in relation to age and sex in moose. *Journal of Mammalogy* **75**:1005–1012.

Cross PC, Bowers JA, Hay CT, Wolhuter J, Buss P, Hofmeyr M, du Toit JT, Getz WM. 2016. Data from: Nonparametric kernel methods for constructing home ranges and utilization distributions. Movebank Data Repository. <https://doi.org/10.5441/001/1.j900f88t>.

Dong X, Fleming CH, Calabrese JM. 2017. ctmm webapp: a graphical user interface for the ctmm R package. Available from <http://ctmm.shinyapps.io/ctmmweb/> (accessed February 2020).

Fahrig L. 2007. Non-optimal animal movement in human-altered landscapes. *Functional Ecology* **21**:1003–1015.

Farhadinia MS, Johnson PJ, Macdonald DW, Hunter LT. 2018. Anchoring and adjusting amidst humans: ranging behavior of Persian leopards along the Iran-Turkmenistan borderland. *PLOS ONE* **13** (e0196602). <https://doi.org/10.1371/journal.pone.0196602>.

Fieberg J. 2007. Kernel density estimators of home range: smoothing and the autocorrelation red herring. *Ecology* **88**:1059–1066.

Fleming CH, Calabrese JM. 2017. A new kernel density estimator for accurate home-range and species-range area estimation. *Methods in Ecology and Evolution* **8**:571–579.

Fleming CH, Calabrese JM, Mueller T, Olson KA, Leimgruber P, Fagan WF. 2014. Non-Markovian maximum likelihood estimation of autocorrelated movement processes. *Methods in Ecology and Evolution* **5**:462–472.

Fleming CH, Fagan WF, Mueller T, Olson KA, Leimgruber P, Calabrese JM. 2015. Rigorous home range estimation with movement data: a new autocorrelated kernel density estimator. *Ecology* **96**:1182–1188.

Fleming CH, Noonan MJ, Medici EP, Calabrese JM. 2019. Overcoming the challenge of small effective sample sizes in home-range estimation. *Methods in Ecology and Evolution* **10**:1679–1689.

Gaston KJ, Jackson SF, Cantu-Salazar L, Cruz-Piñón G. 2008. The ecological performance of protected areas. *Annual Review of Ecology, Evolution, and Systematics* **39**:93–113.

Grafen A. 1989. The phylogenetic regression. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences* **326**:119–157.

Gurarie E, Ovaskainen O. 2015. Characteristic spatial and temporal scales unify models of animal movement. *The American Naturalist* **178**:113–123.

Hall P, Hart JD. 1990. Convergence rates in density estimation for data from infinite-order moving average processes. *Probability Theory and Related Fields* **87**:253–274.

Harvey PH, Pagel MD. 1991. The comparative method in evolutionary biology. Oxford University Press, Oxford, United Kingdom.

Hilbers J, Schipper A, Hendriks A, Verones F, Pereira H, Huijbregts M. 2016. An allometric approach to quantify the extinction vulnerability of birds and mammals. *Ecology* **97**:615–626.

Hill J, DeVault T, Wang G, Belant J. 2020. Anthropogenic mammal mortality increases with the human footprint. *Frontiers in Ecology and the Environment* **18**:13–18.

Hill JE, DeVault TL, Belant JL. 2019. Cause-specific mortality of the world's terrestrial vertebrates. *Global Ecology and Biogeography* **28**:680–689.

Hirt MR, Grimm V, Li Y, Rall BC, Rosenbaum B, Brose U. 2018. Bridging scales: allometric random walks link movement and biodiversity research. *Trends in Ecology & Evolution* **33**:701–712.

Jetz W, Carbone C, Fulford J, Brown JH. 2004. The scaling of animal space use. *Science* **306**:266–268.

Johansson Ö, Rauset GR, Samelius G, McCarthy T, Andrén H, Tumur-sukh L, Mishra C. 2016. Land sharing is essential for snow leopard conservation. *Biological Conservation* **203**:1–7.

Kays R, Crofoot MC, Jetz W, Wikelski M. 2015. Terrestrial animal tracking as an eye on life and planet. *Science* **348**:aaa2478.

Kays R, Hirsch BT. 2015. Data from: Stink or swim: techniques to meet the challenges for the study and conservation of small critters that hide, swim or climb and may otherwise make themselves unpleasant. Movebank Data Repository. <https://doi.org/10.5441/001/1.8d8385j0>.

Kelt DA, Van Vuren DH. 2001. The ecology and macroecology of mammalian home range area. *The American Naturalist* **157**:637–645.

Lindblad-Toh K, et al. 2005. Genome sequence, comparative analysis and haplotype structure of the domestic dog. *Nature* **438**:803–819.

Linnell JD, Andersen R, Kvam T, Andrén H, Liberg O, Odden J, Moa PF. 2001. Home range size and choice of management strategy for lynx in Scandinavia. *Environmental Management* **27**:869–879.

Litvaitis JA, Sherburne JA, Bissonette JA. 1986. Bobcat habitat use and home range size in relation to prey density. *Journal of Wildlife Management* **50**:110–117.

Liu F-GR, Miyamoto MM, Freire NP, Ong PQ, Tennant MR, Young TS, Gugel KF. 2001. Molecular and morphological supertrees for eutherian (placental) mammals. *Science* **291**:1786–1789.

Lukas D, Clutton-Brock TH. 2013. The evolution of social monogamy in mammals. *Science* **341**:526–530.

Matthee CA, van Vuuren BJ, Bell D, Robinson TJ. 2004. A molecular supermatrix of the rabbits and hares (Leporidae) allows for the identification of five intercontinental exchanges during the Miocene. *Systematic Biology* **53**:433–447.

McMahon LA, Rachlow JL, Shipley LA, Forbey JS. 2017. Data from: Evaluation of micro-GPS receivers for tracking small-bodied mammals. Movebank Data Repository. <https://doi.org/10.5441/001/1.b0g6rq1t>.

McNab BK. 1963. Bioenergetics and the determination of home range size. *The American Naturalist* **97**:133–140.

Morato RG, et al. 2018. Jaguar movement database: a GPS-based movement dataset of an apex predator in the neotropics. *Ecology* **99**:1691–1691.

Noonan MJ, et al. 2018. Data from: A comprehensive analysis of autocorrelation and bias in home range estimation. Dryad Digital Repository. <https://doi.org/10.5061/dryad.v5051j2>.

Noonan MJ, et al. 2019. A comprehensive analysis of autocorrelation and bias in home range estimation. *Ecological Monographs* **89**:e01344.

Noonan MJ, Newman C, Markham A, Bilham K, Buesching CD, Macdonald DW. 2018. In situ behavioral plasticity as compensation for weather variability: implications for future climate change. *Climatic Change* **149**:457–471.

Paradis E, Schliep K. 2019. ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* **35**:526–528.

Pawitan Y. 2001. In all likelihood: statistical modelling and inference using likelihood. Clarendon Press, Oxford, United Kingdom.

Pe'er G, et al. 2014. Toward better application of minimum area requirements in conservation planning. *Biological Conservation* **170**:92–102.

Pereira HM, Daily GC, Roughgarden J. 2004. A framework for assessing the relative vulnerability of species to land-use change. *Ecological Applications* **14**:730–742.

Perelman P, et al. 2011. A molecular phylogeny of living primates. *PLOS Genetics* **7**:e1001342.

Pinheiro J, Bates D, DebRoy S, Sarkar D; R Core Team. 2018. nlme: linear and nonlinear mixed effects models. R package version 3.1-137. Available from <https://CRAN.R-project.org/package=nlme> (accessed February 2020).

Price SA, Bininda-Emonds ORP, Gittleman JL. 2005. A complete phylogeny of the whales, dolphins and even-toed hoofed mammals (Cetartiodactyla). *Biological Reviews of the Cambridge Philosophical Society* **80**:445–473.

Rootes WL, Chabreck RH. 1993. Reproductive status and movement of adult female alligators. *Journal of Herpetology* **27**:121–126.

Steiner CC, Ryder OA. 2011. Molecular phylogeny and evolution of the Perissodactyla. *Zoological Journal of the Linnean Society* **163**:1289–1303.

Swihart RK, Slade NA. 1985. Testing for independence of observations in animal movements. *Ecology* **66**:1176–1184.

Tamburello N, Côté IM, Dulvy NK. 2015. Energy and the scaling of animal space use. *The American Naturalist* **186**:196–211.

Tucker MA, et al. 2018. Moving in the Anthropocene: global reductions in terrestrial mammalian movements. *Science* **359**:466–469.

Tucker MA, Ord TJ, Rogers TL. 2014. Evolutionary predictors of mammalian home range size: body mass, diet and the environment. *Global Ecology and Biogeography* **23**:1105–1114.

Uhlenbeck GE, Ornstein LS. 1930. On the theory of the Brownian motion. *Physical Review* **36**:823–841.

Van Eeden LM, Crowther MS, Dickman CR, Macdonald DW, Ripple WJ, Ritchie EG, Newsome TM. 2018. Managing conflict between large carnivores and livestock. *Conservation Biology* **32**:26–34.

West GB, Brown JH, Enquist BJ. 1997. A general model for the origin of allometric scaling laws in biology. *Science* **276**:122–126.

Wilman H, Belmaker J, Simpson J, de la Rosa C, Rivadeneira MM, Jetz W. 2014. Eltontraits 1.0: species-level foraging attributes of the world's birds and mammals. *Ecological Archives* E095-178. *Ecology* **95**:2027–2027.

Wilson MC, et al. 2016. Habitat fragmentation and biodiversity conservation: key findings and future challenges. *Landscape Ecology* **31**:219–227.

Winner K, Noonan MJ, Fleming CH, Olson KA, Mueller T, Sheldon D, Calabrese JM. 2018. Statistical inference for home range overlap. *Methods in Ecology and Evolution* **9**:1679–1691.