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# Effects of Freshwater Salinization and Biotic Stressors on Amphibian Morphology

Jacquelyn L. Lewis<sup>1</sup>, Jonathan J. Borrelli<sup>1</sup>, Devin K. Jones<sup>2</sup>, and Rick A. Relyea<sup>1</sup>

Organisms are commonly exposed to numerous stressors that induce behavioral, physiological, or morphological changes in some combination. At northern temperate latitudes, de-icing agents (primarily sodium chloride, NaCl) are a major stressor to species in freshwater ecosystems. Species-specific responses to road salt toxicity range from lethal to sublethal effects, but it remains unclear how these effects interact with biotic stressors. Morphology can be quite sensitive to environmental changes, yet we know little about how it is affected by road salt exposure. We exposed Wood Frog tadpoles (Rana sylvatica) to two road salt formulations (NaCl and a mixture of NaCl, MgCl<sub>2</sub>, and KCl), each at three concentrations (200, 600, and 1000 mg Cl<sup>-</sup>/L), crossed with three biotic stressor levels (predator cue, competition, and a no-stressor control). We then measured the impacts on relative morphology (snout-vent length, body width, forelimb length, forelimb width, hindlimb length, hindlimb width) of the emerging metamorphs. Salt concentration and biotic stressors both impacted relative morphology, but their effects did not interact. Exposure to road salts increased relative snout-vent length (SVL) and body width. In contrast, competition induced relatively shorter SVL and forelimb length while predator cues induced relatively longer hindlimbs and narrower forelimbs. This is the first discovery that road salts can induce changes in amphibian morphology and that these effects are independent of changes induced by biotic stressors. Future research should examine the effects on overwintering success and future fitness in amphibians as well as the impacts of salt on the morphology of other aquatic taxa that are being exposed to freshwater salinization.

RGANISMS are commonly exposed to a wide range of biotic and abiotic stressors. Although exposure stressors such as parasites, competitors, and predators are ubiquitous within natural systems (Relyea and Hoverman, 2003; Paull and Johnson, 2014; Relyea, 2018), anthropogenic stressors, such as habitat alteration and use of synthetic chemicals, have increasingly introduced novel environmental conditions (Collins and Russell, 2009; Novarro et al., 2018). Stressor exposure can induce changes in behavior (Carr, 2002; Hintz and Relyea, 2017), physiology (Robison et al., 2018), morphology (Relyea, 2012; Payette and Sullivan, 2019), or some combination (Woodley and Lacy, 2010; Lewis and Sullivan, 2020) that allows individuals to cope with and allay stressors. However, these plastic responses can also compromise their performance and fitness, such as reducing ability to forage, defend territories, or find mates (Watson et al., 2004; Ghalambor et al., 2007; Wack et al., 2013). Given the pervasive contamination of natural systems (Malaj et al., 2014), it is vital to understand how contaminants might influence responses to environmental stressors.

In northern temperate zones, salinization is a growing concern to freshwater ecosystems and can act as a major stressor on freshwater organisms (Herbert et al., 2015). Deicing agents, primarily sodium chloride (NaCl), enter freshwater systems by infiltrating ground water and surface runoff (Marsalek, 2003). Although some have noted that the primary reasons for clearing and salting roads are for the economy and safety, salinization of adjacent aquatic habitats can affect ecosystem health and biodiversity (Karraker et al., 2008; Herbert et al., 2015).

Spring-breeding amphibians might be at higher risk to road salt runoff as many species use vernal pools following the influx of water to low-lying areas during snow melt (Karraker et al., 2008). Laboratory studies that estimate mean lethal concentrations (LC<sub>50</sub>) have shown species-specific responses to salt, including high sensitivity (LC<sub>50</sub> = 1000-2000 mg Cl<sup>-</sup>/ L) in Wood Frogs (Rana sylvatica) and Spotted Salamanders (Ambystoma maculatum), intermediate sensitivity (LC<sub>50</sub> = 2500–3500 mg Cl<sup>-</sup>/L) in Spring Peepers (*Pseudacris crucifer*) and Green Frogs (Rana clamitans), and low sensitivity ( $LC_{50} =$  $\sim$ 4000 mg Cl<sup>-</sup>/L) in American Toads (*Anaxyrus americanus*; Collins and Russell, 2009). Similarly, Karraker et al. (2008) found reduced embryonic and larval survival in Spotted Salamanders at moderate (500 μS; roughly 150 mg Cl<sup>-</sup>/L) and high (3000 μS; roughly 950 mg Cl<sup>-</sup>/L) conductivities and in Wood Frogs at high conductivities. Environmentally realistic concentrations of road salt also increase juvenile mortality (Dananay et al., 2015) and physical abnormalities of Wood Frogs while reducing time to metamorphosis, weight, and activity (Sanzo and Hecnar, 2006). While laboratory studies are an important first step for the investigation of the direct lethal and sublethal effects of road salts, they lack the complexity organisms face in natural aquatic communities.

Impacts of contaminants often have interactive effects with biotic stressors, such as predation and competition. Several studies indicate interactive effects of predation stress and environmental stressors, including pesticides (Boone and Semlitsch, 2001; Relyea and Mills, 2001; Boone and James, 2003; Relyea, 2003, 2004, 2012), fertilizers (Ortiz-Santaliestra et al., 2010), and UV-B radiation (Alton et al., 2010). For example, a number of studies have shown that predator cues make pesticides more deadly (Relyea and Mills, 2001; Relyea, 2003, 2004). Similarly, competitive stress can make herbicides significantly more lethal (Jones et al., 2011) and has interactive effects with insecticides (Boone and Bridges, 1999; Boone and Semlitsch, 2002; Boone and James, 2003),

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fertilizers (Ortiz-Santaliestra et al., 2012), and coal-combustion waste (Roe et al., 2006).

There is growing interest in whether salt exposure might also interact with biotic stressors, especially with respect to sublethal effects. For example, there is no interactive effect of salt and predator cues on Rainbow Trout (*Oncorhynchus mykiss*) activity and growth, but the two stressors do have interactive effects on zooplankton reproduction (*Daphnia pulex;* Hintz and Relyea, 2017). In two species of amphibians reared under combinations of salts and biotic stressors (i.e., competition and predator cues), no salt-by-stressor interaction was found on survival, activity, time to metamorphosis, or mass at metamorphosis (Jones et al., 2017). Despite growing interest, the few studies that have examined interactions between salt exposure and biotic stressors have primarily focused on behavior and life history traits.

In addition to behavior and life history traits, contaminant exposure and biotic stressors can influence morphological and physiological traits. In tadpoles, for example, pesticides can cause skeletal and muscle abnormalities (Bernabò et al., 2008, 2011), alter gill morphology (Bernabò et al., 2008, 2011), and increase incidents of malformation (Lavorato et al., 2013; Bernabò et al., 2016; reviewed by Pinelli et al., 2019). Surprisingly, the herbicide Roundup® induces relatively deeper tails in the same direction and magnitude as predator cues (Relyea, 2012, 2018). Predation stress can also increase relative leg length of newly metamorphic frogs (Relyea, 2001; Relyea and Hoverman, 2003) and decreases tail regeneration in salamanders (Payette and Sullivan, 2019). In contrast, competitive stress through an elevated density of tadpoles produces smaller frog metamorphs with relatively shorter limbs and bodies (Relyea and Hoverman, 2003). Furthermore, in natural ponds, Wood Frog absolute snoutvent length (SVL) increased with elevated road salt concentrations (Dananay et al., 2015). Nonetheless, it remains unknown how exposure to road salts will influence the morphology of aquatic taxa.

Our goal was to examine the individual and combined effects of road salts (two types at three concentrations) and biotic stressors (predator cue, competition, no-stressor control) on the relative morphology of metamorphosed amphibians. Based on past studies, we expected predation stress to increase relative leg length and competition to decrease relative SVL and leg length, with salt possibly interfering with these responses. As absolute SVL seems to increase correspondingly to elevated road salt concentrations in natural ponds, we expected similar results for relative SVL which is standardized to account for deviation in mass. However, given the lack of prior studies on road salts, we had no a priori predictions for how tadpole exposure to different road salts and concentrations would alter other morphological measurements.

## **MATERIALS AND METHODS**

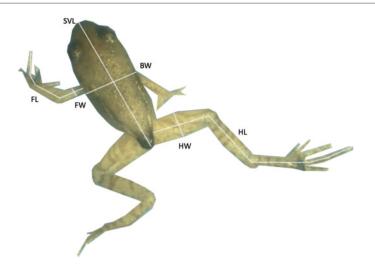
Experimental design.—To determine effects of road salt and biotic stressors on amphibian morphology, we measured external morphology of recently metamorphosed Wood Frogs preserved from Jones et al. (2017). In brief, Jones et al. (2017) conducted a factorial experiment that used seven salt treatments: 1) a no-salt control, 2) three concentrations of NaCl (95–100% pure; 200, 600, and 1000 mg Cl<sup>-</sup>/L), and 3) three concentrations of a salt mixture containing chloride-

based alternatives (NaCl, MgCl $_2$ , and KCl; 200, 600, and 1000 mg Cl $^-$ L). The seven salt treatments were crossed with two biotic stressors (predator cue, competition, and a no-stressor control). The 21 treatment combinations were replicated four times for a total of 84 experimental units.

To simulate a natural wetland, we used mesocosms, outdoor experimental systems which mimic natural conditions and serve as a bridge between field studies and highly controlled laboratory experiments. Each 90 L outdoor mesocosm served as an experimental unit and was filled with 82 L of tap water, leaf litter, and a homogenized aliquot of pond water containing periphyton, phytoplankton, and zooplankton. Tap water was left for several days to allow for the chlorine to off-gas prior to the addition of leaf litter and pond water. Newly hatched American Toad larvae and newly hatched Wood Frog larvae were also added. Initial Wood Frog and American Toad masses were 147±10 and 38±3 mg, respectively (mean ± SE; Jones et al., 2017); both species were at Gosner developmental stage 25 (Gosner, 1960). Competitor stress was manipulated by doubling the number of Wood Frog and toad tadpoles in the respective mesocosms, such that 20 total individuals of each species (density of 24 individuals/m<sup>2</sup>) were added to competition treatments while ten individuals of each species (density of 12 individuals/m<sup>2</sup>) were added to the predator-cue and no-stressor treatments. These densities are well within the densities observed in natural wetlands (Relyea et al., unpubl. data). Furthermore, the original experiment showed significant effects of competitive environment on Wood Frog responses to increased tadpole densities (Jones et al., 2017), indicating that even if the densities chosen do not represent true high and low densities, the higher density was enough to increase competition for resources in the mesocosms. Each mesocosm contained a single predator cage; to manipulate predator stress, we added a single dragonfly larva (Anax junius) to the appropriate cages. The dragonfly larvae were fed approximately 300 mg of larval Wood Frogs three times per week. Previous studies indicated that larval Wood Frogs show antipredator behavior and morphological changes to caged dragonfly larvae that are fed this biomass of prey (Schoeppner and Relyea, 2008).

Wood Frogs were removed from the mesocosms daily as they metamorphosed, which we defined as having four legs and any amount of tail. We housed the metamorphs in the laboratory until tail sorption resulted in tail lengths of <2 mm, at which time they were euthanized and preserved in a 10% formalin solution. The experiment ended at day 49 when the last individual metamorphosed. Biotic stressors had significant effects on amphibian survival, time to metamorphosis, and mass at metamorphosis, but there was no effect of salt or salt-by-environment interactions (Jones et al., 2017). While there was a significant effect of environment on survival, 94-98% of amphibians remained alive to the end of the experiment (Jones et al., 2017). Therefore, we did not consider any effect of survival on morphology. In the current study, we examined the impacts of the 21 treatment combinations on the relative morphology of the Wood Frogs.

Morphometric analysis.—Morphometric data for each Wood Frog were obtained using an Olympus SZX16 microscope and cellSens software. To limit exposure to formaldehyde, frogs were removed from preservation vials and placed in tap water. Frogs were then towel dried, weighed, and photo-



**Fig. 1.** The morphological dimensions that were measured on Wood Frog metamorphs: SVL = snout-vent length, BW = body width, FL = forelimb length, FW = forelimb width, HL = hindlimb length, and HW = hindlimb width.

graphed. For photographs, individuals were pinned to a dissecting tray with their dorsal surface facing up. We then used the software to measure SVL, body width, forelimb length, forelimb width, hindlimb length, and hindlimb width (Fig. 1). SVL was defined as the length from the cloaca to tip of the nose. Body width was measured from the bottom of one armpit to the other. Forelimb length and hindlimb length were both measured from the start of the limb to the tip of the longest toe. Forelimb width and hindlimb width were taken at the widest part of the limbs, perpendicular to the length measurement. The person measuring the animals was not aware of the treatments.

Statistical analysis.—To evaluate the effect of road salt treatment, environment stressor, and their possible interaction on relative morphology, we ran a multivariate analysis of covariance (MANCOVA) with salt and environment as main effects, mass as a covariate, and all possible interactions. We log-transformed the mass and morphological data so relationships between mass and the morphological measurements were linear. We used tank means to eliminate pseudoreplication. We confirmed that all treatments had parallel slopes (a key assumption when assessing massadjusted morphology) by confirming that mass did not interact with any of the treatment main effects or their interaction. Logged tank means met the assumptions of normality and homoscedasticity. To evaluate which body measurements were causing the multivariate effect, we performed subsequent analyses of covariance (ANCOVAs) for each of the six body measurements, while excluding all of the mass-by-treatment interactions since they were not significant in the MANCOVA. When main effects were significant, we conducted a Dunnett test to compare the effects of each salt treatment to the no-salt control and each biotic stressor to the no-stressor control. Statistical analyses were all conducted in R version 3.6.1 (R Core Team, 2019).

### RESULTS

MANCOVA identified significant effects of road salt treatment, environment (biotic stressors), and the mass covariate,

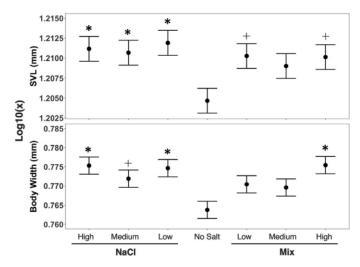
**Table 1.** Results of the multivariate analysis of covariance (MANCOVA) for log-transformed morphological variables of post-metamorphic Wood Frogs. The model was analyzed using salt treatment and tadpole environment as main effects, mass as a covariate, and all of the possible interactions. Removing the mass-by-treatment interactions in the MANCOVA had no qualitative effect on the effects of salt, environment, or their interaction.

	df	Wilks	P
Salt	6	0.194	0.001
Environment	2	0.004	< 0.001
Mass	1	0.039	< 0.001
Salt × Environment	12	0.174	0.315
Salt × Mass	6	0.468	0.686
Environment × Mass	2	0.834	0.846
Salt $\times$ Environment $\times$ Mass	12	0.204	0.539
Residuals	42		

but there was no salt-by-environment interaction (Table 1). The subsequent ANCOVAs indicated that the salt treatments affected relative SVL and body width (Table 2; Fig. 2). Relative SVL increased in all concentrations of NaCl (low: P = 0.008; medium: P = 0.039; high: P = 0.021); it also marginally increased in the low (P = 0.060) and high (P = 0.069) concentrations of the salt mixture. Body width increased (or marginally increased) in all concentrations of NaCl (low: P = 0.005; medium: P = 0.060; and high: P = 0.003); it also increased when exposed to the highest concentration of the salt mixture (P = 0.002). In terms of actual amount of

**Table 2.** Results of the analyses of covariance (ANCOVA) for each of the six log-transformed Wood Frog metamorph body measurements (SVL, body width, forelimb length, forelimb width, hindlimb length, hindlimb width). Each ANCOVA included salt treatment and tadpole environment as main effects and log-transformed mass as a covariate. Interactions were not included in the ANCOVAs since they were not significant in the MANCOVA (Table 1).

	df	F	P
Snout-vent length			
Salt	6	2.379	0.037
Environment	2	2.544	0.085
Mass	1	526.480	< 0.001
Body width			
Salt	6	3.473	0.004
Environment	2	0.923	0.402
Mass	1	231.514	< 0.001
Forelimb length			
Salt	6	1.542	0.177
Environment	2	2.918	0.060
Mass	1	154.994	< 0.001
Forelimb width			
Salt	6	0.892	0.506
Environment	2	2.861	0.064
Mass	1	192.841	< 0.001
Hindlimb length			
Salt	6	0.946	0.468
Environment	2	3.731	0.029
Mass	1	217.926	< 0.001
Hindlimb width			
Salt	6	0.673	0.672
Environment	2	0.512	0.602
Mass	1	211.930	< 0.001



**Fig. 2.** The log10 values of relative SVL and body width of Wood Frogs in response to seven salt treatments. Data points represent Ismeans ( $\pm 1$  SE). Using Dunnett tests to compare each salt treatment against the no-salt control, asterisks represent significant differences (P < 0.05) and crosses (+) indicate marginally significant differences (0.05 < P < 0.1).

morphological change (using non-logged values), NaCl lengthened SVL by 1.4 to 1.7% and body width by 1.9 to 2.8%. The salt mixture lengthened SVL by 1.2 to 1.3% and body width by 2.8%. When considering jumping performance, the ratio of hindlimb to SVL decreased at most by 1.3% in both salt formulations and across concentration.

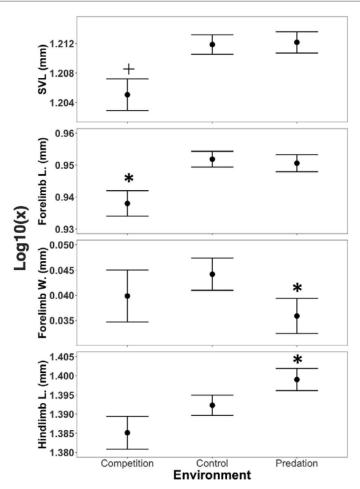
The ANCOVAs also indicated that environment marginally affected SVL while significantly affecting forelimb length, forelimb width, and hindlimb length (Table 2; Fig. 3). Compared to the no-stressor control, competition induced a marginally significant decrease in relative SVL (P=0.060) and a significant decrease in forelimb length (P=0.036). In contrast, predator cues induced significantly narrower forelimbs (P=0.040) and longer hindlimbs (P=0.047). In real terms, competition shortened relative SVL by 1.6% and forelimb length by 3.1% while predation narrowed forelimbs by 1.8% and lengthened hindlimbs by 1.5%.

#### DISCUSSION

Secondary salinization of freshwater systems is an increasingly problematic global scenario (Cañedo-Argüelles et al., 2019). To understand how road salt contamination might influence species' responses to natural stressors, we explored the effects of road salt and biotic stressors on Wood Frog morphology and found that both factors induced morphological changes.

We discovered that competition induces relatively shorter SVL and forelimbs while predator cues induce relatively narrow forelimbs and longer hindlimbs. Furthermore, road salt induced relatively longer SVL and body width. However, we did not observe any salt-by-environment interactions. This is consistent with studies looking at other sublethal effects in Rainbow Trout (Hintz and Relyea, 2017), American Toads, and Wood Frogs (Jones et al., 2017), both of which found no interaction between salt and biotic stressors.

The induced changes we observed are likely linked to morphological changes occurring in the tadpole stage. For instance, tadpoles reared in low resources or high competi-



**Fig. 3.** The log10 values of relative SVL, forelimb length, forelimb width, and hindlimb length of Wood Frog metamorphs in response to larval exposure to three biotic stressors. Data points represent Ismeans ( $\pm 1$  SE). Using Dunnett tests to compare each stressor environment treatment against the no-stressor control, asterisks represent significant differences (P < 0.05) and crosses (+) indicate marginally significant differences (0.05 < P < 0.1).

tion develop relatively small tails and large bodies (Relyea, 2002, 2004; Relyea and Auld, 2005; Van Buskirk, 2009). In considering the subsequent effects as metamorphs, Southern Leopard metamorphs (*Rana sphenocephala*) reared under competition as tadpoles exhibit relatively longer hindlimbs (Emerson, 1986). Relyea and Hoverman (2003) found that larval competition induced Treefrog (*Hyla versicolor*) metamorphs to develop 1% shorter relative SVL and 3% shorter relative forelimbs. These results align with our study of Wood Frogs, with metamorphs induced to have 1.6% shorter SVL and 3.1% shorter forelimbs after larval exposure to competition.

Tadpole exposure to predator cues has a contrasting effect. Many species of tadpoles develop relatively deep tails and small bodies in response to predator cues (Relyea, 2001; Van Buskirk, 2002, 2009; Richter-Boix et al., 2007). When such Wood Frogs emerge as metamorphs, we found they have 1.5% longer hindlimbs. Similarly, Relyea (2001) found that larval predation stress increased relative hindlimb length by 3.4% and forelimb length by 2.9% in Wood Frog metamorphs. However, Relyea (2001) found that relative forelimb length increased by 2.9% whereas forelimb length decreased by 1.8% in our study.

Our results demonstrate carry-over effects across metamorphosis indicating larval conditions have the potential to shape post-metamorphic life. Carry-over effects from environmental stressors early in ontogeny to later in life are found across taxa including insects (Taylor, 1988; Anholt, 1990, 1991; De Moed et al., 1997; Block and Stoks, 2005; Koenraadt et al., 2010; Stoks and Córdoba-Aguilar, 2012), mammals (Ravelli et al., 1976), birds (Haywood and Perrins, 1992; Merila and Svensson, 1997), marine invertebrates (Woollacott et al., 1989; Pechenik et al., 1998), and fishes (Brönmark and Pettersson, 1994; Royle et al., 2006; Roussel, 2007; Chivers et al., 2008). In damselflies, for example, larval competition results in later emergence and lower mass (Anholt, 1990), both of which had negative consequences for later adult fitness (Anholt, 1991; Block and Stoks, 2005; reviewed by Stoks and Córdoba-Aguilar, 2012). Predation stress in fishes induces morphological changes that are not entirely reversible (Brönmark and Pettersson, 1994; Chivers et al., 2008). For example, Crucian Carp (Carassius carassius) increase body depth in response to Northern Pike (Esox lucius) chemical cues and do not return to the same body depth as control even after 180 days in the absence of predation stress (Brönmark and Pettersson, 1994). Across a range of taxa, early ontogeny stress may have irreversible effects in later life stages.

To our knowledge, this is the first study to report on the effects of road salts on multiple morphological traits. We noted an increase in relative SVL across road salt type and concentration, which is similar to Dananay et al.'s (2015) report of a positive correlation between absolute SVL and road salt concentration in Wood Frogs from ponds in northeastern Ohio. In contrast, Kearney et al. (2016) used seawater instead of road salt and found that exposure to high larval salt concentrations (12% seawater) decreases mass and absolute SVL in freshwater Brown Treefrog (*Litoria ewingii*) populations. Overall, the two morphological changes that we observed were similar for each of the two salt formulations and were generally similar across the three concentrations used. This suggests there may be some generalities in how various road salts induce metamorph morphology.

A small number of amphibian studies have examined how road salts affect life history traits. For instance, Albecker and McCoy (2019) found that American Green Treefrogs (H. cinerea) in salt-adapted populations metamorphized earlier and smaller compared to salt-naïve inland populations, but they maintained a constant size across experimentally elevated salt concentrations while inland frogs metamorphized much smaller in high salt concentrations than in lower concentrations. Jones et al. (2017) reported no effect of road salt on tadpole survival, activity, time to metamorphosis, or mass at metamorphosis, but indicated a significant reduction in toad activity in high NaCl (1000 mg Cl-/L) treatments. Plant studies have shown elevated salt leading to phenotypic plasticity with changes in the number of leaves, leaf size (Richards et al., 2005), root structure (Echeverria et al., 2008), and plant architecture (Suter and Widmer, 2013).

Our results highlight the lasting effects of salinization on the post-metamorphic stages of animals. Similarly, other studies indicate salt to have persistent effects. For example, Karraker and Gibbs (2011) reported reduced mass by 33% in Spotted Salamanders (*Ambystoma maculatum*) that were exposed to high salt concentrations (945 mg Cl<sup>-</sup>/L) with mass reduction continuing even when returned to low salt

concentrations. Furthermore, Lambert et al. (2017) found that elevated road salt during larval development can masculinize Wood Frog metamorphs. Salt concentrations of 867 mg Cl<sup>-</sup>/L decreased the ratio of female metamorphs in their experimental population by 10%. In contrast, Matlaga et al. (2014) did not report a lag effect of salt exposure with American Bullfrogs (Lithobates catesbeianus) as embryos reared in NaCl (7, 100, 500, and 1000 mg Cl-/L) were not more vulnerable to future predation events. Outside of amphibians, Atlantic Salmon (Salmo salar) eggs within the first 24 h after fertilization showed global transcriptional changes when exposed to 5000 mg Cl<sup>-</sup>/L of road salt (Tollefsen et al., 2015). Although it has not been determined whether the transcriptional changes have adverse effects, they are suggested to interfere with osmoregulation, ion regulation, oxidative stress, metabolism, renal function, and development in the embryos, which would likely span across ontogeny. While the research is limited, early exposure to road salt appears to have persistent effects across life stages.

The morphological changes induced following larval exposure to predation, competition, and road salt in Wood Frog metamorphs are relatively small, so it is important to consider whether these changes have ecological consequences. For example, Brady et al. (2019) reported increased body mass and farther jumping in female Wood Frogs from populations in close proximity to roads with high salt concentrations. The morphological changes in our study ranged from 1.2-2.8% for road salt and 1.5-3.1% for biotic stressors. Previous studies report a minimum of a 10% change in the ratios of relative hindlimb length to SVL between species before jumping is significantly affected (Stokely and Berberian, 1953; Zug, 1972; Emerson, 1978, 1986). Although reporting on 18 different anuran species, it is unclear how these studies might apply to the jumping ability of Wood Frogs, and, to our knowledge, there have been no jumping performance studies in Wood Frogs. Certainly, much more work needs to be done on this question, including whether sublethal salt exposures affect other traits.

Conclusion.—Our study discovered significant effects of biotic stressors and road salts during the larval period on the morphology of emerging metamorphs, indicating the importance of studies spanning across life stages. Future studies should examine whether similar morphological changes are also inducible in other species of amphibians and whether other aquatic species experience altered morphology from salt exposures. If so, then it is important to know what concentrations and salt formulations (many of which may contain chemical additives such as anti-caking agents) have an effect, the underlying mechanisms that cause these induced changes, and the short- and long-term impacts of these morphological changes on the performance of the induced individuals.

#### **DATA ACCESSIBILITY**

All data are available from the Dryad Digital Repository: https://doi.org/10.5061/dryad.8gtht76kx. Unless otherwise indicated in the figure caption, the published images and illustrations in this article are licensed by the American Society of Ichthyologists and Herpetologists for use if the use includes a citation to the original source in accordance with the Creative Commons Attribution CC BY License.

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#### LITERATURE CITED

- Albecker, M. A., and M. W. McCoy. 2019. Local adaptation for enhanced salt tolerance reduces non-adaptive plasticity caused by osmotic stress. Evolution 73:1941–1957.
- Alton, L. A., R. S. Wilson, and C. E. Franklin. 2010. Risk of predation enhances the lethal effects of UV-B in amphibians. Global Change Biology 16:538–545.
- Anholt, B. R. 1990. An experimental separation of interference and exploitative competition in larval damselfly. Ecology 71:1483–1493.
- Anholt, B. R. 1991. Measuring selection on a population of damselflies with a manipulated phenotype. Evolution 45: 1091–1106.
- Bernabò, I., E. Brunelli, C. Berg, A. Bonacci, and S. Tripepi. 2008. Endosulfan acute toxicity in *Bufo bufo* gills: ultrastructural changes and nitric oxide synthase localization. Aquatic Toxicology 86:447–456.
- Bernabò, I., L. Gallo, E. Sperone, S. Tripepi, and E. Brunelli. 2011. Survival, development, and gonadal differentiation in *Rana dalmatina* chronically exposed to chlorpyrifos. Journal of Experimental Zoology Part A: Ecological Genetics and Physiology 315A:314–327.
- Bernabò, I., A. Guardia, R. Macirella, S. Sesti, A. Crescente, and E. Brunelli. 2016. Effects of long-term exposure to two fungicides, pyrimethanil and tebuconazole, on survival and life history traits of Italian tree frog (*Hyla intermedia*). Aquatic Toxicology 172:56–66.
- Block, M. D., and R. Stoks. 2005. Fitness effects from egg to reproduction: bridging the life history transition. Ecology 86:185–197.
- Boone, M. D., and C. M. Bridges. 1999. The effect of temperature on the potency of carbaryl for survival of tadpoles of the green frog (*Rana clamitans*). Environmental Toxicology and Chemistry 18:1482–1484.
- Boone, M. D., and S. M. James. 2003. Interactions of an insecticide, herbicide, and natural stressors in amphibian community mesocosms. Ecological Applications 13:829– 841.
- Boone, M. D., and R. D. Semlitsch. 2001. Interactions of an insecticide with larval density and predation in experimental amphibian communities. Conservation Biology 15: 11
- Boone, M. D., and R. D. Semlitsch. 2002. Interactions of an insecticide with competition and pond drying in amphibian communities. Ecological Applications 12:307–316.
- Brady, S. P., F. J. Zamora-Camacho, F. A. A. Eriksson, D. Goedert, M. Comas, and R. Calsbeek. 2019. Fitter frogs from polluted ponds: the complex impacts of human-altered environments. Evolutionary Applications 12:1360–1370.

- Brönmark, C., and L. B. Pettersson. 1994. Chemical cues from piscivores induce a change in morphology in crucian carp. Oikos 70:396.
- Cañedo-Argüelles, M., B. Kefford, and R. Schäfer. 2019. Salt in freshwaters: causes, effects and prospects—introduction to the theme issue. Philosophical Transactions of the Royal Society B: Biological Sciences 374:20180002.
- Carr, J. A. 2002. Stress, neuropeptides, and feeding behavior: a comparative perspective. Integrative and Comparative Biology 42:582–590.
- Chivers, D. P., X. Zhao, G. E. Brown, T. A. Marchant, and M. C. O. Ferrari. 2008. Predator-induced changes in morphology of a prey fish: the effects of food level and temporal frequency of predation risk. Evolutionary Ecology 22:561–574.
- Collins, S. J., and R. W. Russell. 2009. Toxicity of road salt to Nova Scotia amphibians. Environmental Pollution 157: 320–324.
- Dananay, K. L., K. L. Krynak, T. J. Krynak, and M. F. Benard. 2015. Legacy of road salt: apparent positive larval effects counteracted by negative postmetamorphic effects in wood frogs. Environmental Toxicology and Chemistry 34:2417–2424.
- De Moed, G. H., G. De Jong, and W. Scharloo. 1997. The phenotypic plasticity of wing size in *Drosophila melanogaster*: the cellular basis of its genetic variation. Heredity 79: 260–267.
- Echeverria, M., A. A. Scambato, A. I. Sannazzaro, S. Maiale, O. A. Ruiz, and A. B. Menéndez. 2008. Phenotypic plasticity with respect to salt stress response by *Lotus glaber*: the role of its AM fungal and rhizobial symbionts. Mycorrhiza 18:317–329.
- Emerson, S. B. 1978. Allometry and jumping in frogs: helping the twain to meet. Evolution 32:551–564.
- Emerson, S. B. 1986. Heterochrony and frogs: the relationship of a life history trait to morphological form. American Naturalist 127:167–183.
- Ghalambor, C. K., J. K. McKay, S. P. Carroll, and D. N. Reznick. 2007. Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. Functional Ecology 21:394–407.
- Gosner, K. L. 1960. A simplified table for staging anuran embryos and larvae with notes on identification. Herpetologica 16:183–190.
- Haywood, S., and C. M. Perrins. 1992. Is clutch size in birds affected by environmental conditions during growth? Proceedings: Biological Sciences 249:195–197.
- Herbert, E. R., P. Boon, A. J. Burgin, S. C. Neubauer, R. B. Franklin, M. Ardón, K. N. Hopfensperger, L. P. M. Lamers, and P. Gell. 2015. A global perspective on wetland salinization: ecological consequences of a growing threat to freshwater wetlands. Ecosphere 6:art206.
- Hintz, W. D., and R. A. Relyea. 2017. A salty landscape of fear: responses of fish and zooplankton to freshwater salinization and predatory stress. Oecologia 185:147–156.
- Jones, D. K., J. I. Hammond, and R. A. Relyea. 2011. Competitive stress can make the herbicide Roundup® more deadly to larval amphibians. Environmental Toxicology and Chemistry 30:446–454.
- Jones, D. K., B. M. Mattes, W. D. Hintz, M. S. Schuler, A. B. Stoler, L. A. Lind, R. O. Cooper, and R. A. Relyea. 2017. Investigation of road salts and biotic stressors on freshwa-

- ter wetland communities. Environmental Pollution 221: 159–167.
- Karraker, N. E., and J. P. Gibbs. 2011. Road deicing salt irreversibly disrupts osmoregulation of salamander egg clutches. Environmental Pollution 159:833–835.
- Karraker, N. E., J. P. Gibbs, and J. R. Vonesh. 2008. Impacts of road deicing salt on the demography of vernal poolbreeding amphibians. Ecological Applications 18:724–734.
- Kearney, B. D., P. G. Byrne, and R. D. Reina. 2016. Anuran developmental plasticity loss: the cost of constant salinity stress. Australian Journal of Zoology 63:331–337.
- Koenraadt, C. J. M., M. Kormaksson, and L. C. Harrington. 2010. Effects of inbreeding and genetic modification on Aedes aegypti larval competition and adult energy reserves. Parasites & Vectors 3:92.
- Lambert, M. R., A. B. Stoler, M. S. Smylie, R. A. Relyea, and D. K. Skelly. 2017. Interactive effects of road salt and leaf litter on wood frog sex ratios and sexual size dimorphism. Canadian Journal of Fisheries and Aquatic Sciences 74: 141–146.
- Lavorato, M., I. Bernabò, A. Crescente, M. Denoël, S. Tripepi, and E. Brunelli. 2013. Endosulfan effects on *Rana dalmatina* tadpoles: quantitative developmental and behavioural analysis. Archives of Environmental Contamination and Toxicology 64:253–262.
- Lewis, J. L., and A. M. Sullivan. 2020. Salamander stress and duress: the relationship between CORT, autotomy and regeneration, and exploratory behaviour. Zoology 139: 125751.
- Malaj, E., P. C. von der Ohe, M. Grote, R. Kühne, C. P. Mondy, P. Usseglio-Polatera, W. Brack, and R. B. Schäfer. 2014. Organic chemicals jeopardize the health of freshwater ecosystems on the continental scale. Proceedings of the National Academy of Sciences of the United States of America 111:9549–9554.
- Marsalek, J. 2003. Road salts in urban stormwater: an emerging issue in stormwater management in cold climates. Water Science and Technology 48:61–70.
- Matlaga, T. H., C. A. Phillips, and D. J. Soucek. 2014. Insensitivity to road salt: an advantage for the American bullfrog? Hydrobiologia 721:1–8.
- Merila, J., and E. Svensson. 1997. Are fat reserves in migratory birds affected by condition in early life? Journal of Avian Biology 28:279–286.
- Novarro, A. J., C. R. Gabor, C. B. Goff, T. D. Mezebish, L. M. Thompson, and K. L. Grayson. 2018. Physiological responses to elevated temperature across the geographic range of a terrestrial salamander. Journal of Experimental Biology 221:jeb178236.
- Ortiz-Santaliestra, M. E., M. J. Fernández-Benéitez, and A. Marco. 2012. Density effects on ammonium nitrate toxicity on amphibians. Survival, growth and cannibalism. Aquatic Toxicology 110–111:170–176.
- Ortiz-Santaliestra, M. E., M. J. Fernández-Benéitez, A. Marco, and M. Lizana. 2010. Influence of ammonium nitrate on larval anti-predatory responses of two amphibian species. Aquatic Toxicology 99:198–204.
- Paull, S. H., and P. T. J. Johnson. 2014. Experimental warming drives a seasonal shift in the timing of host-parasite dynamics with consequences for disease risk. Ecology Letters 17:445–453.
- Payette, W. I., and A. M. Sullivan. 2019. The effect of predator kairomones on caudal regeneration by Allegheny

- Mountain Dusky Salamanders (*Desmognathus ochrophaeus*). Canadian Journal of Zoology 97:502–509.
- Pechenik, J. A., D. E. Wendt, and J. N. Jarrett. 1998. Metamorphosis is not a new beginning. BioScience 48: 901–910.
- Pinelli, C., A. Santillo, G. Chieffi Baccari, S. Falvo, and M. M. Di Fiore. 2019. Effects of chemical pollutants on reproductive and developmental processes in Italian amphibians. Molecular Reproduction and Development 86:1324–1332.
- R Core Team. 2019. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org/
- Ravelli, G. P., Z. A. Stein, and M. W. Susser. 1976. Obesity in young men after famine exposure in utero and early infancy. The New England Journal of Medicine 295:349– 353.
- Relyea, R. A. 2001. The lasting effects of adaptive plasticity: predator-induced tadpoles become long-legged frogs. Ecology 82:1947–1955.
- Relyea, R. A. 2002. Competitor-induced plasticity in tadpoles: consequences, cues, and connections to predator-induced plasticity. Ecological Monographs 72:523–540.
- Relyea, R. A. 2003. Predator cues and pesticides: a double dose of danger for amphibians. Ecological Applications 13: 1515–1521.
- Relyea, R. A. 2004. Synergistic impacts of malathion and predatory stress on six species of North American tadpoles. Environmental Toxicology and Chemistry 23:1080–1084.
- Relyea, R. A. 2012. New effects of Roundup on amphibians: predators reduce herbicide mortality; herbicides induce antipredator morphology. Ecological Applications 22:634–647.
- Relyea, R. A. 2018. The interactive effects of predator stress, predation, and the herbicide Roundup. Ecosphere 9: e02476.
- Relyea, R. A., and J. R. Auld. 2005. Predator- and competitor-induced plasticity: how changes in foraging morphology affect phenotypic trade-offs. Ecology 86: 1723–1729.
- Relyea, R. A., and J. T. Hoverman. 2003. The impact of larval predators and competitors on the morphology and fitness of juvenile treefrogs. Oecologia 134:596–604.
- Relyea, R. A., and N. Mills. 2001. Predator-induced stress makes the pesticide carbaryl more deadly to gray treefrog tadpoles (*Hyla versicolor*). Proceedings of the National Academy of Sciences of the United States of America 98: 2491–2496.
- Richards, C. L., S. C. Pennings, and L. A. Donovan. 2005. Habitat range and phenotypic variation in salt marsh plants. Plant Ecology 176:263–273.
- Richter-Boix, A., G. A. Llorente, and A. Montori. 2007. A comparative study of predator-induced phenotype in tadpoles across a pond permanency gradient. Hydrobiologia 583:43–56.
- Robison, A. L., T. Chapman, and J. R. Bidwell. 2018. Predation cues influence metabolic rate and sensitivity to other chemical stressors in fathead minnows (*Pimephales promelas*) and *Daphnia pulex*. Ecotoxicology 27:55–68.
- Roe, J. H., W. A. Hopkins, S. E. DuRant, and J. M. Unrine. 2006. Effects of competition and coal-combustion wastes on recruitment and life history characteristics of salaman-

- ders in temporary wetlands. Aquatic Toxicology 79:176–184.
- Roussel, J.-M. 2007. Carry-over effects in brown trout (*Salmo trutta*): hypoxia on embryos impairs predator avoidance by alevins in experimental channels. Canadian Journal of Fisheries and Aquatic Sciences 64:786–792.
- Royle, N. J., J. Lindström, and N. B. Metcalfe. 2006. Effect of growth compensation on subsequent physical fitness in green swordtails *Xiphophorus helleri*. Biology Letters 2:39– 42.
- Sanzo, D., and S. J. Hecnar. 2006. Effects of road de-icing salt (NaCl) on larval wood frogs (*Rana sylvatica*). Environmental Pollution 140:247–256.
- Schoeppner, N. M., and R. A. Relyea. 2008. Detecting small environmental differences: risk-response curves for predator-induced behavior and morphology. Oecologia 154: 743–754.
- Stokely, P. S., and J. F. Berberian. 1953. On the jumping ability of frogs. Copeia 1953:187.
- Stoks, R., and A. Córdoba-Aguilar. 2012. Evolutionary ecology of *Odonata*: a complex life cycle perspective. Annual Review of Entomology 57:249–265.
- Suter, L., and A. Widmer. 2013. Phenotypic effects of salt and heat stress over three generations in *Arabidopsis* thaliana. PLoS ONE 8:e80819.
- Taylor, M. F. J. 1988. Field measurement of the dependence of life history on plant nitrogen and temperature for a herbivorous moth. Journal of Animal Ecology 57:873–891.
- Tollefsen, K. E., Y. Song, M. Kleiven, U. Mahrosh, S. Meland, B. O. Rosseland, and H.-C. Teien. 2015.

- Transcriptional changes in Atlantic salmon (*Salmo salar*) after embryonic exposure to road salt. Aquatic Toxicology 169:58–68.
- Van Buskirk, J. 2002. A comparative test of the adaptive plasticity hypothesis: relationships between habitat and phenotype in anuran larvae. American Naturalist 160:87– 102.
- Van Buskirk, J. 2009. Natural variation in morphology of larval amphibians: phenotypic plasticity in nature? Ecological Monographs 79:681–705.
- Wack, C. L., M. K. Ratay, and S. K. Woodley. 2013. Effects of corticosterone on locomotory activity in red-legged salamanders. Herpetologica 69:118–126.
- Watson, R. T., A. Mathis, and R. Thompson. 2004. Influence of physical stress, distress cues, and predator kairomones on the foraging behavior of Ozark zigzag salamanders, *Plethodon angusticlavius*. Behavioural Processes 65:201–209.
- Woodley, S. K., and E. L. Lacy. 2010. An acute stressor alters steroid hormone levels and activity but not sexual behavior in male and female Ocoee salamanders (*Desmog-nathus ocoee*). Hormones and Behavior 58:427–432.
- Woollacott, R. M., J. A. Pechenik, and K. M. Imbalzano. 1989. Effects of duration of larval swimming period on early colony development in *Bugula stolonifera* (Bryozoa: Cheilostomata). Marine Biology 102:57–63.
- Zug, G. R. 1972. Anuran locomotion: structure and function.
  I. Preliminary observations on relation between jumping and osteometrics of appendicular and postaxial skeleton.
  Copeia 1972:613–624.