

Disease or drought: environmental fluctuations release zebra from a potential pathogen-triggered ecological trap

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1 **Abstract**

2 When a transmission hotspot for an environmentally persistent pathogen establishes in
3 otherwise high-quality habitat, the disease may exert a strong impact on a host population.
4 However, fluctuating environmental conditions lead to heterogeneity in habitat quality and
5 animal habitat preference, which may interrupt the overlap between selected and risky habitats.
6 We evaluated spatiotemporal patterns in anthrax mortalities in a plains zebra (*Equus quagga*)
7 population in Etosha National Park, Namibia, incorporating remote-sensing and host telemetry
8 data. A higher proportion of anthrax mortalities of herbivores was detected in open habitats than
9 in other habitat types. Resource selection functions showed that the zebra population shifted
10 habitat selection in response to changes in rainfall and vegetation productivity. Average to high
11 rainfall years supported larger anthrax outbreaks, with animals congregating in preferred open
12 habitats, while a severe drought forced animals into otherwise less preferred habitats, leading to
13 few anthrax mortalities. Thus, the timing of anthrax outbreaks was congruent with preference for
14 open plains habitats and a corresponding increase in pathogen exposure. Given shifts in habitat
15 preference, the overlap in high-quality habitat and high-risk habitat is intermittent, reducing the
16 adverse consequences for the population.

17 **Keywords**

18 Anthrax, disease dynamics, ecological trap, environmental transmission, habitat selection,

19 transmission hotspot

20 **1. Introduction**

21 Habitat quality is context-dependent [1], where consumers distribute in response to resource
22 dynamics on a landscape [2-5]. This habitat heterogeneity in turn affects disease risk due to
23 uneven distributions in species diversity of hosts, vectors and reservoirs [6-8], parasite loads [8,
24 9], and abiotic variation [7, 8]. Moving across diverse and dynamic landscapes, mobile hosts
25 may encounter different rates of pathogen exposure across habitats [10, 11]. Bridging disease
26 dynamics and animal habitat use therefore may help understand variation in transmission
27 dynamics [12].

28 Reservoir dynamics of environmentally transmitted pathogens are likely to depend upon
29 spatial structure, which would in turn be affected by habitat heterogeneity. Aggregation of animal
30 hosts in preferred habitats may “seed” the environment with a higher density of pathogen
31 propagules, maintaining and spreading infection within higher resource quality habitats [13].
32 Since pathogens are unlikely to be detectable by hosts, transmission hotspots could establish in
33 otherwise high-quality habitat if animals are attracted to habitat with heightened disease risk
34 [14]. This habitat would be considered an “ecological trap” if habitat selection preferences lead
35 to lower fitness [15-17]. Ecological trap theory has mainly focused on the effects of predation,
36 food scarcity or anthropogenic disturbance on offspring survival [16-18]. However, habitats with

high concentrations of environmentally transmitted pathogens might also have potential to form ecological traps [13, 14].

This study aims to understand the effects of habitat quality and disease exposure on a susceptible and mobile host population, coupling habitat heterogeneity in disease risk, dynamics in habitat quality, and host habitat use. We evaluate whether a population of plains zebra (*Equus quagga*) susceptible to anthrax experiences an overlap in high-quality habitat and transmission hotspots, which could potentially lead to the formation of an ecological trap. Anthrax is a highly lethal, acute to peracute disease caused by the bacterium, *Bacillus anthracis*. Given its lethality, anthrax has potential fitness consequences for the zebra population, by reducing average fitness of individuals. Although an earlier demographic study concluded that anthrax may be limiting growth of this zebra population [19], we cannot assess whether an anthrax transmission hotspot is also an ecological trap, because we did not measure fitness differences among habitats.

Anthrax only transmits via the environment, and not directly between hosts [20], and hence provides an opportunity to evaluate the spatial variation in pathogen exposure. Transmission relies upon contact with long-lived spores in soil environmental reservoirs. These reservoirs are generated from host disease mortality and subsequent exposure of susceptible hosts through foraging at *B. anthracis*-laden carcass sites [21-24]. Thus, anthrax is a good candidate for investigating how an infectious disease could turn preferred habitat into a disease transmission

55 hotspot.

56 Here we explore the overlap between host habitat selection and areas of heightened anthrax
57 risk, and how preference and risk change with variation in resource availability. We hypothesized
58 that 1) habitat quality is variable in time (i.e. due to amount of annual precipitation), 2) the
59 ranking of available habitats by quality and attractiveness is not static, but varies based on
60 context, and 3) for an environmentally persistent and highly lethal pathogen with only
61 environmental transmission, animals are subject to an overlap in habitats of both high quality and
62 high disease risk—but only sometimes, depending on environmental conditions. Periods of
63 environmental fluctuation that allow reprieve from the transmission hotspot will reduce exposure
64 and the number of pathogen-induced mortalities affecting the population. We first assessed
65 variation in risk of contacting *B. anthracis* by habitat type. We then tested the relationships
66 among habitat dynamics, zebra resource selection among habitats with differential risk, and
67 anthrax mortalities, to evaluate how fluctuating habitat quality affects host habitat use and
68 pathogen exposure, and in turn disease dynamics.

69 **2. Methods**

70 **(a) Study area and periods**

71 Our study was conducted during two time periods (2009 – 2010; 2018 – 2020) in Etosha
72 National Park (ENP), a fenced 22,270-km² reserve located in northern Namibia (figure 1a). ENP

is a semi-arid savannah, with three seasons: cool dry season in May – August, hot semi-dry season in September – December, and hot wet season in January – April. Rainfall is strongly seasonal and occurs mainly between November and April, with the greatest monthly rainfall occurring in January and February [25]. There is a west-east rainfall gradient, increasing from an average of 200 to 450 mm/yr. Average annual rainfall in the central area is 358.0 ± 126.7 mm (mean \pm SD; Okaukuejo station 1954 – 2020). Rainfall is recorded by rainfall years (e.g., July 2009 – June 2010 is the rainfall year 2010), not calendar years. The specific dates of study were May 2009 – August 2010 and October 2018 – April 2020, during which we collected telemetry data from collared zebras. Precipitation in 2010 and 2020 was around or above the average (389.9 and 440.5 mm at Okaukuejo station, respectively), but 2019 was by far the driest year on record in ENP (83.7 mm; Okaukuejo station; figure S1a). The 2019 drought was the most severe drought in Namibia in the last 90 years [26].

Much of ENP is covered by mopane (*Colophospermum mopane*) shrubveld or treeveld, and large salt pans, with open grasslands along the pans. There are seven basic vegetation types described, including bare ground, grassland, steppe, grass savannah, shrub savannah, low tree savannah and high tree savannah [27, 28]. For this study, we grouped these seven categories into four basic habitat types: bare areas (i.e. salt pans), open plains (grassland, grass savannah and steppe), shrublands (shrub savannah), and woodlands (high tree and low tree savannah; figure

1a). These habitat types were used to evaluate resource fluctuation, disease risk and resource selection.

(b) Anthrax risk by habitat

Anthrax is endemic in ENP where plains zebra is the most common host detected, constituting more than 50% of cases [29]. An estimated cause of death for every observed mortality is assigned by an investigation of the carcass remains by park staff or researchers, and blood swabs for disease diagnosis are collected regardless of suspected cause of death. Anthrax mortality in this study was defined as laboratory-confirmed cases from bacterial culture and suspected cases based on symptoms if diagnostic samples were not collected. Signs of suspected anthrax mortalities include no evidence of predation, blood exudation and lack of clotting, and rigidly extended fore legs or oedematous swelling [30]. Zebra anthrax mortalities occur annually with strong seasonality in cases, which peak in the late wet season (March – April) [29, 31], and case numbers are positively correlated with annual rainfall [29] (figure S1b). Anthrax transmission relies on environmental reservoirs created by positive carcasses [21, 22]. Hence, more anthrax carcasses in an area represents a higher risk of exposure.

We used animal mortalities detected by opportunistic surveillance [32] to evaluate anthrax risk by habitat type. Opportunistic mortality surveillance can be biased toward certain landscape features such as access along the road network or vegetation density [33]. Surveillance may also

109 be biased toward detecting certain types of mortality sources based on how long the carcass
110 remains on the landscape. Given the challenges in detecting carcasses equally across the
111 landscape, we focus on the ratio of anthrax cases to other mortalities, as an index of disease risk
112 among habitats. This method assumes that detection rates of carcasses do not differ among
113 habitat types based on the cause of death. We probed this assumption by comparing a subset of
114 the mortality data during periods of heightened surveillance effort that would reduce detection
115 biases. During the periods represented in this study, there was an additional layer of mortality
116 surveillance based on sites where global positioning system (GPS)-tagged scavengers clustered
117 [34-36]. This additional effort yielded mortality records in locations that otherwise would go
118 undetected, and provided a better estimate of the number of anthrax mortalities, and their
119 temporal and spatial distribution during the study periods. The patterns detected during
120 heightened surveillance effort (figure S2) reflected those in the larger, opportunistic dataset, and
121 thus we feel that the patterns of relative anthrax risk among habitats are robust to differences in
122 surveillance effort.

123 We compared anthrax mortalities to mortalities from other natural causes (e.g., predation,
124 old age, starvation), to investigate anthrax risk in a habitat type. Using mortalities with recorded
125 GPS positions for herbivore species that had at least one anthrax case from 1998 to April 2020,
126 we summed the numbers of anthrax and other mortalities by habitat for all herbivore species.

Bare areas were excluded from these comparisons because animals infrequently used these areas, and no mortality surveillance occurred on the salt pans. In total, there were 737 anthrax mortalities, and 28.5% of them were suspected cases. We used chi-square tests of independence to examine differences in ratios of anthrax to other mortalities between habitat types, and further examined the differences by herbivore functional foraging type (grazing, mixed feeding and browsing). For the total mortalities and every foraging type, we conducted three chi-square tests with two habitat types selected for each, and corrected p values with Bonferroni correction.

(c) Dynamics in zebra anthrax cases and habitat quality

We counted accumulated numbers of zebra anthrax mortalities during the seasons involved in the study periods (2009 – 2010; 2018 – 2020), when there was enhanced surveillance, to assess spatiotemporal patterns in anthrax mortality in the zebra population. We summed the numbers of anthrax mortalities within each season, and further separated them based on habitat types.

We used a remotely-sensed index of vegetation primary production, the fraction of photosynthetically active radiation (FPAR) from Moderate Resolution Imaging Spectroradiometer (Terra MODIS; MOD15A2H) [37] to assess habitat dynamics and variation in quality during the study periods. FPAR is widely used to model vegetation biomass and productivity [27, 38]. Although it is subject to the change of foliage resulting in different values

across habitat types [27, 39], FPAR is a better predictor of grass biomass in ENP than other remotely-sensed data sources [27]. The spatial and temporal resolution was 500×500 m and 8-days starting at the 1st day of each year. We extracted FPAR values for the study area (excluding salt pans) and sampling periods, and calculated averages for each habitat type and season.

(d) Zebra habitat use

Zebra preferentially select shorter, more palatable grasses in open plains [28, 40], but in ENP these grasses get depleted in the dry season. To determine host habitat selection changes in response to resource fluctuation, we collected telemetry data with GPS collars (African Wildlife Tracking, Pretoria, South Africa) on adult zebras (9 females Jul 2009 – Aug 2010, 8 males and 10 females Oct 2018 – Aug 2019, and 6 males and 11 females Sep 2019 – Apr 2020) captured in central-eastern ENP.

We divided days into morning (06:00-12:00; GMT+1), afternoon (12:00-18:00) and night (18:00-06:00), and thinned the data by extracting readings closest to 9:00, 15:00 and 24:00 for the three periods of a day for each individual, to reduce autocorrelation in the telemetry data. The distance between locations of two consecutive readings of the thinned data could potentially be far enough for an individual to switch habitats (figure S3). If readings for an individual were fewer than 180 in a season (i.e. two months of sampling), the individual's data for that season were removed. Of thinned readings, 6.8% were in areas where vegetation types were not

available (figure 1), and these readings were excluded from habitat selection analyses.

We calculated seasonal habitat use for the population, as the percentage of readings in a habitat type by season. We then evaluated habitat selection with resource selection functions (RSFs), based on a use-availability framework [41] to account for the habitat availability in the area. These RSFs correspond to second-order selection, or how individuals select ranges compared to the habitats available in the overall population range [42]. Used versus available (not-used) habitat types were then compared with logistic regressions. To define an area with sufficient habitat connectivity from which to generate available locations, we applied a 99.9% kernel density home range estimation to the overall thinned readings from study animals (24,916 readings from 35 individuals) throughout the entire study (clipped by park fenceline; figure 1). The available points were generated with 10 times the thinned readings. Individual-based RSFs were performed to compare the habitats of used and available points [43] in each season, with habitat type as a covariate. We used three orthogonal contrast coding variables for the four-level categorical habitat covariate. The three contrast coding variables compared the preferences for 1) vegetated habitats over bare areas, for 2) open habitats (open plains) over closed habitats (shrublands and woodlands) and for 3) shrublands over woodlands. A regression coefficient represented relative selection strength of an individual [44], which in this case showed habitat preference for a specific habitat comparing to the other habitat type(s). To evaluate whether the

RSF findings were robust to the thinning approach selected, we compared these results with RSF results for fixed intervals of one to 24 hours. The results of the thinning approach we applied were consistent with fixed intervals of one to eight hours (figure S4).

We fit medians of relative selection strengths by season ($N = 9$ seasons) for the three different comparisons to FPAR with linear regressions, to evaluate the relationships between habitat selection and habitat dynamics. Because FPAR varied with habitat types, we used average FPAR at open habitats as an index of habitat dynamics.

In addition to habitat preference, we investigated how consistently zebras used open habitats, to evaluate how likely an individual would be to die in the open habitats after infected there. We explored the probability that an individual exposed in open habitats would remain in that habitat for the duration of the incubation period (figure S5). From this we conclude that although daily movement distances can be relatively long (out to 9 km/hr; figure S3) [45], the average distances are much smaller (the median across individuals and seasons: 0.26 km/hr; figure S3), and zebras have the highest probability of remaining in open habitats when anthrax cases are most prevalent (75% in wet season; figure S5).

(e) Integrating dynamics in habitats, host selection and anthrax mortality

We evaluated whether seasonal variation in habitat dynamics or host habitat selection could predict the number of anthrax mortalities. We fit anthrax case numbers by season ($N = 9$ seasons)

with linear regression separately to FPAR and the median of relative selection strength. We chose the relative selection strength corresponding to selection of high versus low risk habitats, based on the results of analysis for anthrax risk by habitat. The numbers of anthrax mortalities were square-root transformed due to overdispersion and small sample size of the dataset. As in the previous analysis, here average FPAR at open habitats was used as the index of habitat dynamics.

All analyses were done in R 3.6.1 [46]. FPAR was downloaded from National Aeronautics and Space Administration (NASA) Land Processes Distributed Active Archive Center by package MODISTsp [47], and values were extracted by package raster [48]. Kernel density range was estimated using package adehabitatHR [49], and clipped by fenceline with package rgeos [50]. Available points were generated with package sf [51]. Package sp [52, 53] was used to retrieve vegetation/habitat types for used and available points. Chi-square tests, Bonferroni correction, logistic regression and linear regression were done using package stats [46].

3. Results

(a) Anthrax risk by habitat

In general, more herbivore mortalities were found in open habitats than shrublands or woodlands, and anthrax mortality risk was highest in open habitats (figure 2). Mortality data from all herbivorous anthrax host species showed relatively more anthrax mortalities than other mortality sources in open plains than in shrublands ($X_1^2 = 14.36, p < 0.001$; figure 2a) or

woodlands ($X_1^2 = 91.54, p < 0.001$; figure 2a), and relatively more anthrax mortalities in shrublands than in woodlands ($X_1^2 = 32.03, p < 0.001$; figure 2a). Comparing host species by functional foraging types, grazing herbivores had a higher proportion of anthrax mortalities in open plains than in woodlands ($X_1^2 = 14.93, p < 0.001$; figure 2b) and a higher proportion in shrublands than in woodlands ($X_1^2 = 10.36, p < 0.01$; figure 2b). Mixed-feeding herbivores also had a higher proportion of anthrax mortalities in open plains than in woodlands ($X_1^2 = 10.36, p < 0.01$; figure 2c). Browsing herbivores rarely died of anthrax, with no significant differences in the proportion of anthrax mortalities between habitats (figure 2d). More browsing herbivore mortalities were detected in closed than in open habitats, in contrast to higher carcass detection in open habitats for other foraging types. These patterns of relative anthrax risk among habitats for the longer time series (1998 – April 2020) were corroborated by patterns observed during the shorter periods of increased surveillance effort (figure S2).

(b) Dynamics in zebra anthrax cases and in habitat quality

Peaks in total zebra anthrax mortalities correlated with seasonal peaks in vegetation productivity in wet seasons (figure 3), with more cases in open habitats than closed habitats at those productivity peaks. However, inter-annual rainfall patterns affected the intensity of these seasonal anthrax outbreaks. Reduced primary production during the 2019 drought corresponded with very few anthrax deaths: there were only 12 zebra anthrax mortalities recorded in 2018 –

2019, with eight in the wet season (figure 3). The average and above average rainfall years had larger anthrax outbreaks but at different scales, with 144 zebra anthrax mortalities in 2009 – 2010 (94 cases in the wet season; figure 3) and 18 zebra anthrax mortalities in the wet season of 2020 (figure 3), respectively.

(c) Zebra habitat use

Zebra habitat selection preferences varied with rainfall amount and fluctuations in primary production, except for a consistent avoidance of bare areas ($R^2 = 0.006$, $t = -0.21$, $p = 0.837$, $N = 9$; figures 4 and 5). Zebra predominantly used open plains over any closed habitats in the wet season of years with average and above average rainfall when there was higher vegetation productivity (figures 4a and 4b). However, during the semi-dry season in 2009 and during the 2019 drought, vegetation productivity was reduced, and zebra primarily used woodlands (figures 4a and 4b). In average and above average rainfall years, zebra showed tendencies to select open habitats over closed habitats and shrublands over woodlands, especially in the wet season (figures 4 and 5). In contrast, zebra selected closed habitats and woodlands during the 2019 drought (figures 4 and 5). Both relative selection strengths for open over closed habitats and for shrublands over woodlands were significantly positively related to FPAR ($R^2 = 0.67$, $t = 3.75$, $p < 0.01$, $N = 9$; $R^2 = 0.63$, $t = 3.43$, $p < 0.05$, $N = 9$; figures 5b and 5c). No obvious sex differences in habitat selection were detected (figure 4), though sample sizes were relatively small.

(d) Integrating dynamics in habitats, host selection and anthrax mortality

The number of anthrax cases recorded in a season can be linked to the amount of primary productivity and the resulting host habitat selection preferences (figure 6). There was a significantly positive relationship between primary production and the square-root transformed number of anthrax cases recorded per season ($R^2 = 0.56$, $t = 2.97$, $p < 0.05$, $N = 9$; figure 6a). The relative selection strength shown by zebras for open habitats over closed habitats also associated with the number of anthrax mortalities ($R^2 = 0.60$, $t = 3.26$, $p < 0.05$, $N = 9$; figures 6b), where the stronger the preference for open habitats, the more anthrax cases were recorded.

4. Discussion

This study related resource dynamics and host habitat preferences to temporal and spatial variation in disease outbreak dynamics. Host habitat selection varied among seasons and rainfall years in response to environmental fluctuations and habitat dynamics. Zebra preferred open habitats with higher anthrax risk in wet seasons and wetter years, and showed correspondingly higher anthrax mortality associated with higher primary production. In dry seasons and drought, zebras shifted their selection preference away from risky habitats, with a corresponding reduction in anthrax mortality. Our results suggest that habitat dynamics and host habitat selection can be used to predict disease outbreaks for environmentally transmitted disease agents.

The associations detected between habitat selection and disease risk are supported by

seasonal differences in zebra diet selection [40] and pathogen exposure [54]. Together, these indicate that a disease transmission hotspot has developed in the open habitat preferred by zebras. Given the long-lived nature of *B. anthracis* spores, pathogen reservoirs can survive periods of low host density, and infect zebras when conditions support their return to this habitat. The short latency of infection and highly lethal nature of this disease [20, 55] is likely to maintain the transmission hotspot through a positive feedback loop. Zebras contracting the disease on the plains will probably die in these areas and create more local infectious zones, especially in the anthrax season (figure S5), and hence enhance pathogen concentrations in the hotspot habitats. Lack of fitness data prevented us from determining whether this transmission hotspot is also an ecological trap. However, since the transmission hotspot and preferred habitat only overlapped when resources were abundant, even if an ecological trap were to form, it would be temporary, and the detrimental effect on the host population reduced due to the reprieve of selecting different habitats in different periods.

Grazing herbivores may travel long distances, seeking areas with forage availability in response to a drought when grass biomass in preferred locations is depleted [56]. The zebra population selected the open plains in the wet season in the average to above-average rainfall years, and it switched to woodlands in the drought. The plains have palatable grasses which zebra preferentially select [40], but they get depleted in the dry season most years, forcing

individuals to use other habitats [45]. During a drought, grass biomass in the open plains does not recover, which prevents its selection by zebra. As a result, they occupy closed habitats where otherwise less palatable vegetation is found [28, 40]. Consequently, zebra use high disease risk habitats when their resources are abundant, and lower risk habitats when the more desirable habitat is depleted.

The spatial structure of exposure risk and host habitat selection are most likely to determine outbreak intensity in the zebra population rather than other potential factors, such as stress due to nutritional deprivation. These results imply that the zebra population never gets a really “excellent” year because it suffers either from a deadly disease or from food scarcity. However, the population also likely benefits from this shifting habitat selection, which staggers two negative impacts. The population is exposed to higher risk areas when resources are abundant and individuals are less stressed than in the dry periods [57]. This suggests that better health as a result of more resource availability may play a factor in preventing massive anthrax outbreaks in locations like ENP with wet-season outbreaks. In dry-season anthrax outbreaks, which can be more common in other locations, hundreds of animals per species can be impacted [58-60], and species-specific mortality rates in these outbreaks can reach higher than 90% [59]. In comparison, in the wet-season outbreak in 2010 (the biggest outbreak in ENP in nearly 40 years), the zebra anthrax mortality rate is estimated to be around 3.9%. This calculation is based upon

observed cases corrected with an estimated total to observed ratio of 3.8 (2.9-8.2) [32] and a population estimate of 9,225 (5,138-13,672) in the 99.9% kernel density range (figure 1; Etosha Ecological Institute unpublished aerial survey data from 2005).

The anthrax outbreak in 2010 was the largest in ENP since the early 1970s, and spilled over to the dry seasons. There was also an outbreak in the wet season 2020, though at a smaller scale. The variation in outbreak sizes between 2010 and 2020 may be attributed to factors such as multi-year rainfall patterns. Though spores can persist in the environment for decades, spore concentrations at a site decay over time, and carcass sites are most infectious within the first few years after host death [21]. Zebra anthrax cases are positively correlated with annual rainfall [29]. Hence, periods of above average rainfall and higher zebra anthrax mortality will increase the number of highly infectious reservoir sites, while dry periods will deplete them. High rainfall years with more zebra anthrax cases preceded the large outbreak recorded in 2010, while drier years with fewer anthrax cases preceded the smaller outbreak in 2020 (figure S1a).

This study provides new insight into spatiotemporal disease dynamics in mobile hosts. The congruence of anthrax outbreaks and host habitat preferences in this study suggests that when disease risk on a landscape is heterogeneous, disease dynamics can be predicted by host habitat selection. Other diseases transmitted via environmental reservoirs such as avian influenza, chronic wasting disease and white-nose syndrome can also have spatial heterogeneity in disease

325 risk as well as transmission hotspots on a landscape [61-63]. For these cases, because disease
326 dynamics can be predicted by habitat use, movement of the host population can be a key
327 component to understanding infection dynamics.

328 Ultimately, a long-term study would be required to monitor the host population growth rate
329 as well as disease dynamics, to determine if this transmission hotspot is also an ecological trap.

330 In addition, further study is needed to evaluate other potential mechanisms reinforcing a
331 transmission hotspot, such as changes in host foraging behaviour [29, 40]. Nevertheless, by
332 investigating changes in habitat quality and animal movements, our study suggests that disease
333 outbreaks can likely be determined by habitat dynamics and host resource selection when there is
334 spatial heterogeneity in exposure to pathogens. Due to shifts in host habitat selection in response
335 to resource dynamics, the detrimental effect of the overlap in high-quality and high-risk habitat
336 on the host population is only intermittent. Thus, a heterogeneous landscape and environmental
337 fluctuations may reduce the impact of an environmentally transmitted disease on a host
338 population. With the fortuitous circumstances of the contrast of rainfall between the two study
339 periods, our study contributes to better understanding disease dynamics in a natural system.

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356 **References**

- 357 1. Johnson MD. 2007 Measuring habitat quality: a review. *Condor* **109**(3), 489-504.
358 (doi:10.1093/condor/109.3.489)
- 359 2. Skagen SK, Knopf FL. 1994 Migrating shorebirds and habitat dynamics at a prairie wetland
360 complex. *Wilson Bull.*, 91-105.

- 361 3. Nielsen SE, McDermid G, Stenhouse GB, Boyce MS. 2010 Dynamic wildlife habitat
362 models: seasonal foods and mortality risk predict occupancy-abundance and habitat
363 selection in grizzly bears. *Biol. Conserv.* **143**(7), 1623-1634.
364 (doi:10.1016/j.biocon.2010.04.007)
- 365 4. Betts MG, Rodenhouse NL, Scott Sillett T, Doran PJ, Holmes RT. 2008 Dynamic occupancy
366 models reveal within-breeding season movement up a habitat quality gradient by a migratory
367 songbird. *Ecography* **31**(5), 592-600. (doi:10.1111/j.0906-7590.2008.05490.x)
- 368 5. McGarigal K, Wan HY, Zeller KA, Timm BC, Cushman SA. 2016 Multi-scale habitat
369 selection modeling: a review and outlook. *Landsc. Ecol.* **31**(6), 1161-1175.
370 (doi:10.1007/s10980-016-0374-x)
- 371 6. Murray KA, Retallick RW, Puschendorf R, Skerratt LF, Rosauer D, McCallum HI, Berger L,
372 Speare R, VanDerWal J. 2011 Assessing spatial patterns of disease risk to biodiversity:
373 implications for the management of the amphibian pathogen, *Batrachochytrium*
374 *dendrobatidis*. *J. Appl. Ecol.* **48**(1), 163-173. (doi:10.1111/j.1365-2664.2010.01890.x)
- 375 7. Ostfeld RS, Glass GE, Keesing F. 2005 Spatial epidemiology: an emerging (or re-emerging)
376 discipline. *Trends Ecol. Evol.* **20**(6), 328-336. (doi:10.1016/j.tree.2005.03.009)
- 377 8. Lambin EF, Tran A, Vanwambeke SO, Linard C, Soti V. 2010 Pathogenic landscapes:
378 interactions between land, people, disease vectors, and their animal hosts. *Int. J. Health*

- 379 *Geogr.* **9**(1), 54. (doi:10.1186/1476-072X-9-54)
- 380 9. Peterson AT. 2006 Ecologic niche modeling and spatial patterns of disease transmission.
- 381 *Emerg. Infect. Dis.* **12**(12), 1822. (doi:10.3201/eid1212.060373)
- 382 10. Folstad I, Nilssen AC, Halvorsen O, Andersen J. 1991 Parasite avoidance: the cause of post-
- 383 calving migrations in *Rangifer*? *Can. J. Zool.* **69**(9), 2423-2429. (doi:10.1139/z91-340)
- 384 11. Altizer S, Bartel R, Han BA. 2011 Animal migration and infectious disease risk. *Science*
- 385 **331**(6015), 296-302. (doi:10.1126/science.1194694)
- 386 12. Dougherty ER, Seidel DP, Carlson CJ, Spiegel O, Getz WM. 2018 Going through the
- 387 motions: incorporating movement analyses into disease research. *Ecol. Lett.* **21**(4), 588-604.
- 388 (doi:10.1111/ele.12917)
- 389 13. Leach CB, Webb CT, Cross PC. 2016 When environmentally persistent pathogens transform
- 390 good habitat into ecological traps. *R. Soc. Open Sci.* **3**(3), 160051.
- 391 (doi:10.1098/rsos.160051)
- 392 14. Hopkins SR, Hoyt JR, White JP, Kaarakka HM, Redell JA, DePue JE, Scullon WH,
- 393 Kilpatrick AM, Langwig KE. 2021 Continued preference for suboptimal habitat reduces bat
- 394 survival with white-nose syndrome. *Nat. Commun.* **12**(1), 1-9. (doi: 10.1038/s41467-020-
- 395 20416-5)
- 396 15. Battin J. 2004 When good animals love bad habitats: ecological traps and the conservation

397 of animal populations. *Conserv. Biol.* **18**(6), 1482-1491. (doi:10.1111/j.1523-

398 1739.2004.00417.x)

399 16. Hale R, Swearer SE. 2016 Ecological traps: current evidence and future directions. *Proc. R.*

400 *Soc. B* **283**(1824), 20152647. (doi:10.1098/rspb.2015.2647)

401 17. Robertson BA, Hutto RL. 2006 A framework for understanding ecological traps and an

402 evaluation of existing evidence. *Ecology* **87**(5), 1075-1085. (doi:10.1890/0012-

403 9658(2006)87[1075:Affuet]2.0.Co;2)

404 18. Patten MA, Kelly JF. 2010 Habitat selection and the perceptual trap. *Ecol. Appl.* **20**(8),

405 2148-2156. (doi:10.1890/09-2370.1)

406 19. Gasaway WC, Gasaway KT, Berry HH. 1996 Persistent low densities of plains ungulates in

407 Etosha National Park, Namibia: testing the food-regulating hypothesis. *Can. J. Zool.* **74**(8),

408 1556-1572. (doi:10.1139/z96-170)

409 20. Turnbull P. (ed.). 2008 *Anthrax in humans and animals*, 4th edn. Geneva, Switzerland: World

410 Health Organization.

411 21. Turner WC, Kausrud KL, Beyer W, Easterday WR, Barandongo ZR, Blaschke E, Cloete CC,

412 Lazak J, Van Ert MN, Ganz HH, et al. 2016 Lethal exposure: An integrated approach to

413 pathogen transmission via environmental reservoirs. *Sci. Rep.* **6**, 27311.

414 (doi:10.1038/srep27311)

- 415 22. Turner WC, Kausrud KL, Krishnappa YS, Crooms JP, Ganz HH, Mapaire I, Cloete CC,
416 Havarua Z, Kusters M, Getz WM, et al. 2014 Fatal attraction: vegetation responses to
417 nutrient inputs attract herbivores to infectious anthrax carcass sites. *Proc. R. Soc. B*
418 **281**(1795). (doi:10.1098/rspb.2014.1785)
- 419 23. Bellan SE, Turnbull PC, Beyer W, Getz WM. 2013 Effects of experimental exclusion of
420 scavengers from carcasses of anthrax-infected herbivores on *Bacillus anthracis* sporulation,
421 survival, and distribution. *Appl. Environ. Microbiol.* **79**(12), 3756-3761.
422 (doi:10.1128/AEM.00181-13)
- 423 24. Getz WM. 2011 Biomass transformation webs provide a unified approach to consumer–
424 resource modelling. *Ecol. Lett.* **14**(2), 113-124. (doi:10.1111/j.1461-0248.2010.01566.x)
- 425 25. Engert S. 1997 Spatial variability and temporal periodicity of rainfall in the Etosha National
426 Park and surrounding areas in northern Namibia. *Madoqua* **20**(1), 115-120.
- 427 26. Shikangalah RN. 2020 The 2019 drought in Namibia: An overview. *J. Namibian Stud.* **27**,
428 37-58.
- 429 27. Tsalyuk M, Kelly M, Getz WM. 2017 Improving the prediction of African savanna
430 vegetation variables using time series of MODIS products. *ISPRS J. Photogramm. Remote*
431 *Sens.* **131**, 77-91. (doi:10.1016/j.isprsjprs.2017.07.012)
- 432 28. le Roux C, Grunow J, Morris J, Bredenkamp G, Scheepers J. 1988 A classification of the

- 433 vegetation of the Etosha National Park. *S. Afr. J. Bot.* **54**, 1-10. (doi:10.1016/S0254-
434 6299(16)31355-2)
- 435 29. Turner WC, Imologhome P, Havarua Z, Kaaya GP, Mfunne JK, Mpofu ID, Getz WM. 2013
436 Soil ingestion, nutrition and the seasonality of anthrax in herbivores of Etosha National
437 Park. *Ecosphere* **4**(1), 1-19. (doi:10.1890/ES12-00245.1)
- 438 30. Hugh-Jones ME, de Vos V. 2002 Anthrax and wildlife. *Rev. Sci. Tech.* **21**(2), 359-383.
439 (doi:10.20506/rst.21.2.1336)
- 440 31. Lindeque PM, Turnbull PC. 1994 Ecology and epidemiology of anthrax in the Etosha
441 National Park, Namibia. *Onderstepoort J. Vet. Res.* **61**(1), 71-83.
- 442 32. Bellan SE, Gimenez O, Choquet R, Getz WM. 2013 A hierarchical distance sampling
443 approach to estimating mortality rates from opportunistic carcass surveillance data. *Methods*
444 *Ecol. Evol.* **4**(4), 361-369. (doi:10.1111/2041-210x.12021)
- 445 33. Huso MM. 2011 An estimator of wildlife fatality from observed carcasses. *Environmetrics*
446 **22**(3), 318-329. (doi:10.1002/env.1052)
- 447 34. Spiegel O, Getz WM, Nathan R. 2013 Factors influencing foraging search efficiency: why
448 do scarce lappet-faced vultures outperform ubiquitous white-backed vultures? *Am. Nat.*
449 **181**(5), E102-E115. (doi:10.1086/670009)
- 450 35. Borchering RK, Bellan SE, Flynn JM, Pulliam JRC, McKinley SA. 2017 Resource-driven

451 encounters among consumers and implications for the spread of infectious disease. *J. R. Soc.*
452 *Interface* **14**(135). (doi:10.1098/rsif.2017.0555)

453 36. Faustino CES. 2020 Movement ecology and conservation: the case of African vultures.
454 Doctoral thesis, University of St Andrews, St. Andrews, UK.

455 37. Myneni R, Knyazikhin Y, Park T. 2015 MOD15A2H MODIS/terra leaf area index/FPAR 8-
456 day L4 global 500 m SIN grid V006. *NASA EOSDIS Land Processes DAAC*.

457 38. Machwitz M, Gessner U, Conrad C, Falk U, Richters J, Dech S. 2015 Modelling the gross
458 primary productivity of west Africa with the regional biomass model RBM+, using
459 optimized 250 m MODIS FPAR and fractional vegetation cover information. *Int. J. Appl.*
460 *Earth Obs. Geoinf.* **43**, 177-194. (doi:10.1016/j.jag.2015.04.007)

461 39. Majasalmi T, Rautiainen M, Stenberg P. 2014 Modeled and measured fPAR in a boreal
462 forest: Validation and application of a new model. *Agric. For. Meteorol.* **189**, 118-124.
463 (doi:10.1016/j.agrformet.2014.01.015)

464 40. Havarua Z, Turner WC, Mfuno JK. 2014 Seasonal variation in foraging behaviour of plains
465 zebra (*Equus quagga*) may alter contact with the anthrax bacterium (*Bacillus anthracis*).
466 *Can. J. Zool.* **92**(4), 331-337. (doi:10.1139/cjz-2013-0186)

467 41. Johnson CJ, Nielsen SE, Merrill EH, McDonald TL, Boyce MS. 2006 Resource selection
468 functions based on use-availability data: theoretical motivation and evaluation methods. *J.*

469 *Wildl. Manage.* **70**(2), 347-357. (doi:10.2193/0022-541X(2006)70[347:RSFBOU]2.0.CO;2)

470 42. Johnson DH. 1980 The comparison of usage and availability measurements for evaluating
 471 resource preference. *Ecology* **61**(1), 65-71. (doi:10.2307/1937156)

472 43. Fieberg J, Matthiopoulos J, Hebblewhite M, Boyce MS, Frair JL. 2010 Correlation and
 473 studies of habitat selection: problem, red herring or opportunity? *Philos. Trans. R. Soc. B*
 474 **365**(1550), 2233-2244. (doi:10.1098/rstb.2010.0079)

475 44. Avgar T, Lele SR, Keim JL, Boyce MS. 2017 Relative selection strength: quantifying effect
 476 size in habitat-and step-selection inference. *Ecol. Evol.* **7**(14), 5322-5330.
 477 (doi:10.1002/ece3.3122)

478 45. Zidon R, Garti S, Getz WM, Saltz D. 2017 Zebra migration strategies and anthrax in Etosha
 479 National Park, Namibia. *Ecosphere* **8**(8). (doi:10.1002/ecs2.1925)

480 46. R Core Team. 2020 R: a language and environment for statistical computing. R Foundation
 481 for Statistical Computing, Vienna, Austria.

482 47. Busetto L, Raghetti L. 2016 MODISsp: An R package for automatic preprocessing of
 483 MODIS Land Products time series. *Comput. Geosci.* **97**, 40-48.
 484 (doi:10.1016/j.cageo.2016.08.020)

485 48. Hijmans RJ. 2020 raster: geographic data analysis and modeling. R package version 3.3-13.

486 49. Calenge C. 2006 The package “adehabitat” for the R software: a tool for the analysis of

487 space and habitat use by animals. *Ecol. Modell.* **197**, 516-519
 488 (doi:10.1016/j.ecolmodel.2006.03.017)

489 50. Bivand R, Rundel C. 2020 rgeos: interface to geometry engine - open source (GEOS). R
 490 package version 0.5-3.

491 51. Pebesma E, Bivand RS. 2018 Simple features for R: standardized support for spatial vector
 492 data. *R J.* **10**, 439-446. (doi:10.32614/RJ-2018-009)

493 52. Pebesma EJ, Bivand RS. 2005 Classes and methods for spatial data in R. *R News* **5**, 9-13.

494 53. Bivand RS, Pebesma E, Gomez-Rubio V. 2013 *Applied spatial data analysis with R*, 2nd edn.
 495 Springer, NY.

496 54. Cizauskas CA, Bellan SE, Turner WC, Vance RE, Getz WM. 2014 Frequent and seasonally
 497 variable sublethal anthrax infections are accompanied by short-lived immunity in an
 498 endemic system. *J. Anim. Ecol.* **83**(5), 1078-1090. (doi:10.1111/1365-2656.12207)

499 55. Easterday WR, Ponciano JM, Gomez JP, Van Ert MN, Hadfield T, Bagamian K, Blackburn
 500 JK, Stenseth NC, Turner WC. 2020 Coalescence modeling of intrainfection *Bacillus*
 501 *anthracis* populations allows estimation of infection parameters in wild populations. *Proc.*
 502 *Natl. Acad. Sci.* **117**(8), 4273-4280. (doi:10.1073/pnas.1920790117)

503 56. Abraham JO, Hempson GP, Staver AC. 2019 Drought-response strategies of savanna
 504 herbivores. *Ecol. Evol.* **9**(12), 7047-7056. (doi:10.1002/ece3.5270)

- 505 57. Cizauskas CA, Turner WC, Pitts N, Getz WM. 2015 Seasonal patterns of hormones,
506 macroparasites, and microparasites in wild African ungulates: the interplay among stress,
507 reproduction, and disease. *PLoS One* **10**(4), e0120800. (doi:10.1371/journal.pone.0120800)
- 508 58. Muturi M, Gachohi J, Mwatondo A, Lekolool I, Gakuya F, Bett A, Osoro E, Bitek A,
509 Thumbi SM, Munyua P. 2018 Recurrent anthrax outbreaks in humans, livestock, and
510 wildlife in the same locality, Kenya, 2014–2017. *Am. J. Trop. Med. Hyg.* **99**(4), 833-839.
511 (doi:10.4269/ajtmh.18-0224)
- 512 59. Clegg SB, Turnbull P, Foggin C, Lindeque P. 2007 Massive outbreak of anthrax in wildlife
513 in the Malilangwe Wildlife Reserve, Zimbabwe. *Vet. Rec.* **160**(4), 113-118.
514 (doi:10.1136/vr.160.4.113)
- 515 60. de Vos V, Bryden H. 1996 Anthrax in the Kruger National Park: temporal and spatial
516 patterns of disease occurrence. *Salisbury Med. Bull.* **87**, 26-31.
- 517 61. Farnsworth ML, Hoeting JA, Hobbs NT, Miller MW. 2006 Linking chronic wasting disease
518 to mule deer movement scales: a hierarchical Bayesian approach. *Ecol. Appl.* **16**(3), 1026-
519 1036. (doi:10.1890/1051-0761(2006)016[1026:Lcwdtm]2.0.Co;2)
- 520 62. Fuller TL, Saatchi SS, Curd EE, Toffelmier E, Thomassen HA, Buermann W, DeSante DF,
521 Nott MP, Saracco JF, Ralph C. 2010 Mapping the risk of avian influenza in wild birds in the
522 US. *BMC Infect. Dis.* **10**(1), 1-13. (doi:https://doi.org/10.1186/1471-2334-10-187)

523 63. Lilley TM, Anttila J, Ruokolainen L. 2018 Landscape structure and ecology influence the
524 spread of a bat fungal disease. *Funct. Ecol.* **32**(11), 2483-2496. (doi:10.1111/1365-
525 2435.13183)

Figures

Figure 1. The study area of Etosha National Park, Namibia in southern Africa. a) The distribution of habitat types. The four habitat types considered included open plains (light grey), shrublands (medium grey), woodlands (dark grey), and bare areas (large salt pans; tan). White represents areas with vegetation type not available, which were removed from this study. The blue circle indicates the location of Okaukuejo station. b) Plains zebra (*Equus quagga*) space use shows the overall thinned readings of zebra telemetry data (yellow) and their 99.9% kernel density range (blue) which were used to generate available (not-used) points for analysis of resource selection functions. c) The percentages of the available points randomly generated within 99.9% kernel density range by habitat type.

Figure 2. Herbivore mortalities by habitat type in Etosha National Park. Mortalities from 1998–April 2020 with GPS locations were assigned to habitat type (open plains, shrublands or woodlands) and cause of death (anthrax or other natural mortalities) for herbivore species which had at least one anthrax mortality. The mortalities are summed for a) all herbivore species, and then grouped by three functional foraging types, b) grazers, including plains zebra (67.6% of total cases), blue wildebeest (*Connochaetes taurinus*; 9.9%) and gemsbok (*Oryx gazella*; 1.1%), c) mixed feeders including springbok (*Antidorcas marsupialis*; 12.3% of total cases) and African elephant (*Loxodonta africana*; 8.5%), and d) browsers including greater kudu (*Tragelaphus*

544 *strepsiceros*; 0.3% of total cases) and black rhino (*Diceros bicornis*; 0.4%). The numbers above
545 bars indicate ratios of anthrax mortalities to other natural causes of death. The asterisks show the
546 significance of chi-square tests comparing the proportions of anthrax mortalities between paired
547 habitat types. One, two and three asterisks represent the Bonferroni corrected p values less than
548 0.05, 0.01 and 0.001, respectively.

549 **Figure 3.** Zebra anthrax mortalities (a, b) and vegetation productivity (c, d) in Etosha National
550 Park, by habitat type and season in 2009–2010 (a, c) and 2018–2020 (b, d). Productivity was
551 measured as the fraction of photosynthetically active radiation (FPAR). Error bars indicate
552 standard deviations of spatial variation. The axes for anthrax mortalities were square-root
553 transformed. The study period 2009–2010 was an average rainfall year; 2018–2020, a severe
554 drought followed by an above average rainfall year.

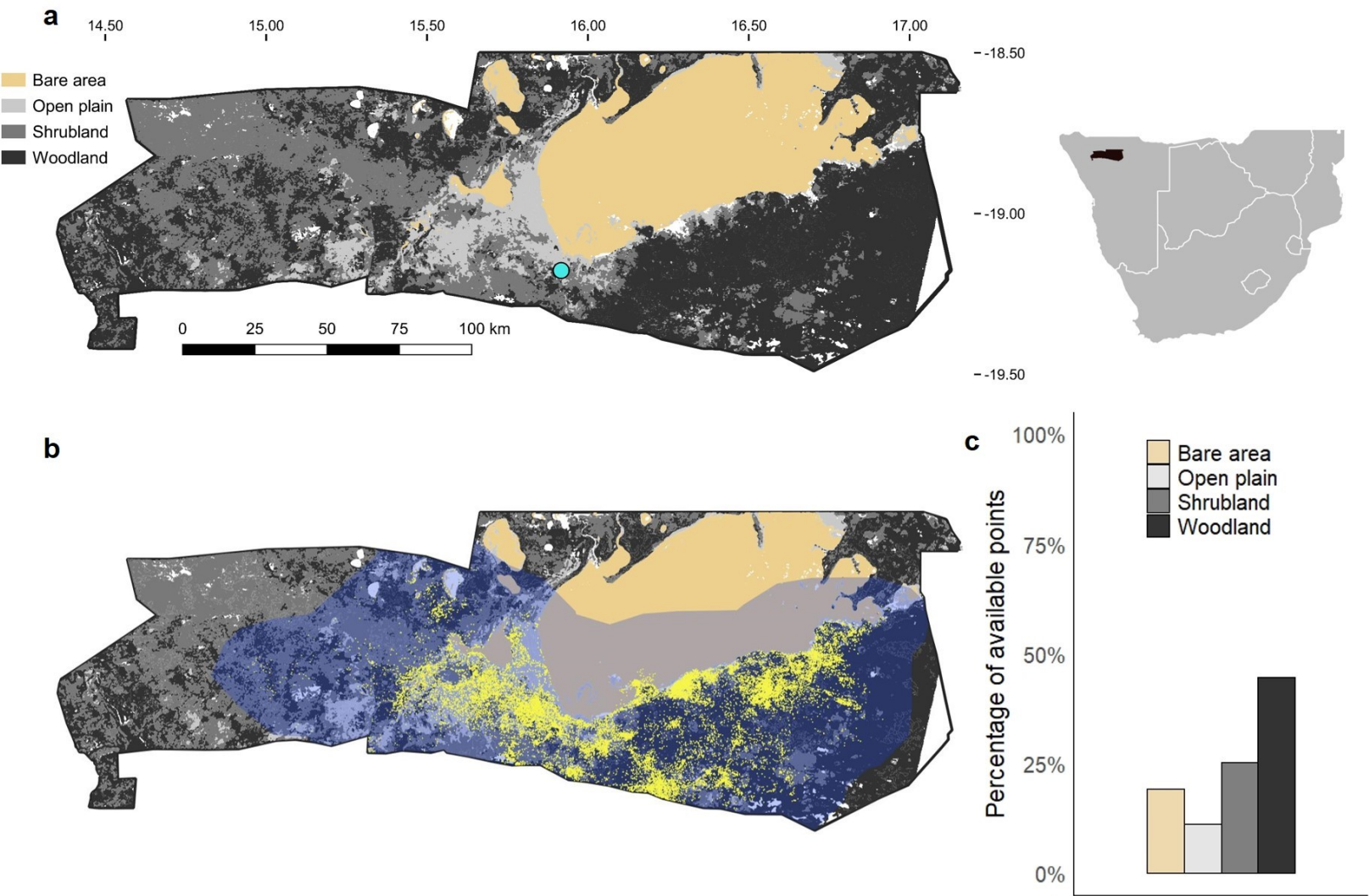
555 **Figure 4.** Inter-annual and seasonal variation in zebra habitat selection in Etosha National Park.
556 a) and b) show the average percentages of zebra selected locations among habitat types by
557 season a) in 2009 – 2010 (an average rainfall year) and b) in 2018 – 2020 (a drought year, 2019,
558 followed by an above average rainfall year, 2020), with error bars representing standard
559 deviations from individual differences. c) to h) show the regression coefficients indicating zebra
560 habitat selection accounting for habitat availability by season and year. The coefficients of three
561 contrast coding variables from seasonal individual-based resource selection functions

represented the relative selection strength (habitat preference) for vegetated habitats over bare areas c) in 2009 – 2010 and d) in 2018 – 2020, for open habitats over closed habitats e) in 2009 – 2010 and f) in 2018 – 2020, and for shrublands over woodlands g) in 2009 – 2010 and h) in 2018 – 2020. Each point is an individual zebra, with males in grey and females in yellow. In 2009 – 2010, all the collared zebras were female. Sample sizes of individual zebras per season ranged from 5 to 17. The boxplots in c) - h) are colour-coded with the seasonal average values of FPAR at open habitats reflecting vegetation productivity differences by season, with grey to purple to blue representing low to high FPAR.

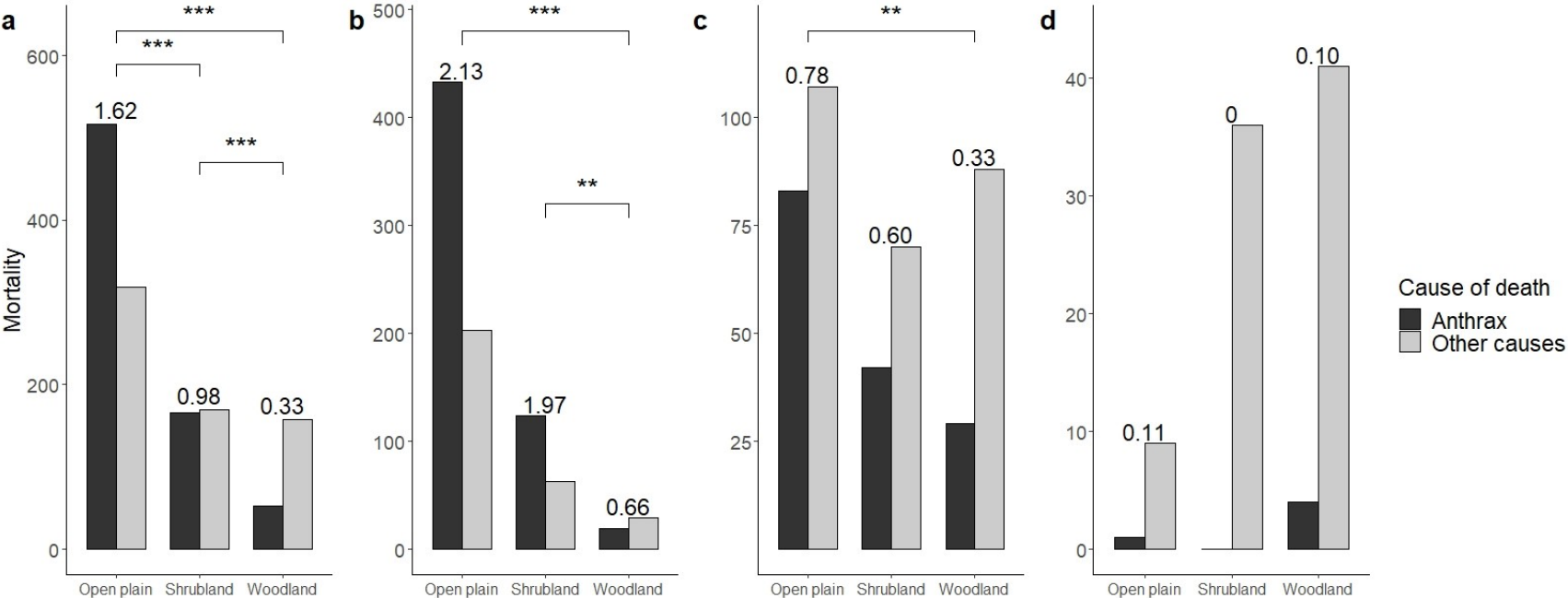
Figure 5. Seasonal relationships between habitat dynamics and zebra habitat selection. Medians of relative habitat selection strengths in relation to average FPAR (a remote-sensing index of vegetation productivity) at open habitats a) for vegetated habitats over bare areas, b) for open habitats over closed habitats, and c) for shrublands over woodlands. An error bar indicates interquartile range from individual variation in each season. Grey dashed lines are best-fitting lines when linear regressions showed significant slopes. Different years/seasons were colour-/shape-coded.

Figure 6. Linear regressions between habitat dynamics, zebra habitat selection, and zebra anthrax cases. Relationships are shown between the square-root transformed anthrax case numbers by season in relation to a) average FPAR (a remote-sensing index of vegetation

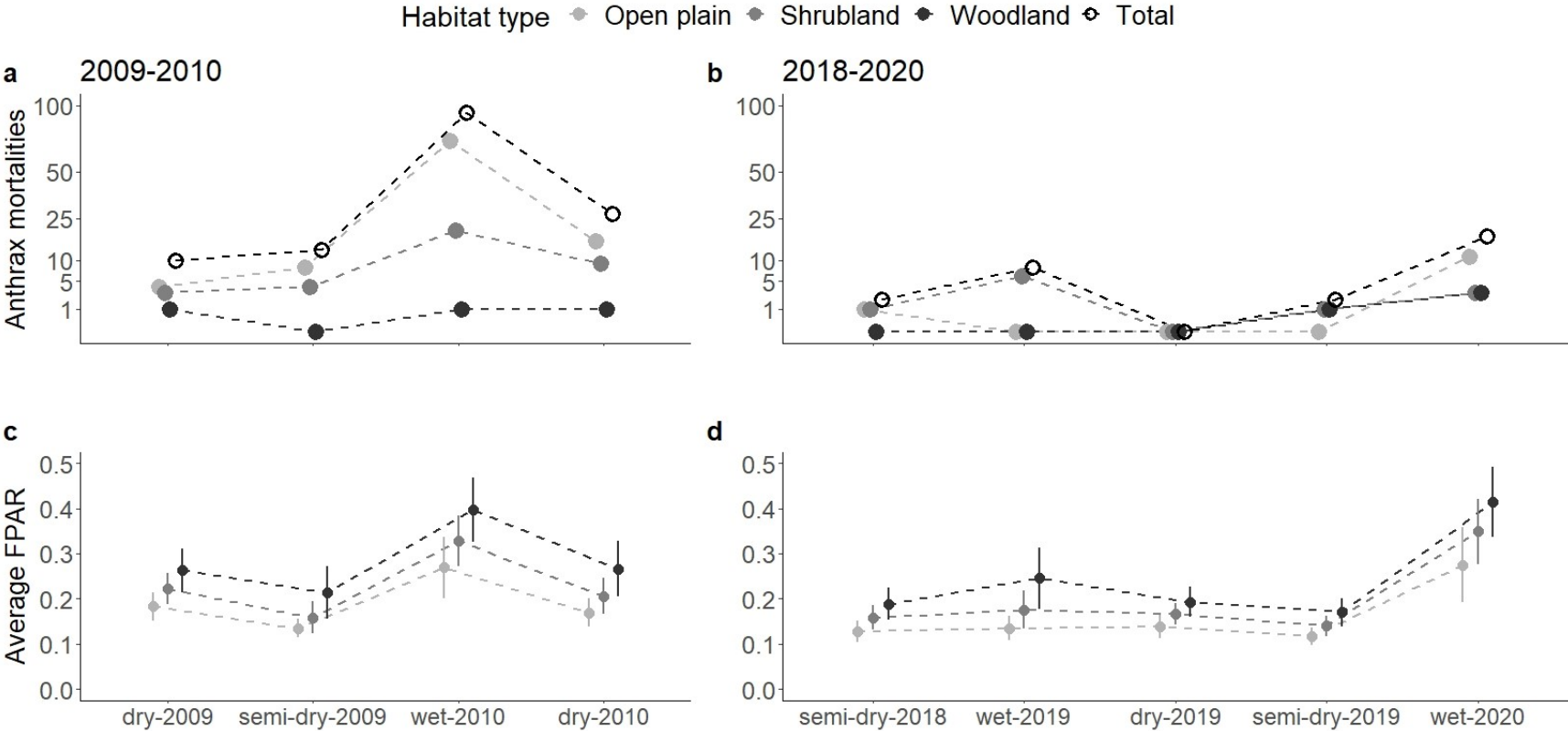
580 productivity) at open habitats, and b) the median of relative selection strengths for open habitats
581 over closed habitats. Error bars indicate interquartile ranges of relative selection strengths from
582 individual variation. Grey dashed lines are best-fitting lines from linear regressions. Different
583 years/seasons were colour-/shape-coded.



586 **Figure 2**

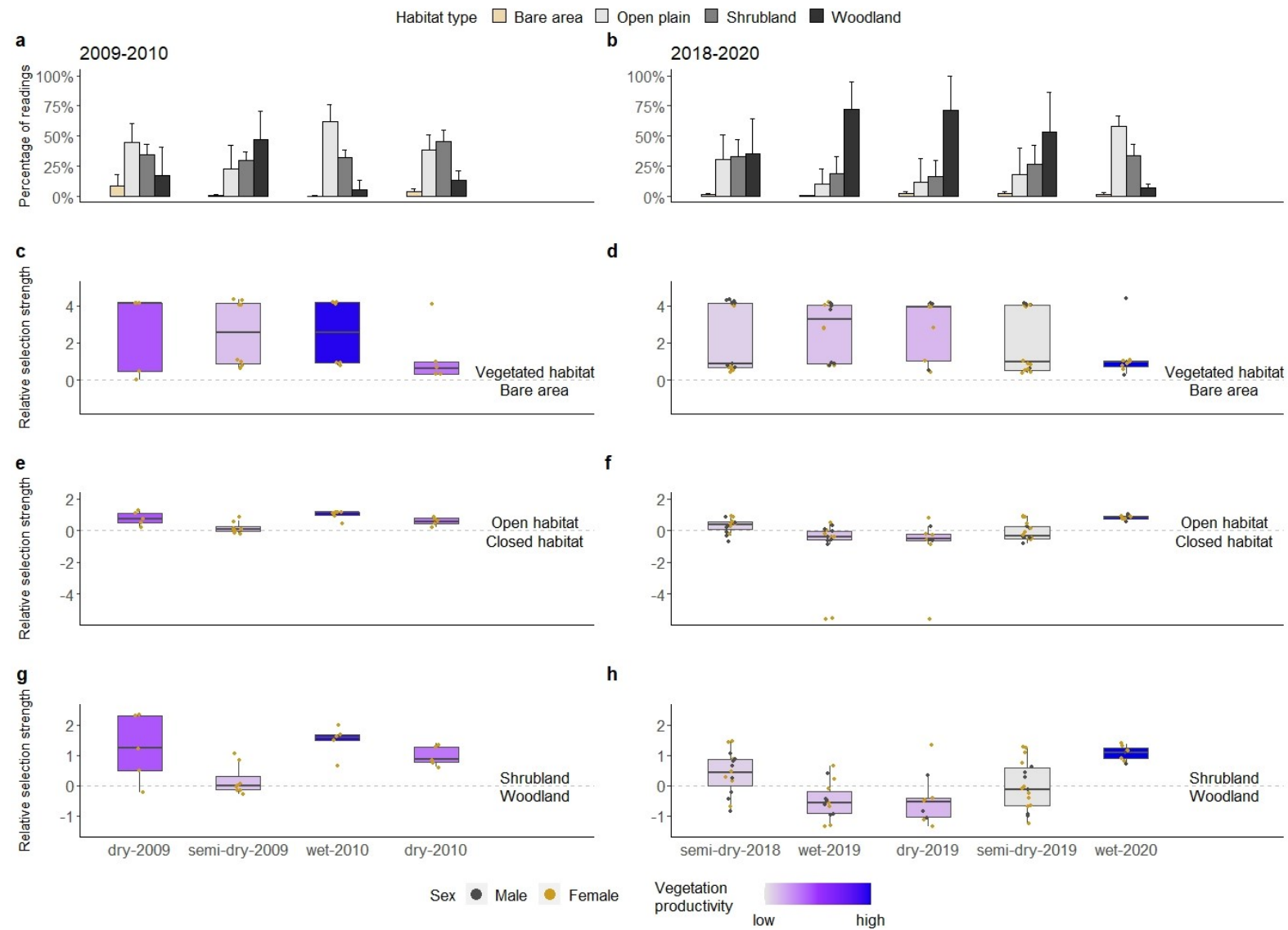


588 **Figure 3**



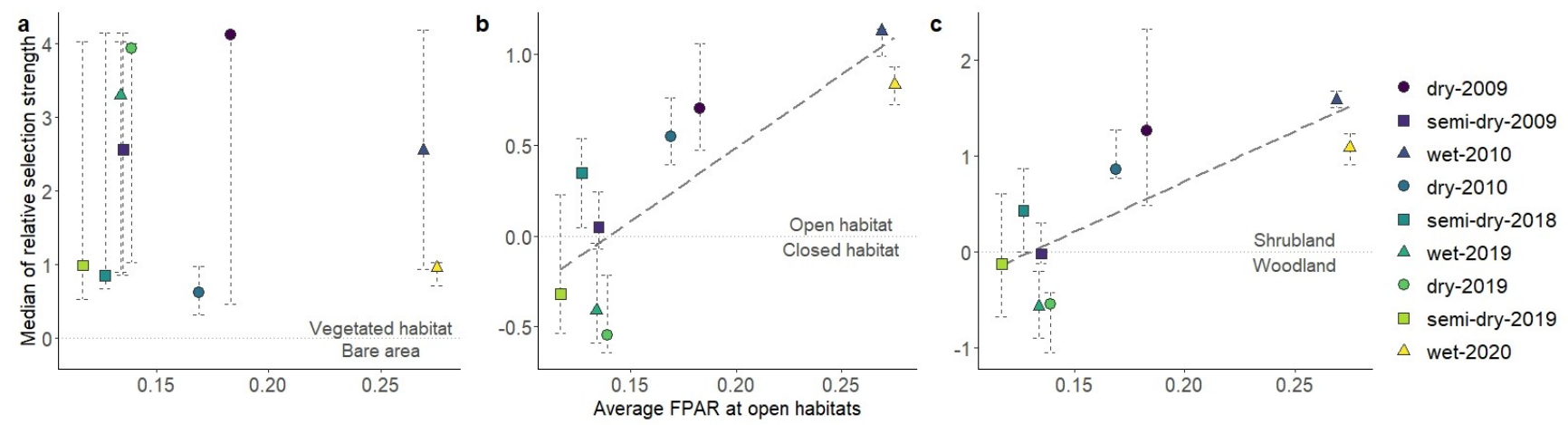
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590 **Figure 4**



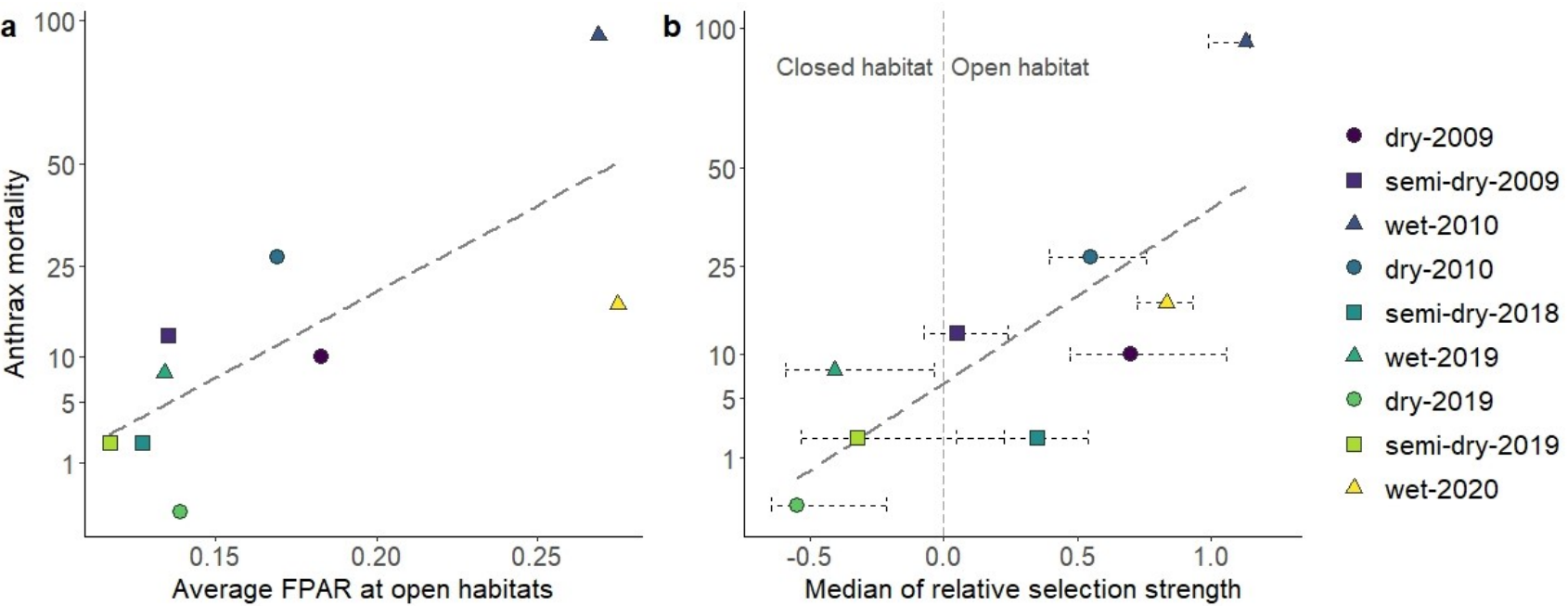
592 **Figure 5**

593



594 **Figure 6**

595



Electronic Supplementary Material

Disease or drought: environmental fluctuations release zebra from a potential pathogen-triggered ecological trap

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Supplementary Materials and Methods

Supplementary References

Supplementary Figures 1-5

Supplementary Materials and Methods

Consistency of habitat selection with different thinning methods

The thinning method we used for plains zebra (*Equus quagga*) telemetry data in the study from Etosha National Park, Namibia (ENP) divided days into morning (06:00-12:00; GMT+1), afternoon (12:00-18:00) and night (18:00-06:00) and extracted readings closest to 9:00, 15:00 and 24:00 for the three periods of a day for each individual. To test whether the results using this thinning method were robust, we compared the results of these resource selection functions (RSFs) to the results of RSFs using fixed intervals thinned to 1, 2, 4, 8, 12 or 24 hours. We only compared the thinning methods using telemetry data from 2009 – 2010, which had a higher frequency of sampling than data in 2018 – 2020. Package `amt` was used to thin the data with fixed intervals [1], with tolerance of 10 minutes. The methods for RSFs then proceeded as described in the main methods section. Results are shown in figure S4.

Probability that an individual infected within open habitats would die within the same habitat

Zebra in ENP can move long distances within a day (see figure S3). We thus investigated how consistently zebra remain in open habitats over time, to evaluate how likely a zebra would be to die in the open habitat if infected there, and how that

varies by season. Though the collared zebra did not die of anthrax, we used non-infected movement patterns to investigate the probability of mortality, assuming the movement patterns do not differ between healthy and sick individuals. A sick individual might be expected to move less, but movement trajectory indices of hippopotamus (*Hippopotamus amphibius*) in Tanzania did not show differences before and after anthrax infection [2]. Because an anthrax-infected animal can die within a few days [3], higher sampling intensity provides a better estimation for short-term movements. We thus used one-hour interval telemetry data from 2009 – 2010, and set every time point when one individual was using open habitats as a start, regardless of where the individual had been previously. We then calculated the average probability that an individual was at open habitats every one hour since the start, up to a time period of ten days, which covers the expected incubation period. These results are shown in figure S5, and indicate that zebras had higher probability remaining in open habitats in the wet/anthrax season.

Supplementary References

1. Signer J, Fieberg J, Avgar T. 2019 Animal movement tools (amt): R package for managing tracking data and conducting habitat selection analyses. *Ecol. Evol.* **9**(2), 880-890. (doi:10.1002/ece3.4823)
2. Stears K, Schmitt MH, Turner WC, McCauley DJ, Muse EA, Kiwango H, Matheyo D, Mutayoba BM. In press. Hippopotamus movements structure the spatiotemporal dynamics of an active anthrax outbreak. *Ecosphere*.
3. Easterday WR, Ponciano JM, Gomez JP, Van Ert MN, Hadfield T, Bagamian K, Blackburn JK, Stenseth NC, Turner WC. 2020 Coalescence modeling of intrainfection *Bacillus anthracis* populations allows estimation of infection parameters in wild populations. *Proc. Natl. Acad. Sci.* **117**(8), 4273-4280. (doi:10.1073/pnas.1920790117)

Supplementary Figures

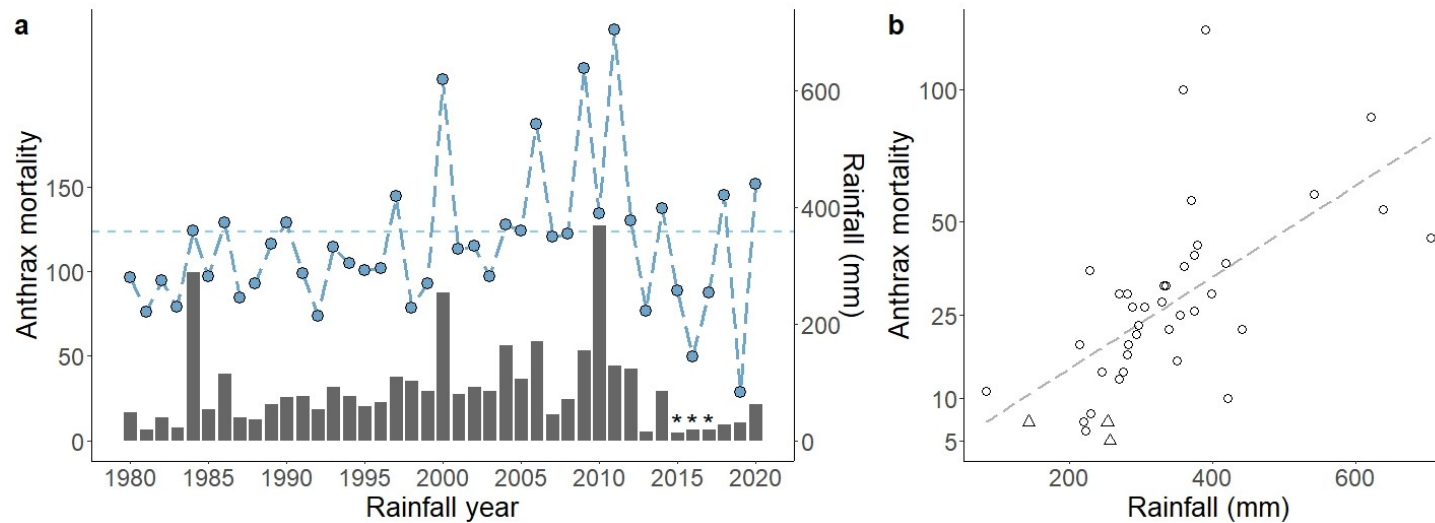


Figure S1.

Anthrax mortalities of plains zebra (*Equus quagga*) and annual rainfall in Etosha National Park. a) Patterns of rainfall and zebra anthrax mortality from 1980 – 2020. The number of zebra anthrax mortalities are shown in grey bars, and blue points and lines are the annual rainfall from Okaukuejo station from rainfall years 1980 to 2020, and the dashed horizontal blue line is average annual rainfall (358.0mm; 1954 – 2020). A rainfall year is from July to June (e.g., July 2009 – June 2010 is the rainfall year 2010). b) The correlation between annual rainfall and zebra anthrax mortalities. The y axis of plot b is square-root transformed due to overdispersion. *Mortality surveillance in years annotated with stars in plot a and shown with triangle shapes in plot b may have been dampened due to staff surveillance effort diverted to rhino protection efforts during a spike in poaching intensity.

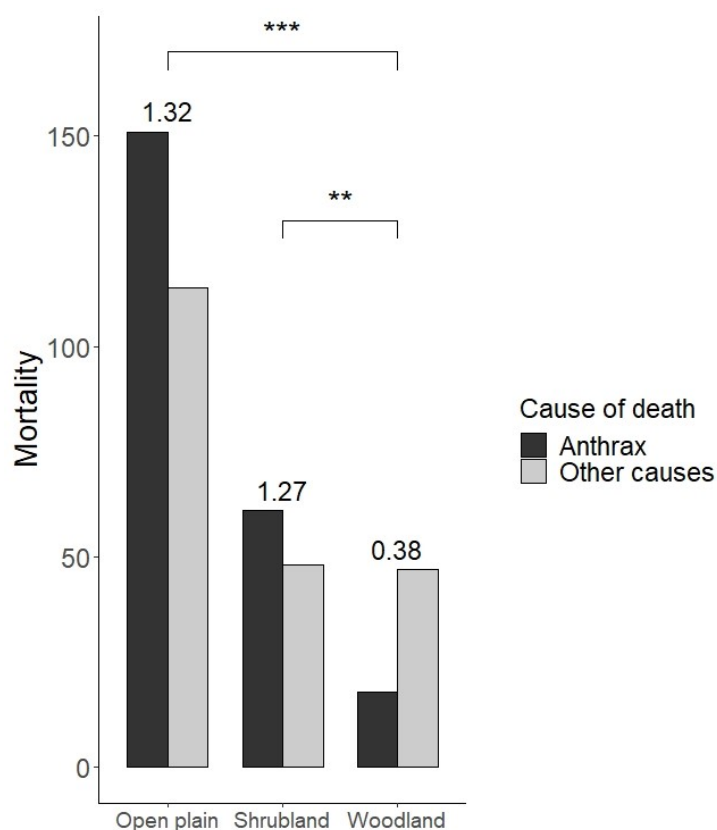


Figure S2.

The numbers of anthrax and non-anthrax natural mortalities during periods of heightened surveillance effort in Etosha National Park. Mortalities are those with global positioning system (GPS) positions collected during the seasons involved in the study periods (May 2009 – August 2010; September 2018 – April 2020), when opportunistic mortality surveillance was supplemented with surveillance efforts from visiting clustered positions of GPS-tagged scavengers. The species represented here are herbivore species with at least one anthrax case between 1998 – 2020 (see figure 2) including plains zebra, blue wildebeest (*Connochaetes taurinus*), gemsbok (*Oryx gazella*), springbok (*Antidorcas marsupialis*), African elephant (*Loxodonta africana*), greater kudu (*Tragelaphus strepsiceros*) and black rhino (*Diceros bicornis*). The numbers above bars indicate ratios of anthrax mortalities to other natural causes of death. The asterisks show the significance of chi-square tests comparing the proportions of anthrax mortalities between paired habitat types. One, two and three asterisks represent the Bonferroni corrected p values less than 0.05, 0.01 and 0.001, respectively.

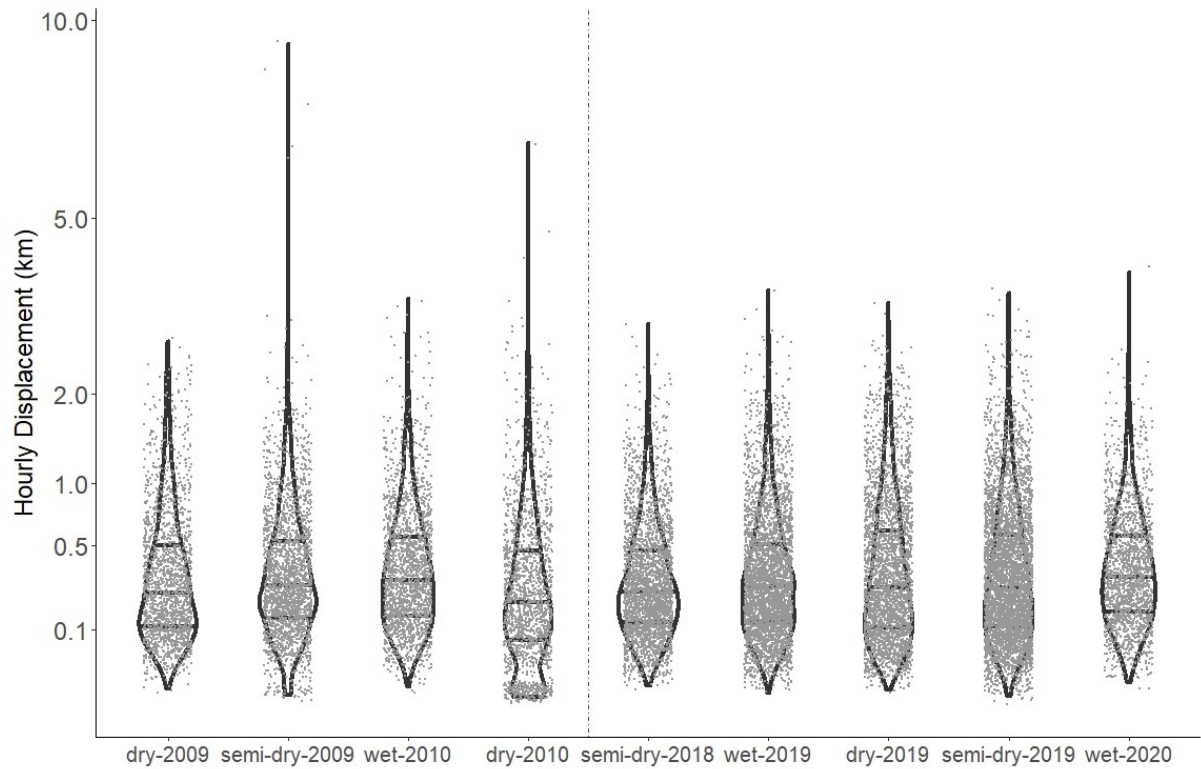


Figure S3.

Spatial displacement between locations of two consecutive time points for plains zebra individuals by season from the telemetry data used for habitat selection analysis in Etosha National Park. Because of different sampling intervals, the displacement was standardized to one-hour interval (velocity per hour). The y axis is square-root transformed due to the high skewness. Horizontal lines from low to high in the violin plots represent 25%, 50% and 75% quantiles.

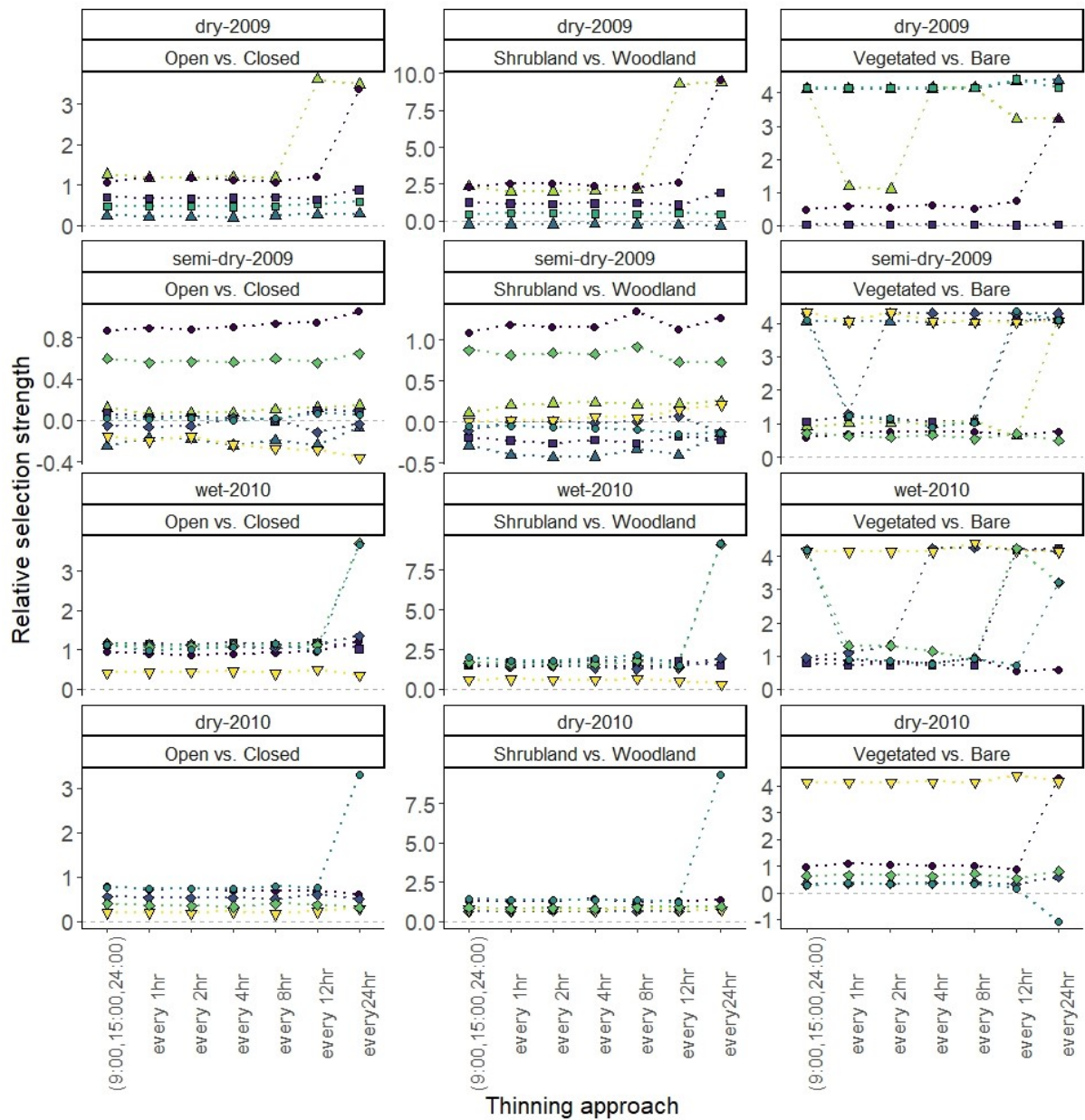


Figure S4.

The relative selection strength of resource selection functions between habitat types calculated from plains zebra telemetry data with different thinning approaches by season in 2009 – 2010. Thinning methods included extracting readings closest to 9:00, 15:00 and 24:00 (the method used in the study; first points) and fixed intervals of 1, 2, 4, 8, 12 and 24 hours. Each shape/colour of point indicates an individual zebra connected with lines, and these shapes/colours are consistent among plots.

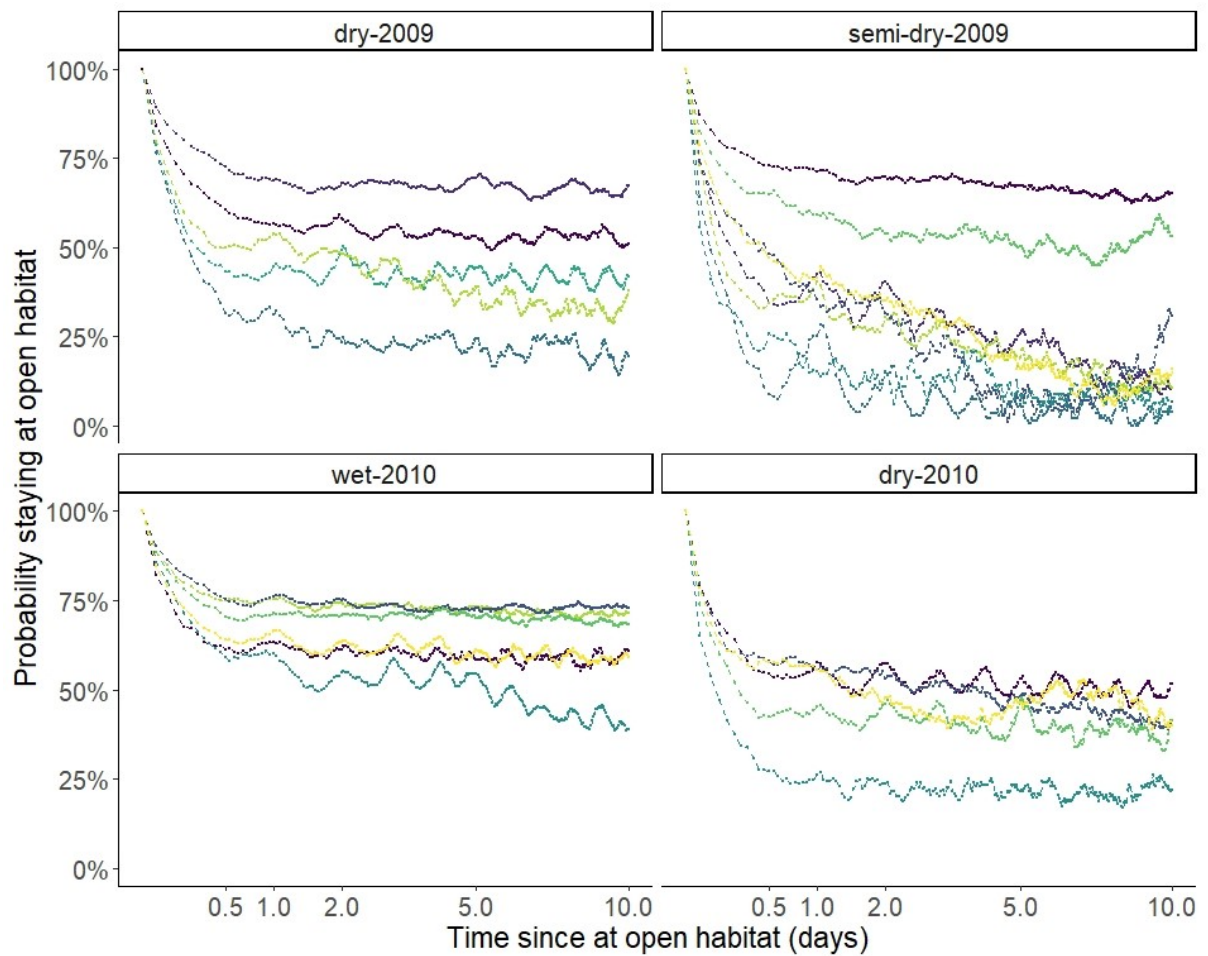


Figure S5.

The average probability that a plains zebra individual in an open habitat remains in the open habitat over time, regardless of where the individual had previously been. These probabilities were investigated by season from zebra movement data in 2009 – 2010 from Etosha National Park, to evaluate how likely a zebra that is exposed to *Bacillus anthracis* in an open habitat would be to die in that same habitat. Coloured lines indicate individual zebras and are standardized across plots. The x axis is square-root transformed to better show the first few days, because the incubation period of anthrax can be very short [3].