

Sex Steroids as Regulators of Gestural Communication

Daniel J. Tobiansky¹ and Matthew J. Fuxjager¹

¹Department of Ecology and Evolutionary Biology, Brown University, Providence, Rhode Island 02912

ORCID numbers: 0000-0001-6381-3453 (D. J. Tobiansky); 0000-0003-0591-6854 (M. J. Fuxjager).

Gestural communication is ubiquitous throughout the animal kingdom, occurring in species that range from humans to arthropods. Individuals produce gestural signals when their nervous system triggers the production of limb and body movement, which in turn functions to help mediate communication between or among individuals. Like many stereotyped motor patterns, the probability of a gestural display in a given social context can be modulated by sex steroid hormones. Here, we review how steroid hormones mediate the neural mechanisms that underly gestural communication in humans and nonhumans alike. This is a growing area of research, and thus we explore how sex steroids mediate brain areas involved in language production, social behavior, and motor performance. We also examine the way that sex steroids can regulate behavioral output by acting in the periphery via skeletal muscle. Altogether, we outline a new avenue of behavioral endocrinology research that aims to uncover the hormonal basis for one of the most common modes of communication among animals on Earth. (*Endocrinology* 161: 1–12, 2020)

Gesture plays a fundamental role in animal communication. As humans, we are intimately aware of this fact, given that individuals from all cultures across the globe use gesture to convey ideas, thoughts, and feelings to others (Fig. 1A; (1)). It is therefore unsurprising that linguists and psychologists have spent decades creating a lexicon to classify the various modes of gestural communication, as well as a framework to understand how each of these modes likely function (2–4). Building on this work is a growing body of mechanistic studies, which probe how the brain and body might control gestural communication. Much of this research suggests that the neural substrates responsible for mediating the perception and production of speech also mediate the gesticulation—voluntary or not—that so often accompanies our everyday conversations (5–7). In many ways, this work only scratches the surface of the intricate mechanisms that underlie gestural communication.

In this mini-review, we explore this topic from the point of view of the endocrine system, assessing how hormones modulate the integration of body movement into animal communication programs. We define gesture as the process by which an individual actively moves its face, limbs, and/or body in a coordinated manner (and often with speech or vocalization) to help facilitate communication (8). This broad definition not only encompasses many of the functional sub-classifications of gesture (eg, sign language, pantomime, co-speech gesture), but also recognizes the fact that gestural communication is found across the entire animal kingdom. In this spirit, we begin our review by focusing on neural and neuroendocrine regulation of co-speech gesture in humans. We then expand our discussion beyond humans and give an overview of the hormonal control of gesture in a wider range of taxa. This comparative approach reveals the diverse ways in which hormone systems are involved in the physiological coordination of signaling with the body alongside sound production or, in some cases, without it.

ISSN Online 1945-7170

© Endocrine Society 2020.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Received 10 February 2020. Accepted 16 April 2020.

First Published Online 20 April 2020.

Corrected and Typeset 25 June 2020.

Abbreviations: DM, dorsomedial nucleus; ICo, intercollicular nucleus; IFG, inferior frontal gyrus; mPOA, medial preoptic area; NRA, nucleus retroambigualis; VMH, ventromedial nucleus of the hypothalamus; PAG, periaqueductal gray; RAm, nucleus retroambigualis.



Figure 1. Examples of gestures in vertebrates. (A) A gesture associated with surprise in humans. This photograph was used by Charles Darwin in his book, *The Expression of the Emotions in Man and Animals* [(114); licensed under Getty Museum Open Content Program]. (B) A territorial gesture of hippopotamus [*Hippopotamus amphibius* (115); credit: Robert A. Tobiansky, with permission]. (C) A mating gesture from a male Victoria's riflebird (*Ptiloris victoriae*), whereby he hold out his wings and move his head side-to-side (31) (credit: Francesco Veronesi licensed under CC BY-SA 2.0). (D) A Bornean rock frog (*Staurois parvus*) producing a foot-flagging gesture to compete with rival males at a breeding site [(116); credit: Vienna Zoo/Doris Preininger, with permission]. (E) A male peacock spider (*Maratus splendens*) performing a mating display in which it raises its abdomen and waves its hind legs [(117); credit: Jurgen Otto licensed under CC BY-NC-ND 2.0]. (F) A springbok (*Antidorcas marsupialis*) performing a stotting display, in which it leaps into the air to notify predators that they have been detected [(118); credit: Yathin sk licensed under CC BY-SA 3.0]. (G) A male white-collared manakin (*Manacus manacus*) in a “beard up” posture, which typically occurs when the male is dancing for the female [(65); credit: Steve Garvie licensed under CC BY-NC-SA 2.0]. (H) A male brown anole (*Anolis sagrei*) gestures to conspecifics and potential predators by extending its dewlap (orange throat patch) in a rhythmic pattern [(119) credit: touterse, licensed under CC BY 2.0].

Sex steroids and communication

Why would hormones influence animal communication, and which hormones are most likely to do so? The answer to the first question centers around the major role of hormone action, which provides context-appropriate regulation of behavior (9, 10). In this way, it makes sense that hormones would also regulate the systems controlling communication because this would ensure that language and/or other signals are produced at the right time and place. The answer to the second question requires an understanding of the function of communication. In this brief review, we focus on communication used to navigate sexual encounters, with the aim of increasing reproductive fitness. For these interactions, we can reasonably expect that hormones underlying sexual behavior modulate communication.

This brings us to sex steroids—namely, androgens, estrogens, and progestins. These hormones are principally produced by the gonads, and mediate sperm production in males and follicular development in females (9). At the same time, these hormones are also released into the bloodstream, where they circulate throughout the body and act on target tissues that express androgen receptors, estrogen receptors, and progesterone receptors. Here, sex steroids act to help facilitate the organization of the reproductive

phenotype (eg, development of the genitals, reproductive tract, secondary sexual characteristics, and neural circuits underlying sexual behavior), as well as the regulation of sexual behavior during adulthood. This collectively means that the same hormones regulate both the expression of sexual traits (morphological and behavioral) and the production of mature gametes. Such a design is by no means an evolutionary accident—rather, this coupling is thought to be adaptive, ensuring that reproductive systems are expressed precisely when gametes are available (10).

Because gestural communication often facilitates reproduction, it stands to reason that sex steroids regulate its production in this context. But, how? We know surprisingly little about this topic (11). One factor that makes the question especially interesting is that gestural communication also has nonreproductive functions. This raises the possibility that, in addition to mediating the performance of sexual gestures, sex steroids also help govern how and/or when animals switch between producing sexual gestures and nonsexual ones.

Regulation of gestural communication in humans by sex steroids

The brain controls the production of gestural communication, determining not only when it is performed, but

also how it is performed. Some of the most interesting work that explores this process is in humans using functional magnetic resonance imaging to determine brain regions associated with different forms of gestural communication. For example, studies in men and women investigating the neural basis of co-speech gestures—the hand movements we tend to make while we speak (12, 13)—find that hand movements during speech are correlated specifically with indices of increased activity in brain areas otherwise linked with language production (Fig. 2) (7, 14). This includes the left inferior frontal gyrus (IFG, or Broca's area), anterior superior temporal gyrus, bilateral posterior superior temporal sulcus, left hippocampus, parahippocampal cortex, and ventral and dorsal premotor areas. These brain regions are involved in word retrieval and speech articulation, as well as motor control of the hands (15, 16). At the same time, co-speech gesture is also associated with increased activity in brain areas that make up the gesture network, which governs pantomime gestures, imitating gestures, and tool use (6, 17). Regions include the premotor and primary motor, left posterior parietal, posterior middle temporal, and middle frontal areas (Fig. 2). Overall, these data point to the existence of gesture and language networks that share a variety of nuclei.

Even less is understood about how sex steroids act on these areas to modulate co-speech gesture. The first hint that such effects may occur comes from work in humans and other nonhuman animals showing that many of the brain regions mentioned above either express steroid hormone receptors (estrogen, androgen, or progesterone receptors) or are connected to brain areas that have

these receptors (18–23). The second hint comes from a small number of studies that suggest sex steroids act in the central nervous system to influence language production (and perception), as well as motor control. For instance, neuroimaging studies in adult men and women reveal that gray matter volume in the left IFG is positively associated with levels of circulating 17β -estradiol (the most bioactive estrogen), but negatively associated with circulating testosterone (24). This suggests that estrogens and androgens are capable of acting in these parts of the brain to induce morphological changes, although the time at which these effects occur (either during development or in adulthood) remains unclear. Other work points to strong organizational effects of sex steroid action on brain regions that underlie language production, as both pre- and postnatal effects of steroids influence language development skills later in life (25, 26). Interestingly, these effects would suggest a manifestation of sex differences in co-speech gesture, yet this does not seem to be the case (eg, (2)).

Sex steroids similarly impact motor centers in the brain, which are also involved in the production of co-speech gesture. Preliminary research, for example, suggests that cancer patients undergoing androgen deprivation therapy exhibit a marked decline in gray matter within the primary motor cortex (27). Moreover, patients with congenital adrenal hyperplasia, a condition in which an enzyme deficiency leads to the overproduction of adrenal androgens, show enhanced performance on gross motor and visuomotor tasks, but diminished performance of fine motor skills (28). These effects may be rooted in sex steroid-dependent regulation of motor

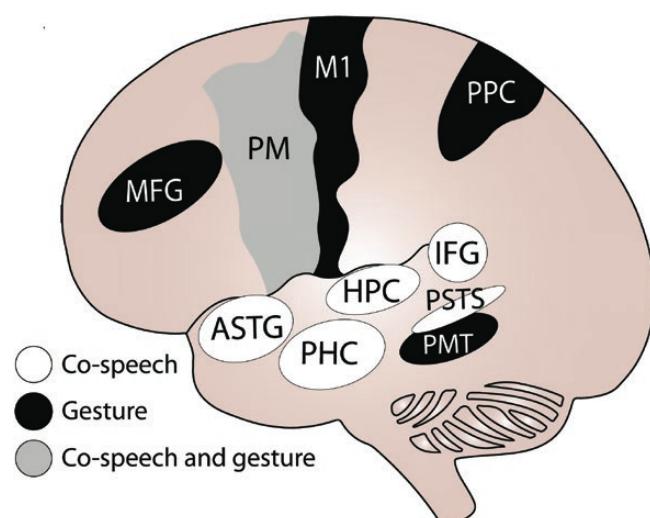


Figure 2. Neural nodes associated with co-speech gesture and other forms of gesture (eg, pantomime) in humans. Neural nodes associated with co-speech gesture (white or gray) include the premotor cortex (PM), hippocampus (HPC), parahippocampal cortex (PHC), left inferior frontal gyrus (IFG), anterior superior temporal gyrus, (ASTG), and bilateral posterior superior temporal sulcus (PSTS). Neural nodes associated with other forms of gesture (eg, pantomime) (black) include the PM, areas of the middle frontal gyrus (MFG), posterior parietal cortex (PPC), the primary motor cortex (M1), and the posterior medial temporal cortex (PMT). The PM is shaded gray to signify its role in both co-speech and gesture.

cortical connectivity during development (29). However, these effects are difficult to reconcile, and thus suggest that regulation of motor command by sex steroid action is a complex process that varies by brain region and across developmental stages.

Although this small body of research implies that sex steroids may exert some influence on the neural basis of co-speech gesture, we must recognize that exceptionally little is known about how these effects are borne out. The first step in addressing this gap is to generate a better understanding of how the brain controls co-speech gesture. This is admittedly a tall order, but ultimately a fascinating question given the degree to which such behavior is ingrained in our communication [even congenitally blind patients produce co-speech gesture (3)]. The next step is to understand how and where sex steroids might act in the brain to modulate this process. As reflected in some of the research cited above, studies in congenital adrenal hyperplasia patients are an obvious place to begin.

Beyond humans: Insights from gestural communication in vertebrates

One might expect that the study of gestural communication in nonhuman animals would quickly dry up. How many species communicate with gesture, and what role could hormones possibly play in this process? The answer to the first part of the question is simple: Nearly all social species use gesture to signal to others, whether they are of the same or different species. Gestures vary tremendously among taxa, ranging from chest pounding to dancing behavior (Fig. 1) (30–36). Often, these displays mediate courtship of potential mates or competitive behavior between rival males (or both). The answer to the second question about hormone regulation of these displays is similarly straightforward: Sex steroids are vital to the production of gestural signals.

Modulation of gesture via sex steroids in the central nervous system

Our understanding of how sex steroids regulate gestural displays starts with classic studies that probe the mechanisms underlying postural and reflexive control necessary for copulation and courtship. One of the best examples comes from work on lordosis in rats. This is not a gesture *per se*; rather, it is a stereotyped mating posture that signifies sexual receptivity and thus mediates successful mating (37). Nonetheless, studies of lordosis behavior in female rodents serve as a template for understanding how hormone systems interact with the brain to influence motor control, which in turn informs our broader understanding of neuroendocrine regulation of gesture. Decades of research shows that a

central node in the activation of lordosis is the ventromedial nucleus of the hypothalamus (VMH) (38, 39). Cells in this nucleus project to a premotor area called the periaqueductal gray (PAG) of the midbrain, which in turn projects to the nucleus retroambiguus (NRA) in the caudal medulla. The NRA sends several bulbospinal projections to spinal motoneurons that innervate much of the body (40). To activate lordosis, estrogens must first prime the VMH by acting through both canonical and noncanonical pathways (37). Subsequent exposure to estrogens and progestins then increases the probability that lordosis occurs in response to sensory input from the female's hind flanks (ie, sensory input from a mounting male) and olfactory information from the vomeronasal organ. Additionally, sex steroid action in these same brain areas also likely mediates a sexually proceptive behavior in female rats known as ear wiggling, which falls well within the definition of a gesture. This behavior signifies willingness to mate to the male and is similarly sensitive to estrogen and progestin priming (41), but far less is known about its underlying neural basis. Nonetheless, this work collectively shows the critical role that sex steroid action plays in setting the neurological stage for refined motor command. Indeed, without estrogen priming and subsequent estrogen/progestin action, lordosis behavior and proceptive gestures fail to manifest (39).

In males, the medial preoptic area (mPOA) is the principal node that mediates sexual behavior, as opposed to the VMH (42, 43). More recent work in birds nicely illustrates how the mPOA (often called the POM in birds) is similarly linked to the systems that govern motor control, given that the mPOA itself does not connect to the spinal cord. Rather, the mPOA projects to the intercollicular nucleus (ICo), a complex of nuclei homologous to the mammalian PAG (44). The cells that receive input from the mPOA then fire on a subnucleus within this complex called the dorsomedial nucleus (DM) (45). To biologists studying display behavior, the DM is an especially interesting part of the brain because its electrical stimulation immediately causes birds (even ones who are anesthetized) to produce vocalizations used for social communication (46). Neuroanatomical work indicates that the DM sends projections to 2 other important areas, the vocal motor nucleus (nXIIts) and the nucleus retroambigualis (RAm) (47). RAm, which is homologous to the mammalian NRA, projects to several spinal motoneurons that innervate thoracic and lumbar expiratory muscles, as well as major cloacal muscles that actuate copulatory reflex movements (48). When testosterone acts via the mPOA, it stimulates the production of different cloacal responses. In male quail, for instance, testosterone modulation of the mPOA

significantly increases the number of rhythmic cloacal sphincter movements in response to a female's presence (49). As such, we again see how central sex steroid action modulates specific movement programs involved in mediating sexual interactions.

As the body of work described above charts out how sex steroid action in the brain can influence basic motor control for sex, we suspect that similar processes underlie the production of gestures used to mediate courtship and male–male competition. In this case, however, regulation of gesture is likely rooted in the functionality of the social decision-making network, which is a group of interconnected nuclei that collectively govern social behavior (Fig. 3) (50, 51). All nodes in this circuit are sensitive to sex steroids, including androgens, estrogens,

and progestins. The network itself includes the VMH, mPOA, and several other hypothalamic nuclei that help control aspects of motivation, arousal, reward, and reinforcement. Importantly, in this list of neural loci is the PAG/ICo, which is thought to serve as the major interface to downstream motoneuron processes. Relative activity of these nuclei helps determine an animal's social output, while steroid hormone action directly mediates activational patterning (52). Importantly, social experience can alter the sensitivity of these brain regions to steroid hormone action, and thus predict meaningful changes in future behavioral interactions (53). Some of the most compelling work to suggest that nodes of the social decision-making network influence gesture and/or posture comes from studies in songbirds (Order:

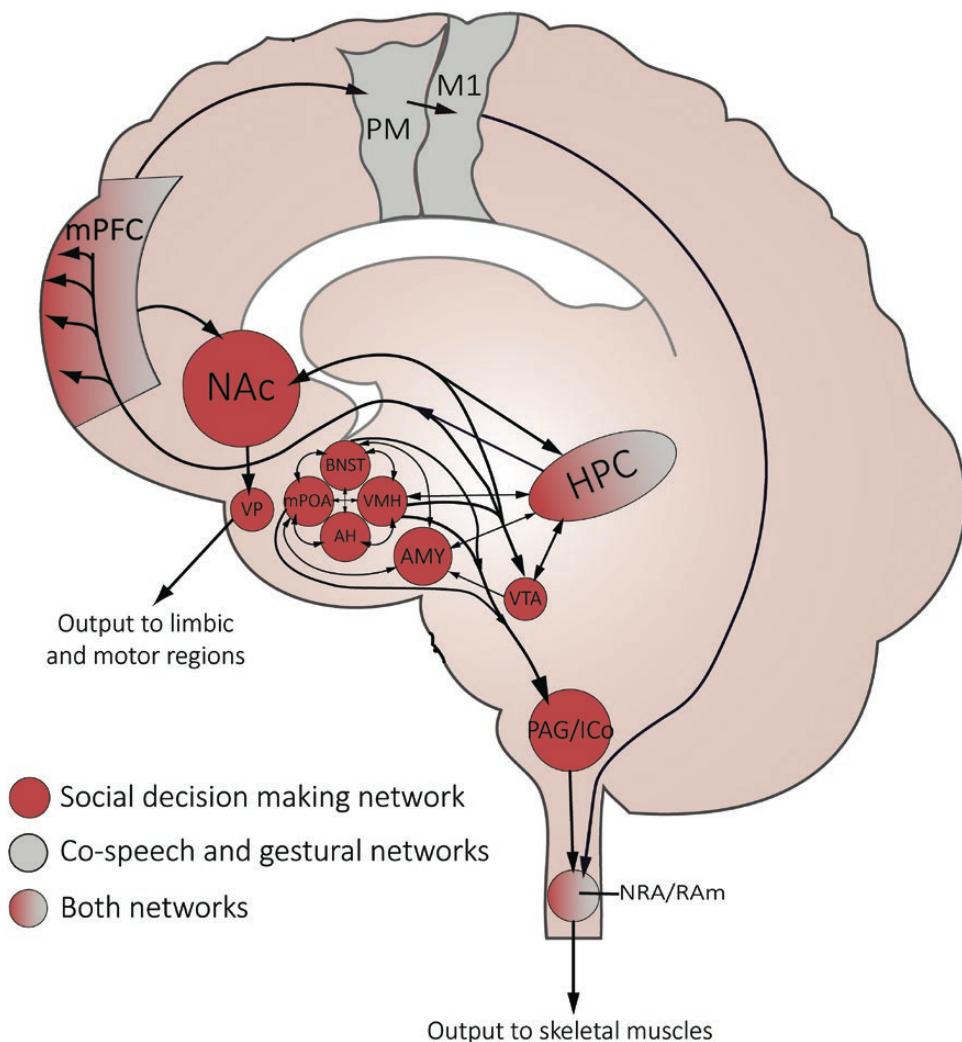


Figure 3. Regions of evolutionary conserved social decision-making network and its interaction with the gesture/co-speech gesture network in the human brain. Select nodes of the social decision making network (red), which include areas from the social behavior network and the motivational/reward circuit (described by 50). Regions involved in co-speech gesture and other forms of gesture (eg, pantomime) are shaded in gray. Nodes that act as an interface between the social decision making network and co-speech/gestural networks are shaded with a gray-red gradient (ie, "Both networks"). Abbreviations: AH, anterior hypothalamus; AMY, amygdala; BNST, bed nucleus of the stria terminalis; HPC, hippocampus; ICo, intercollicular nucleus; M1, primary motor cortex; mPFC, medial prefrontal cortex; mPOA, medial preoptic area; NAC, nucleus accumbens; NRA, nucleus retroambiguus; PAG, periaqueductal gray; RAm, nucleus retroambigualis; VMH, ventromedial hypothalamus; VP, ventral pallidum; VTA, ventral tegmental area.

Passeriformes). For example, work in canaries (*Serinus canaria*) shows that a nucleus within the song control system—the robust nucleus of the arcopallium (RA, a telencephalic nucleus)—directly innervates both the DM (part of the ICo) and RAM (between which there are also connections; see earlier) (54). Integration between these 2 networks is thought to underlie the functional coupling of social display and other relevant behavior, such as motor control of the cloaca and copulation solicitation displays. More importantly, this work provides a template to envision how the nervous systems can integrate aspects of motor control for different types of behavior (eg, song and postural or gestural displays) in a context-appropriate manner.

This research in canaries is important for a few other reasons. First, it again highlights the significance of the hindbrain nuclei (NRA/RAM) for motor skills involved in sexual display. The nuclei are typically viewed as respiratory premotor nuclei, but they are also involved in the execution of other tasks that require alteration of respiratory patterning such as vomiting and abdominal straining (55). This, alongside the fact that NRA/RAM are involved in sexual postures (40, 56), has led others to argue that both nuclei play a broad role in motor control by operating as behavioral pattern generators with numerous connections throughout the nervous system (55). Inputs to NRA/RAM, like the VMH, are sensitive to sex steroids, and studies suggest that estrogenic action within these inputs can trigger neuronal outgrowth of axons from the NRA/RAM down into the spinal cord (57, 58). Thus, sex steroids appear capable of acting centrally to modulate how the brain can send commands to various parts of the spinal cord. Research in male ruffed grouse (*Bonasa umbellus*) provides a potential link between these effects and gestural display. Males of this species perform a territorial and courtship display, in which they stand erect on a log in the forest and rapidly flap their wings up and down to generate a low-frequency drumming noise that booms through the forest (59). Histological studies suggest that this behavior is associated with greater expression of aromatase in certain regions of the VMH (60); thus, enhanced estrogenic production and action in this nucleus may augment NRA/RAM connectivity with spinal interneurons and motoneurons.

Second, the work with canaries also suggests that other brain areas outside the social decision-making network likely contribute to display behavior, and thus gestural displays. The RA is part of the avian arcopallium, which has clear premotor functions. For example, studies show that wing and leg movements are associated with increased expression of immediately

early genes in certain arcopallial nuclei (61), while other work indicates that parts of the arcopallium project to the pontomedullary reticular formation in the brainstem (62, 63). If this latter area is electrically stimulated then basic locomotory patterns (eg, walking, wing flapping) are evoked (64). Research also provides intriguing support for the idea that these pathways are under androgenic modulation in species that perform elaborate gestural displays. Golden-collared manakins (*Manacus vitellinus*), for example, express high levels of androgen receptor throughout much of the arcopallium. These tropical birds produce a courtship display in which they rapidly jump around the forest floor, snapping their wings together in mid-air to generate a loud, firecracker-like pop (65). Avian species that do not produce these displays express little to no androgen receptor in the arcopallium (66), suggesting that increased androgenic sensitivity within this brain region is related to the bird's unusual display routine. Interestingly, one of the species that does not express androgen receptor in the arcopallium is the ruffed grouse (see earlier (60)); thus, androgenic action in the arcopallium itself is not always associated with gesture.

Another important level of the nervous system to consider is the spinal cord. Motoneurons innervating the wing muscles of golden-collared manakins express high levels of androgen receptor, particularly when compared to other birds that do not produce gestural displays with their wings (67). This suggests that androgens help mediate display behavior by acting directly on the motoneurons that relay information from the central nervous system to the muscle itself. Similarly, work in the Japanese quail (*Coturnix japonica*) shows that the motoneurons innervating the cloacal muscles contain high levels of estrogen receptor, which stands in contrast to the dearth of estrogen receptor in most other regions of the spinal cord's ventral horns (68). At the same time, the dorsal horns of the bird's spinal cord make their own estrogens, likely feeding the receptor population in the cloacal motoneurons (69). This suggests that estrogens may act locally within the lower spinal cord to help mediate motor control of the cloaca during sexual interactions.

Additional research suggests that sex steroids are important regulators of neural functioning within the spinal cord. Spinal interneurons, for instance, express estrogen and androgen receptor (70–72). These cells are critical to the control of muscle synergies, which represent the independent, modular movement programs that make up complex behavioral output (73, 74). Thus, if sex steroids regulate the morphology and connectivity of spinal interneurons either during development or in

adulthood, they may have a profound effect on the ontogeny and/or activation of gestural displays. There is a deep and rich literature that explores the neural basis of muscle synergies and their implications for behavioral control, but this work is seldom addressed in the field of behavioral endocrinology, and our understanding of steroid control of gestures could benefit greatly from incorporating these ideas.

Considering the work described above, it is clear that we still know little about how the brain controls animal gestural displays, or how sex steroids act on the brain to modulate this behavior. Studies are needed to pinpoint specific brain regions or neural circuits associated with gestural control, independent of other phenomena (eg, vocalizing, locomotion). From here, we can begin to assess how actions of estrogen, androgens, and progestins modulate these brain nuclei to influence gesture. Other intriguing questions will undoubtedly emerge from studies in this area, such as how central steroid action influences the development of gestural displays (75). Might such effects account for sex differences in gestural behavior? Indeed, future research that embraces this approach promises to redefine our thinking about adaptive motor command and how its mechanisms are embedded in the central networks that underlie the basic elements of social behavior—sex, courtship, territoriality, and cooperation.

Steroidal modulation of gestures via the periphery

We must also recognize that sex steroids act peripherally to influence gestural communication. This process can be conceptualized as steroid modulation of substrates that respond to “instructions” sent from the brain about how to move the body in a specific manner. The idea makes more sense if we consider the nature of the gestures that many animals produce for communication, especially those that demand performance skills and abilities that would not normally be attributed to the species in question (eg, bouts of extreme speed, strength, endurance, or any combination thereof) (76). So how might this work? There are 2 major ways for sex steroids to peripherally regulate the motor systems underlying animal performance.

First, sex steroids might act directly on the musculoskeletal system to adjust how the striated muscle and its associated structures actuate movement. In particular, androgenic steroids, like testosterone, play a powerful role in the regulation of muscle, which expresses high levels of androgen receptor compared with many other tissues in the body (77, 78). Androgens, therefore, induce a variety of effects on muscle fibers, increasing their size, fiber

type composition, and ability to handle calcium ions (79–83). Estrogens and progestins are also known to act on muscle, although their effects are much less clear. Some research suggests that these 2 hormones mediate aspects of muscle performance, like endurance (84, 85). At the same time, other work implies that estrogenic hormones help buffer muscle tissues from use-related damage by directly mediating myocytic repair (86).

Research that directly ties gestural displays to sex steroid action at the level of muscle focuses specifically on androgenic systems. For example, in a fish called the blue-banded goby (*Lythrypnus dalli*), males defend nest sites and court females by performing swim displays, in which they repeatedly extend and retract their pelvic and dorsal fins. The muscles that actuate these fin movements contain high levels of androgen receptor compared with other major muscles that power swimming (87). Moreover, levels of androgen receptor in these same fin muscles are positively associated with the rate at which individuals perform their display, suggesting that androgenic regulation augments muscular control of fin movements related to sexual signaling. Additionally, studies in golden-collared manakins (the acrobatic displaying bird mentioned earlier) shows that activation of androgen receptor in the wing musculature is necessary for most complex gestural signals. If androgen receptors in these tissues are blocked by a peripherally selective androgen receptor antagonist, then males produce fewer gestural displays and slow down the signals they do broadcast to females (88). These effects have been traced back to the muscle that actuates the bird’s rapid wing movements, on which androgens act to increase twitch speeds to ‘superfast’ levels (89). These effects are also observed at the molecular level. For instance, in manakins, androgens upregulate the transcription (messenger ribonucleic acid) of both parvalbumin, a myocytic calcium buffer, and insulin-like growth factor 1, a growth factor that stimulates muscle growth (80). Studies in other taxa similarly show that androgens completely remodel the transcriptomic profile of muscle in a way that undoubtedly affects multiple aspects of performance, such as speed, endurance, force production. (90, 91). Comparative work in birds even shows that the impact of androgens on muscle can vary not only among different tissues, but also across different species (92).

Of course, the role of androgen–muscle interactions in behavior is much more complex than stated above. We would argue that the literature is beginning to assess how androgens help reorient organismal behavior within larger, multifaceted performance landscapes, rather than altering singular aspects of speed, strength, endurance, etc. This idea is anchored by studies showing

that most performance elements are tied together in complex trade-off schema (93–95). Speed, for example, often comes at a cost to force production and the resistance to fatigue (96). In this way, androgens might act on muscle to balance it along this trade-off. In golden-collared manakins, for example, androgenic action not only increases the speed of display muscle, but also likely induces its hypertrophy (89). The latter effects may therefore recoup strength needed to power locomotion, which is otherwise depleted by the substantial increase in speed. If so, then androgens are acting as modulators of performance trade-offs, thereby helping individuals mitigate putative locomotory costs that would accrue as a result of an evolutionary push for an effective (eg, fast) display.

The second way that peripheral androgenic action might regulate mechanisms of gestural control is by indirect modulation of the nervous system (97, 98). This idea originates from work in male rodents, which shows that androgens act on the musculature of the penis to maintain the morphology of the motoneurons that connect these tissues to the spinal cord (99). This occurs because androgens upregulate neurotropic and growth factors, which retrogradely travel from the periphery to the spinal cord (100, 101). There, these proteins initiate a host of changes to the motoneuron, such as increasing its dendritic arborization and soma size (99, 102). Such features of the motoneuron impact its functionality and capacity for integration (103–105).

Unfortunately, little work has been carried out to explore how peripheral steroid action influences central control of behavior. Recent work in frogs, however, suggests that it likely plays a role. Species like the Bornean rock frog (*Staurois parvus*) have evolved intricate waving displays with their hindlimbs to compliment acoustic communication in especially noisy environments, such as under a waterfall. Androgens help activate these displays likely by acting on the musculature that extends, rotates, and retracts the femur (106). Because these displays are believed to be about the slow execution of a gesture that requires great skill and motor command, it is thought the androgenic regulation of muscle helps maintain a spinal phenotype that supports the ability to produce the signal (ie, foot flagging behavior) (106, 107).

Going forward, there are many gaps to fill with regard to our knowledge about sex steroids and their ability to act via the musculoskeletal system to modulate gestural displays. Hints from endocrinological research that estrogens and progestins modulate muscle performance should be examined in a behavioral context. Are the effects of these hormones on select muscles necessary to generate certain types of display movements? If so, why?

How do these hormones change muscle physiology in a functionally meaningful manner? Equally important is a more thorough understanding of how sex steroid-mediated signaling from the muscle to the spinal cord regulates behavior. We have known about this phenomenon for decades, yet it has gained little steam in terms of informing our understanding of behavioral modulation. How might changes to spinal cord morphology induced by peripheral sex steroid action alter motor skills and/or muscle synergies that guide complex movements used to communicate?

Steroid hormones systems and the evolution of gestural communication

An emergent view from the research described above is that sex steroid systems act as channel through which gestural signals can evolve. Accordingly, selection for specific gestural patterns and routines may proceed through concurrent changes to the systems by which sex steroids regulate motor command (107–109). Support for this idea comes from comparative work that shows a positive relationship between expression of androgen receptor in specific target tissues and species variation in gestural display complexity. In *Anolis* lizards, for example, species that exhibit greater rates of territorial push-up display express higher levels of androgen receptor in their forearm muscles (110). Push-up rate was not related to muscle fiber size or body size, and these effects accounted for the shared evolutionary history among the taxa. Overall, this work points to a clear link between the properties of gestural display and the evolution of the androgenic system in the muscles that actuate it. Research in other taxa point to similar effects (106, 111), including work in tropical birds (112). This latter study is important because, while it shows a positive relationship between levels of androgen receptor transcription and taxonomic variation in gestural display complexity, it also reveals that such coevolutionary relationships do not exist with regard to estrogen receptor. Moreover, this study also looks at androgen and estrogen receptor in the spinal cord, and similarly fails to uncover evidence of a coevolutionary relationship between these transcripts and display variation. Thus, the evolutionary linkage between gesture and sex steroid systems appears to be specific to androgenic signaling and muscle.

Building on this idea is work that explores how the androgen receptor itself might evolve. While steroid receptors are thought to be highly conserved, recent work shows that androgen receptor protein in birds does vary across taxa (113). In particular, there are 2 avian families (Class: Aves) that show substantial deviation in the polarity and hydrophobicity of select androgen

receptor domains—manakins (family: Pipridae) and hummingbirds (family: Trochilidae). Many species within these families produce extraordinary gestural displays, pointing to yet another link between the evolution of the androgenic system and elaborate physical displays. Although it is currently unclear how these precise structural changes to the androgen receptor influence its ability to induce signaling, studies of the androgen receptor's biochemistry imply that such modifications likely impact post-translational regulation of the protein's functional potency (113). To this end, if sexual selection for gestural displays drives the evolution of these changes to the androgen receptor, we must recognize that this might influence other androgen-dependent processes unrelated to social display.

Conclusions

In sum, we briefly reviewed the literature that currently provides the basis of our understanding about the relationship between the endocrine system and gestural communication in vertebrates. Most of this work focuses on regulation through sex steroids, acting on pathways in the brain that mediate sexual reflexes and postures. Such research is just beginning to extrapolate these ideas to nonmodel systems, which have evolved highly complex gestural displays that incorporate dance, acrobatics, and other movements requiring a high degree of fine motor control. It appears that the key to understanding these processes lies in untangling the neural pathways that integrate facets of sociality with motor control systems. However, sex steroids also appear to act via the spinal cord and muscular system to prime the body thereby allowing it to appropriately respond to “instructions” from the brain. We end the review with a brief reflection on the evolution of these displays—namely, the idea that changes to the mechanisms underlying steroid action throughout the neuraxis can help precipitate adaptive modifications to gestural communication systems. Thus, sex steroid systems currently appear to be a primary conduit for the evolution of sexual movement programs.

Additional Information

Correspondence: Daniel J. Tobiansky, Department of Ecology and Evolutionary Biology, Brown University, Providence, RI 02912. Email: daniel_tobiansky@brown.edu

Disclosure Summary: The authors have nothing to disclose.

Data Availability: Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

References

1. McNeill D. *Hand and Mind: What Gestures Reveal About Thought*. Chicago, IL: University of Chicago Press; 1992.
2. Gillespie M, James AN, Federmeier KD, Watson DG. Verbal working memory predicts co-speech gesture: evidence from individual differences. *Cognition*. 2014;132(2):174-180.
3. Krauss RM. Why do we gesture when we speak? *Curr Dir Psychol Sci*. 1998;7(2):54-54.
4. Goldin-Meadow S, Nusbaum H, Kelly SD, Wagner S. Explaining math: gesturing lightens the load. *Psychol Sci*. 2001;12(6):516-522.
5. Holle H, Gunter TC, Rüschemeyer SA, Hennenlotter A, Iacoboni M. Neural correlates of the processing of co-speech gestures. *Neuroimage*. 2008;39(4):2010-2024.
6. Króliczak G, Frey SH. A common network in the left cerebral hemisphere represents planning of tool use pantomimes and familiar intransitive gestures at the hand-independent level. *Cereb Cortex*. 2009;19(10):2396-2410.
7. Marstaller L, Burianová H. A common functional neural network for overt production of speech and gesture. *Neuroscience*. 2015;284:29-41.
8. Kendon A. *Gesture: Visible Action as Utterance*. Cambridge, UK: Cambridge University Press; 2004.
9. Adkins-Regan E. *Hormones and Animal Social Behavior: Monographs in Behavior and Ecology*. Princeton, NJ: Princeton University Press; 2005.
10. Crews D, Moore MC. Evolution of mechanisms controlling mating behavior. *Science*. 1986;231(4734):121-125.
11. Emmorey K, Ozyurek A. Language in our hands: neural underpinnings of sign language and co-speech gesture. In: Gazzaniga MS, Mangun R, eds. *The cognitive neurosciences*. 5th ed. Cambridge, MA: MIT Press; 2014:657-666.
12. Valenzeno L, Alibali MW, Klatzky R. Teachers' gestures facilitate students' learning: a lesson in symmetry. *Contemp Educ Psychol*. 2003;28(2):187-204.
13. Alibali MW, Heath DC, Myers HJ. Effects of visibility between speaker and listener on gesture production: some gestures are meant to be seen. *J Mem Lang*. 2001;44(2):169-188.
14. Hostetter AB, Alibali MW. Visible embodiment: gestures as simulated action. *Psychon Bull Rev*. 2008;15(3):495-514.
15. Binder JR, Desai RH, Graves WW, Conant LL. Where is the semantic system? A critical review and meta-analysis of 120 functional neuroimaging studies. *Cereb Cortex*. 2009;19(12):2767-2796.
16. Indefrey P, Levelt WJ. The spatial and temporal signatures of word production components. *Cognition*. 2004;92(1-2):101-144.
17. Vingerhoets G, Vandekerckhove E, Honoré P, Vandemaele P, Achten E. Neural correlates of pantomiming familiar and unfamiliar tools: action semantics versus mechanical problem solving? *Hum Brain Mapp*. 2011;32(6):905-918.
18. Mani S. Progestin receptor subtypes in the brain: the known and the unknown. *Endocrinology*. 2008;149(6):2750-2756.
19. Osterlund MK, Hurd YL. Estrogen receptors in the human forebrain and the relation to neuropsychiatric disorders. *Prog Neurobiol*. 2001;64(3):251-267.
20. Pérez SE, Chen EY, Mufson EJ. Distribution of estrogen receptor alpha and beta immunoreactive profiles in the postnatal rat brain. *Brain Res Dev Brain Res*. 2003;145(1):117-139.
21. Puy L, MacLusky NJ, Becker L, Karsan N, Trachtenberg J, Brown TJ. Immunocytochemical detection of androgen receptor in human temporal cortex characterization and application of polyclonal androgen receptor antibodies in frozen and paraffin-embedded tissues. *J Steroid Biochem Mol Biol*. 1995;55(2):197-209.
22. Sarrieau A, Mitchell JB, Lal S, Olivier A, Quirion R, Meaney MJ. Androgen binding sites in human temporal cortex. *Neuroendocrinology*. 1990;51(6):713-716.

23. Shughrue PJ, Merchenthaler I. Distribution of estrogen receptor β immunoreactivity in the rat central nervous system. *J Comp Neurol*. 2001;436(1):64–81.
24. Witte AV, Savli M, Holik A, Kasper S, Lanzenberger R. Regional sex differences in grey matter volume are associated with sex hormones in the young adult human brain. *Neuroimage*. 2010;49(2):1205–1212.
25. Schaadt G, Hesse V, Friederici AD. Sex hormones in early infancy seem to predict aspects of later language development. *Brain Lang*. 2015;141:70–76.
26. Whitehouse AJ, Maybery MT, Hart R, et al. Fetal androgen exposure and pragmatic language ability of girls in middle childhood: implications for the extreme male-brain theory of autism. *Psychoneuroendocrinology*. 2010;35(8):1259–1264.
27. Chao HH, Hu S, Ide JS, et al. Effects of androgen deprivation on cerebral morphometry in prostate cancer patients – an exploratory study. *Plos ONE*. 2013;8(8):e72032.
28. Collaer ML, Brook CG, Conway GS, Hindmarsh PC, Hines M. Motor development in individuals with congenital adrenal hyperplasia: strength, targeting, and fine motor skill. *Psychoneuroendocrinology*. 2009;34(2):249–258.
29. Venkatesan C, Kritzer MF. Perinatal gonadectomy affects corticocortical connections in motor but not visual cortex in adult male rats. *J Comp Neurol*. 1999;415(2):240–265.
30. Miles MC, Cheng S, Fuxjager MJ. Biogeography predicts macroevolutionary patterning of gestural display complexity in a passerine family. *Evolution*. 2017;71(5):1406–1416.
31. Miles MC, Fuxjager MJ. Synergistic selection regimens drive the evolution of display complexity in birds of paradise. *J Anim Ecol*. 2018;87(4):1149–1159.
32. Miles MC, Fuxjager MJ. Phenotypic diversity arises from secondary signal loss in the elaborate visual displays of toucans and barbets. *Am Nat*. 2019;194(2):152–167.
33. Miles MC, Schuppe ER, Ligon RM, Fuxjager MJ. Macroevolutionary patterning of woodpecker drums reveals how sexual selection elaborates signals under constraint. *Proc R Soc B* 2018;285(1873):20172628.
34. How MJ, Hemmi JM, Zeil J, Peters R. Claw waving display changes with receiver distance in fiddler crabs, *Uca perplexa*. *Anim Behav*. 2008;75:1015–1022.
35. Johnson MA, Wade J. Behavioural display systems across nine *Anolis* lizard species: sexual dimorphisms in structure and function. *Proc Biol Sci*. 2010;277(1688):1711–1719.
36. Lindsay WR, Houck JT, Giuliano CE, Day LB. Acrobatic courtship display coevolves with brain size in manakins (Pipridae). *Brain Behav Evol*. 2015;85(1):29–36.
37. Pfaff D. Cellular and molecular mechanisms of female reproductive behaviors. *Physiol Reprod*. 1994;2:107–220.
38. Blaustein JD. Neuroendocrine regulation of feminine sexual behavior: lessons from rodent models and thoughts about humans. *Annu Rev Psychol*. 2008;59:93–118.
39. Kauffman AS, Micevych PE. Female sexual behavior and hormones in mammals. In: Choe JC, ed. *Encyclopedia of Animal Behavior*. UK: Elsevier; 2019:403–419.
40. Vanderhorst VG, Holstege G. Caudal medullary pathways to lumbosacral motoneuronal cell groups in the cat: evidence for direct projections possibly representing the final common pathway for lordosis. *J Comp Neurol*. 1995;359(3):457–475.
41. Micevych PE, Meisel RL. Integrating neural circuits controlling female sexual behavior. *Front Syst Neurosci*. 2017;11:42.
42. Nutsch VL, Will RG, Robison CL, Martz JR, Tobiansky DJ, Dominguez JM. Colocalization of mating-induced Fos and D2-Like dopamine receptors in the medial preoptic area: influence of sexual experience. *Front Behav Neurosci*. 2016;10:75.
43. Tobiansky DJ, Hattori T, Scott JM, Nutsch VL, Roma PG, Dominguez JM. Mating-relevant olfactory stimuli activate the rat brain in an age-dependent manner. *Neuroreport*. 2012;23(18):1077–1083.
44. Kingsbury MA, Kelly AM, Schrock SE, Goodson JL. Mammal-like organization of the avian midbrain central gray and a reappraisal of the intercollicular nucleus. *Plos ONE*. 2011;6(6):e20720.
45. Riters LV, Alger SJ. Neuroanatomical evidence for indirect connections between the medial preoptic nucleus and the song control system: possible neural substrates for sexually motivated song. *Cell Tissue Res*. 2004;316(1):35–44.
46. Wild J, Li D, Eagleton C. Projections of the dorsomedial nucleus of the intercollicular complex (DM) in relation to respiratory-vocal nuclei in the brainstem of pigeon (*Columba livia*) and zebra finch (*Taeniopygia guttata*). *J Compar Neurol*. 1997;377(3):392–413.
47. Wild JM. The avian nucleus retroambigualis: a nucleus for breathing, singing and calling. *Brain Res*. 1993;606(2):319–324.
48. Wild JM, Balthazart J. Neural pathways mediating control of reproductive behavior in male Japanese quail. *J Comp Neurol*. 2013;521(9):2067–2087.
49. Riters LV, Ahsil P, Balthazart J. Effects of brain testosterone implants on appetitive and consummatory components of male sexual behavior in Japanese quail. *Brain Res Bull*. 1998;47(1):69–79.
50. O'Connell LA, Hofmann HA. The vertebrate mesolimbic reward systems and social behavior network: a comparative synthesis. *J Compar Neurol*. 2011;519(18):3599–3639.
51. Goodson JL, Evans AK, Lindberg L, Allen CD. Neuro-evolutionary patterning of sociality. *Proc Biol Sci*. 2005;272(1560):227–235.
52. Maney DL, Goode CT, Lange HS, Sanford SE, Soloman BJ. Estradiol modulates neural responses to song in a seasonal songbird. *J Compar Neurol*. 2008;511(2):173–186.
53. Fuxjager MJ, Forbes-Lorman RM, Coss DJ, Auger CJ, Auger AP, Marler CA. Winning territorial disputes selectively enhances androgen sensitivity in neural pathways related to motivation and social aggression. *Proc Natl Acad Sci U S A*. 2010;107(27):12393–12398.
54. Wild JM, Botelho JF. Involvement of the avian song system in reproductive behaviour. *Biol Lett*. 2015;11(12):20150773.
55. Kirkwood PA, Ford TW. Do respiratory neurons control female receptive behavior: a suggested role for a medullary central pattern generator? *Prog Brain Res*. 2004;143:105–114.
56. Vanderhorst VG, Holstege G. Nucleus retroambigualis projections to lumbosacral motoneuronal cell groups in the male cat. *J Comp Neurol*. 1997;382(1):77–88.
57. VanderHorst VG. Nucleus retroambigualis-spinal pathway in the mouse: Localization, gender differences, and effects of estrogen treatment. *J Compar Neurol*. 2005;488(2):180–200.
58. VanderHorst VG, Holstege G. Estrogen induces axonal outgrowth in the nucleus retroambigualis-lumbosacral motoneuronal pathway in the adult female cat. *J Neurosci*. 1997;17(3):1122–1136.
59. Garcia M, Charrier I, Iwaniuk AN. Directionality of the drumming display of the ruffed grouse. *Condor*. 2012;114(3):500–506.
60. Corfield JR, Harada N, Iwaniuk AN. Aromatase expression in the brain of the ruffed grouse (*Bonasa umbellus*) and comparisons with other galliform birds (Aves, Galliformes). *J Chem Neuroanat*. 2013;47:15–27.
61. Feenders G, Liedvogel M, Rivas M, et al. Molecular mapping of movement-associated areas in the avian brain: a motor theory for vocal learning origin. *Plos ONE*. 2008;3(3):e1768.
62. Dubbeldam J. The neural substrate for ‘learned’ and ‘nonlearned’ activities in birds: a discussion of the organization of bulbar reticular premotor systems with side-lights on the mammalian situation. *Cells Tissues Organs*. 1998;163(3):157–172.
63. Sholomenko GN, Funk GD, Steeves JD. Avian locomotion activated by brainstem infusion of neurotransmitter agonists and antagonists. I. Acetylcholine excitatory amino acids and substance P. *Exp Brain Res*. 1991;85(3):659–673.
64. Steeves JD, Sholomenko GN, Webster DM. Stimulation of the pontomedullary reticular formation initiates locomotion in decerebrate birds. *Brain Res*. 1987;401(2):205–212.
65. Fusani L, Barske J, Day LD, Fuxjager MJ, Schlenger BA. Physiological control of elaborate male courtship: female choice

- for neuromuscular systems. *Neurosci Biobehav Rev*. 2014;46(Pt 4):534-546.
66. Fusani L, Donaldson Z, London SE, Fuxjager MJ, Schlinger BA. Expression of androgen receptor in the brain of a sub-oscine bird with an elaborate courtship display. *Neurosci Lett*. 2014;578:61-65.
67. Fuxjager MJ, Schultz JD, Barske J, et al. Spinal motor and sensory neurons are androgen targets in an acrobatic bird. *Endocrinology*. 2012;153(8):3780-3791.
68. Evrard HC, Balthazart J. Localization of oestrogen receptors in the sensory and motor areas of the spinal cord in Japanese quail (*Coturnix japonica*). *J Neuroendocrinol*. 2002;14(11):894-903.
69. Evrard HC, Balthazart J. Aromatase (estrogen synthase) activity in the dorsal horn of the spinal cord: functional implications. *Ann Ny Acad Sci*. 2003;1007(1):263-271.
70. Fan X, Kim H-J, Warner M, Gustafsson J-Å. Estrogen receptor β is essential for sprouting of nociceptive primary afferents and for morphogenesis and maintenance of the dorsal horn interneurons. *Proc Natl Acad Sci U S A*. 2007;104(34):13696-13701.
71. Hamson DK, Jones BA, Watson NV. Distribution of androgen receptor immunoreactivity in the brainstem of male rats. *Neuroscience*. 2004;127(4):797-803.
72. Todd A. Ablation of spinal cord estrogen receptor-expressing interneurons reduces chemically-induced modalities of pain and itch. *J Compar Neurol*. 2020;1-15.
73. Bizzelli E, Cheung VC. The neural origin of muscle synergies. *Front Comput Neurosci*. 2013;7:51.
74. Hart CB, Giszter SF. A neural basis for motor primitives in the spinal cord. *J Neurosci*. 2010;30(4):1322-1336.
75. Chiver I, Schlinger BA. Sex differences in androgen activation of complex courtship behaviour. *Anim Behav*. 2017;124:109-117.
76. Beyers J, Heberts E, Podos J. Female mate choice based upon male motor performance. *Anim Behav*. 2010;79(4):771-778.
77. Michel G, Baulieu EE. Androgen receptor in rat skeletal muscle: characterization and physiological variations. *Endocrinology*. 1980;107(6):2088-2098.
78. Feng NY, Katz A, Day LB, Barske J, Schlinger BA. Limb muscles are androgen targets in an acrobatic tropical bird. *Endocrinology*. 2010;151(3):1042-1049.
79. Brantley RK, Marchaterre MA, Bass AH. Androgen effects on vocal muscle structure in a teleost fish with inter- and intra-sexual dimorphism. *J Morphol*. 1993;216(3):305-318.
80. Fuxjager MJ, Barske J, Du S, Day LB, Schlinger BA. Androgens regulate gene expression in avian skeletal muscles. *Plos ONE*. 2012;7(12):e51482.
81. Holmes MM, Bartrem CL, Wade J. Androgen dependent seasonal changes in muscle fiber type in the dewlap neuromuscular system of green anoles. *Physiol Behav*. 2007;91(5):601-608.
82. Regnier M, Herrera AA. Changes in the contractile properties by androgen hormones in sexually dimorphic muscles of male frogs (*Xenopus laevis*). *Journal of Physiology (Cambridge)*. 1993;461(1):565-581.
83. Sassoone DA, Gray GE, Kelley DB. Androgen regulation of muscle fiber type in the sexually dimorphic larynx of *Xenopus laevis*. *J Neurosci*. 1987;7(10):3198-3206.
84. Schneider BS, Fine JP, Nadolski T, Tiidus PM. The effects of estradiol and progesterone on plantarflexor muscle fatigue in ovariectomized mice. *Biol Res Nurs*. 2004;5(4):265-275.
85. Nagai S, Ikeda K, Horie-Inoue K, et al. Estrogen modulates exercise endurance along with mitochondrial uncoupling protein 3 downregulation in skeletal muscle of female mice. *Biochem Biophys Res Commun*. 2016;480(4):758-764.
86. Enns DL, Tiidus PM. The influence of estrogen on skeletal muscle: sex matters. *Sports Med*. 2010;40(1):41-58.
87. Schuppe ER, Pradhan DS, Thonkulpitak K, Drilling C, Black M, Grober MS. Sex differences in neuromuscular androgen receptor expression and sociosexual behavior in a sex changing fish. *Plos ONE*. 2017;12(5):e0177711.
88. Fuxjager MJ, Longpre KM, Chew JG, Fusani L, Schlinger BA. Peripheral androgen receptors sustain the acrobatics and fine motor skill of elaborate male courtship. *Endocrinology*. 2013;154(9):3168-3177.
89. Fuxjager MJ, Miles MC, Goller F, Petersen J, Yancey J. Androgens support male acrobatic courtship behavior by enhancing muscle speed and easing the severity of its tradeoff with force. *Endocrinology*. 2017;158(11):4038-4046.
90. Wyce A, Bai Y, Nagpal S, Thompson CC. Research resource: the androgen receptor modulates expression of genes with critical roles in muscle development and function. *Mol Endocrinol*. 2010;24(8):1665-1674.
91. Yoshioka M, Boivin A, Bolduc C, St-Amand J. Gender difference of androgen actions on skeletal muscle transcriptome. *J Mol Endocrinol*. 2007;39(2):119-133.
92. Fuxjager MJ, Lee JH, Chan TM, et al. Hormones, genes and athleticism: effect of androgens on the avian muscular transcriptome. *Mol Endocrinol*. 2016;30(2):254-271.
93. Rome LC, Lindstedt SL. The quest for speed: muscles built for high-frequency contractions. *News Physiol Sci*. 1998;13(6):261-268.
94. Herrel A, Podos J, Vanhooydonck B, Hendry AP. Force-velocity trade-off in Darwin's finch jaw function: a biomechanical basis for ecological speciation? *Funct Ecol*. 2009;23(1):119-125.
95. Vanhooydonck B, Van Damme R, Aerts P. Speed and stamina trade-off in lacertid lizards. *Evolution*. 2001;55(5):1040-1048.
96. Rome LC, Cook C, Syme DA, et al. Trading force for speed: why superfast crossbridge kinetics leads to superlow forces. *Proc Natl Acad Sci U S A*. 1999;96(10):5826-5831.
97. Schlinger BA, Paul K, Monks DA. Muscle, a conduit to brain for hormonal control of behavior. *Horm Behav*. 2018;105:58-65.
98. Sengelaub DR, Forger NG. The spinal nucleus of the bulbocavernosus: firsts in androgen-dependent neural sex differences. *Horm Behav*. 2008;53(5):596-612.
99. Rand MN, Breedlove SM. Androgen alters the dendritic arbors of SNB motoneurons by acting upon their target muscles. *J Neurosci*. 1995;15(6):4408-4416.
100. Verhovshek T, Cai Y, Osborne MC, Sengelaub DR. Androgen regulates brain-derived neurotrophic factor in spinal motoneurons and their target musculature. *Endocrinology*. 2010;151(1):253-261.
101. Verhovshek T, Sengelaub DR. Androgen action at the target musculature regulates brain-derived neurotrophic factor protein in the spinal nucleus of the bulbocavernosus. *Dev Neurobiol*. 2013;73(8):587-598.
102. Araki I, Harada Y, Kuno M. Target-dependent hormonal control of neuron size in the rat spinal nucleus of the bulbocavernosus. *J Neurosci*. 1991;11(10):3025-3033.
103. Elias LA, Chaud VM, Kohn AF. Models of passive and active dendrite motoneuron pools and their differences in muscle force control. *J Comput Neurosci*. 2012;33(3):515-531.
104. Heckman CJ, Lee RH, Brownstone RM. Hyperexcitable dendrites in motoneurons and their neuromodulatory control during motor behavior. *Trends Neurosci*. 2003;26(12):688-695.
105. Mendell LM. The size principle: a rule describing the recruitment of motoneurons. *J Neurophysiol*. 2005;93(6):3024-3026.
106. Mangiameli LA, Fuxjager MJ, Schuppe ER, Taylor RS, Hödl W, Preininger D. Increased androgenic sensitivity in the hind limb neuromuscular system marks the evolution of a derived gestural display. *Proc Natl Acad Sci U S A*. 2016;113(20):5664-5669.
107. Mangiameli LA, Fuxjager MJ. Insight into the neuroendocrine basis of signal evolution: a case study in foot-flagging frogs. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol*. 2018;204(1):61-70.
108. Fuxjager MJ, Schlinger BA. Perspectives on the evolution of animal dancing: a case study in manakins. *Curr Opin Behav Sci*. 2015;6:7-12.

109. Fuxjager MJ, Schuppe ER. Androgenic signaling systems and their role in behavioral evolution. *J Steroid Biochem Mol Biol*. 2018;184:47–56.
110. Johnson MA, Kircher BK, Castro DJ. The evolution of androgen receptor expression and behavior in *Anolis* lizard forelimb muscles. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol*. 2018;204(1):71–79.
111. Schuppe ER, Fuxjager MJ. Phenotypic variation reveals sites of evolutionary constraint in the androgenic signaling pathway. *Horm Behav*. 2019;115:104538.
112. Fuxjager MJ, Eaton J, Lindsay WR, et al. Evolutionary patterns of adaptive acrobatics and physical performance predict expression profiles of androgen receptor – but not oestrogen receptor – in the forelimb musculature. *Funct Ecol*. 2015;29(9):1197–1208.
113. Schuppe ER, Miles MC, Fuxjager MJ. Evolution of the androgen receptor: perspectives from human health to dancing birds. *Mol Cell Endocrinol*. 2020;499:110577.
114. Darwin C. *The Expression of the Emotions of Man and Animals*. London, UK: John Murray; 1872.
115. Blowers TE, Waterman JM, Kuhar CW, Bettinger TL. Social behaviors within a group of captive female *Hippopotamus amphibius*. *J Ethol*. 2010;28(2):287–294.
116. Hödl W, Amézquita A. In: Ryan MJ, ed. *Anuran Communication*. UK: Smithsonian; 2001.
117. Girard MB, Kasumovic MM, Elias DO. Multi-modal courtship in the peacock spider, *Maratus volans* (O.P.-Cambridge, 1874). *Plos ONE*. 2011;6(9):e25390.
118. FitzGibbon CD, Fanshawe JH. Stotting in Thomson's gazelles: an honest signal of condition. *Behav Ecol Sociobiol*. 1988;23(2):69–74.
119. Ord TJ, Blumstein DT, Evans CS. Intrasexual selection predicts the evolution of signal complexity in lizards. *Proc Biol Sci*. 2001;268(1468):737–744.