



Making the most of your electrons: Challenges and opportunities in characterizing hybrid interfaces with STEM

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Inspired by the unique architectures composed of hard and soft materials in natural and biological systems, synthetic hybrid structures and associated hard-soft interfaces have recently evoked significant interest. Soft matter is typically dominated by structural fluctuations even at room temperature, while hard matter is governed by rigid mechanical behavior. This dichotomy offers considerable opportunities to leverage the disparate properties offered by these components across a wide spectrum spanning from basic science to engineering insights with significant technological overtones. Such hybrid structures, which include polymer nanocomposites, DNA functionalized nanoparticle superlattices, and metal organic frameworks to name a few, have delivered promising insights into the technologically relevant applications such as catalysis, environmental remediation, optoelectronics, and medicine.

The interfacial structure between the hard and soft phases demonstrates features across a variety of length scales and often strongly influence the functionality of hybrid systems. While scanning/transmission electron microscopy (S/TEM) has proven to be a valuable tool for acquiring intricate molecular and nanoscale details of these interfaces, the unusual nature of hybrid composites presents a suite of challenges that make assessing or establishing structure–property relationships especially difficult. There are additional considerations at all stages of sample analysis from preparing electron-transparent samples to obtaining sufficient contrast to resolve the interface between dissimilar materials given the dose sensitivity of soft materials.

We discuss each of these challenges and supplement a review of recent developments in the field with additional experimental investigations and simulations to present solutions for attaining a nano or molecular-level understanding of these interfaces. These solutions present a host of opportunities for investigating the role interfaces play in this unique class of functional materials.

1. Introduction

The unusual hierarchical architectures composed of hard and soft materials in natural and biological systems have inspired a surge in interest related to the synthesis of hybrid

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nanostructures. These hard/soft interfaces (HSI) are ubiquitous across multiple length-scales (down to the nano or molecular scale) in nature and play a critical role in ensuring favorable properties under a variety of environmental conditions. For instance, bones are composed of hard hydroxyapatite and soft collagen in order to provide the necessary structural support to protect internal organs [1,2]. Similar interfaces in teeth between hard enamel and soft dentin provide the extraordinary mechanical strength and toughness that teeth display [3–5]. These types of interfaces are also found in the case of nacre, or mother of pearl, which exhibits excellent mechanical strength and resilience in part due to the underlying architecture composed of hard aragonite and soft biopolymer [6,7].

In materials science, the composite structure archetype has been leveraged for successfully improving properties of structural materials. Hard, brittle materials can be made tougher and more resilient by introducing softer fibrous or particulate species into the underlying matrix [8,9]. More recently a similar approach has been deployed at the nanoscale. Examples include functionalized nanoparticles [10,11], DNA-mediated nanoparticle superlattices [12,13], OD/2D nanocomposites [14], and metal–organic framework - nanoparticle composites [15–17]. These hybrid materials have numerous applications including in supercapacitors [18], flame retardants [19], catalysis [20,21], environmental remediation [22,267], optoelectronics [23,24], batteries [25], photovoltaic cells [26], medicine [11,14], and wearable technologies [27].

1.1. Hard/Soft Interfaces (HSI): structure dictates performance

Although there is great diversity in these materials and their applications, hybrid structures are unified in the fact that many of their exceptional and exotic phenomena arise from the bridging of two dissimilar materials. This unusual HSI region can consist of an abrupt interface or a slowly graded interphase. Examples of abrupt soft/hard interfaces include OD/2D core-shell architectures [24]. In this case, the hard nanoparticle core and soft layered shell interface can be atomically sharp and devoid of any buffer region. The types of bonds present at these HSI dictate the level of charge and photocarrier injection present as well as the magnitude of the diffuse interface scattering that phonons face during heat dissipation [28].

Alternatively, polymer nanocomposites can demonstrate gradually evolving interfacial regions, or interphases separating the hard and soft components. In this area, which can extend on the order of hundreds of nanometers, the soft polymer undergoes chemical and physical changes near the hard material inclusion [29–31].

Atomic and molecular-scale structures provide information regarding the nature of the chemical bonds present between different constituent materials. This information thus helps explain the level of load transfer and stress concentrations that arise when these materials undergo deformation processes [32].

1.2. Challenges associated with STEM analysis of HSI

While the behavior and performance of materials can result from structures encompassing a wide variety of length-scales, in many cases, the critical or deterministic features tend to be on the nano or molecular-scale. Both conventional transmission electron

microscopy (CTEM) and scanning transmission electron microscopy (STEM) are indispensable tools for understanding HSI in these materials. The spatial resolution of the spectroscopic and analytical techniques concomitant with STEM, such as energy dispersive X-ray spectroscopy (EDS) and electron energy loss spectroscopy (EELS), make it possible to identify local chemical, vibrational, electronic, and/or magnetic fluctuations of materials [33–36]. Here, we focus solely on STEM as the multimodal nature of its analytical and imaging techniques offer practical tools for probing these interfaces at the critical length scales [37–40].

STEM requires thin, electron transparent samples, so careful sample preparation is essential to preserve the sample integrity and avoid spurious signals [41]. Even when their natural structure is preserved, specimen damage resulting from electron radiation can take on many forms including knock-on damage (the displacement of atoms from the crystal lattice), radiolysis (inelastic ionization), charging, and/or heating [42,43]. In general, soft materials tend to be more prone to electron beam damage than hard materials. For instance, in the case of a hybrid inorganic/organic perovskite material, replacement of the inorganic Cs^+ ions with organic CH_3NH_3^+ ions leads to a structure that can withstand a multiple order of magnitude decrease in the dose rate ($100 \text{ e}^- \text{ \AA}^{-2} \text{ s}^{-1}$ to $4 \text{ e}^- \text{ \AA}^{-2} \text{ s}^{-1}$) and cumulative dose ($\sim 1000 \text{ e}^- \text{ \AA}^{-2}$ to $\sim 1 \text{ e}^- \text{ \AA}^{-2}$) at room temperature [44–46]. Metal organic frameworks (MOFs), another hybrid structure, can withstand a room temperature dose on the order of $10\text{--}20 \text{ e}^- \text{ \AA}^{-2}$ [47]. As such, the softer material limits the overall dose when imaging HSI and makes it difficult to obtain adequate signal from both constituents present as highlighted in Fig. 1 [48].

The accelerating voltage also plays a role on the damage threshold as soft materials are particularly prone to radiolysis [49], which scales with decreasing electron beam energy, while hard materials are particularly prone to knock-on damage [50],

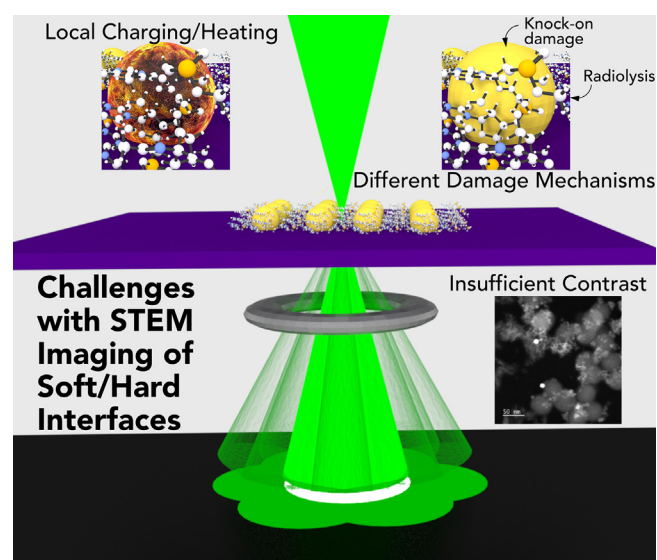


FIG. 1

Challenges associated with imaging Hard/Soft Interfaces with STEM: These include local charging/heating, a discrepancy in beam damage mechanisms, and inadequate image contrast between the different material components.

which scales with increasing electron beam energy. This makes it challenging to entirely avoid beam damage in the critical interphase and interfacial regions. Nonetheless because the obtainable spatial resolution depends on the specimen stability prior to degradation of the measured signal [43], mitigating these sources of specimen damage while simultaneously boosting signal is essential in order to acquire high resolution images of hybrid interfaces.

Some of the most commonly employed tactics to mitigate beam-induced damage and/or boost signal include staining [51–53] and electron microscopy at cryogenic temperatures (cryo-EM) [54]. Unfortunately, neither of these proven methodologies adequately addresses the additional complexity introduced when a hard component is present. In the case of staining, the heavy metal scattering agent obscures the internal structure of the object, often compromising resolution [55]. On the other hand, cryo-EM has been a revolutionary advance that minimizes secondary effects from the initial electron-sample interaction and leads to an overall decrease in beam damage in soft materials [56,57]. However, this method alone does not address the discrepancy in contrast between hard phases and soft phases.

1.3. Review outline and scope

In this review, we discuss the role that STEM can play in interrogating HSI by summarizing recently developed techniques and proposing solutions for addressing challenges associated with imaging HSI. It is intended for material scientists, chemists, and physicists interested in applying STEM techniques to unravel the complex chemical and physical structure of hybrid materials. We first discuss the inherent complexities associated with preparing thin, hybrid composite specimens and a few specialized methods that perform well despite aforementioned constraints. We then discuss how recent experimental advances throughout the entire experimental workflow beginning with sample preparation, followed by imaging and post-processing methodologies, provide a route to attaining improved contrast and image quality from hard/soft interfaces (Fig. 2). We then detail the use of STEM tomography to attain rich three-dimensional information and the use of *in situ* approaches to explore the dynamical evolution of such interfaces. Finally, we discuss ongoing advances and best practices related to microscopy data management that will unlock more opportunities to attain holistic sample information in the future.

2. Specialized sample preparation

Sufficiently thin samples that accurately represent their bulk counterparts must be carefully prepared in order to fully access the variety of signals and information available through STEM. Although simple drop-casting methods can be used for hybrid composite systems such as nanoparticle-DNA hybrids [62,63], preparation of HSI samples can generally be quite challenging. The mismatch in mechanical properties present at HSI creates the need for refined sample preparation methods in order to adequately preserve these interfaces for subsequent microanalysis. Here we provide a breakdown of two specific techniques that

have recently received much attention for site-specific isolation of HSI: ultramicrotomy and focused ion beam (FIB) milling.

2.1. Ultramicrotomy: generating cross-sections of diverse samples

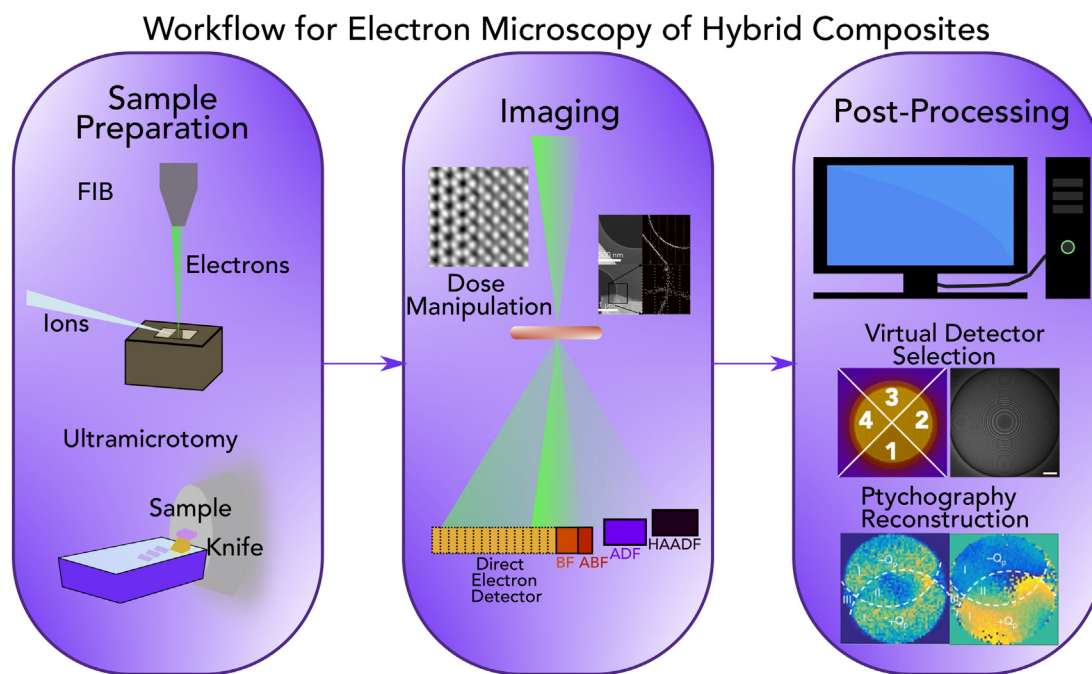
Ultramicrotomy is traditionally used for the analysis of cells and biological tissue embedded in an epoxy resin [64,65], however, many hard [66,67], soft [68], and hybrid materials [69,22][267] can also be prepared with this method [70]. This technique uses an ultramicrotome in conjunction with a glass or diamond knife to produce ultrathin (40–200 nm) cross-sections of material. Biological samples are chemically processed through a series of aldehydes and osmium tetroxide, before being dehydrated and embedded in an epoxy resin. The solidification of this matrix produces a rigid sample for sectioning. A similar technique of embedment with epoxy resin can be used with hybrid systems such as MOFs or nanoparticle-DNA conjugates. Alternatively, bulky materials exhibiting glass transition temperatures (T_g) above room temperature can be mounted onto the ultramicrotome and directly sectioned.

In situations where the soft constituent displays T_g below room temperature, ultramicrotomy can be employed at cryogenic temperatures or with an ultrasonic diamond knife. While the former variant can initiate compression artifacts, the use of an ultrasonic diamond knife, which utilizes a piezo-electric crystal to oscillate the diamond blade in the x-direction relative to the orientation of the block face, can be operated at room temperature and typically minimizes compression artifacts [71]. Large differences in hardness between hard and soft components can further lead to artifacts such as tears, where the harder material is pulled out of the matrix, displacement of softer materials in the membrane or chatter, in which lines form in the specimen parallel to the knife edge [72]. Generally, these artifacts can be mitigated by carefully varying the cutting speed, the cutting angle, or the orientation of the knife blade [72].

2.2. Focused Ion Beam (FIB): site-specific thinning

FIB milling is another versatile method for preparing a broad variety of hybrid composite samples. This includes carbon fibers in epoxy matrix, where the use of FIB makes it possible to preserve the important interphase region that dictates the mechanical performance of such composites [73–76]. For biological hybrid composites, FIB has been exploited to prepare lamella of teeth, bones, and nacre [5,77,78]. In this case, the use of FIB enables subsequent high-resolution imaging of chemical gradients. For electronic architectures, FIB has been employed to isolate HSI present in photovoltaics [79] and flexible electronics [80]. Moreover, the ability to micromachine samples on the nanometer length scale has made FIB attractive for fabricating and positioning samples on specialized grids for 3D tomography as well as *in situ* analysis (Sections 4.1 and 4.2) [81,82].

This method involves milling and isolating a region of interest with a nanometer-scale ion probe. The lamella is then cut free and welded onto a TEM grid where it is thinned with the ion beam such that it is electron transparent for STEM analysis [83–85]. As the grazing incidence angle of the ion beam used for sample thinning leads to milling rates that are largely material independent, FIB is especially useful for preparing specimen

**FIG. 2**

Recent advances throughout the entire electron microscopy workflow. For sample preparation, these include advances in focused ion beam and ultramicrotomy techniques (Section 2). Manipulating the electron dose and direct electron detectors present two opportunities while imaging to preserve structural details in HSI. Finally, post-processing techniques such as the use of virtual detectors or ptychography reconstructions present solutions for addressing these prevailing challenges (Section 3). Images from [58–61].

from heterogeneous hybrid samples [83]. Additionally, this methodology provides site-specific, highly uniform, thin sections, which are quite ideal for ensuing STEM imaging and EELS analysis [85]. Finally, through advances in cryogenic sample preparation, the soft constituents can be sectioned in their vitrified state, which is useful for minimizing the amount of beam damage present at a HSI [86–88].

The milling process, however, can introduce various sample artifacts. For instance, charged ions can cause surface amorphization or become preferentially implanted forming defects in the soft and/or hard components of the sample. This effect can, however, largely be reduced if a low energy (50 eV–1 keV) and low current milling process with gallium or argon ions is performed immediately afterwards [89–91]. Another concern is redeposition of atoms on the sample surface following removal by the ion beam. This can be largely reduced by carefully maintaining a beam current compatible with both hard and soft constituents at each step of the process as well as using a local barrier to preserve the area of interest [92,93]. Finally, FIB milling can be quite time consuming, which limits its utility in isolating macro inter-phase regions within hybrid samples.

3. Advanced STEM methods for analysis of soft/hard structures

Recent STEM developments present new opportunities for probing and analyzing hybrid structures and HSI. In the following sections, we will first discuss the advantages provided by direct electron detectors (DEDs), a major hardware advance. We will then discuss a variety of analytical methods enabled by this tech-

nology to enhance interfacial contrast while maintaining structural integrity.

3.1. Direct detectors: reduced threshold dose and improved contrast

The introduction of DEDs has revolutionized the understanding of nanoscale features in biological and soft material systems over the past decade [94,95,57]. DEDs have brought to bear massive improvements in both detection efficiency and noise floors by eliminating the need for electron-photon conversion via a traditional scintillator and fiber optical plate setup, substantially increasing the detective quantum efficiency [96–98]. It is now possible to image beam-sensitive materials with reduced electron beam fluxes, which has had profound implications for attaining atomic resolution information of highly sensitive samples, such as MOFs and the related chemical organic frameworks (COFs) [99–101].

The high speed, microsecond range readout of DEDs makes it a practical tool to record multidimensional datasets, such as spectral data or a convergent beam electron diffraction (CBED) pattern at each probe position [102–104]. The ability to record a 2D diffraction pattern at each 2D probe position produces a four-dimensional dataset, which is referred to as 4D-STEM. Because CBED patterns contain rich chemical and physical phase information about a sample, this technique unlocks vast structural information about a specimen while allowing for the reconstruction of traditional imaging modalities. 4D STEM has grown in concert with the popularity of DEDs and will be further discussed in Section 3.2 [105].

3.2. Advances in STEM acquisition and reconstruction methods: simulated & experimental data

In traditional STEM, physical detectors are used to selectively capture forward scattered electrons falling within a pre-defined angular range. Transmitted electrons displaced that are mostly forward scattered are captured by a circular detector to generate bright field (BF)-STEM images. Annular dark field (ADF)-STEM refers to a class of techniques that include high angle annular dark field (HAADF)-STEM, where images are generated using an annular detector to capture transmitted electrons scattered outside this bright field range, including . Electrons from the primary beam are scattered to large angles as a result of largely incoherent and elastic interactions with the sample.

In BF-STEM mode, electron signal is the result of the complex interplay between thickness, diffraction, and compositional effects. Conversely, in ADF-STEM mode, electron signal is dominated by elastic Rutherford scattering and is more easily interpretable [106]. Since the Coulomb interaction between atomic cores and incident electrons increases as the effective nuclear charge increases, ADF signal intensity is related to the atomic number (Z) of the constituent atoms in the sample. As a rule of thumb, ADF signal intensity is proportional to Z^α , where α lies between 1.2 and 1.8 depending on the sample, microscope conditions, and collection angle [107,108].

These imaging modes can be described in terms of contrast transfer functions (CTFs) or the relationship between the sample and the resulting image. In the case of non-linear imaging techniques such as BF-STEM, the relationship between the recorded image and the object functions is non-trivial due to the various contrast mechanisms discussed before, as shown by its CTF. In the case of a technique such as ADF-STEM with a more straightforward CTF, the image intensity is approximately a linear convolution of the object function with the electron probe. [109–111] Unfortunately, ADF-STEM is far less sensitive to low Z elements and leads to significant variation in ADF signal intensity for materials with disparate chemical compositions. As a result, this substantial image contrast limits the ability to see fine features within adjacent materials at this heterojunction.

Moreover, when the electron beam interacts with the sample, both its phase and amplitude are modified. For traditional imaging techniques, only the amplitude is captured, so much of the phase information is lost. To this end, there is a need for alternative more efficient imaging methods that are linear with the object transmission function and capture phase information. In the following sections, we discuss recent developments associated with a few of the most promising phase contrast imaging modalities that make it possible to simultaneously image heavy and light atoms.

As mentioned above, with a 4D-STEM dataset, BF and ADF images can be similarly generated by employing a user-defined "virtual" detector during post-imaging analysis. This virtual detector, or binary mask, is applied to each diffraction pattern in order to preferentially select reciprocal space data falling within a specified collection angle range. Instead of collecting a single intensity value at every pixel position with a traditional monolithic BF/ADF detector, the 4D-STEM approach enables retention of the relationship between probe position and scatter-

ing angle distribution. Thus with a 4D dataset, the microscopist has the ability to construct BF and DF images of various collection angles by varying the virtual detectors following the experimental session.

To highlight this point, we present a model hybrid composite system in Fig. 3 composed of nanoparticles (hard) integrated within an epoxy matrix (soft) from which a 4D-STEM dataset was collected. By constructing a circular mask to capture electron signal deflected within the bright field disk (0 to 3.5 mrad), we were able to create the BF-STEM images seen in Fig. 3A. Similarly, by creating an annular mask to capture electron signal deflected outside the bright field disk (3.5 to 25 mrad), the ADF image presented in Fig. 3B was produced. As ADF-STEM offers limited sensitivity to low Z elements, the large discrepancy in signal between the nanoparticles and the neighboring matrix in Fig. 3B makes it challenging to ascertain the fine features present at this interfacial region.

This approach can be taken a step further by tailoring collection radii to the scattering of various elements in a sample to create a map of different phases within a sample. Fig. 3E shows the results of this strategy employed on the same dataset from Fig. 3A-C overlaid on a bright field image. In this case, the ability to differentiate between elements is somewhat limited because there is overlap in the distribution of their scattering angles in reciprocal space and because the dataset is noisy, a constraint imposed by the soft epoxy matrix. Nonetheless, the relative consistency between the EDS map (Fig. 3F) and the phase map suggests that these methods can complement each other in identifying compositional heterogeneities. Additionally, forward scattering-based classifications offer advantages over EDS methods in that they demonstrate a high collection efficiency, which leads to faster data collection.

There is substantially more information than shown in Fig. 3 that can be obtained with 4D data. Because CBED patterns are quite sensitive, subtle changes in the pattern position and intensity on the detector can be related to the local lattice spacing [112–114]. This sensitivity makes it possible to identify variations in local structure and strain fields at HSI. We will discuss some of these techniques in later sections, which are also summarized in Table 1. Ophus et al. provides a comprehensive review of 4D STEM techniques [103].

3.2.1. Annular Bright Field (ABF) Imaging: a route to visualizing heavy and light atoms

In annular bright field (ABF)-STEM, an annular detector collects electrons scattered to the outer edge of the bright field disk [115,116]. This method is compatible either with physical or virtual annular detectors. Due to the relationship between electron channeling effects and electron intensity in this region of the bright field disk, this method allows for simultaneously observing both heavy and light elements as described through three generalized situations detailed by Findlay et al. For instance, in a situation where the electron probe is placed between adjacent atomic columns, a uniformly intense bright field disk with minimal dark field intensity would be generated. If instead this probe were aligned with a column of light elements, a greater proportion of electrons would be scattered to the dark field and the center-most bright field regions due to electron channeling

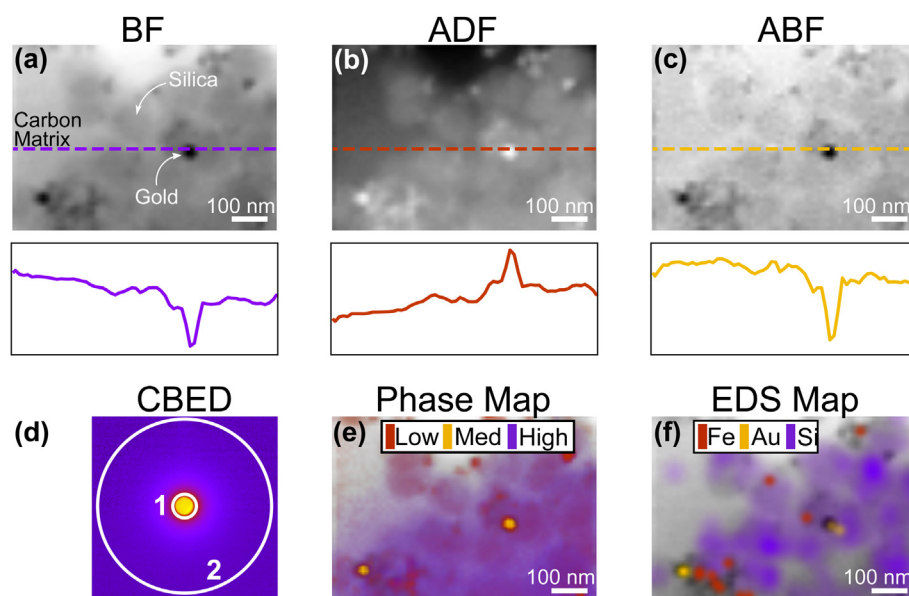


FIG. 3

Imaging of model hybrid composite system composed of Au, Fe₃O₄, and SiO₂ nanoparticles embedded in a carbon matrix with a probe convergence semi-angle of 3.5 mrad to achieve a Kossell-Möndstedt pattern, which provides good separation of disks for easier phase differentiation. (A) BF (<3.5 mrad, detector is area within ring 1 as shown in (D)). (B) ADF (3.5–25 mrad, detector is area outside the central disk and between ring 1 and 2 as shown in (D)). (C) ABF (2.3–3.5 mrad), detector is outer third of area within ring 1 (D)). Normalized line profiles in A–C show change in contrast and signal to noise. (D) CBED averaged across entire sample area. (E) Phase map superimposed on BF image. Color indicates sensitivity to phase defined by low (3.5–5.3 mrad), medium (5.8–7 mrad), and high (8.8–26 mrad) collection angle ranges using virtual detectors. Counts in arbitrary units. (F) EDS map superimposed on BF image. Color indicates normalized X-ray counts. EDS has a small collection angle, making it a dose inefficient method and leading to a sparser dataset that misses some of the nanoparticles. However, with a phase map it is more difficult to distinguish materials with a similar Z or amorphous components, such as in the case of SiO₂ and C.

TABLE 1

Discussion of advantages and limitations associated with various STEM acquisition and reconstruction techniques.

Technique	Advantages	Limitations
BF	Appreciable signal from soft materials	Multiple contrast transfer mechanisms limits analysis
ADF	Linear imaging technique makes it possible to quantitatively discern chemical structure	- Insensitive to low Z elements - Dose-inefficient method
ABF	Appreciable signal from soft and hard materials	- Presence of diffraction contrast sources can make it difficult to quantitatively discern chemical structure - Dose-inefficient method
DPC	- Appreciable signal from soft and hard materials - Dose-efficient method - Approximately linear imaging technique - Linear approximations can break down with increasing sample thickness	- Diffraction contrast can generate imaging artifacts - Requires fast electron detector or quadrant detector
Ptychography	- Appreciable signal from soft and hard materials - Dose-efficient method - Allows for improving spatial resolution beyond the resolution limit set by microscope lenses - Approximately linear imaging technique in the case of weak phase objects	- Typically requires specimen thickness on the order of few atomic layers - Can be computationally expensive depending on reconstruction algorithms - Requires fast electron detector for high resolution
MIDI	- Approximately linear imaging technique yields improved signal from lower spatial frequencies common in amorphous materials - Appreciable signal from soft and hard materials - Dose-efficient method	- Practical challenges with phase plate preparation and alignment - Requires fast electron detector for high resolution

effects, which leads to a reduction in electron intensity to the outer area of the bright field region. Similarly, if the probe were aligned with a column of heavy elements, a greater degree of electron scattering into the dark field region yields an overall

decrease throughout the entire bright field region [117]. While a more comprehensive description of image formation mechanisms that serve as the foundation for ABF signal in hard and soft materials is provided by other references [118,119], the intensity

variations in the outer area of the bright field region associated with this conceptual model suggests that unlike ADF, this method yields appreciable electron signal from both hard and soft materials [120].

From Fig. 3C, we find that this imaging modality does indeed boost signal and structural detail from the soft components in the sample (the epoxy matrix). An optimized collection angular range consisting of the outer third of the central disk produces substantial imaging contrast of these components within the sample [121,118,117]. This enhanced contrast has made ABF popular for imaging lithium ions and there exist several recent reports and review articles dedicated to this topic [122–126]. ABF has similarly made it possible to image carbon shells on metallic nanoparticles [127] and the presence of hydrogen atoms in a YH_2 crystal [128].

ABF presents limitations, however, when addressing HSI. For instance, because ABF signal intensity mainly arises due to coherently scattered electrons, variations in crystal orientation or strain make it challenging to quantitatively assess the chemical nature of various constituents [129]. Furthermore, as previously shown, the non-linear nature of this imaging modality leads to contrast reversals across specific thickness, tilt and defocus ranges [119,129]. Also, because the only electrons retained for image reconstruction are those that fall within a narrow band of polar scattering angles within the bright field disk, the signal-to-noise ratio (SNR) of this method lags other phase contrast imaging techniques, making this a dose inefficient method. Moreover, information associated with higher spatial frequencies is lost as well. One method to mitigate these issues would be to combine this signal with signal from incoherently scattered electrons as in the incoherent bright field (IBF)-STEM method. This method allows for imaging hybrid samples with thicknesses exceeding 100 nm [130,131]. Nonetheless, the collection of both coherent and incoherent electrons can produce data interpretation challenges, which are mitigated with emerging phase contrast techniques discussed in the following sections.

3.2.2. Differential phase contrast/first moment STEM: a means to detecting subtle phase shifts

Because the phase component of the sample transmission function can impart a physical shift on the beam illumination at the detector plane, another phase contrast approach would be to examine methods that characterize this subtle spatial variation. To this end, the similar methods proposed by Dekkers and de Lang, differential phase contrast (DPC), in 1974 and Waddell and Chapman, first moment STEM (FM-STEM), soon thereafter have recently received interest, as the introduction of DEDs have made these methods practically employable [132,133]. The former method involves measuring the difference in electron signal captured within opposite regions on a divided detector, such as quadrant detector, at each probe position to calculate deflections in the transmitted beam in x and y directions as a function of probe position. The latter method involves measuring the intensity center of mass (I_{COM}) of the beam illumination at the detector plane. In recent years, it has been proven that the momentum transfer that the electron probe experiences and calculated through FM-STEM is linearly related to the gradient of the phase of the specimen transmission function.[134] Mean-

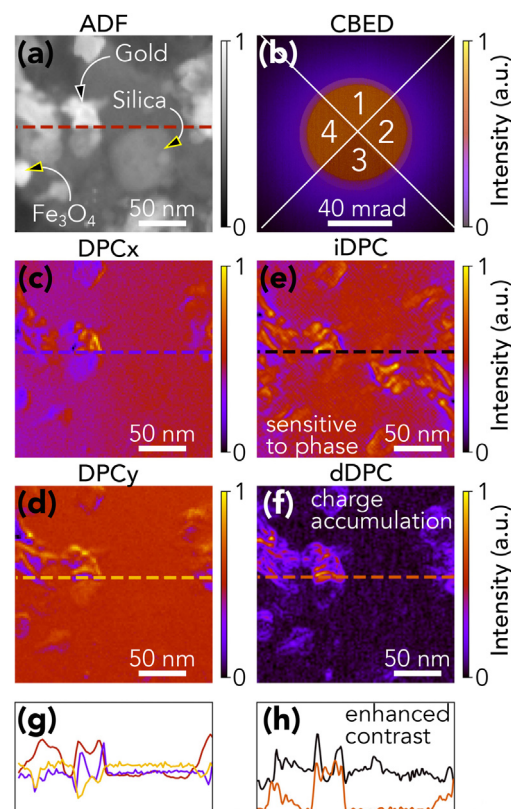


FIG. 4

DPC imaging of sample of Au, Fe_3O_4 , and SiO_2 nanoparticles embedded in a carbon matrix with a probe convergence semi-angle of 30 mrad to obtain to achieve optimal spatial resolution. The dotted lines in the image represent regions over which intensity line profiles were acquired. (A) ADF (3.5–25 mrad). (B) Quadrant detector employed to produce differential signal. Electrons captured in opposite detectors were subtracted from one another. (C) DPC_x (Detector 2 – Detector 4). (D) DPC_y (Detector 1 – Detector 3). The DPC images provide much greater contrast between the soft, lightly scattering components and the background. (E) iDPC image produced by integrating the DPC signal across the bright field disk. iDPC provides an accurate representation of the phase component imparted by the sample. (F) dDPC image produced by taking the divergence of the DPC signals. dDPC is proportional to the projected charge density within the sample. iDPC and dDPC both provide improved interfacial detail. (G) Normalized line profiles show change in contrast and signal to noise between the ADF, DPC_x , and DPC_y images. (H) Normalized line profiles show change in contrast and signal to noise between iDPC and dDPC images. The enhanced signal in both iDPC and dDPC at the edge of the nanostructures may result from the accumulation of charge or carbon buildup at the matrix/nanostructure interface.

while, the signal captured by DPC serves as a less computationally expensive, useful approximation for this gradient.

DPC images of hard and soft nanostructures embedded in an epoxy matrix are presented in Fig. 4. Compared to ADF, it is apparent that recovering the phase component without sacrificing signal in the bright field disk through DPC reduces image contrast between both hard and soft components. These images were constructed by filtering the 4D dataset with the virtual quadrant detectors seen in Fig. 4B. Recently Lazic et al., proposed a variant to these methods by showing that the integrated COM and DPC signals across the 2D bright field detector (iCOM and iDPC) are linearly related to the phase of the transmission function of the sample [134]. In the context of HSI, this purely phase

image has made it possible to identify low Z elements, such as O and even H [135–138]. The divergence of the COM and DPC signals, dCOM and dDPC, are proportional to the projected charge density within the sample and are particularly useful in the context of atomic resolution imaging and modeling electrostatic interactions at interfacial regions. iDPC and dDPC images of the nanoparticle–matrix sample are provided in Fig. 4E–F. As these methods produce appreciable signal from both hard and soft materials, a distinct improvement in relative contrast is evident in both images. The enhanced signal in both iDPC and dDPC at the edges of the nanostructures likely results from the accumulation of charge at the matrix/nanostructure interface that may result from carbon buildup during imaging as discussed previously by Lazic et al. [134].

These methods have emerged as a popular tool for imaging hybrid structures such as zeolites [139–141] and complex oxides [142–144]. Further, the dose-efficient nature of this method allows for obtaining adequate signal from sensitive materials within the sample's dose limit [145]. Although DPC and COM-based techniques offer significant advantages over ABF including dose efficiency and a CTF that is far easier to interpret, sources of diffraction contrast can also contribute to probe deflection and introduce imaging artifacts [146]. Additionally, although iDPC is a highly implementable approximation of iCOM signal, increases in thickness can cause this approximation to fail and generate contrast reversal effects in areas where the defocused probe interacts with the sample. This increase in plural and inelastic scattering with increasing sample thickness is responsible for this breakdown in the approximately linear relationship between iDPC contrast and the sample's transmission function [138,147]. Additionally, iDPC and iCOM remain largely empirical techniques and a comprehensive understanding of contrast mechanisms in different systems requires continued exploration.

3.2.3. Ptychography: a dose efficient method for phase retrieval

The original motivation to use electron ptychography in STEM was to improve spatial resolution beyond the limit set by the microscope lenses [148,105]. Nowadays, the most important outcome of this computational imaging method is that it produces a complex image encompassing both amplitude and phase of the exit electron wave after interaction with the sample [149].

In practical terms, ptychography retrieval methods are conducted by taking the Fourier transform of the CBED pattern captured at each position in real space. In the case where the resultant diffracted beams in the 4D dataset overlap one another, the interference pattern formed in the overlapping region contains both phase and amplitude information that can be distinguished from one another by integrating selected areas of this matrix [150,149]. There exist several algorithms for recovering the phase information with each offering various advantages depending on the materials complexity and electron dose. These include the extended ptychographic iterative engine (ePIE), an iterative Fourier ptychography method that starts from initial guesses for the probe and object functions followed by a series of forward scattering calculations to determine the probe and the exit-wave [151,152,105]. Additionally, non-iterative algorithms involving only Fourier transforms and deconvolutions,

such as Wigner-distribution deconvolution (WDD) method [153] and the single sideband (SSB) method [150], which assumes a weak phase object, have been developed. One advantage of WDD and SSB methods is the possibility of including residual aberrations to further tune the reconstruction quality [149,60].

Additionally, the application of extremely low electron doses (e.g. $< 1\text{e}^-/\text{\AA}^2$) still results in scattering events that are distributed across many pixels in the detector. Ptychography can be efficiently applied to retrieve these events and consequently reconstruct the object function by making use of the entire bright field region [153,150,154,155]. Defocusing the probe can also lower the total amount of electron dose applied to cover the region of interest by reducing the number of probe positions necessary [154]. Moreover, recent developments associated with focused probe ptychography make it possible to complement ptychography reconstructions with high resolution ADF images as well as EDS and potentially EELS maps to attain compositional information in addition to structural details of hybrid structures [60,156].

Thanks to advances with DEDs, electron ptychography has been employed on a wide variety of materials systems. Since applied to study silicon by [148], it has since been used for materials such as graphene [157], GaN [158], doped BiFeO₃ [158], halide perovskites [159], and complex carbon nanotube conjugates [60]. These studies highlight the advantages of ptychography for hybrid materials, as both heavy and light elements can be imaged simultaneously. Researchers are pushing the boundaries of how to more efficiently reconstruct information using lower doses for beam sensitive materials, such as with binary imaging or compressive sensing, which makes it useful for imaging HSI [160,161,154]. The ePIE method is very effective when using defocused probe datasets which allows further electron dose reduction and was recently used to reconstruct data obtained from biological samples at doses as low as $5.7\text{e}^-/\text{\AA}^2$ [162].

Here, to demonstrate the applicability and the benefits of ptychographic reconstruction in systems containing HSI, we utilize a model system of a gold/carbon interface. The 4D dataset was simulated as described in S1.4 without probe aberrations, and the ptychography reconstruction was performed using the SSB method [157]. In our example, we highlight two major characteristics that can be leveraged using electron ptychography: electron dose efficiency and resolution improvement. Fig. 5 shows how SSB ptychographic reconstructions of standard and low electron dose images can enhance structural details of both hard and soft components compared to ADF images. This is due to high efficiency information transfer across a broad spectrum of spatial frequencies in the case of ptychography as compared to ADF.

Nonetheless, there are some significant drawbacks for this method. For instance, ptychography reconstructions require caution when dealing with samples that violate the weak phase object condition and are likely to exhibit dynamical scattering effects. To overcome this issue, novel methods based on multi-slice ptychography as well as closely related techniques including optimum bright field have been developed [163,164]. Moreover,

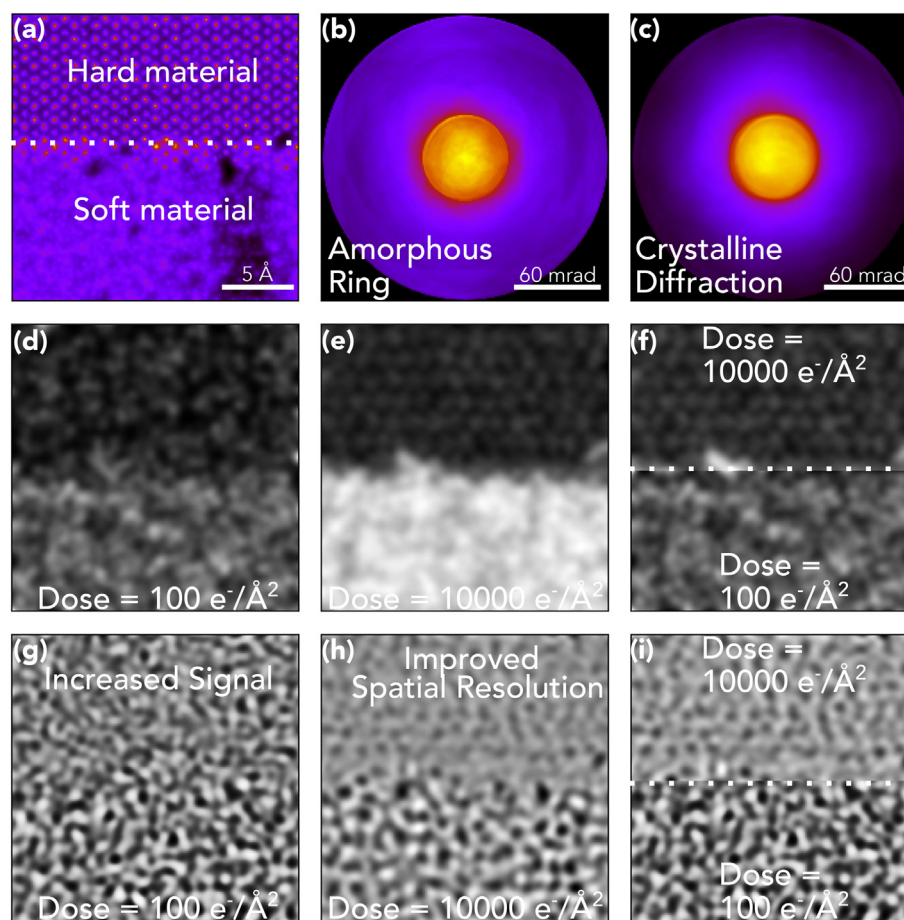


FIG. 5

STEM simulations of gold/carbon matrix interface: (A) The projected potential of the sample. (B-C) The corresponding simulated diffraction patterns taken from the hard (B) and soft (C) regions with a probe convergence semi-angle of 30 mrad to increase the level of overlap between diffracted disks and increase spatial resolution, respectively. (D-E) Simulated ADF images taken with a relatively low dose ($100 \text{ e}^-/\text{\AA}^2$) (D), and a higher dose ($10,000 \text{ e}^-/\text{\AA}^2$) (E). (F) Adaptive sampling method where the low dose is applied to the soft component and the higher dose is applied to the hard component to preserve its structure during ADF imaging. (G-H) Simulated ptychography reconstructions at these low (G) and high (H) dose values produce improved interfacial contrast and spatial resolution from the hard material region. (I) Similar adaptive sampling method applied in (F) followed by ptychography reconstruction.

generating reconstructions of the large 4D datasets commonly collected for ptychography can require significant computational power and storage space. This necessitates improvements in the STEM data workflow which we will discuss in Section 4.3.

3.2.4. Modifying incident electrons: phase plates

Phase plates, mostly in the form of the Zernike and the Volta geometries, are well known tools in cryo-TEM to enhance phase contrast in images without the need for a significantly defocused beam [165–167]. These plates impose a phase shift on the electron wave and as such, the resultant interference between transmitted and diffracted electron beams produces an intensity variation that can be linked to the phase component introduced by the sample. Phase plates have predominately been used with TEM techniques, but recent reviews have proposed opportunities for expanding their use [168–171,268,269]. There has been growing interest in using a similar approach in STEM to collect phase information by manipulating the shape of the electron beam, including creating vortices, concentric Fresnel rings, and bullseye patterns [112,114,172–174].

The rising popularity and availability of the direct detector in combination with the phase plate has led to further developments in imaging HSI. A near linear phase imaging technique called Matched Illumination Detector Interferometry (MIDI-STEM) incorporates a virtual detector whose geometry matches that of the Fresnel phase plate [61]. The strength of the MIDI-STEM technique is in its ability to image heavy and light elements at HSI due to its alternating ring geometry that enhances the transfer of low spatial frequency information within the allowable dose limits set by the soft material. A similar approach was used by Tomita et al., who inserted an amplitude Fresnel zone plate into the probe forming aperture of a microscope in a STEM configuration [176]. The phase component stored at high spatial frequencies of these images can be even further enhanced with the incorporation of both a pre-specimen phase plate and ptychography, as shown by PMIDI-STEM (Ptychography MIDI-STEM) [175]. Overall these phase plate techniques are powerful and their dose efficient nature allows for discerning fine features in hard and soft materials simultaneously.

Here we use simulations to demonstrate some of the advantages of using a combined phase plate, direct detector approach.

We focus on the step-edge junction that forms when MoTe₂ is stacked upon a graphene substrate such as has been demonstrated for photodetector applications [177].

For the purpose of this simulation, an electron beam was first convolved with a Fresnel phase plate at the probe forming aperture. This created a beam with rings of alternating phases of 0 and $\pi/2$. Following the interaction of the modified probe with the sample, the resultant 4D dataset was filtered through a virtual detector exactly matching the illumination pattern incident on the sample. Fig. 6A shows the phase of the control probe as compared to the modified probe in Fig. 6B. The CTF of the modified probe is shown in Fig. 6C and shows strong information transfer at low spatial frequencies.

Compared to the projected potential (Fig. 6D), the conventional dark field images (Fig. 6E) show the expected challenges with attaining sufficient electron signal from both carbon and the heavier Mo and Te atoms at the same time. However, the resulting image captured with a Fresnel plate in Fig. 6F shows significantly improved relative contrast between the hard and soft components. This is emphasized by the line profiles in Fig. 6G.

Although phase plates have the potential to be an incredibly useful imaging technique, there exist challenges associated with aligning these apertures in the microscope and characterizing the initial probe. Moreover, thin film phase plates are prone to carbon buildup over time, which changes the phase shift and decreases the signal to noise. Other phase modification techniques relying on lasers [178] or magnetic fields [179] are being developed to overcome these challenges.

3.3. Dynamic/sparse imaging: image reconstruction from under-sampled datasets

3.3.1. Compressive sensing: image reconstruction through inpainting

As described previously, the soft material sets an upper limit on dose at a HSI, placing restrictions on overall signal to noise. One approach that is used for minimizing the electron dose and damage that soft materials experience is to deliberately sparsely image a sample and then employ a compressive sensing technique to reconstruct a complete image [180]. Sparse imaging offers a host of advantages for hybrid samples, including a reduction in the time over which the electron interacts with the sample. Although, hard and soft materials display differences in terms of damage mechanisms, a decrease in overall dose uniformly preserves the sample.

Compressive sensing is a broad technique rooted in many imaging fields but lends itself especially well to STEM (and SEM) because of its sequential acquisition nature. Here, we will focus on inpainting electron microscopy images to infill missing portions of the under sampled dataset.

While theory papers often rely on virtual image reduction, a physical beam blanker or externally controlled scanning coils are employed in practice. The relative importance of various factors such as acquisition time, location precision, beam stability, and propensity for scanning distortions can dictate which method is used [181–183]. Researchers are continuing to test more exploratory methods for data acquisition, which inspire methods such as adaptive sampling that are particularly relevant for HSI as discussed in the next section. Likewise, there exist numerous approaches for reconstructing images from sparse

datasets that run the gamut from image filtering to advanced machine learning algorithms [183,184]. The simplest approach is interpolation using data from nearest neighbors [185]. However, many algorithms have been developed to improve on this tactic by including learned information about the sample. One common method is a beta factor process analysis (BPFA) approach [186], where a dictionary learning Bayesian model fills in missing pixels probabilistically from known elements for image restoration [182,187–189,185]. Additionally, there exist inpainting methods for suppressing the noise associated with undersampled data [182]. Recent work to develop more advanced approaches using convolutional neural networks or deep learning algorithms assign weights to particular features in an image in order to assist in identification and reconstruction [190].

In the context of heterogeneous hybrid samples, many of these compressive sensing techniques lend themselves especially well to this type of sample due to the ability to detect abrupt interfaces [191,58]. Moreover, compressive sensing can also be combined with the multimodal signals associated with STEM to interrogate interfaces with analytical STEM techniques, including EDS, EELS and cathodoluminescence [58,189,192]. This approach can also be used with 3D reconstructions, which will be discussed in more detail in Section 4.1. As with many advanced approaches, one limitation of this technique is the off-line computation time needed to collect and analyze data. Despite the many benefits of inpainting, especially for materials with a beam sensitive component, this technique does not fundamentally improve contrast between hard and soft materials. One way to improve the relative contrast with this method is with a smart dwell time approach, which is discussed in the next section.

3.3.2. Adaptive dwell time: intelligent sampling of the specimen

One of the main challenges in imaging HSI is that the soft material sets an upper limit for the dose that can be applied and the SNR that can be generated from the hard material. Scanning electron microscopy techniques have the advantage that they can decouple acquisition parameters with spatial position, making it possible to apply different doses to different areas of the sample depending on the local beam sensitivity. An example is shown in Fig. 5F, and it is possible to imagine how such a scheme can be used in combination with ptychography (Fig. 5I) or inpainting (discussed below).

This concept of adaptive dwell time has received recent interest in the microscopy community [193]. For example Timischl used a dynamic dwell time in a SEM based on signal statistics, which ultimately decreases dwell time for brighter pixels [194]. Another SEM method uses a two pass system, where the initial sample area is evaluated and then areas with high spatial frequency information are scanned again for more detailed analysis [195]. In an atomic resolution STEM setup Stevens et al., used a similar adaptive sampling strategy where they applied an increased dose to regional maxima in a ZnSe sample and generated atomic resolution with doses on the order of $10 \text{ e}^-/\text{\AA}^2$ [183]. An adaptive dwell time approach has also been demonstrated for EELS and EDS data collection with a multi-objective autonomous dynamic sampling (MOADS) method. This on-

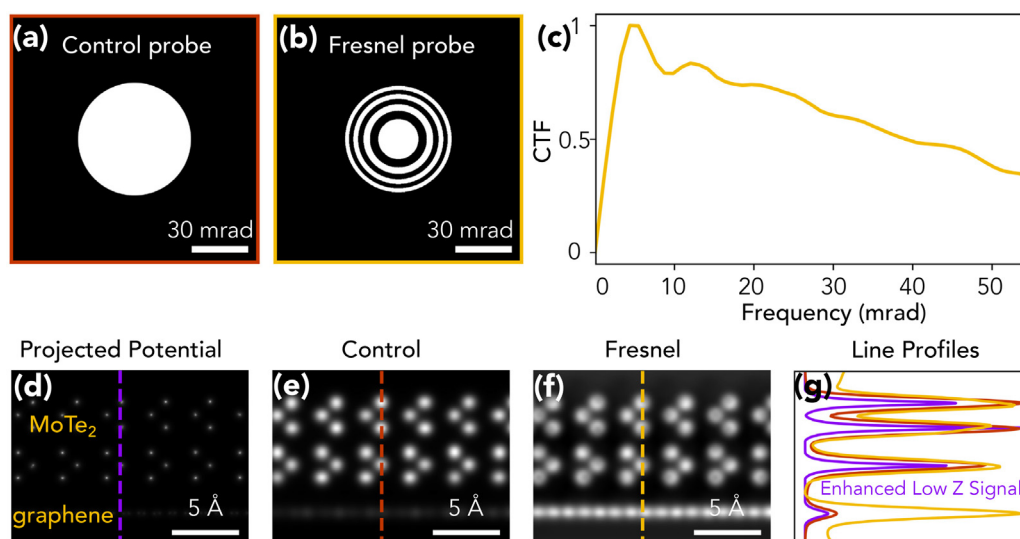


FIG. 6

Phase plate study of a molybdenum ditelluride/graphene layered heterostructure: Phase of the (A) control and (B) Fresnel probe before sample interaction. (C) The CTF for the modified probe. (D) Projected potential of MoTe₂/graphene sample. Conventional (E) dark field (35–120 mrad) image of the area. (F) Image of same region with Fresnel probe, showing improved contrast between heavy and light elements. (G) line profile comparing contrast from (D)–(F).

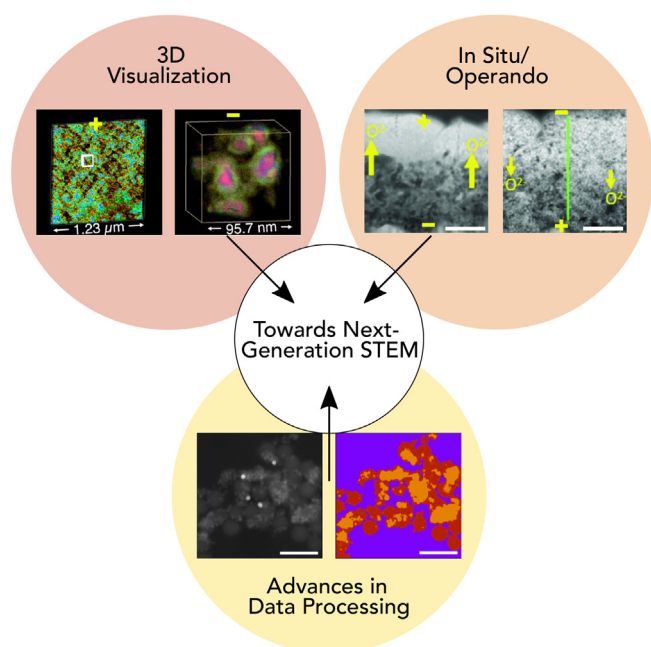


FIG. 7

A subset of emerging opportunities in various parts of the STEM workflow: 3D reconstruction using tomographic methods, *in situ/operando* analysis, and advances in the data processing pipeline. Examples include quantitative, three-dimensional imaging of chromatin structure [232], observing electrically induced oxygen diffusion in inorganic/organic halide perovskites [233], and development of machine learning algorithms for rapid classification of image features.

the-fly dynamic approach reduces sample acquisition time and beam radiation while producing detailed elemental maps [58].

These dynamic sampling techniques are far from routine practice, yet it is clear how they will be beneficial for hybrid materials. With the combination of interface detection, intelligent sparse data collection, adaptive dwell time, and inpainting, hybrid materials can be carefully characterized.

4. Emerging opportunities & outlook

To this point, we have discussed how a variety of recent advances associated with sampling and detection of STEM signals have enabled high-resolution imaging of HSI. Although we have mainly focused on 2D projections, these concepts can be extended further through the use of electron tomography to interrogate complex 3D systems. They can also be applied in tandem with *in situ/operando* STEM methods to understand the behavior of these interfaces when stimulated. Although these methods have the potential to provide a much more complete understanding of an HSI than conventional techniques, these investigations often generate large multidimensional datasets and can rely on computationally expensive reconstructions. These practical considerations are critical for widespread implementation of these methods. We conclude this section by discussing recent advances and best practices related to data management. An overview of this section is presented in Fig. 7.

4.1. 3D reconstruction of hard/soft interfaces: electron tomography

STEM images represent a 2D projection of a 3D structure, which in addition to creating complications in imaging and diffraction analysis, fundamentally results in missing information [106]. There are many techniques to reconstruct 3D information associated with a specimen, such as the use of STEM tomography for understanding the structure and properties of intricate nanostructures [196,197]. This technique involves capturing 2D images at a wide range of tilt angles, which are used to reconstruct a 3D representation of a region of interest.

The captured electron signal for tomography must meet the projection requirement such that it is a monotonic function that scales with a physical property of the system, introducing key questions about contrast that have been discussed throughout this review [198]. BF-STEM signal meets this criteria in the case of an amorphous material as mass thickness serves as the main

contrast mechanism [199]. When a crystalline component is introduced, however, the projection criteria is no longer fulfilled due to the presence of a diffraction contrast term within the BF signal [200]. In this case, ADF signal is the natural alternative since Z contrast meets the projection requirement [201].

Moreover, various groups have recently demonstrated that ADF signal can be used as the basis for atomic electron tomography (AET), a class of techniques that provides three-dimensional structural information from crystalline and amorphous materials with atomic resolution [202–206]. By pairing new iterative algorithms with an aberration corrected STEM using a direct electron detection scheme, an unprecedented level of spatial identification is now achievable, such as the spatial identification of defects, including grain boundaries, dislocations, and vacancies [202–204,207].

ADF electron tomography reconstructions of hybrid composites suffer from the same contrast challenges detailed in Section 3.2, making it difficult to examine materials with both heavy and light elements. As such, tomographic reconstructions employing advanced phase contrast techniques are needed to thoroughly understand interfacial morphology. Recently, a phase contrast atomic resolution tomography technique using high resolution TEM has been demonstrated [208]. This approach provides the ability to identify the location of light atoms, such as lithium, carbon, and oxygen in three dimensions. This TEM technique suggests that STEM methods described earlier, such as ptychography or use of a phase plate, combined with 3D tomographic reconstructions, can provide rich atomic scale information about hybrid materials [209].

Due to limitations in the number of angular projections that can be acquired and the maximum tilt angle (70°) attainable, tomography produces undersampled data. Although algorithms such as weighted back projection (WBP) and simultaneous iterative reconstruction technique (SIRT) have made it possible to mitigate the missing wedge effect and reconstruct a wide variety of hard and soft materials [210], they tend to be quite susceptible to streaking artifacts and blurring in the direction of the missing angular range. In recent years, similar CS algorithms to those discussed previously have been applied to retrieve the optimal undersampled dataset. Through non-linear compressive sensing electron tomography (CS-ET) algorithms, such as total variation minimization (TVM), BPFA, or 3D wavelet inpainting [211–214], promising results have been demonstrated. These algorithms have been able to effectively inpaint the missing angular range and reduce the presence of blurring artifacts [215,212,216,217,211]. Additionally, these methods have proven to deliver high fidelity reconstructions with greater definition from structures such as nanoparticles, than those constructed using traditional reconstruction techniques such as WBP or SIRT, while requiring fewer projections [216]. This, of course, is quite attractive due to the beam sensitive nature of many soft materials. As a result, these algorithms offer another route in addition to traditional cryo-tomography for preserving HSI [218,219].

Additionally, the sampling strategy itself can be varied by continuously rotating the sample in controlled rotational tomography (CORT) to create a sparsely sampled projection. Using CORT in conjunction with aforementioned CS-ET techniques

Li et al., were able to create reconstructions of beam-sensitive samples that were highly consistent with the ground truth structures [220].

Alternatively, a serial defocus approach can provide 3D images of thicker samples, including hybrid nanocomposites [221]. Although ptychography is typically limited to ultra-thin samples, Gao et al. recently demonstrated that an inverse multi-slice ptychography method yields the complex 3D transmission function of a thick sample. This alternative is useful as the presence of multiple scattering events can lead to the captured signal violating the projection requirement [222]. Because ptychography is a low dose method, this approach allows for reconstruction of beam-sensitive structures with minimal loss of 3D resolution [223].

Single particle analysis (SPA), a popular method in the cryo-EM community for analyzing and building 3D reconstructions of biological molecules, is yet another technique that may be valuable in the context of imaging HSI [224–227]. This technique involves imaging, classifying, and stitching together many identical molecules with different orientations using a class averaging approach. SPA can be performed with TEM or STEM, but the simpler contrast transfer function in STEM modes means fewer samples are needed for a reconstruction. This method is promising for understanding the interfacial structure in systems such as functionalized nanoparticles or MOFs, where identical geometries are readily accessible [228,229]. Similar constructs combining the STEM-ADF and spectral signals have also received recent interest [230,231].

4.2. *In Situ/Operando STEM: implications for soft/hybrid interfaces*

The capability to detect a bevy of signals from highly localized volumes with microsecond temporal resolution makes STEM an incredibly useful tool for probing real time phenomena. During an *in situ* experiment, an external stimuli is applied to a system, and the cascading effects are monitored. It can be difficult to implement these experiments in practice, as the external stimulation mechanisms need to be compatible with the high vacuum environment and electron dose common to STEM. Recent advances in specialized holders have created opportunities for studying the impact of heating [234], mechanical deformation [235] optical stimulation [233], electrical biasing [236,237] as well as the impact of liquid environments [238,239].

This type of study is closely linked to the previous discussion of HSI, as changes often occur at the interfacial region between two materials. Moreover, the structural dynamics of interest are commonly accompanied by the migration of light atoms. There exist numerous papers demonstrating electrochemical diffusion of lithium and the structural evolution this induces at the electrode/electrolyte interface [240,241]. Similarly, optical stimulation can induce oxygen migration from the electron transport layer of the photovoltaic cell into the inorganic/organic halide perovskite active layer [233]. Oxygen evolution and reincorporation has also been found to play a key role in explaining hysteretic behavior in oxide-based resistive random-access memory (ReRAM) [242]. Additionally, in liquid cell STEM, the spatial resolution achievable is limited by the SNR, thus achieving appreciable contrast from these light atoms becomes even more

important [238]. Directly imaging these atoms with phase contrast techniques would simplify the resultant analysis and give greater insight into the presence of intermediary steps during this reaction process.

As *in situ* experiments commonly require extended electron exposures, DEDs and the dose-efficient methods described previously are quite valuable in limiting sample damage throughout this time frame. Through the high frame rates possible with DEDs as well as the ability to capture a series of sub-frames and align them together, it is possible to record dynamical processes, such as deformation during tensile loading [243] or translational motion of a catalytic nanoparticle with millisecond temporal resolution and improved SNR [244–246]. Because information related to internal fields is captured in the phase component of the specimen transmission function, techniques such as differential phase contrast provide the ability to spatially characterize the in-plane electric or strain field responsible for the structural evolution [243,247]. Together these developments make it significantly easier to identify the impact of various stimuli on a nanoscale system.

4.3. Practical data acquisition, processing, and handling advances to improve the STEM workflow

The rise of artificial intelligence ecosystems and associated machine learning algorithms has accelerated innovation in a wide variety of scientific disciplines including materials discovery [248]. Artificial intelligence has already begun to play an important role in enhancing understanding of materials through electron microscopy [249–252,193,253,254,190], and we expect that in the coming years, the latest data analysis tools and techniques will revolutionize electron microscopy in ways that leave it better positioned to address major materials challenges. In this section, we discuss the potential impact that advances in this area such as real-time data processing, automated microscopy modules, and improvements in data storage and processing workflows would have in the context of understanding HSI.

4.3.1. Automated microscopy modules for streamlining data acquisition and analysis

Just as other characterization methodologies such as X-ray crystallography have become increasingly automated in recent years [255], the development of automated microscopy workstation would similarly streamline the data acquisition and analysis processes. We envision a paradigm where the microscopist would first image a few relevant and interesting regions, such as HSI, before leaning on a machine learning algorithm to explore an extensive worldwide microscopy database and procure the best course of action for further analysis. This would include defining a design space of possible microscope parameters that would be optimized for the particular tool through an auto-alignment procedure. It would also include identifying a set of ideal microscopy techniques for most effectively interrogating the sample of interest. This latter aspect could eliminate the implicit biases researchers may exhibit towards techniques and methodologies with which they are most familiar. By building a deeper connection between the human user and artificial intelligence, STEM is positioned to become a highly sought-after tool for probing and understanding intricate structure–property relationships.

4.3.2. Real-time processing of STEM data to obtain optimal datasets

While the current workflow for constructing STEM phase contrast images from 4D-STEM datasets provides a high level of understanding of interfacial features, these methods often requires substantial offline processing time. Beyond practical value associated with reducing acquisition and analysis time, having real-time data processing, on-the-fly imaging allows for acquisition of better datasets. This construct would allow microscopists to more directly find optimal experimental conditions and regions of interest. The imaging techniques discussed here each have different sets of ideal conditions forenhancing information transfer. With conventional techniques such as BF and ADF, these ideal conditions for information transfer are more readily identifiable than with phase techniques such as DPC or ptychography. The recent advent of detectors with live processing modules will allow for real-time processing of various phase contrast techniques [256,257]. This type of processing couples nicely with the automated engine discussed in the previous section to dynamically adjust to unexpected observations during a single microscope session.

4.3.3. Standardized framework for microscopy data management and processing

Finally, a standardized approach based on the FAIR (Findable, Accessible, Interoperable, Reusable) guidelines for storing large datasets can dramatically accelerate materials understanding [258]. A standardized naming scheme can avoid headaches for the microscopist following a completed session and is valuable in helping computational algorithms draw trends between sample properties and experimental conditions [259].

Accessing microscopy data is also a challenge as the sheer volume of generated data makes USB hard drives an impractical storage solution. Even cloud storage services may be impractical depending on the data transfer rate provided for transporting files from the acquisition computer to the data cloud. One potential solution would be to create a centralized storage location within a local high-performance computing cluster that users would be able to access for subsequent analysis [260].

The captured data can be made more interoperable through continued development of open-source scripts and applications for processing data. These tools can enhance the scientific accuracy of analysis as the user is privy to all data processing steps. Although there exist a variety of applications for performing particular analytical routines, we hope to see a continued development of computational ecosystems such as py4dSTEM [261], pyXEM [262], LiberTEM [263], pycroscopy [264], pixStem [265] and hyperspy [266], etc. that serve as “one-stop shops” compatible with high performance computing clusters for streamlining analysis. The acceptance of open, hierarchical data formats that allow access to data subsets without having to store the entire dataset in RAM and support compression, such as the sparse HDF5 format, would increase interoperability as well.

Finally, data reusability requires microscopists to publish experimental datasets in repositories and is a key step to enable an automated microscopy engine. Moreover, it serves as a way for the field-at-large to perform quality control and maintain scientific integrity beyond traditional peer review.

5. Summary

Hybrid composites are a compelling class of materials offering considerable opportunity across a wide array of applications. Moreover, to facilitate further development in this field, it is crucial to understand the chemical and physical properties of these composites, especially at their interfaces. In this article, we have discussed the challenges associated with achieving sufficient image contrast in STEM, while preserving structural integrity, when analyzing the interfacial regions between hard and soft components. Enabled by recent advances, we have identified a number of solutions for mitigating these concerns and attaining a nano or atomic-level understanding of these interfaces. Furthermore, by combining these STEM solutions with tomography and *in situ/in operando* methods, rich structure and property information is realizable as well. As such, the multimodality of STEM represents a powerful method for understanding and enhancing the functionality and performance of this emerging class of composite materials.

Data availability

The raw/processed data required to reproduce these findings cannot be shared at this time due to technical or time limitations. The datasets are now available here: <https://zenodo.org/record/4901550#.YLqWFpNKhpQ>

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <https://doi.org/10.1016/j.mattod.2021.05.006>.

References

- [1] M.J. Glimcher, *Med. Mineral. Geochem.* 64 (2006) 223–282.
- [2] K.A. Hing, *Trans. R. Soc. A* 362 (2004) 2821–2850.
- [3] C.P. Lin, W.H. Douglas, S.L. Erlandsen, *J. Histochem. Cytochem.* 41 (1993) 381–388.
- [4] S.J. Marshall et al., *J. Eur. Ceram. Soc.* 23 (2003) 2897–2904.
- [5] K.A. DeRoche et al., *Nature* 583 (2020) 66–71.
- [6] X.D. Li et al., *Nano Lett.* 4 (2004) 613–617.
- [7] X. Li, Z.H. Xu, R. Wang, *Nano Lett.* 6 (2006) 2301–2304.
- [8] G. Mittal, K.Y. Rhee, V. Miskovic-Stankovic, D. Hui, *Compos. Part B* 138 (2018) 122–139.
- [9] M.S. Asl et al., *Ceram. Int.* 44 (2018) 7334–7348.
- [10] K. Saha et al., *Chem. Rev.* 112 (2012) 2739–2779.
- [11] E.C. Dreaden et al., *Chem. Soc. Rev.* 41 (2012) 2740–2779.
- [12] Q.Y. Lin et al., *Science* 359 (2018) 669–672.
- [13] R.J. Macfarlane et al., *Science* 334 (2011) 204–208.
- [14] V. Nandwana et al., *ACS Appl. Nano Mater.* 1 (2018) 1949–1958.
- [15] Q. Yang et al., *Angew. Chem., Int. Ed. Engl.* 55 (2016) 3685–3689.
- [16] C.H. Kuo et al., *J. Am. Chem. Soc.* 134 (2012) 14345–14348.
- [17] G. Lu et al., *Nat. Chem.* 4 (2012) 310–316.
- [18] S. Chen et al., *ACS Nano* 4 (2010) 2822–2830.
- [19] J.W. Gilman et al., *Chem. Mater.* 12 (2000) 1866–1873.
- [20] X.Y. Zhang et al., *J. Mater. Chem.* 20 (2010) 2801–2806.
- [21] L. Jia et al., *J. Phys. Chem. C* 115 (2011) 11466–11473.
- [22] V. Nandwana et al., *Ind. Eng. Chem. Res.* 59 (2020) 10945–10954.
- [23] Y. Li et al., *Nano Lett.* 18 (2018) 7104–7110.
- [24] Y. Li et al., *ACS Nano* 11 (2017) 10321–10329.
- [25] D.W. Wang et al., *ACS Nano* 3 (2009) 1745–1752.
- [26] M. Kulbak, D. Cahen, G. Hodes, *J. Phys. Chem. Lett.* 6 (2015) 2452–2456.
- [27] S. Gong et al., *Adv. Mater.* 32 (2020). e1902278.
- [28] R. Yang, G. Chen, M.S. Dresselhaus, *Nano Lett.* 5 (2005) 1111–1115.
- [29] D. Ciprari, K. Jacob, R. Tannenbaum, *Macromolecules* 39 (2006) 6565–6573.
- [30] Y. Wang et al., *Compos. Sci. Technol.* 162 (2018) 146–155.
- [31] H. Zhang et al., *Acta Mater.* 54 (2006) 1833–1842.
- [32] S. Zhandarov, E. Mader, *Compos. Sci. Technol.* 65 (2005) 149–160.
- [33] F.S. Hage et al., *Phys. Rev. Lett.* 122 (2019). 016103.
- [34] F.S. Hage et al., *Sci. Adv.* 4 (2018) eaar7495.
- [35] O.L. Krivanek et al., *Nature* 464 (2010) 571–574.
- [36] X.Z. Yu et al., *Nature* 465 (2010) 901–904.
- [37] J.A. Scholl, A.L. Koh, J.A. Dionne, *Nature* 483 (2012) 421–427.
- [38] J.C. Meyer et al., *Nat. Mater.* 10 (2011) 209–215.
- [39] F. Wang et al., *J. Am. Chem. Soc.* 133 (2011) 18828–18836.
- [40] P. Yin et al., *Angew. Chem., Int. Ed. Engl.* 55 (2016) 10800–10805.
- [41] J. Mayer et al., *MRS Bull.* 32 (2007) 400–407.
- [42] R.F. Egerton, *Micron* 119 (2019) 72–87.
- [43] R.F. Egerton, *Ultramicroscopy* 127 (2013) 100–108.
- [44] D. Zhang et al., *Science* 359 (2018) 675–679.
- [45] Y. Yu et al., *Nano Lett.* 16 (2016) 7530–7535.
- [46] Q. Chen et al., *Adv. Mater.* 32 (2020) 1907619.
- [47] D. Zhang et al., *Science* 359 (2018) 675–679.
- [48] R. Henderson, *Q. Rev. Biophys.* 28 (1995) 171–193.
- [49] R.F. Egerton, P. Li, M. Malac, *Micron* 35 (2004) 399–409.
- [50] K. Urban, *Phys. Status Solidi A* 56 (1979) 157–168.
- [51] H.P. Erickson, *Biol. Proced. Online* 11 (2009) 32–51.
- [52] S. Mikula, W. Denk, *Nat. Methods* 12 (2015) 541–546.
- [53] S. Mikula, J. Binding, W. Denk, *Nat. Methods* 9 (2012) 1198–1201.
- [54] X. Li et al., *Nat. Methods* 10 (2013) 584–590.
- [55] L.E. Franken et al., *Small* 16 (2020) 1906198.
- [56] A.C. Walls et al., *Nature* 531 (2016) 114–117.
- [57] S.Q. Zheng et al., *Nat. Methods* 14 (2017) 331–332.
- [58] K.A. Hujak et al., *Micron* 108 (2018) 31–40.
- [59] S. de Graaf et al., *Sci. Adv.* 6 (2020) eaay4312.
- [60] H. Yang et al., *Nat. Commun.* 7 (2016) 12532.
- [61] C. Ophus et al., *Nat. Commun.* 7 (2016) 10719.
- [62] G.L. Wang, R.W. Murray, *Nano Lett.* 4 (2004) 95–101.
- [63] A. Kumar et al., *Adv. Mater.* 13 (2001) 341–344.

- [64] E.L. Que et al., *Integr. Biol.* 9 (2017) 135–144.
- [65] K. Choudhuri et al., *Nature* 507 (2014) 118–123.
- [66] X. Zhang et al., *Microsc. Microanal.* 25 (2019) 831–839.
- [67] F. Zhang et al., *Chem. Mater.* 32 (2020) 4292–4302.
- [68] N.W. Kennedy et al., *PLoS One* 15 (2020). e0226395.
- [69] S. Wang et al., *Nat. Commun.* 11 (2020) 2495.
- [70] S.A. Thomas et al., *Environ. Sci. Technol.* 52 (2018) 10030–10039.
- [71] D. Studer, H. Gnaegi, *J. Microsc.* 197 (2000) 94–100.
- [72] J. Ayache et al., *Media 2* (2010).
- [73] Q. Wu et al., *Chin. J. Aeronaut.* 28 (2015) 1529–1538.
- [74] H. Mucha et al., *J. Electron. Microsc.* 54 (2005) 43–49.
- [75] M.G. Huson et al., *Mater. Chem. Phys.* 168 (2015) 193–200.
- [76] S. Kim, V.P. Dravid, *J. Microsc.* 198 (2000) 124–133.
- [77] L.A. Giannuzzi et al., *Int. J. Oral Surg.* 65 (2007) 737–747.
- [78] S. Younis et al., *Cryst. Growth Des.* 12 (2012) 4574–4579.
- [79] T. Supasai et al., *Sol. Energy* 136 (2016) 515–524.
- [80] Y. Lu et al., *ACS Appl. Mater. Interfaces* 11 (2019) 12819–12829.
- [81] K. Grandfield, A. Palmquist, H. Engqvist, *Ultramicroscopy* 127 (2013) 48–52.
- [82] X.L. Zhong et al., *Microsc. Microanal.* 22 (2016) 1350–1359.
- [83] J. Mayer et al., *MRS Bull.* 32 (2007) 400–407.
- [84] M.W. Phaneuf, *Micron* 30 (1999) 277–288.
- [85] L.A. Giannuzzi, F.A. Stevie, *Micron* 30 (1999) 197–204.
- [86] J. Mahamid et al., *J. Struct. Biol.* 192 (2015) 262–269.
- [87] S. Rubino et al., *J. Struct. Biol.* 180 (2012) 572–576.
- [88] M.J. Zachman et al., *Nature* 560 (2018) 345–349.
- [89] Z. Huang, *J. Microsc.* 215 (2004) 219–223.
- [90] B.M. Tracy, R.H. Doremus, *J. Biomed. Mater. Res.* 18 (1984) 719–726.
- [91] Y. Zheng et al., *Scripta Mater.* 116 (2016) 49–52.
- [92] C. Du, J.P.M. Hoefnagels, L.I.J.C. Bergers, *Exp. Mech.* 57 (2017) 1249–1263.
- [93] T. Vermeij, E. Plancher, C.C. Tasan, *Ultramicroscopy* 186 (2018) 35–41.
- [94] Z. Wang et al., *Nat. Commun.* 5 (2014) 4808.
- [95] J. Park et al., *Science* 349 (2015) 290–295.
- [96] M. Battaglia et al., *Nucl. Instrum. Methods Phys. Res., Sect. A* 608 (2009) 363–365.
- [97] G. McMullan et al., *Ultramicroscopy* 109 (2009) 1126–1143.
- [98] G. McMullan et al., *Ultramicroscopy* 147 (2014) 156–163.
- [99] Y. Zhu et al., *Nat. Mater.* 16 (2017) 532–536.
- [100] K.W. Nam et al., *Nat. Commun.* 10 (2019) 4948.
- [101] D. Zhang et al., *Science* 359 (2018) 675–679.
- [102] C. Ophus et al., *Microsc. Microanal.* 20 (2014) 62–63.
- [103] C. Ophus, *Microsc. Microanal.* 25 (2019) 563–582.
- [104] B. Plotkin-Swing et al., *Ultramicroscopy* 217 (2020). 113067.
- [105] Y. Jiang et al., *Nature* 559 (2018) 343–349.
- [106] C.B. Carter, D.B. Williams, *Transmission electron microscopy: Diffraction, imaging, and spectrometry*, Springer, 2016.
- [107] Z.W. Wang et al., *Phys. Rev. B* 84 (2011). 073408.
- [108] P. Nellist, S. Pennycook, *Adv. Imaging Electron Phys.* 113 (2000) 147–203.
- [109] E.J. Kirkland, *Advanced Computing in Electron Microscopy*, Springer, 2020.
- [110] E. Yucelen, I. Lazic, E.G.T. Bosch, *Sci. Rep.* 8 (2018) 2676.
- [111] J.M. Cowley, *Ultramicroscopy* 2 (1976) 3–16.
- [112] S.E. Zeltmann et al., *Ultramicroscopy* 209 (2020). 112890.
- [113] T.C. Pekin et al., *Ultramicroscopy* 176 (2017) 170–176.
- [114] X. Hong, S.E. Zeltmann, B.H. Savitzky, L.R. DaCosta, A. Mueller, A.M. Minor, K. Bustillo, Ophus C. Multibeam, *Microsc. Microanal.* 27 (2021) 129–139.
- [115] H. Rose, *Optik* 39 (1974) 416–436.
- [116] S.D. Findlay et al., *Appl. Phys. Lett.* 95 (2009). 191913.
- [117] S.D. Findlay et al., *Microscopy (Oxford, U.K.)* 66 (2017) 3–14.
- [118] E. Okunishi et al., *Microsc. Microanal.* 15 (2009) 164–165.
- [119] S.D. Findlay et al., *Ultramicroscopy* 110 (2010) 903–923.
- [120] P. Gao et al., *Ultramicroscopy* 184 (2018) 177–187.
- [121] R. Ishikawa et al., *Nat. Mater.* 10 (2011) 278–281.
- [122] Y. Oshima, S. Lee, K. Takayanagi, *Microscopy (Oxford, U.K.)* 66 (2017) 15–24.
- [123] S.D. Findlay et al., *Microscopy (Oxford, U.K.)* 66 (2017) 3–14.
- [124] Y.X. Tong, Q.H. Zhang, L. Gu, *Chin. Phys. B* 27 (2018). 066107.
- [125] S. Lee et al., *Ultramicroscopy* 125 (2013) 43–48.
- [126] L. Gu et al., *J. Am. Chem. Soc.* 133 (2011) 4661–4663.
- [127] W.-H. Lai et al., *Adv. Funct. Mater.* 29 (2019) 1807340.
- [128] R. Ishikawa et al., *Nat. Mater.* 10 (2011) 278–281.
- [129] D. Zhou et al., *Ultramicroscopy* 160 (2016) 110–117.
- [130] P. Ercius, D. Muller, *Microsc. Microanal.* 15 (2009) 238–239.
- [131] P. Ercius et al., *Appl. Phys. Lett.* 88 (2006). 243116.
- [132] N. Dekkers, H. De Lang, *Optik* 41 (1974) 452–456.
- [133] E. Waddell, J. Chapman, *Optik* 54 (1979) 83–96.
- [134] I. Lazic, E.G.T. Bosch, S. Lazar, *Ultramicroscopy* 160 (2016) 265–280.
- [135] E.G.T. Bosch, I. Lazic, S. Lazar, *Microsc. Microanal.* 22 (2016) 306–307.
- [136] J. Bak, H. Bin Bae, S.Y. Chung, *Nat. Commun.* 10 (2019) 2713.
- [137] E. Yucelen, I. Lazic, E.G.T. Bosch, *Sci. Rep.* 8 (2018) 2676.
- [138] S. de Graaf et al., *Sci. Adv.* 6 (2020). eaay4312.
- [139] L. Liu et al., *Nat. Mater.* 18 (2019) 866–873.
- [140] A. Carlsson et al., *Microsc. Microanal.* 24 (2018) 122–123.
- [141] N. Kosinov, E.J.M. Hensen, *Adv. Mater.* 31 (2020). e2002565.
- [142] A. Kumar, et al., *arXiv preprint arXiv:1911.05853*.
- [143] D. Song et al., *Adv. Funct. Mater.* 29 (2019) 1903843.
- [144] W. Li et al., *Adv. Funct. Mater.* 30 (2020) 2001984.
- [145] I. Lazic, E.G. Bosch, In *Chapter Three – Analytical Review of Direct Stem Imaging Techniques for Thin Samples*, in: P.W. Hawkes, (Ed.), *Advances in Imaging and Electron Physics*, Elsevier, 2017, vol. 199, pp 75–184.
- [146] I. MacLaren et al., *Ultramicroscopy* 154 (2015) 57–63.
- [147] E.G.T. Bosch, I. Lazic, *Ultramicroscopy* 207 (2019). 112831.
- [148] P.D. Nellist, B.C. McCallum, J.M. Rodenburg, *Nature* 374 (1995) 630–632.
- [149] C.M. O’Leary et al., *Ultramicroscopy* 221 (2021). 113189.
- [150] J. Rodenburg, B. McCallum, P. Nellist, *Ultramicroscopy* 48 (1993) 304–314.
- [151] M.J. Humphry et al., *Nat. Commun.* 3 (2012) 730.
- [152] A.M. Maiden, J.M. Rodenburg, *Ultramicroscopy* 109 (2009) 1256–1262.
- [153] J. Rodenburg, R. Bates, *Philos. Trans. R. Soc. A* 339 (1992) 521–553.
- [154] P.M. Pelz et al., *Sci. Rep.* 7 (2017) 9883.
- [155] A.J. D’Alfonso et al., *J. App. Phys.* 119 (2016). 054302.
- [156] B. Song et al., *Phys. Rev. Lett.* 121 (2018). 146101.
- [157] T.J. Pennycook et al., *Ultramicroscopy* 151 (2015) 160–167.
- [158] H. Yang et al., *J. Phys.: Conf. Ser.* 644 (2015). 012032.
- [159] R. dos Reis et al., *App. Phys. Lett.* 112 (2018). 071901.
- [160] C.M. O’Leary et al., *App. Phys. Lett.* 116 (2020). 124101.
- [161] A. Stevens et al., *Phys. Lett.* 113 (2018). 033104.
- [162] L. Zhou et al., *Nat Commun* 11 (2020) 2773.
- [163] S. Gao et al., *Nat. Commun.* 8 (2017) 163.
- [164] K. Ooe et al., *Ultramicroscopy* 113133 (2020).
- [165] K. Kanaya et al., *J. App. Phys.* 29 (1958) 1046–1049.
- [166] R. Danev, K. Nagayama, *Ultramicroscopy* 88 (2001) 243–252.
- [167] A. Löfgren et al., *Ultramicroscopy* 164 (2016) 62–69.
- [168] R. Danev, W. Baumeister, *Curr. Opin. Struct. Biol.* 46 (2017) 87–94.
- [169] H.-W. Wang, X. Fan, *Curr. Opin. Struct. Biol.* 58 (2019) 175–182.
- [170] J. Verbeeck, H. Tian, A. Béché, *Ultramicroscopy* 113 (2012) 83–87.
- [171] R.M. Glaeser, *Rev. Sci. Instrum.* 84 (2013). 111101.
- [172] J. Verbeeck, H. Tian, P. Schattschneider, *Nature* 467 (2010) 301–304.
- [173] K. Saitoh et al., *J. Electron Microsc.* 61 (2012) 171–177.
- [174] T.R. Harvey et al., *Phys. Rev. App.* 10 (2018). 061001.
- [175] H. Yang et al., *Ultramicroscopy* 171 (2016) 117–125.
- [176] M. Tomita et al., *Ultramicroscopy* 218 (2020). 113089.
- [177] W. Yu et al., *Small* 13 (2017) 1700268.
- [178] O. Schwartz et al., *Nature Methods* 16 (2019) 1016–1020.
- [179] A. Béché, R. Juchtmans, J. Verbeeck, *Ultramicroscopy* 178 (2017) 12–19.
- [180] E.J. Candès, *Proc. Int. Congr. Math.* (2006).
- [181] A. Béché, B. Goris, B. Freitag, J. Verbeeck, *App. Phys. Lett.* 108 (2016). 093103.
- [182] K. Hujsak et al., *Microsc. Microanal.* 22 (2016) 778–788.
- [183] A. Stevens et al., *App. Phys. Lett.* 112 (2018). 043104.
- [184] J.M. Ede, R. Beanland, *Sci. Rep.* 10 (2020) 8332.
- [185] P. Trampert et al., *Ultramicroscopy* 191 (2018) 11–17.
- [186] M. Zhou et al., *NIPS* 9 (2009) 2295–2303.
- [187] A. Stevens et al., *Microscopy* 63 (2013) 41–51.
- [188] L. Kovarik et al., *App. Phys. Lett.* 109 (2016). 164102.
- [189] A. Zobelli et al., *Ultramicroscopy* 212 (2020). 112912.
- [190] P. Potocek et al., *Microsc. Microanal.* 26 (2020) 403–412.
- [191] G.M.D.P. Godaliyadda et al., *Imaging* 4 (2018) 1–16.
- [192] E. Monier et al., *Ultramicroscopy* 215 (2020). 112993.
- [193] S.R. Spurgeon et al., *Nat. Mater.* (2020) 1–6.
- [194] A. Timischl, *Scanning: J. Scanning Microsc.* 36 (2014) 317–326.
- [195] T. Dahmen et al., *Sci. Rep.* 6 (2016) 25350.
- [196] D. Ciprari, K. Jacob, R. Tannenbaum, *Macromolecules* 39 (2006) 6565–6573.
- [197] P.A. Midgley, M. Weyland, *Ultramicroscopy* 96 (2003) 413–431.
- [198] M. Weyland, P.A. Midgley, *Mater. Today* 7 (2004) 32–40.
- [199] C. Kubel et al., *Microsc. Microanal.* 11 (2005) 378–400.
- [200] A.J. Koster et al., *J. Phys. Chem. B* 104 (2000) 9368–9370.
- [201] S. Bals, G. Van Tendeloo, C. Kisielowski, *Adv. Mater.* 18 (2006) 892.
- [202] R. Xu et al., *Nat. Mater.* 14 (2015) 1099–1103.
- [203] B. Goris et al., *Nat. Mater.* 11 (2012) 930–935.
- [204] M.C. Scott et al., *Nature* 483 (2012) 444–U91.

- [205] J. Miao, P. Ercius, S.J. Billinge, *Science* 353 (2016) aaf2157.
- [206] Y. Yang et al., *Nature* 592 (2021) 60–64.
- [207] X. Tian et al., *Nat. Mater.* 19 (2020) 867–873.
- [208] D. Ren et al., *Ultramicroscopy* 208 (2020). 112860.
- [209] J. Zhou et al., *MRS Bull.* 45 (2020) 290–297.
- [210] H. Friedrich, M.R. McCartney, P.R. Buseck, *Ultramicroscopy* 106 (2005) 18–27.
- [211] B. Goris et al., *Ultramicroscopy* 113 (2012) 120–130.
- [212] A. Stevens, et al., *Microscopy* (Oxford, U.K.) 63 (2014) 41–51.
- [213] G. Wang et al., *Environ. Modell. Software* 30 (2012) 139–142.
- [214] J. Zhang, D. Zhao, W. Gao, *IEEE Trans. Image Process* 23 (2014) 3336–3351.
- [215] R. Leary et al., *Ultramicroscopy* 131 (2013) 70–91.
- [216] Z. Saghi et al., *Nano Lett.* 11 (2011) 4666–4673.
- [217] J.M. Thomas et al., *J. Colloid Interface Sci.* 392 (2013) 7–14.
- [218] C.V. Iancu et al., *Nat. Protoc.* 1 (2006) 2813–2819.
- [219] V. Lucic, A. Rigort, W. Baumeister, *J. Cell. Biol.* 202 (2013) 407–419.
- [220] Y. Li, K.A. Hujtsak, V. Backman, V.P. Dravid, *Microsc. Microanal.* 24 (2018) 502–503.
- [221] H.G. Brown, et al., arXiv preprint arXiv:2011.07652.
- [222] S. Gao et al., *Nat. Commun.* 8 (2017) 163.
- [223] S. Gao et al., *Microsc. Microanal.* 25 (2019) 1802–1803.
- [224] Y. Cheng et al., *Cell* 161 (2015) 438–449.
- [225] Y. Cheng, *Cell* 161 (2015) 450–457.
- [226] J. Frank et al., *Ultramicroscopy* 3 (1978) 283–290.
- [227] FRANK, J. Single-Particle Cryo-Electron Microscopy, 69–72.
- [228] D. Lyumkis, *J. Bio. Chem.* 294 (2019) 5181–5197.
- [229] R.M. Glaeser, *Annu. Rev. Biophys.* 48 (2019) 45–61.
- [230] Y.-H. Pan et al., *J. Struct. Biol.* 166 (2009) 22–31.
- [231] Y.-C. Wang et al., *Nano Lett.* 19 (2019) 732–738.
- [232] Y. Li et al., *bioRxiv* (2019) 636209.
- [233] H.J. Jung et al., *Adv. Mater.* 30 (2018). e1802769.
- [234] R. Sinclair, T. Itoh, R. Chin, *Microsc. Microanal.* 8 (2002) 288–304.
- [235] D. Ge et al., *Acta Mater.* 52 (2004) 3921–3927.
- [236] A.A. Murthy et al., *ACS Nano* 14 (2020) 1569–1576.
- [237] S. Kim et al., *ACS Nano* 12 (2018) 7335–7342.
- [238] N. de Jonge, L. Houben, R.E. Dunin-Borkowski, F.M. Ross, *Nat. Rev. Mater.* 4 (2019) 61–78.
- [239] K. He, T. Shokuhfar, R. Shahbazian-Yassar, *J. Phys. Condens. Matter* 31 (2019). 103001.
- [240] W.J. Liu et al., *Journal of Materials Chemistry A* 5 (2017) 145–154.
- [241] A. Nie et al., *Nano Lett.* 15 (2015) 610–615.
- [242] D. Cooper et al., *Adv. Mater.* 29 (2017) 1700212.
- [243] A. Azizi et al., *Nat. Commun.* 5 (2014) 4867.
- [244] J. Park et al., *Science* 349 (2015) 290–295.
- [245] X. Li et al., *Nat. Methods* 10 (2013) 584–590.
- [246] L. Jones et al., *Microscopy* 67 (2018) i98–i113.
- [247] A.A. Murthy, et al., arXiv preprint arXiv:2012.13842.
- [248] B. Meredig, *Chem. Mater.* 31 (2019) 9579–9581.
- [249] R. Sadre, et al., arXiv preprint arXiv:2012.05322 2020.
- [250] C.-H. Lee et al., *Nano Lett.* 20 (2020) 3369–3377.
- [251] C. Zhang et al., *Ultramicroscopy* 210 (2020). 112921.
- [252] P. Lee, J.D. West, B. Howe, *IEEE Trans. Big Data* 4 (2018) 117–129.
- [253] I. Arganda-Carreras et al., *Bioinformatics* 33 (2017) 2424–2426.
- [254] M. Ziatdinov et al., *ACS Nano* 11 (2017) 12742–12752. PMID:29215876.
- [255] I. Bruno et al., *Crystallography and Databases. Data Sci. J.* (2017) 16.
- [256] M. Nord et al., *Microsc. Microanal.* 26 (2020) 653–666.
- [257] P.M. Pelz, et al., arXiv preprint arXiv:2104.06336.
- [258] M.D. Wilkinson et al., *Sci. Data* 3 (2016). 160018.
- [259] J.A. Taillon et al., *Microsc. Microanal.* (2021) 1–17.
- [260] A. Andreev, D.E. Koo, *Microsc. Today* 28 (2020) 42–45.
- [261] B.H. Savitzky et al., *Microsc. Microanal.* 27 (2021) 1–32.
- [262] D.N. Johnstone, et al., pyxem/pyxem: pyxem 0.10.1. 2020.<https://doi.org/10.5281/zenodo.3667613>.
- [263] A. Clausen, et al., LiberTEM/LiberTEM: 0.5.1. 2020.<https://doi.org/10.5281/zenodo.3982290>.
- [264] S. Somnath, et al., pycroscopy/pycroscopy: 0.60.7-beta. 2020.<https://doi.org/10.5281/zenodo.4008763>.
- [265] M. Nord, pixstem, 2020. URL: <https://pixstem.org/>.
- [266] F. de la Peña, et al., hyperspy/hyperspy: Release v1.6.1. 2020. URL: <https://doi.org/10.5281/zenodo.4294676>.
- [267] S. Ribet, PNAS 118 (2021), <https://doi.org/10.1073/pnas.2102583118>. In press.
- [268] M. Malac, Phase plates in the transmission electron microscope: operating principles and applications, *Microscopy* 70 (1) (2021), <https://doi.org/10.1093/jmicro/dfaa070>. In press.
- [269] S. Hettler, A. Arenal, *Ultramicroscopy* (2021).