

Research



Cite this article: Silbiger NJ, Donahue MJ, Lubarsky K. 2020 Submarine groundwater discharge alters coral reef ecosystem metabolism. *Proc. R. Soc. B* **287**: 20202743. <https://doi.org/10.1098/rspb.2020.2743>

Received: 3 November 2020

Accepted: 23 November 2020

Subject Category:

Global change and conservation

Subject Areas:

ecology, environmental science

Keywords:

submarine groundwater discharge, coral reefs, ecosystem functioning, ecosystem metabolism, biological feedbacks, biogeochemistry

Author for correspondence:

Nyssa J. Silbiger

e-mail: nyssa.silbiger@csun.edu

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.5225973>.

Submarine groundwater discharge alters coral reef ecosystem metabolism

Nyssa J. Silbiger¹, Megan J. Donahue² and Katie Lubarsky³

¹Biology Department, California State University, Northridge, CA 91330, USA

²Hawai'i Institute of Marine Biology, University of Hawai'i at Mānoa, Kāne'ohe, HI 96744, USA

³Scripps Institution of Oceanography, University of California, San Diego, CA, 92037, USA

NJS, 0000-0003-4916-3217; MJD, 0000-0002-7529-001X; KL, 0000-0001-9642-6118

Submarine groundwater discharge (SGD) influences near-shore coral reef ecosystems worldwide. SGD biogeochemistry is distinct, typically with higher nutrients, lower pH, cooler temperature and lower salinity than receiving waters. SGD can also be a conduit for anthropogenic nutrients and other pollutants. Using Bayesian structural equation modelling, we investigate pathways and feedbacks by which SGD influences coral reef ecosystem metabolism at two Hawai'i sites with distinct aquifer chemistry. The thermal and biogeochemical environment created by SGD changed net ecosystem production (NEP) and net ecosystem calcification (NEC). NEP showed a nonlinear relationship with SGD-enhanced nutrients: high fluxes of moderately enriched SGD (Wailupe low tide) and low fluxes of highly enriched SGD (Kūpikipiki'ō high tide) increased NEP, but high fluxes of highly enriched SGD (Kūpikipiki'ō low tide) decreased NEP, indicating a shift toward microbial respiration. pH fluctuated with NEP, driving changes in the net growth of calcifiers (NEC). SGD enhances biological feedbacks: changes in SGD from land use and climate change will have consequences for calcification of coral reef communities, and thereby shoreline protection.

1. Background

Submarine groundwater discharge (SGD) is a natural and understudied feature of many near shore ecosystems that may play a key role in ecosystem functioning. SGD is the flow of water—meteoric, marine or a composite of both—from the land through the marginal seabed and into the coastal ocean [1,2]. SGD is a widespread phenomenon on near shore coral reefs as evidenced by reports from Hawai'i [3–6], Mo'orea [7,8], Florida [9], Israel [10], the Great Barrier Reef [11], Jamaica [12], Mexico [13] and Japan [14]. While there is an extensive body of literature describing high fluxes of SGD to coral reefs and other coastal ecosystems [15,16], there is a critical need to better understand the biological and ecological consequences of SGD [17].

SGD can be a source of high nutrient, low pH and cool water to coastlines [6,11,18,19], which can benefit oligotrophic systems [20] or put coral reefs at risk of eutrophication [15], depending on the watershed conditions and local environmental context. While SGD is natural, it can also act as a conduit for anthropogenic pollutants and lead to reef degradation [21]. Additionally, SGD can be fresh or saline and often has different concentrations of total alkalinity (TA), silicate and dissolved organic compounds than seawater [6,19,22]. Because each of these biogeochemical parameters can affect biological and ecological processes on coral reefs, it is important to understand how the unique environments created by SGD affect coral reef ecosystems.

The altered biogeochemistry in coastal waters from SGD can affect organismal physiology and species interactions that will ultimately lead to changes in key ecosystem functions, such as net ecosystem calcification (NEC) and net ecosystem production (NEP). For example, chronic nutrient loading of coastal waters can destabilize reefs by shifting competitive dominance away from corals and other

calcifiers toward fleshy algae, and by increasing bioerosion rates through enhanced growth of bioeroding invertebrates and endoliths [18,21,23–27]. Conversely, slight increases in nutrients can be beneficial to reefs: a recent *in situ* experiment showed that *Porites lobata* grew fastest at intermediate levels of nutrient-rich SGD [18]. Nutrient loading from SGD can also lead to shifts in macroalgal composition and biomass [28] and microbial community composition to enrich copiotrophic and pathogenic taxa [29–31]. At an ecosystem scale, added nutrients can directly increase NEP rates on fringing reefs [32] or decrease NEP and/or NEC when coupled with other geochemical parameters associated with SGD [33,34]. Further, nutrient loading at concentrations associated with SGD can alter the quality and quantity of organic matter released by benthic communities [29].

SGD-associated changes in carbonate chemistry (e.g. CO₂, pH, TA, dissolved inorganic carbon [DIC]) and temperature will also affect reef dynamics and ecosystem functioning. Lower pH (or high CO₂) typically leads to decreased growth rates of corals and coralline algae [35,36], higher bioerosion rates [37–39] and lower NEC rates [40]. TA or DIC from SGD may be higher or lower than the receiving seawater, depending on the conditions of the aquifer, and altered TA will affect carbonate buffering. Where TA of SGD is elevated relative to the receiving waters, SGD may buffer reefs from ocean acidification, as calcifiers are more resistant to declining pH at higher concentrations of TA [22,41]. In locations where TA of SGD is lower than the receiving waters, SGD may make reefs more susceptible to ocean acidification. Nutrient and carbonate parameters can also interact: elevated nutrients can strengthen the negative relationship between pH and bioerosion [42] and disrupt the relationship between calcification and aragonite saturation state [43]. Increased temperature variability from the SGD could also affect community production and calcification rates [44]. Further, SGD could create biological feedbacks, where changes in production rates from the elevated nutrient concentrations will change local pH conditions, which can ultimately lead to changes in NEC [43,45]. Notably, seasonality, oceanographic conditions [46] and anthropogenic stressors (e.g. nearby cesspools or injection wells) [21] can also affect the relationships between SGD and ecosystem functions. As SGD is a common feature of near shore reef habitats, understanding the mechanisms by which SGD affects ecosystem functioning is a critical need for coral reef management.

Here, we use Bayesian structural equation models to better understand how SGD directly affects the biogeochemical and thermal environment in a shallow reef flat, and how this unique environment directly and indirectly affects key ecosystem functions. We focus on two sites in Hawai'i with differing SGD fluxes, oceanographic conditions and anthropogenic impacts. We hypothesize that SGD will drive a biological feedback loop, whereby SGD indirectly affect NEP and NEC through altered nutrient, temperature and pH values; however, the strength of the relationships will differ between the sites due to differences in local environmental conditions (e.g. aquifer biogeochemistry, residence times, exposure). Fluxes and the biogeochemistry of SGD are highly linked to changes in precipitation, land use, sea-level rise, tidal amplitude and groundwater usage [47]. As the drivers of SGD flux are rapidly changing, we must improve our understanding of the biogeochemical and ecological consequences of SGD to better predict how both natural and human-induced changes in SGD will affect reef ecosystem functioning in the future.

(a) Study area

Maunalua Bay is an 8 km embayment on the southeast side of O'ahu with known inputs of SGD [6,48,49]. There are nine watersheds that feed into Maunalua Bay, each with unique hydrology and history of human impacts [50]. Our study is based at sites adjacent to two of these watersheds: Kūpikipiki'ō (also known as Black Point) and Wailupe (electronic supplementary material, figure S1). Both sites are shallow reef flats (<1 m depth at mean lower low water): the reef flat at Wailupe is approximately 500 m wide, with fine sediment close to shore that gives way to carbonate platforms in a spur and groove pattern, and the reef flat at Kūpikipiki'ō is approximately 200 m wide and has a mix of isolated basalt boulders and carbonate shelves [18]. Both sites are dominated by macroalgae (approx. 50% at Wailupe and approximately 15% at Kūpikipiki'ō) with very low coral cover (<1% at both sites) [28,51]. The dominant algal taxa at Kūpikipiki'ō were *Pterocladia* sp., turf and *Bryopsis pennata*; at Wailupe, the community was dominated by *Acanthophora spicifera*, *Gracilaria salicornia*, *Halimeda discoidea* and *Lyngbya* sp. The dominant corals at both sites were *Porites lobata* and *Montipora* spp., although they were in very low abundance. Kūpikipiki'ō has a freshened SGD flux of $11\,700 \pm 5900 \text{ m}^{-3} \text{ d}^{-1}$ (mean $\pm$ SD) over a 30-day period, with approximately 4.4% of the total recharge due to wastewater effluent from nearby cesspools; Wailupe has a freshened SGD flux of $9100 \pm 2700 \text{ m}^{-3} \text{ d}^{-1}$, with 0.7% of the total recharge is due to wastewater [33,48]. Further, the SGD from Kūpikipiki'ō has substantially higher nutrient concentrations in the groundwater than Wailupe (approx. 170 versus 70 $\mu\text{mol l}^{-1}$ [6,48]), with wastewater accounting for 54 to 95% of the total N and P loads, and higher N:P ratios in the source water [48] (electronic supplementary material, figure S2). The contribution of wastewater to nutrient concentrations in Wailupe is negligible. The oceanographic conditions are also quite different between the two sites: the reef flat at Kūpikipiki'ō is narrowed and the site is more exposed and has shorter residence times than Wailupe [33], but tidal forcing is the dominant driver of diel SGD fluxes and diel current speed and direction at both sites [52]. The differences in hydrology, biogeochemistry and oceanography between Kūpikipiki'ō and Wailupe could lead to differing effects of SGD on ecosystem functioning.

2. Materials and methods

(a) Sampling design

Detailed sampling design, collection and processing of water samples can be found in [18], which we briefly summarize here. Twenty water sampling locations were established at each site ($n = 40$ total) in an approximate grid, scaled to the width of the reef flat, for biogeochemistry measurements (electronic supplementary material, figure S1). At each location, we collected discrete diel water samples across four timepoints (during daytime and nighttime high and low tides) in the spring and fall (totalling eight samples per location, 320 samples overall). Sampling dates were 18 April 2015 and 28 September 2015 at Wailupe, and 2 May 2015 and 26 October 2015 at Kūpikipiki'ō. Circulation on the shallow reef flat is primarily wind-driven [52], and northeast trade winds prevailed during all four sampling events, and wave heights ranged from 1.1 to 1.45 m. All sampling events happened during spring tide events. Water samples for pH, TA, nitrate + nitrite (NO₃⁻ + NO₂⁻), phosphate (PO₄³⁻) and silicate (SiO₃⁴⁻) were hand collected directly above the benthos within 30 min of

high or low tide in acid-washed HDPE bottles. pH was measured immediately using a tris calibrated Orion ROSS Ultra pH/ATC Triode following Dickson SOP 6a [53]. TA samples were immediately preserved with 50% saturated HgCl_2 in deionized water, stored in a cool dark place and later analysed on a Mettler Toledo T-50 autotitrator following Dickson SOP 3b [53] (precision = $2.68 \mu\text{Eq}$, accuracy = $0.48\% \pm 0.33\%$ SD). Inorganic nutrient samples (SiO_3^{4-} , $\text{NO}_3^- + \text{NO}_2^-$, PO_4^{3-}) were immediately filtered through pre-combusted GF/F filters ($0.7 \mu\text{m}$) and stored in a -20°C freezer until further processing. Nutrient samples were processed at the SOEST Laboratory for Analytical Biogeochemistry at the University of Hawai'i at Mānoa using a Seal Analytical AA3 nutrient autoanalyser (reported error [coefficient of variance]: 0.5% for SiO_3^{4-} , 0.3% for $\text{NO}_3^- + \text{NO}_2^-$, and 0.2% for PO_4^{3-}). Temperature was recorded every 15 min on HOBO TidbiT v2 loggers and temperature values were extracted to match the time-discrete water samples were collected.

(b) Characterizing relative submarine groundwater discharge

Silicate is a common tracer for SGD [54] and was used to calculate the relative amount of SGD in the water column at each location during the time of collection. Notably, silicate is a non-conservative tracer and can be influenced by uptake from siliceous phytoplankton or benthic organisms, like sponges [55]. However, a strong significant relationship between silicate and radon (Rn , a common conservative tracer for SGD [56], $r^2 = 0.98$, $p < 0.0001$) from prior data at the same sites [6] indicates that biological uptake of SiO_3^{4-} is minimal at Maunalua (electronic supplementary material, figure S3). Therefore, SiO_3^{4-} can be used as a tracer for SGD at our research sites. The standardized measure of SGD, presented as %SGD, was calculated as

$$\% \text{SGD} = \frac{\text{Si}_{\text{mix}} - \text{Si}_{\text{SW}}}{\text{Si}_{\text{GW}} - \text{Si}_{\text{SW}}},$$

where Si_{mix} is the concentration at each location, Si_{SW} is the ambient concentration in seawater ($1.03 \mu\text{mol l}^{-1}$, the lowest concentration during the sampling points) and Si_{GW} is the concentration of the mean groundwater endpoint ($810 \mu\text{mol l}^{-1} \pm 52$ (SD) for Wailupe ($n = 6$) and 740 ± 13 (SD) $\mu\text{mol l}^{-1}$ for Kūpikipiki'ō ($n = 9$); values from Richardson *et al.* [33]). The associated salinities were 4.9 ± 0.2 (SD), 2.0 ± 0.2 (SD) and 35.2 for Kūpikipiki'ō, Wailupe and the open ocean endmembers, respectively.

(c) Calculating net ecosystem production and net ecosystem calcification

To account for changes in geochemical parameters due to mixing between groundwater and seawater, we normalized all geochemical data to a common reference concentration using the following equation:

$$C_1 = C_{\text{mix}} + (C_{\text{mix}} - C_{\text{SGD}}) \left(\frac{\text{Si}_{\text{mix}} - 1.17}{\text{Si}_{\text{SGD}} - \text{Si}_{\text{mix}}} \right),$$

where C_1 is the silicate-normalized concentration at the reference value ($1.17 \mu\text{mol l}^{-1}$), C_{mix} is the measured concentration (groundwater-marine mixture), C_{SGD} is the average groundwater endmember concentration, Si_{mix} is the measured concentration (groundwater-marine mixture) and Si_{SGD} is the average concentration of the groundwater endmember [33]. The reference value was taken from the Hawaii Ocean Time-series at Station ALOHA in 2015 [57]. Groundwater endmembers for TA and DIC for Kūpikipiki'ō were 2946 ± 8 (SD) $\mu\text{mol kg}^{-1}$ and 3038 ± 8 (SD) $\mu\text{mol kg}^{-1}$, respectively, and at Wailupe were 1754 ± 35 (SD) $\mu\text{mol kg}^{-1}$ and 1779 ± 40 (SD) $\mu\text{mol kg}^{-1}$ [33].

We estimated the NEC and NEP potential [58] for each location $\times$ timepoint by comparing the change in TA or DIC from each water sample with open ocean values from Station ALOHA [57]. NEC potential was calculated as $\Delta\text{TA}/2$, while NEP potential was calculated as ΔDIC . ΔTA was divided by two because 1 mol of CaCO_3 is produced per 2 mol of TA uptake [59]. Differences in TA or DIC are commonly used in the literature as a proxy for whether reefs are net calcifying/producing (TA or DIC depletion) or net dissolving/respiring (TA or DIC repletion) [58,60]. We use the terms NEC and NEP for brevity throughout the remainder of the paper. Positive and negative $\Delta\text{TA}/2$ represent net calcification and net dissolution, respectively. Likewise, positive and negative ΔDIC represent net photosynthesis and net respiration, respectively. DIC was calculated from pH_T and TA using seacarb [61]. The mean $\pm$ SE error propagation of DIC calculated from TA (error of $5 \mu\text{mol kg}^{-1}$) and pH_T (error of 0.01) is $9.6 \pm 0.05 \mu\text{mol kg}^{-1}$. We did not calculate true rates (normalized to residence time) because Maunalua Bay is a shallow embayment with a complex hydrodynamic environment [52], and small uncertainties in residence time measurements can lead to highly inaccurate NEP and NEC calculations [62].

(d) Model description

We used Bayesian structural equation models (SEM) to uncover the pathways and feedback loops by which SGD affects ecosystem functioning. SEMs are useful tools for evaluating multivariate hypotheses under a flexible statistical modelling framework [63]. We specify the SEM using five equations that describe the influence of SGD on NEP and NEC as mediated by inorganic nutrients, temperature and pH, which can vary with season, day/night, or tide. The system of equations, with associated hypotheses, are as follows:

Model 1: $\log(N + N) \sim \log(\text{SGD})$.

Model 1 represents that nitrogen concentration increases directly with %SGD and that nitrogen concentration is primarily a function of mixing [48].

Model 2: temperature $\sim \log(\text{SGD}) \times \text{season} \times \text{day or night}$.

Model 2 represents our hypothesis that temperature decreases as a function of %SGD and that the relationship can vary with time of day and season (i.e. the difference between SGD, temperature and surface water temperature varies by season and by time of day and their interaction).

Model 3: $\text{pH} \sim \log(\text{SGD}) + \text{NEP}$.

Model 3 represents our hypotheses that pH is a function of mixing between surface water and SGD (which has a distinct pH signature from surface water) and, critically, that there is a feedback between NEP and pH.

Model 4: $\text{NEP} \sim \log(N + N) \times \text{tide} \times \text{day or night} + \text{season} \times \text{temperature}$.

Model 4 represents our understanding that NEP is fuelled by nutrients (i.e. nitrate + nitrite) and that this response to higher nitrogen concentration is stronger during the day (when light is available for photosynthesis). Notably, phosphate can also drive NEP rates. However, $\text{NO}_3^- + \text{NO}_2^-$ and PO_4^{3-} were highly collinear (Pearson's $R = 0.95$ at Wailupe and 0.96 at site Kūpikipiki'ō; electronic supplementary material, figure S2); therefore, we only included $\text{NO}_3^- + \text{NO}_2^-$ in the model to represent inorganic nutrients. We also hypothesized that the relationship may interact with tidal mixing (higher concentrations of nutrients coupled with increased organic matter are released during low tide). In addition, we represent the hypothesis that NEP increases as a function of

temperature, which is enhanced under longer daylength (season).

Model 5: $NEC \sim pH + \text{temperature}$.

Model 5 reflects our mechanistic understanding that NEC increases with pH and with temperature.

(e) Model fitting and analysis

The SEMs were run for Kūpikipiki'ō and Wailupe separately. All data were scaled and centred for the analysis; SGD and $\text{NO}_3^- + \text{NO}_2^-$ were log-transformed because they were highly left-skewed, and two extreme outliers were removed from the analysis. There were 15 samples missing from the dataset (5 from Wailupe and 10 from Kūpikipiki'ō) due to sea conditions for a total sample size of 149 and 155 at Kūpikipiki'ō and Wailupe, respectively. The Bayesian SEM models were run using the Hamiltonian Monte Carlo algorithm from STAN in the R package *brms* [64,65]. We ran three parallel chains of length 2000, with a warm-up of 1000, and a thinning parameter of 1. The posterior was modelled from a multivariate normal distribution with relatively uninformative priors for all parameters (student t distribution (d.f. = 3, $\mu = 0$, $\sigma = 10$)). We assessed model convergence by visually checking all trace plots, ensuring that the chains were well-mixed and calculating Gelman–Rubin statistics [66] for all parameters (which were all = 1). To assess model fit, we used posterior predictive checks where we visually inspected the relationship between the empirical distribution and 10 posterior draws using the *pp_check* function in the *tidybayes* package [67]. All results are reported as medians with two-tailed 95% Bayesian credible intervals (essentially Bayesian confidence intervals). Effect sizes with credible intervals that do not overlap zero are considered to be statistically significant.

To characterize the sensitivity of ecosystem metabolism to changes in SGD, we simulated data where we uniformly increased and decreased the relative amount of SGD at each sampling location $\times$ timepoint by 50%. We chose a $\pm 50\%$ change in SGD for the sensitivity analysis to capture the natural range in SGD fluxes experienced at both sites ($11\,700 \pm 5900 \text{ m}^{-3} \text{ d}^{-1}$ and $9100 \pm 2700 \text{ m}^{-3} \text{ d}^{-1}$ [48]). This simulated data was then used to generate predictions for how each state variable (i.e. NEC, NEP, pH, N + N and temperature) would change as a result of increased SGD from the posteriors. All data and R code are available at <https://github.com/njsilbiger/MaunaloaSEM> and at Zenodo (doi:10.5281/zenodo.4281383).

3. Results and discussion

Overall, our study showed that SGD significantly altered the local biogeochemical and thermal environment, which ultimately led to changes in NEC and NEP. We also showed evidence of SGD fuelling biological feedbacks, where changes in NEP as a function of nutrients and temperature led to changes in pH, which ultimately drove NEC. However, the strength (and sometimes direction) of the relationships between SGD, temperature, nitrogen concentration, pH, NEP and NEC differed between the two sites (figure 1; electronic supplementary material, figure S4, S5), highlighting that local oceanographic conditions and aquifer geochemistry can alter the effect of SGD on coral reefs.

(a) Direct and indirect effect of submarine groundwater discharge on nitrogen, temperature and pH

For nutrients, there was a strong positive relationship between %SGD and $\text{NO}_3^- + \text{NO}_2^-$ concentration at both sites, but the effect size was 1.5 times higher at Kūpikipiki'ō than Wailupe

(figure 1a; electronic supplementary material, figure S4A and S5A). The difference in slopes between these two sites is likely to be due to the substantially higher nutrient concentrations in the groundwater at Kūpikipiki'ō relative to Wailupe (169 versus $69 \mu\text{mol l}^{-1}$; [48]). Many studies show high nitrogen fluxes as a result of SGD [19]. High nutrient fluxes from SGD could be a critical source of exogenous nutrients necessary for sustaining high coastal production rates [20], or it could lead to eutrophication risk if associated with anthropogenic pollution [15]. Notably, approximately 14% of global coral reefs are at risk of eutrophication from SGD [15]. Given the major differences in nitrogen fluxes from SGD between the two sites in our study, and the fact that the nutrients at Kūpikipiki'ō are heavily impacted by wastewater [48], the nutrient-rich SGD could lead to different biological responses at each site.

The presence of SGD also decreased water temperature at both sites, which is consistent with studies showing that fresh SGD is typically cooler than ambient seawater [16]. The effect of SGD on temperature was stronger at Wailupe than at Kūpikipiki'ō (figure 1b; electronic supplementary material, figure S4B and S5B), where the 2°C difference between groundwater and ambient seawater was only evident during the day. At night, the differences in temperature as a function of groundwater were undetectable at Kūpikipiki'ō. The stronger effect of SGD on temperature during both the day and night at Wailupe is probably due to differences in mixing rates between the two sites. Wailupe has longer residence times (mixing rates are lower) than Kūpikipiki'ō [33], probably enhancing the difference in temperature between groundwater and ambient seawater. Further, there was no significant interaction between season and %SGD on the temperature at either site, but there was a significant main effect of season: fall was on average 2°C warmer than spring at both sites (electronic supplementary material, table S1, figure S4B).

There was a small, but a significant direct effect of %SGD on pH at both sites (figure 1c; electronic supplementary material, figure S4C and S5C). However, the indirect biological effect of pH from changes in NEP was 5.5 and 8.1 times higher than the direct effects on pH from freshwater intrusion at Kūpikipiki'ō and Wailupe, respectively (figure 1c; electronic supplementary material, figure S4 and S5D). SGD typically has lower pH values than seawater [11,22], which was evident at Wailupe. However, the direct effect of lower pH from SGD mixing with surface water was completely masked by the biological feedbacks at Kūpikipiki'ō, where the net effect of SGD on pH was positive. The site differences in the relationship between SGD and pH are probably due to differences in NEP (the maximum and range in NEP was higher at Kūpikipiki'ō than Wailupe; electronic supplementary material, table S1). Prior studies have shown that NEP can have an overwhelming effect on pH relative to other physical drivers [45] and that nutrient addition could augment biological feedbacks in coastal ecosystems [43,68]. Additionally, vastly different TA values between the source groundwater in Kūpikipiki'ō and Wailupe could be affecting the relationship between %SGD and pH as well [6,33]. The TA at Kūpikipiki'ō is considerably higher than ambient seawater (approx. 3000 versus $2300 \mu\text{mol kg}^{-1}$), where at Wailupe it is lower (approx. $1750 \mu\text{mol kg}^{-1}$). TA affects the buffering capacity of seawater, or how much CO_2 seawater can absorb before decreasing pH, where higher TA seawater has a higher buffering capacity than lower TA [69]. The positive and negative relationships between %SGD

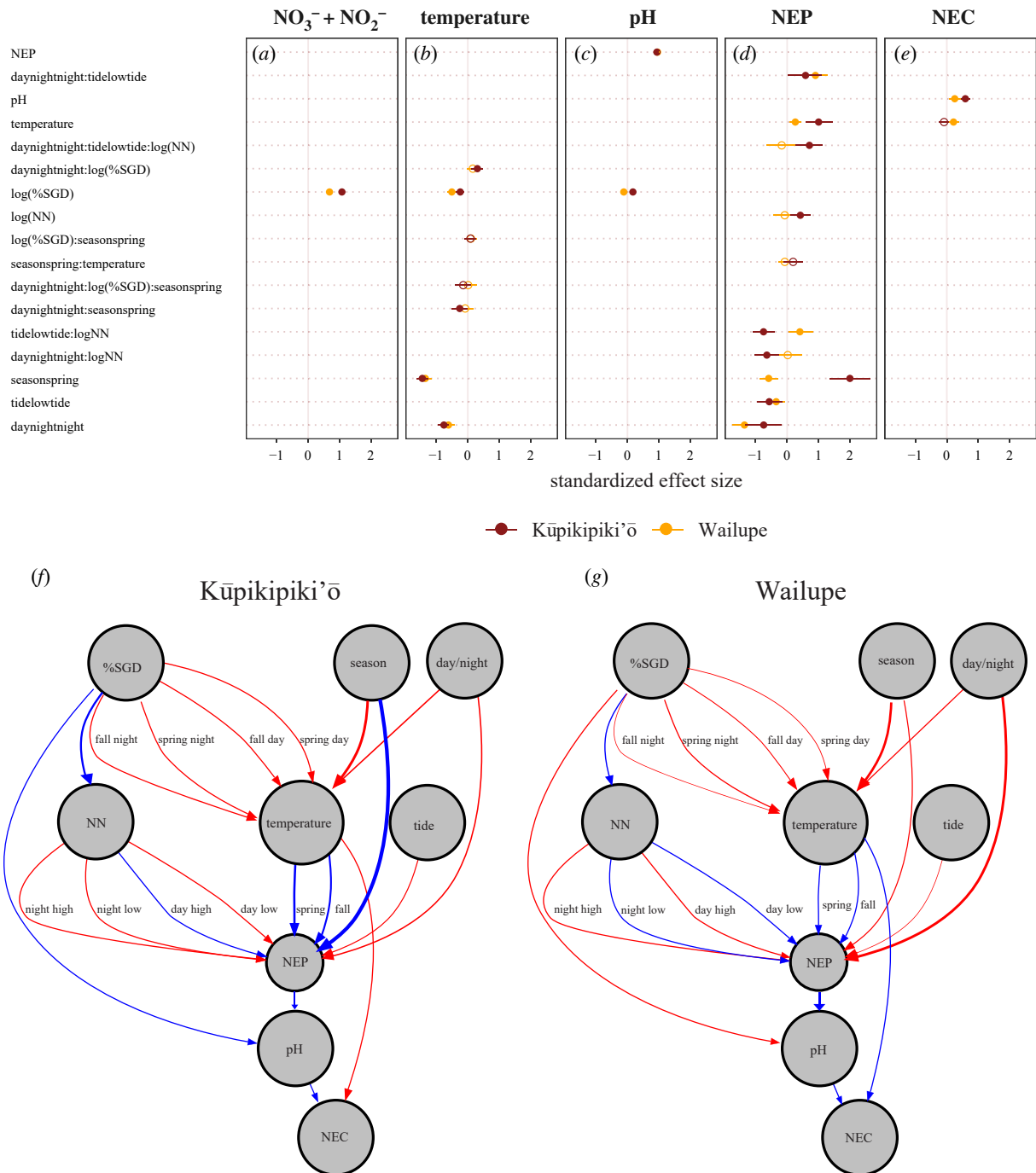


Figure 1. (a–e) Coefficient plot and (f,g) directed acyclic graphs (DAG) from structural equation models. (a–e) Standardized effect sizes for parameters in models 1–5 (left to right), where each point is the median value $\pm$ 95% Bayesian credible interval (BCI). Values with BCIs that do not overlap zero are statistically significant. Open symbols in the figure are not significant. Kūpikipiki'ō and Wailupe values are represented in maroon and gold, respectively. Subsets F and G are DAGs for Kūpikipiki'ō (left) and Wailupe (right). Colours of the arrows represent positive (blue) or negative (red) relationships and the thickness of the lines are correlated with the effect size.

and pH at Kūpikipiki'ō and Wailupe, respectively, could also be influenced by the higher and lower TA concentrations in the groundwater relative to seawater.

(b) Direct and indirect effect of submarine groundwater discharge on net ecosystem production and net ecosystem calcification

The unique thermal and biogeochemical environment created by SGD drove changes in NEP and NEC at both sites. For NEP, there was a strong positive relationship between NEP

and temperature, though the effect was 3.8 times stronger at Kūpikipiki'ō than Wailupe (figure 1d; electronic supplementary material, figure S4F and S5F). There was also a significant seasonal effect (though, notably with opposite patterns between the sites), probably due to seasonal differences in circulation leading to greater differences wave energy and surface currents between the two sites, with higher wave energy and stronger offshore flow at Kūpikipiki'ō than Wailupe [52].

The relationship between $\text{NO}_3^- + \text{NO}_2^-$ and NEP was mediated by day/night and tidal cycle as indicated by the significant three-way interactions, though the relationships

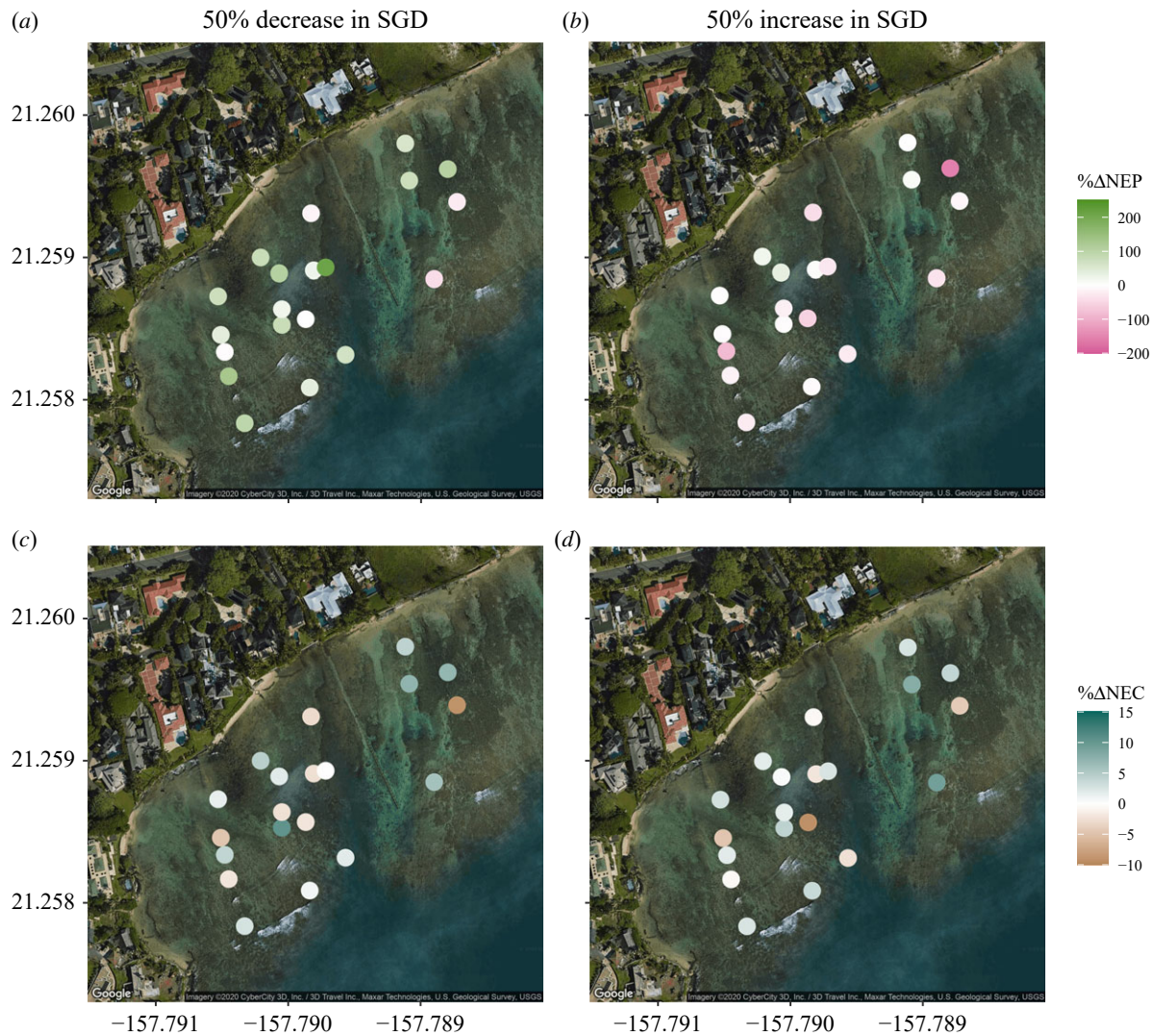


Figure 2. Maps of predicted change (%) in (a,b) NEP and (c,d) NEC at a (a,c) 50% decrease and (b,d) 50% increase in SGD in Kūpikipikī'ō. Dots are median predictions across all eight timepoints for each sampling location.

were different between the sites (figure 1d; electronic supplementary material, figure S4E and S5E). At Kūpikipikī'ō, there was a negative relationship between $\text{NO}_3^- + \text{NO}_2^-$ and NEP at night, regardless of the tide. This relationship is probably driven by increased microbial respiration at night. During the day, however, there was a positive effect of $\text{NO}_3^- + \text{NO}_2^-$ on NEP during high tide, but a negative effect during low tide. At high tide, the SGD was still able to increase the $\text{NO}_3^- + \text{NO}_2^-$ concentrations by $1.4 \mu\text{mol l}^{-1}$, which probably benefitted producers. During low tide, $\text{NO}_3^- + \text{NO}_2^-$ concentrations were 23 times higher than high tide (up to $32.4 \mu\text{mol l}^{-1}$), and, notably, with a significant amount of wastewater [48] and higher DOM concentrations [6], which may have benefitted microbial respiration more than gross photosynthesis. At Wailupe, tide had the opposite effect on the relationship between NEP and $\text{NO}_3^- + \text{NO}_2^-$. There was no relationship between $\text{NO}_3^- + \text{NO}_2^-$ and NEP during high tide, and there was a positive relationship during low tide (electronic supplementary material, figure S5E). Again, this may have been due to differences in nitrogen concentrations between high and low tide at Wailupe, where the nitrogen concentrations are significantly lower than Kūpikipikī'ō (max daytime $\text{NO}_3^- + \text{NO}_2^-$ at Wailupe was $0.68 \mu\text{mol l}^{-1}$ and $7.92 \mu\text{mol l}^{-1}$ at high and low tide, respectively).

The differences in the relationship between nutrients and NEP are consistent with prior studies at these sites. La Valle *et al.* [28] showed that total producer biomass increased with $\text{NO}_3^- + \text{NO}_2^-$ concentration at Wailupe, but not Kūpikipikī'ō. Several factors could be driving the differences in these relationships between our two sites. First, the N:P ratio in the source water is two times higher at Kūpikipikī'ō than Wailupe (electronic supplementary material, figure S2) and the balance between nitrogen and phosphate affects the ability of organisms to take-up nutrients [70]. The differences in macroalgal community composition between the two sites could affect the relationship between NEP and nutrients as the different dominant species have different growth rates [28]. Additionally, differences in the prevalence of human-derived nitrogen and covarying parameters such as organic matter—both of which are higher at Kūpikipikī'ō [6,48]—could be responsible for the different relationship between SGD and NEP at Kūpikipikī'ō and Wailupe. Further, Gill [71] points out the importance of nonlinear relationships between nutrient concentrations and biological responses (with a focus on coral growth) on coral reefs. He shows that, at low nutrient concentrations, reefs are likely to benefit from increases in nutrients, whereas at high concentrations the nutrient enrichment is more likely to be detrimental; Lubarsky *et al.* [18] found evidence of this pattern in coral growth at these sites in Maunaloa.

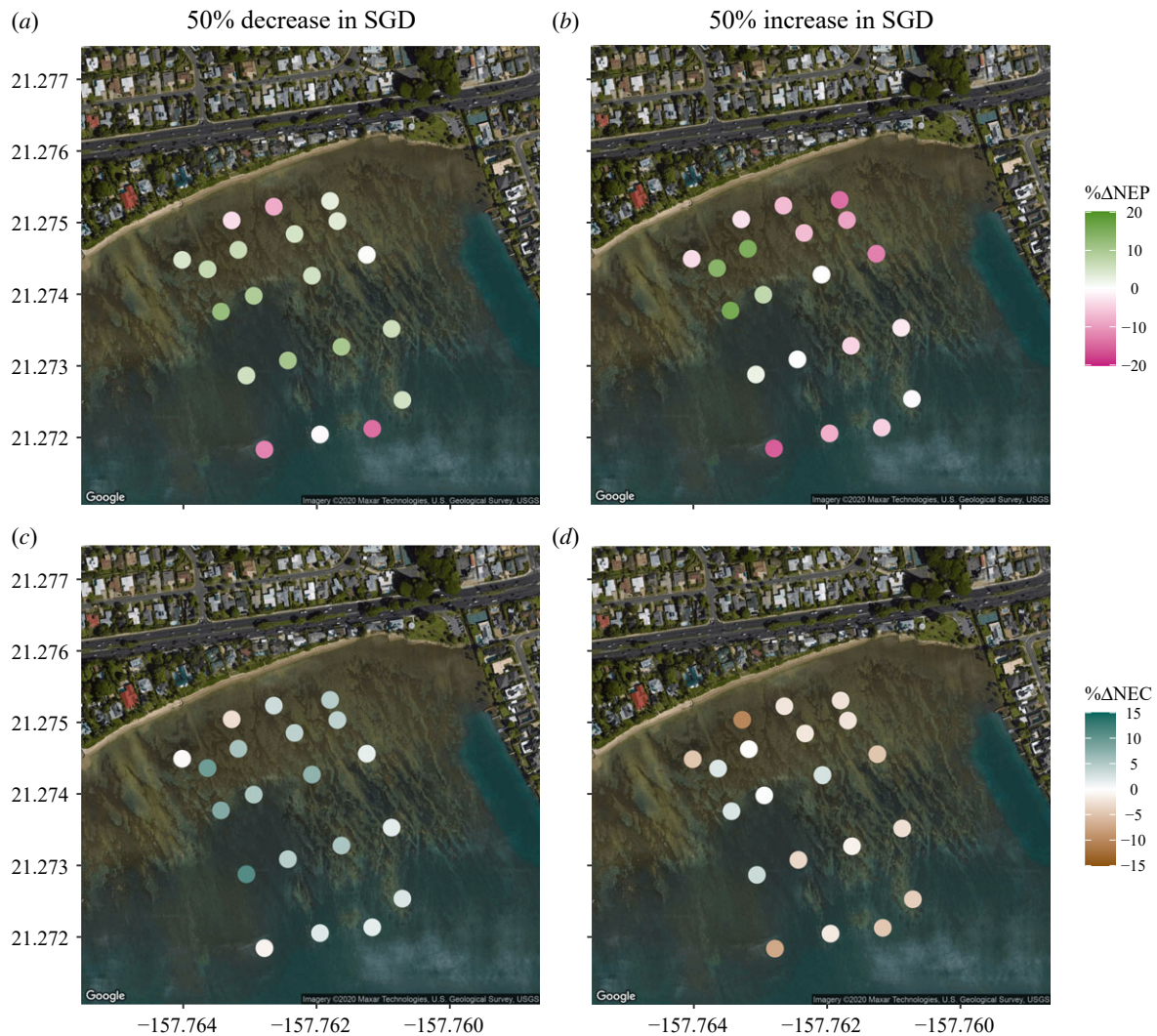


Figure 3. Maps of predicted change (%) in (a,b) NEP and (c,d) NEC at a (a,c) 50% decrease and (b,d) 50% increase in SGD in Wailupe. Dots are median predictions across all eight timepoints for each sampling location.

NEC was also affected by the biogeochemical and thermal environment created by SGD. There was a significant positive relationship between pH and NEC at both sites, although the effect size was 2.5 times higher at Kūpikipiki'ō than Wailupe (figure 1e; electronic supplementary material, figure S4G and S5G). There was also a significant effect of temperature on NEC at Wailupe, but not Kūpikipiki'ō (figure 1e; electronic supplementary material, figure S4H and S5H). These results are consistent with the expectation that natural increases pH and temperature (except during extreme heating events) increase NEC on coral reefs [39,72,73]. Notably, the stronger effect size at Kūpikipiki'ō is opposite of what we would expect based on Silbiger *et al.* [43], which showed that nutrient enrichment in a laboratory setting weakened the relationship between aragonite saturation state (which is positively correlated with pH) and NEC. However, other factors in the groundwater, such as the highly depleted total alkalinity at Wailupe, could be mediating this relationship. Regardless of the differences in effect sizes, this study provides evidence for the strong indirect effect of SGD on NEC. Specifically, SGD augments biological feedbacks: SGD increases nutrient concentrations, which fuels both production and microbial respiration, altering the local pH environment (increasing pH when NEP is positive and decreasing pH when NEP is negative) and ultimately leading to changes in NEC. NEC (i.e. the net growth of calcifiers, such as corals, on the reef) is important

for creating reef structure; therefore, any declines in NEC as a result of changing SGD could lead to loss of reef-associated biodiversity and shoreline protection [74].

There are a few important limitations to consider when interpreting the results of our study. First, our sampling design prioritized high spatial resolution over high temporal resolution. Water samples were collected over two diel cycles aimed at capturing the most extreme groundwater fluxes throughout the day (high tide and low tide) and year (dry and wet season). While more frequent sampling would probably reduce the uncertainty in our models, prior studies at Maunalua Bay consistently show similar relationships between SGD and many of the parameters analysed in the current study [6,28,33,48]. Second, we calculated NEC and NEP potential because of the complex hydrodynamic environment at Maunalua Bay. While changes in DIC and TA are commonly used in the literature as a proxy for net calcification/net production [58,60,72], future studies at Maunalua should include high-resolution oceanographic data to better understand NEP and NEC rates, and how they are affected by SGD.

(c) Sensitivity analysis of submarine groundwater discharge on ecosystem functioning

The fluxes and biogeochemistry of SGD change with precipitation, land use, sea-level rise, tidal amplitude and groundwater

usage [47], making it critical to better understand how SGD affects reef ecosystem functioning. We used our SEM models to test how a 50% change (both decrease and increase) in SGD would affect ecosystem metabolism (figures 2 and 3; electronic supplementary material, figures S6 and S7). We found that the effects of increasing SGD on NEP and NEC were highly spatially variable, probably due to differences in the distance from SGD seeps and local oceanographic conditions. Overall, the magnitude of change (both positive and negative) in both NEP and NEC was higher at Kūpikipiki'ō than Wailupe. Specifically, SGD had an order of magnitude higher effect on NEP at Kūpikipiki'ō than Wailupe, where a 50% increase in SGD led to a −15.4 to 34.5% change (percentage change are median values over the eight sampling timepoints for each location) in NEP at Kūpikipiki'ō and −15.4 to 15.6% change in NEP at Wailupe across all 20 sampling locations. When we decreased SGD by 50% in the model, there was a −42.4 to 208% and −14.0 to 12% change in NEP at Kūpikipiki'ō and Wailupe, respectfully, across all sampling points. The large differences in sensitivity for NEP between the two sites could be in response to the substantially higher nutrient concentrations in the SGD at Kūpikipiki'ō than Wailupe. NEC was much less sensitive to changes in SGD than NEP at both sites. A 50% increase in SGD led to a −9.16 to 9.12% and −9.96 to 3.39% change in NEC at Kūpikipiki'ō and Wailupe, respectfully, whereas a 50% decrease in SGD led to a −9.01 to 10.3% and −2.62 to 11.1% change in NEC across all sample locations. The lower sensitivity of NEC to SGD than NEP is probably due to the lower coral cover (approx. 1%) relative to macroalgal cover (approx. 15–50%) at both sites [28,51].

Importantly, the SEM models did not have a direct link between SGD and NEC or SGD and NEP; yet our model predicted that both NEP and NEC would change with an increase in %SGD. These predictions provide further evidence for (i) the cascading and indirect effect of SGD on ecosystem functioning, where the effect of SGD on ecosystem metabolism is mediated by changes in nutrients, temperature and pH, and (ii) the context dependency of SGD, as evidenced by the nonlinear relationships between SGD and ecosystem metabolism and the substantial differences in sensitivity of SGD between the two sites. As hydrologists and climate modellers advance our understanding for how SGD fluxes will change in the future, our modelling approach

can be used to predict the downstream effects of altered SGD fluxes on ecosystem functioning.

Our study provides evidence that SGD leads to cascading changes in biogeochemistry and ecosystem metabolism, and that the effects of SGD on ecosystem functioning are altered by human disturbance, hydrology and oceanographic context. SGD is ubiquitous along coastlines and is directly impacted by humans through climate change and land use; yet, we have only scratched the surface on our understanding of how SGD directly and indirectly affects biological processes. These types of studies are necessary to better manage watersheds and help prepare for how coral reefs or any coastal ecosystem may change in the future. More studies should examine the relationships between SGD and ecological processes under different environmental conditions to gain a broader understanding of the mechanisms by which SGD affects ecosystem functioning.

Data accessibility. All data and R code are available at <https://github.com/njsilbiger/MaunaloaSEM> and at Zenodo (<https://doi.org/10.5281/zenodo.4281383>)

Authors' contributions. N.J.S., M.J.D. and K.L. conceived the idea. N.J.S., M.J.D. and K.L. collected the data. N.J.S. and M.J.D. statistically analysed the data. N.J.S. led the writing of the manuscript. N.J.S., M.J.D. and K.L. edited and approved of the manuscript.

Competing interests. We declare we have no competing interests.

Funding. This research was funded in part by the National Science Foundation Award 1924281 to N.J.S., NSF 1923877 to M.J.D. and the University of Hawai'i SeaGrant Program (NA14OAR4170071-R/SB-10) to M.J.D. Some data were obtained via the Hawaii Ocean Time-series HOT-DOGS application at University of Hawai'i at Mānoa, which was funded by National Science Foundation Award 1756517. This paper is funded in part by a grant from the National Oceanic and Atmospheric Administration, Project R/SB-10, which is sponsored by the University of Hawaii Sea Grant College Program, SOEST, under Institutional grant no. NA14OAR4170071 from NOAA Office of Sea Grant, Department of Commerce. The views expressed herein are those of the authors and do not necessarily reflect the views of NOAA or any of its subagencies.

Acknowledgements. We thank our collaborator C. Nelson for advice and comments on this manuscript, and field/laboratory support from several volunteers including J. Kuwabara, A. Kirby, E. Nalley, J. Zill, C. Counsell, J. Caldwell, I. Caldwell, C. Couch, H. Shiu, E. Lenz, J. Davidson, H. Putnam, B. Day, N. Jones, K. Remple and F. La Valle. This is contribution number 315 from the CSUN Marine Biology Program, number 1835 from the Hawai'i Institute of Marine Biology, number 11191 from SOEST and UNIH-SEA-GRANT-JC-14-09.

References

- Lee J, Kim G. 2015 Dependence of coastal water pH increases on submarine groundwater discharge off a volcanic island. *Estuar. Coast. Shelf Sci.* **163**, 15–21. (doi:10.1016/j.ecss.2015.05.037)
- Moosdorf N, Stieglitz T, Waska H, Dürr HH, Hartmann J. 2014 Submarine groundwater discharge from tropical islands: a review. *Grundwasser* **20**, 53–67. (doi:10.1007/s00767-014-0275-3)
- Garrison GH, Glenn CR, McMurtry GM. 2003 Measurement of submarine groundwater discharge in Kahana Bay, O'ahu, Hawai'i. *Limnol. Oceanogr.* **48**, 920–928. (doi:10.4319/lo.2003.48.2.0920)
- Street JH, Knee KL, Grossman EE, Paytan A. 2008 Submarine groundwater discharge and nutrient addition to the coastal zone and coral reefs of leeward Hawai'i. *Mar. Chem.* **109**, 355–376. (doi:10.1016/j.marchem.2007.08.009)
- Swarzenski PW, Dulai H, Kroeger KD, Smith CG, Dimova N, Storlazzi CD, Prouty NG, Gingerich SB, Glenn CR. 2017 Observations of nearshore groundwater discharge: Kahekili Beach Park submarine springs, Maui, Hawaii. *J. Hydrol. Regional Stud.* **11**, 147–165. (doi:10.1016/j.ejrh.2015.12.056)
- Nelson CE *et al.* 2015 Fluorescent dissolved organic matter as a multivariate biogeochemical tracer of submarine groundwater discharge in coral reef ecosystems. *Mar. Chem.* **177**, 232–243. (doi:10.1016/j.marchem.2015.06.026)
- Knee KL, Crook ED, Hench JL, Leichter JJ, Paytan A. 2016 Assessment of submarine groundwater discharge (SGD) as a source of dissolved radium and nutrients to Moorea (French Polynesia) coastal waters. *Estuaries Coasts* **39**, 1651–1668. (doi:10.1007/s12237-016-0108-y)
- Hagedorn B, Becker MW, Silbiger NJ. 2020 Evidence of freshened groundwater below a tropical fringing reef. *Hydrogeol. J.* **28**, 2501–2517. (doi:10.1007/s10040-020-02191-1)
- Corbett DR, Chanton J, Burnett W, Dillon K, Rutkowski C, Fourqurean JW. 1999 Patterns of groundwater discharge into Florida Bay. *Limnol. Oceanogr.* **44**, 1045–1055. (doi:10.4319/lo.1999.44.4.1045)

10. Shellenbarger GG, Monismith SG, Genin A, Paytan A. 2006 The importance of submarine groundwater discharge to the near shore nutrient supply in the Gulf of Aqaba (Israel). *Limnol. Oceanogr.* **51**, 1876–1886. (doi:10.4319/lo.2006.51.4.1876)
11. Cyronak T, Santos IR, Erler DV, Maher DT, Eyre BD. 2014 Drivers of pCO₂ variability in two contrasting coral reef lagoons: the influence of submarine groundwater discharge. *Global Biogeochem. Cycles* **28**, 2013GB004598. (doi:10.1002/2013GB004598)
12. D'Elia CF, Webb KL, Porter JW. 1981 Nitrate-rich groundwater inputs to discovery bay, Jamaica: a significant source of N to local coral reefs? *Bull. Mar. Sci.* **31**, 903–910.
13. Beddows PA, Smart PL, Whitaker FF, Smith SL. 2002 Density stratified groundwater circulation on the Caribbean Coast of the Yucatan Peninsula, Mexico. In *Hydrogeology and biology of post-paleozoic carbonate aquifers* (eds JB Martin, CM Wicks, ID Sasowsky), pp. 129–134. Leesburg, VA: Karst Waters Institute.
14. Blanco AC, Watanabe A, Nadaoka K, Motooka S, Herrera EC, Yamamoto T. 2011 Estimation of nearshore groundwater discharge and its potential effects on a fringing coral reef. *Mar. Pollut. Bull.* **62**, 770–785. (doi:10.1016/j.marpolbul.2011.01.005)
15. Luijendijk E, Gleeson T, Moosdorf N. 2020 Fresh groundwater discharge insignificant for the world's oceans but important for coastal ecosystems. *Nat. Commun.* **11**, 1260. (doi:10.1038/s41467-020-15064-8)
16. Moore WS. 2010 The effect of submarine groundwater discharge on the ocean. *Ann. Rev. Mar. Sci.* **2**, 59–88. (doi:10.1146/annurev-marine-120308-081019)
17. Lecher AL, Mackey KRM. 2018 Synthesizing the effects of submarine groundwater discharge on marine biota. *Hydrology* **5**, 60. (doi:10.3390/hydrology5040060)
18. Lubarsky K, Silbiger NJ, Donahue MJ. 2018 Effects of submarine groundwater discharge on coral accretion and bioerosion on two shallow reef flats. *Limnol. oceanogr.* **63**, 1660–1676. (doi:10.1002/lno.10799)
19. Paytan A, Shellenbarger GG, Street JH, Gonneea ME, Davis K, Young MB, Moore WS. 2006 Submarine groundwater discharge: an important source of new inorganic nitrogen to coral reef ecosystems. *Limnol. Oceanogr.* **51**, 343–348. (doi:10.4319/lo.2006.51.1.0343)
20. Kim G, Kim J-S, Hwang D-W. 2011 Submarine groundwater discharge from oceanic islands standing in oligotrophic oceans: implications for global biological production and organic carbon fluxes. *Limnol. Oceanogr.* **56**, 673–682. (doi:10.4319/lo.2011.56.2.0673)
21. Prouty NG, Cohen A, Yates KK, Storlazzi CD, Swarzenski PW, White D. 2017 Vulnerability of coral reefs to bioerosion from land-based sources of pollution. *J. Geophys. Res. C: Oceans* **122**, 9319–9331. (doi:10.1002/2017JC013264)
22. Cyronak T, Santos IR, Erler DV, Eyre BD. 2013 Groundwater and porewater as major sources of alkalinity to a fringing coral reef lagoon (Muri Lagoon, Cook Islands). *Biogeosciences* **10**, 2467. (doi:10.5194/bg-10-2467-2013)
23. Fabricius KE. 2005 Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis. *Mar. Pollut. Bull.* **50**, 125–146. (doi:10.1016/j.marpolbul.2004.11.028)
24. Szmant AM. 2002 Nutrient enrichment on coral reefs: is it a major cause of coral reef decline? *Estuaries* **25**, 743–766. (doi:10.1007/BF02804903)
25. Lapointe BE. 1997 Nutrient thresholds for bottom-up control of macroalgal blooms on coral reefs in Jamaica and southeast Florida. *Limnol. Oceanogr.* **42**, 1119–1131. (doi:10.4319/lo.1997.42.5_part_2.1119)
26. Smith J, Smith C, Hunter C. 2001 An experimental analysis of the effects of herbivory and nutrient enrichment on benthic community dynamics on a Hawaiian reef. *Coral Reefs* **19**, 332–342. (doi:10.1007/s003380000124)
27. D'Angelo C, Wiedenmann J. 2014 Impacts of nutrient enrichment on coral reefs: new perspectives and implications for coastal management and reef survival. *Curr. Opin. Environ. Sustain.* **7**, 82–93. (doi:10.1016/j.cosust.2013.11.029)
28. La Valle F, Thomas FIM, Nelson CE. 2019 Macroalgal biomass, growth rates, and diversity are influenced by submarine groundwater discharge and local hydrodynamics in tropical reefs. *Mar. Ecol. Prog. Ser.* **621**, 51–67. (doi:10.3354/meps12992)
29. Quinlan ZA *et al.* 2018 Fluorescent organic exudates of corals and algae in tropical reefs are compositionally distinct and increase with nutrient enrichment. *Limnol. Oceanogr. Lett.* **3**, 331–340. (doi:10.1002/lol2.10074)
30. Kelly LW *et al.* 2014 Local genomic adaptation of coral reef-associated microbiomes to gradients of natural variability and anthropogenic stressors. *Proc. Natl Acad. Sci. USA* **111**, 10 227–10 232. (doi:10.1073/pnas.1403319111)
31. Haas AF *et al.* 2016 Global microbialization of coral reefs. *Nat. Microbiol.* **1**, 16042. (doi:10.1038/nmicrobiol.2016.42)
32. McMahon A, Santos IR. 2017 Nitrogen enrichment and speciation in a coral reef lagoon driven by groundwater inputs of bird guano. *J. Geophys. Res. C: Oceans* **122**, 7218–7236. (doi:10.1002/2017JC012929)
33. Richardson CM, Dulai H, Popp BN, Ruttenberg K, Fackrell JK. 2017 Submarine groundwater discharge drives biogeochemistry in two Hawaiian reefs. *Limnol. Oceanogr.* **62**, S348–S363. (doi:10.1002/lno.10654)
34. Prouty NG, Yates KK, Smiley N, Gallagher C, Cheriton O, Storlazzi CD. 2018 Carbonate system parameters of an algal-dominated reef along West Maui. *Biogeosciences* **15**, 2467–2480. (doi:10.5194/bg-15-2467-2018)
35. Kroeker KJ, Kordas RL, Crim RN, Singh GG. 2010 Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. *Ecol. Lett.* **13**, 1419–1434. (doi:10.1111/j.1461-0248.2010.01518.x)
36. Pandolfi JM, Connolly SR, Marshall DJ, Cohen AL. 2011 Projecting coral reef futures under global warming and ocean acidification. *Science* **333**, 418–422. (doi:10.1126/science.1204794)
37. Silbiger NJ, Donahue MJ. 2015 Secondary calcification and dissolution respond differently to future ocean conditions. *Biogeosciences* **12**, 567–578. (doi:10.5194/bg-12-567-2015)
38. Silbiger NJ, Guadalupe O, Thomas FIM, Donahue MJ. 2016 A novel μ CT analysis reveals different responses of bioerosion and secondary accretion to environmental variability. *PLoS ONE* **11**, e0153058. (doi:10.1371/journal.pone.0153058)
39. Andersson AJ, Gledhill D. 2013 Ocean acidification and coral reefs: effects on breakdown, dissolution, and net ecosystem calcification. *Ann. Rev. Mar. Sci.* **5**, 321–348. (doi:10.1146/annurev-marine-121211-172241)
40. Albright R, Takeshita Y, Koweek DA, Ninokawa A, Wolfe K, Rivlin T, Nebuchina Y, Young J, Caldeira K. 2018 Carbon dioxide addition to coral reef waters suppresses net community calcification. *Nature* **555**, 516–519. (doi:10.1038/nature25968)
41. Kleypas JA, Langdon C. 2006 Coral Reefs and Changing Seawater Carbonate Chemistry. In *Coral reefs and climate change: science and management*, pp. 73–110. Arlington, VA: American Geophysical Union.
42. DeCarlo TM, Cohen AL, Barkley HC, Cobban Q, Young C, Shamberger KE, Brainard RE, Golbuu Y. 2015 Coral macrobioerosion is accelerated by ocean acidification and nutrients. *Geology* **43**, 7–10. (doi:10.1130/G36147.1)
43. Silbiger NJ, Nelson CE, Remple K, Sevilla JK, Quinlan ZA, Putnam HM, Fox MD, Donahue MJ. 2018 Nutrient pollution disrupts key ecosystem functions on coral reefs. *Proc. Biol. Sci.* **285**, 20172718. (doi:10.1098/rspb.2017.2718)
44. Courtney TA *et al.* 2017 Environmental controls on modern scleractinian coral and reef-scale calcification. *Sci Adv* **3**, e1701356. (doi:10.1126/sciadv.1701356)
45. Silbiger NJ, Sorte CJB. 2018 Biophysical feedbacks mediate carbonate chemistry in coastal ecosystems across spatiotemporal gradients. *Sci. Rep.* **8**, 796. (doi:10.1038/s41598-017-18736-6)
46. Li L, Barry DA, Stagnitti F, Parlange J-Y. 1999 Submarine groundwater discharge and associated chemical input to a coastal sea. *Water Resour. Res.* **35**, 3253–3259. (doi:10.1029/1999WR900189)
47. Santos IR, Eyre BD, Huettel M. 2012 The driving forces of porewater and groundwater flow in permeable coastal sediments: a review. *Estuar. Coast. Shelf Sci.* **98**, 1–15. (doi:10.1016/j.ecss.2011.10.024)
48. Richardson CM, Dulai H, Whittier RB. 2017 Sources and spatial variability of groundwater-delivered nutrients in Maunaloa Bay, O'ahu, Hawai'i. *J. Hydrol. Regional Stud.* **11**, 178–193. (doi:10.1016/j.ejrh.2015.11.006)
49. Swarzenski PW, Dulaiova H, Dailer ML, Glenn CR, Smith CG, Storlazzi CD. 2013 A Geochemical and

- Geophysical Assessment of Coastal Groundwater Discharge at Select Sites in Maui and O'ahu, Hawai'i. In *Groundwater in the coastal zones of Asia-Pacific*, pp. 27–46. Dordrecht, The Netherlands: Springer.
50. Wolanski E, Martinez JA, Richmond RH. 2009 Quantifying the impact of watershed urbanization on a coral reef: Maunalua Bay, Hawaii. *Estuar. Coast. Shelf Sci.* **84**, 259–268. (doi:10.1016/j.ecss.2009.06.029)
 51. La Valle FF, Kantar MB, Nelson CE. In press. Coral reef benthic community structure is associated with the spatiotemporal dynamics of submarine groundwater discharge chemistry. *Limnol. Oceanogr.* **9999**, 1–13. (doi:10.1002/lno.11596)
 52. Storlazzi CD, Katherine Presto M, Logan JB, Field ME. 2010 *Coastal circulation and sediment dynamics in Maunalua Bay, Oahu, Hawaii, measurements of waves, currents, temperature, salinity, and turbidity; November 2008–February 2009*. Open-File Report. Reston, VA: US Geological Survey. (doi:10.3133/ofr20101217)
 53. Dickson AG, Sabine CL, Christian JR. 2007 *Guide to best practices for ocean CO₂ measurements*. Silver Spring, MD: NOAA.
 54. Oehler T, Tamborski J, Rahman S, Moosdorf N, Ahrens J, Mori C, Neuholz R, Schnetger B, Beck M. 2019 DSI as a tracer for submarine groundwater discharge. *Front. Mar. Sci.* **6**, 563. (doi:10.3389/fmars.2019.00563)
 55. Maldonado M, Riesgo A, Bucci A, Tzlerb KRÜ. 2010 Revisiting silicon budgets at a tropical continental shelf: silica standing stocks in sponges surpass those in diatoms. *Limnol. Oceanogr.* **55**, 2001–2010. (doi:10.4319/lno.2010.55.5.2001)
 56. Dulaiova H, Gonnessa ME, Henderson PB, Charette MA. 2008 Geochemical and physical sources of radon variation in a subterranean estuary—Implications for groundwater radon activities in submarine groundwater discharge studies. *Mar. Chem.* **110**, 120–127. (doi:10.1016/j.marchem.2008.02.011)
 57. Karl D. 2019 Niskin bottle water samples and CTD measurements from the Hawaii Ocean Time-Series cruises from 1988–2016 (HOT project). *Biol. Chem. Oceanogr. Data Manage. Office.* (doi:10.1575/1912/bco-dmo.3773.1)
 58. Cyronak T *et al.* 2018 Taking the metabolic pulse of the world's coral reefs. *PLoS ONE* **13**, e0190872. (doi:10.1371/journal.pone.0190872)
 59. Chisholm JRM, Gattuso J-P. 1991 Validation of the alkalinity anomaly technique for investigating calcification of photosynthesis in coral reef communities. *Limnol. Oceanogr.* **36**, 1232–1239. (doi:10.4319/lno.1991.36.6.1232)
 60. Yeakel KL, Andersson AJ, Bates NR, Noyes TJ, Collins A, Garley R. 2015 Shifts in coral reef biogeochemistry and resulting acidification linked to offshore productivity. *Proc. Natl Acad. Sci. USA* **112**, 14 512–14 517. (doi:10.1073/pnas.1507021112)
 61. Gattuso J-P, Epitalon J-M, Lavigne H, Orr J. 2019 seacarb: seawater carbonate chemistry. R package version 3.2. 12.
 62. Courtney TA, Andersson AJ. 2019 Evaluating measurements of coral reef net ecosystem calcification rates. *Coral Reefs* **38**, 997–1006. (doi:10.1007/s00338-019-01828-2)
 63. Grace JB, Bollen KA. 2008 Representing general theoretical concepts in structural equation models: the role of composite variables. *Environ. Ecol. Stat.* **15**, 191–213. (doi:10.1007/s10651-007-0047-7)
 64. Bürkner PC. 2017 An R package for bayesian multilevel models using Stan. *J. Stat. Softw.* **80**, 1–28. (doi:10.18637/jss.v080.i01)
 65. Bürkner P-C. 2018 Advanced Bayesian Multilevel Modeling with the R package brms. *The R Journal*, **10**, 395–411. (doi:10.32614/RJ-2018-017)
 66. Gelman A, Rubin DB. 1992 Inference from iterative simulation using multiple sequences. *Stat. Sci.* **7**, 457–472. (doi:10.1214/ss/1177011136)
 67. Kay M. 2018 tidybayes: tidy data and geoms for Bayesian models. (doi:10.5281/zenodo.1308151)
 68. Cai W-J *et al.* 2011 Acidification of subsurface coastal waters enhanced by eutrophication. *Nat. Geosci.* **4**, 766–770. (doi:10.1038/ngeo1297)
 69. Zeebe RE, Wolf-Gladrow D. 2001 *CO₂ in seawater: equilibrium, kinetics, isotopes*. Bremerhaven, Germany: Gulf Professional Publishing.
 70. Atkinson MJ. 2011 Biogeochemistry of Nutrients. In *Coral reefs: an ecosystem in transition* (eds Z Dubinsky, N Stambler), pp. 199–206. Dordrecht, The Netherlands: Springer Netherlands.
 71. Gil MA. 2013 Unity through nonlinearity: a unimodal coral–nutrient interaction. *Ecology* **94**, 1871–1877. (doi:10.1890/12-1697.1)
 72. Shamberger KEF, Feely RA, Sabine CL, Atkinson MJ, DeCarlo EH, Mackenzie FT, Drupp PS, Butterfield DA. 2011 Calcification and organic production on a Hawaiian coral reef. *Mar. Chem.* **127**, 64–75. (doi:10.1016/j.marchem.2011.08.003)
 73. Silverman J, Lazar B, Erez J. 2007 Effect of aragonite saturation, temperature, and nutrients on the community calcification rate of a coral reef. *J. Geophys. Res.* **112**, 1. (doi:10.1029/2006jc003770).
 74. Woodhead AJ, Hicks CC, Norström AV, Williams GJ, Graham NAJ. 2019 Coral reef ecosystem services in the Anthropocene. *Funct. Ecol.* **33**, 1023–1034. (doi:10.1111/1365-2435.13331)