Meso-tetra(dioxanyl)porphyrins: Neutral, low molecular weight, and chiral porphyrins with solubility in aqueous solutions

Xu-Liang Jiang*ab, Dinusha Damunupolaa◊ and Christian Brückner⁰*a

^aDepartment of Chemistry, University of Connecticut, 55 N Eagleville Rd., Storrs, CT 06269-3060, USA ^bSchool of Pharmaceutical Engineering, Shenyang Pharmaceutical University, Shenyang, 110016, China

Received 11 April 2021 Accepted 21 May 2021

Dedicated to Professor Jonathan S. Lindsey on the occasion of his 65th birthday

ABSTRACT: The synthesis of the low-molecular weight, neutral, yet water-soluble porphyrin *meso*-tetra(dioxan-2-yl)porphyrin is described. The key intermediate dioxan-2-carbaldehyde is accessible in either racemic or in stereo-pure forms from commercially available starting materials in three steps. Using 4×1 or 2 + 2-type syntheses provide the porphyrin in modest yields. While the racemic aldehyde created an intractable mixture of diastereomers, the enantiopure aldehyde created a single enantiomer of the target porphyrin. The porphyrin was spectroscopically characterized. As its free base or zinc complex, it showed excellent solubility properties in organic and aqueous solvents, though free water-solubility was not achieved. The work expands on the availability of chiral porphyrins and neutral porphyrins with considerable solubility in aqueous solution.

KEYWORDS: *meso*-alkylporphyrins, water-soluble porphyrins, 4×1 and 2 + 2-type porphyrin syntheses, water-soluble porphyrins.

INTRODUCTION

The utilization of porphyrins and hydroporphyrins in, for example, cancer photodynamic therapy [1–4], photoantifungal [5] and -antimicrobials [6], as fluorescence labels [7], fluorescence or photoacoustic imaging agents [4, 8-15], or *in vivo* chemosensors [16, 17] requires their solubility in aqueous biological media. Aqueous solubility is frequently also wanted for their utilization in a number of technical applications, such as (photo)catalysis [18–20] or chemosensing [21]. While water solubility of the porphyrins can be mediated by surfactants or their incorporation into liposomes [22–24], their inherent aqueous

solubility is often more practical and thusly more desirable.

Early and effective efforts to impart water-solubility onto *meso*-arylporphyrins, the most commonly used class of fully synthetic porphyrins [25], relied on the introduction of cationic or anionic *meso*-aryl substituents, such as pyridinium-, ammonium-, phosphonate-, sulfonate-, or carboxylate-functionalities [26]. Examples are the classic tetrasulphonated porphyrin 1, the tetracationic tetrakis-pyridinium porphyrins, but also newer examples, like octacarboxylate 2 (Fig. 1) [27]. These porphyrins are generally prepared *via* Adler-type syntheses of suitably (neutral) precursors, followed by their direct sulfonation, quaternization, or modification with esters and subsequent saponification, respectively.

While successful in generating water-soluble porphyrins, these methods have drawbacks; for instance, the sulfonation reaction condition in particular is harsh. Lindsey and co-workers presented a series of structurally

Copyright © World Scientific Publishing Company

[♦]SPP full member in good standing.

^{*}Correspondence to: (XLJ) tel.: +86-24-43520243, email: xuliangjiang@syphu.edu.cn (CB) tel.: +1-860-486-2743, email: c.bruckner@uconn.edu.

Fig. 1. Examples of literature-known water-soluble porphyrins.

(CH2CH2O)4CH3

H₃C(OH₂CH₂C)₄

diverse swallowtail porphyrins [28] (and hydroporphyrins [29]) made by total syntheses that elegantly circumvented such harsh post-macrocycle synthesis functional group manipulations to fashion water-soluble derivatives, such as compound 3, shown to be suitable for bio-conjugation to monoclonal antibodies [30, 31]. We also note the small molecular weight of this compound. However, the chromatographic purification of polyionic porphyrins can be difficult [26].

An alternative method to impart water-solubility into charge-neutral porphyrins late in their synthetic sequence is their conjugation to polyethylene glycols (PEG) moieties [26, 32, 33]. One example among the *meso*-aryl series is PEG-based dendrimer-modified compound **4** [26]. Aqueous media solubility could also be achieved, *inter alia*, by conjugation of the porphyrins to oxygen-rich polar molecules like cyclodextrans [34],

(unprotected) sugars [35, 36], or PEGs [26]. However, in order to achieve the high overall oxygen contents needed to achieve water-solubility, their molecular weight increased, often drastically (cf. compound 4).

(CH₂CH₂O)₄CH₃

H₃C(OH₂CH₂C)₄

MW = 3490.43 g/mol

The lowest molecular weight porphyrin, archetype porphin $C_{20}H_{14}N_4$, exhibits low solubility in any solvent [37, 38]. *Meso*-substitution, on the other hand, generally is efficient in preventing porphyrin macrocycle π - π aggregation and thusly improving their solubility. This suggests the replacement of the *meso*-aryl groups by small, possibly non-aromatic oxygen/heteroatomrich entities, such as furanyl- [39, 40], hydropyranyl-, morpholinyl-, or dioxanyl-moieties, *etc.*, or even just short PEG chains. Only an astonishingly small number such, and related, systems have been described, among them (water-soluble and unprotected) *meso*-carbohydrate-substituted porphyrins [41–46]. On the

downside, the chemical stability of some pyranosylsubstituted porphyrins was described as being low [41] or the solubility of the hydroxy-protected derivatives in aqueous solution were not specified [46]. Known *meso*furanyl- [39, 40] and -(1,3-dithian-2-yl)porphyrins [47] are not sufficiently water-soluble. Outside of the patent literature [48], *meso*-dioxanyl-substituted porphyrins have not been described. However, we are also aware that seemingly simple porphyrins may show unexpected solubility or conformational effects [47, 49–51].

This contribution describes our preliminary results on the formation and properties of the $4 \cdot 1$ and 2 + 2-type syntheses and aqueous solubility of *meso*-tetra(dioxanyl) porphyrin, a novel small molecular weight porphyrin with considerable solubility in aqueous solution.

RESULTS AND DISCUSSION

Synthesis of dioxanylaldehyde 9

Crucial starting materials in the syntheses of *meso*-substituted dipyrromethanes, some 2 + 2 syntheses of porphyrins utilizing these dipyrromethanes, and $4 \cdot 1$ -type syntheses of porphyrins are suitable aldehydes carrying the future *meso*-substituent. A suitable aldehyde, dioxan-2-yl-carbaldehyde 9, prepared in steps from commercially available racemic epichlorohydrin 5^{rac} (Scheme 1), is known [52]. In short, reaction of epichlorohydrin 5^{rac} with 2-chloroethanol generated

the ring opened ether 6 that underwent a base-induced ring-closure to form epoxide 7^{rac} . Under more forcing base conditions, it underwent an intramolecular epoxide opening, forming the intermediate alcohol dioxanyl-2-hydroxymethyl 8^{rac} [52, 53]. The final step is the oxidation of the alcohol to the final aldehyde. Instead of the Swern oxidation described by Cushman and co-workers [53], we resorted to an oxidation using an oxoammonium salt (Bobbitt's salt) for the simplicity of this oxidation protocol and the absence of malodorous side products [54, 55]. Most significant in this reaction sequence is the fact that starting from enantiomerically pure (and also commercially available) epichlorohydrins $5^s/5^R$, the enantiomerically pure dioxanylaldehydes $9^R/9^S$, respectively, can also be prepared [53].

4×1 Synthesis of *meso*-tetradioxanylporphyrins 11

Reaction of the racemic dioxanylcarbaldehydes 9^{rac} under Adler conditions [56] failed to produce any porphyrin. Under Lindsey conditions [57] (Scheme 1), we retrieved traces of a purple product with a porphyrintype UV-vis spectrum, the expected composition for the target *meso*-tetradioxanylporphyrin 11^{rac} (as probed by HR-MS, ESI+ mode), but the ¹H NMR spectroscopic analyses of this fraction indicated the presence of many products we found intractable. This was attributed to the presence of a host of diastereomers. It was finally decided to simplify the syntheses and abandon the use of the racemic mixture of the aldehyde (9^{rac}).

Scheme 1. Synthetic route toward racemic dioxanylcarbaldehydes 9 from commercially available epichlorohydrins 5 and their conversion along 4 • 1 or 2 + 2-type synthetic routes to title porphyrin 11. ^aYield from literature [53]. ^bYield from literature [52].

 Indeed, the use of chiral aldehyde 9^R under Lindsey conditions generated a single isomer of a pink porphyrin 11^S in 2–3% yield (Scheme 1). Balaban and co-workers prepared a range of *meso*-alkylporphyrins using Montmorillonite clay as catalysts in the presence of 3 Å molecular sieves [58]. When these conditions were applied to the chiral aldehyde 9^R , the same pink products 11^S were obtained as a pure fraction possessing identical and porphyrin-like UV-vis spectrum (Fig. 2a) in improved, albeit still modest, yields of 3–6%. Nonetheless, this allowed the preparation of about 40 mg of this porphyrin per batch at a half-gram aldehyde scale. For the characterization of porphyrin 11^S , see below.

2 + 2 Synthesis of meso-tetradioxanylporphyrins 10^S

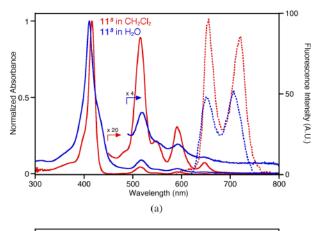
The 2 + 2-type syntheses of porphyrins are typically applied for the synthesis of A_2B_2 -type systems [59], but it was felt that this approach toward the target A_4 -porphyrin could be advantageous. The synthesis of the dipyrromethane building block 10^S from the corresponding enantio-pure aldehyde 9^R under the optimized Lindsey conditions that also avoided any chromatographic workup [60] was straight-forward and yielded the product as an off-white powder in 35% yield (400 mg per 580 mg aldehyde). Its 1H and ^{13}C NMR spectra showed all the diagnostic signals for the pyrrolyland dioxanyl-moieties, as well as the hydrogen atom at the central sp 3 -hybridized carbon atom (for details, see ESI). This dipyrromethene was used without further purification.

A 2 + 2 synthesis using dipyrromethane 10^{S} and aldehyde 9^{R} under the standard acid-catalyzed conditions [61, 62] generated the corresponding porphyrin 11^{S} in 23% yield (Scheme 1). Based on the amount of aldehyde required over the two steps, this yield is only slightly higher than in the best $4 \cdot 1$ synthesis of the porphyrin, but the reactions are robust, more readily scalable, and the product porphyrin is readily isolated. No premature decomposition during the handling, zinc insertion (see below), or chromatographic purification of the porphyrin was noticed.

The insertion of the zinc into the free base 11st proceeded, as expected, under mild and standard conditions [63] (Scheme 1). Metal insertion resulted in the expected changes in the optical spectra of the porphyrin (Fig. 2b). Metal insertion is also indicated by the disappearance of the inner NH proton signals in the ¹H NMR of the product (see ESI for details).

Spectroscopic characterization of the *meso*-tetradioxanylporphyrins and its zinc (II) complex

The UV-vis and fluorescence emission spectra of free base dioxanylporphyrin 11^s and its zinc complex 11^sZn in CH₂Cl₂ are, as expected, typical for a porphyrin/metalloporphyrin [64] (Fig. 2, Table 1): Sharp Soret



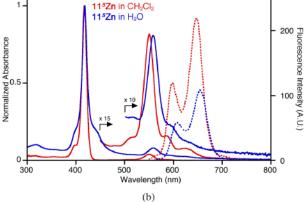


Fig. 2. UV-vis absorption (solid traces) and fluorescence emission spectra (broken traces) of 11^s (A) and 11^sZn (B) in CH_2Cl_2 (red traces) and pure, oxygenated H_2O (blue traces); $\lambda_{excitation} = \lambda_{Soret}$. The multiplication factors listed also reflect the concentration increase.

bands and, for the free base, four Q-bands with the λ_{max} band at 647 nm, and a two-band emission with a small Stoke's shift. The very weak absorbance of the longest wavelength of absorbance was noted at 647 nm. The corresponding peaks of the spectra in H_2O are only slightly shifted, suggesting no major aggregation, although the ratio of the intensity of the Soret band to the side bands is reduced. As could have been expected [31, 65], the fluorescence quantum yield of the compounds in (oxygenated) H_2O is only about 50% of that of the compounds in CH_2Cl_2 .

The ¹H NMR spectrum of the dioxanylporphyrin 11^s shows the diagnostic signals for a 4-fold symmetric *meso*-substituted porphyrin (Fig. 3): a sharp singlet (at 9.96 ppm, 4H) assigned to the •-protons and the much high-field shifted inner NH proton signal, here at -2.8 ppm (1H). This also suggests the absence of any diastereomers (including atropisomers, at 25°C). Because of the vicinity to the diatropic ring current, all of the dioxanyl protons are much more shifted and better resolved than those in the aldehyde or dipyrromethane (see ESI). Supported by H,H-COSY spectra (see ESI),

Table 1. UV-vis absorption and fluorescence emission peaks and fluorescence quantum yeilds of 11s and 11sZn in CH2Cl2 and H2O.

Compound	Solvent	UV-vis Absorbance [nm]	Fluorescence Emission [nm]	Fluorescence Quantum Yield
11 ^s	CH ₂ Cl ₂	416 (Soret), 515, 546 590, 647 nm	656, 720 nm	0.030a
11 ^s	H ₂ O (air)	415 (Soret), 519, 547, 592, 649 nm	650, 706 nm	0.014 ^a
11 ^s Zn	CH_2Cl_2	417 (Soret), 551 nm	598, 648 nm	0.036^{b}
11 ^s Zn	H ₂ O (air)	418 (Soret), 558 nm	654, 610 nm	0.017^{b}

^ameso-tetraphenylporphyrin in CH_2Cl_2 used as standard ($\phi = 0.13$); ^b[meso-tetraphenylporphyrinato]zinc(II) in CH_2Cl_2 used as standard ($\phi = 0.033$) [66].

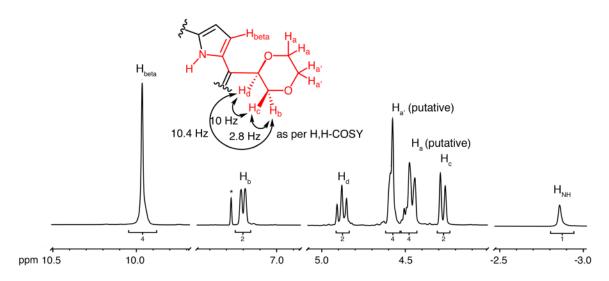


Fig. 3. ¹H NMR (400 MHz, CDCl₃) spectrum of meso-tetra((S)-1,4-dioxanyl)porphyrin 11^S.

the dd at 4.9 ppm (2H) was assigned to the proton at the *ipso*-carbon, the much-separated d at 7.19 and 4.28 ppm (2H each) to diastereotopic methylene protons at the C3-position, and the two multiplets at 4.58 and 4.45 ppm (4H each) to the two diastereotopic remaining methylene protons (at C5 and C6).

Aqueous solubility of the meso-tetradioxanylporphyrins 11^{s} and 11^{s} Zn

In their solid forms, the *meso*-tetradioxanylporphyrins 11^s and 11^sZn do not dissolve in aqueous solutions. But both 11^s and 11^sZn readily dissolve in methanol and we successively added aliquots of concentrated (33 and 50 mg/mL, respectively) solutions of the porphyrins in methanol to water to access the solubility in aqueous solutions. Methanol concentration after addition of aliquots was maintained well below 0.2% to avoid its influence on solubility of the porphyrin in water. Using UV-vis spectroscopy, we observed at which concentration we could detect the first signs of turbidity (elevation of baseline) and/or changes in the UV-vis

that could indicate aggregation (see ESI). Free base dioxanylporphyrin 11^S exhibited an aqueous solubility of, conservatively estimated, • than 0.05 mg/mL and likely exceeding 0.1 mg/mL (1.5 • 10^4 mol/L) (at which point the aqueous solution contained less than 0.2% MeOH). The corresponding zinc complex proved to be about twice as soluble, with a solubility exceeding > 0.20 mg/mL (2.8 • 10^4 mol/L).

CONCLUSIONS

The key aim of this study — the synthesis of a low-molecular weight, neutral, yet water-soluble porphyrin — could be accomplished. Thus, the synthesis of *meso*-tetra(dioxan-2-yl)porphyrin using the key intermediate dioxan-2-carbaldehyde is possible. Unlike the sugar-derived *meso*-pyranose-based porphyrins, however, the synthesis of the title compound did not require any protection group strategies and it is chemically stable. The key intermediate dioxan-2-carbaldehyde can be synthesized in stereoselective fashion from chiral and

commercially available starting materials in The presence of one stereocenter in each meso-substituents poses challenges and opportunities. The challenges arise from the creation of a complex mixture of a number of diastereomeric porphyrins upon utilization of a racemic mixture of the aldehyde. The opportunities arise from the ability to generate the enantiomerically pure isomers of the porphyrin, as demonstrated for the tetra-((S)-1,4dioxanyl)porphyrin 11^s and its zinc complex 11^sZn (though direct evidence of their stereochemical purity was not shown). The neutral products show considerable (for a porphyrin) solubility in aqueous solutions (1-3 • 10⁻⁴ mol/L range).

The preparation of the enantiomeric analogues to 11^R/11^RZn, the study of their chiroptic properties, the implications of the ability to readily prepare a pair of enantiomers, the biological uptake of the (chiral) *meso*tetra(dioxan-2-yl)porphyrins [67], their conversion to the corresponding hydroporphyrins, as well as the synthesis of other porphyrinoids (such as 5,15-disubstituted porphyrins [62], patterned porphyrins [68], or corroles [69]) incorporating the *meso*-dioxan-2-yl groups are currently being pursued in our group.

Acknowledgments

This work was supported by the US National Science Foundation (NSF) through grant CHE-1800361 (to CB) and the China Scholarship Council (to XJ).

Supporting information

A reproduction of the experimental data (such as ¹H, ¹³C NMR spectra, FT-IR, MS spectra, and excitation emission spectra) of all novel compounds described is given in the supplementary material (Figs S1–S21). This material is available free of charge *via* the Internet at https://www.worldscientific.com/doi/suppl/10.1142/S108842462150070X.

REFERENCES

- 1. Sternberg ED, Dolphin D and Brückner C. *Tetrahedron* 1998; **54**: 4151–4202.
- 2. Dolmans DEJGJ, Fukumura D and Jain RK. *Nature Rev. Cancer* 2003; **3**: 380–387.
- Brandis AS, Salomon Y and Scherz A. In *Chloro-phylls and Bacteriochlorophylls*, Grimm B, Porra RJ, Rüdinger W, Scheer H. (Eds.) Springer: Dordrecht, NL, 2006; pp. 485–494.
- 4. Ethirajan M, Chen Y, Joshi P and Pandey RK. *Chem. Soc. Rev.* 2011; **40**: 340–362.
- Donnelly RF, McCarron PA and Tunney MM. Microbiol. Res. 2008; 163: 1–12.
- 54 6. Wainwright M. *Photodiagn. Photodyn. Ther.* 2009;55 6: 167–169.
- Jiang J, Taniguchi M and Lindsey JS. New J. Chem.
 2015; 39: 4534–4550.

 Stender AS, Marchuk K, Liu C, Sander S, Meyer MW, Smith EA, Neupane B, Wang G, Li J, Cheng J-X, Huang B and Fang N. *Chem. Rev.* 2013; 113: 2469–2527.

- 9. Pandey RK, James N, Chen Y and Dobhal MP. *Top. Heterocycl. Chem.* 2008; **14**: 41–74.
- Weissleder R and Pittet MJ. Nature 2008; 452: 580–589.
- Kim C, Favazza C and Wang LV. Chem. Rev. 2010; 110: 2756–2782.
- 12. Luo S, Zhang E, Su Y, Cheng T and Shi C. *Biomaterials* 2011; **32**: 7127–7138.
- 13. Wang LV and Hu S. Science 2012; 335: 1458-1462.
- 14. Weber J, Beard PC and Bohndiek SE. *Nat. Methods* 2016; **13**: 639–650.
- Luciano M, Erfanzadeh M, Zhou F, Zhu H, Bornhütter T, Röder B, Zhu Q and Brückner C. *Org. Biomol. Chem.* 2017; 15: 972–983.
- Spencer JA, Ferraro F, Roussakis E, Klein A, Wu J, Runnels JM, Zaher W, Mortensen LJ, Alt C, Turcotte R, Yusuf R, Cote D, Vinogradov SA, Scadden DT and Lin CP. *Nature* 2014; 508: 269–273.
- Lemon CM, Karnas E, Han X, Bruns OT, Kempa TJ, Fukumura D, Bawendi MG, Jain RK, Duda DG and Nocera DG. J. Am. Chem. Soc. 2015; 137: 9832–9842.
- Barona-Castano JC, Carmona-Vargas CC, Brocksom TJ and de Oliveira KT. *Molecules* 2016; 21: 310.
- 19. Rybicka-Jasinska K, Ciszewski LW and Gryko D. *Adv. Synth. Catal.* 2016; **358**: 1671–1678.
- Rybicka-Jasinska K, Shan W, Zawada K, Kadish KM and Gryko D. J. Am. Chem. Soc. 2016; 138: 15451–15458.
- Worlinsky JL, Halepas S and Brückner C. *Org. Biomol. Chem.* 2014; 12: 3991–4001.
- Paszko E and Senge MO. Curr. Med. Chem. 2012;
 19: 5239–5277.
- 23. Huynh E, Lovell JF, Helfield BL, Jeon M, Kim C, Goertz DE, Wilson BC and Zheng G. *J. Am. Chem. Soc.* 2012; **134**: 16464–16467.
- Lovell JF, Jin CS, Huynh E, Jin H, Kim C, Rubinstein JL, Chan WCW, Cao W, Wang LV and Zheng G. *Nat. Mater.* 2011; 10: 324–332.
- 25. Lindsey JS. In *The Porphyrin Handbook*, Vol. 1, Kadish KM, Smith KM, Guilard R. (Eds.) Academic Press: San Diego, 2000; pp. 45–118.
- Luciano M and Brückner C. Molecules 2017; 22: 980.
- 27. Guldi DM, Zilbermann I, Anderson G, Li A, Balbinot D, Jux N, Hatzimarinaki M, Hirsch A and Prato M. *Chem. Commun.* 2004: 726–727.
- 28. Thamyongkit P, Speckbacher M, Diers JR, Kee HL, Kirmaier C, Holten D, Bocian DF and Lindsey JS. *J. Org. Chem.* 2004; **69**: 3700–3710.
- 29. Borbas KE, Ruzie C and Lindsey JS. *Org. Lett.* 2008; **10**: 1931–1934.

2 3

4

5

6

7

8

9

10

11

12

13

14 15

16

17

18 19

20

21

22

23 24

25

26

27

28

29

30

31

32

33

34

35

36

37

38 39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54 55

56

57

- 30. Borbas KE, Mroz P, Hamblin MR and Lindsey JS. 1 2 Bioconjugate Chem. 2006; 17: 638-653.
- 3 31. Borbas KE, Chandrashaker V, Muthiah C, Kee HL, 4 Holten D and Lindsey JS. J. Org. Chem. 2008; 73: 5 3145-3158.
- 6 32. Hambright P. In The Porphyrin Handbook, Vol. 3, 7 Kadish KM, Smith KM, Guilard R. (Eds.) Aca-8 demic Press: San Diego, 2000; pp. 129-210.

9

- 33. Pisarek S, Maximova K and Gryko D. Tetrahedron 10 2014; 70: 6685-6715.
- 11 34. Králová J, Kejík Z, Bríza T, Poucková P, Král A, 12 Martásek P and Král V. J. Med. Chem. 2010; 53: 13 128-138.
- 14 35. Singh S, Aggarwal A, Bhupathiraju NV, Arianna 15 G, Tiwari K and Drain CM. Chem. Rev. 2015; 115: 16 10261-10306.
- 17 36. Moylan C, Scanlan EM and Senge MO. Curr. Med. 18 Chem. 2015; 22: 2238-2348.
- 19 37. Senge MO and Davis M. J. Porphyrins Phthalocya-20 nines 2010; 14: 557-567.
- 21 38. Devillers CH, Fleurat-Lessard P and Lucas D. In 22 Handbook of Porphyrin Science, Vol. 37, Kadish 23 KM, Smith KM, Guilard R. (Eds.) World Scientific 24 Publishing Company: Singapore, 2016; pp. 75–231.
- 25 39. Gupta I and Ravikanth M. Tetrahedron Lett. 2002; 26 **43**: 9453-9455.
- 27 40. Gupta I and Ravikanth M. Tetrahedron 2003; 59: 28 6131-6139.
- 29 41. Maillard P, Huel C and Momenteau M. Tetrahedron 30 Lett. 1992; 33: 8081-8084.
- 31 42. Cornia M, Menozzi M, Ragg E, Mazzini S, Sca-32 rafoni A, Zanardi F and Casiraghi G. Tetrahedron 33 2000; 56: 3977-3983.
- 43. Cornia M, Binacchi S, Del Soldato T, Zanardi F and 34 35 Casiraghi G. J. Org. Chem. 1995; 60: 4964-4965.
- 36 44. Cornia M, Casiraghi G, Binacchi S, Zanardi F and 37 Rassu G. J. Org. Chem. 1994; 59: 1226-1230.
- 38 45. Casiraghi G, Cornia M, Rassu G, Del Sante C and 39 Spanu P. Nat. Prod. Lett. 1992; 1: 45-50.
- 40 46. Vedachalam S, Choi B-H, Pasunooti KK, Ching KM, Lee K, Yoon HS and Liu X-W. MedChem-41 42 Comm 2011; 2: 390-396.
- 43 47. Wacker P, Dahms K, Senge MO and Kleinpeter E. J. 44 Org. Chem. 2007; 72: 6224-6231.
- 45 48. Kim M, Ye J, Yoo B, Yoon J, Hwang J, Kim S and 46 Jeong J. Porphyrin-Based Transition Metal Com-47 plexes and Use as Dopants in Organic LEDs; Patent 48 US20200006677, 2020. 49

- 49. Veyrat M, Ramasseul R, Turowska-Tyrk I, Scheidt WR, Autret M, Kadish KM and Marchon J-C. Inorg. Chem. 1999; 38: 1772-1779.
- 50. Veyrat M, Ramasseul R, Marchon J-C, Turowska-Tyrk I and Scheidt WR. New J. Chem. 1995; 19: 1199-1202.
- 51. Senge MO and Davis M. Acta Crystallogr., Sect. E: Struct. Rep. Online 2010; E66: m790.
- 52. Pallavicini M, Valoti E and Villa L. Enantiomer 2001; 6: 267-273.
- 53. Kim HY, Kuhn RJ, Patkar C, Warrier R and Cushman M. Bioorg. Med. Chem. 2007; 15: 2667-2679.
- 54. Bobbitt JM, Brückner C and Merbouh N. Org. React. 2009; 74: 106-424.
- 55. Bobbitt JM, Bartelson AL, Bailey WF, Hamlin TA and Kelly CB. J. Org. Chem. 2014; 79: 1055-1067.
- Adler AD, Longo FR, Finarelli JD, Goldmacher J, Assour J and Korsakoff L. J. Org. Chem. 1967; 32:
- 57. Lindsey JS and Wagner RW. J. Org. Chem. 1989; **54**: 828-836.
- 58. Plamont R, Kikkawa Y, Takahashi M, Kanesato M, Giorgi M, Chan Kam Shun A, Roussel C and Balaban TS. Chem.—Eur. J. 2013; 19: 11293-11300.
- 59. Gryko DT and Tasior M. Tetrahedron Lett. 2003; **44**: 3317–3321.
- 60. Laha JK, Dhanalekshmi S, Taniguchi M, Ambroise A and Lindsey JS. Org. Process Res. Dev. 2003; 7: 799-812.
- 61. Lee C-H and Lindsey JS. Tetrahedron 1994; 50: 11427-11440.
- 62. Boyle RW, Brückner C, Posakony J, James BR and Dolphin D. Org. Synth. 1999; 76: 287-293.
- 63. Buchler JW. In *The Porphyrins*, Vol. 1, Dolphin D. (Ed.), Academic Press: New York, 1978; pp. 389-483.
- 64. Gouterman M. In *The Porphyrins*, Vol. 3, Dolphin D. (Ed.) Academic Press: New York, 1978; pp. 1–165.
- 65. Turro NJ, Ramamurthy V and Scaiano JC. Modern Molecular Photochemistry of Organic Molecules; University Science Books: California, 2010.
- 66. Taniguchi M, Lindsey JS, Bocian DF and Holten D. J. Photochem. Photobiol., C 2021; 46: 100401.
- 67. Trivedi ER, Harney AS, Olive MB, Podgorski I, Moin K, Sloane BF, Barrett AG, Meade TJ and Hoffman BM. Proc. Natl. Acad. Sci. USA. 2010; **107**: 1284–1288.
- 68. Lindsey JS. Acc. Chem. Res. 2010; 43: 300-311.
- 69. Briñas RP and Brückner C. Synlett 2001: 442–444.

50

51

52

53

54

55 56

57