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### Inversion breakpoints and the evolution of supergenes

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#### **Abstract**

The coexistence of discrete morphs that differ in multiple traits is common within natural populations of many taxa. Such morphs are often associated with chromosomal inversions, presumably because the recombination suppressing effects of inversions help maintain alternate adaptive combinations of alleles across the multiple loci affecting these traits. However, inversions can also harbor selected mutations at their breakpoints, leading to their rise in frequency in addition to (or independent from) their role in recombination suppression. In this review, we first describe the different ways that breakpoints can create mutations. We then critically examine the evidence for the breakpoint-mutation and recombination suppression hypotheses for explaining the existence of discrete morphs associated with chromosomal inversions. We find that the evidence that inversions are favored due to recombination suppression is often indirect. The evidence that breakpoints harbor mutations that are adaptive is also largely indirect, with the characterization of inversion breakpoints at the sequence level being incomplete in most systems. Direct tests of the role of suppressed recombination and breakpoint mutations in inversion evolution are thus needed. Finally, we emphasize how the two hypotheses of recombination suppression and breakpoint mutation can act in conjunction, with implications for understanding the emergence of supergenes and their evolutionary dynamics. We conclude by discussing how breakpoint characterization could improve our understanding of complex, discrete phenotypic forms in nature.

Keywords: chromosomal inversion, linkage, mutation, recombination, genome evolution.

#### Supergenes and complex phenotypes

The origin and maintenance of discrete morphs within natural populations has long fascinated evolutionary biologists (Darwin, 1862; Fisher, 1930; Ford, 1971). Because of the discrete nature of the phenotypic differences observed, this variation was often assumed to have a genetic origin, making its study of primary interest for early geneticists (see Chapter 6 of Ford 1971 for an overview of genetic programs of the time). Moreover, the diverse nature of the traits that usually differ between these morphs (color, morphology, sexual-compatibility, etc.), led to the hypothesis that the genetic basis of this discrete variation is a functional unit containing multiple linked loci, termed a "Super-gene" (spelled supergene hereafter; see glossary) by Darlington and Mather in 1949 (Darlington & Mather, 1949; Fisher, 1930; Ford, 1965; Mather, 1955). Indeed, as early as 1930 Fisher had anticipated that genetic linkage between multiple selected loci could evolve under natural selection (Fisher, 1930) and Ford later suggested that linkage could be increased via translocation or chromosomal inversions (Ford, 1971). Chromosomal inversions can indeed strongly reduce recombination between the genomic segments they cover, a property leading to their discovery in 1921 by Sturtevant (Sturtevant, 1921). Specifically, single crossing-over events within the rearranged region in heterokaryotypic individuals produce non-viable or unfit gametes with unbalanced gene content (Hoffmann & Rieseberg, 2008; Kirkpatrick, 2010; Liehr et al., 2019). Recombination between chromosomal rearrangements is still possible when double crossing-over events occur, making recombination more likely to happen at the center of inversions than near breakpoints (Hoffmann & Rieseberg, 2008; Stump et al., 2007). Nonetheless, such double crossing-over events are relatively rare, estimated to occur with a frequency of the order of 10<sup>-4</sup> to 10<sup>-3</sup> per generation in *Drosophila* (Stevison, Hoehn, & Noor, 2011).

Examples of discrete morphs associated with a region of suppressed recombination (*i.e.*, candidate supergenes; see glossary) are accumulating rapidly. In the majority of cases, suppressed recombination is due to an inversion (Schwander, Libbrecht, & Keller, 2014). However, in some instances, suppressed recombination is due to other mechanisms such as close physical proximity of highly diverged DNA sequences (Tetsuyuki Entani et al., 2003; Ushijima et al., 2003), proximity to a chromosomal centromere (T. Entani et al., 1999), or hemizygosity (Li et al., 2016). These findings have thus piqued interest in the evolution of supergenes and inversions, forming the central topic of the current review. Because there are already several excellent general reviews of supergenes and inversions (Faria, Johannesson, Butlin, & Westram, 2019; Schwander et al., 2014; Wellenreuther & Bernatchez, 2018), we focus here specifically on distinguishing two hypotheses for their evolution, a topic that has received less attention.

There are currently two core hypotheses for how and why supergenes and inversions evolve. The first and generally most accepted hypothesis is that inversions are selected due to their effect on suppressing recombination between sets of epistatic or locally adapted genes (*i.e.*, supergene hypothesis; see glossary) (Charlesworth & Barton, 2018; Kirkpatrick, 2010; Kirkpatrick & Barton, 2006). The second hypothesis is that inversions create adaptive mutations at their breakpoints (*i.e.* breakpoint-mutation hypothesis; see glossary; Fig. 1A) (Dobzhansky, 1947, 1970; Kirkpatrick, 2010), leading to their rise in frequency via selection on breakpoint variants. As we discuss in more detail below, these two hypotheses are not mutually exclusive and might combine to drive inversion and supergene evolution. Specifically, inversions could be selected due to their effect on suppressing recombination between an adaptive breakpoint variant and an adaptive variant within the rearrangement (*i.e.* breakpoint-linkage hypothesis; see glossary; Fig. 1A). In a similar manner, breakpoint

mutations could be favored because of their property to suppress recombination outside of the inversion (R. B. Corbett-Detig, 2016), locking together multiple variants favored in particular combinations, inside and/or outside the inversion.

Distinguishing among these hypotheses (Fig. 1A) is important because it provides insight into the relative significance of novel mutation, standing genetic variation, and recombination in adaption, thus addressing a major issue in evolution concerning the relationship between adaptation and genome rearrangement (Fig. 1B). However, empirical testing of these hypotheses can be a challenging task as it requires the quantification and localization of adaptive variants in regions of suppressed recombination (here inversions). When multiple traits are associated with an inversion, a role for recombination suppression between multiple variants controlling different traits is usually assumed. However, a breakpoint mutation can be pleiotropic and affect multiple traits (Fisher, 1930), making the identification of adaptive variants within the region of suppressed recombination necessary.

While the supergene hypothesis and the breakpoint-linkage hypothesis seem similar, they imply different evolutionary histories for supergenes (Fig. 1B). Under the supergene hypothesis, divergently selected variants evolve first and subsequently become locked together in favorable combinations because of an inversion (Charlesworth & Barton, 2018; Kirkpatrick & Barton, 2006). Under the breakpoint-linkage hypothesis two main evolutionary scenarios for supergene emergence are possible. In the first scenario, inversions first establish because they generate an adaptive breakpoint mutation when they form. Only subsequently might inversions evolve into supergenes by the emergence of new variants within them (Navarro & Barton, 2003). In the second scenario, inversions establish because they couple an already existing adaptive variant with a new adaptive breakpoint mutation. This last

exist by facilitating the initial rise in frequency of an inversion. Indeed, under the supergene hypothesis, theory implies that for new inversions to establish, they must capture most segregating variants at divergently selected loci in favorable combinations (Charlesworth & Barton, 2018; Kirkpatrick & Barton, 2006). However, an inversion with an adaptive breakpoint mutation may not need to lock all or a majority of the segregating variation in favorable combinations to initially rise in frequency due to the advantageous breakpoint mutation. If the inversion persists for a sufficient amount of time because of a selected breakpoint mutation, this may subsequently allow for the sorting out of any initially captured maladapted alleles (e.g., via double crossing-over events or noncrossover gene conversion) to create a supergene with a now higher selective value.

In this review, we critically examine the evidence for the supergene and breakpoint-mutation hypotheses, and their potential combination (*i.e.* the breakpoint-linkage hypothesis). We begin with a discussion of the mechanisms by which breakpoints can create adaptive mutations, as this is often a less appreciated facet of inversions than is recombination suppression. We then empirically review well-known cases of candidate supergenes (Box 2 – Fig. 1). Our collective findings lead us to propose that characterization of breakpoints at the sequence level could improve our understanding of supergenes. Specifically, a common molecular mechanism generating inversions also generates duplications at both ends of the inverted DNA segment (Ranz et al., 2007). This offers a unique opportunity to potentially date inversions independently from their content to test if they emerged before or after the divergently selected variants they lock together via suppressed recombination. Thus, breakpoint characterization could be important to understand the origin and maintenance of complex, discrete phenotypic forms in nature.

#### Molecular mechanisms by which inversion breakpoints can create mutations

The idea that inversions might be selected because of an adaptive mutation at one of their breakpoints is not new (Dobzhansky, 1947). However, this hypothesis is yet to receive highly focused study in supergene evolution, despite known molecular mechanisms that are likely to create breakpoint mutations when inversions form. Two main processes can generate chromosomal inversions, ectopic recombination and staggered breaks. Ectopic recombination generates inversions via a recombination event between two homologous sequences oriented head-to-head along a chromosome (Box 1 for further details; Box 1 – Fig. 1A). Staggered breaks can generate inversions via the complete detachment of a DNA segment from a chromosome and its subsequent reattachment in the opposite orientation at the same position in the chromosome (Box 1; Box 1 – Fig. 1B). In the *Drosophila melanogaster* species subgroup, the group best characterized for inversion breakpoints at present (R. B. Corbett-Detig, 2016), the majority of inversions are generated by staggered breaks (Ranz et al., 2007). While ectopic recombination leaves little molecular trace, staggered breaks can generate duplications at both breakpoints of the newly formed inversion (Box 1; Box 1 – Fig. 1B). As explained in more detail below, these breakpoint duplications provide a possible means to accurately date inversions, independently from the divergently selected variants they contain and, thus, could potentially allow us to resolve among the competing scenarios for supergene emergence (Fig. 1B). Consequently, if staggered breaks are primarily responsible for generating inversions in groups in addition to *D. melanogaster* flies, this could be a helpful tool for characterizing the emergence of supergenes (for details see the section Future directions).

Both ectopic recombination and staggered breaks can generate important phenotypic effects, in principle, by changing the expression level of a gene (or set of genes) at breakpoints (Fig. 2A). However, while differential expression of genes at breakpoints is common, it is not always observed (Said et al., 2018). Furthermore, differential expression is usually not restricted to genes at breakpoints, but is also detected for many genes inside and outside inversions (Cheng, Tan, Hahn, & Besansky, 2018; Lavington & Kern, 2017; Said et al., 2018). This suggests that the differential expression of genes at breakpoints potentially works in conjunction with other genetic changes to affect phenotypic variation, or is sometimes a by-product of genome rearrangement rather than being the actual cause for observed phenotypic variation. Breakpoints can also create chimeric genes (see glossary; Fig. 2B), which might sometimes have a beneficial effect, as documented now in a diverse set of taxa (Carvunis et al., 2012; Guillén & Ruiz, 2012; Jones et al., 2012; Stewart & Rogers, 2019). For example, in Drosophila mojavensis a chimeric gene at an inversion breakpoint is associated with the detoxification of toxic compounds in cacti (Guillén & Ruiz, 2012). Another example is found in the threespine stickleback Gasterosteus aculeatus, where a gene located at an inversion breakpoint on chromosome XI harbors different exon sequences depending on the inversion orientation, and is associated with differences in muscular development between lake and oceanic phenotypes (Jones et al., 2012). While speculative, an inversion breakpoint could create a new pleiotropic chimeric gene that affects multiple traits. Moreover, staggered breaks often generates duplications at both breakpoints in the derived sequence (Box1 – Fig. 1), allowing for the creation of chimeric genes without loss of the donor genes (Fig. 2B). This property will also facilitate neo-functionalization of such duplicated genes.

As we will see below, direct evidence that breakpoint mutations drive supergene evolution is lacking, but the mechanisms by which they might do so are established. With these

considerations in mind, we now turn to evaluating the role of suppressed recombination and breakpoint mutation in the evolution of candidate supergenes associated with chromosomal inversions.

The supergene hypothesis: what is the evidence for recombination suppression among loci controlling selected trait variation?

A core prediction of the supergene hypothesis (*i.e.*, that suppressed recombination is advantageous) is that multiple, linked genetic variants control trait variation that differs between inversion forms. In contrast, if variation is controlled by a single locus with pleiotropic effects on several traits, suppressed recombination is not necessarily favored or required to explain the existence of discrete morphs that differ in these traits.

One line of indirect evidence supporting the supergene hypothesis is that inversions of intermediate size usually reach higher frequencies than small inversions, suggesting that they are selectively favored more frequently than small inversions (Cáceres, Barbadilla, & Ruiz, 1997; R. B. Corbett-Detig, 2016; Wellenreuther & Bernatchez, 2018). This is expected if larger inversions are more likely to lock physically distant selected variants together via suppressed recombination. In contrast, if the selective nature of an inversion solely stems from a breakpoint mutation, shorter inversions should be favored because they have less chance of capturing recessive deleterious mutations when they form (Cáceres et al., 1997; R. B. Corbett-Detig, 2016; Kirkpatrick, 2010).

Despite these observations, direct evidence for the supergene hypothesis is still lacking in most systems. Indeed, our review of the literature revealed that we still know very little about the number, location and identity of selected variants within candidate supergenes associated with inversions. This is not surprising given that determining the number of

adaptive loci within a region of suppressed recombination is difficult, because suppressed recombination frustrates attempts at fine-scale genetic mapping. The presence of two or more selected loci in a supergene was historically inferred from genetic crosses aimed at detecting rare recombinants (i.e., intermediate phenotypic forms) (Cain & Sheppard, 1954; Mather, 1950). For example, Mather described a genetic model where multiple selected loci locked within an inversion control heterostyly in Primula sinensis, a classic example of a putative supergene (Mather, 1950). However, recent work using modern genomic approaches suggests that Mather's simple genetic model is erroneous and needs modification (Li et al., 2016). Specifically, a 178 kilo base-pair indel containing five predicted genes is associated with the different floral types (Li et al., 2016), with one floral type being hemizygous for this DNA segment and the other completely lacking it (Table 1) (Li et al., 2016). A revision of the classic genetic model also occurred for the iconic land snail Cepaea nemoralis (Table 1), where cryptic shell color-pattern was long thought to be associated with five loci locked within an inversion (Cain & Sheppard, 1954). Recent work showed an absence of recombination in the putative supergene, casting doubt on the number and linkage relationships of loci controlling shell color (Gonzalez, Aramendia, & Davison, 2019). These recent developments urge for more rigorous tests on the true number of selected variants within supergenes and their precise genomic location(s).

To our knowledge, the existence of multiple selected loci within a supergene associated with an inversion has been demonstrated in only six systems, three involving plants, two involving animals, and one involving a fungus. Of these six, three involve segregation distorters (Table 1; Box 2). In addition, indirect but fairly convincing evidence exist for two other animal systems. Details of all the examples mentioned above are presented in Box 2. Clearly, further

work determining the number of loci within inversions and candidate supergenes that contribute to trait variation is warranted. In this regard, emerging techniques that allow functional genetic manipulation of structural variants (e.g., CRISPR-Cas9) hold promise for making such progress (Hopkins, Tyukmaeva, Gompert, Feder, & Nosil; Kraft et al., 2015).

The breakpoint-mutation hypothesis: what is the evidence that functional variants reside in or near inversion breakpoints?

A core assumption of the breakpoint-mutation hypothesis is that functional variants reside in or near inversion breakpoints. We found evidence consistent with this hypothesis in three systems (*Papilio polytes, Solenopsis invicta* and *Timema cristinae*), but expect this number to increase when breakpoint characterization is completed in more systems (Table 1).

In the mimetic butterfly *Papilio polytes*, an inversion breakpoint disrupts a transcriptional regulator and affects the expression in embryonic wings of three neighboring genes potentially involved in wing color-pattern development (*UXT*, *U3X* and *prospero*; Table 1) (Nishikawa et al., 2015). In the fire ant *Solenopsis invicta*, a region of suppressed recombination generated via three overlapping inversions (Yan et al., 2020) is associated with different social structures of colonies (Wang et al., 2013). A breakpoint of the second inversion disrupts a gene (SI2.2.0\_02248) which is a strong candidate to explain differences in colony social structure, and the disrupted allele is associated with increased expression of this particular gene (Huang, Dang, Chang, & Wang, 2018; Yan et al., 2020). In the stick insect *Timema cristinae* a large deletion occurring at one breakpoint of a putative inversion deleted multiple loci known to affect multiple aspects of color in another species of the genus (Villoutreix et al., 2020).

Interestingly, with the exception of *Timema cristinae*, indications exist of putative selected variants within inversions at locations other than the breakpoints. Therefore, support for a strict version of the breakpoint-mutation hypothesis is scarce, with only one example potentially supporting it (*T. cristinae*). Even in the case of *T. cristinae*, genes controlling adaptive traits other than coloration, which combine with color alleles to determine fitness, could exist in locations in the inversion away from breakpoints. We discuss the implication of these observations in the following section.

# Mixed hypotheses: The breakpoint-linkage hypothesis and its implications for supergene emergence

As mentioned previously, the supergene and breakpoint-mutation hypotheses are not mutually exclusive. The selective advantage of an inversion could come from its property of impeding recombination between variants within the inversion and a favorable breakpoint mutation. We refer to this mixed hypothesis as the breakpoint-linkage hypothesis (see glossary), which predicts that loci controlling trait variation will be localized both at one (or both) breakpoint(s) and other regions within the inversion.

In theory, the breakpoint-linkage hypothesis could involve different processes and dynamics than those predicted by the classic supergene hypothesis. A supergene composed of a selected locus within an inversion and an adaptive breakpoint mutation suggests two main evolutionary scenarios. In the first, an inversion could originally establish because of an adaptive breakpoint mutation (Guerrero, Rousset, & Kirkpatrick, 2012), and then subsequently evolve into a supergene by the emergence of additional genetic variants within it (Navarro & Barton, 2003) (Fig. 1B). This scenario could be initiated in allopatry, where an inversion with an adaptive breakpoint sweeps to high frequency or fixation and is selected for

its recombination suppressing properties only later upon secondary contact (or introgression) and gene flow. In this case, allopatry allows for the evolution and capture of alternative sets of favorable alleles in alternate arrangements, bypassing the difficulties of supergene emergence when gene flow occurs and maladaptive alleles are segregating as standing variation within populations (Feder, Gejji, Powell, & Nosil, 2011).

In the second scenario, pre-existing genetic variants in the region where the inversion will occur could become associated with a new adaptive breakpoint mutation (Fig. 1B). Although formal modeling is required, it is possible that such a coupling could relax some of the restrictions of a model based strictly on suppressed recombination (Charlesworth & Barton, 2018; Kirkpatrick & Barton, 2006). For example, such coupling might mean that the initial inversion does not need to capture all or the majority of divergently selected variants in a favored combination in order to spread. Instead, variant sorting could occur after the establishment of the inversion, by double cross-over events (or noncrossover gene conversion (Korunes & Noor, 2017)) that disassociate adaptive and maladaptive alleles. In principle, the rise in frequency of the inversion and its equilibrium frequency could thus be dependent on the sum of adaptive values of the breakpoint and the variants it originally captures.

The breakpoint-linkage hypothesis predicts that loci controlling trait variation will be localized at one or both breakpoint(s) and within the inversion. Some empirical data are at least consistent with this prediction. For example, the butterfly (*P. polytes*) and fire ant (*S. invicta*) examples mentioned in the previous section as instances where functional loci reside in or near inversion breakpoints could involve the breakpoint-linkage hypothesis (Table 1). In *P. polytes*, the inversion associated with the breakpoint mutation, besides affecting the

expression of the three genes *UXT*, *U3X* and *prospero* potentially involved in mimetic wing color-pattern (*UXT*, *U3X*) and *prospero*), also contains a locus (away from any breakpoint) shown to affect mimetic wing color-pattern (*doublesex*) (Kunte et al., 2014; Nishikawa et al., 2015). In the fire ant *S. invicto*, in addition to the strong candidate gene for odor reception (*S12.2.0\_02248*) located at an inversion breakpoint, the region of suppressed recombination also contains (away from any breakpoints) a strong candidate gene for odor generation/reception (*Gp-9*), a mechanism at the root of social structure differences in fire ants (Wang et al., 2013).

Variations on the breakpoint-linkage hypothesis are possible. For example, another type of mixed model is one where the selective advantage of breakpoints stems from their property to suppress recombination outside of the inversion (R. B. Corbett-Detig, 2016), Recombination suppression outside of inversions could be achieved by a breakpoint disrupting a 'sensitive site' (a locus believed to be necessary to produce normal crossing-over frequencies in the surrounding genomic region) (R. B. Corbett-Detig, 2016). This could lock together multiple variants in particular combinations, inside and/or outside the inversion. This mechanism may be important in meiotic drive systems, where multiple selected loci are located within a region of suppressed recombination containing multiple, non-overlapping inversions. *Mus musculus* t-haplotypes and the *Drosophila melanogaster* SD locus could involve such a mixed model. In these two systems, individual inversions appear to very rarely contain more than one selected locus (Herrmann & Bauer, 2012; Larracuente & Presgraves, 2012) (See Box 2 for details).

While functional work is needed to better elucidate the role of different loci in controlling trait variation (*P. polytes* and *S. invicta*), or recombination suppression (*M. musculus* and *D.* 

melanogaster), these four systems provide some preliminary support for mixed hypotheses. Regardless of whether recombination suppression is most relevant for genes inside versus outside of an inversion or for both, these systems highlight the possibility for breakpoint mutations generated by a new chromosomal rearrangement to be integral members of the group of selected loci that form a supergene. The vast majority of candidate supergene systems still lack a complete characterization of inversion breakpoints (often one breakpoint is characterized at the DNA sequence level but not the other; Table 1). It is possible that more examples supporting the breakpoint-linkage hypothesis will emerge as breakpoints are better characterized in a greater number of systems.

#### **Future directions**

Future work could usefully focus in two directions. First, better information on the number, location, and identify of loci controlling trait variation within inversions is required. One way that this can be achieved is via genetic crosses to create recombinant forms, such as for the examples discussed in Box 2. Although the number of selected loci in an inversion determined through genetic crosses will tend to underestimate the actual number (because insufficient recombination may occur in these crosses to separate variants in close physical proximity), the results can still indicate whether or not multiple loci or variants are involved, thus confirming the existence of a supergene. As an alternative to controlled crosses, genome wide association mapping can be applied in species that are close relatives to those with discrete morphs and inversions, but that lack inversion polymorphism (Villoutreix et al., 2020). This will yield more precise estimates, but needs to be complemented with functional manipulation in the species with the inversion. Ultimately, functional manipulations such as those offered by CRISPR-Cas9 may be required to precisely and firmly establish the identities and numbers of variants

functionally contributing to trait variation, possibly by 'engineering' recombinant alleles to validate the independent effects of individual mutations on a phenotype (Hopkins et al., 2020; Kraft et al., 2015). Such gene editing may be particularly required for identification of causal genetic variation at breakpoints, as many variants will be in complete association in these regions due to the lack of recombination (Hoffmann & Rieseberg, 2008; Matzkin, Merritt, Zhu, & Eanes, 2005).

Second, once the selected variants within a region of suppressed recombination are identified, further work can be implemented to understand how the supergene emerged (Fig. 1B). More specifically, when variants are located inside the inversion boundaries, one can test if the variants predate or postdate the inversion itself. The usual procedure to compare inversion age to the age of the selected variants within them is to compare their times to the most recent common ancestor (TMRCA) using coalescence theory (Lee et al., 2017). Inversion age is estimated by calculating TMRCA on the inverted haplotypes only, while ages for loci within the inversion are obtained comparing all haplotypes (inverted and non-inverted). While being the current standard, this method suffers from several caveats. First, it does not date the inversion totally independently from the variants it contains. Second, it can overestimate inversion age relative to the selected variants within it and could thus result in the erroneous conclusion that the variants appeared after the inversion. The reason for such over-estimation is that double crossing-over at neutral loci within the rearranged region will reintroduce older alleles from non-inverted haplotypes into the inverted haplotypes, thus increasing the TMRCA for the inversion, and therefore the estimated age of the inversion. Duplications at breakpoints generated by staggered breaks, if neutral, could help circumvent these caveats. Indeed, breakpoint duplications would allow the inversion to be dated independently from its content, and the inversion age estimated using these duplications would not be as biased because double crossing-over events between non-inverted and inverted DNA sequences very rarely involve sequences at breakpoints (Hoffmann & Rieseberg, 2008). As a proof of concept, a breakpoint duplication was successfully used to date inversion age in *H. numata* butterflies (Jay et al., 2018).

Nonetheless, even this method using breakpoint duplications is not without difficulties for at least two reasons. First, it requires sequence-level characterization of breakpoints, a difficult endeavor because breakpoints often reside in highly repetitive regions of the genomes (Russell B. Corbett-Detig et al., 2019; da Silva et al., 2019), which are difficult to assemble using short-read sequencing technologies (Fig. 3A). Second, obtaining accurate age estimates for loci experiencing selection is difficult, and breakpoint duplications may not be always neutral. Still, promise for progress exists in sequence-level characterization of breakpoints.

Specifically, creative approaches such as those applied by Corbett-Detig and colleagues in Drosophila (Fig. 3B) (Russell B. Corbett-Detig, Cardeno, & Langley, 2012; Russell B. Corbett-Detig & Hartl, 2012) may help in characterizing breakpoints at the sequence level in other taxa. Longread technologies such as Nanopore (Jain, Olsen, Paten, & Akeson, 2016) and PacBio (Eid et al., 2009) should also be of great help for such characterization.

#### **Concluding remarks**

We outlined how inversions can facilitate supergene evolution by suppressed recombination, by inducing adaptive mutations at breakpoints or by a combination of these two mechanisms.

Surprisingly, definitive evidence for the existence of multiple selected loci within candidate supergenes associated with an inversion exist in only six systems. Thus, direct evidence for the supergene hypothesis is still scarce, although indirect lines of evidence in several systems are consistent with the hypothesis. Little is known about the emergence of supergenes,

specifically if divergently-selected variants predate inversions or if they accumulated within inversions after the inversion originated. Observations in multiple candidate supergene systems suggest a joint role for suppressed recombination and adaptive breakpoint mutation in supergene evolution. Thus, further tests of the breakpoint-linkage hypothesis are warranted. Distinguishing and confirming the role of breakpoints in supergene evolution will require characterization of inversion breakpoints at the sequence level, which has yet to be completed in most systems. We propose that further study of inversion breakpoints could improve our understanding of the origin and maintenance of complex, discrete phenotypic forms in nature, and the dynamics by which supergenes emerge. Although we focused here on morphs, similar processes could apply to the evolution of other discrete units of diversity such as ecotypes, species, and sexes. 

#### Glossary:

- 2 <u>Breakpoint-linkage hypothesis:</u> the selective advantage of an inversion stems from both a
- favorable effect of a mutation at one of its breakpoints and the recombination suppression
- 4 between this mutation and at least one other selected variant within the inversion.
- 5 <u>Breakpoint-mutation hypothesis (also referred as 'position effect hypothesis' in the</u>
- 6 literature): the selective advantage of an inversion stems from a favorable effect associated
- 7 with a mutation at one or both of its breakpoints.
- 8 <u>Candidate supergene:</u> a case where multiple co-varying and discrete phenotypes are
- 9 associated with a region of suppressed recombination (usually an inversion). The existence
- of multiple loci associated with these traits, within the region of suppressed recombination,
- must be demonstrated to advance the example from a candidate to a bonafide supergene.
- 12 <u>Chimeric gene</u>: a new gene constituted of sequences not previously associated together, for
- example, as parts of two genes or parts of genes and non-coding DNA. Here, we consider
- cases where the different DNA segments are brought together by an inversion breakpoint
- 15 (see Fig. 2B).
- 16 <u>Chromosomal inversion:</u> a chromosomal rearrangement in which a segment of a
- 17 chromosome is reversed end to end. This causes the inverted segment to have an opposite
- sequence order from the ancestral sequence and reduces recombination in individuals
- 19 heterozygous for the inversion. Recombination is especially reduced near chromosomal
- inversion breakpoints.
- 21 <u>Chromosomal inversion breakpoint</u> (shortened to 'inversion breakpoint' or simply
- 22 'breakpoint' in the main text and hereafter): the position(s) on a chromosome where DNA

23	sequence orientation shifts after the occurrence of a chromosomal inversion (i.e., the end of
24	an inversion where the genome shifts from inverted back to collinear). An inversion
25	generally has two breakpoints.
26	(Chromosomal) breakpoint mutation: sequence modification(s) at breakpoints resulting
27	from a chromosomal inversion. Mutations may also occur near breakpoints after an
28	inversion has occurred, however, here, we refer to changes associated with the creation of
29	the rearrangement itself.
30	<u>Discrete morphs:</u> individuals of the same species that exhibit discrete phenotypic
31	differences (e.g., in morphology, color, behavior, physiology, etc.). Here, we focus on cases
32	where morphs are differentiated by multiple co-varying phenotypic traits such that multiple
33	genes may be involved.
34	Supergene: a group of genes affecting different phenotypes so seldom separated by
35	crossing-over that in effect they operate as a single genetic entity (i.e., a non-recombining
36	locus). This end may be achieved by close genetic proximity on a chromosome (i.e., tight
37	physical linkage), by inclusion within an inversion, or by other molecular mechanisms
38	impeding crossing-over. This definition is modified from that of E. B. Ford (Ford, 1965).
39	Supergene hypothesis: supergenes evolve because suppressed recombination within them
40	maintains favorable allelic combinations at multiple loci that control different traits, thereby
41	favoring suppressed recombination.

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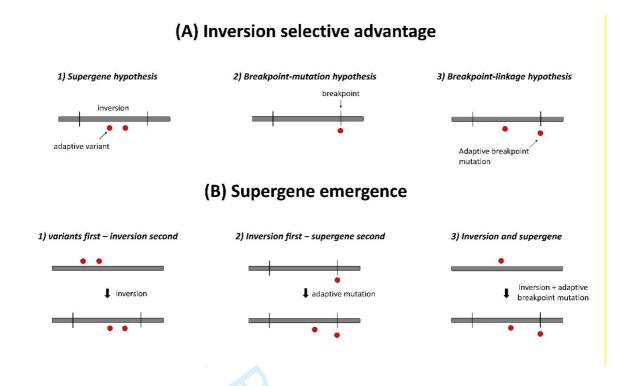


Figure 1 – Hypotheses underlying selective advantages to inversions and scenarios for supergene emergence: A. Hypotheses explaining a selective advantage to inversions. B. Scenarios for supergene emergence. Grey squares represent a chromosome; red dots are adaptive variants affecting a selected trait; black bars are inversion breakpoints.

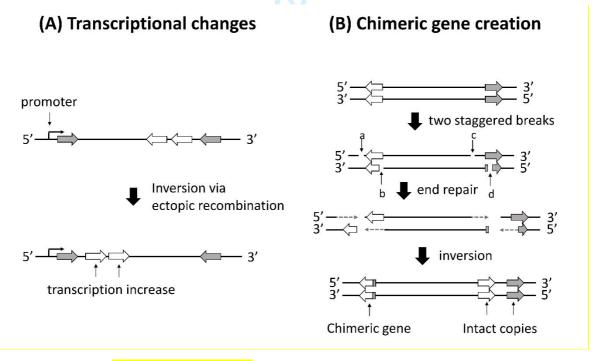


Figure 2 – Possible mechanisms creating mutations at inversion breakpoints: A. An inversion generated through ectopic recombination affects the transcription level of two genes by changing their positions in the genome relative to a promoter. B. An inversion generated through staggered breaks generates a new chimeric gene, while avoiding the loss of the donor genes because of duplication. Grey and black arrows represent genes, with

- orientation indicated via arrow direction; a, b, c, d represent single strand breaks, resulting
- in two staggered breaks.

## (A) Misassembly of inversion breakpoint duplications

## (B) Characterizing breakpoints sequences

Proximal breakpoint

Actual sequence of the derived conformation: end duplication end duplication

Reference inversion genome Assembly (short reads, 10X) Informative reads for breakpoint sequence assembly

Distal breakpoint

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Figure 3 - Characterizing breakpoints at the DNA sequence level: A. Using short-read sequencing technologies can lead to misassembly of inversion breakpoints, for example due to duplications generated through staggered breaks (Ranz et al., 2007). Grey and black arrows represent genes, with orientation indicated by the direction of arrows. B. A method to characterize breakpoint sequences using a single reference genome and short read data, as developed by Corbett-Detig and colleagues (Russell B. Corbett-Detig et al., 2012). Black arrows symbolize aligned reads on the reference genome, while grey arrows symbolize their unaligned paired reads. Breakpoint sequence characterization is possible by using the black and grey read pairs to generate *de novo* assembly of the breakpoint region. Figure redrawn and modified from (Russell B. Corbett-Detig et al., 2012) with the permission of the authors.

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83 Box 1 - Inversion breakpoints can create a diversity of types of genetic variation via 84 mutation. 85 We here discuss how inversions are formed and the associated effects of induced mutations 86 on genetic variation at breakpoints. Two known molecular mechanisms can generate 87 inversions: (1) ectopic recombination and, (2) staggered breaks (Box 1. - Fig1) (Ling & Cordaux, 88 2010; Ranz et al., 2007). 89 The first mechanism occurs from recombination between homologous sequences, usually 90 transposable elements (Ling & Cordaux, 2010) or paralogous genes, that are oriented headto-head in the original sequence, subsequently resulting in inversion of the sequence located 92 between these elements (Box 1 – Fig. 1A). This mechanism leaves little molecular trace and 93 generates cut-and-paste type breakpoints. The second mechanism involves staggered breaks 94 occurring in two positions of the original sequence (Ranz et al., 2007). Such breaks are usually 95 staggered, meaning that they will result in fragments with stretches of single stranded DNA 96 at their extremities (Box 1. - Fig. 1B). The DNA repair mechanism that synthesizes the reverse 97 complement of these single stranded stretches then joins the DNA sequences back together 98 in a non-homologous way, sometimes reinserting the DNA segment in the opposite 99 orientation, creating an inversion. When the breaks are heavily staggered (i.e., with long 100 single strand stretches), this results in inversions with duplicated sequences at both breakpoints in the derived sequence (Box 1. - Fig. 1B) (Ranz et al., 2007). In contrast, when 102 the breaks are blunt or slightly staggered, cut-and-paste type breakpoints are created in the 103 derived sequence, with no or small duplications, respectively. 104 Despite these clear mechanisms of inversion formation, we still know very little about genetic 105 variation at breakpoints and the mutations that breakpoints generate (Table 1). This stems

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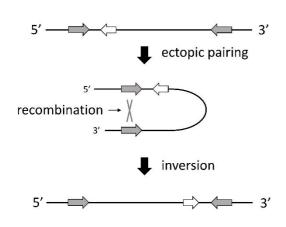
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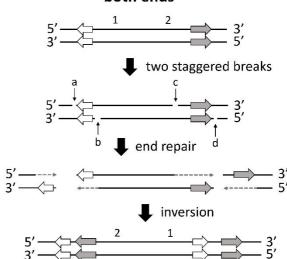
largely from the fact that inversion breakpoints often lie in very repetitive regions of the genome (Russell B. Corbett-Detig et al., 2019; da Silva et al., 2019) which are difficult to sequence and assemble and thus challenging to study (Fig. 3A). Nonetheless, some progress has been made, particularly in the *Drosophila melanogaster* species subgroup (R. B. Corbett-Detig, 2016; Russell B. Corbett-Detig & Hartl, 2012; McBroome, Liang, & Corbett-Detig, 2020; Ranz et al., 2007). In this system, staggered breaks appear to be the most common mechanism generating inversions, very often resulting in duplications at breakpoints (Ranz et al., 2007). We note, however, that locating breakpoints is not sufficient to identify their phenotypic effects. Obtaining the derived DNA sequence at breakpoints is necessary to try and understand possible phenotypic effects. Understanding such effects from DNA sequence modification is however not always a straightforward task, as the disruption of a gene sequence by a breakpoint can sometimes lead to increases in expression of this gene, as seen in fire ants (Yan et al., 2020). Nonetheless, phenotypic effects may be expected, for example, because both ectopic recombination and staggered breaks can change the expression level of a set of genes (Fig. 2A). In addition, staggered breaks can have another potential effect, in which the duplication of sequences leads to the creation of a new chimeric gene. This occurs via merging the initial part of a gene with another coding sequence, such as another gene or pseudo-gene (Fig. 2B). This phenomenon can happen without the loss of the originally disrupted genes (via their duplication), mitigating fitness costs. Thus, there are several possibilities by which breakpoints can create genetic variation with likely phenotypic effects. The observation that similar genomic regions are often re-used repeatedly as breakpoints for different inversions, including in different taxa, raises the

- possibility for a recurrent and even somewhat predictable role for breakpoints in evolution
- 130 (Pevzner & Tesler, 2003).

### (A) Ectopic recombination. Cut-andpaste breakpoints



# (B) Staggered breaks. Duplication at both ends



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Box 1 - Figure 1 - Known molecular mechanisms generating inversions: White and grey arrows represent genes, with their orientation indicated by the direction of the arrows. A. Ectopic recombination between similar elements (transposable elements or duplicated genes) in opposite orientation in a DNA segment leads to the inversion of the interspacing sequence. B. Fours single strand breaks (a, b, c, and d) lead to two staggered breaks. Through end repair and reattachment in the opposite orientation an inversion is generated with sequence duplication at both breakpoints (black and grey genes). The number above the DNA stretch are used here to illustrate the change in orientation of the sequence.

141	Box 2 - Determining the number of loci within inversions that contribute to selected trait
142	variation
143	Despite seventy years of research on supergenes, definitive evidence for the existence of
144	multiple selected variants within a region of suppressed recombination containing an
145	inversion exists only for a few systems, as described below.
146	Evidence for multiple loci controlling trait variation within an inversion has been
147	demonstrated in <i>Boechera stricta</i> (Box 2 – Fig. 1A), a perennial plant from the Rocky
148	Mountains. Here, two subspecies adapted to different climates have come into secondary
149	contact (Lee et al., 2017). The subspecies differ in multiple traits and through crosses between
150	different genotypes lacking the inversion it was demonstrated that three different QTLs (i.e.
151	Quantitative trait loci) are associated with these divergent traits and located within the
152	inversion region (Box 2 – Fig. 2B). Interestingly, independent of the genotype at these three
153	QTLs, the orientation of the DNA segment at the rearranged region explained a significant
154	amount of variance for these traits, without any apparent effect of the inversion breakpoints.
155	The molecular mechanism for this result is still unknown. The inversion is relatively young and
156	originated after secondary contact, likely locking together pre-existing selected variants (Lee
157	et al., 2017), but this interpretation is debated (Charlesworth & Barton, 2018).
158	In the Mimulus guttatus species complex, the existence of multiple selected loci within an
159	inversion has also been confirmed by genetic crosses, where two life strategies (perennial vs
160	annual) are maintained through divergent selection (Lowry & Willis, 2010). Specifically,
161	through crosses between the <i>Mimulus guttatus</i> annual type and a closely related perennial
162	species with a collinear gene order, Mimulus tilingii, Coughlan and Willis (Coughlan & Willis,

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2018) identified two QTLs within the inversion associated with the traits experiencing divergent selection. In the Brassicaceae self-incompatibility locus (S-locus), it was shown that two selected loci reside within a region of suppressed recombination containing inversions (Goubet et al., 2012; Takayama & Isogai, 2005). At the phenotypic level, self-incompatibility alleles are equivalent to mating types, where a plant stigma can recognize its own pollen via a specific interaction between a protein in the pollen coat and a receptor in the plant stigma, preventing fertilization (Durand et al., 2020; Takayama & Isogai, 2005). With this system, plants cannot self-fertilize and cannot be fertilized by pollen from plants expressing a similar mating type. Through knock-outs and gains of function experiments, the genes coding for the pollen protein (SCR) and the stigma receptor (SRK) have been identified (Schopfer, Nasrallah, & Nasrallah, 1999; Silva et al., 2001; Takasaki et al., 2000; Takayama & Isogai, 2005; Takayama et al., 1987; Takayama et al., 2000). These two genes are present in different positions and orientations in different S-locus haplotypes, suggesting that many inversions have occurred at the S-locus (Goubet et al., 2012). The S-locus region displays little sequence conservation (likely because of extensive degeneration), frustrating attempts at characterizing the exact breakpoints of these inversions or the ancestral orientation of the region (Goubet et al., 2012). The existence of multiple selected loci within a region of suppressed recombination containing inversions has also been verified in a spore killer system (Neurospora intermedia spore killer element Sk-2) and two meiotic drive systems (Mus musculus t-haplotypes and Drosophila melanogaster SD locus). In these three systems, a combination of 'killer' loci (named driver loci hereafter) and 'protective' loci (named response loci hereafter) kills spores

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or gametes with specific genotypes, resulting in increased transmission (up to 99%) of the spore or gamete killing element. Suppressed recombination between driver and response loci is necessary for the maintenance of drive systems over many generations (Hammond, Rehard, Xiao, & Shiu, 2012), qualifying them as supergene examples. The existence of two selected loci (one driver locus and one response locus) was demonstrated in Neurospora intermedia (element Sk-2) through gene knock-outs and complementation (Hammond, Rehard, Xiao, & Shiu, 2012; Rhoades et al., 2019). In this system, a succession of non-overlapping inversions interspaced with repeats tightly link the driver locus with the response locus (Svedberg et al., 2018). The existence of five selected loci (four driver loci and one response locus) (Herrmann & Bauer, 2012) was also demonstrated in Mus musculus t-haplotypes meiotic drive through the generation of rare recombinants via genetic crosses and mutagenesis (Herrmann & Bauer, 2012; Lyon, 1984; Lyon & Meredith, 1964). Multiple interspaced non-overlapping inversions are present in the t-haplotype locus but none contain both a driver and the response locus, suggesting that recombination suppression is achieved via additional factors in this system (Kelemen & Vicoso, 2018). A similar technique to the one employed in the t-haplotype was used to decipher the genetic basis of SD meiotic drive in *Drosophila melanogaster* and confirmed the existence of five loci involved in this drive system (four driver loci and one response locus) (Hartl, 1974; Larracuente & Presgraves, 2012). Deletion of chromosomal segments via mutagenesis confirmed these results and allowed for fine mapping of the different loci (Ganetzky, 1977; Larracuente & Presgraves, 2012). As is the case for the *t-haplotype* locus, multiple interspaced non-overlapping inversions are present in the SD locus (different driver alleles are associated with different inversions), and inversions rarely contain a driver locus together with the

response locus, suggesting that additional factors also prevent recombination in this system (Larracuente & Presgraves, 2012).

Less direct, but reasonably strong, evidence for multiple selected loci within an inversion has also been reported in two insect systems (Table 1). Here, the approach was to compare the genetic basis of traits in closely related taxa that exhibit versus lack inversion polymorphism.

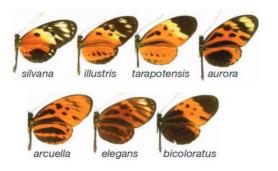
In the polymorphic butterfly *Heliconius numata* a region of suppressed recombination generated via multiple adjacent inversions controls variation in mimetic color morphs. The

In the polymorphic butterfly *Heliconius numata* a region of suppressed recombination generated via multiple adjacent inversions controls variation in mimetic color morphs. The same genomic region is known to contain at least three recombining loci associated with color variation in other species of the genus (Box 1 – Fig. 1D) (Huber et al., 2015; Mathieu Joron et al., 2011; Van Belleghem et al., 2017). It is therefore likely that the adjacent inversions in *H. numata* harbor multiple variants at these three loci that recombine in other *Heliconius* species. A similar example stems from *Timema*, a genus of stick insect that is endemic to the western USA and Mexico and that shows within-population variation for cryptic coloration (i.e., green versus melanistic morphs). In several species, morphs are highly discrete and controlled by a region of suppressed recombination on linkage group eight (named *'Mel-Stripe'*) (Nosil et al., 2018; Villoutreix et al., 2020). However, a recent study in a species displaying more quantitative color variation (*Timema chumash*) and a lack of suppressed recombination revealed that multiple genetic variants, and likely multiple genes within *Mel-Stripe*, control color variation (Villoutreix et al., 2020).

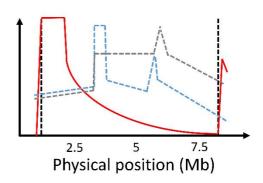
# (A) Boechera stricta



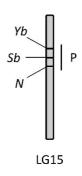
(C) H. numata aposematic morphs



# (B) Multiple QTL within *Bs1* inversion



# (D) H. numata colour supergene



Box 2 - Figure 1 – Flagship examples of supergenes: A. Boechera stricta subspecies types. The East subspecies has smaller leaves with more trichomes. Photo credit: Cheng-Ruei Lee.

B. Multiple QTLs were detected in the Bs1 inversion. The red, blue dashed and grey-dashed lines correspond to different composite traits obtained through discriminant function analysis. See Lee et al., 2017 for more details. The two black-dashed lines correspond to the position of the inversion breakpoints. Breakpoints are of the cut-and-paste type for the Bs1 inversion. Figure redrawn and modified with the permission of the authors. C. Heliconius numata aposematic morphs, involved in Müllerian mimicry rings with other butterflies. D. Aposematic morphs in H. numata are associated with adjacent inversions (spanning the genomic region indicated by P) on linkage group 15 (LG15), encompassing at least three loci (Yb, Sb and N) associated with mimetic color morphs in H. melpomene.

<b>#</b>	Species	Multiple traits?	Selection on phenotypes?	Map to an inversion?	Multiple loci?	Selection on recomb. sup.?	Adaptive breakpoint mutation?	Breakpoints sequence?	Variants first?
Aves									
1	Ruff - Philomachus pugnax	Yes. (Kupper et al., 2016; Lamichhaney et al., 2016)	Likely. (Kupper et al., 2016; Lamichhaney et al., 2016)	Yes. (Kupper et al., 2016; Lamichhaney et al., 2016)	Likely. (Kupper et al., 2016; Lamichhaney et al., 2016)	Not tested.	No. Counter- selected.  (Kupper et al., 2016; Lamichhane y et al., 2016)	Yes. (Kupper et al., 2016; Lamichhane y et al., 2016)	Not known.
2	White- throated sparrow - Zonotrichia albicollis	Yes. (Tuttle et al., 2016)	Yes. (Tuttle et al., 2016)	Yes. (Thomas et al., 2008; Thorneycroft, 1966; Throneycroft, 1975; Tuttle et al., 2016)	Likely. (Tuttle et al., 2016)	Not tested.	Not known.	No.	Not known.
Fungi									

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3	Neurospora intermedia Sk-2 spore killer	Yes.  (Hammond et al., 2012;  Rhoades et al., 2019)	Yes. (Turner & Perkins, 1979)	Yes. (Svedberg et al., 2018)	Yes.  (Hammond et al., 2012; Rhoades et al., 2019)	Not tested.	No.	Yes. (Svedberg et al., 2018)	Likely. (Svedber g et al., 2018)
Inve	rtebrates								
4	Anopheles – Anopheles gambiae	<mark>Likely.</mark> (Cheng et al., 2018)	Likely.  (Fouet, Gray, Besansky, & Costantini, 2012; Gray, Rocca, Costantini, & Besansky, 2009; Rocca, Gray, Costantini, & Besansky, 2009)	Yes.  (Cheng et al., 2018; Coluzzi, Sabatini, della Torre, Di Deco, & Petrarca, 2002; Coluzzi, Sabatini, Petrarca, & Di Deco, 1979; Fouet et al., 2009; Rocca et al., 2009)	Likely.  (Ayala et al., 2019; Cheng et al., 2012)	Not tested.	No.	Partly characterize d. (Cheng et al., 2018; Sharakhov et al., 2006)	Not known.
5	Grove snail - Cepaea nemoralis	Yes. (Cain & Sheppard, 1950, 1952)	Likely.  (Cain & Sheppard, 1950, 1954; Cameron & Cook, 2012)	Not known. (Gonzalez et al., 2019)	Not known.	Not known.	Not known.	No.	Not known.

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9	Fire ant - Solenopsis invicta	Yes. (Wang et al., 2013).	<mark>Likely.</mark> (Ross & Keller, 1995)	Yes. (Wang et al., 2013)	Likely. (Wang et al., 2013)	Not tested.	Likely. (Huang et al., 2018; Yan et al., 2020)	Yes. (Yan et al., 2020)	Not known.
10	Timema walking sticks - Timema cristinae	Yes. (Comeault et al., 2015)	Yes. (Cristina P. Sandoval, 1994; C. P. Sandoval & Nosil, 2005)	Likely. (Villoutreix et al., 2020)	Likely. (Villoutreix et al., 2020)	Not tested.	Likely. (Villoutreix et al., 2020)	Partly characterize d.  (Nosil et al., 2018; Villoutreix et al., 2020)	Not known.
Mam	<del>mals</del>			16/					
11	Commom mice – Mus musculus t-haplotypes	Yes. (Herrmann & Bauer, 2012)	Yes. (Chesley & Dunn, 1936; Herrmann & Bauer, 2012)	Yes.  (Herrmann & Bauer, 2012;  Kelemen & Vicoso, 2018)	Yes. (Herrmann & Bauer, 2012; Lyon, 1984)	Yes. (Herrmann & Bauer, 2012; Lyon, 1984)	No.	No.  (Kelemen & Vicoso, 2018)	Likely.  (Kelemen & Vicoso, 2018)
12	Humans – Homo sapiens	Yes. (Puig, Casillas, Villatoro, & Caceres, 2015;	Likely. (Stefansson et al., 2005)	Yes. (Puig et al., 2015; Stefansson et al., 2005)	Likely. (Puig et al., 2015)	Not tested.	Likely. (de Jong et al., 2012)	Yes. (Stefansson et al., 2005)	Not known.

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Plan	<u>ts</u>	Stefansson et al., 2005)							
13	Mustards- Brassicaceae Self- incompatibil ity (S-locus)	Yes. (Takayama & Isogai, 2005)	Yes. (De Nettancourt, 2001)	Yes. (Goubet et al., 2012)	Yes.  (Schopfer, Nasrallah, & Nasrallah, 1999; Silva et al., 2001; Takasaki et al., 2000; Takayama & Isogai, 2005; Takayama et al., 1987; Takayama et al., 2000)	Likely. (Durand et al., 2020)	No. (Goubet et al., 2012)	No. (Goubet et al., 2012)	<mark>Not</mark> known.
14	Drummond' s rockcress - Boechera stricta	Yes. (Lee et al., 2017)	Yes.  (Anderson, Lee, & Mitchell-Olds, 2011; Anderson, Lee, Rushworth, Colautti, & Mitchell-Olds,	Yes. (Lee et al., 2017)	Yes. (Lee et al., 2017)	Yes, but debated. (Charleswort h & Barton, 2018; Lee et al., 2017)	No. (Lee et al., 2017).	Yes. (Lee et al., 2017).	Yes. (Lee et al., 2017)

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			2013; Lee et al., 2017)						
15	Common yellow monkeyflow er -Mimulus guttatus species complex	Yes. (Coughlan & Willis, 2018; Lowry & Willis, 2010)	Yes. (Lowry & Willis, 2010)	Yes. (Coughlan & Willis, 2018; Lowry & Willis, 2010)	Yes. (Coughlan & Willis, 2018)	Not tested.	Not known.	No.	Not known.
<mark>16</mark>	Primrose - Primula sp.	Yes. (Darwin, 1862)	Likely. (Piper & Charlesworth, 2008)	No. (Li et al., 2016)	Likely. (Li et al., 2016)	Not tested.	N.A. Indel. (Li et al., 2016)	Yes. (Li et al., 2016)	N. A.
Tele	<mark>osts</mark>				Uh				
<b>17</b>	Atlantic Cod – Gadus morhua.	Yes. (Berg et al., 2016; Sinclair- Waters et al., 2018)	Likely. (Berg et al., 2016; Sinclair- Waters et al., 2018)	Yes.  (Berg et al., 2016;  Kirubakaran et al., 2016;  Sinclair-Waters et al., 2018)	Likely.  (Berg et al., 2016; Kirubakaran et al., 2016; Sinclair- Waters et al., 2018)	Not tested.	Not known.	No.	Not known.

18	Threespine sticklebacks - Gasterosteu s aculeatus	Yes. (Jones et al., 2012)	Likely. (Jones et al., 2012)	Yes. (Jones et al., 2012)	Likely.  (Albert et al., 2007; Colosimo et al., 2004)	Not tested.	Yes. (Jones et al., 2012)	Yes. (Jones et al., 2012)	Not known.
<b>19</b>	Steelhead/r ainbow trout – Oncorhynch us mykiss	Yes.  (Nichols, Edo, Wheeler, & Thorgaard, 2008; Pearse, Miller, Abadia-Cardoso, & Garza, 2014)	Not tested.	Likely.  (Nichols et al., 2008; O'Malley, Sakamoto, Danzmann, & Ferguson, 2003; Pearse et al., 2014; Robison, Wheeler, Sundin, Sikka, & Thorgaard, 2001)	Likely. (Lemopoulos, Uusi-Heikkilä, Huusko, Vasemägi, & Vainikka, 2018)	Not tested.	Not known.	No.	Not known.

Table 1 – Examples of candidate supergene. #: example number. Species: common and latin names. Multiple traits: Do morphs differ by multiple traits? Selection on phenotypes: Has selection on these traits been shown? Map to an inversion: Are these traits associated with an inversion? Multiple loci: Has the existence of multiple loci controlling the traits been demonstrated? Selection on recomb. sup.: Has the selective advantage of an inversion been demonstrated to come from its property of suppressing recombination between multiple selected loci? Adaptive breakpoint mutation: Is their evidence for selection acting on one or both of the breakpoints in the inversion? Breakpoint sequence: Have the breakpoints been characterized at the sequence level? Variants first?: Do adaptive variants in the putative supergene

- predate the emergence of the inversion? In each column, the term 'likely' represent cases were lines of evidence exist to support an item, but
- definitive evidence is currently lacking.



252	Bibliography
253	
254	Albert, A. Y. K., Sawaya, S., Vines, T. H., Knecht, A. K.,
255	Miller, C. T., Summers, B. R., Schluter, D. (2007).
256	The genetics of adaptative shape shift in stickleback:
257	Pleiotropy and effect size. <i>Evolution</i> , 62(1), 76-85.
258	Anderson, J. T., Lee, C. R., & Mitchell-Olds, T. (2011). Life-
259	History Qtls and Natural Selection on Flowering Time in
260	Boechera Stricta, a Perennial Relative of Arabidopsis.
261	Evolution, 65(3), 771-787. doi:10.1111/j.1558-
262	5646.2010.01175.x
263	Anderson, J. T., Lee, C. R., Rushworth, C. A., Colautti, R. I.,
264	& Mitchell-Olds, T. (2013). Genetic trade-offs and
265	conditional neutrality contribute to local adaptation. <i>Mol</i>
266	Ecol, 22(3), 699-708. doi:10.1111/j.1365-
267	294X.2012.05522.x
268	Ayala, D., Zhang, S., Chateau, M., Fouet, C., Morlais, I.,
269	Costantini, C., Besansky, N. J. (2019). Association
270	mapping desiccation resistance within chromosomal
271	inversions in the African malaria vector Anopheles
272	gambiae. <i>Mol Ecol, 28</i> (6), 1333-1342.
273	doi:https://doi.org/10.1111/mec.14880
274	Benson, W. W. (1972). Natural-Selection for Mullerian
275	Mimicry in Heliconius-Erato in Costa-Rica. Science,
276	176(4037), 936-939. doi:DOI
277	10.1126/science.176.4037.936
278	Berg, P. R., Star, B., Pampoulie, C., Sodeland, M., Barth, J.
279	M. I., Knutsen, H., Jentoft, S. (2016). Three
280	chromosomal rearrangements promote genomic
281	divergence between migratory and stationary ecotypes of
282	Atlantic cod. Scientific Reports, 6(23246).
283	doi:10.1038/srep23246

- Cáceres, M., Barbadilla, A., & Ruiz, A. (1997). INVERSION 284 LENGTH AND BREAKPOINT DISTRIBUTION IN 285 THE DROSOPHILA BUZZATII SPECIES COMPLEX: 286 IS INVERSION LENGTH A SELECTED TRAIT? 287 Evolution, 51(4), 1149-1155. 288 doi:https://doi.org/10.1111/j.1558-5646.1997.tb03962.x 289 Cain, A. J., & Sheppard, P. M. (1950). Selection in the 290 polymorphic land snail Cepæa nemoralis. Heredity, 4(3), 291 275-294. doi:10.1038/hdy.1950.22 292 Cain, A. J., & Sheppard, P. M. (1952). The effects of natural 293 selection on body colour in the land snail Cepaea 294 nemoralis. *Heredity*, 6(2), 217-231. 295 doi:10.1038/hdy.1952.22 296 Cain, A. J., & Sheppard, P. M. (1954). NATURAL 297 SELECTION IN CEPAEA. Genetics, 39(1), 89-116. 298 Cameron, R., & Cook, L. (2012). CORRELATED 299 PHENOTYPIC RESPONSES TO HABITAT 300 DIFFERENCE IN CEPAEA NEMORALIS (L.). FOLIA 301 MALACOLOGICA, 20(4), 255-263. 302 Carvunis, A.-R., Rolland, T., Wapinski, I., Calderwood, M. 303 A., Yildirim, M. A., Simonis, N., ... Vidal, M. (2012). 304 Proto-genes and de novo gene birth. *Nature*, 487(7407), 305 370-374. doi:10.1038/nature11184 306 Charlesworth, B., & Barton, N. H. (2018). The Spread of an 307 Inversion with Migration and Selection. Genetics, 308 208(1), 377-382. 309 Cheng, C. D., Tan, J. C., Hahn, M. W., & Besansky, N. J. 310 (2018). Systems genetic analysis of inversion 311 polymorphisms in the malaria mosquito Anopheles 312 gambiae. Proc Natl Acad Sci USA, 115(30), E7005-313
- E7014. doi:10.1073/pnas.1806760115 Cheng, C. D., White, B. J., Kamdem, C., Mockaitis, K., 315 Costantini, C., Hahn, M. W., & Besansky, N. J. (2012). 316

314

317	Ecological Genomics of Anopheles gambiae Along a
318	Latitudinal Cline: A Population-Resequencing Approach.
319	Genetics, 190(4), 1417-1432.
320	doi:10.1534/genetics.111.137794
321	Chesley, P., & Dunn, L. C. (1936). THE INHERITANCE OF
322	TAILLESSNESS (ANURY) IN THE HOUSE MOUSE.
323	Genetics, 21(5), 525-536.
324	Chouteau, M., Llaurens, V., Piron-Prunier, F., & Joron, M.
325	(2017). Polymorphism at a mimicry supergene
326	maintained by opposing frequency-dependent selection
327	pressures. Proc Natl Acad Sci USA, 114(31), 8325-
328	8329. doi:10.1073/pnas.1702482114
329	Clarke, C. A., & Sheppard, F. R. S. (1972). The genetics of
330	the mimetic butterfly Papilio polytes L. Philosophical
331	Transactions of the Royal Society of London. B,
332	Biological Sciences, 263(855), 431-458.
333	doi:10.1098/rstb.1972.0006
334	Colosimo, P. F., Peichel, C. L., Nereng, K., Blackman, B. K.,
335	Shapiro, M. D., Schluter, D., & Kingsley, D. M. (2004).
336	The Genetic Architecture of Parallel Armor Plate
337	Reduction in Threespine Sticklebacks. Plos Biology,
338	2(5), e109. doi:10.1371/journal.pbio.0020109
339	Coluzzi, M., Sabatini, A., della Torre, A., Di Deco, M. A., &
340	Petrarca, V. (2002). A polytene chromosome analysis of
341	the Anopheles gambiae species complex. Science,
342	298(5597), 1415-1418. doi:10.1126/science.1077769
343	Coluzzi, M., Sabatini, A., Petrarca, V., & Di Deco, M. A.
344	(1979). Chromosomal differentiation and adaptation to
345	humn environments in the Anopheles gambiae complex.
346	Royal Society of Tropical Medicine and Hygiene, 73(5),
347	483-497.
348	Comeault, Aaron A., Flaxman, Samuel M., Riesch, R.,
349	Curran, E., Soria-Carrasco, V., Gompert, Z., Nosil, P.

350	(2015). Selection on a Genetic Polymorphism
351	Counteracts Ecological Speciation in a Stick Insect.
352	Current Biology, 25(15), 1975-1981.
353	doi:10.1016/j.cub.2015.05.058
354	Corbett-Detig, R. B. (2016). Selection on Inversion
355	Breakpoints Favors Proximity to Pairing Sensitive Sites
356	in Drosophila melanogaster. <i>Genetics</i> , 204(1), 259-265.
357	doi:10.1534/genetics.116.190389
358	Corbett-Detig, R. B., Cardeno, C., & Langley, C. H. (2012).
359	Sequence-Based Detection and Breakpoint Assembly of
360	Polymorphic Inversions. Genetics, 192(1), 131-137.
361	doi:10.1534/genetics.112.141622
362	Corbett-Detig, R. B., & Hartl, D. L. (2012). Population
363	Genomics of Inversion Polymorphisms in Drosophila
364	melanogaster. PLoS Genet, 8(12), e1003056.
365	doi:10.1371/journal.pgen.1003056
366	Coughlan, J. M., & Willis, J. H. (2018). Dissecting the role of
367	a large chromosomal inversion in life history divergence
368	throughout the Mimulus guttatus species complex. Mol
369	Ecol, 28(6), 1343-1357. doi:doi:10.1111/mec.14804
370	Darlington, C. D., & Mather, K. (1949). The elements of
371	Genetics. London: George Allen & Unwind LTD.
372	Darwin, C. (1862). On the Two Forms, or Dimorphic
373	Condition, in the Species of Primula, and on their
374	remarkable Sexual Relations. Journal of the Proceedings
375	of the Linnean Society of London. Botany, 6(22), 77-96.
376	doi:10.1111/j.1095-8312.1862.tb01218.x
377	de Jong, S., Chepelev, I., Janson, E., Strengman, E., van den
378	Berg, L. H., Veldink, J. H., & Ophoff, R. A. (2012).
379	Common inversion polymorphism at 17q21.31 affects
380	expression of multiple genes in tissue-specific manner.
381	BMC Genomics, 13(458). doi:10.1186/1471-2164-13-
382	458

De Nettancourt, D. (2001). *Incompatibility and incongruity in* 383 wild and cultivated plants. Berlin: Springer-Verlag. 384 Dobzhansky, T. G. (1947). Adaptive Changes Induced by 385 Natural Selection in Wild Populations of Drosophila. 386 Evolution, 1(1/2), 1-16. doi:10.2307/2405399 387 Dobzhansky, T. G. (1970). Genetics of the Evolutionary 388 Process: Columbia University Press. 389 Durand, E., Chantreau, M., Le Veve, A., Stetsenko, R., Dubin, 390 M., Genete, M., . . . Castric, V. (2020). Evolution of self-391 incompatibility in the Brassicaceae: Lessons from a 392 textbook example of natural selection. Evolutionary 393 Applications, 13(6), 1279-1297. 394 doi:https://doi.org/10.1111/eva.12933 395 Eid, J., Fehr, A., Gray, J., Luong, K., Lyle, J., Otto, G., . . . 396 Turner, S. (2009). Real-Time DNA Sequencing from 397 Single Polymerase Molecules. Science, 323(5910), 133-398 138. doi:10.1126/science.1162986 399 Entani, T., Iwano, M., Shiba, H., Che, F.-S., Isogai, A., & 400 Takayama, S. (2003). Comparative analysis of the self-401 incompatibility (S-) locus region of Prunus mume: 402 identification of a pollen-expressed F-box gene with 403 allelic diversity. Genes to Cells, 8(3), 203-213. 404 doi:https://doi.org/10.1046/j.1365-2443.2003.00626.x 405 Entani, T., Iwano, M., Shiba, H., Takayama, S., Fukui, K., & 406 Isogai, A. (1999). Centromeric localization of an S-407 RNase gene in Petunia hybrida Vilm. Theoretical and 408 Applied Genetics, 99(3), 391-397. 409 doi:10.1007/s001220051249 410 Faria, R., Johannesson, K., Butlin, R. K., & Westram, A. M. 411 (2019). Evolving Inversions. Trends Ecol Evol, 34(3), 412 239-248. doi:10.1016/j.tree.2018.12.005 413 Feder, J. L., Gejji, R., Powell, T. H. Q., & Nosil, P. (2011). 414 ADAPTIVE CHROMOSOMAL DIVERGENCE 415

416	DRIVEN BY MIXED GEOGRAPHIC MODE OF
417	EVOLUTION. Evolution, 65(8), 2157-2170.
418	doi:10.1111/j.1558-5646.2011.01321.x
419	Fisher, R. A. (1930). The Genetical Theory of Natural
420	Selection: Oxford.
421	Ford, E. B. (1965). Genetic Polymorphism. London: Studies
422	Faber & Faber.
423	Ford, E. B. (1971). <i>Ecological Genomics</i> (3rd ed.). London:
424	Chapman and Hall LTD.
425	Fouet, C., Gray, E., Besansky, N. J., & Costantini, C. (2012).
426	Adaptation to Aridity in the Malaria Mosquito
427	Anopheles gambiae: Chromosomal Inversion
428	Polymorphism and Body Size Influence Resistance to
429	Desiccation. PLoS One, 7(4), e34841.
430	doi:10.1371/journal.pone.0034841
431	Ganetzky, B. (1977). On the components of segregation
432	distortion in Drosophila melanogaster. Genetics, 86(2 Pt.
433	1), 321-355.
434	Gonzalez, D. R., Aramendia, A. C., & Davison, A. (2019).
435	Recombination within the Cepaea nemoralis supergene is
436	confounded by incomplete penetrance and epistasis.
437	Heredity, 123(2), 153-161. doi:10.1038/s41437-019-
438	0190-6
439	Goubet, P. M., Bergès, H., Bellec, A., Prat, E., Helmstetter,
440	N., Mangenot, S., Castric, V. (2012). Contrasted
441	Patterns of Molecular Evolution in Dominant and
442	Recessive Self-Incompatibility Haplotypes in
443	Arabidopsis. <i>PLoS Genet</i> , 8(3), e1002495.
444	doi:10.1371/journal.pgen.1002495
445	Gray, E. M., Rocca, K. A. C., Costantini, C., & Besansky, N.
446	J. (2009). Inversion 2La is associated with enhanced
447	desiccation resistance in Anopheles gambiae. <i>Malaria</i>
448	Journal, 8. doi:10.1186/1475-2875-8-215

449	Guerrero, R. F., Rousset, F., & Kirkpatrick, M. (2012).
450	Coalescent patterns for chromosomal inversions in
451	divergent populations. Philosophical Transactions of the
452	Royal Society B-Biological Sciences, 367(1587), 430-
453	438.
454	Guillén, Y., & Ruiz, A. (2012). Gene alterations at Drosophila
455	inversion breakpoints provide prima facie evidence for
456	natural selection as an explanation for rapid
457	chromosomal evolution. BMC Genomics, 13(1), 53.
458	doi:10.1186/1471-2164-13-53
459	Hammond, T. M., Rehard, D. G., Xiao, H., & Shiu, P. K. T.
460	(2012). Molecular dissection of Neurospora Spore killer
461	meiotic drive elements. Proceedings of the National
462	Academy of Sciences, 109(30), 12093-12098.
463	doi:10.1073/pnas.1203267109
464	Hartl, D. L. (1974). GENETIC DISSECTION OF
465	SEGREGATION DISTORTION. I. SUICIDE
466	COMBINATIONS OF SD GENES. Genetics, 76(3),
467	477-486.
468	Herrmann, B. G., & Bauer, H. (2012). The mouse t-haplotype:
469	a selfish chromosome – genetics, molecular mechanism,
470	and evolution. In J. Piálek, M. Macholán, P. Munclinger,
471	& S. J. E. Baird (Eds.), Evolution of the House Mouse
472	(pp. 297-314). Cambridge: Cambridge University Press.
473	Hiraizumi, Y., & Crow, J. F. (1960). HETEROZYGOUS
474	EFFECTS ON VIABILITY, FERTILITY, RATE OF
475	DEVELOPMENT, AND LONGEVITY OF
476	DROSOPHILA CHROMOSOMES THAT ARE
477	LETHAL WHEN HOMOZYGOUS. Genetics, 45(8),
478	1071-1083.
479	Hoffmann, A. A., & Rieseberg, L. H. (2008). Revisiting the
480	Impact of Inversions in Evolution: From Population
481	Genetic Markers to Drivers of Adaptive Shifts and

```
Speciation? Annual Review of Ecology, Evolution, and
482
         Systematics, 39(1), 21-42.
483
          doi:10.1146/annurev.ecolsys.39.110707.173532
484
    Hopkins, D. P., Tyukmaeva, V. I., Gompert, Z., Feder, J. L.,
485
          & Nosil, P. (2020). Functional genomics offers new tests
486
          of speciation hypotheses. Trends Ecol Evol, 35(11), 968-
487
         971.
488
    Huang, Y.-C., Dang, V. D., Chang, N.-C., & Wang, J. (2018).
489
          Multiple large inversions and breakpoint rewiring of
490
          gene expression in the evolution of the fire ant social
491
          supergene. Proceedings of the Royal Society B:
492
         Biological Sciences, 285(1878), 20180221.
493
          doi:10.1098/rspb.2018.0221
494
    Huber, B., Whibley, A., Poul, Y. L., Navarro, N., Martin, A.,
495
         Baxter, S., . . . Joron, M. (2015). Conservatism and
496
         novelty in the genetic architecture of adaptation in
497
          Heliconius butterflies. Heredity, 114(5), 515-524.
498
         doi:10.1038/hdy.2015.22
499
    Jain, M., Olsen, H. E., Paten, B., & Akeson, M. (2016). The
500
          Oxford Nanopore MinION: delivery of nanopore
501
          sequencing to the genomics community. Genome
502
         Biology, 17(1), 239. doi:10.1186/s13059-016-1103-0
503
    Jay, P., Whibley, A., Frézal, L., Rodríguez de Cara, M. Á.,
504
         Nowell, R. W., Mallet, J., . . . Joron, M. (2018).
505
          Supergene Evolution Triggered by the Introgression of a
506
          Chromosomal Inversion. Current Biology, 28(11), 1839-
507
          1845.e1833. doi:10.1016/j.cub.2018.04.072
508
    Jones, F. C., Grabherr, M. G., Chan, Y. F., Russell, P.,
509
          Mauceli, E., Johnson, J., . . . Kingsley, D. M. (2012). The
510
          genomic basis of adaptive evolution in threespine
511
          sticklebacks. Nature, 484, 55-61.
512
          doi:10.1038/nature10944
513
```

514	https://www.nature.com/articles/nature10944#supplementary-
515	information
516	Joron, M., Frezal, L., Jones, R. T., Chamberlain, N. L., Lee, S.
517	F., Haag, C. R., ffrench-Constant, R. H. (2011).
518	Chromosomal rearrangements maintain a polymorphic
519	supergene controlling butterfly mimicry. Nature,
520	477(7363), 203-206. doi:10.1038/nature10341
521	Joron, M., Wynne, I. R., Lamas, G., & Mallet, J. (1999).
522	Variable selection and the coexistence of multiple
523	mimetic forms of the butterfly Heliconius numata.
524	Evolutionary Ecology, 13(7-8), 721-754.
525	doi:10.1023/A:1010875213123
526	Kapan, D. D. (2001). Three-butterfly system provides a field
527	test of mullerian mimicry. Nature, 409(6818), 338-340.
528	doi:10.1038/35053066
529	Kelemen, R. K., & Vicoso, B. (2018). Complex History and
530	Differentiation Patterns of the t-Haplotype, a Mouse
531	Meiotic Driver. Genetics, 208(1), 365-375.
532	doi:10.1534/genetics.117.300513
533	Kirkpatrick, M. (2010). How and Why Chromosome
534	Inversions Evolve. <i>Plos Biology</i> , 8(9), e1000501.
535	doi:10.1371/journal.pbio.1000501
536	Kirkpatrick, M., & Barton, N. (2006). Chromosome
537	Inversions, Local Adaptation and Speciation. Genetics,
538	173(1), 419-434. doi:10.1534/genetics.105.047985
539	Kirubakaran, T. G., Grove, H., Kent, M. P., Sandve, S. R.,
540	Baranski, M., Nome, T., Andersen, O. (2016). Two
541	adjacent inversions maintain genomic differentiation
542	between migratory and stationary ecotypes of Atlantic
543	cod. <i>Mol Ecol</i> , 25(10), 2130-2143.
544	doi:10.1111/mec.13592
545	Korunes, K. L., & Noor, M. A. F. (2017). Gene conversion
546	and linkage: effects on genome evolution and speciation.

547	<i>Mol Ecol</i> , 26(1), 351-364.
548	doi:https://doi.org/10.1111/mec.13736
549	Kraft, K., Geuer, S., Will, Anja J., Chan, Wing L., Paliou, C.,
550	Borschiwer, M., Andrey, G. (2015). Deletions,
551	Inversions, Duplications: Engineering of Structural
552	Variants using CRISPR/Cas in Mice. Cell Reports, 10(5),
553	833-839. doi:https://doi.org/10.1016/j.celrep.2015.01.016
554	Kunte, K., Zhang, W., Tenger-Trolander, A., Palmer, D. H.,
555	Martin, A., Reed, R. D., Kronforst, M. R. (2014).
556	doublesex is a mimicry supergene. Nature, 507(7491),
557	229-232. doi:10.1038/nature13112
558	Kupper, C., Stocks, M., Risse, J. E., dos Remedios, N.,
559	Farrell, L. L., Mcrae, S. B., Burke, T. (2016). A
560	supergene determines highly divergent male reproductive
561	morphs in the ruff. <i>Nature Genetics</i> , 48(1), 79-83.
562	doi:10.1038/ng.3443
563	Lamichhaney, S., Fan, G. Y., Widemo, F., Gunnarsson, U.,
564	Thalmann, D. S., Hoeppner, M. P., Andersson, L.
565	(2016). Structural genomic changes underlie alternative
566	reproductive strategies in the ruff (Philomachus pugnax).
567	Nature Genetics, 48(1), 84-88. doi:10.1038/ng.3430
568	Larracuente, A. M., & Presgraves, D. C. (2012). The Selfish
569	Segregation Distorter Gene Complex of Drosophila
570	melanogaster. Genetics, 192(1), 33-53.
571	doi:10.1534/genetics.112.141390
572	Lavington, E., & Kern, A. D. (2017). The Effect of Common
573	Inversion Polymorphisms In(2L) and In(3R)Mo on
574	Patterns of Transcriptional Variation in Drosophila
575	melanogaster. G3: Genes Genomes Genetics, 7(11),
576	3659-3668. doi:10.1534/g3.117.1133
577	Lee, C. R., Wang, B. S., Mojica, J. P., Mandakova, T., Prasad,
578	K. Y. S. K., Goicoechea, J. L., Mitchell-Olds, T.
579	(2017). Young inversion with multiple linked QTLs

580	under selection in a hybrid zone. <i>Nature Ecology &amp;</i>
581	Evolution, 1(5). doi:10.1038/s41559-017-0119
582	Lemopoulos, A., Uusi-Heikkilä, S., Huusko, A., Vasemägi,
583	A., & Vainikka, A. (2018). Comparison of Migratory and
584	Resident Populations of Brown Trout Reveals Candidate
585	Genes for Migration Tendency. Genome Biology and
586	Evolution, 10(6), 1493-1503. doi:10.1093/gbe/evy102
587	Li, J., Cocker, J. M., Wright, J., Webster, M. A., McMullan,
588	M., Dyer, S., Gilmartin, P. M. (2016). Genetic
589	architecture and evolution of the S locus supergene in
590	Primula vulgaris. <i>Nature Plants, 2</i> (12), 16188.
591	doi:10.1038/nplants.2016.188
592	Liehr, T., Weise, A., Mrasek, K., Ziegler, M., Padutsch, N.,
593	Wilhelm, K., & Al-Rikabi, A. (2019). Recombinant
594	Chromosomes Resulting From Parental Pericentric
595	Inversions—Two New Cases and a Review of the
596	Literature. Frontiers in Genetics, 10(1165).
597	doi:10.3389/fgene.2019.01165
598	Ling, A., & Cordaux, R. (2010). Insertion Sequence
599	Inversions Mediated by Ectopic Recombination between
600	Terminal Inverted Repeats. <i>PLoS One</i> , 5(12), e15654.
601	doi:10.1371/journal.pone.0015654
602	Lowry, D. B., & Willis, J. H. (2010). A Widespread
603	Chromosomal Inversion Polymorphism Contributes to a
604	Major Life-History Transition, Local Adaptation, and
605	Reproductive Isolation. <i>Plos Biology</i> , 8(9), e1000500.
606	Lyon, M. F. (1984). Transmission ratio distortion in mouse t-
607	haplotypes is due to multiple distorter genes acting on a
608	responder locus. Cell, 37(2), 621-628. doi:10.1016/0092-
609	8674(84)90393-3
610	Mallet, J., & Barton, N. H. (1989). Strong Natural-Selection
611	in a Warning-Color Hybrid Zone. Evolution, 43(2), 421-
612	431. doi:Doi 10.2307/2409217

613	Mather, K. (1950). The Genetical Architecture of Heterostyly
614	in Primula sinensis. Evolution, 4(4), 340-352.
615	doi:10.2307/2405601
616	Mather, K. (1955). Polymorphism as an Outcome of
617	Disruptive Selection. Evolution, 9(1), 52-61.
618	doi:10.2307/2405357
619	Merrill, R. M., Wallbank, R. W. R., Bull, V., Salazar, P. C.
620	A., Mallet, J., Stevens, M., & Jiggins, C. D. (2012).
621	Disruptive ecological selection on a mating cue.
622	Proceedings of the Royal Society B-Biological Sciences,
623	279(1749), 4907-4913. doi:10.1098/rspb.2012.1968
624	Navarro, A., & Barton, N. H. (2003). Chromosomal
625	Speciation and Molecular DivergenceAccelerated
626	Evolution in Rearranged Chromosomes. Science,
627	300(5617), 321-324. doi:10.1126/science.1080600
628	Nichols, K. M., Edo, A. F., Wheeler, P. A., & Thorgaard, G.
629	H. (2008). The genetic basis of smoltification-related
630	traits in Oncorhynchus mykiss. Genetics, 179(3), 1559-
631	1575. doi:10.1534/genetics.107.084251
632	Nishikawa, H., Iijima, T., Kajitani, R., Yamaguchi, J., Ando,
633	T., Suzuki, Y., Fujiwara, H. (2015). A genetic
634	mechanism for female-limited Batesian mimicry in
635	Papilio butterfly. <i>Nature Genetics</i> , 47(4), 405-409.
636	doi:10.1038/ng.3241
637	Nosil, P., Villoutreix, R., de Carvalho, C. F., Farkas, T. E.,
638	Soria-Carrasco, V., Feder, J. L., Gompert, Z. (2018).
639	Natural selection and the predictability of evolution in
640	Timema stick insects. <i>Science</i> , <i>359</i> (6377), 765-770.
641	doi:10.1126/science.aap9125
642	O'Malley, K. G., Sakamoto, T., Danzmann, R. G., &
643	Ferguson, M. M. (2003). Quantitative Trait Loci for
644	Spawning Date and Body Weight in Rainbow Trout:
645	Testing for Conserved Effects Across Ancestrally

646	Duplicated Chromosomes. <i>Journal of Heredity</i> , 94(4),
647	273-284. doi:10.1093/jhered/esg067
648	Pearse, D. E., Miller, M. R., Abadia-Cardoso, A., & Garza, J.
649	C. (2014). Rapid parallel evolution of standing variation
650	in a single, complex, genomic region is associated with
651	life history in steelhead/rainbow trout. Proceedings of the
652	Royal Society B-Biological Sciences, 281(1783).
653	doi:10.1098/rspb.2014.0012
654	Piper, J., & Charlesworth, B. (2008). The evolution of distyly
655	in Primula vulgaris. Biological Journal of the Linnean
656	Society, 29(2), 123-137. doi:10.1111/j.1095-
657	8312.1986.tb01827.x
658	Puig, M., Casillas, S., Villatoro, S., & Caceres, M. (2015).
659	Human inversions and their functional consequences.
660	Briefings in Functional Genomics, 14(5), 369-379.
661	doi:10.1093/bfgp/elv020
662	Ranz, J. M., Maurin, D., Chan, Y. S., von Grotthuss, M.,
663	Hillier, L. W., Roote, J., Bergman, C. M. (2007).
664	Principles of Genome Evolution in the Drosophila
665	melanogaster Species Group. <i>Plos Biology</i> , 5(6), e152.
666	doi:10.1371/journal.pbio.0050152
667	Rhoades, N. A., Harvey, A. M., Samarajeewa, D. A.,
668	Svedberg, J., Yusifov, A., Abusharekh, A.,
669	Hammond, T. M. (2019). Identification of rfk-1, a
670	Meiotic Driver Undergoing RNA Editing in Neurospora.
671	Genetics, 212(1), 93-110.
672	doi:10.1534/genetics.119.302122
673	Robison, B. D., Wheeler, P. A., Sundin, K., Sikka, P., &
674	Thorgaard, G. H. (2001). Composite Interval Mapping
675	Reveals a Major Locus Influencing Embryonic
676	Development Rate in Rainbow Trout (Oncorhynchus
677	mykiss). Journal of Heredity, 92(1), 16-22.
678	doi:10.1093/jhered/92.1.16

679	Rocca, K. A. C., Gray, E. M., Costantini, C., & Besansky, N.
680	J. (2009). 2La chromosomal inversion enhances thermal
681	tolerance of Anopheles gambiae larvae. Malaria Journal,
682	8. doi:10.1186/1475-2875-8-147
683	Ross, K. G., & Keller, L. (1995). Ecology and Evolution of
684	Social-Organization - Insights from Fire Ants and Other
685	Highly Eusocial Insects. Annual Review of Ecology and
686	<i>Systematics</i> , 26, 631-656. doi:DOI
687	10.1146/annurev.es.26.110195.003215
688	Said, I., Byrne, A., Serrano, V., Cardeno, C., Vollmers, C., &
689	Corbett-Detig, R. (2018). Linked genetic variation and
690	not genome structure causes widespread differential
691	expression associated with chromosomal inversions.
692	Proceedings of the National Academy of Sciences,
693	115(21), 5492-5497. doi:10.1073/pnas.1721275115
694	Sandler, L., & Hiraizumi, Y. (1960). MEIOTIC DRIVE IN
695	NATURAL POPULATIONS OF DROSOPHILA
696	MELANOGASTER. IV. INSTABILITY AT THE
697	SEGREGATION-DISTORTER LOCUS. Genetics,
698	<i>45</i> (9), 1269-1287.
699	Sandoval, C. P. (1994). Differential visual predation on
700	morphs of Timema cristinae (Phasmatodeae:Timemidae)
701	and its consequences for host range. Biological Journal
702	of the Linnean Society, 52(4), 341-356.
703	doi:doi:10.1111/j.1095-8312.1994.tb00996.x
704	Sandoval, C. P., & Nosil, P. (2005). Counteracting selective
705	regimes and host preference evolution in ecotypes of two
706	species of walking-sticks. <i>Evolution</i> , 59(11), 2405-2413.
707	Schopfer, C. R., Nasrallah, M. E., & Nasrallah, J. B. (1999).
708	The Male Determinant of Self-Incompatibility in
709	Brassica. <i>Science</i> , 286(5445), 1697-1700.
710	doi:10.1126/science.286.5445.1697

Schwander, T., Libbrecht, R., & Keller, L. (2014). Supergenes 711 and Complex Phenotypes. Current Biology, 24(7), R288-712 R294. doi:10.1016/j.cub.2014.01.056 713 Sharakhov, I. V., White, B. J., Sharakhova, M. V., Kayondo, 714 J., Lobo, N. F., Santolamazza, F., . . . Besansky, N. J. 715 (2006). Breakpoint structure reveals the unique origin of 716 an interspecific chromosomal inversion 2La in the 717 Anopheles gambiae complex. Proceedings of the 718 National Academy of Sciences, 103(16), 6258-6262. 719 doi:10.1073/pnas.0509683103 720 Silva, N. F., Stone, S. L., Christie, L. N., Sulaman, W., 721 Nazarian, K. A. P., Burnett, L. A., . . . Goring, D. R. 722 (2001). Expression of the S receptor kinase in self-723 compatible Brassica napus cv. Westar leads to the allele-724 specific rejection of self-incompatible Brassica napus 725 pollen. Molecular Genetics and Genomics, 265(3), 552-726 559. doi:10.1007/s004380100446 727 Sinclair-Waters, M., Bradbury, I. R., Morris, C. J., Lien, S., 728 Kent, M. P., & Bentzen, P. (2018). Ancient chromosomal 729 rearrangement associated with local adaptation of a 730 postglacially colonized population of Atlantic Cod in the 731 northwest Atlantic. Mol Ecol, 27(2), 339-351. 732 doi:10.1111/mec.14442 733 Stefansson, H., Helgason, A., Thorleifsson, G., 734 Steinthorsdottir, V., Masson, G., Barnard, J., . . . 735 Stefansson, K. (2005). A common inversion under 736 selection in Europeans. Nature Genetics, 37(2), 129-137. 737 doi:10.1038/ng1508 738 Stevison, L. S., Hoehn, K. B., & Noor, M. A. F. (2011). 739 Effects of Inversions on Within- and Between-Species 740 Recombination and Divergence. Genome Biology and 741 Evolution, 3, 830-841. doi:10.1093/gbe/evr081 742

- Stewart, N. B., & Rogers, R. L. (2019). Chromosomal
- rearrangements as a source of new gene formation in
- Drosophila yakuba. *PLoS Genet, 15*(9), e1008314.
- doi:10.1371/journal.pgen.1008314
- 547 Stump, A. D., Pombi, M., Goeddel, L., Ribeiro, J. M. C.,
- Wilder, J. A., Torre, A. D., & Besansky, N. J. (2007).
- Genetic exchange in 2La inversion heterokaryotypes of
- Anopheles gambiae. *Insect Molecular Biology*, 16(6),
- 703-709. doi:https://doi.org/10.1111/j.1365-
- 752 2583.2007.00764.x
- Sturtevant, A. H. (1921). A Case of Rearrangement of Genes
- in Drosophila. *Proc Natl Acad Sci U S A*, 7(8), 235-237.
- doi:10.1073/pnas.7.8.235
- Svedberg, J., Hosseini, S., Chen, J., Vogan, A. A., Mozgova,
- I., Hennig, L., . . . Johannesson, H. (2018). Convergent
- evolution of complex genomic rearrangements in two
- fungal meiotic drive elements. Nature Communications,
- 9(1), 4242. doi:10.1038/s41467-018-06562-x
- Takasaki, T., Hatakeyama, K., Suzuki, G., Watanabe, M.,
- Isogai, A., & Hinata, K. (2000). The S receptor kinase
- determines self-incompatibility in Brassica stigma.
- Nature, 403(6772), 913-916. doi:10.1038/35002628
- 765 Takayama, S., & Isogai, A. (2005). SELF-
- INCOMPATIBILITY IN PLANTS. Annual Review of
- 767 Plant Biology, 56(1), 467-489.
- doi:10.1146/annurev.arplant.56.032604.144249
- Takayama, S., Isogai, A., Tsukamoto, C., Ueda, Y., Hinata,
- K., Okazaki, K., & Suzuki, A. (1987). Sequences of S-
- glycoproteins, products of the Brassica campestris self-
- incompatibility locus. *Nature*, *326*(6108), 102-105.
- doi:10.1038/326102a0
- Takayama, S., Shiba, H., Iwano, M., Shimosato, H., Che, F.-
- S., Kai, N., ... Isogai, A. (2000). The pollen determinant

of self-incompatibility in Brassica campestris. 776 Proceedings of the National Academy of Sciences, 97(4), 777 1920-1925. doi:10.1073/pnas.040556397 778 Thomas, J. W., Cáceres, M., Lowman, J. J., Morehouse, C. B., 779 Short, M. E., Baldwin, E. L., . . . Martin, C. L. (2008). 780 The Chromosomal Polymorphism Linked to Variation in 781 Social Behavior in the White-Throated Sparrow 782 (Zonotrichia albicollis) Is a Complex Rearrangement and 783 Suppressor of Recombination. Genetics, 179(3), 1455-784 1468. doi:10.1534/genetics.108.088229 785 Thorneycroft, H. B. (1966). Chromosomal Polymorphism in 786 the White-Throated Sparrow, Zonotrichia albicollis 787 (Gmelin). Science, 154(3756), 1571-1572. 788 Throneycroft, H. B. (1975). A Cytogenetic Study of the 789 White-Throated Sparrow, Zonotrichia albicollis 790 (Gmelin). Evolution, 29(4), 611-621. 791 doi:10.2307/2407072 792 Turner, B. C., & Perkins, D. D. (1979). SPORE KILLER, A 793 CHROMOSOMAL FACTOR IN NEUROSPORA 794 THAT KILLS MEIOTIC PRODUCTS NOT 795 CONTAINING IT. *Genetics*, 93(3), 587-606. 796 Tuttle, E. M., Bergland, A. O., Korody, M. L., Brewer, M. S., 797 Newhouse, D. J., Minx, P., . . . Balakrishnan, C. N. 798 (2016). Divergence and Functional Degradation of a Sex 799 Chromosome-like Supergene. Current Biology, 26(3), 800 344-350. 801 Ushijima, K., Sassa, H., Dandekar, A. M., Gradziel, T. M., 802 Tao, R., & Hirano, H. (2003). Structural and 803 Transcriptional Analysis of the Self-Incompatibility 804 Locus of Almond: Identification of a Pollen-Expressed 805 F-Box Gene with Haplotype-Specific Polymorphism. 806 The Plant Cell, 15(3), 771-781. doi:10.1105/tpc.009290 807

808	Van Belleghem, S. M., Rastas, P., Papanicolaou, A., Martin,
809	S. H., Arias, C. F., Supple, M. A., Papa, R. (2017).
810	Complex modular architecture around a simple toolkit of
811	wing pattern genes. Nature Ecology & Evolution, 1,
812	0052. doi:10.1038/s41559-016-0052
813	http://www.nature.com/articles/s41559-016-
814	0052#supplementary-information
815	Villoutreix, R., de Carvalho, C. F., Soria-Carrasco, V.,
816	Lindtke, D., De-la-Mora, M., Muschick, M., Nosil, P
817	(2020). Large-scale mutation in the evolution of a gene
818	complex for cryptic coloration. Science, 369(6502), 460-
819	466. doi:10.1126/science.aaz4351
820	Wang, J., Wurm, Y., Nipitwattanaphon, M., Riba-Grognuz,
821	O., Huang, Y. C., Shoemaker, D., & Keller, L. (2013). A
822	Y-like social chromosome causes alternative colony
823	organization in fire ants. <i>Nature</i> , 493(7434), 664-668.
824	doi:10.1038/nature11832
825	Wellenreuther, M., & Bernatchez, L. (2018). Eco-
826	Evolutionary Genomics of Chromosomal Inversions.
827	Trends Ecol Evol, 33(6), 427-440.
828	doi:10.1016/j.tree.2018.04.002
829	Yan, Z., Martin, S. H., Gotzek, D., Arsenault, S. V., Duchen,
830	P., Helleu, Q., Keller, L. (2020). Evolution of a
831	supergene that regulates a trans-species social
832	polymorphism. Nature Ecology & Evolution, 4, 240-249
833	
834	