

RESEARCH ARTICLE

Saltwater and nutrient legacies reduce net ecosystem carbon storage despite freshwater restoration: insights from experimental wetlands

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Net ecosystem carbon balance is a comprehensive assessment of ecosystem function that can test restoration effectiveness. Coastal peatlands are globally important carbon sinks that are vulnerable to carbon loss with saltwater intrusion. It is uncertain how wetland carbon stocks and fluxes change during freshwater restoration following exposure to saltwater and elevated nutrients. We restored freshwater to sawgrass (*Cladium jamaicense*) peat monoliths from freshwater marshes of the Everglades (Florida, U.S.A.) that had previously been exposed to elevated salinity (approximately 9 ppt) and phosphorus (P) loading (1 g P m⁻² year⁻¹) in wetland mesocosms. We quantified changes in water and soil physicochemistry, plant and soil carbon and nutrient standing stocks, and net ecosystem productivity during restoration. Added freshwater immediately reduced porewater salinity from >8 to approximately 2 ppt, but elevated porewater dissolved organic carbon persisted. Above- and below-ground biomass, leaf P concentrations, and instantaneous rates of gross ecosystem productivity (GEP) and ecosystem respiration (ER) remained elevated from prior added P. Modeled monthly GEP and ER were higher in marshes with saltwater and P legacies, resulting in negative net ecosystem productivities that were up to 12× lower than controls. Leaf litter breakdown rates and litter P concentrations were 2× higher in marshes with legacies of added saltwater and P. Legacies of saltwater and P on carbon loss persisted despite freshwater restoration, but recovery was greatest for freshwater marshes exposed to saltwater alone. Our results suggest that restoration in nutrient-limited freshwater wetlands exposed to saltwater intrusion and nutrient enrichment is a slow process.

Key words: biogeochemistry, carbon, Florida Coastal Everglades, Long Term Ecological Research, phosphorus, saltwater intrusion, sea-level rise, soil elevation

Implications for Practice

- Whether or not freshwater restoration can reverse impacts of saltwater intrusion in coastal ecosystems is a fundamental uncertainty.
- Legacies of elevated saltwater and nutrients can persist and impede ecosystem structure and net ecosystem carbon storage.
- Sustained freshwater restoration may be an essential management strategy needed to protect vulnerable coastal ecosystems exposed to sea-level rise and saltwater intrusion.

et al. 2020). Sea-level rise is a global-scale disturbance that is rapidly increasing saltwater intrusion in coastal wetland ecosystems (Herbert et al. 2015; Tully et al. 2019). Low-lying coastal ecosystems become particularly vulnerable to saltwater intrusion, resulting in a shift in chemical and hydrological regimes (Herbert et al. 2015; Osland et al. 2016; Dessu et al. 2018). Coastal wetland ecosystems are important global carbon sinks that are directly threatened by climate change, land-use change,

Author contributions: DL, JK, MK, MR, SR contributed ideas, data collection, and writing; DL analyzed data.

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doi: 10.1111/rec.13524

Supporting information at:

<http://onlinelibrary.wiley.com/doi/10.1111/rec.13524/supplinfo>

and interactions of both with saltwater intrusion from sea-level rise (Kirwan & Megonigal 2013; Tully et al. 2019). The fate of coastal ecosystems as sea-level rise and saltwater intrusion increase will be determined by the ability of these ecosystems to store carbon (Kominoski et al. 2018). Forecasting whether coastal ecosystems continue to develop, transition to another vegetated state, or slowly or abruptly decline to open water, requires a mechanistic understanding of how saltwater intrusion affects carbon production, storage, and export as sea levels rise (Morris et al. 2002; Jiang et al. 2012; Chambers et al. 2019).

The relative sensitivity of marsh plants to saltwater intrusion is species-specific and dependent on antecedent conditions and interacting stressors (Flynn et al. 1995; Breithaupt et al. 2020). Recent experimental manipulations of saltwater intrusion have revealed consistent declines in plant productivity (Herbert et al. 2018; Wilson et al. 2018, 2019; Solohin et al. 2020), declines in soil elevation and carbon storage (Charles et al. 2019; Solohin et al. 2020), and increases in soil organic matter breakdown (Charles et al. 2019; Servais et al. 2020). In addition, nutrient enrichment (elevated phosphorus [P]) stimulates sawgrass (*Cladium jamaicense*) above- and belowground biomass, regardless of the presence of saltwater (Charles et al. 2019; Wilson et al. 2019). Phosphorus enrichment also increases root productivity and leaf and root litter decomposition, regardless of the presence of saltwater (Charles et al. 2019; Servais et al. 2019). Compared to the subsidy effect of P, salinity increases osmotic stress and ionic concentrations (Herbert et al. 2015; Tully et al. 2019), which causes root mortality and soil elevation loss (Charles et al. 2019) and desorption of dissolved carbon and nutrients from peat soils (Servais et al. 2019; Wilson et al. 2019). Control wetlands, without added saltwater or P, had negligible changes in soil elevation during the study period (Charles et al. 2019). Carbon losses (e.g. leached dissolved organic carbon, increased microbial, and plant respiration) exceeded carbon production and retention (i.e. gains; e.g. primary

production, root productivity, and aboveground biomass growth), resulting in consistent soil elevation declines with exposure to saltwater with and without added P (Table 1). This decrease in soil elevation (23 mm year⁻¹) is an order of magnitude higher than global (2.8 mm year⁻¹) and local (7.7 mm year⁻¹) estimates of sea-level rise (Watson et al. 2015; Dessu et al. 2018).

Declines in net ecosystem carbon storage are expected in freshwater peatlands exposed to saltwater intrusion without restored or increased freshwater. The Florida Everglades is the largest subtropical freshwater peatland in the United States that is being restored through reestablishment of historic freshwater flows (NASEM 2018; Sarker et al. 2020). More than a century of draining and building of canals has reduced the original size of the historic Everglades by half (McVoy 2011). These changes resulted in reductions of freshwater flows to the southern Everglades, making it increasingly vulnerable to saltwater intrusion (Davis & Ogden 1994). The Everglades is an extremely phosphorus-limited, upside-down estuary, whereby marine water pulses stimulate wetland productivity (Childers et al. 2006), but paradoxically freshwater and brackish marshes occupying the leading edge of saltwater intrusion are sensitive to elevated salinities (Charles et al. 2019; Wilson et al. 2019). Coupled with recent findings of persistent elevation loss in wetlands exposed to elevated salinity (Stagg et al. 2018; Charles et al. 2019; Solohin et al. 2020 and references therein), wetlands exposed to salinity and P may have strong internal feedbacks that reinforce a persistent degraded state (sensu Søndergaard et al. 2001; Suding et al. 2004). Freshwater pulses into brackish soils can decrease osmotic stress and promote methanogenesis (Chambers et al. 2013) by removing accumulated Cl⁻, HS⁻, and SO₄²⁻ (Jolly et al. 2008). Freshwater pulses decrease salinity and increase primary production in coastal swamp forests (Middleton et al. 2015). Freshwater restoration will need to maintain and enhance soil elevation gains and net ecosystem

Table 1. Summarized published results from a saltwater × phosphorus (P) manipulation study that quantified the responses of freshwater marshes to saltwater intrusion. Freshwater sawgrass (*Cladium jamaicense*) peat monoliths were exposed to freshwater (control, FW), elevated P (FWP), elevated saltwater (SW), and elevated saltwater and P (SWP) from 7 February 2015 to 12 February 2017. A dominant positive (+) or negative (-) effect is from a two-way ANOVA test. When both saltwater and P effects were found (i.e. either main or interactive effects), the stronger main effect was presented. Two-way ANOVA and Tukey's HSD post hoc comparison were used to determine a difference at a significance level of $\alpha = 0.05$. Superscripted letters represent results from Tukey's HSD test ($p < 0.05$). Tukey's HSD test was not performed on gross ecosystem productivity and ecosystem respiration. (SRP, soluble reactive phosphorus; DOC, dissolved organic carbon; ER, ecosystem respiration; GEP, gross ecosystem productivity).

Response variable	FW	FWP	SW	SWP	p	Effect	Reference
Porewater salinity (ppt)	0.46 ^a	0.44 ^a	8.3 ^b	8.0 ^b	<0.05	Salt (+)	Servais et al. (2019)
Porewater SRP ($\mu\text{mol L}^{-1}$)	0.15 ^a	0.20 ^a	0.38 ^b	0.39 ^b	<0.001	Salt (+)	Wilson et al. (2019)
Porewater DOC ($\mu\text{mol L}^{-1}$)	1862 ^a	1981 ^a	3281 ^b	3268 ^b	<0.001	Salt (+)	Wilson et al. (2019)
Porewater NH ₄ ⁺ ($\mu\text{mol L}^{-1}$)	6.6 ^a	8.8 ^a	36.5 ^b	28.6 ^b	<0.001	Salt (+)	Wilson et al. (2019)
Sawgrass aboveground biomass (g C m^{-2})	294 ^a	846 ^b	398 ^a	627 ^{ab}	<0.01	Phosphorus (+)	Wilson et al. (2019)
Sawgrass root productivity ($\text{g C m}^{-2} \text{ year}^{-1}$)	81.3 ^{ab}	179.2 ^a	49.1 ^b	126.5 ^{ab}	<0.01	Phosphorus (+)	Charles et al. (2019)
Sawgrass leaf litter breakdown (k day^{-1})	0.0014 ^a	0.002 ^b	0.0015 ^a	0.002 ^b	<.001	Phosphorus (+)	Charles et al. (2019)
Sawgrass root (0–7.5 cm) breakdown (k day^{-1})	0.0009 ^a	0.0011 ^b	0.0008 ^a	0.0012 ^b	<0.01	Phosphorus (+)	Servais et al. (2019)
Sawgrass belowground biomass (g C m^{-2})	456 ^a	599 ^b	252 ^c	30 ^c	<0.05	Salt (-)	Charles et al. (2019)
Soil elevation change (cm year^{-1})	-0.04 ^{ab}	0.88 ^a	-2.75 ^b	-2.29 ^b	<0.001	Salt (-)	Charles et al. (2019)
GEP ($\mu\text{mol C m}^{-2} \text{ s}^{-1}$)	6.6 ^a	15.8 ^b	7.4 ^a	12.3 ^b	<.001	Phosphorus (+)	Wilson et al. (2019)
ER ($\mu\text{mol C m}^{-2} \text{ s}^{-1}$)	1.4 ^a	3.2 ^b	1.4 ^a	2.7 ^b	<.001	Phosphorus (+)	Wilson et al. (2019)

carbon storage to promote development trajectories in coastal ecosystems (Nyman et al. 2006; Neubauer 2008).

Our objective was to test whether freshwater restoration, aimed at remediating the effects of saltwater and P legacies, could reestablish wetland ecosystem development trajectories that increase net ecosystem carbon storage. We used sawgrass-peat monoliths collected from freshwater wetlands that were inundated with brackish water (approximately 9 ppt) and enriched with P ($1 \text{ g P m}^{-2} \text{ year}^{-1}$) during the manipulation experiment preceding our restoration experiment. We tested the following questions: (1) Can restoring freshwater to freshwater sawgrass marshes offset legacies of elevated saltwater and P?, (2) What biogeochemical mechanisms are altered by freshwater restoration?, and (3) Can freshwater restoration enhance net ecosystem carbon storage and transform wetlands exposed to elevated saltwater and nutrients from carbon sources to carbon sinks?

Methods

Experimental Design

We used sawgrass-peat monoliths placed in polycarbonate boxes ($3,500 \text{ cm}^2$ surface area \times 53 cm height) in an outdoor mesocosm facility at Florida Bay Interagency Science Center, Key Largo, Florida (Fig. 1). In July 2014, intact plant-soil monoliths were transported from a nutrient-unenriched freshwater marsh wetland in the Everglades to the facility. The monoliths were inundated under the freshwater condition to allow acclimation for 7 months; an augmentation of seawater and P was initiated on 7 February 2015 and maintained until 12 February 2017 to quantify responses to long-term exposure to elevated saltwater and P. Details on collection site characteristics, material handling, acclimation, incubation, and manipulation are provided elsewhere (Charles et al. 2019; Servais et al. 2019; Wilson et al. 2019). Briefly, freshwater was collected

from the C-111 Canal ($25^{\circ}17'31.74''\text{N}$, $80^{\circ}27'21.59''\text{W}$). Saltwater was pumped directly from Florida Bay. Freshwater and diluted seawater (salinity: approximately 9 ppt) were added manually 2–3 times per week to keep surface soils inundated. Phosphoric acid ($0.45 \text{ mg P day}^{-1}$) was pumped directly into the P-added plots. The manipulation experiment lasted for 24 months and included a total of 24 experimental plots, including the control ($n = 6$) and three legacy treatments ($n = 6$ per treatment): (i) freshwater (the control or FW), (ii) freshwater and added P (FWP_L), (iii) added saltwater (SW_L), and (iv) added saltwater and P (SWP_L).

Immediately following the manipulation experiment, we drained, rinsed, and refilled all holding containers with freshwater. Freshwater was manually added to each container three times per week from 12 February 2017 to 19 June 2018, maintaining water depth above the soil surface at approximately 10 cm.

Surface and Porewater Physicochemistry

Physicochemical conditions were monitored ($n = 6$ replicates per legacy treatment) monthly at surface water and porewater. A porewater sipper with an aquarium air stone (4 cm long \times 1 cm diameter) was inserted to 15 cm depth near the middle of each monolith prior to the manipulation experiment. Sulfide concentration in porewater (HS^- ; $n = 3$ per legacy treatment) was measured until not detected (2 March, 3 July, and 1 August 2017). Salinity, conductivity, and temperature were surveyed with a YSI Model 600 XL (Xylem, Inc., Yellow Springs, OH, U.S.A.). Water samples were collected using a 60-mL syringe, filtered through a 0.7-μm glass fiber filter, and stored at -20°C until analysis. Water chemistry was analyzed at the Southeast Environmental Research Center Nutrient Analysis Laboratory (SERC-NAL). SERC-NAL followed internal and external quality assurance practices and is NELAC certified for non-potable water-General Chemistry under State Lab ID E76930. Filtered

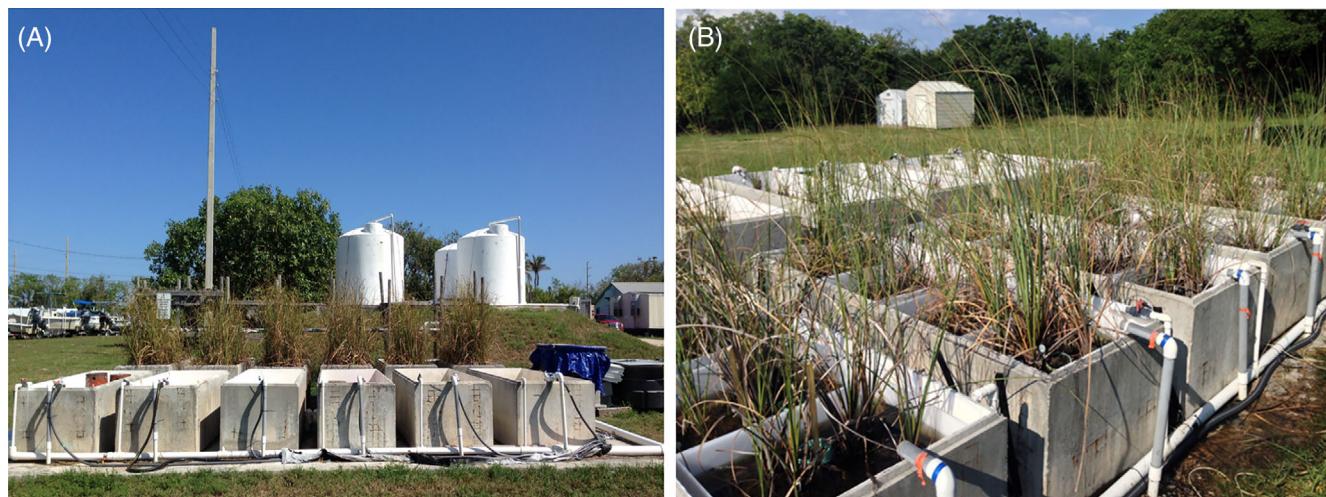


Figure 1. Experimental wetland mesocosm facility, Key Largo, Florida, U.S.A. (A) Image illustrates the overall footprint of the facility with raised water headtanks in the background and concrete basins with and without wetland soils and sawgrass (*Cladium jamaicense*) plants in the foreground. (B) Image illustrates how experimental sawgrass (*Cladium jamaicense*) wetland mesocosms were in closed-bottom, self-contained polycarbonate bins that enabled replication within and among treatments.

samples were analyzed for dissolved organic carbon (DOC), ammonium (NH_4^+), soluble reactive phosphorus (SRP), and HS^- . The concentration of SRP was analyzed on an Alpkem RFA 300 autoanalyzer (OI Analytical, College Station, TX, U.S.A.), and DOC (and total organic carbon [TOC]) was analyzed with a Shimadzu 5,000 TOC Analyzer (Shimadzu Corporation, Columbia, MD, U.S.A.). Porewater HS^- concentrations were quantified following standard methods (McKee et al. 1988). The concentration of total phosphorus (TP) was analyzed using the high-temperature persulfate digestion method (Solórzano & Sharp 1980).

Organic Carbon Flux and Elemental Concentrations

Sawgrass aboveground biomass was measured in February, May, and July of 2017 and February, April, and June of 2018 using a non-destructive allometric method (Daoust & Childers 1998). The decomposition of sawgrass litter was calculated from mass loss and expressed as a first-order decay constant (Davis 1991). Standing dead *Cladium jamaicense* leaf litter was removed from plants from all mesocosms. Litter was oven-dried at 40°C for 1 week and placed into 1-mm mesh bags ($n = 2$ per replicate mesocosm per sampling event). Litter bags were incubated on surface soils within each mesocosm beginning 11 April 2017 for 155 and 317 days. Collected sample bags were immediately placed on ice and returned to the lab. Incubated litter was rinsed with deionized water, oven-dried at 60°C until mass stabilized, and subsequently weighed for individual mass remaining. A decay constant, k (day $^{-1}$), was calculated over sampling intervals as a first-order decay constant using the following:

$$y = e^{-kt}$$

where y is the proportion of mass remaining at time t .

After completion of the freshwater restoration manipulation, samples for C, N, and P content were collected for live sawgrass leaf, live fine root, and soil ($n = 6$ replicates per legacy treatment). The live sawgrass leaf and root were identified based on color and texture grown from short shoots (<40 cm). We cut soil cores into a rectangular cuboid (56 cm 2 surface area \times 25 cm height). Biomass and soil samples were oven-dried at 60°C until mass stabilized. Dried samples were ground using a ball mill (8000-D, Spex SamplePrep, Metuchen, New Jersey, U.S.A.) and subsequently analyzed for C and N content using a CHN Analyzer (Carlo Erba 1,500, Milan, Italy) and P content using the ash/acid extraction method followed by the ascorbic acid method for spectrophometric analysis (Solórzano & Sharp 1980). Soil bulk density was determined by dividing dry weight by core volume.

Ecosystem Metabolism

Monthly CO₂ fluxes were measured ($n = 4$ replicates per legacy treatment) around midday (approximately 10:00 to approximately 15:00) from May 2017 to June 2018. We used transparent polycarbonate chambers consisting of a bottom collar, the main chamber (0.2 m 2 \times 120 cm height), and a top lid (0.2 m 2 \times 30 cm height). We manipulated multiple levels of photosynthetically active

radiation (PAR) to mimic diel changes in PAR by shading chambers with different layers of plastic mesh. Gas samples were measured on a CO₂/H₂O analyzer (LI-840A, LI-COR Biosciences, Lincoln, NE, U.S.A.). Net ecosystem productivity (NEP) was calculated as the linear slope of CO₂ concentration over time for the 30–100% light levels, and ecosystem respiration (ER) was calculated as the slope of CO₂ concentration under the dark condition (assumed to be similar to nighttime ER). Gross ecosystem productivity (GEP) was estimated by summing NEP and ER. We used nonlinear least square models for fitting monthly CO₂ fluxes with environmental variables. Briefly, we fitted hyperbolic relationships between GEP and PAR, and exponential relationships between ER and temperature per plot on each sampling event. Fluxes of GEP and ER were modeled at a 15-min frequency by using the empirically derived parameters and climatic data (PAR, temperature). See Supplement S1 for additional details.

Statistical Analyses

All statistical analyses were conducted using R v.1.0.143 (Core Team 2017). Differences in water physicochemistry, aboveground biomass, and short-term carbon flux rate among legacy treatments were tested through linear mixed-effects models using the R package “nlme” (Pinheiro et al. 2018). The model was to determine the main effect of saltwater legacy, P legacy, time (sampling date), and their interactions with a random effect of plot number. When an interactive effect of saltwater legacy and date was detected, a pairwise comparison within each sampling event was assessed with a least-squares ANOVA using the “lsmeans” package (Lenth 2018) to identify the effectiveness of the saltwater legacy treatments (i.e. SW_L, SWP_L) and the P legacy treatments (i.e. FWP_L, SWP_L) within sampling dates. Legacy effects on the overall mean of belowground biomass, soil bulk density, leaf litter breakdown, modeled carbon flux, and carbon and nutrient content were determined using a two-way ANOVA. We then assessed mean differences among the control and legacy treatments using a least-square means multiple comparison test adjusted to Tukey’s HSD. Log-transformed variables were used for meeting the normality of residuals and the homogeneity of variances using visual inspection with a normal probability plot. The critical level of significance was set at $\alpha = 0.05$.

Mann–Kendall and Theil–Sen slope tests were employed to assess trajectories of physicochemical conditions, aboveground biomass, and short-term carbon flux. All values were analyzed without data transformation because the non-parametric methods have advantages of no strict rules required for residuals with non-normality, time-series data with serial dependence, and missing data (Sen 1968; Hirsch & Slack 1984; Theil 1992). We applied the Mann–Kendall test of the null hypothesis that there is no monotonic trend at a significance level of 0.05. When the null hypothesis was rejected ($p < 0.05$), a slope of an existing monotonic trend was estimated with the Theil–Sen slope. The modified Mann–Kendall test determined the statistical significance of the Theil–Sen slope after accounting for serial autocorrelation using the R packages “Kendall” (McLeod 2015) and “fume” (Santander Meteorology Group 2012). The slope of lines over a pair of all available data points was calculated as

the median of all possible pairwise slopes between each and another monthly data. To ease the comparison among legacy treatments, we provided a percent change (% change year⁻¹) from a mean of sample data over the observation period (Yue & Hashino 2003).

Differences in modeled ecosystem metabolism were determined using pairwise comparison. Modeled metabolism outputs were subtracted between the FW and other legacy treatments (e.g. $\Delta GPP_{(n=1-1,000,FW-FWP)} = GPP_{(n=1-1,000,FW)} - GPP_{(n=1-1,000,FWP)}$) for all combinations (i.e. FW versus FWP_L, SW_L, and SWP_L). The differences were either positive or negative values with the mean of zero if the modeled metabolism in the FW and legacy treatment plots were very similar. Statistical difference was defined when zero lies above or below 2.5% of a probability distribution (i.e. a significance level of 0.05). This approach is conceptually straightforward and avoids a strict assumption of normal distribution in other parametric tests and has been frequently used to find differences in resampling methods (Crowley 1992; Neubauer 2013).

Results

Surface and Porewater Chemistries

Adding freshwater reduced salinities from 9.2 to 2.8 ppt in porewater and from 8.5 to 2.9 ppt in surface water in SW_L and SWP_L

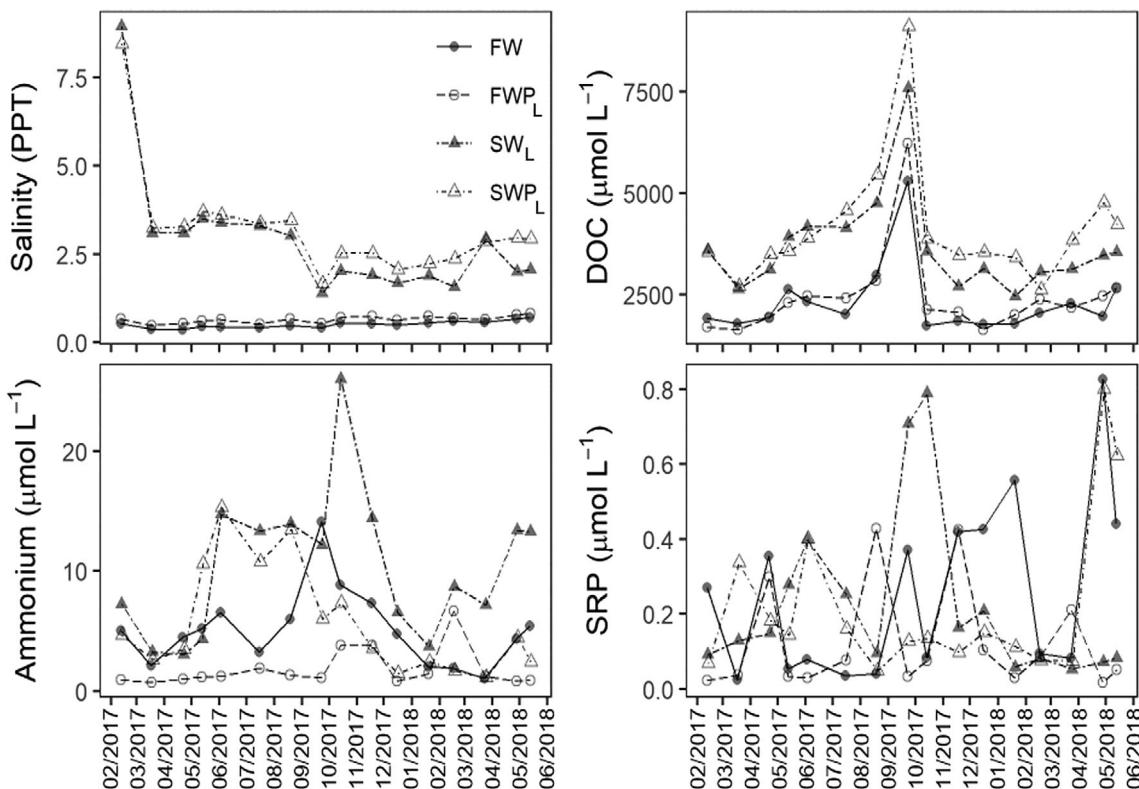


Figure 2. Changes in porewater physicochemistry during freshwater restoration of freshwater sawgrass (*Cladium jamaicense*) peat wetland mesocosms from freshwater (control, FW), and legacy elevated saltwater and phosphorus (P) treatments [elevated P (FWP_L), elevated saltwater (SW_L), and elevated saltwater and P (SWP_L)]. Error bars are omitted for clarity.

(Fig. 2, Table 2). Salinities were maintained at low brackish levels (approximately 2.8 ppt) until September 2017, when Hurricane Irma's rainfall further decreased salinity by approximately 55% in the saltwater legacy plots. A long-term decline in salinity was observed in the SW_L (approximately -5% year⁻¹) (Table 2). Added freshwater rapidly reduced porewater HS⁻ concentrations from 0.22 mmol L⁻¹ in the SW_L and 0.12 mmol L⁻¹ in the SWP_L within the first 2 months (Table 2).

Porewater NH₄⁺ concentrations were higher in the SW_L than in the FWP_L (ANOVA, $F_{1,21} = 8.1, p < 0.01$) and P legacies (ANOVA, $F_{1,21} = 8.9, p < 0.01$) (Table 2). Higher porewater NH₄⁺ concentrations were observed in SW_L and SWP_L than FW from June to October 2017 due to the saltwater legacy (Table 2). In contrast, the P legacy decreased porewater NH₄⁺ concentrations from June to December 2017 (Table 2), suggesting biotic uptake during the wet growing season. When the saltwater and P legacies coexisted (i.e. SWP_L), the effect of P legacy not only offset the saltwater legacy but also had a lasting impact, as NH₄⁺ concentrations declined over the study period in the SWP_L (Table 2). Similar to saltwater legacy effects on porewater NH₄⁺ concentrations, porewater DOC concentrations were higher and varied over time in SW_L and SWP_L due to an interactive effect of saltwater legacy and time (ANOVA, $F_{14,293} = 2.4, p < 0.01$) (Fig. 2, Table 2).

Table 2. Mean and standard error of physicochemical variables ($n = 6$ per treatment), biomass ($n = 4$ per treatment), and short-term metabolism ($n = 6$ per treatment) during freshwater restoration of freshwater sawgrass (*Cladium jamaicense*) peat wetland mesocosms from freshwater (control, FW), and legacy elevated saltwater and phosphorus (P) treatments [elevated P (FWP_L) elevated saltwater (SW_L), and elevated saltwater and P (SWP_L)]. A temporal trend (%-change year⁻¹) was estimated using the Mann–Kendall (MK) and Theil–Sen (T–S) slope tests. A trend (i.e. slope coefficient) can be interpreted as a percent change from the overall average. Bolded trends represent an increase (+) or decrease (−) at a significance level of $p < 0.05$. No trend analysis was conducted for HS[−] concentration, belowground biomass, bulk density, and leaf litter breakdown. Superscripted letters and bolded values represent increases ($p < 0.05$) among control and legacy treatment responses using two-way ANOVA and Tukey's HSD. (TP: Total phosphorus).

	FW						FWP _L						SW _L						SWP _L									
	Mean			T-S			MK			Mean			T-S			MK			Mean			T-S			MK			
	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	Mean	T-S	MK	
Surface water																												
Salinity (ppt)	0.54 ± 0.01 ^a	5.3	<0.001	0.63 ± 0.02 ^a	4.2	<0.001	2.2 ± 0.1^b	8.6 ± 0.4^b	−5.1	0.01	2.5 ± 0.07^b	8.7 ± 0.4^b	−2.2	−2	0.15													
Conductivity (S cm ^{−1})	0.98 ± 0.02 ^a	4.6	<0.001	1.1 ± 0.03 ^a	3.9	<0.001			−5	<0.01																	0.14	
DOC (µM)	1800 ± 50	2	0.01	1800 ± 50	1.6	0.08	2,100 ± 80	−3.4	0.14	2,300 ± 70	−1.7	0.43																
NH ₄ ⁺ (µM)	1.7 ± 0.3	5.1	0.08	2.4 ± 0.4	−3.1	0.76	3.6 ± 0.8	5.7	0.43	1.4 ± 0.07	−4.3	0.07																
SRP (µM)	0.23 ± 0.05	9.5	0.01	0.22 ± 0.04	4.1	0.42	0.1 ± 0.01	2.9	0.69	0.095 ± 0.02	−6.6	0.04																
TP (µM)	1.2 ± 0.1	1.1	0.20	0.97 ± 0.06	−7.7	0.07	0.73 ± 0.05	1.8	0.62	1.1 ± 0.05	−3.7	0.20																
Porewater																												
Salinity (ppt)	0.51 ± 0.01 ^a	4.7	<0.001	0.65 ± 0.02 ^a	2.9	<0.01	2.5 ± 0.1^b	9.1 ± 0.5^b	−5.7	0.02	2.8 ± 0.1^b	9.3 ± 0.4^b	−2.8	−2.6	0.34													
Conductivity (S cm ^{−1})	1.0 ± 0.06 ^a	4.3	<0.001	1.2 ± 0.03 ^a	2.9	<0.01																					0.31	
DOC (µM)	2,300 ± 100 ^a	−0.1	0.69	2,500 ± 100 ^a	0.7	0.06	3,700 ± 200^b	11 ± 2^b	−1.3	0.79	4,200 ± 300^b	5.8 ± 1^{ab}	0.2	0.72														
NH ₄ ⁺ (µM)	5.1 ± 0.6 ^{ab}	−4.5	0.55	1.9 ± 0.5 ^a	1.3	0.55	1.9	0.49																		0.04		
SRP (µM)	0.25 ± 0.05	8.2	<0.01	0.13 ± 0.03	−0.4	0.82	0.24 ± 0.05	−6.3	0.09	0.23 ± 0.06	−3.7	0.37																
HS [−] (mM)	0.01 ± 0.001 ^a	—	—	0.005 ± 0.001 ^a	—	—	0.22 ± 0.04 ^b	—	—	0.12 ± 0.01 ^b	—	—														—		
Aboveground biomass (g C m ^{−2})	296 ± 52.4 ^a	8.6	0.13	868 ± 123^{bc}	10.5	0.45	467 ± 56.6 ^{ac}	8.4	0.26	1,421 ± 128 ^b	23.6	0.02																
Belowground biomass (g C m ^{−2})	1,492 ± 146 ^a	—	—	2,226 ± 231^b	—	—	—	—	—	1881 ± 136 ^{ab}	—	—	2096 ± 165^b	—	—													
Soil bulk density (g C cm ^{−3})	0.14 ± 0.01	—	—	0.14 ± 0.01	—	—	0.17 ± 0.01	—	—	0.18 ± 0.01	—	—	0.18 ± 0.01	—	—											—		
Leaf litter breakdown (k day ^{−1})	0.001 ± 0.0004 ^a	—	—	0.0011 ± 0.0003 ^a	—	—	0.0017 ± 0.004 ^{ab}	—	—	—	—	—	0.0026 ± 0.0005^b	—	—										—			
GEP(µmol C m ^{−2} s ^{−1})	7.53 ± 0.43 ^a	−0.3	0.55	20.1 ± 0.66^b	−0.2	0.38	11.6 ± 0.52 ^a	−0.8	0.26	21.0 ± 0.76 ^b	0.6	0.09														0.01		
ER (µmol C m ^{−2} s ^{−1})	2.28 ± 0.15 ^a	0.5	0.38	7.47 ± 0.26^b	0	0.92	4.30 ± 0.17 ^a	0.3	0.52	9.28 ± 0.38^b	0.9	0.09														<0.01		

Surface water and porewater SRP were not different among legacy treatments (Table 2). In all legacy treatments, there were no temporal trends in SRP concentrations in porewater; however, there was a decline in SRP concentrations in surface water in the SWP_L (Table 2). Hurricane Irma caused rapid increases in porewater DOC concentrations in all control and legacy treatments; whereas hurricane-induced increases in pore NH₄⁺ and SRP concentrations were only observed in the SW_L.

Above- and belowground Biomass and Elemental Concentrations

The SW_L had increased aboveground growth in the last 6 months (from February to June 2018) of freshwater restoration (LSMEANS, $p < 0.05$). An interactive effect of saltwater legacy and date (ANOVA, $F_{5,105} = 3.16, p < 0.05$) indicated a shift of saltwater legacy effect over time. The main effect of P legacy (ANOVA, $F_{1,21} = 33.6, p < 0.001$) remained effective throughout the study. Aboveground biomass was lower in FWP_L than SWP_L, and aboveground biomass increased in SWP_L but not in FWP_L wetlands (Table 2). A shift of saltwater legacy observed in aboveground biomass was also observed for belowground biomass. During freshwater restoration, the P legacy (ANOVA, $F_{1,21} = 7.8, p < 0.05$) resulted in the highest belowground biomass in FWP_L followed, in order of decreasing biomass, by SWP_L, SW_L, and FW (Table 2).

Carbon concentrations did not vary in sawgrass leaf and fine root among legacy treatments, whereas nutrient concentrations varied due to legacy effects of salt and P (Table 3). Different legacies affected nitrogen concentrations in sawgrass leaf (saltwater legacy, ANOVA, $F_{1,21} = 5.37, p < 0.05$) and root (P legacy, ANOVA, $F_{1,21} = 5.30, p < 0.05$). Regardless of the differing legacies, nitrogen concentrations in leaf and root were lowered in all legacy treatments relative to FW (Table 3). Although P concentrations in root did not vary among legacy treatments, P concentrations were lowered in all legacy

treatments relative to FW (Table 3). By contrast, leaf P concentrations were highest in FWP_L and not different among the other legacy treatments or FW (Table 3). There were no interactive effects between the saltwater and P legacies (ANOVA, $F_{1,20} = 0.68, p > 0.05$) on leaf P contents; however, the saltwater legacy (ANOVA, $F_{1,21} = 11.0, p < 0.01$) decreased and the P legacy (ANOVA, $F_{1,21} = 9.7, p < 0.01$) increased leaf P contents. As a result of the opposite effects of the legacies, nutrient use efficiency of sawgrass leaf, calculated as a molar ratio of C to P content, was the highest in SW_L (7454) followed by SWP_L (6693), FW (6512), and FWP_L (4995), implying higher efficiency for the use of carbon and P in saltwater legacy plots than the freshwater plots.

Leaf Litter Breakdown and Elemental Concentrations

The highest litter k occurred in SWP_L and was approximately twice that of FW and FWP_L, whereas the intermediate breakdown rate in SW_L was not different from any other plots (Table 2). Litter breakdown rates were higher in the saltwater legacy than in the freshwater plots (ANOVA, $F_{1,21} = 8.89, p < 0.01$), where the P and interactive P and saltwater legacies were absent.

After the 305 days of incubation, carbon and nitrogen concentrations in sawgrass leaf litter did not vary among legacy treatments; however, leaf litter P concentrations were the highest in SWP_L followed by FWP_L, FW, and SW_L due to the P legacy (ANOVA, $F_{1,21} = 16.7, p < 0.001$) (Table 3). Phosphorus concentrations were approximately 2 \times higher in live sawgrass leaf than leaf litter in FW, as well as FWP_L and SW_L, whereas P concentrations were similar between the live leaves and leaf litter in the SWP_L (Table 3).

Soil Elemental Concentrations

The saltwater legacy (ANOVA, $F_{1,21} = 9.4, p < 0.01$) increased soil bulk density by 25% in the saltwater legacy treatments;

Table 3. Mean ($n = 6$ per treatment) and standard error of carbon (C), nitrogen (N), and P content in sawgrass (*Cladium jamaicense*) leaves, sawgrass leaf litter, soil, and sawgrass roots during freshwater restoration of freshwater sawgrass (*C. jamaicense*) peat wetland mesocosms from freshwater (control, FW), and legacy elevated saltwater and phosphorus (P) treatments [elevated P (FWP_L), elevated saltwater (SW_L), and elevated saltwater and P (SWP_L)]. The live samples were collected on the last day (01 June 2018) of the experiment, while the leaf litter samples were collected upon the termination of litter decomposition experiment (305th day, 12 February 2018). Superscripted letters and **bolded** values represent significant differences ($p < 0.05$) among control and legacy treatment responses using two-way ANOVA and Tukey's HSD.

	FW	FWP _L	SW _L	SWP _L
Sawgrass leaf				
C (mg g ⁻¹)	438 ± 4.9	439 ± 4.8	431 ± 8.2	448 ± 7.3
N (mg g ⁻¹)	6.6 ± 0.5^a	5.9 ± 0.5 ^{ab}	5.7 ± 0.2 ^{ab}	4.9 ± 0.3^b
P (μg g ⁻¹)	179 ± 11^a	230 ± 13^b	151 ± 8.8^a	176 ± 28^a
Sawgrass leaf litter				
C (mg g ⁻¹)	474 ± 4.6	488 ± 12	485 ± 8.3	474 ± 7.6
N (mg g ⁻¹)	6.2 ± 0.4	6.1 ± 0.6	6.9 ± 0.4	6.9 ± 0.3
P (μg g ⁻¹)	78.8 ± 13^a	145 ± 15 ^{ab}	76.4 ± 6.6^a	192 ± 55^b
Soil				
C (mg g ⁻¹)	453 ± 6.9	462 ± 7.8	437 ± 6.9	431 ± 5.6
N (mg g ⁻¹)	26 ± 0.9	26 ± 1	27 ± 0.6	26 ± 0.8
P (μg g ⁻¹)	456 ± 26	455 ± 32	430 ± 21	543 ± 57
Sawgrass root				
C (mg g ⁻¹)	860 ± 78	965 ± 12	942 ± 11	931 ± 3.6
N (mg g ⁻¹)	6.5 ± 0.59^a	5.6 ± 0.52 ^{ab}	5.6 ± 0.22 ^{ab}	4.4 ± 0.48^b
P (μg g ⁻¹)	185 ± 25	178 ± 11	158 ± 25	134 ± 6.7

however, there were no differences in bulk density among legacy treatments (Table 2). Soil carbon, nitrogen, and P concentrations did not vary among control and legacy treatments (Table 3).

Instantaneous Ecosystem Metabolism

There were a total of 13 gas flux measurements in 16 subsampled plots ($n = 4$ per legacy treatment) per event except during overcast days (the number of plots per event [n] = 16 for 7 months, $n = 15$ for 3 months, $n = 12$ for 2 months, $n = 8$ for 1 month). Overall, average monthly GEP rates ranged from 6.3 ± 0.81 (standard error) $\mu\text{mol m}^{-2} \text{ s}^{-1}$, measured in FW in

June 2017, to $24.5 \pm 5.7 \mu\text{mol m}^{-2} \text{ s}^{-1}$, measured in SWP_L in November 2017 (Table 2). The P legacy (ANOVA, $F_{1,13} = 39.8, p < 0.001$) resulted in higher short-term GEP rates in FWP_L and SWP_L than FW and SW_L, whereas the saltwater legacy had no effect on GEP (ANOVA, $F_{1,13} = 2.5, p > 0.05$) (Table 2).

The use of temperature-derived ER was not feasible because a logarithmic scale of ER rates was not correlated with air temperature in any plots ($n = 15, R^2 < 0.21, p > 0.05$) except for one FWP_L plot ($R^2 = 0.73, p < 0.01$). We assumed constant respiration rates in each month for all plots except for one FWP_L plot where the air temperature adjusted short-term ER rates. Overall, average monthly ER rates ranged from $1.4 \pm 0.29 \mu\text{mol m}^{-2} \text{ s}^{-1}$,

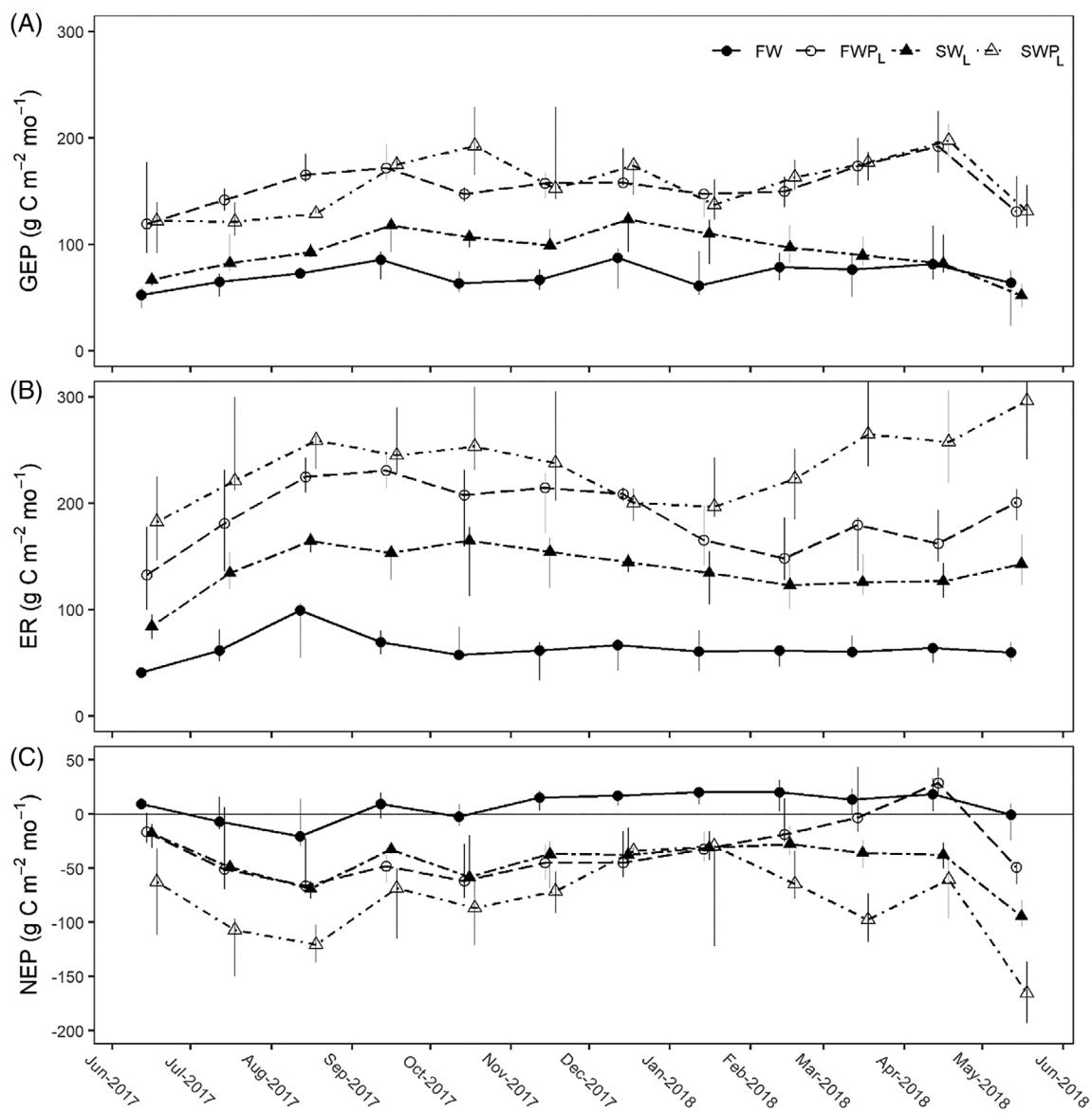


Figure 3. Values of modeled (A) gross ecosystem productivity (GEP) at maximum light levels, (B) ecosystem respiration (ER), and (C) net ecosystem productivity (NEP) during freshwater restoration of freshwater sawgrass (*Cladium jamaicense*) peat wetland mesocosms from freshwater (control, FW), and legacy elevated saltwater and phosphorus (P) treatments [elevated P (FWP_L), elevated saltwater (SW_L), and elevated saltwater and P (SWP_L)]. Points are medians, and error bars are 25th and 75th percentiles of 1,000 model outputs.

measured in SW_L in May 2017, to $12 \pm 1.3 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$, measured in SWP_L in June 2018 (data not shown). Both the P legacy (ANOVA, $F_{1,13} = 67.3$, $p < 0.001$) and saltwater legacy (ANOVA, $F_{1,13} = 11.4$, $p < 0.01$) stimulated short-term ER rates with higher rates in FWP_L and SWP_L than FW and SW_L (Table 2). A steady increase in short-term ER rates (Mann-Kendall, $p < 0.01$) was observed in SWP_L (Table 2).

Modeled Ecosystem Metabolism

There were differences in monthly modeled (see Supplement S1 for details) GEP rates in FWP_L and SWP_L compared to FW (Fig. 3A). These differences are not surprising, given that the P legacy plots had higher aboveground biomass (Table 2). Monthly modeled GEP rates in SW_L differed ($p < 0.05$) from FW only in 2 months during the wet season. Overall, there were effects of P legacy (ANOVA, $F_{1,34} = 269$, $p < 0.001$) and saltwater legacy (ANOVA, $F_{1,34} = 10.8$, $p < 0.01$) on modeled GEP rates. The P legacy approximately doubled modeled GEP rates in FWP_L compared to FW, whereas the saltwater legacy resulted in approximately 30% higher modeled GEP rates in SW_L than FW (Fig. 3(A)). Yet the P legacy effect appeared to offset the saltwater legacy when the two legacies coexisted, as modeled GEP rates in SWP_L and FWP_L were similar (Fig. 3(A)).

Modeled ER rates were different between the control and all legacy treatments with the highest rate in the SWP_L followed by FWP_L , SW_L , and FW (Fig. 3(B)). The saltwater legacy (ANOVA, $F_{1,34} = 24.7$, $p < 0.001$) approximately doubled the average modeled ER rate in SW_L compared to FW. The P legacy (ANOVA, $F_{1,34} = 454$, $p < 0.001$) approximately tripled the average modeled ER rate in FWP_L compared to FW. In contrast to the negligible saltwater legacy with P legacy on modeled GEP, the combined saltwater and P legacies (ANOVA, $F_{1,33} = 28.4$, $p < 0.001$) approximately quadrupled the average modeled ER rate in SWP_L compared to FW.

Higher modeled GEP than ER rates in FW resulted in net autotrophy, indicating that the system was a net sink of carbon annually (Fig. 3C). In contrast, all other legacy treatments were a net source of carbon (i.e. net heterotrophy). The P legacy (ANOVA, $F_{1,34} = 26.7$, $p < 0.001$) and the saltwater legacy (ANOVA, $F_{1,34} = 5.41$, $p < 0.05$) resulted in a similar magnitude of net heterotrophy (i.e. a system was releasing more carbon than it was retaining) in FWP_L and SW_L (Fig. 3C). There were no interactive effects between the saltwater and the P legacies (ANOVA, $F_{1,33} = 1.4$, $p > 0.05$); however, the additive effect of saltwater and P legacies reduced modeled NEP rates in SWP_L compared to FWP_L and SW_L by nearly 2× (Fig. 3C).

Discussion

Our results demonstrate saltwater and P legacies persisted following a continuous exposure (approximately 2 years) to saltwater, despite freshwater reintroduction (approximately 1 year), and maintained a net carbon release that would delay the recovery of net ecosystem carbon storage. The severity and duration of exposure to elevated saltwater and P preceding our freshwater restoration manipulation impacted the recovery of

ecosystem processes, resulting in a carbon imbalance in the saltwater-exposed plots. We observed P legacies, continuously stimulating biotic growth and litter breakdown processes during freshwater restoration. Some plant responses recovered from prior saltwater exposure, especially when saltwater and P co-occurred. Notably, sawgrass root biomass increased the most during freshwater restoration in wetlands with a saltwater legacy. A reduction in salt stress, likely lowering ionic strength and toxicity, restored sawgrass above- and belowground biomass with a lag of approximately one year. Increased carbon and nitrogen availability also enhanced microbial SRP uptake and decomposition of plant litter, especially in the sediment-water interface, where enhanced nutrient recycling would supply emergent plant growth. However, this positive feedback mechanism disrupted the ecosystem carbon balance due to a net increase in autotrophic and heterotrophic respiration relative to the amount of carbon inputs. We summarize and interpret how the saltwater and P legacies differentially influenced ecosystem components and metabolic pathways to understand the recovery trajectory of restored wetlands and its implication for freshwater restoration in the Florida Everglades.

Saltwater and P Legacies on Net Ecosystem Carbon Storage

Although elevated saltwater previously reduced sawgrass growth, sawgrass exposed to elevated saltwater had higher growth during freshwater restoration. Freshwater restoration increased sawgrass aboveground biomass in the legacy saltwater and legacy saltwater and P treatments. These increases in restored sawgrass growth and increased biomass are likely explained by increased nutrient use efficiency (i.e. a molar C:P ratio in sawgrass leaves), which increased 56–70% in the saltwater legacy plots compared to 7% in FW plots. Additionally, belowground biomass increased in the legacy saltwater and legacy saltwater and P treatments during restoration. Although not measured in this study, increased growth in recovering *Cladium jamaicense* plants could have been facilitated by arbuscular mycorrhiza (Jayachandran & Shetty 2003), which can reduce salt stress through nutrient and water acquisition and enhance photosynthetic efficiency (Evelin et al. 2009 and references therein). Our results suggest that freshwater *C. jamaicense* growth and biomass were temporarily inhibited by elevated saltwater but were resilient once the salinity stress was mitigated by freshwater restoration.

Freshwater restoration must take into account how legacies of saltwater and nutrients may affect ER and GEP differently. Some degree of carbon imbalance was expected, especially during early restoration because enhanced plant respiration often comes before recovering photosynthesis under stress (Flexas et al. 2005). However, this imbalance persisted throughout our study, in part because the saltwater legacy increased autotrophic and heterotrophic respiration relative to GEP. The disproportionate saltwater effect on ER might be caused by a reduction in nutrient resorption efficiency for photosynthesis (i.e. a nutrient-conserving mechanism that transfers nutrients from senesced to green leaves, [Aerts 1996]) and an enhancement of respiration from nutrient-enriched litter. This phenomenon was observed in sawgrasses grown in P-enriched soils

($>500 \mu\text{g g}^{-1}$ P) (Richardson et al. 1999) due to enhanced microbial P assimilation and immobilization into the surface soil or leaf litter (Debusk & Reddy 2005; Pivničková et al. 2010). In our study, average soil P content exceeded the enrichment threshold ($543 \mu\text{g g}^{-1}$ P) only in the SWP legacy treatment. Also, P concentrations were higher in leaf litter than live sawgrass leaves in marshes with previous exposure to elevated saltwater and P.

Previous studies have highlighted the geochemical and biological pathways that can enhance the role of internal P loading in response to P enrichment. There are multiple mechanistic pathways of P legacies in wetlands, including growth-dependent P storage in photosynthetic communities (Newman et al. 1996; Noe et al. 2002; Gaiser 2009), redox-dependent P retention in flocculent sediments (Reddy et al. 1999), redox-dependent microbial uptake (Newman & Reddy 1993), pH-mediated P release (Dierberg et al. 2002), anoxia-promoted P release (Parsons et al. 2017), macrophyte translocation of P from soil to water column (Noe & Childers 2007), and faunal regeneration of P from leaf litter or plankton to higher trophic levels (Schindler et al. 1993; Hagerthey et al. 2014). In P-limited ecosystems like the Everglades, P legacies can affect the storage and cycling of P even after nutrient inputs are reduced or removed (Childers et al. 2019; Sarker et al. 2020). Therefore, nutrient legacies might persist longer than saltwater legacies, and the differential reduction in the multiple disturbance legacies complicates predictions for recovery of carbon balance in coastal ecosystems (Tully et al. 2019).

Freshwater Restoration and Net Ecosystem Carbon Storage

Our measurements of net autotrophic conditions (net carbon sink) in control wetlands were within the high end of reported NEP values ($-32\text{--}91 \text{ g C m}^{-2} \text{ year}^{-1}$) from a short-hydroperiod, freshwater sawgrass marsh (Zhao et al. 2019). In contrast, net heterotrophic conditions (net carbon source) measured from legacy treatment wetlands were an order of magnitude lower than the lowest reported in situ rates of NEP from the same short-hydroperiod, freshwater sawgrass marsh (Zhao et al. 2019). Our results suggest that legacy effects of saltwater and P in Everglades marshes might cause degraded wetlands to be persistent as net carbon sources.

Elevation loss from saltwater intrusion appears to be a reinforcing positive feedback on reducing net ecosystem productivity and net ecosystem carbon storage. Our findings of net ecosystem heterotrophy in sawgrass wetlands that were previously exposed to elevated saltwater with and without added P, and that subsequently all lost soil elevation, underscore the importance of soil elevation to net ecosystem carbon storage. Assuming constant soil carbon contents and bulk densities, modeled NEP rates were extrapolated to a change in soil elevation in experimental monoliths. For example, average NEP in FW plots ($7.6 \text{ g C m}^{-2} \text{ mo}^{-1}$) was divided by soil carbon content ($0.45 \text{ g C g-soil}^{-1}$) and bulk density ($0.14 \text{ g-soil cm}^{-3}$), resulting in 1.4 mm year^{-1} in the 2018 water year. The rate is within a range of vertical accretion rates (1.6 mm year^{-1}) in unenriched portions of central and southern Everglades where sawgrass was the dominant emergent vegetation

(Craft & Richardson 1993, 1998). In the legacy plots, net heterotrophic rates translate into a loss of soil elevation at a rate of $-6.3 \text{ mm year}^{-1}$ in FWP, $-7.1 \text{ mm year}^{-1}$ in SW and $-12.5 \text{ mm year}^{-1}$ in SWP. Assuming a 150 cm ground level, constant elevation changes, and a local sea-level rise (7.7 mm year^{-1} [Dessu et al. 2018]), it will take 60 years to reach a 30-cm soil thickness, below which soil depths cannot support roots' structural integrity. Yet our prediction is likely conservative because other confounding factors, such as gravity compaction and seasonal hydrologic cycle, can accelerate soil organic carbon loss (Cahoon et al. 2011; Day et al. 2011).

The lack of seasonality in carbon fluxes within our mesocosm study was likely caused by constant (approximately 10 cm) water inundation. This stationarity condition differs from the historical Everglades, where water depths fluctuated seasonally from below-ground levels to a high of about 45 cm in sawgrass ridges (McVoy 2011). A comparison of sawgrass biomass across a hydrologic gradient provides insight into the optimal condition for sawgrass growth; an annual average water depth of 10–20 cm results in the highest growth (Childers et al. 2006). Flooding might suppress soil organic carbon mineralization (Neubauer et al. 2000; Wilson et al. 2018) during the dry season when the soil surface is typically re-aerated. Thus, our experimental design might have a marginally positive impact on sawgrass productivity but underestimate microbial respiration on an annual basis.

Nutrient enrichment and saltwater intrusion often lead to plant community shifts in wetlands. Our model prediction did not consider a shift in the macrophyte community from sawgrass to other emergent vegetation (e.g. *Typha domingensis*, *Eleocharis interstincta*) because macrophytes did not change their diversity or composition in response to nutrient enrichment during our mesocosm study, similar to other in situ studies (Craft et al. 1995; Chiang et al. 2000). However, previous studies documented marsh-mangrove transition zones (ecotones) encroaching inland with saltwater in the coastal Everglades (Ross et al. 2000). Increased seawater inundation favors the growth and expansion of mangroves that build up organic compounds belowground fast to keep up with rising sea levels (McKee et al. 2007). For example, in mangrove wetlands, soil accretion rate ranges from 2 to 8 mm year^{-1} (McKee 2011; Krauss et al. 2017). Therefore, future studies should address the potential change of soil accretion rate associated with a vegetation community transition (e.g. Breithaupt et al. 2020).

Freshwater restoration can reduce the rates of saltwater intrusion, while also supporting the recovery of carbon balance in wetlands with previous exposure to saltwater. The capacity of ecosystems to retain stored carbon is enhanced where slow biogeochemical processes predominate (Newman et al. 2017; Ratajczak et al. 2017). However, rapid carbon losses with disturbance are difficult to regain in oligotrophic ecosystems (Kominoski et al. 2018; Gaiser et al. 2020). The remnant ecosystems of the Florida Everglades are highly hydrologically modified and engineered, yet they are undergoing rapid freshwater restoration (Childers et al. 2019; Sarker et al. 2020). Reestablishing freshwater discharge and restoring historical flow pattern are critical to mitigate saltwater intrusion (Kaplan et al. 2010)

and are a fundamental goal of Everglades restoration (NASEM 2018). Although we found little evidence of short-term (16 months) recovery of carbon balance in marshes with collapsed soils, freshwater restoration efforts in coastal ecosystems is critical for preventing stored carbon loss in more intact ecosystems. Our study found that saltwater and P legacies transformed freshwater peat wetlands into net carbon sources, despite more than a year of freshwater restoration, suggesting slow recovery of net ecosystem carbon storage in nutrient-limited freshwater wetlands exposed to saltwater intrusion. Our modeled results of reduced NEP in wetland mesocosms at low concentrations of porewater salinity (approximately 2 ppt) from legacy saltwater treatments are similar to field measurements made at similar porewater salinities including: field-based salinity dosing studies in freshwater marshes of the Everglades (Wilson et al. 2018), freshwater tidal marshes of the Altamaha River (Herbert et al. 2018), and tidal brackish marshes in the Mobile Bay (Wilson et al. 2015) and Delaware River estuaries (Weston et al. 2011). Sustained freshwater restoration must reverse legacies of saltwater intrusion and nutrient enrichment to increase net ecosystem carbon storage that is necessary to protect coastal ecosystems exposed to sea-level rise.

Acknowledgments

Logistical support was provided by the Florida Coastal Everglades Long Term Ecological Research (FCE-LTER) program, Everglades National Park, and the South Florida Water Management District. This research was supported by the National Science Foundation through the FCE-LTER program (DEB-1237517). We thank Stephen Davis and two anonymous reviewers who provided helpful feedback on previous versions on this manuscript. This manuscript is contribution number 1032 from the Southeast Environmental Research Center in the Institute of Environment at Florida International University.

LITERATURE CITED

Aerts R (1996) Nutrient resorption from senescing leaves of perennials: are there general patterns? *Journal of Ecology* 84:597–608

Breithaupt JL, Smoak JM, Bianchi TS, Vaughn DR, Sanders CJ, Radabaugh KR, Osland MJ, Feher LC, Lynch JC, et al. (2020) Increasing rates of carbon burial in Southwest Florida coastal wetlands. *Journal of Geophysical Research: Biogeosciences* 125:e2019JG005349

Cahoon DR, Perez BC, Segura BD, Lynch JC (2011) Elevation trends and shrink–swell response of wetland soils to flooding and drying. *Estuarine, Coastal and Shelf Science* 91:463–474

Chambers LG, Osborne TZ, Reddy KR (2013) Effect of salinity-altering pulsing events on soil organic carbon loss along an intertidal wetland gradient: a laboratory experiment. *Biogeochemistry* 115:363–383

Chambers LG, Steinmuller HE, Breithaupt JL (2019) Toward a mechanistic understanding of “peat collapse” and its potential contribution to coastal wetland loss. *Ecology* 100:e02720

Chapin FS, Woodwell GM, Randerson JT, Rastetter EB, Lovett GM, Baldocchi DD, et al. (2006) Reconciling carbon-cycle concepts, terminology, and methods. *Ecosystems* 9:1041–1050

Charles SP, Kominoski JS, Troxler TG, Gaiser EE, Servais S, Wilson BJ, et al. (2019) Experimental saltwater intrusion drives rapid soil elevation and carbon loss in freshwater and brackish Everglades marshes. *Estuaries and Coasts* 42:1868–1881

Chiang C, Craft CB, Rogers DW, Richardson CJ (2000) Effects of 4 years of nitrogen and phosphorus additions on Everglades plant communities. *Aquatic Botany* 68:61–78

Childers DL, Gaiser E, Ogden LA (2019) The Coastal Everglades: the dynamics of social-ecological transformation in the South Florida landscape. New York, NY: Oxford University Press

Childers DL, Iwaniec D, Rondeau D, Rubio G, Verdon E, Madden CJ (2006) Responses of sawgrass and spikerush to variation in hydrologic drivers and salinity in Southern Everglades marshes. *Hydrobiologia* 569:273–292

Core Team R (2017) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria

Craft CB, Richardson CJ (1993) Peat accretion and N, P, and organic C accumulation in nutrient-enriched and unenriched Everglades peatlands. *Ecological Applications* 3:446–458

Craft CB, Richardson CJ (1998) Recent and long-term organic soil accretion and nutrient accumulation in the Everglades. *Soil Science Society of America Journal* 62:834–843

Craft CB, Vymazal J, Richardson CJ (1995) Response of everglades plant communities to nitrogen and phosphorus additions. *Wetlands* 15:258–271

Crowley PH (1992) Resampling methods for computation-intensive data analysis in ecology and evolution. *Annual Review of Ecology and Systematics* 23: 405–447

Daoust RJ, Childers DL (1998) Quantifying aboveground biomass and estimating net aboveground primary production for wetland macrophytes using a non-destructive phenometric technique. *Aquatic Botany* 62:115–133

Davis S, Ogden JC (1994) Everglades: the ecosystem and its restoration. Boca Raton, FL: CRC Press

Davis SM (1991) Growth, decomposition, and nutrient retention of *Cladium jamaicense* Crantz and *Typha domingensis* Pers. in the Florida Everglades. *Aquatic Botany* 40:203–224

Day JW, Kemp GP, Reed DJ, Cahoon DR, Boumans RM, Suhayda JM, Gambrell R (2011) Vegetation death and rapid loss of surface elevation in two contrasting Mississippi delta salt marshes: the role of sedimentation, autocompaction and sea-level rise. *Ecological Engineering* 37:229–240

Debusk WF, Reddy KR (2005) Litter decomposition and nutrient dynamics in a phosphorus enriched Everglades marsh. *Biogeochemistry* 75:217–240

Dessu SB, Price RM, Troxler TG, Kominoski JS (2018) Effects of sea-level rise and freshwater management on long-term water levels and water quality in the Florida Coastal Everglades. *Journal of Environmental Management* 211:164–176

Dierberg FE, DeBusk TA, Jackson SD, Chimney MJ, Pietro K (2002) Submerged aquatic vegetation-based treatment wetlands for removing phosphorus from agricultural runoff: response to hydraulic and nutrient loading. *Water Research* 36:1409–1422

Evelin H, Kapoor R, Giri B (2009) Arbuscular mycorrhizal fungi in alleviation of salt stress: a review. *Annals of Botany* 104:1263–1280

Flexas J, Galmes J, Ribas-Carbo M, Medrano H (2005) The effects of water stress on plant respiration. Pages 85–94. In: Lambers H, Ribas-Carbo M (eds) *Plant respiration: from cell to ecosystem. Advances in Photosynthesis and Respiration* Springer Netherlands, Dordrecht

Flynn KM, McKee KL, Mendelsohn IA (1995) Recovery of freshwater marsh vegetation after a saltwater intrusion event. *Oecologia* 103:63–72

Gaiser E (2009) Periphyton as an indicator of restoration in the Florida Everglades. *Ecological Indicators* 9:S37–S45

Gaiser EE, Bell DM, Castorani MCN, Childers DL, Groffman PM, Jackson CR, et al. (2020) Long-term ecological research and evolving frameworks of disturbance ecology. *Bioscience* 70:141–156

Hagerthey SE, Cook MI, Kobza RM, Newman S, Bellinger BJ (2014) Aquatic faunal responses to an induced regime shift in the phosphorus-impacted Everglades. *Freshwater Biology* 59, 1389–1405

Herbert ER, Boon P, Burgin AJ, Neubauer SC, Franklin RB, Ardón M, Hopfensperger KN, Lamers LPM, Gell P (2015) A global perspective on

wetland salinization: ecological consequences of a growing threat to freshwater wetlands. *Ecosphere* 6:1–43

Herbert ER, Schubauer-Berigan J, Craft CB (2018) Differential effects of chronic and acute simulated seawater intrusion on tidal freshwater marsh carbon cycling. *Biogeochemistry* 138:137–154

Hirsch RM, Slack JR (1984) A nonparametric trend test for seasonal data with serial dependence. *Water Resources Research* 20:727–732

Jayachandran K, Shetty KG (2003) Growth response and phosphorus uptake by arbuscular mycorrhizae of wet prairie sawgrass. *Aquatic Botany* 76:281–290

Jiang J, DeAngelis DL, Smith TJ III, The SY, Koh HL (2012) Spatial pattern formation of coastal vegetation in response to hydrodynamics of soil pore-water salinity: a model study. *Landscape Ecology* 27:109–119

Jolly ID, McEwan KL, Holland KL (2008) A review of groundwater–surface water interactions in arid/semi-arid wetlands and the consequences of salinity for wetland ecology. *Ecohydrology* 1:43–58

Kaplan D, Muñoz-Carpentra R, Wan Y, Hedgepeth M, Zheng F, Roberts R, Service RR (2010) Linking river, floodplain, and vadose zone hydrology to improve restoration of a coastal river affected by saltwater intrusion. *Journal of Environmental Quality* 39:1570–1584

Kirwan ML, Megonigal JP (2013) Tidal wetland stability in the face of human impacts and sea-level rise. *Nature* 504:53–60

Kominoski JS, Gaiser EE, Baer SG (2018) Advancing theories of ecosystem development through long-term ecological research. *Bioscience* 68:554–562

Krauss KW, Cormier N, Osland MJ, Kirwan ML, Stagg CL, Nestlerode JA, Russell MJ, et al. (2017) Created mangrove wetlands store belowground carbon and surface elevation change enables them to adjust to sea-level rise. *Scientific Reports* 7:1–11

Lenth R (2018) Lsmeans: least-squares means.

McKee KL (2011) Biophysical controls on accretion and elevation change in Caribbean mangrove ecosystems. *Estuarine, Coastal and Shelf Science* 91:475–483

McKee KL, Cahoon DR, Feller IC (2007) Caribbean mangroves adjust to rising sea level through biotic controls on change in soil elevation. *Global Ecology and Biogeography* 16:545–556

McKee KL, Mendelsohn IA, Hester MW (1988) Reexamination of pore water sulfide concentrations and redox potentials near the aerial roots of *Rhizophora mangle* and *Avicennia germinans*. *American Journal of Botany* 75:1352–1359

McLeod AI (2015) CRAN—Package. Kendall rank correlation and Mann–Kendall trend test. Version 2.2.

McVoy CW (2011) Landscapes and hydrology of the predrainage Everglades. Gainesville, FL: University Press of Florida

Middleton B, Johnson D, Roberts B (2015) Hydrologic remediation for the Deep-water horizon incident drove ancillary primary production increase in coastal swamps. *Ecohydrology* 8:838–850

Morris JT, Sundareshwar PV, Nietz CT, Kjerfve B, Cahoon DR (2002) Responses of coastal wetlands to rising sea level. *Ecology* 83: 2869–2877

National Academies of Sciences, Engineering, and Medicine (2018) Progress Toward Restoring the Everglades: The Seventh Biennial Review.

Neubauer SC (2008) Contributions of mineral and organic components to tidal freshwater marsh accretion. *Estuarine, Coastal and Shelf Science* 78:78–88

Neubauer SC, Miller WD, Anderson IC (2000) Carbon cycling in a tidal freshwater marsh ecosystem: a carbon gas flux study. *Marine Ecology Progress Series* 199:13–30

Neubauer SC (2013) Ecosystem responses of a tidal freshwater marsh experiencing saltwater intrusion and altered hydrology. *Estuaries and Coasts* 36: 491–507

Newman S, Grace JB, Koebel JW (1996) Effects of nutrients and hydroperiod on *Typha*, *Cladium*, and *Eleocharis*: implications for Everglades restoration. *Ecological Applications* 6:774–783

Newman S, Osborne TZ, Hagerthey SE, Saunders C, Rutcher K, Schall T, Reddy KR (2017) Drivers of landscape evolution: multiple regimes and their influence on carbon sequestration in a sub-tropical peatland. *Ecological Monographs* 87:578–599

Newman S, Reddy KR (1993) Alkaline phosphatase activity in the sediment-water column of a hypereutrophic Lake. *Journal of Environmental Quality* 22:832–838

Noe GB, Childers DL (2007) Phosphorus budgets in Everglades wetland ecosystems: the effects of hydrology and nutrient enrichment. *Wetlands Ecology and Management* 15:189–205

Noe GB, Childers DL, Edwards AL, Gaiser E, Jayachandran K, Lee D, et al. (2002) Short-term changes in phosphorus storage in an oligotrophic Everglades wetland ecosystem receiving experimental nutrient enrichment. *Biogeochemistry* 59:239–267

Nyman JA, Walters RJ, Delaune RD, Patrick WH (2006) Marsh vertical accretion via vegetative growth. *Estuarine, Coastal and Shelf Science* 69:370–380

Osland MJ, Enwright NM, Day RH, Gabler CA, Stagg CL, Grace JB (2016) Beyond just sea-level rise: considering macroclimatic drivers within coastal wetland vulnerability assessments to climate change. *Global Change Biology* 22:1–11

Parsons CT, Rezanezhad F, O'Connell DW, Van Cappellen P (2017) Sediment phosphorus speciation and mobility under dynamic redox conditions. *Biogeoosciences* 14:3585–3602

Pinheiro J, Bates D, DebRoy S, Sarkar D (2018) NLme: linear and nonlinear mixed effects models.

Pivnickyová B, Rejmáková E, Snyder JM, Šantrúčková H (2010) Heterotrophic microbial activities and nutritional status of microbial communities in tropical marsh sediments of different salinities: the effects of phosphorus addition and plant species. *Plant and Soil* 336:49–63

Ratajczak Z, D'Odorico P, Collins SL, Bestelmeyer BT, Isbell FI, Nippert JB (2017) The interactive effects of press/pulse intensity and duration on regime shifts at multiple scales. *Ecological Monographs* 87:198–218

Reddy KR, Kadlec RH, Flagg E, Gale PM (1999) Phosphorus retention in streams and wetlands: a review. *Critical Reviews in Environmental Science and Technology* 29:83–146

Richardson CJ, Ferrell GM, Vaithianathan P (1999) Nutrient effects on stand structure, resorption efficiency, and secondary compounds in everglades sawgrass. *Ecology* 80:2182–2192

Ross MS, Meeder JF, Sah JP, Ruiz RL, Telesnicki GJ (2000) The southeast Saline Everglades revisited: 50 years of coastal vegetation change. *Journal of Vegetation Science* 11:101–112

Santander Meteorology Group (2012) CRAN—Package fume. Version 1.0.

Sarkar SK, Kominoski JS, Gaiser EE, Scinto LJ, Rudnick DT (2020) Quantifying effects of increased hydroperiod on wetland nutrient concentrations during early phases of freshwater restoration of the Florida Everglades. *Restoration Ecology* 28:1561–1573

Schindler DE, Kitchell JF, He X, Carpenter SR, Hodgson JR, Cottingham KL (1993) Food web structure and phosphorus cycling in lakes. *Transactions of the American Fisheries Society* 122:756–772

Sen PK (1968) Estimates of the regression coefficient based on Kendall's tau. *Journal of the American Statistical Association* 63:1379–1389

Servais S, Kominoski JS, Charles SP, Gaiser EE, Mazzei V, Troxler TG, Wilson BJ (2019) Saltwater intrusion and soil carbon loss: testing effects of salinity and phosphorus loading on microbial functions in experimental freshwater wetlands. *Geoderma* 337:1291–1300

Servais S, Kominoski JS, Coronado-Molina C, Bauman L, Davis SE, Gaiser EE, et al. (2020) Effects of saltwater pulses on soil microbial enzymes and organic matter breakdown in freshwater and brackish coastal wetlands. *Estuaries and Coasts* 43:814–830

Solohin E, Widney SE, Craft CB (2020) Declines in plant productivity drive loss of soil elevation in a tidal freshwater marsh exposed to saltwater intrusion. *Ecology* 101:e03148

Solórzano L, Sharp JH (1980) Determination of total dissolved phosphorus and particulate phosphorus in natural waters. *Limnology and Oceanography* 25:754–758

Søndergaard M, Jensen PJ, Jeppesen E (2001) Retention and internal loading of phosphorus in shallow, eutrophic lakes. *The ScientificWorld Journal* 1:427–442

Stagg CL, Baustian MM, Perry CL, Carruthers TJ, Hall CT (2018) Direct and indirect controls on organic matter decomposition in four coastal wetland communities along a landscape salinity gradient. *Journal of Ecology* 106:655–670

Suding KN, Gross KL, Houseman GR (2004) Alternative states and positive feedbacks in restoration ecology. *Trends in Ecology & Evolution* 19:46–53

Theil H (1992) A rank-invariant method of linear and polynomial regression analysis. Pages 345–381. In: Raj B, Koerts J (eds) Henri Theil's contributions to economics and econometrics: econometric theory and methodology. Advanced Studies in Theoretical and Applied Econometrics Springer Netherlands, Dordrecht

Tully K, Gedan K, Epanchin-Niell R, Strong A, Bernhardt ES, BenDor T, et al. (2019) The invisible flood: the chemistry, ecology, and social implications of coastal saltwater intrusion. *Bioscience* 69:368–378

Watson CS, White NJ, Church JA, King MA, Burgette RJ, Legresy B (2015) Unabated global mean sea-level rise over the satellite altimeter era. *Nature Climate Change* 5:565–568

Weston NB, Vile MA, Neubauer SC, Velinsky DJ (2011) Accelerated microbial organic matter mineralization following salt-water intrusion into tidal freshwater marsh soils. *Biogeochemistry* 102:135–151

Wilson BJ, Mortazavi B, Kiene RP (2015) Spatial and temporal variability in carbon dioxide and methane exchange at three coastal marshes along a salinity gradient in a northern Gulf of Mexico estuary. *Biogeochemistry* 123:329–347

Wilson BJ, Servais S, Charles SP, Mazzei V, Gaiser EE, Kominoski JS, Richards JH, Troxler TG (2019) Phosphorus alleviation of salinity stress: effects of saltwater intrusion on an Everglades freshwater peat marsh. *Ecology* 100:e02672

Wilson BJ, Servais S, Mazzei V, Kominoski JS, Hu M, Davis SE, et al. (2018) Salinity pulses interact with seasonal dry-down to increase ecosystem carbon loss in marshes of the Florida Everglades. *Ecological Applications* 28:2092–2108

Yue S, Hashino M (2003) Long term trends of annual and monthly precipitation in Japan. *Journal of the American Water Resources Association* 39:587–596

Zhao J, Malone SL, Oberbauer SF, Olivas PC, Schedlbauer JL, Staudhammer CL, Starr G (2019) Intensified inundation shifts a freshwater wetland from a CO₂ sink to a source. *Global Change Biology* 25:3319–3333

Supporting Information

The following information may be found in the online version of this article:

Supplement S1. Ecosystem metabolism

Coordinating Editor: Siobhan Fennessy

Received: 31 December, 2020; First decision: 28 April, 2021; Revised: 31 July, 2021; Accepted: 10 August, 2021