# Warming during maternal generations delays offspring germination in native and nonnative species

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#### 4 ABSTRACT

As environmental conditions shift due to global warming and other human-caused environmental 5 6 changes, plastic responses in phenological traits like germination or flowering time may become increasingly important. While phenological plasticity is a common response to global warming, 7 with many populations exhibiting earlier germination or flowering in warmer years, warming 8 9 may also result in transgenerational plasticity, especially on early life stages. In other words, seeds produced by mothers inhabiting warmer environments may germinate faster (or slower) 10 than seeds produced by mothers inhabiting ambient or cooler environments. Here, we use seeds 11 collected from a field warming experiment to examine how germination and early growth differ 12 in response to ambient vs. warmed (+3°C) temperatures experienced by both maternal and 13 14 offspring generations. Because nonnative species are often more phenotypically plastic than native species and because a variety of life history traits and environmental factors affect the 15 evolution of both within- and transgenerational plasticity, we include multiple invasive and 16 17 native plant species in our study. On average, warming experienced during maternal generations delayed germination by ~0.2 days/°C, although species varied in the magnitude of response. In 18 19 contrast, warming during the offspring generation tended to advance germination by  $\sim 0.1$ 20 days/°C. Nonnative species demonstrated higher germination success than native species, but we detected no differences in germination timing between native and nonnative species or that 21 22 native and nonnative species differed in either within- or transgenerational plasticity, although 23 species (independent of native status) did exhibit differing degrees of within- and

transgenerational plasticity in germination timing and early growth. This study suggests that
temperatures experienced by maternal plants can influence their offspring's germination
phenology, potentially even more so than temperatures experienced in the offspring's immediate
environment.

**KEY WORDS:** biological invasions, climate change, germination, maternal effects, plasticity

#### **30 INTRODUCTION**

Phenotypic plasticity (i.e., the ability to vary in phenotype under different environmental 31 conditions; Pigliucci [2008]) may allow species to express advantageous phenotypes across a 32 broad range of environmental conditions (Baker 1965). Because phenotypic plasticity is a major 33 response to global warming, it potentially contributes to species success under future 34 35 environmental conditions (Matesanz et al. 2010, Merilä and Hendry 2014). For instance, increased allocation to root mass increases water acquisition under drought (Sultan and Bazzaz 36 1993; Heschel et al. 2004), and advancing phenology under increasing temperature or nutrient 37 availability can help many plants escape stressful conditions or increase reproduction (Cohen 38 1976, Menzel et al. 2006, Power et al. 2006, Franks et al. 2007, Gugger et al. 2015, 39 Lustenhouwer et al. 2017). Thus, (within-generational) plastic responses may be beneficial for 40 plant performance under changing environments and reduce potential fitness consequences of 41 global change (Hendry et al. 2008, Nicotra et al. 2010, Lázaro-Nogal et al. 2015). 42 43 Transgenerational plasticity (i.e., the influence of the parental generation's environment on offspring phenotypes), like within-generational plasticity, can affect fitness (survival and 44 fecundity) (Uller 2008, Snell-Rood 2013, Vayda et al. 2018) and population persistence 45 46 (Donelan et al. 2020). For example, transgenerational plasticity increases desiccation tolerance in dog ticks (Yoder et al. 2006), drought tolerance in *Impatiens capensis* and *Polygonum persicaria* 47 48 (Riginos et al. 2007, Sultan et al. 2009, Herman and Sultan 2011, Herman et al. 2012), thermal 49 tolerance in minnows and sticklebacks (Salinas and Munch 2012, Shama and Wegner 2014), dispersal in marine bryzoans (Burgess and Marshall 2011), and egg production in butterflies 50 51 (Steigenga and Fischer 2007). Transgenerational plasticity may provide a more rapid response to 52 novel environmental conditions than within-generational plasticity because maternal plants can

provision their offspring in a way that minimizes the stress their offspring experience (Donohue
and Schmitt 1998, Mousseau and Fox 1998, Dyer et al. 2010). In contrast, within-generational
plasticity is inherently delayed as species sense a cue and respond appropriately, so offspring still
experience stress (Weinig 2000, Chevin et al. 2010).
Both transgenerational plasticity and within-generational plasticity can affect the same
trait such that the offspring's phenotype depends on both the offspring environment and the
environment their parents experienced. In plants, for example, germination often demonstrates

60 both within- and transgenerational plasticity to temperature and light conditions experienced

61 during maternal and offspring generations in species including *Plantago lanceolata* (Lacey 1996,

62 Lacey and Herr 2000), Leucanthemopsis alpina (Bernareggi et al. 2016), Brassica rapa

63 (Wadgymar et al. 2018), and *Arabidopsis thaliana* (Blödner et al. 2007, Whittle et al. 2009,

64 Donohue 2009, Auge et al. 2017), and in *Campanula americana* seeds sown in light gaps had

65 greater germination and survival than seeds sown in shade, but only if their mother also grew in a

66 light gap (Galloway and Etterson 2007). Although transgenerational plasticity can be

67 maladaptive if environmental stress causes inferior offspring production or if parental and

offspring environments are not well-matched (Stearns 1992, Marshall and Uller 2007, Munday et

al. 2013, Munday 2014), adaptive transgenerational plasticity is predicted to evolve if the

70 parental environment is predictive of the offspring environment (Kingsolver and Huey 1998,

Herman et al. 2012, 2014, Burgess and Marshall 2014, Leimer and McNamara 2015, Colicchio

and Herman 2020). Transgenerational plasticity therefore might promote rapid responses (and

73 possible adaptation) to directional environmental change if parents can accurately convey

information about novel conditions to their offspring (Donelson et al. 2018, Bell and Hellmann

75 2019).

Although phenotypic plasticity is a common response to global change, the extent of 76 plastic responses often varies across species (Henn et al. 2018). A variety of life-history traits 77 such as self-compatibility may affect the evolution of within- and transgenerational plasticity 78 (Dury and Wade 2019), and several hypotheses speculate that phenotypic plasticity may 79 facilitate biological invasions (Sultan 2001, Wolkovich and Cleland 2011), enabling them to 80 81 colonize and establish in novel climates (Schlichting and Levin 1986). Indeed, nonnative species often demonstrate greater within-generational plasticity than native species (Richards et al. 2006; 82 83 Davidson et al. 2011; but see Godoy et al. 2011). For example, nonnative species show greater increases in survival and growth than native species in response to nutrient addition and high 84 light (Milberg et al. 1999, Leishman and Thomson 2004, Gleason and Ares 2004, Brock et al. 85 2005) and stronger advances in leaf-out and flowering time in response to warming (Crawley et 86 al. 1996, DeFalco et al. 2007, Resasco et al. 2007, Xu et al. 2007, Godoy et al. 2009, Willis et al. 87 2010, Pearson et al. 2012, Wolkovich et al. 2013, Wilsey et al. 2015, 2018, Zettlemoyer et al. 88 89 2019). Such within-generational plasticity likely affects a species' establishment and spread, i.e., its "invasiveness" (van Kleunen and Richardson 2007), and could contribute to invasion success 90 under global warming. 91

Similar to the explanations for greater within-generational plasticity in nonnative species, we hypothesize that nonnative species may also express greater transgenerational plasticity than native species for two reasons. First, transgenerational plasticity can more rapidly increase stress tolerance and fitness in offspring than within-generational plasticity. This could promote establishment of nonnative species in stressful habitats or result in more rapid population spread for widespread nonnatives (i.e., invasive species). For example, in nonnative *Cyperus esculentus*, maternal responses to nutrient-poor soil conditions promoted greater propagule dispersal in the

next generation (Dyer et al. 2010). Second, nonnative species are more often self-compatible
than native species (Baker 1955, Razanajatovo et al. 2016), and selfing makes transgenerational
plasticity more likely to evolve (Dury and Wade 2019). Together, this suggests that nonnative
species may be particularly likely to have greater transgenerational plasticity.

Here, we use seeds collected from a field warming experiment at the W.K. Kellogg 103 104 Biological Station (KBS) to examine transgenerational and within-generation plasticity in response to warming in a suite of grassland species. Although we use the general term 105 106 "transgenerational plasticity", which includes non-genetic inheritance, parental effects, carry-107 over effects, intergenerational effects, seed provisioning, and epigenetic transmission (Donelson et al. 2018), we specifically test the effect of the maternal environment on offspring phenotypes, 108 or maternal effects (Roach and Wulff 1987). We focus on early phenological and growth traits 109 because both within- and transgenerational phenological plasticity (i.e., shifts in the timing of 110 life-history events like germination and flowering) may be especially important responses for 111 112 species success under climate change (Jump and Peñuelas 2005; Merilä and Hendry 2014; Bonamour et al. 2019) and because germination timing can be plastic (see above) and is linked to 113 plant fitness (Kalisz 1986, Donohue 2002, Donohue et al. 2010, Cochrane et al. 2015, Leverett et 114 115 al. 2018). Similarly, offspring growth often demonstrates transgenerational plasticity (Agrawal 2001; Galloway and Etterson 2007; Sultan et al. 2009; Latzel et al. 2010), potentially providing 116 117 offspring an advantage in stressful conditions (Moles and Westoby 2006; Herman and Sultan 118 2011). Because nonnative species exhibit greater plasticity in flowering time than native species in this (Zettlemoyer et al. 2019) and other systems (Grman and Suding 2010, Wainwright and 119 120 Cleland 2013, Balshor et al. 2017, Wilsey et al. 2018) and to test whether patterns of within- and 121 transgenerational plasticity in germination and early growth are generalizable across native and

nonnative species, we include multiple native and nonnative species in this study. This work
extends studies investigating transgenerational plasticity in germination timing in response to
temperature, which has largely been tested in single species, by examining whether
transgenerational plasticity to warming is consistent across species and whether it differs
between native and nonnative taxa. We ask: Do germination and early growth demonstrate
within- or transgenerational plasticity and if so, do native and nonnative species differ in their
within- or transgenerational responses to warming temperatures?

#### **129 MATERIALS & METHODS**

To assess the roles of within- and transgenerational plasticity and their interaction on 130 germination and early growth of grassland species in response to warming, we planted seeds 131 from maternal plants that had been grown under an experimental warming simulation in the field 132 into warmed and ambient growth chamber conditions in a full factorial design (maternal 133 temperature environment  $\times$  offspring temperature environment). Although each generation 134 135 experienced different degrees of environmental control (i.e., maternal plants were grown in the field while offspring were grown in potting soil in a growth chamber), this design is common for 136 testing maternal effects on germination (Galloway and Etterson 2007). 137

*Maternal temperature environment*— The simulated warming array in the field uses infrared heaters to elevate temperatures 3°C above ambient temperatures, matching regional predictions for climate warming in this area by the end of the 21<sup>st</sup> century (0.3°C-4.8°C) (Stocker et al. 2013). Warmed plots also have lower soil moisture (percent water content) than ambient plots (warming  $\chi^2_{1,6}$ =34.02, p<0.0001; ambient = 16.2% vs. warmed = 9.31% water). The array has run over the growing season (April-October) since 2008. In spring 2012, we planted 52 species (25 native and 27 nonnative) into the old field community in each plot (n=3

replicates/species/plot  $\times$  4 plots/warming treatment), although only the 24 species (13 native and 145 11 nonnative) that successfully produced viable seeds in both warming treatments are used here 146 (see below). Study species were forb and grass species found in old fields or grasslands 147 (Schultheis et al. 2015). We define native species as species naturally occurring in Michigan 148 prior to European settlement and nonnative species as species introduced to Michigan from 149 150 outside the United States. For a full description of the simulated warming experiment, see Zettlemoyer et al. (2019). In this system, nonnative species exhibit advanced flowering 151 152 phenology relative to native species and accelerate flowering in response to warming more so 153 than native species, although prior studies did not investigate other phenological stages like germination timing (Zettlemoyer et al. 2019). However, because the study species are perennial, 154 we focused on early growth stages and did not grow plants to flowering for this experiment (see 155 below). 156

To determine which plants produced viable seeds, we conducted germination trials using 157 158 seeds from plants collected from the simulated warming experiment in the greenhouse (n=41 species; 21 native and 20 nonnative) (greenhouse conditions were 29°C with a 16-hour 159 photoperiod). Seeds were collected from the field in 2013 and stored in a cool, dark cabinet until 160 161 2019. However, we did not account for variable dormancy requirements among species (e.g., stratification), potentially limiting germination and restricting our experiment to those species 162 163 with lax germination requirements. We planted seeds in low-nutrient potting media (Sunshine 164 Mix LP5) and monitored daily seedling emergence over six weeks. 24 species (13 native and 11 nonnative) successfully germinated in the greenhouse. We included all species with at least 20% 165 166 germination in an attempt to boost the number of native species included in our study. For each 167 of those 24 species, we selected seeds from 3-5 individuals grown under ambient field maternal

conditions and another 3-5 individuals grown under warmed field maternal conditions. When
possible, we elected to use seeds from maternal plants from different field plots. We ended up
with 116 total maternal plants spanning the 24 species (3-5 individuals per maternal environment
per species).

*Offspring temperature environment*— To set growth chamber/offspring temperature 172 173 conditions, we collected daily maximum and minimum temperatures from 01-Apr-2013 thru 30-June-2013 (approximately the time period when germination occurs in the field) from the 174 175 CLIMOD database (http://climod2.nrcc.cornell.edu/). We used these day and night temperature 176 extremes observed in the field dataset to program daily temperature curves for each growth chamber (Percival Scientific, Perry, IA, USA), with the warmed chamber set to be consistently 177 3°C warmer than the ambient chamber. Day length was set at 14 hours, roughly matching 178 photoperiod in the field. This design allows us to separate effects of maternal temperatures (i.e., 179 temperatures experienced during seed maturation in the field) vs. offspring temperatures (i.e., 180 181 temperatures experienced post-dispersal in the growth chamber) (Burghardt et al. 2015). We planted three seeds from each maternal plant into separate conetainers (SC10 Ray 182 Leach conetainers, Stuewe & Sons Inc., Tangent, OR, USA) filled with low-nutrient potting 183 184 media (Sunshine Mix LP5, BFG Supply, Kalamazoo, MI, USA) and placed them into ambient and warmed growth chambers (n=3 replicates  $\times$  116 maternal plants  $\times$  2 chamber temperatures = 185 186 696 seedlings). We rotated trays between chambers every three days to minimize chamber 187 effects and watered as needed to maintain similar moisture levels between treatments. Germination (indicated as cotyledon emergence) was recorded daily. We measured seedling 188 189 height (the longest leaf; cm) at the end of the experiment. 190 Data analysis

To examine whether native and nonnative species' germination and early growth 191 (seedling height) demonstrate within- or transgenerational plasticity in response to temperature, 192 we used generalized linear mixed models fit in the lme4 package in R v.3.0.2 (Bates et al. 2015, 193 R Core Development Team 2015). We conducted three models with three separate response 194 variables: (1) germination success (1=yes, 0=no; binomial distribution), (2) days to germination 195 196 (negative binomial distribution for overdispersed count data; Lindén and Mäntyniemi 2011), excluding ungerminated seeds, and (3) seedling height (cm; Gaussian distribution). For 197 198 germination success, we included maternal temperature environment ( $E_M$ ; ambient vs. warmed 199 field conditions), offspring temperature environment (E<sub>0</sub>; ambient vs. warmed chamber conditions), status (native vs. nonnative), and their interactions as predictor variables. We 200 included species (nested within status) and field plot (nested within maternal environment) as 201 random effects. For days to germination and seedling height, we only included species that 202 203 successfully germinated in all temperature combinations (n=3 native and 5 nonnative species). 204 Due to resulting low sample sizes, we could not test interactions between maternal and offspring environments. We instead ran two models for each response variable, one for (i) 205 transgenerational plasticity and another for (ii) within-generational plasticity. Models for (i) 206 207 transgenerational plasticity included status,  $E_M$ , and status  $\times E_M$  as predictor variables and species (nested within status) and plot (nested within  $E_M$ ) as random effects. Models for (ii) 208 209 within-generational plasticity were identical but included  $E_0$  instead of  $E_M$ . For seedling height 210 models, because seedling height depends on germination time, we first regressed height against days to germination for each species. We subsequently used the residuals as the response 211 212 variable for the third model, thereby removing variation in height due to differences in

213 germination phenology. We provide the results of models using height as a response variable in214 Appendix Table A1 (results are qualitatively similar).

215 We used similar models to examine variation in within- and transgenerational plasticity in germination success, germination timing, and seedling height among species regardless of 216 status. For germination success, we included species, E<sub>M</sub>, E<sub>O</sub>, and their interactions as predictor 217 218 variables, with plot (nested in E<sub>M</sub>) as a random effect. For days to germination and seedling height (residuals), we again only included the eight species that successfully germinated in all 219 220 temperature combinations. Like above, we ran two models each for days to germination and 221 seedling height, one for (i) transgenerational and another for (ii) within-generational plasticity. These models, respectively, included (i) species,  $E_M$ , and species  $\times E_M$  and (ii) species,  $E_0$ , and 222 species  $\times$  E<sub>O</sub> as predictor variables. We included field plot (nested in E<sub>M</sub>) as a random effect in 223 all models. Following significant interactions between species  $\times E_M$  and species  $\times E_O$  for days to 224 225 germination and seedling height (see Results), we conducted individual species models. For 226 species with sufficient sample sizes, we examined the effects of  $E_M$ ,  $E_0$ , and the interaction of  $E_M \times E_O$  on days to germination and seedling height. For species with low sample sizes where 227  $E_M \times E_O$  could not be estimated, we removed the interaction and only tested the main effects of 228 229 maternal and offspring temperatures. Field plot (nested in  $E_M$ ) was included as a random effect in all individual species models. Individual species models were not conducted for germination 230 231 success because we detected no evidence for species-specific responses to temperature (see 232 Results).

Finally, to investigate whether and how plasticity in germination timing is associated with plasticity in other phenological stages, we compared both within- and transgenerational plasticity in germination timing found here to within-generational plasticity in flowering time

from Zettlemover et al. (2019) (again using only the eight species that germinated in every 236 temperature combination). Within-generational phenological plasticity was calculated as the 237 difference in mean offspring phenotype ( $\bar{X}$ ; i.e., germination timing or flowering time) when 238 offspring experienced warmed vs. ambient temperatures (Valladares et al. 2006): 239 240 Within – generational phenological plasticity =  $X_{offspring.warmed} - X_{offspring.ambient}$ Transgenerational phenological plasticity was calculated as the difference in mean offspring 241 phenotype when maternal plants experienced warmed vs. ambient temperatures: 242 243 Transgenerational phenological plasticity =  $X_{maternal.warmed} - X_{maternal.ambient}$ We used differences for phenological plasticity because proportional responses for 244 phenological estimates would cause later flowering individuals to appear to be less plastic when 245 they could in fact be advancing flowering time as much as early flowering individuals. 246 We calculated within-and transgenerational plasticity in early growth as the proportional 247 difference in mean offspring phenotype (seedling height) when offspring or maternal plants 248 (respectively) experienced warmed vs. ambient temperatures: 249 Within – generational plasticity in early growth =  $\frac{X_{offspring.warmed} - X_{offspring.ambient}}{X_{offspring.ambient}}$ , and 250  $Transgenerational \ plasticity \ in \ early \ growth = \frac{X_{maternal.warmed} - X_{maternal.ambient}}{X_{maternal.ambient}}$ 251 We averaged responses to temperature within generations because we detected no 252 interactions between maternal and offspring temperatures (see Results). We then estimated 253 254 Pearson's correlations among within-generational plasticity in flowering time, withingenerational plasticity in germination timing, transgenerational plasticity in germination timing, 255 within-generational plasticity in early growth (seedling height), transgenerational plasticity in 256 early growth, and mean early growth. 257

258 **RESULTS** 

Nonnative species tended to be approximately twice as likely to germinate than native species (status  $\chi^2_{1,0.005}$ =3.30, p=0.06; Table A2; Fig. 1). We detected no evidence that temperatures experienced by either maternal or offspring generations affected germination success in native or nonnative species or across species (all E<sub>M</sub> × status {or species} and E<sub>O</sub> × status {or species} interactions, p>0.5, Table A2-A3).

264 Species varied in their germination timing responses to warming during maternal generations (E<sub>M</sub> × species  $\chi^2_{7,202}$ =17.56, p=0.02; Table A3). On average, warmer temperatures 265 experienced during the maternal generation delayed germination by  $0.20 \pm 0.1$  days/°C relative 266 to ambient temperatures (E<sub>M</sub>  $\chi^{2}_{1,5.86}$ =3.58, p=0.05; Fig. 2a). Three native species, Achillea 267 millefolium, Panicum virgatum, and Coreopsis lanceolata, and two nonnative species, Dactylis 268 glomerata and Gaillardia pulchella, demonstrated this pattern, with all other species not 269 responding significantly to maternal temperature environments (E<sub>M</sub>: AM  $\chi^{2}_{1,13}$ =3.09, p=0.07; CL 270  $\chi^{2}_{1,3,24}$ =5.91, p=0.01; PV  $\chi^{2}_{1,5}$ =4.09, p=0.04; DG  $\chi^{2}_{1,16,81}$ =4.65, p=0.03; GP  $\chi^{2}_{1,3}$ =5.66, p=0.02; 271 Table 1; Fig. 2b). 272

In contrast, germination timing responses to offspring warming were much weaker and 273 did not vary substantially across species (E<sub>0</sub>: effect size =  $-0.11 \pm -0.10$  days/°C earlier under 274 warmed relative to ambient temperatures;  $\chi^{2}_{1,190,37}=1.14$ , p=0.2; Table A2; Fig. 2c; species × E<sub>0</sub> 275  $\chi^{2}_{7,202}$ =7.31, p=0.4; Table A3). Native *Coreopsis lanceolata* and nonnative *Dactylis glomerata*, 276 Hypericum perfoliatum, and Phleum pratense germinated earlier under warmed relative to 277 278 ambient offspring temperatures, but no other species responded significantly to offspring temperatures (EO: CL  $\chi^2_{1,2.01}$ =11.17, p=0.0008; DG  $\chi^2_{1,47.74}$ =2.15, p=0.1; HP  $\chi^2_{1,18.27}$ =4.15, 279 p=0.04; PP  $\chi^{2}_{1,44.73}$ =2.44, p=0.1; Table 1; Fig. 2d). We detected no consistent differences in 280

germination phenology between native vs. nonnative species in response to temperaturesexperienced by either generation (Table A2).

Warming during both maternal and offspring generations affected plant height, although 283 the direction of effects varied across species (species  $\times$  E<sub>M</sub>:  $\chi^{2}_{7,184.68}$ =32.82, p<0.0001; species  $\times$ 284 E<sub>0</sub>:  $\chi^{2}_{7,184,22}$ =13.57, p=0.05; Table A3; Fig. 3). On average, seedlings tended to grow more 285 286 rapidly than expected under warmed relative to ambient offspring environments (E<sub>0</sub>:  $\chi^{2}_{1,192.93}$ =3.23, p=0.07; Fig. 3c; Table A2), with three nonnative species, *Dactylis glomerata*, 287 Gaillardia pulchella, and Poa compressa, growing significantly more rapidly under warmed 288 offspring conditions (DG:  $\chi^{2}_{1,53}$ =6.41, p=0.01; GP  $\chi^{2}_{1,3}$ =6.35, p=0.01; PC  $\chi^{2}_{1,41.09}$ =5.55, p=0.02; 289 Table 1; Fig. 3d). However, native C. lanceolata and nonnative Hypericum perforatum grew 290 more slowly under warmed temperatures (CL  $\chi^{2}_{1,2.10}=22.59$ , p<0.0001; HP  $\chi^{2}_{1,18.72}=7.29$ , 291 p=0.007).. Seedling height responses to maternal temperature conditions were weaker, with only 292 293 native Coreopsis lanceolata responding significantly to the maternal temperature environment. It 294 grew more slowly than expected (i.e., seedlings were shorter than expected after controlling for differences in height due to variation in germination phenology) under ambient relative to 295 warmed maternal environments ( $\chi^{2}_{1,3,40}=78.65$ , p<0.0001). As with germination timing, we 296 detected no evidence that native and nonnative taxa differed in within- or transgenerational 297 plasticity in height. 298

299 Correlations among plasticity in germination and flowering phenology

Within-generational plasticity in germination timing, transgenerational plasticity in germination timing, and plasticity in flowering time were not correlated (Fig. A2), with one possible exception. Within-generational plasticity in germination timing was negatively correlated with within-generational plasticity in early-growth (r=-0.84, p=0.006; Fig A2f),

304 suggesting that species that are more plastic in their germination timing in response to

305 temperatures experienced as offspring are less plastic in their growth rates.

## 306 **DISCUSSION**

## 307 *Effects of warming temperatures on germination phenology*

Consistent with the few prior investigations of maternal warming, we found that warming 308 309 experienced during the maternal generation, on average, delayed germination in the offspring generation. For example, warming during maternal generations results in delayed bud burst in 310 311 *Populus nigra* (Dewan et al. 2018), and shorter winters as a result of warming temperatures can 312 also delay offspring germination (Walck et al. 2011). Similarly, stressful or unfavorable conditions during maternal generations delay germination in species such as Arabidopsis 313 thaliana (Donohue et al. 2005), Pinus pinaster (Cendán et al. 2013), and Banksia species 314 (Cochrane et al. 2014) (but see Moriuchi et al. 2016, Walter et al. 2016). If warmed 315 environments are similarly stressful, then delayed germination in response to warming 316 317 temperatures experienced by maternal plants may reflect this stress response. Such responses could be adaptive and a strategy to avoid future stressful conditions (i.e., predictive plasticity or 318 predictive germination) (Cohen 1967, Gremer et al. 2016), if delayed germination reduces the 319 320 likelihood of experiencing conditions with catastrophic fitness effects (e.g., frost; Milbau et al. 2009). Delaying germination in stressful environments is also predicted to reduce the risk of all 321 322 seeds germinating into an unfavorable environment, thereby reducing temporal variation in 323 fitness (Clauss and Venable 2000, Simons 2011, Gremer and Venable 2014, Gremer et al. 2016). However, these hypotheses require future work investigating whether delayed germination in 324 325 response to warming parental environments corresponds with higher fitness under warming 326 temperatures.

In contrast to our finding of delayed germination in response to maternal warming, 327 warming during the offspring generation has minimal effects, but tended to act in the opposite 328 direction by advancing germination phenology as might be expected given many species' 329 temperature cues for germination. The three species responding significantly to warming during 330 the offspring generation all responded by advancing germination. Other studies frequently detect 331 332 advancing germination (Milbau et al. 2009, Zettlemoyer et al. 2017), leaf-out (Basler & Körner 2014, Geng et al. 2020) and flowering under warmer conditions (Fitter and Fitter 2002, 333 Parmesan and Yohe 2003, Thackeray et al. 2016). Earlier germination can provide earlier access 334 335 to resources like space, water, and light via priority effects (Wolkovich and Cleland 2011, Wainwright et al. 2012), allows for earlier growth relative to other species in the community 336 (Dickson et al. 2012, Fridley 2012, Wilsey et al. 2015), and can increase chances of surviving to 337 reproduction (Leverett et al. 2018) and enhance plant fitness (Verdú and Traveset 2005). 338 However, as climate change continues to alter local environmental conditions in a directional 339 340 manner, mismatched responses between maternal and offspring generations, as detected here, could reduce plant fitness by counteracting one another and minimizing the phenological 341 342 response.

343 *Correlations among plasticity in germination and flowering phenology* 

Overall, our results suggest that plasticity in one life-history stage does not predict plasticity in another. If plasticity in germination timing and flowering time are truly not correlated, these traits could evolve separately in response to local environmental conditions as opposed to representing a correlated response to warming temperatures (Burgess et al. 2007). However, multiple species demonstrated consistent patterns in phenological plasticity across lifehistory stages; *Coreopsis lanceolata, Hypericum perforatum, Dactylis glomerata,* and *Phleum* 

pratense all tended to advance flowering under warming (Zettlemoyer et al. 2019) and also 350 tended to advance germination timing under warmed offspring environments. Some of these 351 species also demonstrated transgenerational plasticity. For one species (P. virgatum), 352 germination responses to maternal warming were in the same direction as flowering responses to 353 warming temperatures (i.e., a tendency for delayed flowering and germination in response to 354 355 warming in both generations), but for the other (C. lanceolata), the transgenerational response opposed both within-generational germination plasticity and flowering plasticity to warming 356 (i.e., maternal warming delayed germination while offspring warming accelerated both 357 358 germination and flowering).

359 Differences between native and nonnative species

In contrast to previous studies detecting earlier and more plastic germination phenology 360 in nonnative species than natives in response to rising temperatures (Gerlach and Rice 2003, 361 Seabloom et al. 2003, Resasco et al. 2007, Xu et al. 2007, Abraham et al. 2009, Grman and 362 363 Suding 2010, Wainwright and Cleland 2013, Balshor et al. 2017, Wilsey et al. 2018; reviewed in Gioria and Pyšek 2017), we detected no differences in germination phenology between the native 364 and nonnative species studied here in response to either maternal or offspring temperatures, 365 366 although we had limited power (eight species) to detect such effects. Similar germination phenology between native and nonnative species has been observed in *Impatiens* species (Laube 367 368 et al. 2015) and between native vs. nonnative populations of *Hieracium pilosella* and *Hypericum* 369 perforatum (Beckmann et al. 2011). Other species characteristics beyond native or invasive status are also predicted to influence the evolution of transgenerational plasticity. For example, 370 371 theory predicts that self-incompatibility should correlate with lower transgenerational plasticity 372 (Dury and Wade 2019). Counter to this prediction, all four species that demonstrated

transgenerational plasticity in our study by delaying germination under maternal warming are
self-incompatible (Table A4). While a greater complement of species would be needed to
rigorously test the association between mating system and transgenerational plasticity, it is
possible that the transgenerational plasticity observed here is a maladaptive stress response,

377 rather than the adaptive transgenerational plasticity modeled in Dury and Wade (2019).

378 Similar to the germination timing results, we also detected no evidence that nonnative 379 species exhibited more plastic growth responses to warming. However, three of the six nonnative 380 species studied here (*Dactylis glomerata, Gaillardia pulchella,* and *Poa compressa*)

demonstrated within-generational plasticity to warming temperatures via greater early growth.
While this result in part supports previous findings suggesting that invasive species' growth and
fitness increase under warming conditions (Parker-Allie et al. 2009; Verlinden and Nijs 2010;
Compagnoni and Adler 2014) potentially providing invasive species with an advantage under
climate change (Hellmann et al. 2008), it also suggests that these growth benefits are not
pervasive and that warming may lead to increased success of some, but certainly not all,
invasives.

Nonnative species had a higher probability of germination than native species regardless 388 389 of temperature, consistent with other studies comparing germination success in native vs. nonnative species (Colautti et al. 2006, Beckmann et al. 2011, Wainwright and Cleland 2013, 390 391 Balshor et al. 2017). On one hand, this could be due to harvesting time: native species flower 392 later in this system, and some of the latest flowering species (e.g., Aster pilosis and Solidago canadensis) likely had less-ripe seeds at collection. We checked for seed viability using 393 394 tetrazolium assays; similar percentages of seeds were viable in native vs. nonnative species (although nonnative species tended to have more viable seeds; Table A4; status  $\chi^2_{1,34}=2.25$ , 395

p=0.14), suggesting that differences in seed viability did not influence our results. Alternatively, 396 this could be due to higher thresholds for breaking dormancy in native species, wherein seeds 397 require a cold period (i.e., vernalization) and an environmental cue that conditions are suitable 398 for germination and growth (Fenner and Thompson 2005, Baskin and Baskin 2014). Future 399 studies should account for species differences in requirements to break dormancy (e.g., 400 401 stratification, diurnal vs. nocturnal temperatures, photoperiod). We did not include a stratification treatment in this study, which could limit germination success (Munir et al. 2001). 402 403 However, as temperatures warm, native species may be less likely to experience sufficient vernalization (Murray et al. 1989, Schwartz and Hanes 2010, Cook et al. 2012, Fu et al. 2015). 404 Additionally, nonnative species may have broader germination requirements than native species 405 (Gioria and Pyšek 2017), so future work should test whether the invasive species have similarly 406 high germination success in field conditions. Ultimately, higher germination success in 407 nonnative species suggests that germination could increase nonnative species' population growth 408 rates relative to native species because germination plays an important role in determining 409 population growth rates (Picó 2012, Leverett et al. 2018). 410

411 *Conclusions* 

We find that maternal warming delays germination and that warmed temperatures experienced during the offspring generation tend to advance germination, although this latter pattern is weaker and not statistically significant. Our results indicate that temperatures experienced by maternal plants can impact their offspring's germination phenology, potentially even more so than temperatures experienced by the offspring themselves. This study furthers our understanding of both within- and transgenerational plasticity to temperature during early life-

history stages, but future work is needed to understand potential benefits or consequences ofdelayed germination under maternal warming.

## 420 DECLARATIONS

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- 428 **Conflicts of interest** The authors have no conflicts of interest to declare.

429 **Data accessibility** – Upon publication, all data for this study will be available as .csv files in the

430 Dryad Digital Repository: doi:10.5061/dryad.n2z34tmx9 (Zettlemoyer and Lau 2021).

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747 TABLES

Table 1. Species-specific models for the effect of maternal and offspring temperatures on 748 germination and early growth. Results of generalized linear mixed models (GLMMs) for 749 750 individual species' germination success (1=yes, 0=no; binomial distribution), (B) germination timing (days to germination; negative binomial distribution), and (C) height (residuals after 751 controlling for germination timing; Gaussian distribution). We included offspring environment 752 (E<sub>0</sub>; ambient vs. warmed chamber conditions), maternal environment (E<sub>M</sub>; ambient vs. warmed 753 field conditions), and their interaction as fixed predictor variables (where possible). Plot (nested 754 in maternal environment) was included as a random effect in each model. "--" indicates that a 755 parameter was not estimated due to low power. \*\*\*p≤0.0001, \*\*p≤0.01, \*p≤0.05, ·p≤0.1 (bold 756 757 indicates  $p \leq 0.1$ ).

Species	Germination timing $\chi^2$			Height (residuals) $\chi^2$		
Natives	Ео	Ем	E <sub>M</sub> × E <sub>0</sub>	Ео	Ем	E <sub>M</sub> × Eo
Arabis glabra (AG)	2e <sup>-04</sup>	3e <sup>-04</sup>	2e <sup>-04</sup>	0.00	3e <sup>-04</sup>	2e <sup>-04</sup>
Achillea millefolium (AM)	0.18	3.09 ·	1.44	0.00	0.00	0.25
Coreopsis lanceolata (CL)	11.18 ***	5.91 *	NA	22.59 ***	78.66 ***	NA
Penstemon hirsutus (PH)						
Panicum virgatum (PV)	0.03	4.09 *		0.00	0.79	
Nonnatives						
Bromus inermis (BI)						
Leucanthemum vulgare (LV)						
Centaurea maculosa (CM)						
Dianthus armeria (DA)	0.62	0.06	0.21	0.09	0.29	1.02
Dactylis glomerata (DG)	0.28	4.65 *	2.15	6.41 *	1.08	0.03
Gaillardia pulchella (GP)	0.32	5.66 *		6.35 *	0.45	

Hypericum perfoliatum (HP)	4.15 *	1.51		7.29 **	0.05	
Melilotus officinalis (MO)						
Poa compressa (PC)	0.25	0.03	0.11	5.55 *	0.19	0.01
Phleum pratense (PP)	2.44 ·	0.47	0.02	0.00	0.00	0.98

#### 759 FIGURE LEGENDS

**Figure 1.** Germination success (proportion of seeds germinated; least square means  $\pm$  standard

rror) under ambient vs. warmed  $(+3^{\circ}C)$  maternal environments (x-axis) and ambient (blue) vs.

762 warmed (orange) offspring environments in native (left) vs. nonnative (right) species.

**Figure 2.** Days to germination under ambient (blue) vs. warmed (+3°C; orange) (a) maternal

real environments or (c) offspring environments. (b) and (d) show species-specific response to

765 maternal and offspring temperatures, respectively. Native species are indicated with asterisks and

766 green lines while nonnative species are indicated with purple lines. An asterisk within a bracket

767 indicates that species differed in germination timing between ambient vs. warmed temperatures

768 (Tukey tests, a=0.05), where \*\*\*p<0.0001, \*p<0.05, §p<0.1. We provide sample sizes in the top 769 species panels.

**Figure 3.** Residuals of seedling height (cm; after removing variation due to days to germination)

under ambient (blue) vs. warmed (+3°C; orange) temperatures experienced during (a) maternal

or (c) offspring generations. (b) and (d) show species-specific response to maternal and offspring

temperatures, respectively. Positive residual values indicate more rapid growth (i.e., taller

seedlings) than expected based on germination timing. Native species are indicated with asterisks

and green lines while nonnative species are indicated with purple lines. An asterisk within a

bracket indicates that species differed in early growth between ambient vs. warmed temperatures

777 (Tukey tests, a=0.05), where \*\*\*p<0.0001, \*p<0.05, p<0.1.

# **FIGURES**

Figure 1.









