Zootaxa 4970 (2): 353–371
https://www.mapress.com/j/zt/

Article

ISSN 1175-5326 (print edition)
ZOOTAXA
ISSN 1175-5334 (online edition)

https://doi.org/10.11646/zootaxa.4970.2.8
http://zoobank.org/urn:lsid:zoobank.org:pub:A10E0734-85A7-4B44-AD45-63142A1CC7D1

Four new *Scopalina* from Southern California: the first Scopalinida (Porifera: Demospongiae) from the temperate Eastern Pacific

THOMAS L. TURNER
Ecology, Evolution, and Marine Biology Department, University of California, Santa Barbara, Santa Barbara, CA 93106
tlturner@ucsb.edu; https://orcid.org/0000-0002-1380-1099

Abstract

Sponges (phylum Porifera) are common inhabitants of kelp forest ecosystems in California, but their diversity and ecological importance are poorly characterized in this biome. Here I use freshly collected samples to describe the diversity of the order Scopalinida in California. Though previously unknown in the region, four new species are described here: *Scopalina nausicae* **sp. nov.**, *S. kuyamu* **sp. nov.**, *S. goletensis* **sp. nov.**, and *S. jali* **sp. nov.**. These discoveries illustrate the considerable uncharacterized sponge diversity remaining in California kelp forests, and the utility of SCUBA-based collection to improve our understanding of this diversity.

Key words: sponges, Scopalina, phylogeny, 28S, cox1

Introduction

The order Scopalinida is young: it was created in 2015 (Morrow & Cárdenas 2015). The independent evolution of this lineage, however, is old. Phylogenies based on ribosomal DNA place the order, together with the freshwater sponges, as the sister clade to all other extant orders in the subclass Heteroscleromopha (Morrow *et al.* 2013). This interesting phylogenetic position motivates further investigation of this group. The order is comprised of only two genera, *Scopalina* and *Svenzea*, though genetic data suggest *Stylissa flabelliformis* may also be in the order (other genotyped *Stylissa* cluster with the order Agelisida (Erpenbeck *et al.* 2006; Morrow *et al.* 2012; Morrow & Cárdenas 2015)). Scopalinida have been found to be sources for novel anti-microbial or anti-tumor compounds, which further motivates efforts to characterize their diversity (Avilés *et al.* 2013; Vicente *et al.* 2015; Wei *et al.* 2007).

Nearly all Scopalinida are described from warm waters in the Mediterranean, Caribbean, South-West Pacific, and Madagascar (van Soest *et al.* 2019). The only previous exceptions to this pattern are the two most recently described species, from the Falkland Islands, which were found by hand-collecting sponges while SCUBA diving (Goodwin *et al.* 2011). Collecting by hand has been shown to be a productive way to discover new sponges from rocky areas in the shallow subtidal (Goodwin *et al.* 2011; Goodwin & Picton 2009), but past sponge surveys in Southern California have primarily been conducted via dredging or by collecting in the intertidal zone (Bakus & Green 1987; Green & Bakus 1994; de Laubenfels 1932; Sim & Bakus 1986). As a result, some common sponges found in California kelp forests — which occur on shallow hard-bottom substrate — remain unknown to science. Kelp forests in California are experiencing rapid changes due to anthropogenic impacts, and considerable work is focused on understanding kelp forest ecology to better predict and/or mitigate these impacts (Caselle *et al.* 2018; Castorini *et al.* 2018; Eger *et al.* 2020; Miller *et al.* 2018; Reed *et al.* 2016). The roles that sponges play in this ecosystem are unknown, but describing the species composition of the system represents a first and necessary step to improving this understanding. To better understand the abundance and distribution of shallow-water marine sponges in California, I have used SCUBA to collect over 300 individuals, mostly from kelp forest habitats in the Santa Barbara Channel. I have previously used this collection to revise the order Tethyida in California (Turner 2020b), but the other sponges remain to be described. Ten of these new samples can be assigned to the order Scopalinida, and are described herein as four new species in the genus *Scopalina*.

Accepted by J. Hooper: 27 Apr. 2021; published: 14 May 2021

Methods

Collections. I collected sponge samples by hand with a knife. Each sample was placed immediately in a plastic bag with copious seawater. After the dive, these bags were put on ice for 2–12 hours. Samples were then preserved in 95% ethanol, which was exchanged for new preservative after 1–3 days, and changed again if it remained cloudy. Samples were vouchered with the California Academy of Sciences in San Francisco; archival information is listed in table 1.

Samples were photographed *in situ* with an Olympus TG5 before collection. I photographed all sponge morphotypes found at each site, so that presence/absence across sites could be used to form hypotheses about sponge distributions and habitat. Table 2 contains information about all locations investigated, together with an estimate of search effort at each location. These locations include intertidal sites, floating docks in marinas, and subtidal sites searched on SCUBA. Subtidal sites were generally shallow (<15 m) rocky reefs, but a few sites were deeper (up to 27 m) and include artificial reefs and oil rigs. Scopalinida were only found at shallow, subtidal, natural reefs. Field photos of all sponges have been archived with vouchers, and also posted as searchable, georeferenced records on the site iNaturalist.org.

Spicules. Spicule preparations were performed by digesting soft tissue subsamples in bleach. With the spicules settled at the bottom of the reaction tube, the bleach was then pipetted off and replaced with distilled water; this was repeated several times. Spicules were imaged using a D3500 SLR camera (Nikon) with a NDPL-1 microscope adaptor (Amscope) attached to a compound triocular microscope. A calibration slide was used to determine the number of pixels per mm, and spicules were then measured using ImageJ (Schneider *et al.* 2012). Spicule length was measured as the longest possible straight line from tip to tip, even when spicules were curved or bent. Spicule width was measured at the widest point. To image spicular architecture, I hand-cut perpendicular sections and, when possible, removed sections of ectosome. Sections were digested in a mixture of 97% Nuclei Lysis Solution (Promega; from the Wizard DNA isolation kit) and 3% 20mg/ml Proteinase K (Promega). This digestion eliminates cellular material while leaving the spongin network intact.

Genotyping. I extracted DNA from some samples with the Qiagen Blood & Tissue kit, and used the Qiagen Powersoil kit on others; downstream results did not differ based on the kit used. At the cox1 locus, a ~1200 bp fragment was amplified with the following primers (LCO1490: 5'-GGT CAA CAA ATC ATA AAG AYA TYG G-3'; COX1-R1: 5'-TGT TGR GGG AAA AAR GTT AAA TT-3'); these amplify the "Folmer" barcoding region and the "co1-ext" region used by some sponge barcoding projects (Rot *et al.* 2006).

Three primer sets were used to amplify portions of the 28S rDNA nuclear locus. Samples were sequenced over the ~800 bp D3-D5 region using primers Por28S-830F (5'- CAT CCG ACC CGT CTT GAA -3') and Por28S-1520R (5'- GCT AGT TGA TTC GGC AGG TG -3') (Morrow *et al.* 2012). Most samples were also sequenced over the ~800 bp D1-D2 region using primers Por28S-15F (5'-GCG AGA TCA CCY GCT GAA T-3') and Por28S-878R (5'-CAC TCC TTG GTC CGT GTT TC-3') (Morrow *et al.* 2012). A few samples were sequenced using primers C2 (5'-GAA AAG AAC TTT GRA RAG AGA GT-3') and D2 (5'-TCC GTG TTT CAA GAC GGG-3') (Chombard *et al.* 1998). The C2-D2 region is a ~50% subset of the D1-D2 region, including the most rapidly evolving region recommended by the sponge barcoding project.

PCR was performed using a Biorad thermocycler (T100); the following conditions were used for the cox1 locus: 95°C for 3 min, followed by 35 cycles of 94°C for 30 sec, 52°C for 30 sec, 72°C for 90 seconds, followed by 72°C for 5 minutes. The 28S C2-D2 region was amplified with the same conditions, except a 57°C annealing temperature and 60 second extension time; the 28S D1-D2 and D3-D5 regions used a 53°C annealing temperature and 60 second extension time. PCR was performed in 50 µl reactions using the following recipe: 24 µl nuclease-free water, 10 µl 5x PCR buffer (Gotaq flexi, Promega), 8 µl MgCl, 1 µl 10mM dNTPs (Promega), 2.5 µl of each primer at 10 µM, 0.75 bovine serum albumin (10 mg/ml, final conc 0.15 mg/ml), 0.25 µl Taq (Gotaq flexi, Promega), 1 µl template. ExoSAP-IT (Applied Biosystems) was used to clean PCRs, which were then sequenced by Functional Biosciences using Big Dye V3.1 on ABI 3730xl instruments. PCR products were sequenced in both directions, and a consensus sequence was constructed using Codon Code v.9 (CodonCode Corporation). Blastn was used to verify that the resulting traces were of sponge origin. All sequences have been deposited in Genbank; accession numbers are listed in table 1.

Genetic analysis. I used the NCBI taxonomy browser to compile data from all samples identified as belonging

TABLE 1. Archival information for vouchered samples and samples with sequence data. CASIZ: California Academy of Sciences Invertebrate Zoology.

Collection location	**Lat**	**Long**	**Date**	**Species**	**Collection ID**	**Mueseum voucher**	**CO1 sequence**	**28S sequence**
Big Rock, Santa Cruz Island, CA	34.05220	-119.57360	1/19/20	Scopalina jali	TLT567	CASIZ 235466	-	MT586556
Naples Reef, Santa Barbara, CA	34.42212	-119.95154	9/26/19	Scopalina jali	TLT324	CASIZ 235467	MT583199	MT586558
Naples Reef, Santa Barbara, CA	34.42212	-119.95154	12/10/19	Scopalina jali	TLT558	CASIZ 235468	-	-
Naples Reef, Santa Barbara, CA	34.42212	-119.95154	7/31/19	Scopalina kuyamu	TLT122	CASIZ 235469	MT583200	MT586559
Elwood Reef, Santa Barbara, CA	34.41775	-119.90150	10/23/19	Scopalina goletensis	TLT307	CASIZ 235470	MT583198	MT586557
Coal Oil Point, Santa Barbara, CA	34.40450	-119.87890	8/30/19	Scopalina nausicae	TLT341	CASIZ 235471	-	MT586560
Isla Vista Reef, Santa Barbara, CA	34.40278	-119.85755	8/1/19	Scopalina nausicae	TLT132	CASIZ 235472	-	MT586561
Arroyo Quemado Reef, Santa Barbara, CA	34.46775	-120.11905	1/7/20	Scopalina nausicae	TLT549	CASIZ 235473	-	-
Goalpost, Point Loma, San Diego, CA	32.69438	-117.26860	2/7/20	Scopalina nausicae	TLT471	CASIZ 235474	MW834351	MT586562

TABLE 2. Sampling effort and locations. Harbor and intertidal sites are listed last. For subtidal sites, the number of sampling dives is listed, and the cumulative dive time as an estimate of search effort.

Region	Location	Lat	Long	Dives	Max depth (m)	Cumulative search time (min)	Scopalina nausicae	Scopalina jali	Scopalina kuyamu	Scopalina goletensis
Monterey	Perkin's Reef	36.62920	-121.92031	1	39	46	-	-	-	-
Monterey	Hopkin's Marine Station	36.62084	-121.90142	1	35	74	-	-	-	-
Monterey	Middle Reef, Pt. Lobos	36.52172	-121.93894	1	48	60	-	-	-	-
Santa Barbara	Arroyo Hondo Reef	34.47182	-120.14262	2	26	52	-	-	-	-
Santa Barbara	Arroyo Quemado Reef	34.46775	-120.11905	7	35	302	collected	-	-	-
Santa Barbara	Tajigus	34.46279	-120.10185	2	26	93	-	-	-	-
Santa Barbara	Refugio	34.46097	-120.06640	1	22	60	-	-	-	-
Santa Barbara	Elwood Junkpile	34.42687	-119.92390	1	40	34	-	-	-	-
Santa Barbara	Naples Reef	34.42212	-119.95154	6	52	279	-	collected	collected	-
Santa Barbara	Elwood Reef	34.41775	-119.90150	3	46	138	-	collected	-	collected
Santa Barbara	Goleta Sewer Pipe	34.41420	-119.82930	1	21	22	-	-	-	-
Santa Barbara	Stearn's Wharf	34.41023	-119.68563	1	20	48	-	-	-	-
Santa Barbara	Coal Oil Point	34.40450	-119.87890	10	37	567	collected	-	-	-
Santa Barbara	UCSB intake pipe	34.40297	-119.84080	1	58	34	-	-	-	-
Santa Barbara	Isla Vista Reef	34.40278	-119.85755	4	40	135	collected	-	-	-
Santa Barbara	Leadbetter Reef	34.39895	-119.69703	1	31	14	-	-	-	-
Santa Barbara	1000 Steps	34.39472	-119.71347	1	22	30	-	-	-	-
Santa Barbara	Mohawk Reef	34.39407	-119.72957	7	31	220	-	-	-	-
Santa Barbara	Carpinteria Reef	34.39163	-119.54169	2	19	61	-	-	-	-
Santa Barbara	Oil Platform C	34.33293	-119.63173	1	76	39	-	-	-	-
Santa Barbara	Oil Platform A	34.33189	-119.61353	1	90	35	-	-	-	-
Santa Cruz Island	Hazzard's	34.06066	-119.82810	1	43	75	-	-	-	-
Santa Cruz Island	Diablo Point	34.05878	-119.75776	1	40	62	-	-	-	-
Santa Cruz Island	Cueva Valdez	34.05498	-119.81960	1	45	75	-	-	-	-

.....continued on the next page

TABLE 2. (Continued)

Region	Location	Lat	Long	Dives	Max depth (m)	Cumulative search time (min)	Scopalina nausicae	Scopalina jali	Scopalina kuyamu	Scopalina goletensis
Santa Cruz Island	The Suburbs	34.05291	-119.58309	1	45	59	-	-	-	-
Santa Cruz Island	Big Rock	34.05220	-119.57360	5	47	299	-	collected	-	-
Santa Cruz Island	Platt's Harbor	34.04765	-119.73537	1	34	86	-	-	-	-
Santa Cruz Island	Saddleback Ridge	34.03817	-119.52470	2	41	134	-	-	-	-
San Miguel Island	Wreck of the Cuba	34.03009	-120.45575	1	34	52	-	-	-	-
San Miguel Island	Inside of Wycoff Ledge	34.02132	-120.38710	1	63	52	-	-	-	-
Anacapa Island	Landing Cove	34.01690	-119.36090	1	50	57	-	-	-	-
Anacapa Island	Portuguese Rock	34.01545	-119.42149	2	85	111	-	-	-	-
Anacapa Island	Landslide Cove	34.01539	-119.43560	1	36	49	-	-	-	-
Anacapa Island	Underwater Arch	34.01399	-119.36224	1	33	45	-	-	-	-
San Miguel Island	Miracle Mile	34.01367	-120.34180	2	65	108	-	-	-	-
Anacapa Island	West End	34.01352	-119.44570	1	38	40	-	collected	-	-
Anacapa Island	Zebra Cove	34.01000	-119.44000	1	42	66	-	-	-	-
Anacapa Island	Underwater Island	34.00720	-119.42959	2	60	102	-	-	-	-
Anacapa Island	Cat Rock	34.00426	-119.42335	1	37	65	-	-	-	-
Santa Cruz Island	Pink Ribbon	33.98967	-119.52470	1	36	60	-	-	-	-
Santa Cruz Island	E. of Marmeda	33.98380	-119.61290	1	49	38	-	-	-	-
Santa Cruz Island	Marmeda	33.98378	-119.63910	1	43	35	-	-	-	-
Santa Rosa Island	Johnson's Lee	33.89966	-120.10735	2	49	65	-	-	-	-
Los Angeles	Redondo barge	33.83833	-118.41040	2	90	49	-	-	-	-
Los Angeles	Flat rock	33.79662	-118.41110	1	26	45	-	-	-	-
Los Angeles	Resort Wall	33.76499	-118.42815	1	69	43	-	-	-	-
Los Angeles	Halfway reef	33.76265	-118.42560	1	74	40	-	-	-	-
Los Angeles	Hawthorne reef	33.74714	-118.42090	1	79	39	-	-	-	-
Catalina Island	Parson's Landing	33.47502	-118.55000	2	55	120	-	-	-	-

.....continued on the next page

TABLE 2. (Continued)

Region	Location	Lat	Long	Dives	Max depth (m)	Cumulative search time (min)	Scopalina nausicae	Scopalina jali	Scopalina kuyamu	Scopalina goletensis
Catalina Island	Eel Cove	33.45662	-118.51120	1	30	30	-	-	-	-
San Diego	La Jolla	32.85227	-117.27239	1	51	67	photographed	-	-	-
San Diego	Six Fathoms	32.71000	-117.26860	1	60	40	-	-	-	-
San Diego	Goalpost	32.69438	-117.26860	1	50	50	collected	-	-	-
San Diego	Dino Head	32.68808	-117.27080	1	84	29	-	-	-	-
San Diego	Train Wheels	32.65205	-117.26243	1	94	30	-	-	-	-
Marin	Tomales Bay Resort & Marina	38.10792	-122.86237	n/a	n/a	10	-	-	-	-
Marin	Sausalito Yacht Harbor	37.85930	-122.48044	n/a	n/a	24	-	-	-	-
Alameda	Jack London Marina	37.79371	-122.27757	n/a	n/a	30	-	-	-	-
San Mateo	Pillar Point Harbor	37.50231	-122.48042	n/a	n/a	20	-	-	-	-
Monterey	Monterey Marina	36.60797	-121.89288	n/a	n/a	20	-	-	-	-
San Luis Obispo	Morro Bay Embarcadero	35.36989	-120.85627	n/a	n/a	20	-	-	-	-
San Luis Obispo	Morro Bay Public Boat Launch	35.35751	-120.85047	n/a	n/a	15	-	-	-	-
Santa Barbara	Santa Barbara Harbor	34.40559	-119.68964	n/a	n/a	150	-	-	-	-
Ventura	Ventura Harbor	34.24801	-119.26550	n/a	n/a	70	-	-	-	-
Los Angeles	Marina del Rey	33.97228	-118.44653	n/a	n/a	30	-	-	-	-
Los Angeles	22nd Street Marina	33.72477	-118.28078	n/a	n/a	20	-	-	-	-
Los Angeles	Rainbow Harbor	33.76184	-118.19111	n/a	n/a	30	-	-	-	-
Orange	Newport Bay: N. Balboa Island	33.60879	-117.89304	n/a	n/a	20	-	-	-	-
Orange	Newport Bay: Balboa Peninsula	33.60375	-117.90052	n/a	n/a	20	-	-	-	-

.....continued on the next page

TABLE 2. (Continued)

Region	Location	Lat	Long	Dives	Max depth (m)	Cumulative search time (min)	Scopalina nausicae	Scopalina jali	Scopalina kuyamu	Scopalina goletensis
Orange	Dana Point Harbor: Embarcadero	33.46090	-117.69156	n/a	n/a	30	-	-	-	-
San Diego	Oceanside harbor	33.20623	-117.39208	n/a	n/a	20	-	-	-	-
San Diego	Mission bay: Ski Beach Launch	32.77359	-117.23277	n/a	n/a	10	-	-	-	-
San Diego	Mission Bay: South Shores Launch	32.76406	-117.21750	n/a	n/a	30	-	-	-	-
San Diego	Mission Bay: Quivera Basin	32.76421	-117.23829	n/a	n/a	40	-	-	-	-
San Diego	San Diego Bay: Shelter Island Marina	32.71094	-117.23423	n/a	n/a	40	-	-	-	-
Marin	Drake's Estero intertidal	38.03523	-122.92903	n/a	n/a	80	-	-	-	-
Santa Cruz	Scott's Creek Intertidal	37.04213	-122.23366	n/a	n/a	60	-	-	-	-
Monterey	Point Pinos Intertidal	36.63447	-121.93923	n/a	n/a	90	-	-	-	-
Monterey	Asilomar State Beach intertidal	36.62744	-121.94098	n/a	n/a	75	-	-	-	-
Monterey	Carmel Point intertidal	36.54488	-121.93300	n/a	n/a	40	-	-	-	-
San Luis Obispo	Hazard Canyon intertidal	35.28959	-120.88415	n/a	n/a	140	-	-	-	-
San Luis Obispo	Cave Landing intertidal	35.17535	-120.72240	n/a	n/a	80	-	-	-	-
Santa Barbara	Elwood Intertidal	34.42395	-119.90604	n/a	n/a	20	-	-	-	-
Santa Barbara	COP Intertidal	34.40666	-119.87750	n/a	n/a	60	-	-	-	-
Ventura	Lechuza Point intertidal	34.03437	-118.86146	n/a	n/a	60	-	-	-	-
Los Angeles	Point Fermin intertidal	33.70664	-118.28595	n/a	n/a	64	-	-	-	-
San Diego	Point La Jolla Intertidal	32.85112	-117.27298	n/a	n/a	60	-	-	-	-
San Diego	Bird Rock intertidal	32.81447	-117.27431	n/a	n/a	50	-	-	-	-

to the order Scopalinida. I also used blastn to search for additional sequences that appeared to fall within this order, but found none. At the 28S locus, data from Genbank was only included in the phylogeny if the highly variable C2-D2 region was included. Sequences at cox1 were included if they contained the Folmer barcoding region. Together, included data are from 18 different publications (Blanquer & Uriz 2008; Erpenbeck *et al.* 2002, 2006, 2007a, b, 2012, 2016; Kandler *et al.* 2019; Lavrov *et al.* 2005; Lavrov & Lang 2005; Montalvo & Hill 2011; Morrow *et al.* 2012, 2013; Nichols 2005; Pett & Lavrov 2015; Riesgo *et al.* 2013; Rot *et al.* 2006; Thacker *et al.* 2013). Sequence alignments were produced in Codon Code v.9 (CodonCode Corporation). Phylogenies were estimated with maximum likelihood using IQ-Tree (Nguyen *et al.* 2015; Trifinopoulos *et al.* 2016). Phylogenies are unrooted, and root placement in figures is based on a published phylogeny that had many more characters and representatives from more sponge orders, but limited taxon sampling within the Scopalinida (Morrow *et al.* 2013). I used the Ultrafast bootstrap (Hoang *et al.* 2018) to measure node confidence. Phylogenies were produced from the IQ-Tree files using the Interactive Tree of Life webserver (Letunic & Bork 2019). Figures were made ready for publication using R (r-project.org) and/or Gimp (gimp.org).

Results

Genetic Results

Figure 1 shows the phylogentic tree of newly collected samples, previously sequenced Scopalinida, and outgroups at the cox1 mitochondrial locus. The four new species from California form a clade with five previously sequenced *Scopalina*, with *Svenzea zeai* as the closest outgroup. No other closely related sequences could be found in Genbank, so these data support inclusion of the new species in the genus *Scopalina*. Figure 2 shows the phylogentic tree at the 28S nuclear locus (the large ribosomal subunit), which is entirely congruent with results at cox1.

Both gene trees place two of the new species, *S. kuyamu* and *S. goletensis*, as the closest relatives to each other. This raises the question of whether these two species may in fact be a single species. However, these vouchers are different at 4.4% of sites at the cox1 locus, and 3.2% of sites at the 28S locus. The magnitude of this difference is similar to species-level divergence among other Porifera. A review of genetic distances at cox1 found an average 4.9% sequence divergence between any individual sponge sequence and the most closely related sequence available from another species (N=57, (Huang *et al.* 2008)). A more recent analysis of 39 species in the poriferan order Suberitida found 3.7% divergence at this same locus (Turner 2020a). Together with the morphological differences detailed below, these data support species status for both *S. kuyamu* and *S. goletensis*.

Systematics

Order Scopalinida Morrow & Cárdenas, 2015

Definition: Encrusting, massive or erect flabellate growth forms; smooth or conulose surface supported by prominent spongin fibres cored with spicules; megascleres styles and/or oxeas, often with telescoped ends; no ectosomal skeleton; tissue contains an unusual cell type filled with refractile granules. (Modified from Morrow and Cárdenas (2015) to include oxeas among megascleres.)

Remarks: As the focus of this paper is alpha taxonomy, rather than a revision of higher taxonomy, I have retained the definition of Morrow and Cárdenas (2015), adding oxeas among the megascleres as the only modification. However, it should be noted that a more thorough revision is needed, and would likely result in further changes. The description of *Svenzea zeai*, for example, does not include prominent spongin fibers.

Family Scopalinidae Morrow *et al.*, 2012

Definition same as order.

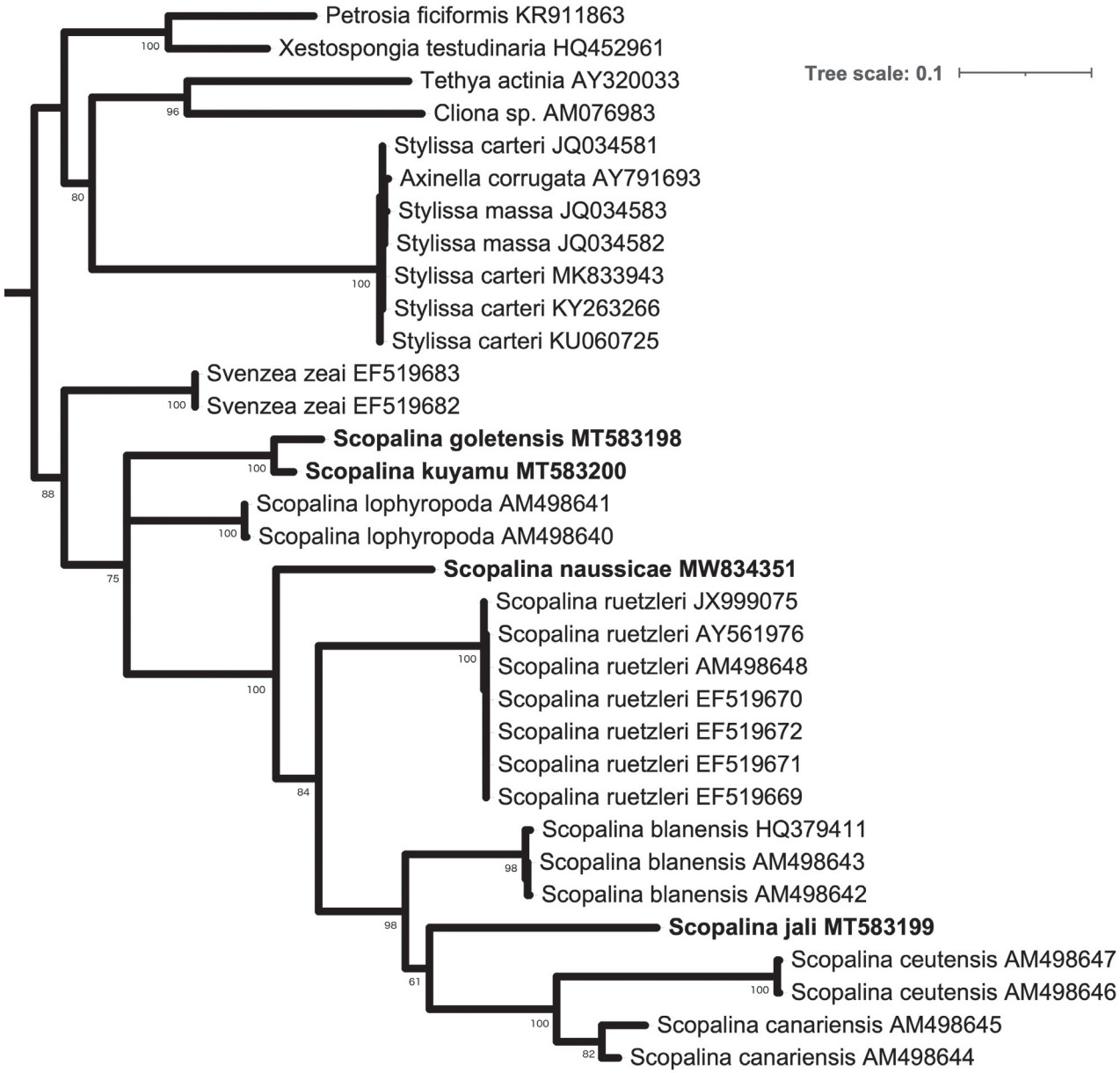

FIGURE 1. Gene tree at the cox1 mitochondrial locus. Bootstrap values are shown for nodes with > 50% support; nodes with < 50% support are collapsed. Genbank accession numbers are shown; those beginning with M, shown in bold, are new. Scale bar indicates substitutions per site. Root placement based on Redmond *et al.* (2013).

Scopalina Schmidt 1862

Definition: Thinly or thickly encrusting; soft and compressible; few or no ectosomal spicules; spongin abundant, with extensions of spongin manifest as mounds or fibers arising from basal spongin plate; these fibers may branch and merge; choanosomal skeleton of spicules or spicule bundles with proximal ends or entire spicule enclosed in spongin; choanosome may have a grainy appearance. Larvae are elongated, conical; anterior region wider than the posterior zone; completely covered by short cilia. (Modified from Blanquer and Uriz 2008).

Diagnosis: *Scopalina* have abundant spongin, while *Svenzea* are described as having limited spongin, primarily at the nodes of a reticulated spicule network. *Svenzea* tend to have shorter spicules, (200–300 µm), whereas in *Scopalina* they mostly range from 400 to 2000 µm (though *S. canariensis* averages only 199 µm). The skeletal architecture of *Svenzea* has been noted as more like that of the haplosclerida than *Scopalina*. *Svenzea* are massive or thickly encrusting, while *Scopalina* are thinly to thickly encrusting.

Stylissa are erect, flabellate, or lobate, rather than possessing encrusting morphologies seen in *Scopalina*. *Stylissa* are noted as having a skeletal architecture like that of the Halichondridae, with many spicules in confusion.

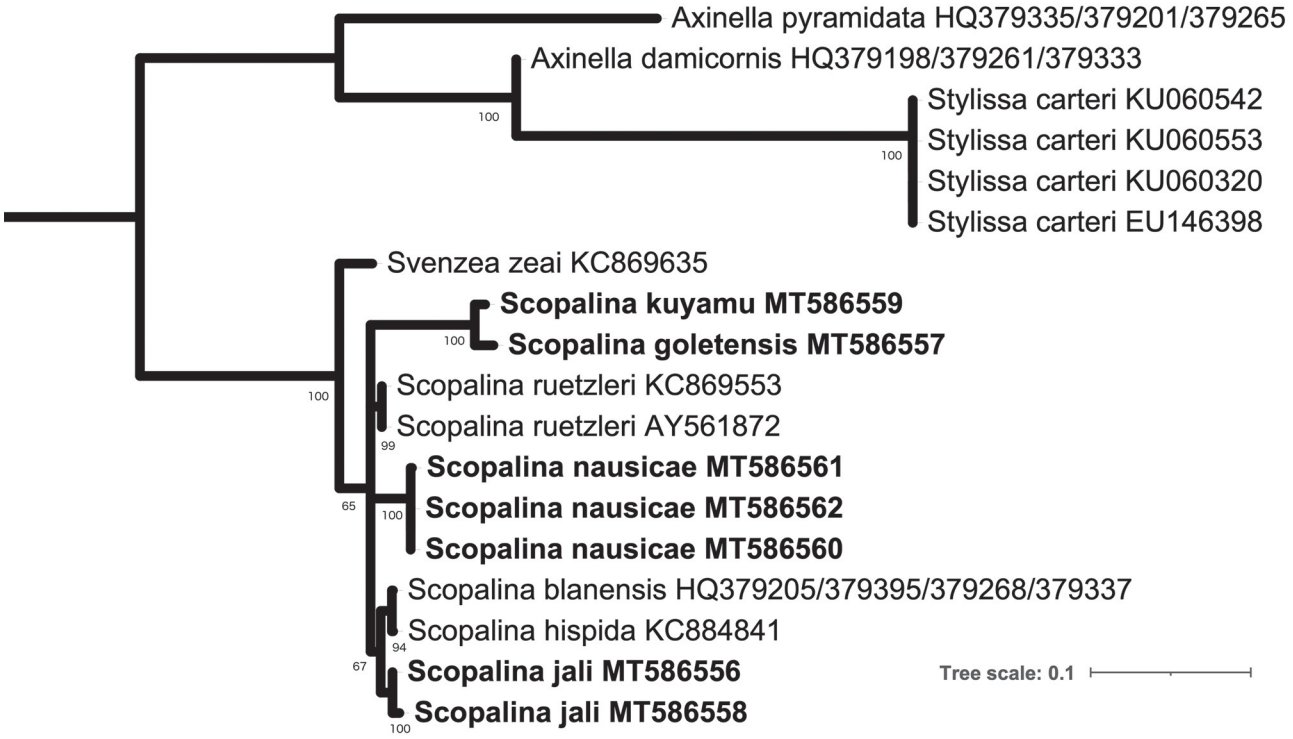

FIGURE 2. Gene tree at the large ribosomal subunit (28S). Bootstrap values are shown for nodes with > 50% support; nodes with < 50% support are collapsed. Genbank accession numbers are shown; those beginning with MT, in bold, are new. Scale bar indicates substitutions per site. Root placement based on Redmond *et al.* (2013).

Scopalina nausicae **sp. nov.**
(Fig. 3)

Material examined. Holotype: (CASIZ 235474) Point Loma, San Diego, California, USA (32.69438, -117.26860), 15 m depth, 2/7/20. Paratypes: (CASIZ 235471) Coal Oil Point, Santa Barbara, California, USA (34.40450, -119.87890), 11 m depth, 8/30/19; (CASIZ 235472) Isla Vista Reef, Santa Barbara, California, USA (34.40278, -119.85755), 12 m depth, 8/1/19; (CASIZ 235473) Arroyo Quemado Reef, Santa Barbara, California, USA (34.46775, -120.11905), 11 m depth, 1/7/20.

Etymology. Named for the fictional character Nausicaä from the film *Nausicaä and the Valley of the Wind*.

Morphology. Encrusting, 2–4 mm thick, up to 10 cm across (Fig. 3). Soft and compressible. Prominent conules 0.5–1.0 mm in height,1.5–3.5 mm apart; spicules protrude at conules, making them microscopically hispid. Scattered oscules 1–2 mm in diameter. In nature, ectosome appears opaque at conules but often lacy and porous between them; ectosome more opaque in collected samples. Ectosome peach colored, choanosome yellow when alive; all tissues fade to beige when preserved in ethanol.

Skeleton. Vertical trunks of spongin, 100–550 μm wide, arise from a basal spongin mat and terminate in surface conules. Secondary branches of spongin 50–100 μm wide arise from primary trunks, branching off at an angle of less than 90 degrees and still extending towards surface. Primary and secondary trunks cored with spicules with pointed ends up; spicules entirely enclosed in spongin or with tips projecting; projecting tips fan out to create a bouquet that pierces the ectosome at conules. An additional type of spongin tract is distinct from those described above: 60–90 μm wide, these tracts branch from primary trunks at approximately 90-degree angles, then meander through the choanosome in a vermiform fashion, sometimes branching; these vermiform tracts do not contain spicules. Basal spongin, spicule-containing spongin trunks, and vermiform tracts are sporadically cored with sediment. Spicule-containing and vermiform spongin tracts are often filled and/or coated with what appear to be algal cells; these are red in preserved tissue.

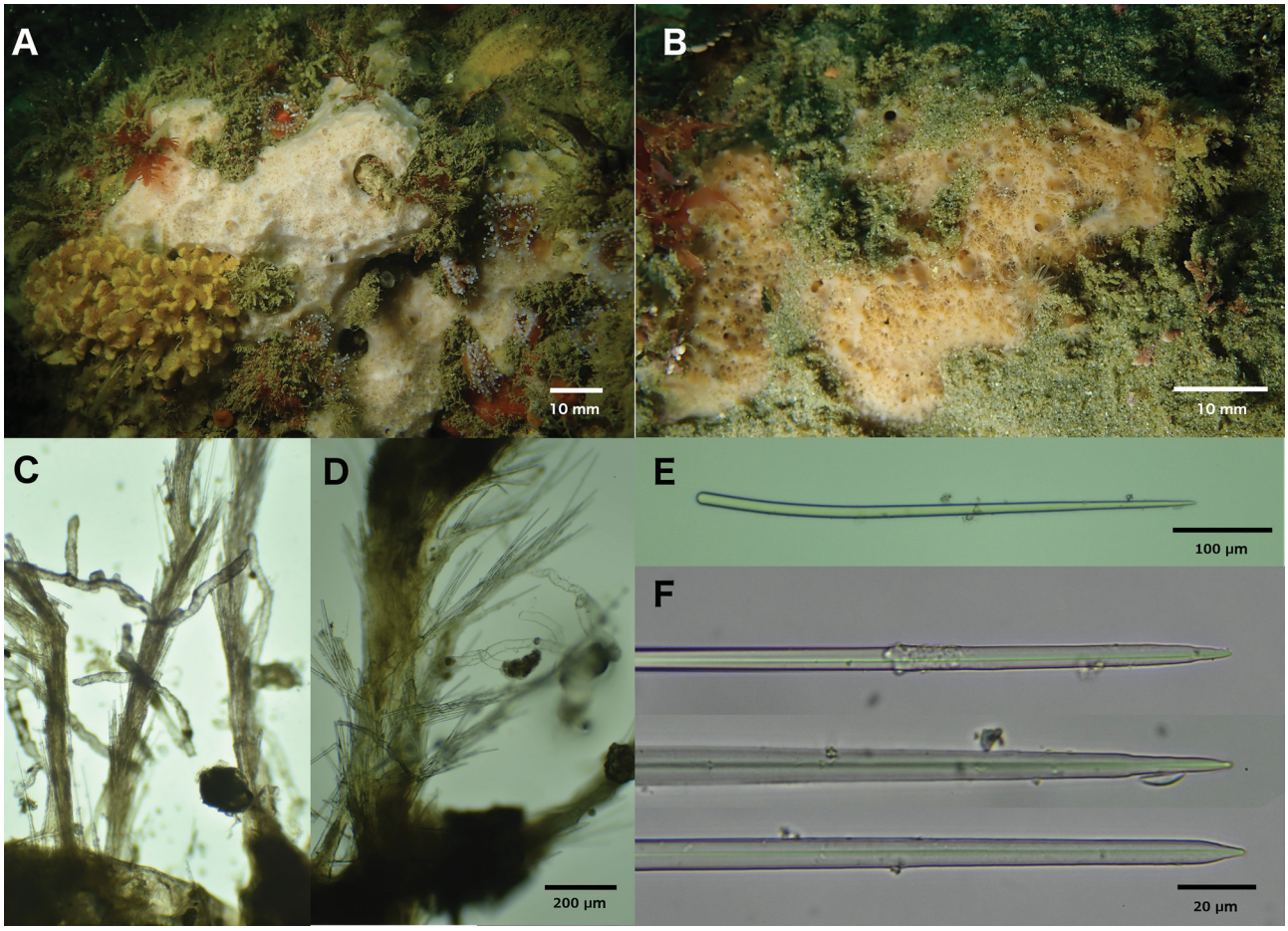

FIGURE 3. *Scopalina nausicae* **sp. nov**. A: Field photo of paratype (CASIZ 235472), Isla Vista Reef. B: Field photo of paratype (CASIZ 235473), Arroyo Quemado Reef. C, D: Skeletal architecture of holotype (CASIZ 235474), Point Loma; scale applied to both. In C, vermiform and primary spongin trunks are coated in apparent algae. In D, only primary spongin trunk showing algae, but vermiform tracts are sporadically cored with sediment. E, F: spicules from paratype (CASIZ 235472).

Spicules. Styles only, usually bent towards the head end, thickest at the head and tapered to a point. Some show "telescoping" (width decreasing in a step-wise fashion) at the pointed end. Average spicule length for each voucher: 454, 483, 505, 532 µm (N=31–40 per sample); total range in spicule length across vouchers 375–623 µm (N=135). Average spicule width at head, for each voucher: 9, 9, 11, 11 µm (N=31–40 per sample); total range in spicule width at head 5–17 µm (N=135).

Distribution and habitat. This species is common on the shallow (5–16 m) rocky reef at Coal Oil Point, Santa Barbara, California. Often found on vertical rock walls or boulders, it can also occur on flatter areas, and has been found partially buried by sand. It was not found at most other locations investigated, but was located in similar habitat at the Arroyo Quemado Reef (near Point Conception) and in the kelp forests in extreme Southern California, off Point Loma and La Jolla, San Diego. It is therefore likely that the specie's range encompasses at least the Southern Californian and Ensenadan biogeographical provinces, bounded by Point Conception in the North and Punta Eugenia in the South (Blanchette *et al.* 2008; Valentine 1966).

Remarks. Skeletal architecture, spiculation, and genotype all conspire to place this species within the *Scopalina*. I was unable to detect the "graininess" said to characterize other Scopalinidae. However, this was hard to assess due to the abundant sediment within the sponge: dark grains were apparent, but appeared to be sediment rather than refractile cells.

Spicule dimensions, skeletal morphology, and genotype all serve to differentiate *S. nausicae* **sp. nov.** from the three other species newly described here. Fourteen other species are currently placed in the genus *Scopalina*, according to the World Porifera Database (van Soest *et al.* 2019). None of these are known from the Eastern Pacific, making them unlikely conspecifics with any of the species described here. The gross morphology of *S. nausicae*

sp. nov. in the field is quite similar to published images of *S. ruetzleri* (Wiedenmayer, 1977) (West Atlantic) and *S. erubescens* (Goodwin *et al.,* 2011) (Faulkland Islands). Spicule length and sponge color also match *S. erubescens* better than other *Scopalina*, making this species the most likely conspecific. In addition to geographic separation, however, *S. erubescens* is larger, more thickly encrusting, and has thicker spicules and spicule bundles. The description of *S. erubescens* also lacks any mention of the vermiform spongin tracts that pervade *S. nausicae* **sp. nov.** (Goodwin *et al.* 2011). *Scopalina ruetzleri* can be excluded as a conspecific based on genetic data at both cox1 and 28S as well as color and habitat (Rützler *et al.* 2003). This species is described as ranging throughout the Caribbean, but was also recently reported from the tropical Eastern Pacific (Carballo *et al.* 2019). This latter report is not accompanied by morphological or genetic information, so comparisons between tropical Pacific *Scopalina* and *S. nausicae* **sp. nov.** await future investigation.

Within its range, it is likely that this sponge can be identified from field photos, as I have seen no other sponge with a similar morphology to date.

***Scopalina kuyamu* sp. nov.**
(Fig. 4)

Material examined. Holotype: (CASIZ 235469) Naples Reef, Santa Barbara, California, USA (34.42212, -119.95154), 12 m depth, 7/31/19.

Etymology. Named for the village of Kuyamu, a community of Barbareño Chumash that once stood onshore at the site where the sponge was discovered.

Morphology. Encrusting, 1–2 mm thick, 6 cm across (Fig. 4). Soft and compressible. Surface hispid due to a profusion of protruding styles. Distinct ectosome not apparent. Peach colored in nature, except for translucent-white varicose channels running along surface. Few scattered oscules, each ~300 μm diameter; smaller pores (approximately 80 μm diameter) abundant and uniformly distributed. Beige when preserved in ethanol.

Skeleton. Basal mat of spongin cored with sediment. Extensions of spongin arise from this mat: most are low mounds, some only 25–50 μm high, but some are fingers 100–300 μm high and cored with sediment. Heads of spicules are embedded in these mounds and fingers, either singly or in bundles of up to 12. Spicules extent vertically and pierce the surface of the sponge.

Spicules. Styles only, usually bent towards the head end, thickest at the head and tapered to a point. Some spicule tips are "telescoping" (width decreasing in a step-wise fashion) at the pointed end. Spicules averaged 1557 μm in length (N=35, range 879–1948 μm); 16 μm in width (N=35, range 11–21).

Distribution and habitat. Only a single individual has been found, on a vertical wall at 12 m depth, at Naples Reef, in Santa Barbara, California. Habitat was rocky reef with abundant bryozoan, sponge, and anthozoan cover, adjacent to year-round kelp forest. Three additional dives at the same location failed to locate other individuals; similar, nearby habitat to the East and West also had considerable search effort, so this species appears to be rare in this area.

Remarks. This sponge is quite genetically and morphologically distinct from *S. nausicae* and *S. jali*. The spicular architecture is fairly similar to *S. goletensis*, though the spicule density is lower. As a result, *S. goletensis* is removable from the substrate as a fairly firm sheet, while *S. kuyamu* peels away in rubbery strips that curl up upon themselves. Also, the spicules average over twice as long in *S. kuyamu*, with non-overlapping size ranges among the spicules measured. These morphological differences seem unlikely to be due to environmental influences, as the two species were collected at the same depth, at very similar reefs, less than 5 km apart. Together with the considerable genetic divergence, these differences support species status for both species.

Among *Scopalina* from other regions, the only species with spicules as large as *S. kuyamu* are *S. lophyropoda* (Schmidt, 1862) (Blanquer & Uriz 2008) (Mediterranean) and *S. bunkeri* (Goodwin *et al.,* 2011) (Falkland Islands). In addition to great geographic distance, *S. lophyropoda* can be excluded based on genetic data (Fig. 1); *S. bunkeri* has a different spicular architecture, gross morphology, and color (Goodwin *et al.* 2011).

It does not seem likely that this species can be identified from field photos alone, though it is difficult to say if there are reliable field marks until more individuals are found.

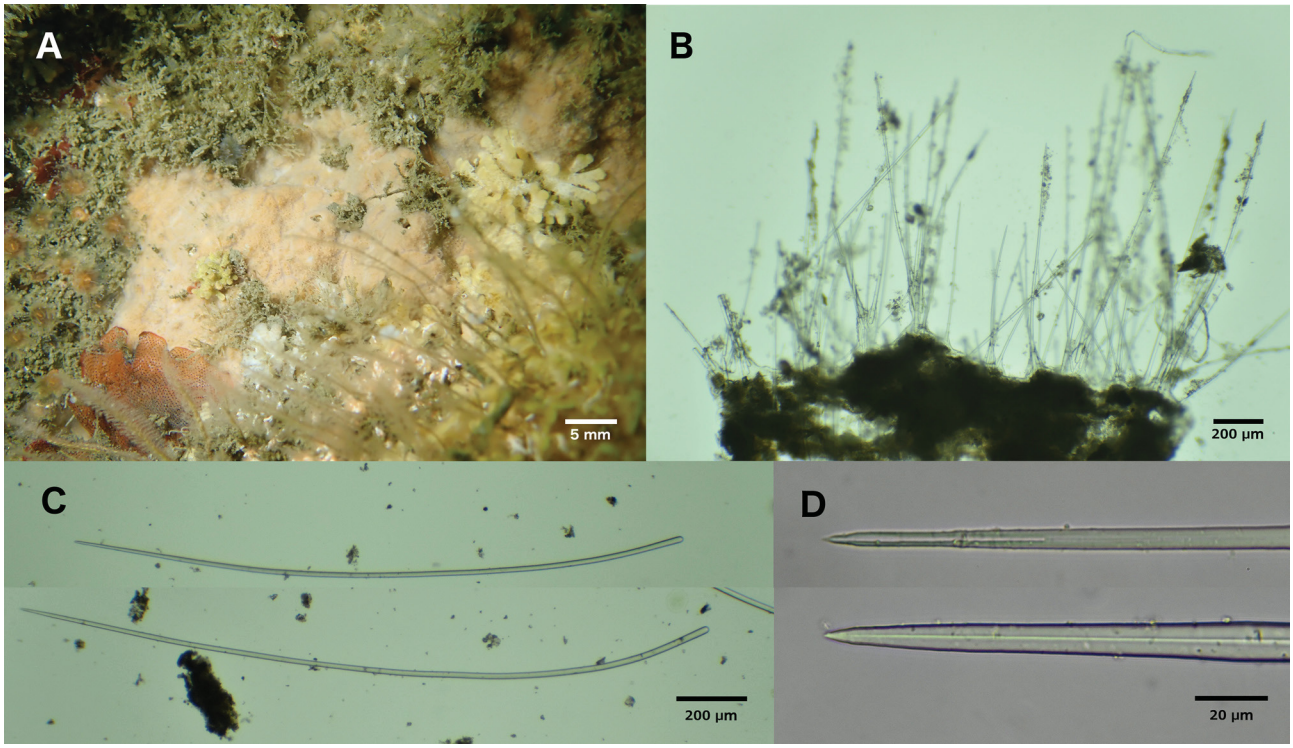

FIGURE 4. *Scopalina kuyamu* **sp. nov.**. All images from holotype. A: field photo. B: Skeletal architecture, showing basal plate cored with copious sediment. C, D spicules.

***Scopalina goletensis* sp. nov.**
(Fig. 5)

Material examined. Holotype: (CASIZ 235470) Elwood Reef, Santa Barbara, California, USA (34.41775, -119.90150), 12 m depth, 10/23/19.

Etymology. Named for the town of Goleta that is onshore from the location where the sponge was discovered.

Morphology. Encrusting, 1.0–1.2 mm thick, approximately 2.5 cm across (figure 5). Firm and incompressible. Surface hispid due to dense profusion of protruding styles. Distinct ectosome not apparent. Beige / cream colored in nature; retained the same color when preserved in ethanol. Surface traced by varicose, translucent channels; pores (approximately 200–300 µm diameter) abundant and uniformly distributed.

Skeleton. Basal mat of spongin cored with sediment. Vertical extensions of spongin 10–600 µm high arise from this mat: none of these were cored with sediment, but loose sediment was abundant throughout the sponge. Heads of some spicules are embedded singly, directly in the basal mat of spongin, but most are embedded as tiered bundles in the vertical extensions of spongin.

Spicules. Styles only, usually slightly bent towards the head end, thickest at the head and tapered to a point. Some spicule tips are "telescoping" (width decreasing in a step-wise fashion) at the pointed end. Spicules averaged 687 µm in length (N=37, range 388–801 µm); 15 µm in width (N=37, range 6–21).

Distribution and habitat. Only a single individual has been found, on a vertical ledge at 12 m depth, at Elwood Reef, in Santa Barbara, California. Habitat was rocky reef with abundant bryozoan, sponge, and anthozoan cover, under a year-round kelp canopy. Considerable search effort at Elwood Reef and nearby locations failed to locate additional individuals, so this species is likely to be rare in this area.

Remarks. This species is most similar to *S. kuyamu,* but is morphologically and genetically distinct, as detailed in the *S. kuyamu* remarks. The spicule dimensions are similar to several species from other regions (*S. azurea* (Bibiloni, 1993)*, S. blanensis* (Blanquer & Uriz, 2008)*, S. hispida* (Hechtel, 1965)), though none of these others is known to have spicules as thick. All but *S. azurea* can also be excluded based on the available genetic data (Figs. 1,

2). Conspecificity with *S. azurea* is unlikely based on geographic isolation, color, and spicular architecture (Bibiloni 1993).

It does not seem likely that this species can be identified from field photos alone, though it is difficult to say if there are reliable field marks until more individuals are found.

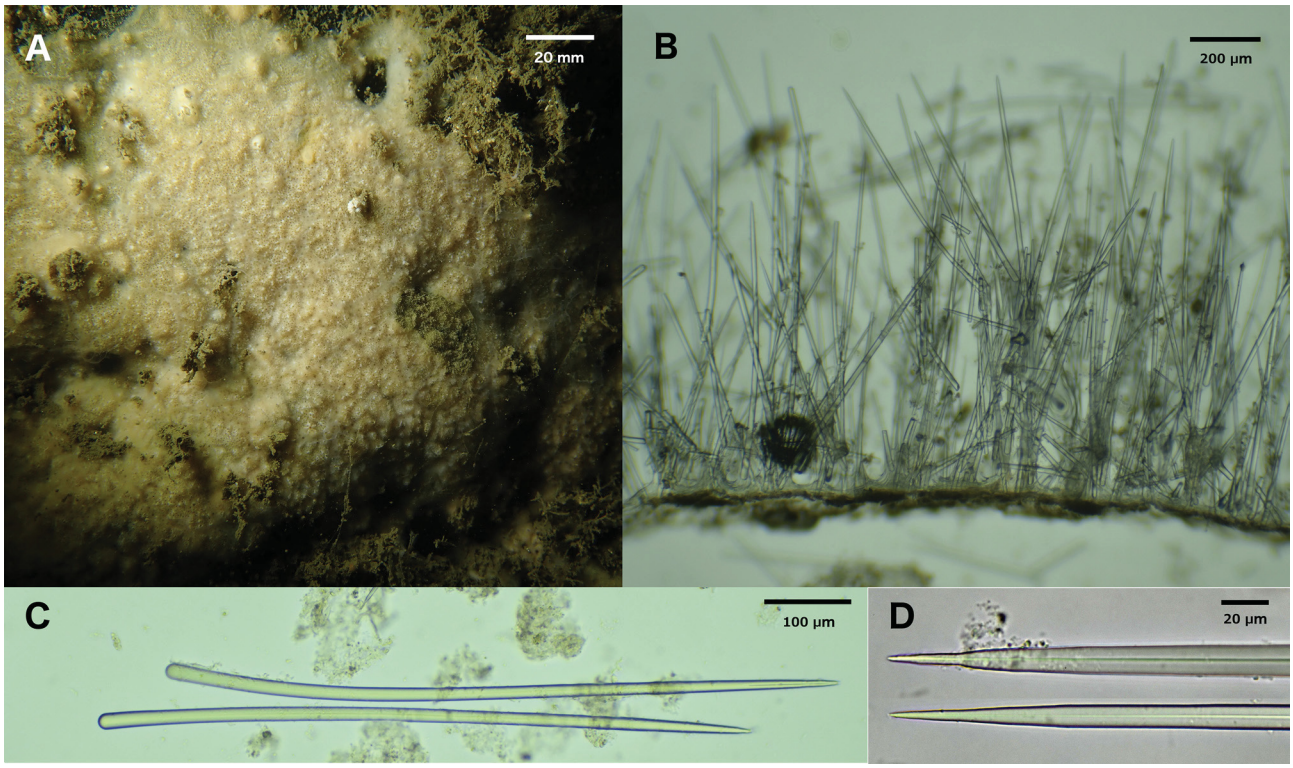

FIGURE 5. *Scopalina goletensis* **sp. nov.**. All images from holotype. A: field photo. B: Skeletal architecture, showing basal plate cored with sediment. C, D spicules.

***Scopalina jali* sp. nov.**
(Fig. 6)

Material examined. Holotype: (CASIZ 235466) Big Rock, Santa Cruz Island, California, USA (34.05220, -119.57360), 12m depth, 1/19/20. Paratypes: (CASIZ 235467) Naples Reef, Santa Barbara, California, USA (34.42212, -119.95154), 11 m depth, 9/26/19. (CASIZ 235468) Naples Reef, Santa Barbara, California, USA (34.42212, -119.95154), 15 m depth, 12/10/19.

Etymology. The ectosome of live specimens *in situ* is reminiscent of a jali: a latticed screen common in Indo-Islamic architecture.

Morphology. Thickly encrusting, 1.0–1.5 cm thick, up to 35 cm across (Fig. 6). Soft, spongey, and very compressible. Ectosome transparent, without spicules; a lattice-like mesh of spongin fibers visible in life; conules present, but very small (100–300 µm in width and height); ectosome more opaque after preservation in ethanol but remains partially transparent and lacy. Color in freshly collected specimens is terra-cotta (reddish-brown); red and orange tones are more muted in field photos, with color appearing to vary from tan to terra-cotta; samples fade to beige when preserved in ethanol. Oscules 10–20 mm in diameter; occur singly; sparse in some samples and denser in others; partially closed by ectosomal membrane in collected samples.

Skeleton. Abundant spongin fibers cored with spicules form a chaotic mesh lattice within choanosome. Larger spongin tracts, 45–65 µm wide, are cored with bundles approximately 5 spicules wide; smaller tracts, 8–20 µm wide, are cored with single spicules. No spicules detected outside of spongin tracts. Considerable silt apparent in tissue sections, but none seen coring spongin tracts.

Spicules. Oxeas only, gently curved; some spicule tips show "telescoping" (width decreasing in a step-wise fashion). Average spicule length for each voucher: 354, 358, 366 µm (N=30–37 per sample); total range in spicule

length across vouchers 219–436 µm (N=100). Average spicule width at widest point, for each voucher: 8, 9, 11 µm (N=30–37 per sample); total range in spicule width at head 2–18 µm (N=100).

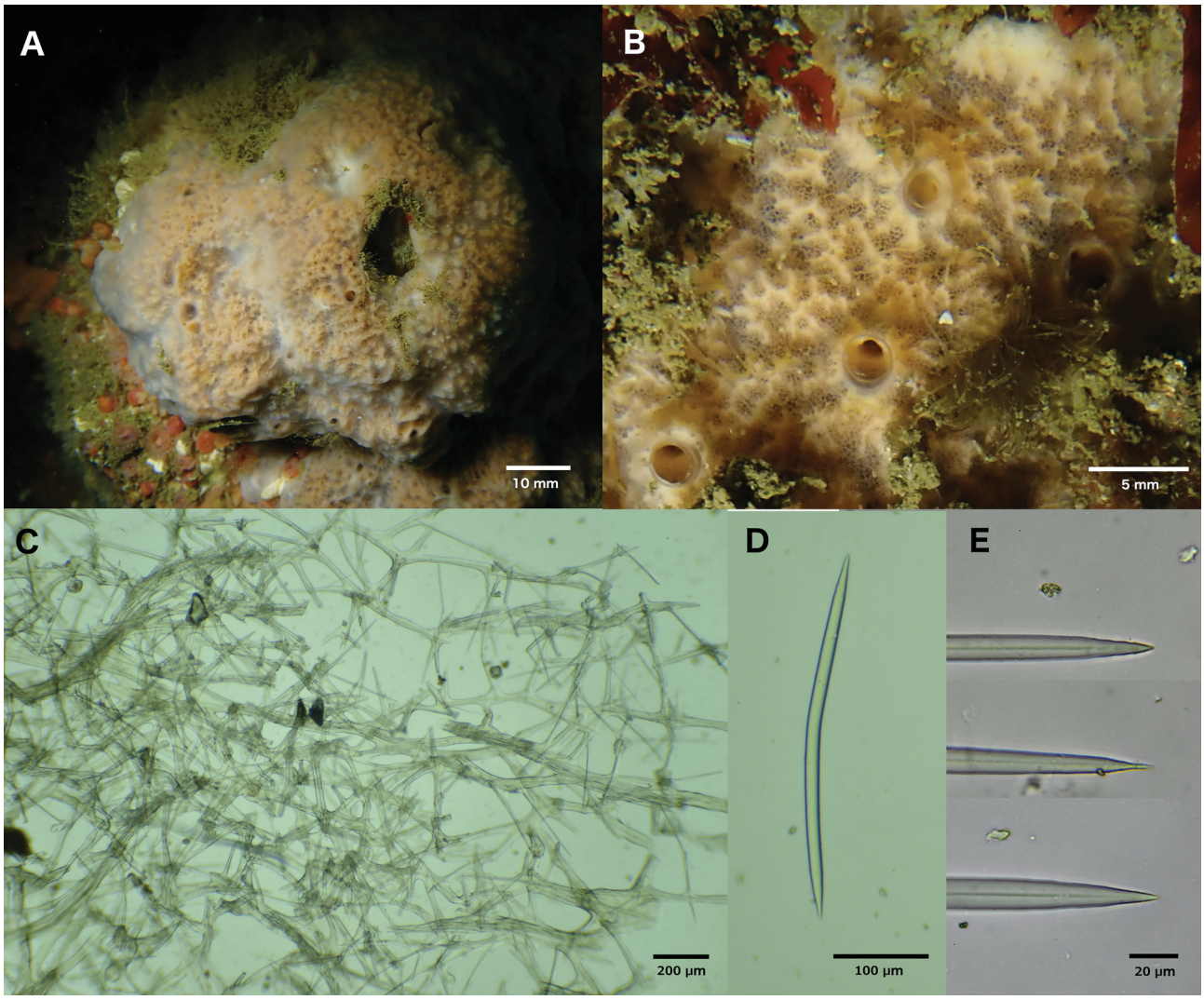

FIGURE 6. *Scopalina jali* **sp. nov.**. A: Field photo of holotype (CASIZ 235466), Santa Cruz Island. B: Field photo of paratype (CASIZ 235467), Naples Reef. C: Skeletal architecture of holotype. In C, vermiform and primary spongin trunks coated in apparent algae. D, E: spicules from paratype (CASIZ 235467).

Distribution and habitat. In the winter of 2019–2020, this sponge was abundant on the shallow (5–17 m) rocky reefs off of Naples Point and the Elwood Bluffs, Santa Barbara, California. The species was not seen in 4 dives at these same locations in the Spring and Summer of 2019, suggesting that the population may vary seasonally or in a boom-and-bust fashion on longer timescales. Consistent with this latter possibility, many large individuals of this species were seen at the Big Rock dive site at Santa Cruz Island in January of 2020, while no individuals were seen in three dives at the same location in November of 2018. The only other probable sighting I am aware of is a photo uploaded to the site iNaturalist (inaturalist.org/observations/41000570). This photo is very likely to be *S. jali*, as no other sponge with this morphology is known in Southern California. The photo is annotated as from Heisler Park, Laguna Beach, from 3/4/2007.

Remarks. Genetic data at two loci confirm that this species is within the *Scopalina*. Abundant spongin fibers cored with simple spicules, telescoping spicule tips, and lack of ectosomal skeleton are all consistent with this placement. The presence of oxeas, rather than styles, required modification of recent definitions of order, family, and genus -- though one species currently placed in *Scopalina* in the World Porifera database also contains only oxeas (*S. agoga* (de Laubenfels, 1954)) and another contains both styles and oxeas (*S. australiensis* (Pulitzer-Finali, 1982)). *Scopalina jali* is differentiated from *S. agoga* by spicule size and the presence of many tangential spicules in the ectosome of *S. agoga*; this previously described species is also known only from Palau (de Laubenfels 1954). The

skeletal architecture of *S. jali* differs markedly from the other California species described herein due to its highly reticulated nature, but this is similar to the published description of the Atlantic species *S. ceutensis* (Blanquer & Uriz, 2008).

It is likely that this sponge can be identified from field photos alone within Southern California, as I have seen no other sponge with a similar morphology to date.

Conclusions

It is remarkable that, among the relatively well-studied kelp forests of the Santa Barbara Channel, I was able to locate 4 new species from an order not previously known to occur in the region. These species were not only undescribed, but apparently unsampled: no previous California survey includes samples matching their description (Lee *et al.* 2007). These results illustrate how much remains to be learned about the sponges of California, and show that SCUBA-based collection efforts can help bridge this gap. Moreover, collection by hand allows for underwater photography of live samples before collection. By comparing photographs of the species described here with photos of all other sponges I have collected, I believe that the two more common species (*S. jali* and *S. nausicae*) are easily recognizable within their range. This assertion is supported by the fact that, after collection of the first samples of each, subsequent samples were correctly identified as conspecifics based on field photos before being confirmed as such using spicules and DNA. This will simplify future efforts to understand the ecology of these species, perhaps by using diver surveys or photo transects.

In contrast, the other two new species (*S. kuyamu* and *S. goletensis*) are thinly encrusting and will be harder to identify based on gross morphology. Each was found only once; as they were both found within the range and habitat I have most thoroughly sampled, it is likely they are uncommon in this area (but could be common in deeper waters). Though describing new species from a single sample is not ideal, I feel confident these samples are not conspecific with any currently named species due to their considerable genetic divergence from other sequenced species, substantial morphological differences from un-genotyped species, and the fact that no previously named species are known from the region.

Much remains to be learned about the Scopalinida. Twenty species were previously known to reside in the order: 14 *Scopalina* species*,* 5 *Svenzea* species, and *Stylissa flabelliformis* (Morrow & Cárdenas 2015). These sponges are known from the Mediterranean and Canary Islands (5 species), Caribbean (6 species), the tropical South-West Pacific (6 species), Madagascar (1 species), and the Falkland Islands (2 species). The addition of four species from Southern California expands this range considerably, and makes the group more accessible to researchers in this geographic region. It is my hope that this will lead to an improved understanding of the ecology and evolution of these interesting and understudied animals.

Acknowledgements

I am grateful for the help and support of many people in UCSB's Marine Science Institute and Diving & Boating Program, especially Robert Miller, Clint Nelson, Christoph Pierre, Frankie Puerzer, Christian Orsini, and H. Mark Page. Steve Lonhart (NOAA) and Shannon Myers (UCSC) were instrumental in facilitating collections in Central California, and the Natural History Museum of Los Angeles' DISCO program facilitated collections in Los Angeles County. I'd also like to thank the editor and two anonymous reviewers for their help in improving the paper.

Funding declaration

Financial support was provided by UCSB and by the National Aeronautics and Space Administration Biodiversity and Ecological Forecasting Program (Grant NNX14AR62A); the Bureau of Ocean Energy Management Environmental Studies Program (BOEM Agreement MC15AC00006); the National Oceanic and Atmospheric Administration in support of the Santa Barbara Channel Marine Biodiversity Observation Network; and the U.S. National Science Foundation in support of the Santa Barbara Coastal Long Term Ecological Research program under Awards OCE-9982105, OCE-0620276, OCE-1232779, OCE-1831937. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

Avilés, E., Rodríguez, A.D. & Vicente, J. (2013) Two rare-class tricyclic diterpenes with antitubercular activity from the Caribbean sponge *Svenzea flava*. Application of vibrational circular dichroism spectroscopy for determining absolute configuration. *Journal of Organic Chemistry*, 78, 11294–11301. https://doi.org/10.1021/jo401846m

Bakus, G.J. & Green, K.D. (1987) The distribution of marine sponges collected from the 1976-1978 Bureau of Land Management Southern California Bight program. *Bulletin of the Southern California Academy of Sciences*, 86, 57–88.

Bibiloni, M.A. (1993) Some new or poorly known sponges of the Balearic Islands (Western Mediterranean). *Scientia Marina*, 57, 307–318.

Blanchette, C., Miner, C., Raimondi, P., Lohse, D., Heady, K. & Broitman, B. (2008) Biogeographical patterns of rocky intertidal communities along the Pacific coast of North America. *Journal of Biogeography*, 35, 1593–1607. https://doi.org/10.1111/j.1365-2699.2008.01913.x

Blanquer, A. & Uriz, M.-J. (2008) "A posteriori" searching for phenotypic characters to described new cryptic species of sponges revealed by molecular markers (*Scopalina*: Dictyonellidae). *Invertebrate Systematics*, 22, 489–502. https://doi.org/10.1071/IS07004

Caselle, J.E., Davis, K. & Marks, L.M. (2018) Marine management affects the invasion success of a non-native species in a temperate reef system in California, USA. *Ecology Letters*, 21, 43–53. https://doi.org/10.1111/ele.12869

Castorini, M.C.N., Reed, D.C. & Miller, R.J. (2018) Loss of foundation species: disturbance frequency outweighs severity in structuring kelp forest communities. *Ecology*, 99, 2442–2454. https://doi.org/10.1002/ecy.2485

Carballo, J., Cruz-Barraza, J., Vega, C., Nava, H. & Chávez-Fuentes, M. (2019) Sponge diversity in Eastern Tropical Pacific coral reefs: an interoceanic comparison. *Scientific Reports*, 9, 9409. https://doi.org/10.1038/s41598-019-45834-4

Chombard, C., Boury-Esnault, N. & Tillier, S. (1998) Reassessment of homology of morphological characters in tetractinellid sponges based on molecular data. *Systematic Biology*, 47, 351–366. https://doi.org/10.1080/106351598260761

Eger, A.M., Marzinelli, E., Gribben, P., Johnson, C.R., Layton, C., Steinberg, P.D., Wood, G., Silliman, B.R. & Vergés, A. (2020) Playing to the positives: Using synergies to enhance kelp forest restoration. *Frontiers in Marine Science*, 7, 544. https://doi.org/10.3389/fmars.2020.00544

Erpenbeck, D., Breeuwer, J., van der Velde, H. & van Soest, R. (2002) Unravelling host and symbiont phylogenies of halichondrid sponges (Demospongiae, Porifera) using a mitochondrial marker. *Marine Biology*, 141, 377–386. https://doi.org/10.1007/s00227-002-0785-x

Erpenbeck, D., Breeuwer, J.A.J., Parra-Velandia, F.J. & van Soest, R.W.M. (2006) Speculation with spiculation?—Three independent gene fragments and biochemical characters versus morphology in demosponge higher classification. *Molecular Phylogenetics and Evolution*, 38, 293–305. https://doi.org/10.1016/j.ympev.2005.11.001

Erpenbeck, D., Duran, S., Rützler, K., Paul, V., Hooper, J. & Wörheide, G. (2007a) Towards a DNA taxonomy of Caribbean demosponges: a gene tree reconstructed from partial mitochondrial CO1 gene sequences supports previous rDNA phylogenies and provides a new perspective on the systematics of Demospongiae. *Journal of the Marine Biological Association of the United Kingdom*, 87, 1563–1570. https://doi.org/10.1017/S0025315407058195

Erpenbeck, D., Hall, K., Alvarez, B., Büttner, G., Sacher, K., Schätzle, S., Schuster, A., Vargas, S., Hooper, J. & Wörheide, G. (2012) The phylogeny of halichondrid demosponges: past and present re-visited with DNA-barcoding data. *Organisms Diversity & Evolution*, 12, 57–70. https://doi.org/10.1007/s13127-011-0068-9

Erpenbeck, D., List-Armitage, S., Alvarez, B., Degnan, B., Wörheide, G. & Hooper, J. (2007b) The systematics of Raspailiidae (Demospongiae: Poecilosclerida: Microcionina) re-analysed with a ribosomal marker. *Journal of the Marine Biological Association of the United Kingdom*, 87, 1571–1576. https://doi.org/10.1017/S0025315407058201

Erpenbeck, D., Voigt, O., Al-Aidaroos, A., Berumen, M., Büttner, G., Catania, D., Guirguis, A., Paulay, G., Schätzle, S. & Wörheide, G. (2016) Molecular biodiversity of Red Sea demosponges. *Marine Pollution Bulletin*, 105, 507–514. https://doi.org/10.1016/j.marpolbul.2015.12.004

Goodwin, C., Jones, J., Neely, K. & Brickle, P. (2011) Sponge biodiversity of the Jason Islands and Stanley, Falkland Islands with descriptions of twelve new species. *Journal of the Marine Biological Association of the United Kingdom*, 91, 275–301. https://doi.org/10.1017/S0025315410001542

Goodwin, C.E. & Picton, B.E. (2009) Demosponges of the genus *Hymedesmia* (Poecilosclerida: Hymedesmidae) from Rathlin Island, Northern Ireland, with a description of six new species. *Zoological Journal of the Linnean Society*, 156, 896–912. https://doi.org/10.1111/j.1096-3642.2008.00498.x

Green, K.D. & Bakus, G.J. (1994) *Taxonomic atlas of the benthic marine fauna of the Western Santa Maria Basin and the*

Western Santa Barbara Channel. Vol. 2. The Porifera. The Santa Barbara Museum of Natural History, Santa Barbara, California, 82 pp.
Hechtel, G.J. (1965) A systematic study of the Demospongiae of Port Royal, Jamaica. *Bulletin of the Peabody Museum of Natural History*, 20, 1–103.
Hoang, D., Chernomor, O., von Haeseler, A., Minh, B. & Vinh, L. (2018) UFBoot2: Improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution*, 35, 518–522.
https://doi.org/10.1093/molbev/msx281
Huang, D., Meier, R., Todd, P.A. & Chou, L.M. (2008) Slow mitochondrial COI sequence evolution at the base of the Metazoan tree and its implications for DNA barcoding. *Journal of Molecular Evolution*, 66, 167–174.
https://doi.org/10.1007/s00239-008-9069-5
Kandler, N., Wooster, M., Leray, M., Knowlton, N., de Voogd, N., Paulay, G. & Berumen, M. (2019) Hyperdiverse macrofauna communities associated with a common sponge, *Stylissa carteri*, shift across ecological gradients in the Central Red Sea. *Diversity*, 11, 18.
https://doi.org/10.3390/d11020018
de Laubenfels, M.W. (1932) The marine and fresh-water sponges of California. *Proceedings of the United States National Museum*, 81, 1–140.
https://doi.org/10.5479/si.00963801.81-2927.1
de Laubenfels, M.W. (1954) *The sponges of the West-Central Pacific*. Oregon State College, Corvallis, Oregon, 320 pp.
https://doi.org/10.5962/bhl.title.6516
Lavrov, D., Forget, L., Kelly, M. & Lang, B. (2005) Mitochondrial genomes of two demosponges provide insights into an early stage of animal evolution. *Molecular Biology and Evolution*, 22, 1231–1239.
https://doi.org/10.1093/molbev/msi108
Lavrov, D. & Lang, B. (2005) Transfer RNA gene recruitment in mitochondrial DNA. *Trends in Genetics*, 21, 129–133.
https://doi.org/10.1016/j.tig.2005.01.004
Lee, W., Elvin, D. & Reiswig, H. (2007) *The sponges of California: A guide and key to the marine sponges of California*. Monterey Bay Sanctuary Foundation, Monterey, California, 395 pp.
Letunic, I. & Bork, P. (2019) Interactive Tree Of Life (iTOL) v4: recent updates and new developments. *Nucleic Acids Research*, 47, W256–W259.
https://doi.org/10.1093/nar/gkz239
Miller, R.J., Lafferty, K.D., Lamy, T., Kui, L., Rassweiler, A. & Reed, D.C. (2018) Giant kelp, *Macrocystis pyrifera*, increases faunal diversity through physical engineering. *Proceedings of the Royal Society B*, 285, 20172571.
https://doi.org/10.1098/rspb.2017.2571
Montalvo, N. & Hill, R. (2011) *Applied Environmental Microbiology*, 77, 7207–7216.
https://doi.org/10.1128/AEM.05285-11
Morrow, C. & Cárdenas, P. (2015) Proposal for a revised classification of the Demospongiae (Porifera). *Frontiers in Zoology*, 12, 1–27.
https://doi.org/10.1186/s12983-015-0099-8
Morrow, C.C., Picton, B.E., Erpenbeck, D., Boury-Esnault, N., Maggs, C.A. & Allcock, A.L. (2012) Congruence between nuclear and mitochondrial genes in Demospongiae: A new hypothesis for relationships within the G4 clade (Porifera: Demospongiae). *Molecular Phylogenetics and Evolution*, 62, 174–190.
https://doi.org/10.1016/j.ympev.2011.09.016
Morrow, C.C., Redmond, N.E., Picton, B.E., Thacker, R.W., Collins, A.G., Maggs, C.A., Sigwart, J.D. & Allcock, A.L. (2013) Molecular phylogenies support homoplasy of multiple morphological characters used in the taxonomy of Heteroscleromorpha (Porifera: Demospongiae). *Integrative and Comparative Biology*, 53, 428–446.
https://doi.org/10.1093/icb/ict065
Nguyen, L.-T., Schmidt, H., von Haeseler, A. & Minh, B. (2015) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology and Evolution*, 32, 268–274.
https://doi.org/10.1093/molbev/msu300
Nichols, S. (2005) An evaluation of support for order-level monophyly and interrelationships within the class Demospongiae using partial data from the large subunit rDNA and cytochrome oxidase subunit I. *Molecular Phylogenetics and Evolution*, 34, 81–96.
https://doi.org/10.1016/j.ympev.2004.08.019
Pett, W. & Lavrov, D. (2015) Cytonuclear interactions in the evolution of animal mitochondrial tRNA metabolism. *Genome Biology and Evolution*, 7, 2089–2101.
https://doi.org/10.1093/gbe/evv124
Pulitzer-Finali, G. (1982) Some new or little-known sponges from the Great Barrier Reef of Australia. *Bollettino dei Musei e degli Istituti Biologici dell'Universitá di Genova*, 48–49, 87–141.
Reed, D., Washburn, L., Rassweiler, A., Miller, R., Bell, T. & Harrer, S. (2016) Extreme warming challenges sentinel status of kelp forests as indicators of climate change. *Nature Communications*, 7, 13757.
https://doi.org/10.1038/ncomms13757
Riesgo, A., Novo, M., Sharma, P., Peterson, M., Maldonado, M. & Giribet, G. (2013) Inferring the ancestral sexuality and re-

productive condition in sponges (Porifera). *Zoologica Scripta*, 43, 101–117.
https://doi.org/10.1111/zsc.12031
Rot, C., Goldfarb, I., Ilan, M. & Huchon, D. (2006) Putative cross-kingdom horizontal gene transfer in sponge (Porifera) mitochondria. *BMC Evolutionary Biology*, 6, 1–11.
https://doi.org/10.1186/1471-2148-6-71
Rützler, K., van Soest, R., & Alvarez, B. (2003) *Svenzea zeai*, a Carribean reef sponge with giant larva, and *Scopalina ruetzleri*: a comparative fine-structural appraoch to classification (Demospongiae, Halichondrida, Dicyonellidae. *Invertebrate Biology*, 122, 203–222.
https://doi.org/10.1111/j.1744-7410.2003.tb00085.x
Schmidt, O. (1862) *Die Spongien des adriatischen Meeres*. Wilhelm Engelmann, Leipzig, 88 pp.
Schneider, C., Rasband, W. & Eliceiri, K. (2012) NIH Image to ImageJ: 25 years of image analysis. *Natue Methods*, 9, 671–675.
https://doi.org/10.1038/nmeth.2089
Sim, C.J. & Bakus, G.J. (1986) Marine sponges of Santa Catalina Island, California. *Occasional Papers of the Allan Hancock Foundation*, 5, 1–23.
van Soest, R.W.M., Boury-Esnault, N., Hooper, J.N.A., Rützler, K., de Voogd, N.J., Alvarez, B., Hajdu, E., Pisera, A.B., Manconi, R., Schönberg, C., Klautau, M., Kelly, M., Vacelet, J., Dohrmann, M., Díaz, M.-C., Cárdenas, P., Carballo, J.L., Ríos, P., Downey, R. & Morrow, C.C. (2019) World Porifera Database. Available from: http://www.marinespecies.org/porifera (accessed 1 September 2019)
https://doi.org/10.14284/359
Thacker, R., Hill, A., Hill, M., Redmond, N., Collins, A., Morrow, C., Spicer, L., Carmack, C., Zappe, M., Pohlmann, D., Hall, C., Diaz, M. & Bangalore, P. (2013) Nearly complete 28S rRNA gene sequences confirm new hypotheses of sponge evolution. *Integrative and Comparative Biology*, 53, 373–387.
https://doi.org/10.1093/icb/ict071
Trifinopoulos, J., Nguyen, L.-T., von Haeseler, A. & Minh, B. (2016) W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Research*, 44, W232–W235.
https://doi.org/10.1093/nar/gkw256
Turner, T.L. (2020a) The marine sponge *Hymeniacidon perlevis* is a globally-distributed invasive species. *Aquatic Invasions*, 15, 542–561.
https://doi.org/10.3391/ai.2020.15.4.01
Turner, T.L. (2020b) The order Tethyida (Porifera) in California: taxonomy, systematics, and the first member of the family Hemiasterellidae in the Eastern Pacific. *Zootaxa*, 4861 (2), 211–231.
https://doi.org/10.11646/zootaxa.4861.2.3
Valentine, J. (1966) Numerical analysis of marine molluscan ranges on extratropical northeastern Pacific shelf. *Limnology and Oceanography*, 11, 198–211.
https://doi.org/10.4319/lo.1966.11.2.0198
Vicente, J., Stewart, A.K., van Wagoner, R.M., Elliott, E., Bourdelais, A.J. & Wright, J.L.C. (2015) Monacyclinones, new angucyclinone metabolites isolated from *Streptomyces* sp. M7_15 associated with the Puerto Rican sponge *Scopalina ruetzleri*. *Marine Drugs*, 13, 4682–4700.
https://doi.org/10.3390/md13084682
Wei, X., Rodríguez, A.D., Wang, Y. & Franzblau, S.G. (2007) Novel ring B abeo-sterols as growth inhibitors of Mycobacterium tuberculosis isolated from a Caribbean Sea sponge, *Svenzea zeai*. *Tetrahedron Letters*, 48, 8851–8854.
https://doi.org/10.1016/j.tetlet.2007.10.070
Wiedenmayer, F. (1977) Shallow-water sponges of the western Bahamas. *Experientia Supplementum*, 28, 1–287.
https://doi.org/10.1007/978-3-0348-5797-0