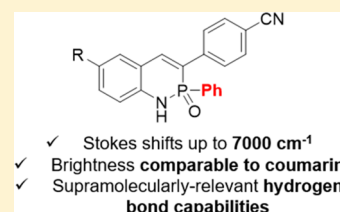


Amplification of the Quantum Yields of 2- $\lambda^5$ -Phosphaquinolin-2-ones through Phosphorus Center ModificationJeremy P. Bard,<sup>†</sup> Hannah J. Bates,<sup>†</sup> Chun-Lin Deng,<sup>†</sup> Lev N. Zakharov,<sup>‡</sup> Darren W. Johnson,<sup>\*,†</sup> and Michael M. Haley<sup>\*,†</sup><sup>†</sup>Department of Chemistry & Biochemistry and the Materials Science Institute, University of Oregon, Eugene, Oregon 97403-1253, United States<sup>‡</sup>CAMCOR, University of Oregon, Eugene, Oregon 97403-1433, United States

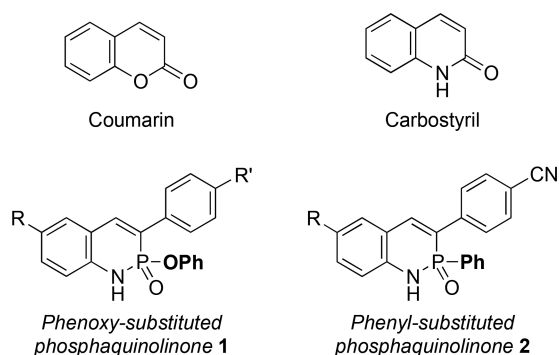
## Supporting Information

**ABSTRACT:** We report the synthesis and characterization of P-phenyl modified phosphorus- and nitrogen-containing phosphaquinolinone heterocycles. The change from –OPh to –Ph results in a marked increase in the quantum yield of the scaffold as well as a moderate red-shifting of the emission. While calculations suggest that  $\pi$  to  $\pi^*$  transitions are dominant, intramolecular charge transfer (ICT) also contributes in the excited state. Solution- and solid-state studies of the dimerization of this new congener to the P-phenoxy variant are also reported, showing retention of the dimerization behavior in this scaffold.



## INTRODUCTION

Small molecule fluorophores are used ubiquitously throughout many different fields, including chemical biology, molecular probe development, and materials for industrial and environmental sensing.<sup>1–4</sup> In many of these applications, a fluorophore must exhibit a few characteristics to be considered optimal: large Stokes shift, high brightness, and red-shifted emission. One such example of a molecule that meets these specifications is coumarin (Figure 1).<sup>5–14</sup> An impressive number of



**Figure 1.** Well-studied coumarin and carbostyryl scaffolds (top) compared to phosphaquinolinone analogues (bottom).

coumarin-containing compounds have been reported throughout the literature that are collected either from in-lab syntheses or by isolation from natural sources.<sup>15,16</sup> This scaffold has been the subject of a variety of synthetic modifications, diversification by adding groups onto the backbone, or incorporation of the coumarin system into larger ring networks.<sup>17–19</sup> Through these modifications, a tremendous breadth of understanding upon the structure–activity relations has been developed<sup>20–24</sup> that have guided the design of many useful derivatives, which

have emerged for applications in chemosensing and many other areas.<sup>25–31</sup>

Alongside the many derivatives of the parent coumarin scaffold, there is the nitrogen-containing structural analogue known as carbostyryl (Figure 1).<sup>32–37</sup> Though not as widely utilized as coumarin, carbostyryl is the subject of many structure–property relationship studies, and it shows promise for use in both pharmaceutical discovery and fluorescence imaging applications.<sup>38,39</sup> These carbostyryl analogues expand on the applications of the coumarin family through modifying the lactone core to a lactam. With further alteration of this core, new applications, functionality, and fluorescent properties are expected from this widely used fluorophore.

Recently, we reported a series of phosphorus- and nitrogen-containing (PN) phosphaquinolinone 1 derivatives (Figure 1).<sup>40,41</sup> This scaffold, which is one of only a handful of similar heterocycles,<sup>42–52</sup> is also an isostere to carbostyryl and coumarin, with the only difference being the replacement of the lactam carbonyl with an isolobal, chiral phosphorus center. We have performed a variety of structure–property studies that looked at the effects of both acene core modification and substitution at various points on the scaffold.<sup>40,41,53–56</sup> In these studies, it was found that the emission wavelength can be moderately red-shifted through careful substitution of various groups on the backbone, affording significant Stokes shifts and modest quantum yields. On the basis of these design principles, this moiety has recently been implemented in a fluorescent receptor for HSO<sub>4</sub><sup>–</sup> in acidic media, showing promise for future applications of this scaffold that take advantage of both its exceptional hydrogen bonding

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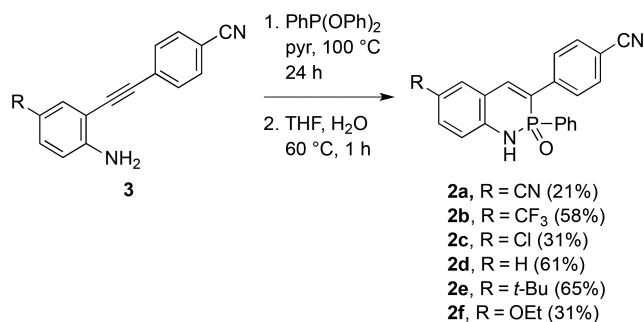
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capabilities and its inherent fluorescence.<sup>54</sup> As clearly shown for the coumarin and carbostyryl motifs, systematic modification of various structural aspects can lead to very useful derivatives. In our continuing efforts to study the 2- $\lambda^5$ -phosphaquinolin-2-one skeleton, the next facet that we wanted to explore was the variation of the group attached to the phosphorus center. Disclosed herein is the substitution of a phenyl ring in place of the standard phenoxy upon the phosphorus center, generating a racemic mixture of heterocycle **2** (Figure 1). We also hypothesize that this would increase the quantum yield through rigidifying the scaffold. With these modifications, the reported compounds could have greater potential for applications in phosphorus-containing chemosensors and fluorophores, expanding on this pre-existing group of molecules.<sup>57–65</sup>

## RESULTS AND DISCUSSION

The synthesis of **2** starts from key arylethynylaniline intermediate **3**, prepared following previously reported methods.<sup>40</sup> Aniline **3** is then reacted with diphenyl phenylphosphonite (PhP(OPh)<sub>2</sub>) in pyridine at 100 °C. Subsequent hydrolysis in THF at 60 °C furnishes phenyl-appended

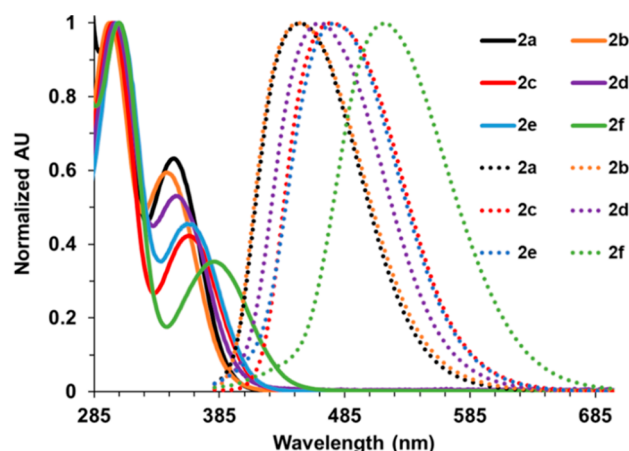
### Scheme 1. Synthesis of Phosphaquinolinones **2**



heterocycles **2** in modest to good yields (Scheme 1). <sup>1</sup>H, <sup>13</sup>C, <sup>31</sup>P, and <sup>19</sup>F NMR spectra were collected for **2** (see Supporting Information), which reveal that the direct attachment of the phenyl ring to the phosphorus center not only splits the signals of the phenyl ring (<sup>3</sup>J<sub>P,H</sub> and <sup>1</sup>J<sub>P,<sup>13</sup>C</sub> values listed in the Experimental Section) but also affords coupling constants of *ca.* 30 Hz (<sup>3</sup>J<sub>P,H</sub>) for the alkene proton signal.

The photophysical properties of **2a–2f** in CHCl<sub>3</sub> are shown in Figure 2 and are compiled in Table 1. All derivatives share a common  $\lambda_{\text{max}}$  at *ca.* 300 nm, and the lowest energy absorption peaks range from 343 to 381 nm. The absorption coefficients for this scaffold stay within the range of  $1.5 \times 10^4$  (for **2a**) to  $2.2 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$  (for **2e** and **2f**). The  $\lambda_{\text{em}}$  values range from 447 (for **2a**) to 515 nm (for **2f**) with Stokes shifts on the order of 6350–7000 cm<sup>−1</sup>. Interestingly, the emission spectra of **2** show a *ca.* 20 nm bathochromic shift from those of the analogous congeners of **1**,<sup>40</sup> and the quantum yields of this scaffold show a dramatic improvement, on the order of a 4–5-fold increase in most cases. Brightness values range from  $6.84 \times 10^3$  (for **2b**) to  $1.14 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$  (for **2a**), which are now on par with several optimized coumarin derivatives.<sup>66</sup>

Fluorescence lifetime measurements were also performed (Figure S6), and the radiative ( $k_r$ ) and nonradiative ( $k_{\text{nr}}$ ) decay rate constants were determined. The  $k_r$  values range from 0.06 to 0.19 ns<sup>−1</sup>, and the  $k_{\text{nr}}$  values vary from 0.06 to 0.18 ns<sup>−1</sup>,



**Figure 2.** Absorption (solid lines) and fluorescence (dotted lines) spectra of **2** in CHCl<sub>3</sub> at 298 K.

showing either equal rates or a slightly larger  $k_{\text{nr}}$  in most cases. These values elucidate a potential explanation for the increased quantum yields when compared to similar values of phosphaquinolinones **1**.<sup>40</sup> For **1**, the  $k_r$  values range from 0.10 to 0.30 ns<sup>−1</sup>, which show similar values, whereas the  $k_{\text{nr}}$  values vary from 0.30 to 3.0 ns<sup>−1</sup>, which are substantially faster in most cases. The diminished ratio of  $k_{\text{nr}}$  to  $k_r$  seen for **2** may suggest that the reduced degrees of freedom may indeed be the cause of the increased quantum yields.

To gain further understanding of these experimental results, the frontier orbitals for heterocycles **2** were calculated (Tables 1 and S1–S7). The narrowest HOMO–LUMO gap is seen for **2f** (3.90 eV), and the largest is for **2a** (4.40 eV). This trend arises because of a higher magnitude of HOMO destabilization than that of the stabilization of the LUMO with more donating substituents (Table S1). These values also follow a similar trend to the optical gaps (Table 1). TD-DFT was then used to examine the  $S_0$  to  $S_1$  transition. It was found that the  $S_0$  to  $S_1$  transition is dominated by the HOMO–LUMO transition (Table S1). Additionally, the distributions of the HOMOs and LUMOs show a slightly more pronounced separation due to a larger HOMO localization at the phosphorus center (Figure S4). These observations suggest that  $\pi$  to  $\pi^*$  transitions are dominant, but there may be some intramolecular charge transfer (ICT) occurring in the excited state. The emission of **2** was then examined in solvents of varying polarities (Figures S7–S13 and Tables S11–S16). In these studies, bathochromic shifting is observed in every case with more polar solvents. Compound **2f** showed the greatest shifting, ranging from emission wavelengths of 504 to 531 nm and Stokes shifts of 6070 and 7070 cm<sup>−1</sup> in cyclohexane and acetonitrile, respectively.

The TD-DFT optimized  $S_1$  state near the Franck–Condon geometry of **2f** shows that the dihedral angle between the parent core and appended 4-cyanophenyl substituent becomes smaller compared to the ground state (Figure 3). Additionally, the C–C bond connecting them is shortened by *ca.* 0.035 Å. The considerable geometric changes lead to a more conjugated system at the  $S_1$  state, in which the HOMO–LUMO energy gap decreases by 0.9 eV (Figure S5); thus, the computed emission wavelength of 541 nm is within the observed emission maxima (Table 1). These computational results could further explain the large Stokes shift for this type of PN-heterocycle.

Table 1. Photophysical Properties and HOMO–LUMO Energy Gaps of Heterocycles 2<sup>a</sup>

compd	$\lambda_{\text{abs}}$ (nm)/ $\epsilon$ (M <sup>-1</sup> cm <sup>-1</sup> )	$\lambda_{\text{em}}$ (nm)/ $\phi$ (%)	Stokes shift (cm <sup>-1</sup> )	$\tau^b$ (ns)	$k_r$ (ns <sup>-1</sup> )	$k_{\text{nr}}$ (ns <sup>-1</sup> )	$\Delta E_{\text{opt}}$ (eV)	$\Delta E_{\text{DFT}}$ (eV) <sup>c</sup>
2a	348/15000	447/76	6360	4.1	0.19	0.06	3.19	4.40
2b	343/18000	449/38	6880	3.5	0.11	0.18	3.18	4.43
2c	360/19000	474/43	6680	4.8	0.09	0.12	3.05	4.23
2d	352/18000	467/50	7000	3.9	0.13	0.13	3.15	4.30
2e	360/22000	475/51	6730	4.2	0.12	0.12	3.04	4.21
2f	381/22000	515/35	6830	6.3	0.06	0.10	2.83	3.90

<sup>a</sup>All values collected in CHCl<sub>3</sub>. <sup>b</sup>Decay curves fitted using a monoexponential fitting model. <sup>c</sup>Calculated at the PBE0/TZVP level of theory.

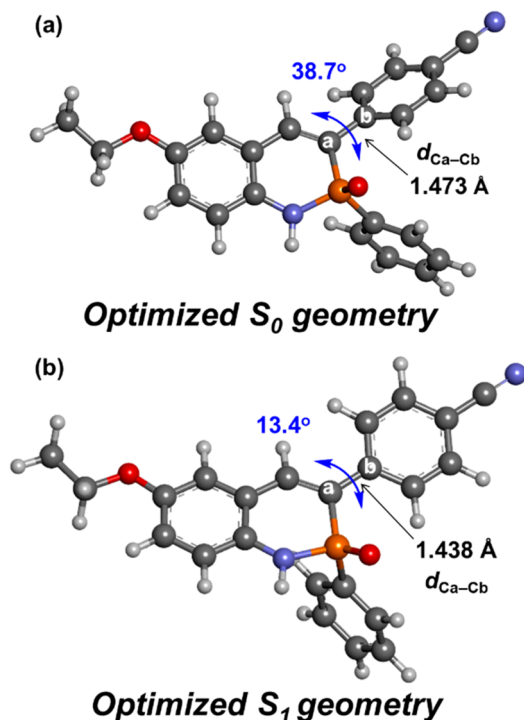


Figure 3. Selected bond length and dihedral angle in the optimized  $S_0$  and  $S_1$  structures of **2f** calculated by DFT and TD-DFT methods at the PCM(CHCl<sub>3</sub>)-PBE0/TZVP level of theory, respectively.

In addition to improved fluorescence properties, we were curious to see how phenyl substitution would affect the strength of hydrogen bond dimerization we typically observe for phosphaquinolones. Variable concentration (VC) NMR experiments were performed in water-saturated CDCl<sub>3</sub> to assess the strength of dimerization (Tables 2 and S17–S21, Figures S14–S23). Heterocycles **2** exhibit dimerization strengths of 22 (for **2e** and **2f**) to 82 M<sup>-1</sup> (for **2b**). While these values are roughly 70–80% smaller than those measured for the analogous congeners of **1**, the strengths of dimerization

Table 2. Dimerization Constants and Energies for **2**

compd	$K_{\text{dim}}$ (M <sup>-1</sup> )	$\Delta G_{\text{dim}}$ (kcal mol <sup>-1</sup> )
2a <sup>a</sup>		
2b	82	−2.6
2c	54	−2.3
2d	24	−1.8
2e	22	−1.8
2f	22	−1.8

<sup>a</sup>Not determined because of minimal solubility in H<sub>2</sub>O-saturated CDCl<sub>3</sub>. Values reported with errors less than 15%.

for **2** again exceed those of many typical head-to-tail hydrogen bonded dimers.<sup>67</sup> This result suggests that this new entry into the phosphaquinolone family can still be implemented in supramolecular systems, as found in **1**.<sup>54</sup>

Single crystals suitable for X-ray diffraction were grown by slowly diffusing pentane into a CHCl<sub>3</sub> solution of **2f**, and the resultant data are shown in Figures 4 and S1–S3. The

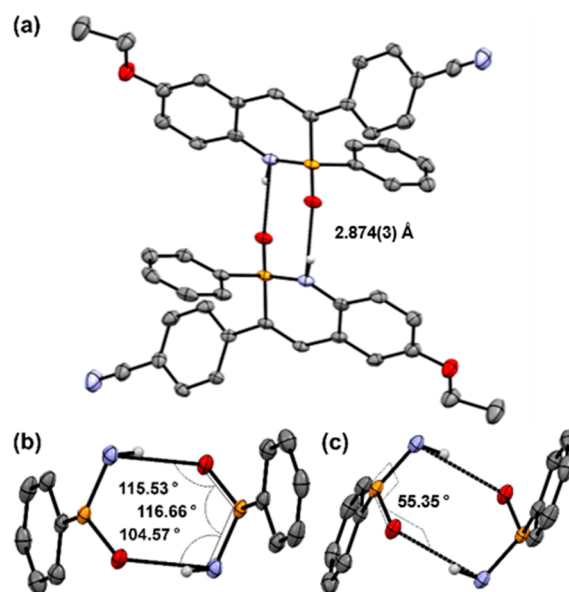


Figure 4. (a) Characteristic PN-heterocycle dimer for **2f** with the O...N distance (Å) shown as well as (b) bond angles and (c) torsional angles formed within monomers upon dimerization. Ellipsoids drawn at 30% probability.

structure of **2f** still features the typical *meso*-dimer between racemates (Figure 4a); however, the N...O distance in the dimer of **2f** (2.874 Å) is longer than those of heterocycles **1** (2.768–2.821 Å),<sup>40</sup> which supports the observation that the molecule should form a weaker dimer in the solution-state as well. This weakened hydrogen bonding interaction can potentially be explained by examining the pseudo six-membered ring formed between the monomers. The N...O–P (115.53°), O–P–N (116.66°), and P–N...O (104.57°) angles formed between the participating atoms in the dimer formation show significant deviation from the ideal 120° orientation (Figure 4b), likely caused by the large O–P–N–H torsional angle of 55.35° (Figure 4c). With an angle so much larger than the analogous angle found in the crystal structures of several derivatives of **1** (*ca.* 30–40°),<sup>40</sup> there is a less ideal orientation for the two monomers to associate, slightly weakening the interaction overall. By comparing optimized geometries of the *meso*-dimer of **2f** and its –OPh analogue, there are some additional steric clashes in **2f** among the C–H



atoms in the phenyl ring and N–H moieties, according to the noncovalent interactions (NCI) plot (Figure S24). Moreover, the natural bond orbital (NBO) analyses predict a total contribution of the  $n_{\text{O}} \rightarrow \sigma_{\text{NH}}^*$  interactions of 23.1 kcal mol<sup>-1</sup> for **2f** and 25.0 kcal mol<sup>-1</sup> for the respective –Oph analogue (Figure S25). Therefore, the strength of dimerization may decrease to some degree due to the weaker primary hydrogen bonding and extra steric hindrance, in agreement with the observed diminished  $K_{\text{dim}}$  for **2**.

## CONCLUSION

In summary, we have shown the effects of the attachment of a phenyl group on the phosphorus center of the phosphaquinolone scaffold. This new class of PN-heterocycles not only has large Stokes shift values (up to 7000 cm<sup>-1</sup>) but also shows a marked 4–5-fold increase in the quantum yield when compared to previously reported phenoxy-substituted compounds. Additionally, this modification retains the strong dimerization strengths of the scaffold in both the solid and solution states. This new modification deepens the fundamental understanding of the phosphaquinolone scaffold and allows for further possibilities in the applications of this scaffold as a biologically or industrially relevant fluorophore, like the coumarin and carbostyryl scaffolds.

## EXPERIMENTAL SECTION

**General.** All air- or water-free reactions were performed under a N<sub>2</sub> atmosphere using Schlenk techniques. Column chromatography was performed using silica gel (240–300 mesh), with solvent systems being referenced to the most abundant solvent. NMR spectra were acquired at room temperature on a Varian Inova 500 instrument (<sup>1</sup>H: 500 MHz, <sup>13</sup>C: 126 MHz, <sup>19</sup>F: 471 MHz, <sup>31</sup>P: 202 MHz) or a Bruker Avance III HD 500 apparatus equipped with a Prodigy multinuclear cryoprobe (<sup>1</sup>H: 500 MHz, <sup>13</sup>C: 126 MHz). <sup>1</sup>H and <sup>13</sup>C chemical shifts ( $\delta$ ) are expressed in parts per million (ppm) relative to residual CHCl<sub>3</sub> shifts (<sup>1</sup>H: 7.26 ppm, <sup>13</sup>C: 77.16 ppm) or residual DMSO shifts (<sup>1</sup>H: 2.50 ppm, <sup>13</sup>C: 39.52 ppm). <sup>31</sup>P and <sup>19</sup>F NMR spectra are referenced to 85% H<sub>3</sub>PO<sub>4</sub> ( $\delta$  0 ppm) and to CFCl<sub>3</sub> ( $\delta$  0 ppm), respectively, as the external standards. UV–vis spectra were recorded using an Agilent Technologies Cary 60 UV–vis spectrophotometer in HPLC-grade CHCl<sub>3</sub>. Fluorescence emission spectra were recorded using a Horiba Jobin Yvon FluoroMax-4 fluorimeter exciting at 365 nm. Quantum yields ( $\phi$ ) were determined through a comparison of the emission and absorption intensities of the analyte to that of a 0.1 M H<sub>2</sub>SO<sub>4</sub>/quinine sulfate solution.<sup>68</sup> Fluorescence lifetime measurements were recorded using a Horiba FluoroHub Single Photon Counting Controller with a TemPro Fluorescence Lifetime System attachment. High-resolution mass spectra (HRMS) were recorded on a Waters XEVO G2-XS mass spectrometer. 2-Ethynylanilines **3a–3f**<sup>40</sup> and phenyl diphenylphosphonite (PhP(OPh)<sub>2</sub>)<sup>69</sup> were prepared as previously described.

**General Synthetic Procedure for Phosphaquinolone 2.** 2-Ethynylaniline **3** (1.0 equiv) and PhP(OPh)<sub>2</sub> (2.0 equiv) were dissolved in pyridine (ca. 0.35 M). The vessel was sealed and heated to 100 °C for 24 h in an oil bath. The mixture was then diluted with toluene, and the solvent was removed *in vacuo*. This was repeated three times to remove all residual pyridine. The crude material was dissolved in THF, and ca. five drops of water were added. The solution was stirred at 60 °C for 1 h before being dried (Na<sub>2</sub>SO<sub>4</sub>), filtered, and concentrated *in vacuo*. The crude mixture was then purified by column chromatography on silica gel. Reported yields are given for >95% pure material (by <sup>1</sup>H NMR spectroscopy), though subsequent recrystallization from hexanes and CH<sub>2</sub>Cl<sub>2</sub> was used to achieve analytically pure material.

**Phosphaquinolone 2a.** Compound **2a** was synthesized from **3a** (462 mg, 1.9 mmol, 1 equiv) and PhP(OPh)<sub>2</sub> (1.11 g, 3.8 mmol, 2 equiv). Column chromatography (1:1 EtOAc:CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  = 0.20) gave

**2a** (149 mg, 21%) as a pale brown solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.28 (d,  $J$  = 3.8 Hz, 1H), 8.14 (d,  $J$  = 2.0 Hz, 1H), 8.05 (d,  $J$  = 30.1 Hz, 1H), 7.82 (ABm,  $J$  = 8.4 Hz, 4H), 7.81–7.77 (m, 1H), 7.69–7.64 (m, 2H), 7.58–7.51 (m, 1H), 7.49–7.43 (m, 2H), 7.21 (d,  $J$  = 8.5 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  143.1 (d,  $J$  = 3.8 Hz), 140.9 (d,  $J$  = 11.7 Hz), 139.3, 135.8, 133.8, 132.6, 132.5 (d,  $J$  = 2.7 Hz), 132.4 (d,  $J$  = 137.0 Hz), 132.2 (d,  $J$  = 10.8 Hz), 128.6 (d,  $J$  = 13.2 Hz), 128.1 (d,  $J$  = 6.1 Hz), 126.9 (d,  $J$  = 115.9 Hz), 119.2, 119.0 (d,  $J$  = 9.9 Hz), 118.5, 117.7 (d,  $J$  = 8.1 Hz), 110.6, 102.1; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.77; HRMS (ASAP) [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>15</sub>N<sub>3</sub>OP 368.0953, found 368.0977.

**Phosphaquinolone 2b.** Compound **2b** was synthesized from **3b** (700 mg, 2.4 mmol, 1 equiv) and PhP(OPh)<sub>2</sub> (1.40 g, 4.8 mmol, 2 equiv). Recrystallization from CH<sub>2</sub>Cl<sub>2</sub> and hexanes gave **2b** (580 mg, 58%) as a yellow solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  10.14 (d,  $J$  = 4.1 Hz, 1H), 8.16 (d,  $J$  = 30.1 Hz, 1H), 8.06 (d,  $J$  = 2.2 Hz, 1H), 7.85 (d,  $J$  = 8.3 Hz, 2H), 7.80 (d,  $J$  = 8.4 Hz, 2H), 7.73–7.63 (m, 3H), 7.53 (td,  $J$  = 7.4, 1.5 Hz, 1H), 7.49–7.43 (m, 2H), 7.25 (d,  $J$  = 8.5 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  142.5 (d,  $J$  = 3.7 Hz), 141.1 (d,  $J$  = 12.0 Hz), 139.9, 132.7 (d,  $J$  = 136.9 Hz), 132.6, 132.4 (d,  $J$  = 2.7 Hz), 132.2 (d,  $J$  = 10.7 Hz), 130.5, 128.5 (d,  $J$  = 13.2 Hz), 128.1 (d,  $J$  = 6.1 Hz), 127.4 (d,  $J$  = 3.7 Hz), 126.5 (d,  $J$  = 116.3 Hz), 124.5 (q,  $J$  = 271.2 Hz), 120.6 (q,  $J$  = 32.3 Hz), 118.6 (d,  $J$  = 13.0 Hz), 118.5, 117.4 (d,  $J$  = 7.9 Hz), 110.5; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.86; <sup>19</sup>F NMR (471 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  –59.92; HRMS (ASAP) [M + H]<sup>+</sup> calcd for C<sub>22</sub>H<sub>15</sub>N<sub>2</sub>OP<sub>2</sub> 411.0874, found 411.0909.

**Phosphaquinolone 2c.** Compound **2c** was synthesized from **3c** (645 mg, 2.6 mmol, 1 equiv) and PhP(OPh)<sub>2</sub> (1.5 g, 5.1 mmol, 2 equiv). Column chromatography (1:1:1 hexanes:EtOAc:CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  = 0.10) followed by two rounds of recrystallization from CH<sub>2</sub>Cl<sub>2</sub> and hexanes gave **2c** (300 mg, 31%) as a yellow solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  9.80 (d,  $J$  = 4.2 Hz, 1H), 7.99 (d,  $J$  = 29.8 Hz, 1H), 7.83 (d,  $J$  = 8.4 Hz, 2H), 7.78 (d,  $J$  = 8.5 Hz, 2H), 7.73 (d,  $J$  = 2.5 Hz, 1H), 7.69–7.57 (m, 2H), 7.56–7.49 (m, 1H), 7.48–7.38 (m, 3H), 7.11 (d,  $J$  = 8.7 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  141.3 (d,  $J$  = 12.0 Hz), 139.4, 138.3 (d,  $J$  = 3.7 Hz), 132.8 (d,  $J$  = 136.6 Hz), 132.5, 132.3, 132.2, 130.7, 129.9, 128.5 (d,  $J$  = 13.1 Hz), 128.1 (d,  $J$  = 6.2 Hz), 126.5 (d,  $J$  = 116.8 Hz), 123.6, 120.2 (d,  $J$  = 12.6 Hz), 118.6, 118.4 (d,  $J$  = 8.1 Hz), 110.3; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.69; HRMS (ASAP) [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>15</sub>N<sub>2</sub>OPCl 377.0611, found 377.0641.

**Phosphaquinolone 2d.** Compound **2d** was synthesized from **3d** (151 mg, 0.69 mmol, 1 equiv) and PhP(OPh)<sub>2</sub> (463 mg, 1.4 mmol, 2 equiv). Column chromatography (1:1:1 hexanes:EtOAc:CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  = 0.20) gave **2d** (144 mg 61%) as a yellow solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.73 (d,  $J$  = 8.1 Hz, 2H), 7.66 (d,  $J$  = 30.4 Hz, 1H), 7.71–7.66 (m, 2H), 7.54 (d,  $J$  = 8.2 Hz, 2H), 7.47–7.42 (m, 2H), 7.38–7.32 (m, 3H), 7.06 (t,  $J$  = 7.5 Hz, 1H), 6.97 (d,  $J$  = 8.0 Hz, 1H), 6.83 (br s, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  141.5 (d,  $J$  = 11.8 Hz), 141.0 (d,  $J$  = 3.0 Hz), 138.8 (d,  $J$  = 4.3 Hz), 132.7 (d,  $J$  = 10.8 Hz), 132.6 (d,  $J$  = 2.8 Hz), 132.5, 132.2 (d,  $J$  = 139.3 Hz), 131.5, 131.2, 128.6 (d,  $J$  = 13.8 Hz), 128.4 (d,  $J$  = 6.2 Hz), 126.0 (d,  $J$  = 119.5 Hz), 121.4, 119.4 (d,  $J$  = 12.1 Hz), 118.8, 117.3 (d,  $J$  = 7.7 Hz), 111.6; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, CDCl<sub>3</sub>)  $\delta$  10.52; HRMS (ASAP) [M + H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>16</sub>N<sub>2</sub>OP 343.1000, found 343.1030.

**Phosphaquinolone 2e.** Compound **2e** was synthesized from **3e** (549 mg, 2.0 mmol, 1 equiv) and PhP(OPh)<sub>2</sub> (1.3 g, 4.0 mmol, 2 equiv). Column chromatography (1:1:1 EtOAc:CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  = 0.25) gave **2e** (520 mg, 65%) as a yellow solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.73 (d,  $J$  = 8.0 Hz, 2H), 7.72–7.67 (m, 2H), 7.68 (d,  $J$  = 30.3 Hz, 1H), 7.53 (d,  $J$  = 8.1 Hz, 2H), 7.48–7.39 (m, 3H), 7.37–7.32 (m, 2H), 6.90 (d,  $J$  = 8.4 Hz, 1H), 6.48 (br s, 1H), 1.35 (s, 9H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  144.4, 141.7 (d,  $J$  = 12.1 Hz), 141.5, 136.4, 132.7 (d,  $J$  = 10.8 Hz), 132.5, 132.5, 132.4 (d,  $J$  = 139.2 Hz), 129.1, 128.5 (d,  $J$  = 13.6 Hz), 128.4 (d,  $J$  = 6.4 Hz), 127.6, 125.8 (d,  $J$  = 119.9 Hz), 119.0, 118.9 (d,  $J$  = 8.4 Hz), 116.9 (d,  $J$  = 7.3 Hz), 111.4, 34.4, 31.5; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, CDCl<sub>3</sub>)  $\delta$

10.44; HRMS (ASAP)  $[M + H]^+$  calcd for  $C_{25}H_{24}N_2OP$  399.1628, found 399.1629.

**Phosphaquinolinone 2f.** Compound **2f** was synthesized from **3f** (430 mg, 1.7 mmol, 1 equiv) and  $PhP(OPh)_2$  (969 g, 3.3 mmol, 2 equiv). Column chromatography (1:1:1 hexanes:EtOAc:CH<sub>2</sub>Cl<sub>2</sub>,  $R_f$  = 0.25) gave **2f** (200 mg, 31%) as a pale yellow solid: mp > 250 °C; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.73–7.65 (m, 2H), 7.71 (d,  $J$  = 8.6 Hz, 2H), 7.59 (d,  $J$  = 30.4 Hz, 1H), 7.53 (d,  $J$  = 8.2 Hz, 2H), 7.47–7.42 (m, 1H), 7.37–7.31 (m, 2H), 6.98–6.93 (m, 2H), 6.88 (d,  $J$  = 8.5 Hz, 1H), 6.60 (br s, 1H), 4.04 (q,  $J$  = 7.0 Hz, 2H), 1.43 (t,  $J$  = 7.0 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (126 MHz, CDCl<sub>3</sub>) δ 153.5, 141.6 (d,  $J$  = 12.0 Hz), 140.7, 132.8 (d,  $J$  = 10.7 Hz), 132.7, 132.6 (d,  $J$  = 3.0 Hz), 132.5, 132.1 (d,  $J$  = 140.4 Hz), 128.5 (d,  $J$  = 13.8 Hz), 128.4 (d,  $J$  = 6.4 Hz), 126.7 (d,  $J$  = 119.9 Hz), 119.9, 119.8 (d,  $J$  = 11.8 Hz), 118.8, 118.2 (d,  $J$  = 7.7 Hz), 114.8, 111.5, 64.3, 15.0; <sup>31</sup>P{<sup>1</sup>H} NMR (202 MHz, CDCl<sub>3</sub>) δ 10.51; HRMS (ASAP)  $[M + H]^+$  calcd for  $C_{23}H_{20}N_2O_2P$  387.1261, found 387.1283.

## ■ ASSOCIATED CONTENT

### ■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.9b02132.

NMR spectra for all new compounds, X-ray structure data and molecular packing, computational details (including the coordinates of optimized structures), and self-dimerization study data (PDF)

Crystallographic information for **2f** (CCDC No. 1944053) (CIF)

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### Notes

The authors declare no competing financial interest.

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