

Sep 24, 2021

🌐 Measuring Coral Skeletal Architecture Using Image Analysis

Rowan Mclachlan¹, Ashruti Patel¹, Andrea G Grottoli¹¹The Ohio State University

2 Works for me

 Sharedx.doi.org/10.17504/protocols.io.bx5bpq2n[Coral Bleaching RCN protocols](#)

Rowan Mclachlan
The Ohio State University

ABSTRACT

Coral morphology is influenced by genetics, the environment, or the interaction of both, and thus is highly variable. This protocol outlines a non-destructive and relatively simple method for measuring Scleractinian coral sub-corallite skeletal structures (such as the septa length, theca thickness, and corallite diameter, etc.) using digital images produced as a result of digital microscopy or from scanning electron microscopy. This method uses X and Y coordinates of points placed onto photomicrographs to automatically calculate the length and/or diameter of a variety of sub-corallite skeletal structures in the Scleractinian coral *Porites lobata*. However, this protocol can be easily adapted for other coral species - the only difference may be the specific skeletal structures that are measured (for example, not all coral species have a pronounced columella or pali, or even circular corallites). This protocol is adapted from the methods described in Forsman et al. (2015) & Tisthammer et al. (2018).

There are 4 steps to this protocol:

- 1) Removal of Organic Tissue from Coral Skeletons
- 2) Imaging of Coral Skeletons
- 3) Photomicrograph Image Analysis
- 4) Calculation of Corallite Microstructure Size

This protocol was written by Dr. Rowan McLachlan and was reviewed by Ashruti Patel and Dr. Andréa Grottoli.

Acknowledgments

Leica DMS 1000 and Scanning Electron Microscopy photomicrographs used in this protocol were acquired at the Subsurface Energy Materials Characterization and Analysis Laboratory (SEMCAL), School of Earth Sciences at The Ohio State University, Ohio, USA. I would like to thank Dr. Julie Sheets, Dr. Sue Welch, and Dr. David Cole for training me on the use of these instruments.

Tisthammer KR, Richmod RH (2018). Corallite skeletal morphological variation in Hawaiian *Porites lobata*. *Coral Reefs* 37, 445–456.
<https://doi.org/10.1007/s00338-018-1670-5>

Forsman Z, Wellington GM, Fox GE, Toonen RJ (2015). Clues to unraveling the coral species problem: distinguishing species from geographic variation in *Porites* across the Pacific with molecular markers and microstructural traits. *PeerJ* 3:e751.
<https://doi.org/10.7717/peerj.751>

DOI

dx.doi.org/10.17504/protocols.io.bx5bpq2n

PROTOCOL CITATION

Rowan McLachlan, Ashruti Patel, Andrea G Grottoli 2021. Measuring Coral Skeletal Architecture Using Image Analysis. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.bx5bpq2n>



KEYWORDS

morphometrics, morphometry, micromorphology, microstructure, corallite, septa, theca, columella, skeleton, aragonite, microcorallite morphology, coral, coral reef

LICENSE

————— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Sep 10, 2021

LAST MODIFIED

Sep 24, 2021

PROTOCOL INTEGER ID

53123

MATERIALS TEXT

For Removal of organic tissue from coral skeletons:

500 ml plastic beakers
sonicator bath
plastic forceps
aluminum weighing pans
laboratory drying oven
deionized water
sodium hypochlorite bleach (3% concentration)

For Imaging of coral skeletons:

Digital Microscope (e.g., Leica DMS 1000) or Scanning Electron Microscope (SEM)

For Photomicrograph Image Analysis

Computer
ImageJ software

Removal of Organic Tissue from Coral Skeletons

- 1 Remove coral fragments from the freezer (if applicable) and place them in individual 500 ml plastic beakers filled with deionized water. Soak coral fragments for 30 minutes at room temperature to remove excess salt and prevent NaCl precipitation.
- 2 Using plastic forceps, transfer coral fragments to new 500 ml beakers filled with ~ 200 ml of 3% sodium hypochlorite bleach (Fig 1A). Take care not to damage fragile corallite micro-skeleton. Soak for 1 hour at room temperature.
- 3 Replace liquid contents of the beakers with fresh bleach solution and arrange beakers in a sonicator bath (Fig. 1B). Sonicate for approximately 30 minutes at room temperature.
- 4 After sonication, visually inspect skeletons. If the coral skeletons are completely white and devoid of tissue, proceed to the next step (Step 5). However, if tissue is still visible (i.e., the coral surface is light brown or green) then sonicate skeletons in fresh bleach solution for an additional 30 minutes (or longer) until all tissue disappears.
- 5 Drain and dispose of bleach, transfer coral skeletons into individual pre-labeled aluminum weighing pans, and dry overnight at 60 °C in a drying oven (Fig. 1C).

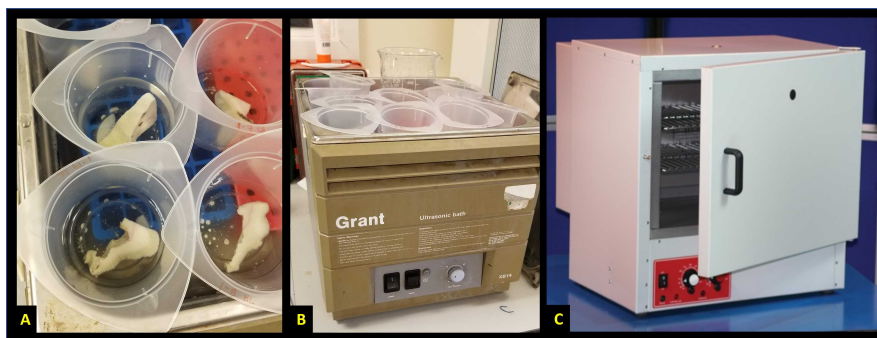


Fig. 1. Photographs of A) *Porites lobata* coral skeletons soaking in 3% bleach solution in individual 500 ml plastic beakers, B) beakers arranged inside a sonicator bath, and C) a laboratory drying oven.

Imaging of Coral Skeletons

- 6 The photomicrographs used in this protocol were produced using a Leica DMS1000 Microscope. For detailed instructions on how to use this instrument, please refer to the User Manual.

[Leica_DMS1000_UserManual_EN.pdf](#)

- 7 Multiple photomicrographs should be taken per individual coral skeleton at a variety of angles and magnifications (Fig. 2). First, take lower magnification images in order to visualize the gross morphology of the skeleton (Fig. 2 A). Next, take medium-magnification images from above (i.e., dorsal view) in order to visualize individual corallites (Fig. 2 B). Finally, take multiple high-magnification images to visualize sub-corallite structures (Fig. 2 C, D). Specific magnification levels vary and will depend on the size of coral fragments and the size of corallites. The angle of the skeleton relative to the microscope should be adjusted between high-magnification photomicrographs as later image-analysis measurements can only be conducted on corallites that were photographed perfectly perpendicular to the field-of-view rather than at an angle.



Fig. 2. Photographs of A) low magnification cross-sectional view of *Porites lobata* coral skeleton slice, B) medium magnification dorsal view with ~60 corallites in field-of-view, and C) high magnification dorsal view with ~8 complete corallites in field-of-view.

Photomicrograph Image Analysis

- 8 Download ImageJ. ImageJ can be downloaded for free from the NIH website: <https://imagej.nih.gov/ij/download.html>.
- 9 Open one high-magnification photomicrograph in ImageJ (Fig. 3).

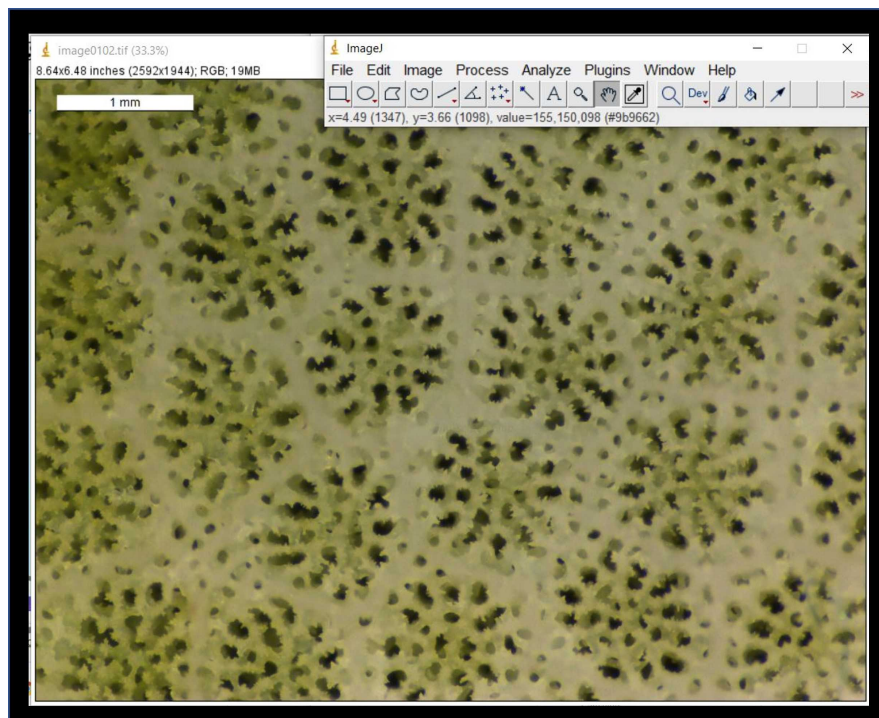


Fig. 3. A high-magnification photomicrograph of *Poites lobata* showing approximately 10 complete corallites. The image is opened in the software ImageJ.

- 10 Set the scale. Using the hand tool and the zoom function (zoom shortcuts: Ctrl and + on PC keyboard, or left-click and scroll on PC mouse), zoom in to the upper left corner of the photomicrograph where the scale bar is located (Fig. 4A). Using the "Straight" tool (fifth button from the left), carefully select the entire 1 mm scale bar (Fig. 4A). Once the scalebar is selected, go to "Analyze" followed by "Set Scale" (Fig. 4A). In the Set Scale window, type "1" into the "Known Distance" field, and type "mm" into the "Unit of length" field (Fig. 4B).

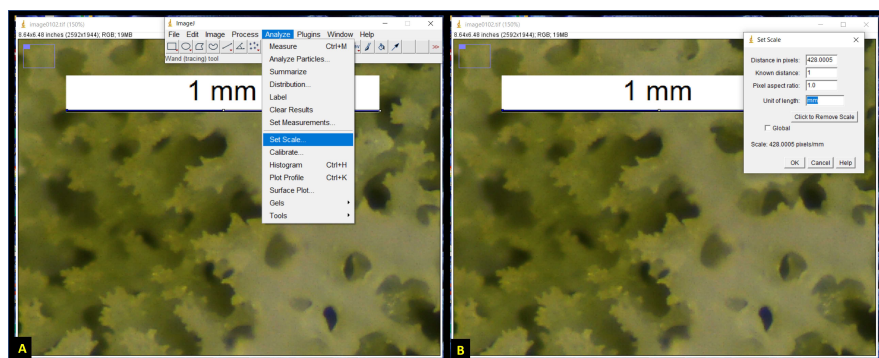


Fig. 4. A) How to select the scale bar and set the scale in ImageJ, and B) the Set Scale window.

- 11 After setting the scale, zoom out again (Fig. 5A), select an appropriately angled corallite, and then zoom in such that a single corallite fills the entire ImageJ window (Fig. 5B).

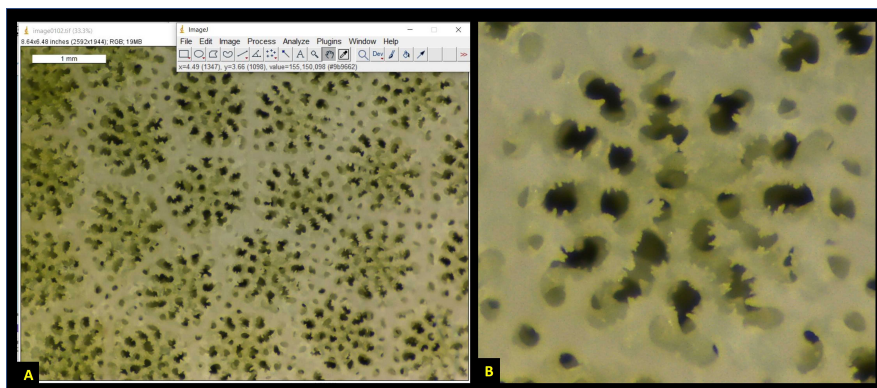


Fig. 5. A) high magnification photomicrograph opened in the software ImageJ, and B) zoomed-in section of photomicrograph such that a single corallite fills the computer screen.

- 12 Make sure to only select corallites which are perpendicular to the field-of-view (Fig. 6). If the cupped-base of the corallite is at an angle relative to the field-of-view, then measurements made from images will not be accurate. You will notice that in Fig. 6A, only the central corallite is appropriately angled, whereas those around it are all tilted.

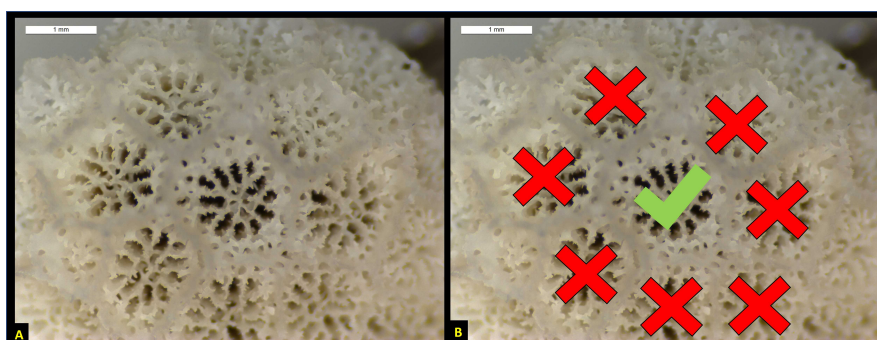


Fig. 6. A) example of a different high magnification photomicrograph of *P. lobata* corallites, and B) the same photomicrograph showing that only the central most corallite (green check mark) is at an appropriate angle for image analysis.

- 13 After selecting an appropriately-angled corallite, next, identify the dorsal directive septa (ubiquitous among *Porites* corals) (Fig. 7). Rotating clockwise from the dorsal directive are lateral pair 1, lateral pair 2, the ventral triplet, lateral pair 3, and finally, lateral pair 4.

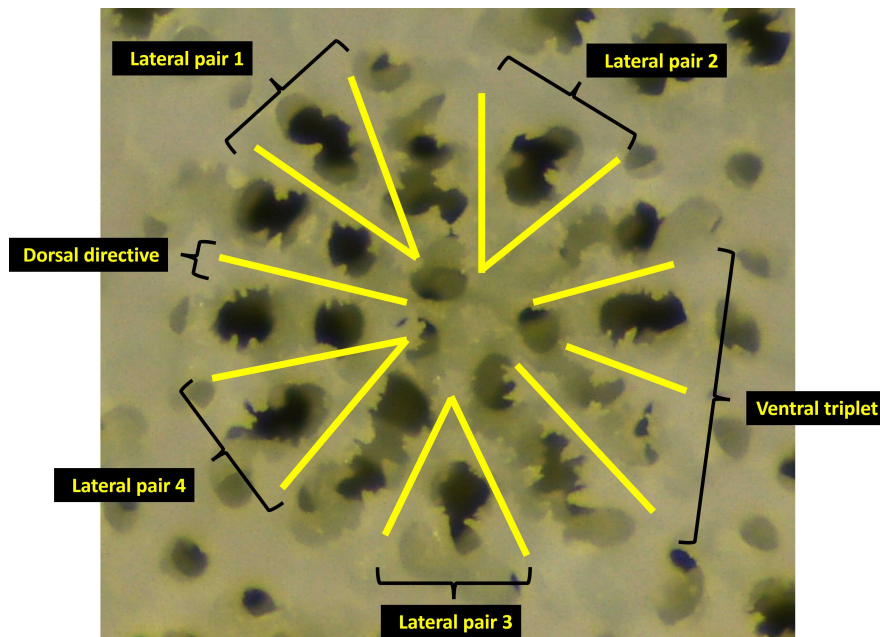


Fig. 7. Labelled photo of a *Porites* corallite highlighting the ubiquitous pattern of septal growth: 1 dorsal directive, 4 lateral pairs and 1 ventral triplet, always in this arrangement.

- 14 Select the "Multi-point" tool (seventh button from the left) (Fig. 8A). Place two points on top of the image. The first point should be placed at the outer end of the dorsal directive septa, and the second point on the inner end of the dorsal directive septa (Fig. 8B). These points will be used to calculate septal-length. Notice in Fig. 8 the small blue numbers next to each point (you may need to enlarge the image in order to see these numbers).

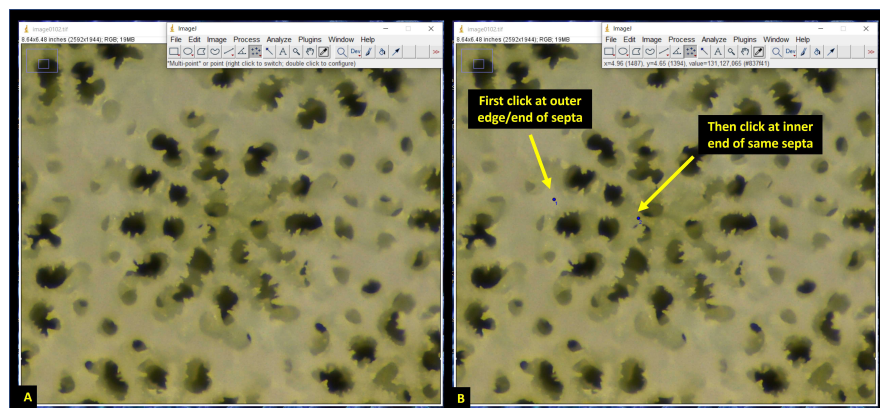


Fig. 8. A) Select the "multi-point" tool from the ImageJ toolbar, and B) place the first point (#1) at the outer end of the dorsal directive septa and place the second point (#2) at the inner end of the same septa.

Take care when placing these septa end-point and try to be consistent within corallites, among corallites, and among different skeleton samples. One method we used to improve consistency was to use the negative space (black holes) as a guide (Fig. 9).

Notice in Fig. 9 how the blue point #1 is placed between the yellow arrows which align with the skeleton holes between septa. If the blue point had instead been placed where the red cross symbol is, it would have been too far out towards the theca wall. Conversely, if the blue point had instead been placed where the orange cross symbol is, it would have been too far inside the calyx area.

Likewise, for the placement of point #2, we used the ring of the fossa (purple circle in Fig. 9) as a guide and tried to

place point 2 on, or as close to this ring as possible.

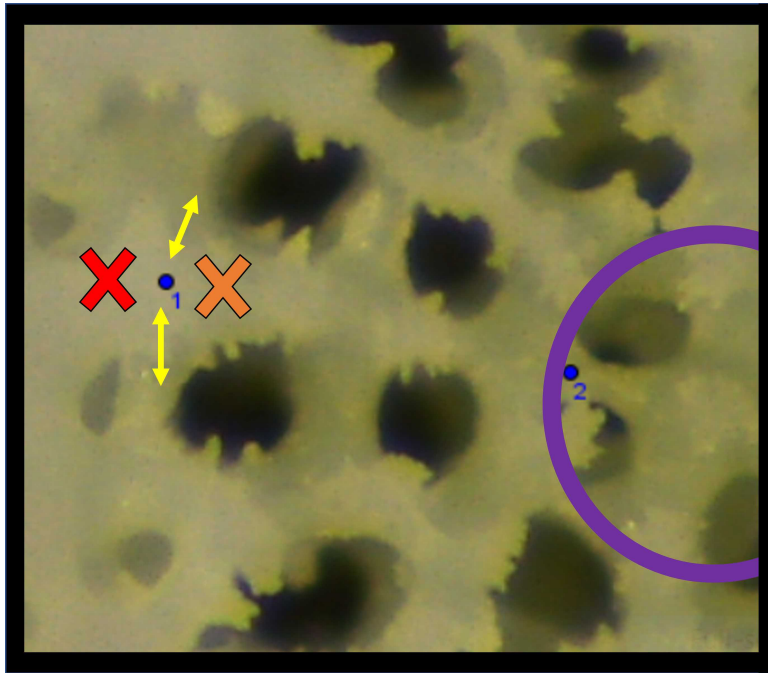


Fig. 9. Diagram showing where the septal endpoints were placed relative to other sub-corallite skeletal structures.

- 15 Repeat this process for all 12 septa (proceeding clockwise from the dorsal directive) until points #1 through #24 have been placed onto the photomicrograph (Fig. 10).

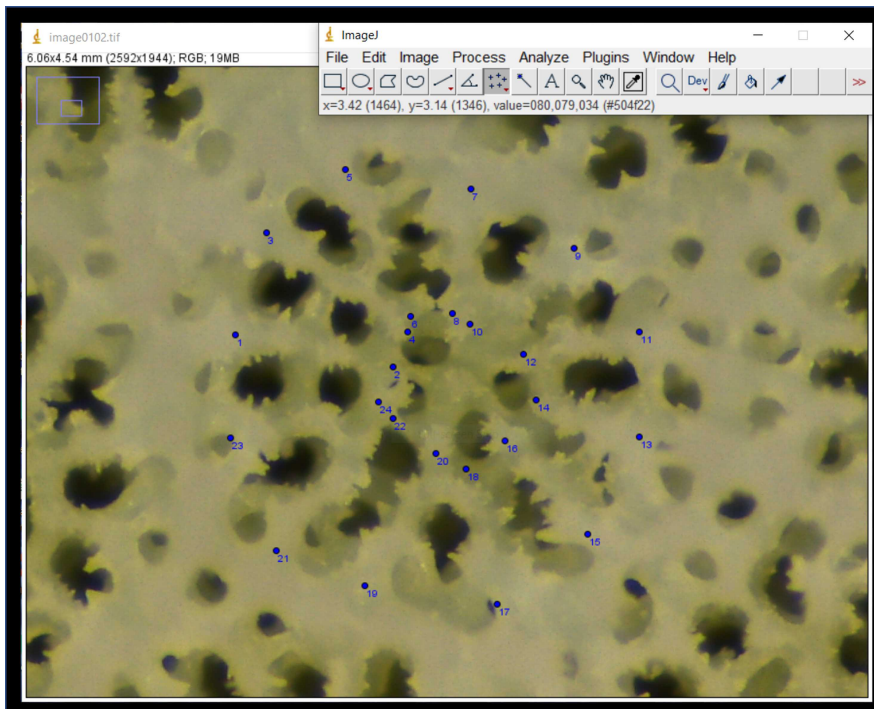


Fig. 10. Placement of 24 septal end points (blue numbered dots) on photomicrograph.

- 16 Next, place an additional 24 points on the photomicrograph on either side of each septa, located at the midpoint (i.e., halfway between the endpoints) (Fig. 11A). These points will be used to calculate septal-width. Proceed clockwise from the dorsal directive septa until points #25 through #48 have been placed (Fig. 11B).

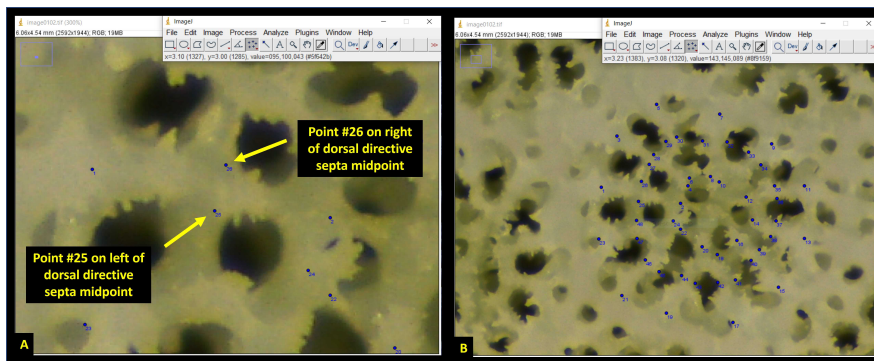


Fig. 11. A) Placement of septa midpoints to the left and right of dorsal directive septa. B) Placement of points #25 through #48 on photomicrograph.

- 17 Next, place an additional 12 points on the photomicrograph opposite from the septa outer-edge points (Fig. 12A). These points should be placed on the inner edge of the theca walls of neighboring corallites (Fig. 12A). The distance between septa outer-edge points and these points will be used to calculate the thickness of the theca wall. Proceed clockwise from the dorsal directive septa until points #49 through #60 have been placed (Fig. 12B).

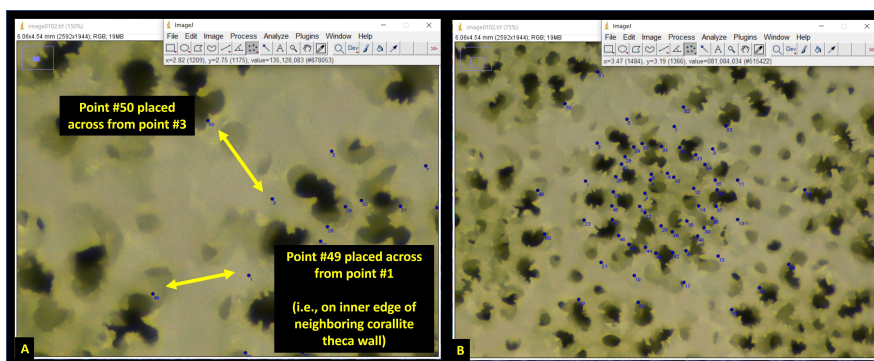


Fig. 12. A) Placement of points opposite the outer-edge endpoint of septa. B) Placement of points #49 through #60 on photomicrograph.

- 18 Next place point #61 at the center of the primary corallite (Fig. 13 green text). Following this, place one point in the center of all the neighboring corallites (only those which share a thecal wall with the primary corallite (Fig. 13 yellow text). The number of neighboring corallites will vary depending upon the individual (anywhere from 4–8 in my experience).

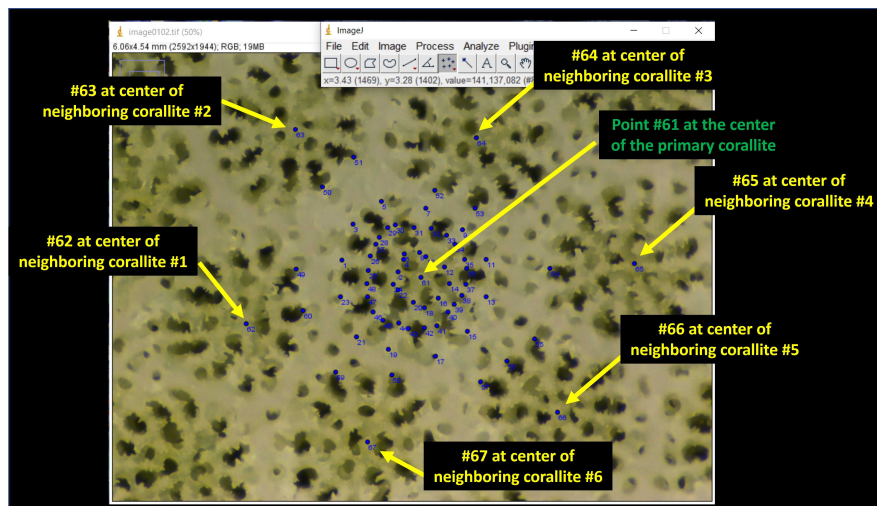


Fig. 13. Placement of point #61 in the center of the primary corallite (green text), and points #62 and upwards in the center of all neighboring corallites (yellow text).

- 19 Next, navigate to the toolbar, press "Analyze" followed by "Measure" [keyboard shortcut is Ctrl + M] (Fig. 14A). This will bring up a Results window of the X and Y coordinates for all 67 points on the photomicrograph (Fig. 14B). Copy and paste these values into an excel spreadsheet.

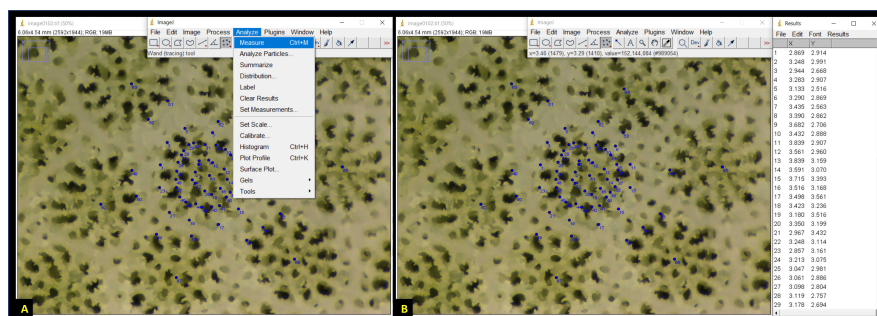


Fig. 14. A) On the ImageJ toolbar, press "Analyze" followed by "Measure". B) Copy & Paste the X and Y coordinates for all 67 points from the Results window which appears.

Calculation of Corallite Microstructure Size

- 20 There are a variety of different measurements that can be made using this data set of X and Y coordinates. Table 1 summarizes which points can be used to calculate the length and width of various corallite microstructures. This is not an exhaustive list, however, and other measurements are possible with the placement of additional points (e.g., pali area, columella diameter, etc.) or from observations (e.g., number of radii, the shape of the ventral triplet, etc.). Please see Forsman et al. (2015) or Tisthammer et al. (2018) for such examples.

A	B	C	D	E	F
Description	Points	Description	Points	Description	Points
Septa length 1	1:2	Septa width 1	25:26	Septa spacing 1	1:3
Septa length 2	3:4	Septa width 2	27:28	Septa spacing 2	3:5
Septa length 3	5:6	Septa width 3	29:30	Septa spacing 3	5:7
Septa length 4	7:8	Septa width 4	31:32	Septa spacing 4	7:9
Septa length 5	9:10	Septa width 5	33:34	Septa spacing 5	9:11
Septa length 6	11:12	Septa width 6	35:36	Septa spacing 6	11:13
Septa length 7	13:14	Septa width 7	37:38	Septa spacing 7	13:15
Septa length 8	15:16	Septa width 8	39:40	Septa spacing 8	15:17
Septa length 9	17:18	Septa width 9	41:42	Septa spacing 9	17:19
Septa length 10	19:20	Septa width 10	43:44	Septa spacing 10	19:21
Septa length 11	21:22	Septa width 11	45:46	Septa spacing 11	21:23
Septa length 12	23:24	Septa width 12	47:48	Septa spacing 12	23:1
Theca thickness 1	1:49	Corallite width 1	1:13	Corallite spacing 1	61:62
Theca thickness 2	3:50	Corallite width 2	3:15	Corallite spacing 2	61:63
Theca thickness 3	5:51	Corallite width 3	5:17	Corallite spacing 3	61:64
Theca thickness 4	7:52	Corallite width 4	7:19	Corallite spacing 4	61:65
Theca thickness 5	9:53	Corallite width 5	9:21	Corallite spacing 5	61:66
Theca thickness 6	11:54	Corallite width 6	11:23	Corallite spacing 6	61:67
Theca thickness 7	13:55	Fossa width 1	2:14	Corallite spacing 7	61:68
Theca thickness 8	15:56	Fossa width 2	4:16	Corallite spacing 8	61:69
Theca thickness 9	17:57	Fossa width 3	6:18		
Theca thickness 10	19:58	Fossa width 4	8:20	Corallite Area	1:3:5:7:9:11:13:15:17:19:21:23
Theca thickness 11	21:59	Fossa width 5	10:22	Fossa Area	2:4:6:8:10:12:14:16:18:20:22:24
Theca thickness 12	23:60	Fossa width 6	12:24		

Table 1. Summary of which points are used for the calculation of various corallite microstructures.

- 21 To calculate the distance between two points, use the following formula:

$$\sqrt{(x_2 - x_1)^2 + (y_2 - y_1)^2}$$

Image source: Forsman et al (2015) DOI: [10.7717/peerj.751](https://doi.org/10.7717/peerj.751)

- 22 **EXAMPLE:** To calculate the length of the dorsal septa (aka "Septa Length 1") we will use the X and Y coordinates of Point #1 and Point #2 (recall Table 1) which were measured and are shown in Figure 14.

- Point 1 (X = 2.869, Y = 2.914)
- Point 2 (X = 3.248, Y = 2.991)

Next, using the equation shown in Step 21:

- Septa Length 1 = $\sqrt{(3.248 - 2.869)^2 + (2.991 - 2.914)^2}$

Therefore, Septa Length 1 = 0.387 cm

- 23 In addition to distances between points, the [area of polygons](#) constructed from multiple points can also be calculated. For instance, Corallite Area or Fossa Area (cm²) can be calculated using the following equation:

The area is then given by the formula

$$\left| \frac{(x_1 y_2 - y_1 x_2) + (x_2 y_3 - y_2 x_3) \dots + (x_n y_1 - y_n x_1)}{2} \right|$$

Image source: <https://www.mathopenref.com/coordpolygonarea.html>