



Biogeochemical and plant trait mechanisms drive enhanced methane emissions in response to whole-ecosystem warming

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Abstract. Climate warming perturbs ecosystem carbon (C) cycling, causing both positive and negative feedbacks on greenhouse gas emissions. In 2016, we began a tidal marsh field experiment in two vegetation communities to investigate the mechanisms by which whole-ecosystem warming alters C gain, via plant-driven sequestration in soils, and C loss, primarily via methane (CH_4) emissions. Here, we report the results from the first 4 years. As expected, warming of 5.1°C more than doubled CH_4 emissions in both plant communities. We propose this was caused by a combination of four mechanisms: (i) a decrease in the proportion of CH_4 consumed by CH_4 oxidation, (ii) more C substrates available for methanogenesis, (iii) reduced competition between methanogens and sulfate-reducing bacteria, and (iv) indirect effects of plant traits. Plots dominated by *Spartina patens* consistently emitted more CH_4 than plots dominated by *Schoenoplectus americanus*, indicating key differences in the roles these common wetland plants play in affecting anaerobic soil biogeochemistry and suggesting that plant composition can modulate coastal wetland responses to climate change.

1 Introduction

Methane (CH_4) is a potent greenhouse gas that contributes to 15 %–19 % of total greenhouse gas radiative forcing (IPCC, 2013) and has a sustained-flux global warming potential that is 45 times that of CO_2 on a 100-year timescale (Neubauer and Megonigal, 2015). Wetlands are the largest natural source of CH_4 to the atmosphere and were recently identified as the largest source of uncertainty in the global CH_4 budget (Saunio et al., 2016). Recent estimates calculate that CH_4

emissions from vegetated coastal wetlands offset 3.6 of the 12.2 million metric tons (MMT) of CO_2 equivalents accumulated by these ecosystems each year (EPA, 2017). Despite this, there is still a substantial knowledge gap regarding how global change factors, such as climate warming, will alter coastal wetland CH_4 emissions (McLeod et al., 2011) even though these feedbacks have the potential to shift coastal wetlands from being a net sink of C to a net source (Al-Haj and Fulweiler, 2020; Bridgman et al., 2006).

The net flux of CH_4 to the atmosphere from any ecosystem represents the balance between the amount of CH_4 produced (methanogenesis), the amount of CH_4 oxidized (methanotrophy), and the rate of CH_4 transport from the soil. In coastal wetlands, methanogenesis occurs through three pathways: (i) hydrogenotrophic methanogenesis (i.e., CO_2 reduction) in which H_2 is the electron donor and CO_2 is the electron acceptor; (ii) acetoclastic methanogenesis, in which acetate (CH_3COOH) is split into CH_4 and CO_2 ; and (iii) methylotrophic methanogenesis in which methylated compounds are converted to CH_4 and CO_2 (Conrad, 2020; Oremland et al., 1982; Schlesinger and Bernhardt, 2020). Rates of methanogenesis are driven by low-redox conditions and substrate availability, while aerobic CH_4 oxidation requires both O_2 and CH_4 as substrates. Roots and rhizomes in wetland ecosystems influence methane-related substrates through at least two mechanisms: (i) deposition of organic compounds that support multiple pathways of heterotrophic microbial respiration, including methanogenesis, and (ii) release of O_2 that simultaneously promotes CH_4 oxidation and regeneration of competing electron acceptors such as Fe(III) and SO_4 (Philippot et al., 2009; Stanley and Ward, 2010). Root exudates, which typically include low-molecular-weight compounds, may either be more readily used by microbes than

existing soil C (Kayranli et al., 2010; Megonigal et al., 1999) or prime microbial use of soil C (Basiliko et al., 2012; Philip-pot et al., 2009; Robroek et al., 2016; Waldo et al., 2019). Root exudates can also decrease CH₄ oxidation by stimulating use of O₂ by other aerobic microbes (Lenzewski et al., 2018; Mueller et al., 2016). Consequently, wetland CH₄ emissions are strongly linked to a wide variety of plant traits that govern the supply of reductive (organic carbon) and oxidative (O₂) substrates to soils (Moor et al., 2017; Mueller et al., 2020).

Although it is understood that wetland plants are a primary control on CH₄ emissions and that much of their influence is mediated through conditions in the rhizosphere (Waldo et al., 2019), there are surprisingly few data, especially from coastal wetlands, that couple plant responses to the dynamics of electron donors (organic C), electron acceptors (O₂, SO₄), and the rates of competing (sulfate reduction vs. methanogenesis) or opposing (CH₄ production vs. CH₄ oxidation) microbial processes. The general lack of process data on wetland CH₄ cycling makes it difficult to forecast ecosystem responses to climate change. For example, the well-documented observation that warming increases wetland methane emissions can be either amplified or dampened depending on changes in plant activity (e.g., primary production) or plant traits (e.g., community composition) (Mueller et al., 2020). Vegetation composition has been shown to be a stronger control on CH₄ emissions than ~1 °C of warming in northern peatlands (Ward et al., 2013), and Chen et al. (2017) proposed that warming effects on plant functional types can drive C flux responses that cannot otherwise be explained by abiotic conditions. In freshwater marshes, plant species and growth trends have also been linked to seasonal shifts in pools of dissolved CH₄ and dissolved inorganic carbon (DIC; Ding et al., 2005; Stanley and Ward, 2010) and methanogenesis dynamics (Sorrell et al., 1997).

Tidal wetlands are particularly good model systems for determining the mechanisms by which warming alters CH₄ emissions. Not only will the CH₄ cycle respond to the direct effects of warming, but the temperature effects on the outcome of competition for electron acceptors are relatively easily observed because of the abundance of SO₄. Thermodynamic theory in which terminal electron acceptors (TEAs) are used in order of decreasing thermodynamic yield is commonly interpreted to mean that a system will support only one form of anaerobic respiration at a time, with acetoclastic and hydrogenotrophic methanogenesis occurring only when pools of more energetically favorable TEAs have been depleted (Conrad, 2020; Schlesinger and Bernhardt, 2020). However, in real systems with spatial and temporal variability in the supply of electron donor substrates and TEAs, all forms of anaerobic metabolism occur simultaneously (Megonigal et al., 2004; Bridgman et al., 2013). Much of this spatial and temporal variation arises from the distribution and activity of roots and rhizomes as mediated by the rhizosphere (Neubauer et al., 2008). Global change factors such as warm-

ing will further affect the spatial distribution of key metabolic substrates. In addition, the relatively limited species diversity in saline tidal wetlands allows species-level effects on CH₄ cycling to be delineated more easily than in diverse freshwater wetlands.

Methane flux measurements are a metric of broader shifts in redox potential and biogeochemical cycling, as they are sensitive to virtually all processes that regulate availability of electron donors and electron acceptors. Emissions are commonly predicted to increase with future climate warming, including from coastal wetlands (Al-Haj and Fulweiler, 2020), but there is minimal prior understanding of the underlying mechanisms, which was the focus of this study. Our objectives were to explore the mechanisms that drive enhanced CH₄ emissions under warming. To accomplish this, we measured monthly CH₄ emissions from 2016 through 2019 and coupled these flux measurements with analysis of porewater biogeochemistry and vegetation biomass and composition.

2 Materials and methods

2.1 Site description and experimental design

The Salt Marsh Accretion Response to Temperature eXperiment (SMARTX) was established in the Smithsonian's Global Change Research Wetland (GCRew) in 2016. GCRew is part of Kirkpatrick Marsh, a microtidal, brackish high marsh on the western shore of the Chesapeake Bay, USA (38°53' N, 76°33' W). Soils are organic (> 80 % organic matter) to a depth of 5 m, which is typical of high marshes in the Chesapeake Bay and elsewhere. The very low mineral content (< 20 %) affects methane dynamics because negligible competition between methanogens and iron-reducing bacteria for electron donors is expected in the absence of a significant pool of poorly crystalline iron oxides (Roden and Wetzel, 1996), as has been documented previously at this site (Weiss et al., 2004). Soil bulk density in the upper 60 cm averages 0.124 g cm⁻³ and ranges from 0.079 to 0.180 g cm⁻³. The relatively uniform bulk density of the soil profile reflects the uniform soil organic matter content and the fact that bulk density becomes largely independent of organic matter and mineral content once organic matter content exceeds 50 % (Holmquist et al., 2018). The marsh is typically saturated to within 5–15 cm of the soil surface, but inundation frequency varies across the site, from 10 %–20 % of high tides in high-elevation areas to 30 %–60 % of high tides in low-elevation areas.

SMARTX consists of six replicate transects, three located in each of the two dominant annual plant communities (Fig. S1). In the C₃-dominated community (herein the “C₃ community”) the C₃ sedge *Schoenoplectus americanus* (herein *Schoenoplectus*) composes more than 90 % of the aboveground biomass (Table 1). In the C₄-dominated community (herein the “C₄ community”), 75 % of the above-

Table 1. Relative contribution to total aboveground biomass from C₃ sedges (*Schoenoplectus americanus*) and C₄ grasses (*Spartina patens* and *Distichlis spicata*) in each plant community. Values are means and SE (*n* = 12).

Year	C ₃ community		C ₄ community	
	% C ₃	% C ₄	% C ₃	% C ₄
2016	93 (3)	8 (3)	8 (2)	76 (6)
2017	91 (3)	9 (3)	10 (3)	64 (4)
2018	95 (1)	5 (1)	15 (4)	65 (7)
2019	93 (2)	4 (1)	23 (5)	56 (6)

ground biomass was initially composed of two C₄ grasses (*Spartina patens* and *Distichlis spicata*, herein *Spartina* and *Distichlis*, respectively). However, by 2019, *Spartina* and *Distichlis* declined to 56 % of the aboveground biomass (Table 1).

Each transect is an active warming gradient consisting of unheated ambient plots and plots that are heated to 1.7, 3.4, and 5.1 °C above ambient. All plots are 2 m × 2 m with a 20 cm-wide buffer around the perimeter. Aboveground plant-surface temperature is elevated via infrared heaters, and soil temperature is elevated via vertical resistance cables (Rich et al., 2015). Soils are heated to a depth of 1.5 m, which is the depth most vulnerable to climate or human disturbance (Pendleton et al., 2012). Aboveground and belowground temperature variation are assessed via thermocouples embedded in acrylic plates at plant canopy level and inserted into the soil, respectively, and the temperature gradient is maintained by integrated microprocessor-based feedback control (Rich et al., 2015). Noyce et al. (2019) provide additional details of the heating system. Warming began on 1 June 2016 and has continued year-round.

2.2 Methane flux measurements

Methane emissions were measured monthly year-round from May 2016 to December 2019 using a static chamber system. One permanent 160 cm² aluminum base was inserted 10 cm into the soil in each plot in April 2016. On each measurement date, clear chambers (40 cm × 40 cm × 40 cm) were gently placed on top of each base and secured with compression clips. Chambers consisted of an aluminum frame with transparent sides made of polychlorotrifluoroethylene film (Honeywell International) and closed-cell foam on the base. Depending on the height of the vegetation at the time of measurement, chambers were stacked up to four high (total height of 40–160 cm) (Fig. S2). The advantage of this stacking method is that it uses the minimal chamber volume necessary, while also allowing for plant growth. After placement, the chambers were left open for at least 10 min, to minimize disturbance effects and allow air inside the chamber to return to ambient conditions. During data collection, chambers

were covered with a transparent polycarbonate top equipped with sampling tubes, a fan to circulate air inside the chambers, a PAR sensor, and thermocouples. The sealed chamber was covered with a foil shroud to block out all light and to minimize changes in temperature and relative humidity during the measurement period. An Ultraportable Greenhouse Gas Analyzer (Los Gatos Research, CA) was used to measure headspace CH₄ concentrations for 5 min. Plots were accessed from permanent boardwalks elevated 15 cm above the soil surface to avoid compressing the surrounding peat and altering diffusive CH₄ emissions. Fluxes were calculated as the slope of the linear regression of CH₄ concentration over time. The 40 fluxes where *p* > 0.05 were assigned a value of one-half the limit of detection of the system (Wassmann et al., 2018; Table S1). This was 1 % of fluxes during the growing season and 4.5 % of fluxes over the remaining months. For 2017–2019, monthly measurements were scaled to annual estimates by regressing CH₄ emissions against daily mean soil temperature and day of year (as a proxy for phenological status). Annual estimates were not calculated for 2016 because flux measurements did not start until May.

2.3 Porewater sampling and analysis

Porewater samples were collected in May, July, and September of each year using stainless-steel “sippers” permanently installed in each plot. Each sipper consisted of a length of stainless-steel tubing, crimped and sealed at the end, with several slits (approximate width 0.8 mm) cut in the bottom 2 cm. The aboveground portion of each sipper was connected to Tygon Masterflex[®] tubing capped with a two-way stopcock. In May 2016, duplicate clusters of sippers were installed in each of the 30 plots at 20, 40, 80, and 120 cm below the soil surface. An additional set of 10 cm deep sippers was installed in 2017. In this study we defined samples from 10–20 cm as “rooting zone” samples and samples from 40–120 cm as “deep peat” samples. On sampling dates, porewater sitting in the sippers was drawn up and discarded, after which 60 mL of porewater from each depth (30 mL from each sipper) was withdrawn and stored in syringes equipped with three-way stopcocks. A 10 mL aliquot of each sample was filtered through a pre-leached 0.45 µm syringe-mounted filter, preserved with 5 % zinc acetate and sodium hydroxide, and frozen for future SO₄ and Cl analysis. Dissolved CH₄ was extracted from 15 mL of porewater in the syringe by drawing 15 mL of ambient air and shaking vigorously for 2 min to allow the dissolved CH₄ to equilibrate with the headspace. Headspace subsamples were then immediately analyzed on a Shimadzu GC-14A gas chromatograph equipped with a flame ionization detector. A total of 3 mL of porewater was used to measure pH using a Fisher Scientific accumet electrode (13-620-290). The remaining porewater was used to measure H₂S and NH₄; those data are not reported here.

SO₄ and Cl were measured on a Dionex ICS-2000 ion chromatography system (2016–2018) or a Dionex Inte-

gation (2019). On the Dionex ICS-2000 samples were separated using an A11 column with 30 mM of KOH as eluent; on the Dionex Integrion samples were separated using an A11 4 μ m fast column with 35 mM KOH. Sulfate depletion ($\text{Sulfate}_{\text{Dep}}$) was calculated based on measured porewater concentrations of SO_4 ($\text{SO}_{4\text{pw}}$) and Cl (Cl_{pw}) and the constant molar ratio of Cl to SO_4 in surface seawater ($R_{\text{sw}} = 19.33$; Bianchi, 2006) using the following equation: $\text{Sulfate}_{\text{Dep}} = \text{Cl}_{\text{pw}}/R_{\text{sw}} - \text{SO}_{4\text{pw}}$. If driven only by seawater inputs, the ratio of Cl to SO_4 would remain constant, but under anaerobic conditions SO_4 can be reduced by sulfate-reducing bacteria, altering this ratio. As a result, SO_4 depletion can be used as a proxy for SO_4 reduction rates.

2.4 Plant biomass measurements

Measurements of *Schoenoplectus*, *Spartina*, and *Distichlis* biomass were conducted during peak biomass of each year (29 July–2 August) as described by Noyce et al. (2019). *Schoenoplectus* biomass was estimated using non-destructive allometric techniques (Lu et al., 2016) in 900 cm^2 quadrats, and *Spartina* and *Distichlis* biomass were estimated through destructive harvest of 25 cm^2 subplots.

2.5 Data analysis

Statistics were conducted in R (version 3.6.3). Methane flux (Fig. S3) and porewater data were log transformed to become normally distributed prior to statistical analyses. The “growing season” was defined as May through September based on *Schoenoplectus* growth trends (Fig. S4). Pearson’s correlations were used to test the relationships between CH_4 flux and soil temperature as well as CH_4 flux and plant biomass. Responses of CH_4 emissions to vegetation type and warming treatment were analyzed using linear mixed models with vegetation community and warming treatment as categorical variables and plot and year as random effects. *P* values were calculated using Satterthwaite’s method, and Tukey’s post hoc tests were used to compare individual means. Porewater data were averaged per year and then analyzed using one-way ANOVAs to determine the effects of warming treatment or plant community, applying Tukey’s HSD test for post hoc analyses.

3 Results

3.1 Environmental conditions, site characteristics, and experiment performance

The growing season of 2016 was the hottest of the 4 years, with growing season temperatures averaging $> 1^\circ\text{C}$ above the other 3 years (Table 2). While 2017 through 2019 had similar summer temperatures, they had very different precipitation regimes: 2018 was much wetter on average and 2019 was slightly drier (Table 2). During all years, temperatures in

Table 2. Growing season (May–September) temperature and precipitation. Temperature data are means (SE) of daily averages from ambient plots, and precipitation is the total from May through September.

Year	Mean aboveground temperature ($^\circ\text{C}$)	Mean soil temperature ($^\circ\text{C}$)	Total precipitation (cm)
2016	24.7 (0.3)	22.4 (0.2)	51.0
2017	22.0 (0.3)	20.7 (0.2)	51.1
2018	23.7 (0.3)	20.8 (0.2)	86.4
2019	23.6 (0.3)	20.9 (0.2)	43.7

the experimental plots were successfully shifted by the target differentials of $+1.7$, $+3.4$, and $+5.1^\circ\text{C}$ above the ambient plots (Fig. 1 (top); Noyce et al., 2019). Porewater pH ranged from 6.4 to 6.8 across the measurement period, with no effect of temperature treatment ($p > 0.1$; data not shown). There was no difference in soil bulk density between the ambient and $+5.1^\circ\text{C}$ plots after 4.5 years of warming ($p = 0.54$; data not shown).

3.2 Methane fluxes

Methane emissions increased with soil temperature ($R^2 = 0.41$, $p < 0.001$) (Fig. 1). Emissions from all treatments had strong seasonal trends; fluxes were highest in the C_3 community in June through August and peak fluxes in the C_4 community were shifted about a month later to July through September (Fig. S3). Whole-ecosystem warming increased CH_4 emissions throughout the growing season ($F_{3,400} = 5.1$, $p = 0.002$; Fig. 2). Across all 4 years, 5.1°C of warming more than doubled growing season emissions, from 624 to 1413 $\mu\text{mol CH}_4 \text{ m}^{-2} \text{ d}$ ($p_{\text{adj}} = 0.02$; Fig. 2).

Mean CH_4 emissions were higher from the C_4 community than from the C_3 community both during the growing season ($F_{1,22} = 13.6$, $p = 0.001$; Fig. 3a) and on an annual basis ($F_{1,22} = 8.5$, $p = 0.008$; Fig. 3b). Mean annual CH_4 emissions ranged from 58 mmol CH_4 (ambient) to 343 $\text{mmol CH}_4 \text{ m}^{-2} \text{ yr}^{-1}$ ($+5.1^\circ\text{C}$) in the C_3 community and from 55 mmol CH_4 (ambient) to 879 $\text{mmol CH}_4 \text{ m}^{-2} \text{ yr}^{-1}$ ($+5.1^\circ\text{C}$) in the C_4 community (Table S2). Under ambient conditions, growing season CH_4 fluxes were almost twice as large from C_4 plots, whereas under low warming (1.7 to 3.4°C) this difference increased to more than 3 times as large (Fig. 3a). From 2017–2019, CH_4 emissions were positively related to *Spartina* and *Distichlis* aboveground biomass across all warming treatments and negatively related to *Schoenoplectus* biomass (Fig. 4a, b). In 2016, however, the direction of those relationships in both plant communities were the exact opposite, with *Spartina* and *Distichlis* biomass negatively related, and *Schoenoplectus* biomass positively related, to CH_4 emissions (Fig. 4a, b).

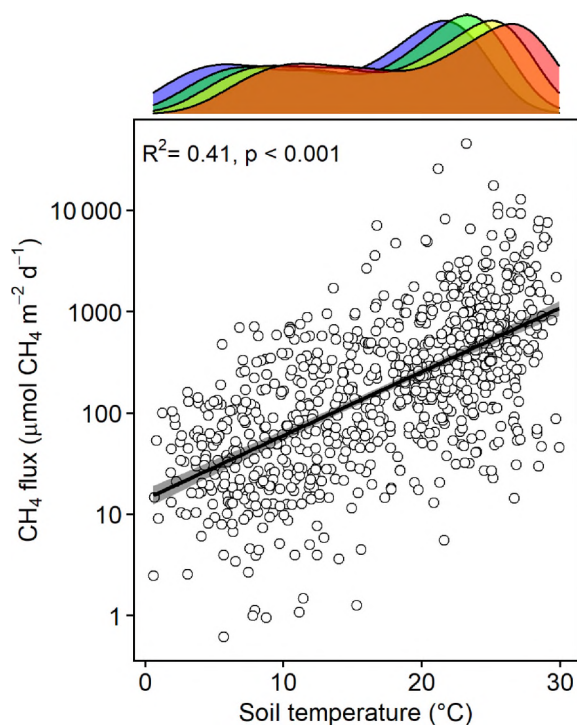


Figure 1. Bottom: CH₄ emissions from each plot versus the soil temperature at the time of measurement. Top: density plot depicting the range of soil temperatures in each treatment, delineated by color: ambient (blue), +1.7 (green), +3.4 (yellow), and +5.1 °C (red).

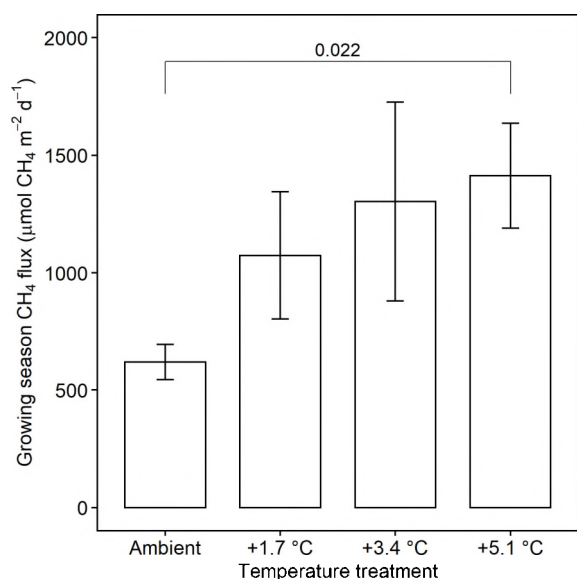


Figure 2. Comparison of CH₄ emissions from each warming treatment during the growing season (May–September). Means include both the C₃ and C₄ community and all years of measurement. Error bars indicate SE. Horizontal bars indicate means that are significantly different and the corresponding *p*_{adj}.

3.3 Porewater chemistry

Under ambient conditions, porewater collected from the C₄ community had more dissolved CH₄ ($F_{1,22} = 18.4$, $p < 0.001$; Fig. 5a, b), less SO₄ ($F_{1,22} = 29.1$, $p < 0.001$; Fig. 6a), and similar salinity ($p = 0.068$) compared to the C₃ community. In the C₃ community, warming increased dissolved CH₄ in both the rooting zone porewater (10–20 cm) ($F_{3,44} = 2.85$, $p = 0.048$; Fig. 5a) and in the deep peat (40–120 cm) ($F_{3,44} = 6.23$, $p = 0.001$; Fig. 5b). Dissolved CH₄ concentrations were relatively similar in the ambient, +1.7, and +3.4 °C treatments but more than doubled with +5.1 °C of warming in both the rooting zone (59 to 125 μmol CH₄ L⁻¹, $p_{\text{adj}} < 0.001$) and the deeper porewater (43 to 1254 μmol CH₄ L⁻¹, $p_{\text{adj}} < 0.001$). In the C₄ community there was minimal effect of warming treatment on porewater in the rooting zone ($F_{3,44} = 0.442$, $p = 0.72$; Fig. 5a), but all levels of warming decreased dissolved CH₄ below 40 cm ($F_{3,44} = 129.3$, $p < 0.001$), with concentrations in the +3.4 and +5.1 plots less than a third of the concentrations in the ambient plots (155 vs. 56 and 40 μmol CH₄ L⁻¹, $p_{\text{adj}} < 0.001$; Fig. 5b).

In the C₃ community, SO₄ concentrations decreased with warming ($F_{3,44} = 3.76$, $p = 0.017$), but warming effects on SO₄ cycling in the C₄ community were more mixed with +3.4 °C increasing SO₄ ($p_{\text{adj}} = 0.048$) but no other treatments having large effects (Fig. 6a). In all plots, the measured concentrations of rooting-zone SO₄ were lower than expected based on salinity (Fig. 6b), indicating that SO₄ reduction occurred. In both plant communities, the +5.1 °C treatments increased this SO₄-depletion effect compared to ambient, though the effect was stronger in the C₃ community ($p < 0.001$) than the C₄ community ($p = 0.04$) (Fig. 6b). Dissolved CH₄ was highest in both plant communities when SO₄ concentrations were < 5 mmol SO₄ L⁻¹ (Fig. S5).

4 Discussion

Soil temperature (both seasonal and experimental) and plant traits were both strong drivers of CH₄ emissions from this site. This follows prior field, mesocosm, and incubation studies across a variety of wetlands, in which temperature has been shown to be a strong predictor of CH₄ emissions (e.g., Al-Haj and Fulweiler, 2020; van Bodegom and Stams, 1999; Christensen et al., 2003; Dise et al., 1993; Fey and Conrad, 2000; Liu et al., 2019; Ward et al., 2013; Yang et al., 2019; Yvon-Durocher et al., 2014) and in which plant functional type has an interacting effect (e.g., Chen et al., 2017; Duval and Radu, 2018; Liu et al., 2019; Mueller et al., 2020; Ward et al., 2013). Methane emissions are a function of the balance between methanogenesis, CH₄ oxidation, and CH₄ transport, so explaining these results requires some combination of stimulation of methanogenesis, reduction of CH₄ oxidation, or increase in CH₄ transport.

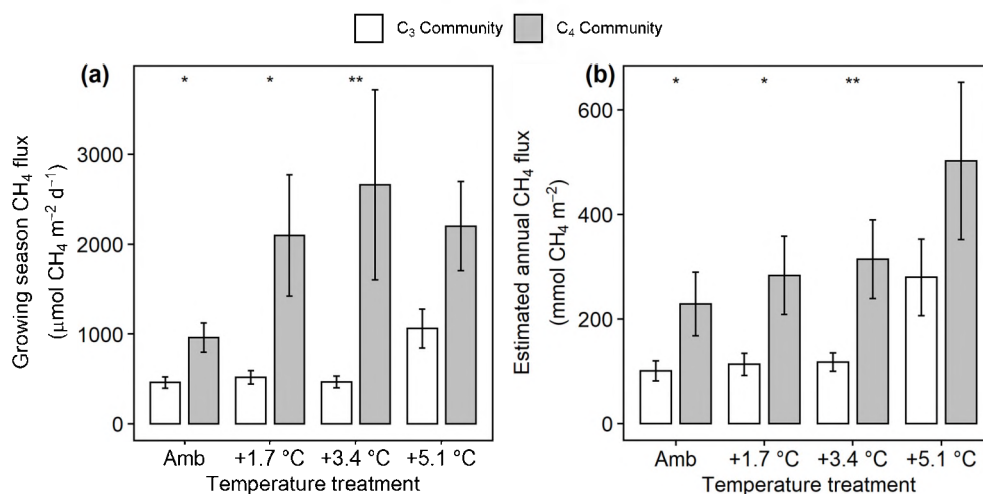


Figure 3. Comparison of CH₄ emissions from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). (a) During the growing season (May–September) and (b) scaled to a year. Means are averaged across all sampling dates for 2017–2019. Error bars indicate SE. Asterisks indicate significant differences between C₃ and C₄ means at a given temperature (* $p_{\text{adj}} < 0.05$, ** $p_{\text{adj}} < 0.01$).

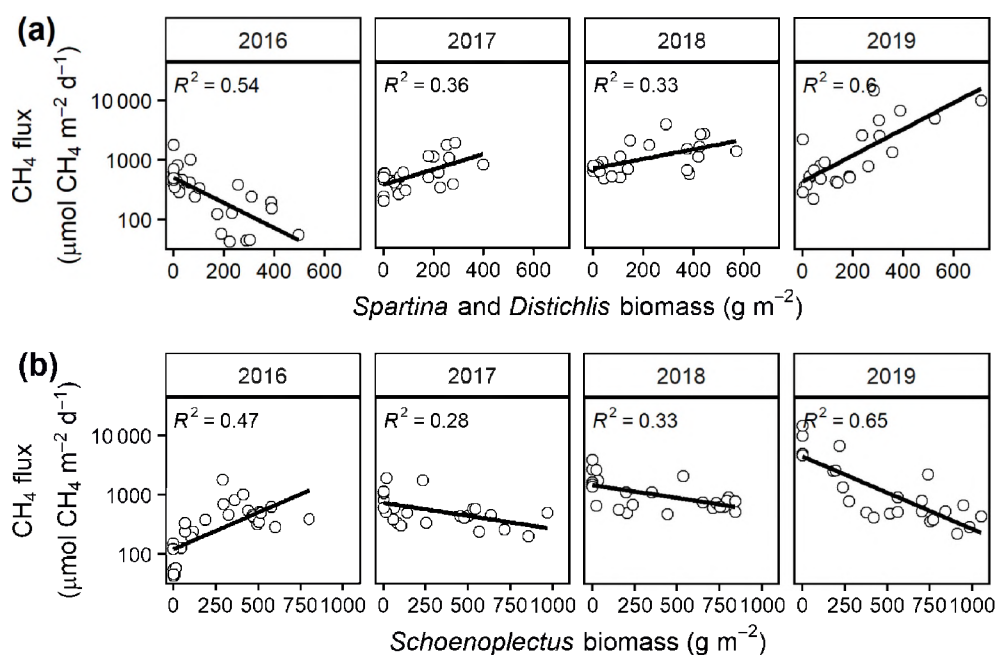


Figure 4. Mean growing season (May–September) CH₄ emissions from each plot versus the biomass of (a) C₃ (*Schoenoplectus*) and (b) C₄ (*Spartina* and *Distichlis*) plants. All regressions are significant at $p = 0.05$.

Prior data from brackish wetlands are limited, but incubation studies of freshwater wetland soils typically show large increases in CH₄ fluxes with warming (van Bodegom and Stams, 1999; Duval and Radu, 2018; Hopple et al., 2020; Inglett et al., 2012; Sihi et al., 2017; Wilson et al., 2016), indicating that warming alters belowground processes. Though there is some evidence that rhizosphere temperature alters CH₄ transport through rice aerenchyma (Hosono and Nouchi, 1997), any transport-driven effects in this ecosys-

tem would be transient unless there were a simultaneous increase in net CH₄ production (i.e., an increase in methanogenesis that was not completely offset by methanotrophy). Instead, we observed a sustained increase in CH₄ emissions, suggesting large shifts in anaerobic metabolism, especially with +5.1 °C of warming. We propose four potential and non-exclusive mechanisms to explain the temperature-driven increase in CH₄ emissions: (1) shifted ratios of CH₄ production to oxidation, (2) increased substrate availability, (3) re-

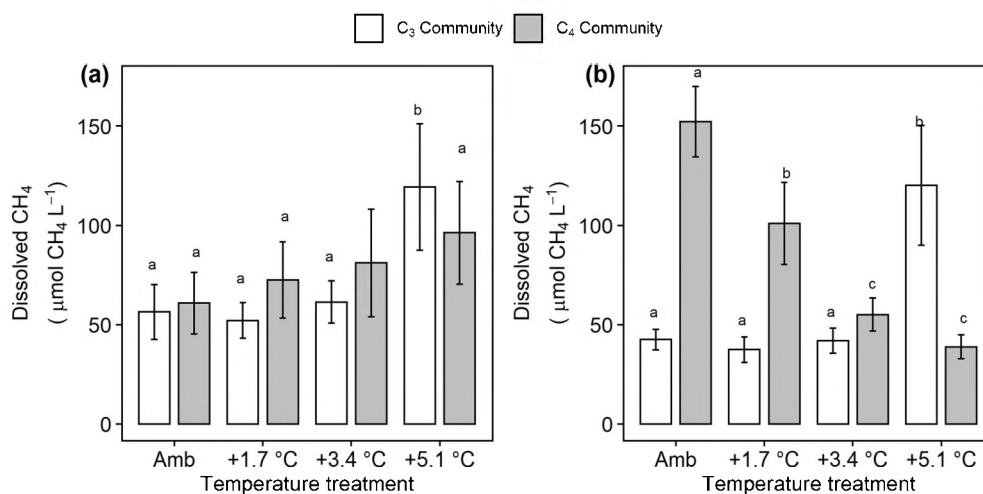


Figure 5. Comparison of dissolved CH₄ from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). **(a)** In the dominant rooting zone (10–20 cm) and **(b)** below the rooting zone (40–120 cm). Means are averaged across all sampling dates for 2016–2019. Error bars indicate SE. Letters indicate temperature treatments that are significantly different from each other ($p_{\text{adj}} < 0.05$) within the same plant community.

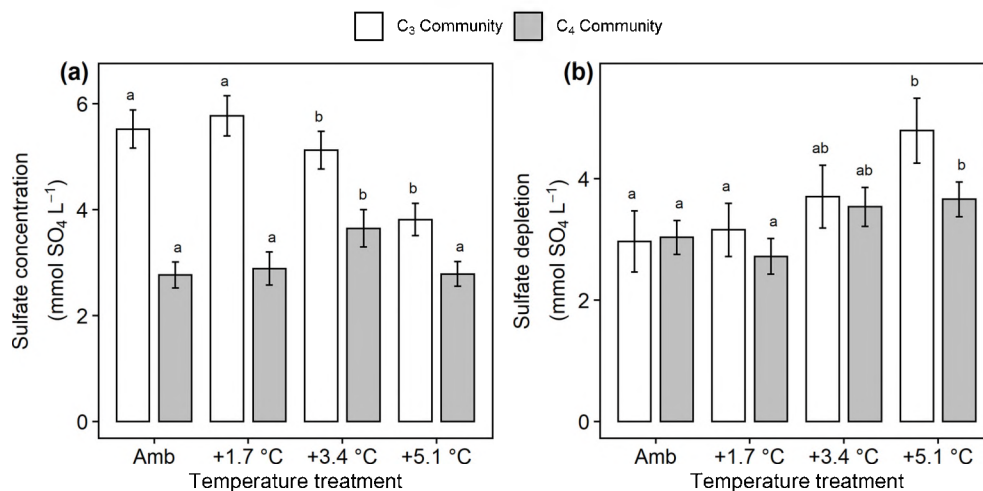


Figure 6. Comparison of sulfate concentrations and estimated sulfate depletion from the C₃ community dominated by *Schoenoplectus* (open bars) and the C₄ community dominated by *Spartina* and *Distichlis* (grey bars). **(a)** Sulfate availability throughout the entire soil profile and **(b)** sulfate depletion in the rooting zone. Means are averaged across all sampling dates for 2016–2019. Error bars indicate SE. Letters indicate temperature treatments that are significantly different from each other ($p_{\text{adj}} < 0.05$) within the same plant community.

duced competition with sulfate reducers for H₂ and organic C, and (4) indirect plant trait effects (Fig. 7).

4.1 Whole-ecosystem warming promotes methanogenesis over CH₄ oxidation

Holding the supply of substrates and transport properties of the system constant, warming is expected to increase rates of CH₄ production relative to CH₄ oxidation due solely to differences in the temperature dependence of each process (Megonigal et al., 2016). In wetland soils, the average Q_{10} of methanogenesis is 4.1 compared to 1.9 for aerobic CH₄

oxidation (Segers, 1998), which means that a system starting with a given initial ratio between the two processes will become increasingly dominated by methanogenesis as soils warm. A corollary to this expected pattern is that the ratio of the two processes should be constant if the Q_{10} responses are similar, an outcome that was supported with in situ measurements of the two processes in a tidal freshwater forested wetland (Megonigal and Schlesinger, 2002). We did not quantify the temperature dependence of CH₄ production and oxidation in the present study, but based on the literature (Segers, 1998) it is likely that methanogenic activity increased more than aerobic methanotrophic activity in direct

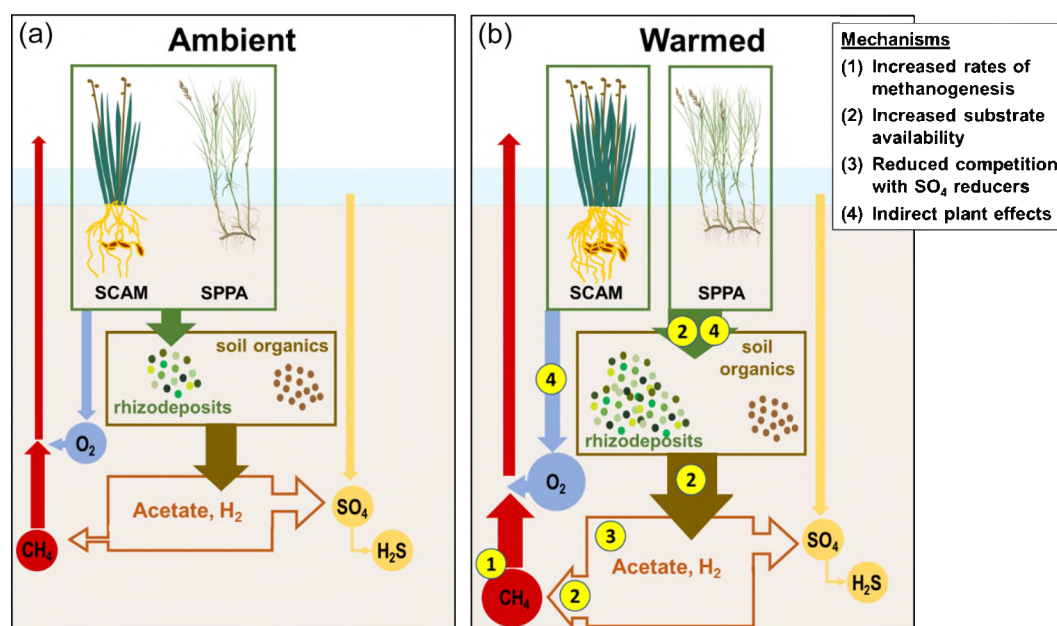


Figure 7. Schematic of mechanisms driving enhanced CH_4 emissions in response to warming. SCAM: *Schoenoplectus americanus*. SPPA: *Spartina patens*. **(a)** Processes under ambient conditions. Plants add organic compounds to the soil, which are transformed into other low-molecular-weight organic compounds. This pool, and processed soil organic matter, support terminal respiration processes dominated by SO_4 reduction over CH_4 production in organic-rich brackish marsh soils. Plants also transport O_2 , which supports oxidation of a fraction of the CH_4 before it can be transported out of the soil. **(b)** Processes under warmed conditions. (1) Rates of CH_4 production increase more than rates of CH_4 oxidation. (2) Substrate availability increases as plants add more rhizodeposits and organic matter is more rapidly fermented to low-molecular-weight organic compounds and H_2 . (3) The pool of electron donors available to methanogens increases as SO_4 reducers become SO_4 limited. (4) The dominant plant species have different effects on these processes, with *S. americanus* driving a net increase in O_2 transport and *S. patens* driving a net increase in rhizodeposits.

response to warming (Fig. 7, mechanism 1). Evidence for this is that rhizosphere pools of porewater CH_4 were highest in the warmest treatment (Fig. 5a); because this occurred despite either no change or an increase in aboveground biomass (Noyce et al., 2019), which would by itself have lowered porewater CH_4 due to venting (plant transport), it indicates that CH_4 production increased relative to the sum of aerobic and anaerobic methane oxidation.

4.2 Whole-ecosystem warming increases substrate availability for methanogens

Methanogenesis can be the terminal step of anaerobic decomposition, but a consortium of microbes is required to break down soil organic matter to electron donor substrates that methanogens can metabolize. The final step in any decomposition pathway involves the flow of electrons from organic matter (electron donors) to a TEA. Under anaerobic conditions, this is accomplished by microbes that tend to specialize in one TEA and compete for organic C as an electron donor (Megonigal et al., 2004). Consequently, the supply of both electron donors and TEAs regulates the multi-step process of anaerobic decomposition and thus ultimately controls CH_4 emissions. For all pathways, methanogenic activity is

typically limited by the supply of electron donors, including low-molecular-weight organic compounds (e.g., acetate; Neubauer and Craft, 2009) and H_2 , a product of organic matter fermentation. We propose that whole-ecosystem warming increases the availability of previously limited C substrates in two aspects (Fig. 7, mechanism 2).

First, warming may directly influence C availability through biochemical kinetics. Even if organic inputs remained constant, warming likely accelerates fermentation of soil organic matter, increasing substrate availability for methanogens. Second, the warmed plots had longer growing seasons than the unheated controls (Noyce et al., 2019). This presumably increased inputs of root exudates and fresh detritus, accelerating all forms of heterotrophic microbial respiration by providing organic material that is decomposed into low-molecular-weight organic C compounds and H_2 (Philipot et al., 2009), stimulating growing season CH_4 emissions from warmed plots. In 2017, we observed that gross primary production was positively correlated with CH_4 emissions and that this effect increased with warming (Fig. S6). Prior studies have also linked CH_4 production or emissions to rates of photosynthesis (Vann and Megonigal, 2003), periods of active growth (Chen et al., 2017; Ward et al., 2013), and plant

senescence, which coincides with a pulse input of labile C from plants to soils (Bardgett et al., 2005).

Temperature-accelerated biochemical kinetics and increased electron donor supply are mechanisms that can increase methanogenesis without necessarily shifting methanogenic pathways. However, shifts in the balance between hydrogenotrophic, acetoclastic, and methylotrophic methanogenesis pathways can be expected with warming. For example, in Arctic permafrost, methylotrophic methanogenesis was found to be more sensitive to warming than the other pathways (de Jong et al., 2018). Such shifts can be quantified with future analyses of H_2 and low-molecular-weight organic compounds (e.g., Bridgman et al., 2013; Yang et al., 2016), isotopic tracing of specific methanogenic pathways (e.g., Blaser and Conrad, 2016; Conrad, 2005; Neumann et al., 2016; Whiticar, 1999), and molecular community analyses (e.g., Bridgman et al., 2013; He et al., 2015; Wilson et al., 2016).

4.3 Whole-ecosystem warming reduces competition with sulfate reducers

While low-molecular-weight organic compounds are electron donors for acetoclastic methanogenic respiration, they are also substrates for other microbial groups such as SO_4 reducers (Megonigal et al., 2004; Ye et al., 2014). As a result, consumption of the limited organic carbon supply by SO_4 reducers should (and often does) limit methanogenic activity, such that terminal microbial respiration is typically dominated by SO_4 reduction in brackish marshes (Sutton-Grier et al., 2011). Similarly, SO_4 reducers are more efficient than methanogens at competing for the H_2 required for CO_2 reduction (Kristjansson et al., 1982). We did not measure rates of SO_4 reduction in this study but can use SO_4 depletion as a proxy; more SO_4 depletion indicates that more SO_4 reduction has occurred. Warming generally increased SO_4 depletion, especially in the plots dominated by *Schoenoplectus* (Fig. 6b). Differences in SO_4 depletion between plots are not driven by SO_4 inputs because the only supply of SO_4 is the tidal flow, which is the same for all plots in each community of the experiment. Instead, higher rates of SO_4 reduction are most likely driven by some combination of electron donor supply and kinetics. While SO_4 reducers likely benefited from the increased availability of electron donors, as described above, the kinetics of SO_4 reduction also respond strongly to temperature (Weston and Joye, 2005).

When SO_4 concentrations drop below a threshold concentration, SO_4 reduction becomes SO_4 -limited, rather than electron-donor-limited (Megonigal et al., 2004). A review of the coastal wetland CH_4 literature estimated this threshold at 4 mmol SO_4 (Poffenbarger et al., 2011), a value that is consistent with patterns of porewater CH_4 and SO_4 at the GCReW site (Keller et al. 2009). As SO_4 and O_2 are the dominant electron-accepting compounds that suppress methanogenesis in this organic soil, this drawdown then re-

leases the methanogens from substrate competition (Fig. 7, mechanism 3). Here, we show that SO_4 is typically below 4 mmol in the +5.1 plots in the C_3 community and in all plots in the C_4 community (Figs. 6, S5). The drawdown of SO_4 may also reduce rates of anaerobic CH_4 oxidation (Hinrichs and Boetius, 2003). Van Hulzen et al. (1999) proposed a multi-phase system in a warming incubation experiment, observing that first methanogens are outcompeted for substrates by other microbes, next CH_4 production increases as the supply of inhibiting TEA decreases, and finally TEA availability is reduced to the point that methanogenesis is controlled only by the supply of electron donors. Warming in this study decreased the time required for the system to pass through the first two phases (van Hulzen et al., 1999). In our experiment, this final phase of increased methanogenic activity occurs when SO_4 concentrations dip below 4 mmol $SO_4 L^{-1}$, which occurs most often in the +5.1 °C plots, especially in the C_3 community. This interpretation is also supported by the long-term record of porewater chemistry from an allied experiment at the site, demonstrating that porewater CH_4 concentrations increase as SO_4 concentrations decrease (Keller et al., 2009).

Methanogens may also have a competitive advantage over SO_4 reducers for electron donor consumption at warmer temperatures (van Hulzen et al., 1999). Sulfate reducers and methanogens have very similar K_M values for acetate, but the K_M for acetoclastic methanogenesis may decrease with temperature whereas K_M values for SO_4 reducers increase with temperature (van Bodegom and Stams, 1999). If this is the case in our system, then warming would allow methanogens to use a greater proportion of the available substrates.

4.4 Plant traits modify warming effects on CH_4 cycling

The three biogeochemical mechanisms we propose to explain a warming-induced increase in CH_4 emissions should interact strongly with plant responses to warming. Relationships between plant functional groups and CH_4 emissions have been demonstrated through field studies in other wetland ecosystems such as peatlands (Bubier et al., 1995; Ward et al., 2013) and in tidal wetland mesocosms (Liu et al., 2019; Martin and Moseman-Valtierra, 2017; Mueller et al., 2020). We provide field evidence that two species with distinct plant traits – *Schoenoplectus* and *Spartina* – have strikingly different effects on CH_4 emissions from brackish wetlands. *Spartina*-dominated communities had consistently higher CH_4 emissions under both ambient and warmed conditions (Fig. 3). In most years, *Schoenoplectus* biomass was negatively correlated with CH_4 emissions, while *Spartina* and *Distichlis* biomass was positively correlated. Vegetation effects are typically strongest during the growing season, when the plants are actively altering rhizosphere biogeochemistry (van der Nat and Middelburg, 1998b; Ward et al., 2013), which is consistent with our observations in this study.

As with warming effects, plant-driven shifts in CH₄ emissions are the result of differing rates of CH₄ production, oxidation, transport, or a combination of these processes, but sustained differences in emissions cannot be attributed only to transport, as discussed previously. Instead, the stimulation of CH₄ emissions is likely due to changes in the plant-mediated supply of electron acceptors and electron donors. In a field environment, differentiating between species-specific effects and underlying environmental conditions can be difficult, but mesocosm studies that control all environmental factors have also found species-specific effects on CH₄ cycling (e.g., Liu et al., 2014). Plants can alter CH₄ cycling by adding O₂ (electron acceptor) or C substrates (electron donors) to the rhizosphere, altering the redox state. We propose that *Schoenoplectus* is a net oxidizer of the rhizosphere and that *Spartina* is a net reducer, and thus their presence and productivity have opposing effects on CH₄ emissions (Fig. 7, mechanism 4).

4.4.1 *Schoenoplectus* oxidizes the rhizosphere, increasing CH₄ oxidation

Species vary in their capacity to support aerobic CH₄ oxidation (van der Nat and Middelburg, 1998b), and *Schoenoplectus* appears to support higher rates of aerobic CH₄ oxidation than *Spartina* (Mueller et al., 2020). *Scirpus lacustris* is morphologically similar to *Schoenoplectus americanus* studied here and has been demonstrated to have substantial rhizosphere oxidation capacity, especially during the growing season (van der Nat and Middelburg, 1998b). Consequently, these plants likely exert stronger control on rates of CH₄ oxidation than rates of methanogenesis (van der Nat and Middelburg, 1998a). We hypothesize that the relatively high capacity of *Schoenoplectus* to transport O₂ held the CH₄ emissions stimulation caused by modest levels of warming (+1.7 to +3.4 °C) to rates similar to under ambient conditions (Fig. 3). At high warming (+5.1 °C), however, *Schoenoplectus* community CH₄ emissions drastically increase (Fig. 3). We suggest that this is due to the combined effects of the three mechanisms discussed previously, namely the differences in the Q_{10} values of CH₄ production and CH₄ oxidation, the increased supply of organic substrate through plant productivity, and the decrease in competition for electron donors due to SO₄ depletion. Collectively, when the ecosystem is warmed above current ambient conditions by 5 °C or more, enhanced stimulation of CH₄ production starts to offset some of the *Schoenoplectus* oxidation effect. This also offers an explanation for the positive correlation between *Schoenoplectus* biomass and CH₄ emissions observed in 2016 as that was the hottest of the 4 years in this study.

4.4.2 *Spartina* reduces the rhizosphere, increasing CH₄ production

The variability in quality and quantity of root exudates between plant functional types is well known to affect microbial community composition and activity (Deyn et al., 2008). Methanogenesis responses to warming in incubation studies are related to the lignin and cellulose content of the peat, which in turn depends on the plant functional type from which the peat developed (Duval and Radu, 2018). Although warming likely increases substrate availability across the whole experiment, the production of labile, low-molecular-weight C substrates through fermentation is less sensitive to temperature above 25 °C than below this threshold (Neubauer and Craft, 2009; Weston and Joye, 2005). Microorganisms may also preferentially use freshly produced (i.e., labile) organic carbon compounds as electron donors (DeLaune et al., 2014), and consequently warming effects on CH₄ production should be strongest in a system where rates of root exudation and turnover are most rapid. We propose that root exudation and turnover explain the positive correlation between plant biomass and CH₄ emissions that we observed in the C₄ community. Multiple years of porewater chemistry at this site show that *Spartina*-dominated communities have higher DOC and dissolved CH₄ than adjacent *Schoenoplectus* communities (Keller et al., 2009; Marsh et al., 2005). Though we did not directly measure root exudation, porewater DOC is partially derived from root exudates and has been used as a proxy to understand the responses of root exudates to global change factors (Dieleman et al., 2016; Fenner et al., 2007; Jones et al., 2009).

We observed a simultaneous increase in dissolved CH₄ at the soil surface and a decrease in dissolved CH₄ at depth in the warmed C₄ plots. As with the observed trends in CH₄ emissions, there are multiple mechanisms that could cause a shift in porewater CH₄ concentrations. Of the four mechanisms outlined above, perhaps the simplest explanation is an increase in labile C at shallow depths and a decrease in deeper soil. This is consistent with DOC depth profiles from this C₄ community in which porewater DOC increases with warming in shallow samples but decreases with warming in deep samples (Fig. S7). This shallowing of peak DOC concentrations could be due to a warming-induced increase in evapotranspiration, leading to slower downward hydrologic transport of DOC-rich surface porewater to lower depths, or a warming-induced shallowing of the root system, leading to a shift in the location of root exudates.

In most years, *Spartina* biomass was positively correlated with CH₄ emissions, supporting our hypothesis that *Spartina* favors net CH₄ production. However, in 2016 *Spartina* biomass and CH₄ emissions were negatively correlated. Prior work at this site has indicated that *Spartina* and *Distichlis* biomass is more negatively affected by hot and dry growing conditions than *Schoenoplectus* (Noyce et al., 2019) due in part because the *Spartina* and *Distichlis* (C₄) communities

are less frequently inundated. The 2016 growing season was substantially warmer than average (Table 2), and the heating treatments were initialized on 1 June of that year, after the annual plants had already established and may have developed adaptations to ambient, rather than elevated, temperature conditions. The combination of these two effects likely led to heat stress, reducing the root exudates supplied to the rhizosphere microbial community (Heckathorn et al., 2013) and thus minimizing the *Spartina* stimulation effect.

4.5 Comparisons with prior data

Methane emissions have been measured at the GCRW site previously, but this study represents the most comprehensive dataset collected to date and is thus particularly useful for advancing the process-based understanding needed to improve prognostic models. Overall, our flux estimates are lower than those reported previously. The earliest CH₄ fluxes were measured in a single month (July) in *Schoenoplectus*-dominated plots and reported to be 331 to 6883 $\mu\text{mol m}^{-2} \text{d}^{-1}$ (Dacey et al., 1994), much higher than our range of 359 to 1651 $\mu\text{mol m}^{-2} \text{d}^{-1}$ for ambient temperature *Schoenoplectus* plots in July. Similarly, Marsh et al. (2005) reported mean growing season (May–October) CH₄ emissions from this site of $846 \pm 111 \mu\text{mol CH}_4 \text{ m}^{-2} \text{d}^{-1}$, whereas we measured $656 \pm 79 \mu\text{mol CH}_4 \text{ m}^{-2} \text{d}^{-1}$ over the same months. Finally, Pastore et al. (2017) estimated average annual fluxes in their *Schoenoplectus*-dominated ambient CO₂ plots as $3.1 \pm 1.7 \text{ g CH}_4 \text{ m}^{-2} \text{yr}^{-1}$, compared to our estimates of $1.6 \pm 0.3 \text{ g CH}_4 \text{ m}^{-2} \text{yr}^{-1}$ for *Schoenoplectus* plots. The different estimates by these studies may be partly due to inter-annual variability as demonstrated in our data where 2018 had substantially higher fluxes than any of the surrounding years (Table S2).

The annual estimates reported here for ambient temperature plots trended lower than published mean CH₄ emissions for mesohaline tidal marshes. Our plots ranged from 0.7 to 9.3 $\text{g CH}_4 \text{ m}^{-2} \text{yr}^{-1}$ (mean = 9.3), compared to the range of 3.3 to 16.4 $\text{g CH}_4 \text{ m}^{-2} \text{yr}^{-1}$ (mean = 16.4) reported by Poffenbarger et al. (2011). This difference may be explained by the fact that there was significant within-class variation in the oligohaline and mesohaline salinity classes that was unexplained, and their assessment was based on too few data points to fully capture the variation that is expected to exist in the mesohaline class. Indeed, subsequent studies have documented fluxes well below 3 $\text{g CH}_4 \text{ m}^{-2} \text{yr}^{-1}$ (Krauss and Whitbeck, 2012) and even negative fluxes (Al-Haj and Fulweiler, 2020). We hypothesize that the low fluxes measured at our site reflect *Schoenoplectus americanus* traits that favor CH₄ oxidation more than CH₄ production and that the high end of our range was limited by the high soil elevation (i.e., deep water table) of areas dominated by *Spartina patens*, offsetting the influence of *S. patens* traits that favor CH₄ production.

4.6 Implications for tidal wetland carbon cycling

Warming accelerates rates of CH₄ emissions from brackish marshes, especially during the growing season. This is driven by both direct and indirect warming effects and mediated by soil biogeochemistry, but the magnitude of the warming effect is also dependent on traits of the plant species that dominate the plant community. Communities dominated by *Spartina patens* increase net CH₄ emissions in response to smaller increments of warming than communities dominated by *Schoenoplectus americanus*. *Spartina*-dominated sites may thus have a higher likelihood of shifting from a net C sink to a net C source under future warming conditions, due to this increased loss of C as CH₄. However, this effect could be mitigated if these high-elevation *Spartina* marshes become dominated by *Schoenoplectus* in response to predicted accelerated sea-level rise (Kirwan and Guntenspergen, 2012). In addition, *Spartina* traits are plastic and influenced by factors such as soil redox conditions (Kludze and DeLaune, 1994), salinity (Crozier and DeLaune, 1996), and water level (Liu et al., 2019), all of which can be expected to change plant-mediated effects on CH₄ biogeochemistry. Further studies are needed to thoroughly assess the range of environmental conditions under which *Spartina* is a net reducer and *Schoenoplectus* is a net oxidizer as proposed by the present study.

Data availability. All data are available from the corresponding author upon request.

Supplement. The supplement related to this article is available online at: <https://doi.org/10.5194/bg-18-2449-2021-supplement>.

Author contributions. GLN and JPM designed the study, GLN collected and analyzed the data, and GLN and JPM wrote the paper.

Competing interests. The authors declare that they have no conflict of interest.

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