

Influence of the dynamically disordered N-terminal tail domain on the amyloid core structure of human Y145Stop prion protein fibrils

1 Zhe Qi¹, Krystyna Surewicz², Witold K. Surewicz², Christopher P. Jaroniec^{1*}

2 ¹Department of Chemistry and Biochemistry, The Ohio State University, Columbus, OH, USA

3 ²Department of Physiology and Biophysics, Case Western Reserve University, Cleveland, OH, USA

4 * **Correspondence:**

5 Christopher P. Jaroniec

6 jaroniec.1@osu.edu

7 **Keywords: amyloid, prion, octarepeat, intrinsically disordered region/protein, magic-angle**
8 **spinning (MAS) solid-state NMR**

9

10 Abstract

11 The Y145Stop mutant of human prion protein (huPrP23-144) is associated with a familial prionopathy
12 and provides a convenient *in vitro* model for investigating amyloid strains and cross-seeding barriers.
13 huPrP23-144 fibrils feature a compact and relatively rigid parallel in-register β -sheet amyloid core
14 spanning ~30 C-terminal amino acid residues (~112-141) and a large ~90-residue dynamically
15 disordered N-terminal tail domain. Here, we systematically evaluate the influence of this dynamic
16 domain on the structure adopted by the huPrP23-144 amyloid core region, by investigating using
17 magic-angle spinning solid-state nuclear magnetic resonance (NMR) spectroscopy a series of fibril
18 samples formed by huPrP23-144 variants corresponding to deletions of large segments of the N-
19 terminal tail. We find that deletion of the bulk of the N-terminal tail, up to residue 98, yields amyloid
20 fibrils with native-like huPrP23-144 core structure. Interestingly, deletion of additional flexible
21 residues in the stretch 99-106 located outside of the amyloid core yields shorter heterogenous fibrils
22 with fingerprint NMR spectra that are clearly distinct from those for full-length huPrP23-144,
23 suggestive of the onset of perturbations to the native structure and degree of molecular ordering for the
24 core residues. For the deletion variant missing residues 99-106 we show that native huPrP23-144 core
25 structure can be “restored” by seeding the fibril growth with preformed full-length huPrP23-144 fibrils.

Introduction

Most peptide and protein molecules are capable of undergoing conformational conversion from their native state into highly ordered, β -sheet rich amyloid fibrils (Dobson, 1999), and for ~50 human proteins such misfolding and amyloid formation can occur under physiological conditions *in vivo* leading to development of disease (Chiti and Dobson, 2006). A number of amyloids have been found to contain large dynamically disordered domains flanking the structured fibril core (Heise *et al.*, 2005; Siemer *et al.*, 2006; Loquet *et al.*, 2009; Helmus *et al.*, 2010; Bibow *et al.*, 2011; Li *et al.*, 2012; Raveendra *et al.*, 2013; Frederick *et al.*, 2014; Isas, Langen and Siemer, 2015; Cervantes *et al.*, 2016; Lin *et al.*, 2017; Murray *et al.*, 2017; Caulkins *et al.*, 2018; Dregni *et al.*, 2019; Fonda *et al.*, 2021), and it has been suggested that the presence of these conformationally flexible domains may be of pathological or functional significance by stabilizing fibril structures and mediating interactions involving protofilaments (Uversky and Fink, 2004; Chiti and Dobson, 2006; Tompa, 2009; van der Wel, 2017; Siemer, 2020). The detailed characterization of dynamically disordered regions in amyloids (and in other large biomacromolecular assemblies) has generally been pursued by multidimensional magic-angle spinning (MAS) nuclear magnetic resonance (NMR) techniques, which are able to visualize these domains directly in hydrated samples at ambient temperature by using experiments based on scalar coupling mediated polarization transfers (Tycko, 2006; van der Wel, 2017; Jaroniec, 2019; Siemer, 2020).

The C-terminally truncated Y145Stop prion protein (PrP23-144) variant is associated with a hereditary prionopathy in humans (Ghetti *et al.*, 1996), and mouse PrP23-144 amyloid fibrils have recently been shown to cause transmissible prion disease in mice (Choi *et al.*, 2016). Importantly, the highly homologous human (hu), mouse (mo) and Syrian hamster (Sha) PrP23-144 proteins (pairwise amino acid, aa, sequence identities of ~90-95%) have also been shown to provide a valuable *in vitro* model for detailed investigation of the structural basis of amyloid strains and transmissibility barriers (Kundu *et al.*, 2003; Vanik, Surewicz and Surewicz, 2004; Jones and Surewicz, 2005; Surewicz, Jones and Apetri, 2006). Our previous structural and dynamic solid-state NMR studies of huPrP23-144 fibrils revealed the presence of a structured ~30-residue parallel in-register β -amyloid core (aa ~112-141) exhibiting limited protein backbone motions on the ~0.1-1 ms time scale located near the C-terminus and a large dynamically disordered ~90-residue N-terminal tail domain (aa ~23-110) (Helmus *et al.*, 2008, 2010, 2011; Theint *et al.*, 2018; Aucoin *et al.*, 2019; Shannon *et al.*, 2019). Additional studies of PrP23-144 amyloids containing mutations and deletions corresponding to different huPrP23-144 core residues enabled these sequence modifications to be correlated with structural and dynamic changes in

the PrP23-144 amyloid core and provided initial insights into mammalian PrP23-144 cross-seeding specificities (Jones *et al.*, 2011; Theint, Nadaud, Aucoin, *et al.*, 2017; Theint, Nadaud, Surewicz, *et al.*, 2017; Dao *et al.*, 2021).

Previous studies of amyloids formed by full-length prion protein (PrP23-231) and the proteinase-K resistant 90-231 fragment of transmissible spongiform encephalopathy associated mammalian PrP deposits (Prusiner, 1998) suggest that the flexible N-terminal domain may play a role in PrP aggregation properties, and prion structure and pathogenesis (Weissmann, 1999; Lawson *et al.*, 2004; Baskakov and Bocharova, 2005; Frankenfield, Powers and Kelly, 2005). The present study aims to assess the influence of the dynamic huPrP23-144 N-terminal region on amyloid assembly and resulting β -core conformation. This is achieved by performing systematic solid-state NMR, atomic force microscopy (AFM) and thioflavin T (ThT) fluorescence studies on fibril samples formed *in vitro* from recombinant huPrP23-144 variants corresponding to deletions of large segments of the N-terminal tail. Overall, we find that the majority of dynamically disordered N-terminal tail residues, including the octarepeat region (aa 51-91) implicated in copper binding and homeostasis (Millhauser, 2007; Aguzzi, Baumann and Bremer, 2008), have little impact on the fibril assembly kinetics and ability of the deletion variants to adopt the native huPrP23-144 amyloid core structure. However, we also find that a stretch of ~ 10 conformationally flexible residues that precede the amyloid core region in huPrP23-144 fibrils appears to play a role in the ability to adopt the native core structure and the degree of molecular ordering within the core.

Results

Previous solid-state NMR studies indicate that the relatively rigid β -core region of huPrP23-144 fibrils consists of residues 112-141 (Helmus *et al.*, 2008, 2010, 2011; Shannon *et al.*, 2019). In contrast residues 23-111 and 142-144 are not observable in conventional cross-polarization magic angle spinning (CP-MAS) solid-state NMR spectra that utilize dipolar coupling-based polarization transfers, consistent with their increased mobility (van der Wel, 2017; Siemer, 2020) while most of these residues can be detected in MAS NMR spectra utilizing polarization transfers mediated via J-couplings (van der Wel, 2017; Siemer, 2020). To assess the potential influence of the dynamically disordered N-terminal domain of huPrP23-144 on the conformation adopted by the amyloid core region we generated a series of fibril samples from large N-terminal domain deletion variants of huPrP23-144 and examined their fibrillization kinetics, morphologies and molecular conformations by using ThT fluorescence, AFM and solid-state NMR, respectively.

The huPrP23-144 deletion variants employed in these studies spanned residues 28-106 – note that the short segment (aa 23-27) containing multiple lysine and arginine residues (as well as the N-terminal GDSP extension present in our huPrP23-144 construct (Helmus *et al.*, 2008) was not deleted in order to ensure the solubility of the different deletion variants. Initially, we investigated the following huPrP23-144 variants: Δ 28-50, Δ 51-91 (corresponding to deletion of the entire octarepeat region) and Δ 92-106 (see Figure 3a for the huPrP23-144 protein sequence and summary of the deletion variants studied). Briefly, fibrils generated from the Δ 28-50 and Δ 51-91 constructs were found to exhibit wild-type (WT) like morphologies and molecular conformations, while significant differences relative to WT were observed for the Δ 92-106 fibrils. To investigate this further we prepared the Δ 92-98 variant, which was found to form WT-like fibrils suggesting that deletion of huPrP23-144 N-terminal residues up to aa 98 does not have a significant impact on formation of the native huPrP23-144 amyloid core structure. Based on these findings we then generated the Δ 28-98 variant corresponding to the deletion of nearly the entire huPrP23-144 N-terminal tail. While the Δ 28-98 construct expressed at reasonable level in rich medium it was found not to express at sufficiently high level in ^{13}C and ^{15}N isotope enriched minimal medium to permit multidimensional solid-state NMR studies, which led us to generate an additional, Δ 28-91, deletion variant. The studies of all the aforementioned huPrP23-144 N-terminal deletion variants are described in additional detail below.

As noted above, all the huPrP23-144 deletion variants investigated in this study (Δ 28-50, Δ 28-91, Δ 28-98, Δ 51-91, Δ 92-98 and Δ 92-106) readily converted to amyloid fibrils in autocatalytic, unseeded reactions carried out in potassium phosphate buffer at pH 6.4. The kinetics of fibril formation were monitored by the standard ThT binding assay (Naiki *et al.*, 1989) revealing a nearly identical ~3-4 h lag phase for WT huPrP23-144 and all the deletion variants at 400 μM protein concentration, in line with the value reported previously for WT huPrP23-144 (Kundu *et al.*, 2003). Representative data for WT, Δ 28-91 and Δ 92-106 huPrP23-144 are shown in Figure 1. Furthermore, we found that addition of a small amount of pre-formed WT huPrP23-144 fibril seeds to the reaction resulted in complete elimination of the lag phase for all deletion variants studied (Figure 1).

Atomic force microscopy was then used to investigate the morphologies of the resulting amyloid fibrils. With exception of the Δ 92-106 fibrils, all the other deletion variants displayed morphologies that were similar to one another as well as to the morphology of WT huPrP23-144 amyloid. Specifically, as shown in Figure 2, these fibrils had highly uniform, micron long, threadlike morphologies with left-handed twist characterized by heights of ~5-6 nm and periodicities of ~30 nm.

In contrast, the $\Delta 92-106$ fibril sample was far more heterogeneous, containing shorter fibrils of varying lengths in the range of tens to hundreds of nanometers as well as considerable amounts of apparently amorphous, non-fibrillar aggregates (Figure 2c).

Finally, in order to compare the protein conformations and degree of molecular ordering for the different fibril samples at the atomic level, we recorded two-dimensional (2D) fingerprint ^{15}N - $^{13}\text{C}\alpha$ chemical shift correlation solid-state NMR spectra for wild-type huPrP23-144 amyloid and all the deletion variants except $\Delta 28-98$. Figure 3 shows the resulting 2D ^{15}N - $^{13}\text{C}\alpha$ spectra, which were found to be effectively identical in terms of NMR signal frequencies, linewidths and relative intensities for WT huPrP23-144 and all deletion variants with exception of $\Delta 92-106$ fibrils; the latter exhibited considerable spectral differences relative to the other samples (Figure 3f). Note that for fibrils formed from the $\Delta 28-98$ variant, which did not express at a sufficiently high level in ^{13}C and ^{15}N isotope labeled minimal media to permit collection of a ^{15}N - $^{13}\text{C}\alpha$ correlation spectrum, a 1D ^{13}C CP-MAS spectrum could be recorded for unlabeled fibrils (Figure 4). Apart from the obviously lower sensitivity associated with the use of a natural abundance sample, this spectrum showed considerable similarity to the reference ^{13}C CP-MAS spectrum recorded for ^{13}C , ^{15}N -labeled WT huPrP23-144 fibrils. Altogether these data indicate that other than $\Delta 92-106$, all the huPrP23-144 deletion variants studied form amyloids that are highly ordered at the atomic level and possess WT-like core structures and core residue backbone motions. On the other hand, the spectra of $\Delta 92-106$ fibrils are indicative of structural perturbation/polymorphism for the core residues and a higher degree of molecular disorder, consistent with the higher degree of sample heterogeneity observed by AFM. Comparison of 2D ^{15}N - $^{13}\text{C}\alpha$ spectra for WT and $\Delta 92-106$ fibrils reveals that they key chemical shift perturbations involve residues ~112-115. This suggests that conformations of the $\beta 1$ -strand and several following core residues are primarily impacted in the deletion variant, in line with earlier studies (Theint, Nadaud, Aucoin, *et al.*, 2017) indicating that several residues in this regime appear to be key for stabilizing the core fold of WT huPrP23-144 amyloid. Remarkably, however, we also find that seeding $\Delta 92-106$ amyloid formation with pre-formed WT huPrP23-144 fibrils results in the structural adaptation of the $\Delta 92-106$ protein to the WT huPrP23-144 amyloid core fold as evidenced by their indistinguishable 2D ^{15}N - $^{13}\text{C}\alpha$ spectra (Figure 3g).

Discussion

Collectively, on the basis of effectively identical fibril assembly kinetics, morphologies and fingerprint solid-state NMR spectra for all huPrP23-144 variants studied containing large deletions up to residue 98, our results indicate that the bulk of the dynamically disordered N-terminal tail domain of huPrP23-144 is not essential for amyloid formation under autocatalytic conditions and ability of the resulting β -core region to adopt a WT-like structure. Remarkably, this finding is strongly correlated with our previously reported data showing that proteinase-K resistant fragments of WT huPrP23-144 amyloid fibrils span residues 97-144, 98-144 and 99-144 (Jones *et al.*, 2011).

Combined with the finding that the protein conformation and degree of molecular ordering appear to be significantly perturbed for the Δ 92-106 deletion variant relative to WT huPrP23-144 and the different large huPrP23-144 deletion variants up to residue 98, our data indicate that aa ~99-106 (and presumably several additional amino acids preceding the structured and relatively rigid fibril core beginning around residue Met-112) play a key role in stabilizing the formation of the characteristic huPrP23-144 β -core structure in spite of their flexible nature and location outside of the structured amyloid core region. Given that the 100-110 stretch of huPrP23-144 contains four positively charged lysine residues (at positions 101, 104, 106 and 110) we speculate that the stabilization of the amyloid core structure in WT huPrP23-144 occurs via electrostatic interactions with the negatively charged C-terminal aspartate residue.

While deletion of amino acids in the 99-106 regime clearly impacts the amyloid core structure and molecular ordering of huPrP23-144 within fibrils formed under autocatalytic conditions as discussed above, we also find that the seeding of fibril formation by the Δ 92-106 deletion variant with pre-formed WT huPrP23-144 fibrils at low concentration leads to the Δ 92-106 proteins adopting a WT-like core structure at the atomic level as revealed by the fingerprint ^{15}N - $^{13}\text{C}\alpha$ NMR spectrum that is effectively identical to the corresponding spectrum for WT huPrP23-144 amyloid. Interestingly, this structural templating process that yields a WT-like fold for Δ 92-106 amyloid was found to not appreciably alter the overall morphology of the Δ 92-106 fibrils as viewed by AFM, with the majority of the sample consisting of relatively short fibrils similar to those observed in the unseeded reaction.

In summary, we systematically evaluated the influence of the dynamically disordered N-terminal tail domain of huPrP23-144 on the structure adopted by the amyloid core region by using deletion mutagenesis combined with magic-angle spinning solid-state NMR spectroscopy. We find that N-terminal huPrP23-144 residues up to aa 98, which coincide with the protein segment most susceptible

to proteinase-K digestion in mature fibrils, can be deleted without impacting the core structure formed in an autocatalytic fibril assembly process. Remarkably, deletion of additional flexible residues (aa 99-106) located outside the amyloid core leads to the formation of amyloid fibrils with a perturbed core structure and reduced degree of molecular ordering in the fibril lattice, most likely caused by the disruption of stabilizing electrostatic interactions involving several lysine side-chains found in this region and the C-terminal aspartate residue. These structural perturbations, however, can be alleviated by catalyzing the amyloid formation with preformed WT huPrP23-144 fibril seeds.

Materials and Methods

Protein expression and purification. The plasmid encoding human PrP23-144 was described previously (Kundu *et al.*, 2003) and plasmids encoding huPrP23-144 deletion variants were generated similarly to our previous study (Jones *et al.*, 2011) by deletion mutagenesis using a QuikChange kit (Stratagene). Uniformly ^{13}C , ^{15}N labeled huPrP23-144 and deletion variants were expressed in *E. coli* BL21 (DE3) cells and purified by nickel affinity chromatography as described in detail in previous studies (Theint, Nadaud, Surewicz, *et al.*, 2017; Dao *et al.*, 2021). The identities and purities of the resulting proteins were routinely confirmed by SDS/PAGE and MALDI mass spectrometry.

Amyloid fibril formation. Amyloid fibrils were prepared under quiescent conditions at 25 °C as described in previous studies (Theint, Nadaud, Aucoin, *et al.*, 2017; Theint, Nadaud, Surewicz, *et al.*, 2017; Dao *et al.*, 2021). Lyophilized huPrP23-144 variants were dissolved in ultrapure water at ~400 μM concentration and 1 M potassium phosphate buffer at pH 6.4 was added to a final concentration of 50 mM. For samples seeded with WT huPrP23-144 amyloid, 2% (mole fraction) of preformed fibrils was added immediately to the reaction after addition of phosphate buffer.

Thioflavin T fluorescence and atomic force microscopy. Kinetics of fibril formation were monitored via the standard ThT fluorescence assay (Naiki *et al.*, 1989) as described in detail in previous studies (Theint *et al.*, 2018). Fibril morphologies were assessed by atomic force microscopy (AFM) as follows. Fibril suspensions were diluted 50-fold in ultrapure water, deposited on freshly cleaved mica substrates (Ted Pella Inc.) for 5 min, rinsed with three 50 μL aliquots of ultrapure water, and allowed to air dry for 1-2 h prior to imaging. Images were collected using a Bruker Dimension Icon AFM in PeakForce

quantitative nanomechanical mapping mode with a ScanAsyst-Air probe and processed with the Bruker NanoScope Analysis software.

Solid-state NMR spectroscopy. Fibril suspensions for solid-state NMR analysis were incubated as described above for 48 h and centrifuged. The fibril pellets were washed three times with 50 mM potassium phosphate pH 6.4 buffer and packed into 3.2 mm zirconia rotors by centrifugation with the final samples containing ~5-10 mg of fibrils. Standard 1D ^{13}C CP-MAS and 2D ^{15}N - $^{13}\text{C}\alpha$ solid-state NMR spectra were recorded on 500 MHz Varian and 800 MHz Bruker spectrometers, equipped with 3.2 mm triple resonance (^1H - ^{13}C - ^{15}N) BioMAS and E^{free} probes, respectively. The sample temperature and MAS frequencies were actively regulated at 5 °C and 11.111 kHz, respectively, and the experimental parameters were similar to those used in our previous solid-state NMR studies of WT huPrP23-144 fibrils (Helmus *et al.*, 2008) with data acquisition times for the 2D ^{15}N - $^{13}\text{C}\alpha$ NMR spectra of 8-16 h per sample. NMR spectra were processed and analyzed using NMRPipe (Delaglio *et al.*, 1995) and Sparky (Goddard and Kneller, 2006), respectively.

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contributions

WKS and CPJ designed the study. ZQ and KS prepared samples and performed experiments. ZQ prepared figures. ZQ, WKS and CPJ wrote the manuscript.

Funding

This work was supported by grants from NIH (R01GM094357 to CPJ and WKS and S10OD012303 to CPJ) and NSF (MCB-1715174 to CPJ).

Acknowledgments

We thank Dr. Sean Smrt for stimulating discussions and insightful comments on the manuscript.

References

- Aguzzi, A., Baumann, F. and Bremer, J. (2008) The prion's elusive reason for being, *Annu. Rev. Neurosci.* 31, 439–477. doi: 10.1146/annurev.neuro.31.060407.125620.
- Aucoin, D. *et al.* (2019) Protein-solvent interfaces in human Y145Stop prion protein amyloid fibrils probed by paramagnetic solid-state NMR spectroscopy, *J. Struct. Biol.* 206(1), 36–42. doi: 10.1016/j.jsb.2018.04.002.
- Baskakov, I. V and Bocharova, O. V (2005) In vitro conversion of mammalian prion protein into amyloid fibrils displays unusual features, *Biochemistry.* 44(7), 2339–2348. doi: 10.1021/bi048322t.
- Bibow, S. *et al.* (2011) The dynamic structure of filamentous tau, *Angew. Chem. Int. Ed.* 50(48), 11520–11524. doi: 10.1002/anie.201105493.
- Caulkins, B. G. *et al.* (2018) Dynamics of the proline-rich C-Terminus of huntingtin exon-1 fibrils, *J. Phys. Chem. B.* 122(41), 9507–9515. doi: 10.1021/acs.jpcb.8b09213.
- Cervantes, S. A. *et al.* (2016) Identification and structural characterization of the N-terminal amyloid core of Orb2 isoform A, *Sci. Rep.* 6, 38265. doi: 10.1038/srep38265.
- Chiti, F. and Dobson, C. M. (2006) Protein misfolding, functional amyloid, and human disease, *Annu. Rev. Biochem.* 75, 333–366. doi: 10.1146/annurev.biochem.75.101304.123901.
- Choi, J.-K. *et al.* (2016) Amyloid fibrils from the N-terminal prion protein fragment are infectious, *Proc. Natl. Acad. Sci. USA* 113(48), 13851–13856. doi: 10.1073/pnas.1610716113.
- Dao, H. H. *et al.* (2021) ¹³C and ¹⁵N chemical shift assignments of A117V and M129V human Y145Stop prion protein amyloid fibrils, *Biomol. NMR Assign.* 15(1), 45–51. doi: 10.1007/s12104-020-09981-4.
- Delaglio, F. *et al.* (1995) NMRPipe: A multidimensional spectral processing system based on UNIX pipes, *J. Biomol. NMR*, 6(3), 277–293. doi: 10.1007/BF00197809.
- Dobson, C. M. (1999) Protein misfolding, evolution and disease, *Trends Biochem. Sci.* 24(9), 329–332. doi: 10.1016/s0968-0004(99)01445-0.
- Dregni, A. J. *et al.* (2019) In vitro 0N4R tau fibrils contain a monomorphic β -sheet core enclosed by dynamically heterogeneous fuzzy coat segments, *Proc. Natl. Acad. Sci. USA* 116(33), 16357–16366. doi: 10.1073/pnas.1906839116.
- Fonda, B. D. *et al.* (2021) Identification of the rigid core for aged liquid droplets of an RNA-binding protein low complexity domain, *J. Am. Chem. Soc.* 143(17), 6657–6668. doi: 10.1021/jacs.1c02424.
- Frankenfield, K. N., Powers, E. T. and Kelly, J. W. (2005) Influence of the N-terminal domain on the aggregation properties of the prion protein, *Prot. Sci.* 14(8), 2154–2166. doi: 10.1110/ps.051434005.
- Frederick, K. K. *et al.* (2014) Distinct prion strains are defined by amyloid core structure and chaperone binding site dynamics, *Chem. Biol.* 21(2), 295–305. doi: 10.1016/j.chembiol.2013.12.013.
- Ghetti, B. *et al.* (1996) Vascular variant of prion protein cerebral amyloidosis with tau-positive

- neurofibrillary tangles: the phenotype of the stop codon 145 mutation in PRNP, *Proc. Natl. Acad. Sci. USA* 93(2), 744–748. doi: 10.1073/pnas.93.2.744.
- Goddard, T. and Kneller, D. (2006) SPARKY 3. San Francisco: University of California.
- Heise, H. *et al.* (2005) Molecular-level secondary structure, polymorphism, and dynamics of full-length alpha-synuclein fibrils studied by solid-state NMR, *Proc. Natl. Acad. Sci. USA* 102(44), 15871–15876. doi: 10.1073/pnas.0506109102.
- Helmus, J. J. *et al.* (2008) Molecular conformation and dynamics of the Y145Stop variant of human prion protein in amyloid fibrils, *Proc. Natl. Acad. Sci. USA* 105(17), 6284–6289. doi: 10.1073/pnas.0711716105.
- Helmus, J. J. *et al.* (2010) Conformational flexibility of Y145Stop human prion protein amyloid fibrils probed by solid-state nuclear magnetic resonance spectroscopy, *J. Am. Chem. Soc.* 132(7), 2393–2403. doi: 10.1021/ja909827v.
- Helmus, J. J. *et al.* (2011) Intermolecular alignment in Y145Stop human prion protein amyloid fibrils probed by solid-state NMR spectroscopy, *J. Am. Chem. Soc.* 133(35), 13934–13937. doi: 10.1021/ja206469q.
- Isas, J. M., Langen, R. and Siemer, A. B. (2015) Solid-state nuclear magnetic resonance on the static and dynamic domains of huntingtin exon-1 fibrils, *Biochemistry*, 54(25), 3942–3949. doi: 10.1021/acs.biochem.5b00281.
- Jaroniec, C. P. (2019) Two decades of progress in structural and dynamic studies of amyloids by solid-state NMR, *J. Magn. Reson.* 306, 42–47. doi: 10.1016/j.jmr.2019.07.015.
- Jones, E. M. *et al.* (2011) Structural polymorphism in amyloids: new insights from studies with Y145Stop prion protein fibrils, *J. Biol. Chem.* 286(49), 42777–42784. doi: 10.1074/jbc.M111.302539.
- Jones, E. M. and Surewicz, W. K. (2005) Fibril conformation as the basis of species- and strain-dependent seeding specificity of mammalian prion amyloids, *Cell* 121(1), 63–72. doi: 10.1016/j.cell.2005.01.034.
- Kundu, B. *et al.* (2003) Nucleation-dependent conformational conversion of the Y145Stop variant of human prion protein: structural clues for prion propagation, *Proc. Natl. Acad. Sci. USA* 100(21), 12069–12074. doi: 10.1073/pnas.2033281100.
- Lawson, V. A. *et al.* (2004) Flexible N-terminal region of prion protein influences conformation of protease-resistant prion protein isoforms associated with cross-species scrapie infection in vivo and in vitro, *J. Biol. Chem.*, 279(14), 13689–13695. doi: 10.1074/jbc.M303697200.
- Li, J. *et al.* (2012) The RIP1/RIP3 necrosome forms a functional amyloid signaling complex required for programmed necrosis, *Cell* 150(2), 339–350. doi: 10.1016/j.cell.2012.06.019.
- Lin, H.-K. *et al.* (2017) Fibril polymorphism affects immobilized non-amyloid flanking domains of huntingtin exon1 rather than its polyglutamine core, *Nat. Comm.* 8, 15462. doi: 10.1038/ncomms15462.

- Loquet, A. *et al.* (2009) Prion fibrils of Ure2p assembled under physiological conditions contain highly ordered, natively folded modules, *J. Mol. Biol.* 394(1), 108–118. doi: 10.1016/j.jmb.2009.09.016.
- Millhauser, G. L. (2007) Copper and the prion protein: methods, structures, function, and disease, *Annu. Rev. Phys. Chem.* 58, 299–320. doi: 10.1146/annurev.physchem.58.032806.104657.
- Murray, D. T. *et al.* (2017) Structure of FUS protein fibrils and its relevance to self-Assembly and phase separation of low-complexity domains, *Cell* 171(3), 615–627.e16. doi: 10.1016/j.cell.2017.08.048.
- Naiki, H. *et al.* (1989) Fluorometric determination of amyloid fibrils in vitro using the fluorescent dye, thioflavin T1, *Anal. Biochem.* 177(2), 244–249. doi: 10.1016/0003-2697(89)90046-8.
- Prusiner, S. B. (1998) Prions, *Proc. Natl. Acad. Sci. USA*, 95(23), 13363–13383. doi: 10.1073/pnas.95.23.13363.
- Raveendra, B. L. *et al.* (2013) Characterization of prion-like conformational changes of the neuronal isoform of Aplysia CPEB, *Nat. Struct. Mol. Biol.* 20(4), 495–501. doi: 10.1038/nsmb.2503.
- Shannon, M. D. *et al.* (2019) Conformational dynamics in the core of human Y145Stop prion protein amyloid probed by relaxation dispersion NMR, *ChemPhysChem* 20(2), 311–317. doi: 10.1002/cphc.201800779.
- Siemer, A. B. *et al.* (2006) Observation of highly flexible residues in amyloid fibrils of the HET-s prion, *J. Am. Chem. Soc.* 128(40), 13224–13228. doi: 10.1021/ja063639x.
- Siemer, A. B. (2020) Advances in studying protein disorder with solid-state NMR, *Solid State Nucl. Magn. Reson.* 106, 101643. doi: 10.1016/j.ssnmr.2020.101643.
- Surewicz, W. K., Jones, E. M. and Apetri, A. C. (2006) The emerging principles of mammalian prion propagation and transmissibility barriers: Insight from studies in vitro, *Acc. Chem. Res.* 39(9), 654–662. doi: 10.1021/ar050226c.
- Theint, T., Nadaud, P. S., Surewicz, K., *et al.* (2017) ¹³C and ¹⁵N chemical shift assignments of mammalian Y145Stop prion protein amyloid fibrils, *Biomol. NMR Assign.* 11(1), 75–80. doi: 10.1007/s12104-016-9723-6.
- Theint, T., Nadaud, P. S., Aucoin, D., *et al.* (2017) Species-dependent structural polymorphism of Y145Stop prion protein amyloid revealed by solid-state NMR spectroscopy, *Nat. Comm.* 8(1), 753. doi: 10.1038/s41467-017-00794-z.
- Theint, T. *et al.* (2018) Structural studies of amyloid fibrils by paramagnetic solid-state nuclear magnetic resonance spectroscopy, *J. Am. Chem. Soc.* 140(41), 13161–13166. doi: 10.1021/jacs.8b06758.
- Tompa, P. (2009) Structural disorder in amyloid fibrils: its implication in dynamic interactions of proteins, *FEBS J.* 276(19), 5406–5415. doi: 10.1111/j.1742-4658.2009.07250.x.
- Tycko, R. (2006) Molecular structure of amyloid fibrils: insights from solid-state NMR, *Q. Rev.*

- 345 *Biophys.* 39(1), 1–55. doi: 10.1017/S0033583506004173.
- 346 Uversky, V. N. and Fink, A. L. (2004) Conformational constraints for amyloid fibrillation: The
- 347 importance of being unfolded, *Biochim. Biophys. Acta* 1698(2), 131–153. doi:
- 348 10.1016/j.bbapap.2003.12.008.
- 349 Vanik, D. L., Surewicz, K. A. and Surewicz, W. K. (2004) Molecular basis of barriers for
- 350 interspecies transmissibility of mammalian prions, *Mol. Cell.* 14(1), 139–145. doi: 10.1016/s1097-
- 351 2765(04)00155-8.
- 352 Weissmann, C. (1999) Molecular genetics of transmissible spongiform encephalopathies, *J. Biol.*
- 353 *Chem.* 274(1), 3–6. doi: 10.1074/jbc.274.1.3.
- 354 van der Wel, P. C. A. (2017) Insights into protein misfolding and aggregation enabled by solid-state
- 355 NMR spectroscopy, *Solid State Nucl. Magn. Reson.* 88, 1–14. doi: 10.1016/j.ssnmr.2017.10.001.
- 356
- 357
- 358

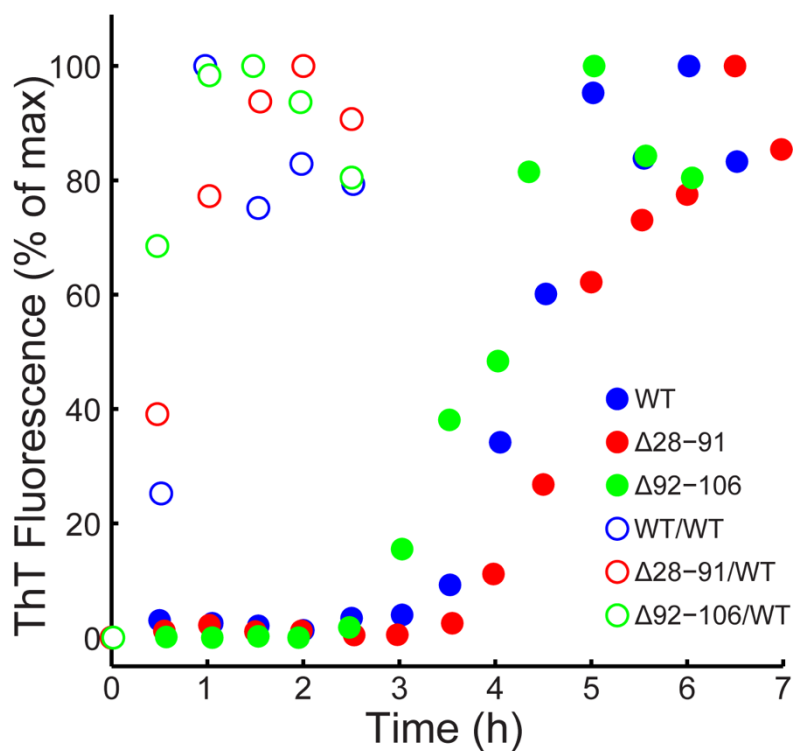


Figure 1. Kinetics of amyloid formation monitored by thioflavin T fluorescence in the absence (filled circles) and presence (open circles) of 2% (mole fraction) of WT huPrP23-144 fibril seeds for WT (blue), $\Delta 28-91$ (red) and $\Delta 92-106$ (green) huPrP23-144.

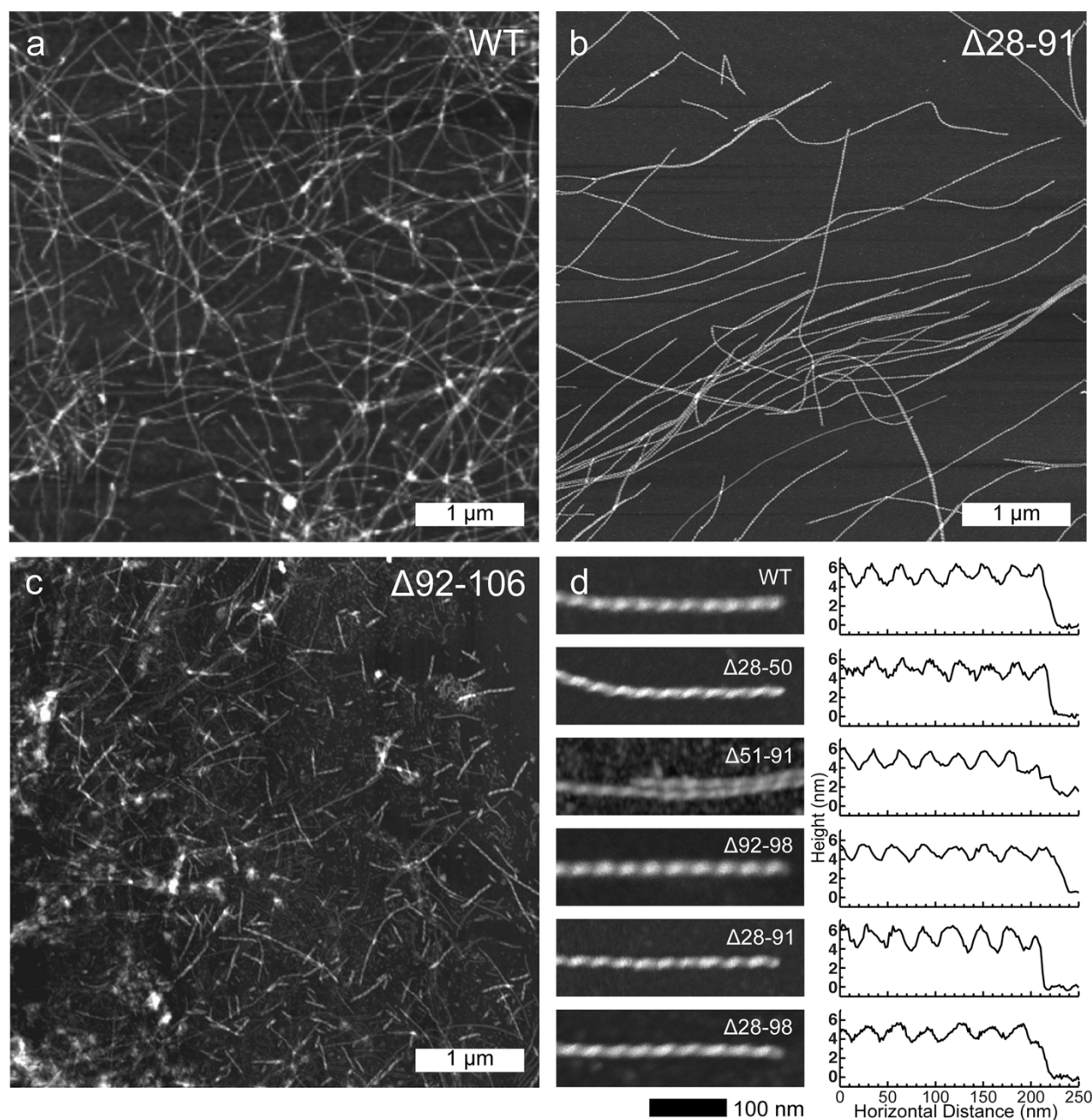


Figure 2. Representative low-resolution atomic force microscopy images for (a) WT, (b) $\Delta 28-91$ and (c) $\Delta 92-106$ huPrP23-144 fibrils (scale bar 1 μm for all panels). (d) High-resolution atomic force microscopy images (left; scale bar 100 nm for all panels) and corresponding height profiles (right) for WT, $\Delta 28-50$, $\Delta 51-91$, $\Delta 92-98$, $\Delta 28-91$, and $\Delta 28-98$ huPrP23-144 fibrils as indicated in the insets.

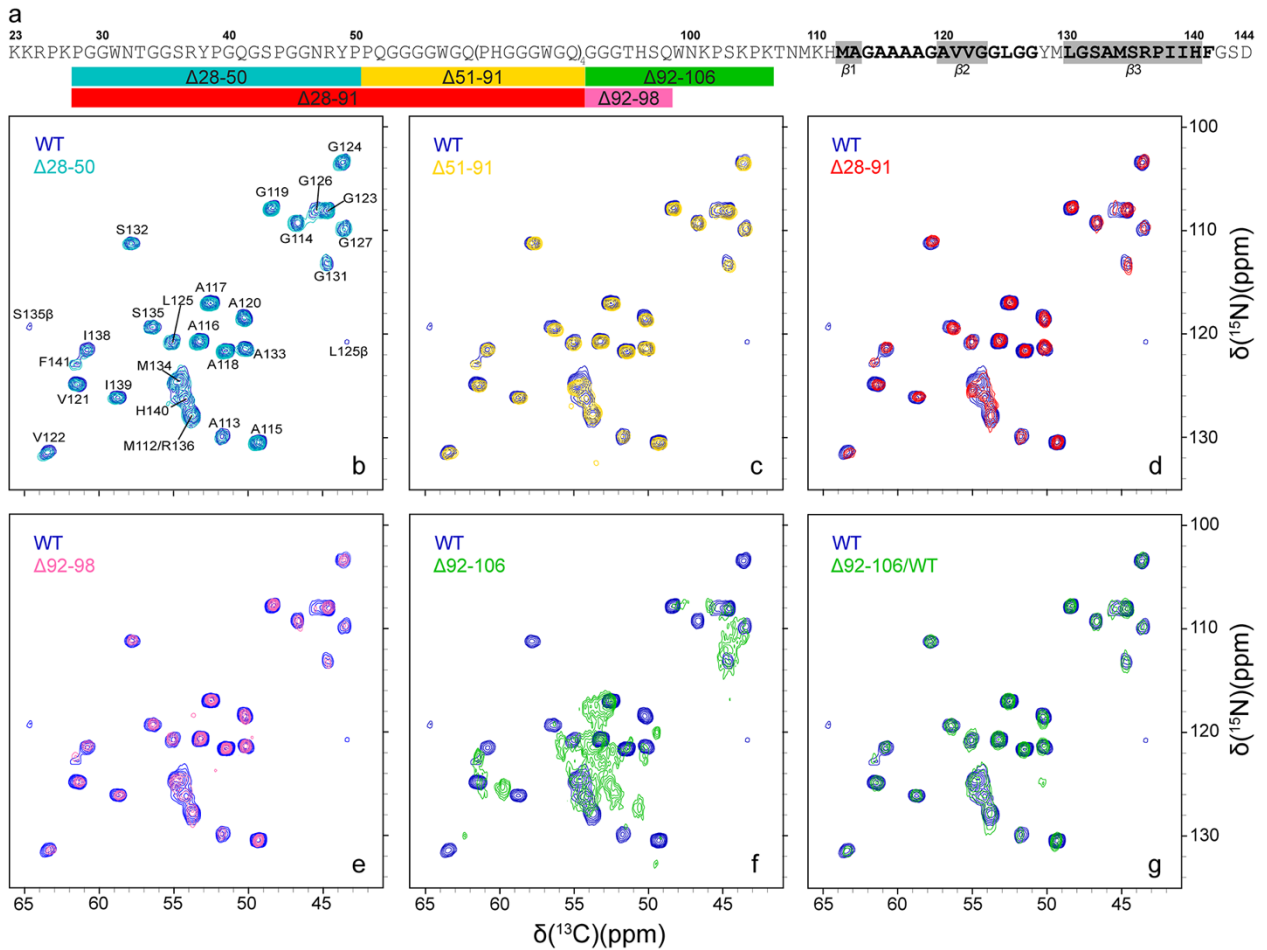


Figure 3. (a) Amino acid sequence of huPrP23-144. Relatively rigid residues comprising the amyloid core detected in conventional solid-state NMR experiments are shown in bold black font, with regions having highest β -strand propensity highlighted in grey rectangles (Theint, Nadaud, Surewicz, *et al.*, 2017), and conformationally flexible residues (Helmus *et al.*, 2008, 2010) are shown in grey font. The rectangles below the amino acid sequence schematically show the key huPrP23-144 large deletion variants that were investigated in this study, with the colors of the rectangles corresponding to the colors of the contours in the solid-state NMR spectra in panels (b)-(g). (b-g) 2D ^{15}N - $^{13}\text{C}\alpha$ chemical shift correlation solid-state NMR spectra of $\Delta 28-50$ (b, cyan), $\Delta 51-91$ (c, gold), $\Delta 28-91$ (d, red), $\Delta 92-98$ (e, magenta), $\Delta 92-106$ (f, green) huPrP23-144 fibrils and $\Delta 92-106$ fibrils seeded with WT huPrP23-144 amyloid (g, green), overlaid with the reference spectrum for WT huPrP23-144 fibrils (blue).

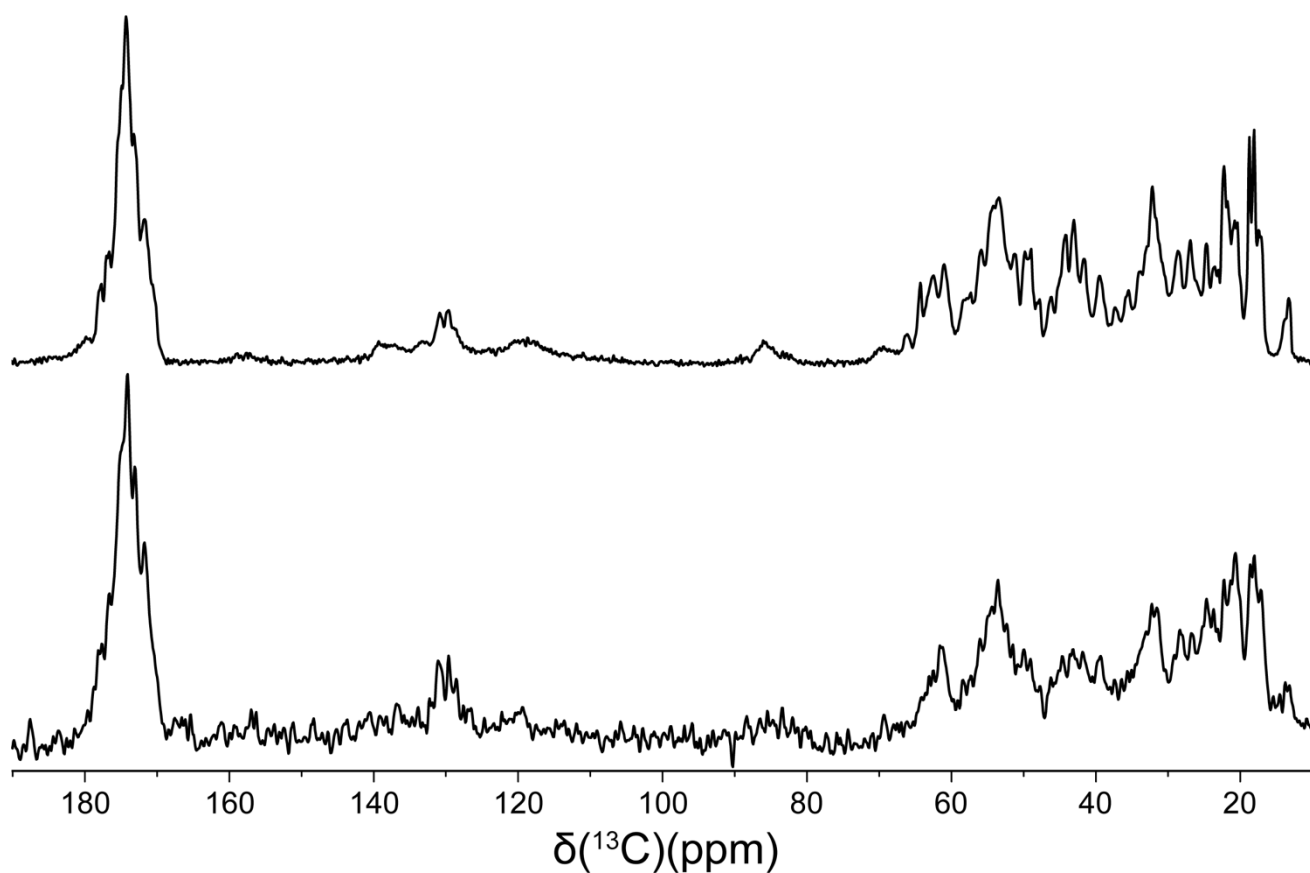


Figure 4. ^{13}C CP-MAS spectra of ^{13}C , ^{15}N WT (top) and natural abundance $\Delta 28-98$ (bottom) huPrP23-144 amyloid fibrils.