Title:

# Linking environmental salinity to respiratory phenotypes and metabolic rate in fishes: a data mining and modelling approach

Running title:

Salinity affects gill and blood phenotypes

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Summary statement:

Freshwater teleosts have lower gill oxygen conductances and higher haemoglobin-oxygen affinities than seawater species; both groups use different combinations of these traits to maintain similar metabolic rates and hypoxia tolerance.

**Abstract**

The gill is the primary site of ionoregulation and gas exchange in teleost fishes. However, those characteristics that benefit diffusive gas exchange (large, thin gills) may also enhance the passive equilibration of ions and water that threaten osmotic homeostasis. Our literature review revealed that gill surface area and thickness were similar in freshwater (FW) and seawater (SW) species; however, the diffusive oxygen ($O_2$) conductance ($G_d$) of the gill was lower in FW species. While a lower $G_d$ may reduce ion losses, it also limits $O_2$ uptake capacity and possibly aerobic performance in situations of high $O_2$ demand (e.g., exercise) or low $O_2$ availability (e.g., environmental hypoxia). We also found that FW fishes had significantly higher haemoglobin (Hb)-$O_2$ binding affinities than SW species, which will increase the $O_2$ diffusion gradient across the gills. Therefore, we hypothesized that the higher Hb-$O_2$ affinities of FW fishes compensate, in part, for their lower $G_d$. Using a combined literature review and modelling approach, our results show that a higher Hb-$O_2$ affinity in FW fishes increases the flux of $O_2$ across their low-$G_d$ gills. In addition, FW and SW teleosts can achieve similar maximal rates of $O_2$ consumption ($M\square O_{2max}$) and hypoxia tolerance ($P_{crit}$) through different combinations of Hb-$O_2$ affinity and $G_d$. Our combined data identified novel patterns in gill and Hb characteristics between FW and SW fishes and our modelling approach provides mechanistic insight into the relationship between aerobic performance and species distribution ranges, generating novel hypotheses at the intersection of cardiorespiratory and ionoregulatory fish physiology.

## Gill oxygen conductance and Hb-$O_2$ binding affinity are associated with salinity

Seawater (SW) and freshwater (FW) environments present different osmoregulatory challenges to resident species. Teleosts in both environments have similar internal ion concentrations (Holmes and Donaldson, 1969) and therefore, are hypo-osmotic compared to ion-rich SW where they risk losing water to the environment, and hyper-osmotic compared to ion-poor FW where they risk losing ions (Evans et al., 2005). The fish gill is the primary site of gas exchange and presents a large, thin epithelium to the surrounding water that facilitates the uptake of oxygen ($O_2$) and the excretion of the metabolic by-products carbon dioxide and ammonia. However, those gill morphometrics that maximize diffusive gas exchange may also increase passive ion and water fluxes that, if unopposed, would lead to osmotic disturbances. These conflicting requirements for ionoregulation and $O_2$ uptake at the fish gill were recognized over 100 years ago by August Krogh (Ege and Krogh, 1914; Krogh, 1937) and later substantiated in seminal work by Randall et al. (1972), Gonzalez and McDonald (1992) and others. Today, the effect is known as the osmorespiratory compromise, and it has been treated in detailed reviews (Gilmour and Perry, 2018; Gonzalez, 2011; Nilsson et al., 2012; Perry, 1998; Sardella and Brauner, 2007; Wood and Eom, 2021). The present study builds on previous work to uncover general patterns among teleost fishes that link the environmental salinity to gill and blood phenotypes and to provide mechanistic insight into the physiology underlying these observations by using a modelling approach.

FW and SW teleosts have evolved divergent strategies to overcome the osmotic challenges imposed by their respective environments, involving anatomical and physiological adaptations at different organ systems. Briefly, FW fishes take up ions at the gills and excrete dilute urine via the kidneys, whereas SW fishes take up water and ions via their digestive tracts and excrete ions via the gills, kidneys and with their faeces (Evans et al., 2005; Grosell, 2010; Marshall and Grosell, 2006). These divergent mechanisms enable FW and SW fishes to maintain water and ion homeostasis in their respective environments and are well-studied, but their effects on gill morphometrics and, thus, $O_2$ uptake capacity, have not been studied broadly. To address this knowledge gap, we conducted a systematic literature review of gill, blood and metabolic characteristics in 355 teleost species in relation to their aquatic habitat (155 FW, 200 SW species; see Appendix for details on literature review and the Supplement for raw data).

Our initial analysis indicated that FW fishes, on average, had significantly smaller mass-specific gill surface areas (GSA) than SW fishes ($P = 0.019$; Fig. 1A), consistent with what has long been reported in the literature. However, we also observed differences in body mass between the investigated FW and SW fishes, which biases this analysis because GSA scales allometrically with body mass. Furthermore, these interspecific comparisons are dominated by a few hyper-diverse taxa (Ostariophysi in FW and Percomorpha in SW; Vega and Wiens, 2012), which may introduce an additional phylogenetic bias. Therefore, to test if salinity affected GSA while taking into account allometric scaling and phylogenetic non-independence of species, we fitted a phylogenetically-corrected linear model to the $\log_{10}$-transformed GSA and body mass data (Fig. 1B), and then calculated the vertical residuals that provided a body mass-independent measure of GSA (Garland et al., 1992). These residuals were independent of salinity ($P = 0.672$; Fig. 1C), indicating that, once corrected for mass and phylogenetic relatedness, teleosts in FW and SW had similar GSAs. These findings illustrate the importance of accounting for body mass and species relatedness in interspecific comparisons of traits that scale allometrically with body mass. In contrast, the blood-water diffusion distance (hereafter, gill thickness; Fig. 1 D) was unaffected by body mass ($P = 0.789$; Fig. E) and, thus, these data were only corrected for the phylogenetic relatedness of the species. As with GSA, we found no significant effect of salinity on gill thickness ($P = 0.140$; Fig. F), indicating that other factors must determine the observed variability.

To explore the interacting effects of GSA and thickness, we calculated the $O_2$ diffusive conductance of the gills ($G_d$) in FW and SW species, which describes the amount of $O_2$ that can diffuse across the gills for a given $PO_2$ gradient. In our study, $G_d$ was calculated from GSA and gill thickness values from the literature, and from the gill characteristics described in Table S1 (Dejours, 1981). We found 33 species with reported values for both GSA and thickness (Fig. 1G) and, after correcting for body mass and phylogenetic relatedness (Fig. 1H), FW fishes had significantly lower $G_d$ values than SW fishes ($P = 0.017$; Fig. 1I). This result should be interpreted in light of the relatively few species with thickness measurements and the fact that some species with extreme gill characteristics may disproportionately influence the analysis (such as the highly active Scombridae in SW and some facultative air-breathing fishes in FW). We tried expanding our analysis to all actinopterygian fishes to obtain additional values for $G_d$ but found that gill thickness values were also largely unavailable. Additional measurements of

gill thickness in other FW and SW fishes are needed to strengthen the relationships between Gd and environmental salinity that we found in our analysis. Nevertheless, the data that are currently available indicate that, even after mass and phylogenetic corrections, FW teleosts have lower $G_d$ than SW teleosts, and if substantiated more broadly this finding may change our understanding of the physiological implications of environmental salinity on gill phenotypes.

Smaller and thicker gills can benefit fishes by reducing the rate of passive ion and water fluxes that can disrupt osmotic homeostasis (Greco et al., 1996; Henriksson et al., 2008; Perry, 1998; Sollid et al., 2003). However, the gill is also the primary site for gas exchange and smaller and thicker gills will result in lower $G_d$. In FW fishes, lower $G_d$ may limit $O_2$-uptake capacity and could impair their performance in situations of high $O_2$ demand (e.g., exercise) or low $O_2$ availability (e.g., environmental hypoxia). Interestingly, our data revealed a potential compensatory mechanism, as FW species had ~2-fold higher haemoglobin (Hb) $O_2$ affinities than SW species (expressed as lower $P_{50}$ values of Hb; the $PO_2$ at which Hb is 50% saturated with $O_2$; $P < 0.001$; Fig. 1L). Hb $P_{50}$ values were corrected for phylogenetic relatedness and a significant temperature effect using a Van't Hoff Plot ($P < 0.001$; Fig. 1K). Because Hb is the principal $O_2$ carrier in the blood and its $O_2$-binding characteristics determine the kinetics of $O_2$ loading at the gills and unloading at the tissues, the higher Hb-$O_2$ affinities of FW species could enhance the diffusion gradient of $O_2$ across the fish gill and theoretically support higher rates of metabolic $O_2$ consumption ($M\square O_2$; a proxy for metabolic rate). These observations led us to hypothesize that the high Hb-$O_2$ affinities of FW species compensate for their lower $G_d$, allowing FW species to maintain similar $M\square O_2$ as SW species during aerobic challenges, such as exercise and hypoxia.

To test this hypothesis, we explored the interacting effects of $G_d$ and Hb $P_{50}$ using a mathematical model (based on Malte and Weber, 1985) that predicts the maximal $M\square O_2$ ($M\square O_{2max}$) that a fish can support over a range of gill and blood characteristics, as well as environmental conditions (see Appendix for detailed model description). Specifically, we tested three predictions derived from our hypothesis: all else being equal, 1) higher Hb-$O_2$ affinities will compensate for lower $G_d$ to maintain arterial $O_2$ transport; 2) FW and SW teleosts will achieve similar $M\square O_{2max}$ using different combinations of $G_d$ and Hb-$O_2$ affinity; and 3) FW and SW teleosts will achieve similar hypoxia tolerances (represented by $P_{crit}$; the lowest water $PO_2$ at which a fish can maintain its standard $M\square O_2$; $M\square O_{2Std}$) using different combinations of $G_d$ and

143 Hb-$O_2$ affinity. We then compared the modelled results with empirical values mined from the
144 literature that revealed novel patterns in the divergent gill and Hb phenotypes of FW and SW
145 teleosts in relation to environmental salinity.

**Prediction 1: Higher Hb-$O_2$ affinities will compensate for lower $G_d$ to maintain arterial $O_2$ transport**

First, we used our $M\dot{O}_{2max}$ model to investigate the effects of gill and blood characteristics on the equilibration of $PO_2$ across the gill epithelium using the lowest, median and highest $G_d$ values for FW and SW fishes that we observed in the literature(using only values that were based on $N > 1$), over the range of Hb-$O_2$ affinities reported in Fig. 1. FW fishes with the lowest $G_d$ were severely diffusion-limited, reflected in a poor equilibration of $PO_2$ between the counter-current flows of water and blood (Fig. 2A). $M\dot{O}_{2max}$ in these fishes was generally low, but the highest values were achieved at the lowest Hb $P_{50}$. Therefore, when all else is equal, a higher Hb-$O_2$ affinity can partially compensate for a low $G_d$ by increasing the $PO_2$ gradient across the epithelium. This finding is consistent with previous work that modelled the diffusion of $O_2$ across the fish gill and found an increased $O_2$ extraction from the water and lower ventilatory requirements at high Hb-$O_2$ affinities (Malte and Weber, 1987). In addition, a reduction in Hb-$O_2$ affinity caused a steep decline in the arterial $O_2$ saturation of Hb ($S_aO_2$), which determines the maximum capacity for blood $O_2$ transport and $M\dot{O}_{2max}$ (Gallaugher et al., 2001). In SW species, the lowest, highestand median values for $G_d$ were higher than in FW fishes. Consequently, $O_2$ uptake at the gills in SW was generally more efficient, as indicated by a nearly complete equilibration of $PO_2$ across the gill epithelium, especially at the lowest Hb $P_{50}$ values (Fig. 2D).

An increase in $G_d$ to median values increased $M\dot{O}_{2max}$ in both FW and SW fishes and shifted the optimal Hb $P_{50}$ to higher values (Fig. 2B and E). Higher $G_d$ also reduced the effect of Hb $P_{50}$ on $S_aO_2$ because $O_2$ uptake became less dependent on the $PO_2$ diffusion gradient. The fact that a high Hb-$O_2$ affinity exerts the greatest benefits in fish with the lowest $G_d$ is consistent with our literature-mined data. Combined, these findings indicate that the higher Hb-$O_2$ affinities observed in FW fishes may have adaptive significance to compensate for their low-$G_d$ gills, an idea that could be substantiated by investigating $G_d$ in additional species.

SW fishes with median $G_d$ achieved full $O_2$ saturation during blood transit through the gill, suggesting that $M\dot{O}_{2max}$ is limited by maximal cardiac output ($Q\dot{}_{max}$), rather than the diffusion characteristics of the gill (Fig. 2 D), and the same was true for those FW and SW species with the highest $G_d$ (Fig. 2 C and F). These findings generally agree with experimental work, indicating that $O_2$-uptake at the gill is not diffusion-limited in normoxia, but rather a

function of maximal perfusion and/or ventilation rates (Daxboeck et al., 1982; Malte and Weber, 1985; Randall and Daxboeck, 1984). However, in >50% of the FW species in our mined data set, $\dot{M}O_{2max}$ may be limited by gill diffusion characteristics, where increases in $\dot{Q}$ are inconsequential. These results must be interpreted with respect to the $\dot{Q}_{max}$ value for rainbow trout that we used in our simulations, which may be higher than those of many low-$G_d$ FW species. Nevertheless, our results highlight systematic differences in cardiorespiratory phenotypes between FW and SW teleosts that have ecological consequences. Those species with the highest $G_d$ may achieve increases in $\dot{M}O_{2max}$ through increases in $\dot{Q}$, enabling more active lifestyles that are not available to low-$G_d$ species. A broader assessment of $G_d$, $\dot{M}O_{2max}$ and $\dot{Q}$ in relation to environmental salinity in teleosts may shed light on the mechanistic basis for our observations and their ecological significance.

Finally, our simulations also showed that the Hb $P_{50}$ values that achieved the highest $S_aO_2$ did not necessarily produce the highest $\dot{M}O_{2max}$. Lower Hb $P_{50}$ values can safeguard $S_aO_2$ and improve $\dot{M}O_{2max}$ in those fish with the lowest $G_d$. However, the higher $G_d$ we observed in SW species would lessen the benefits of a lower Hb $P_{50}$, potentially enabling them to exploit a higher range of Hb $P_{50}$ values that promote the offloading of $O_2$ at the tissues, which we explored in more detail.

**Prediction 2: FW and SW teleosts will achieve similar $M\dot{O}_{2max}$ using different combinations of $G_d$ and Hb-$O_2$ affinity**

The optimal Hb $P_{50}$ is a compromise between the physiological requirements for $O_2$ loading at the gas exchange surface and unloading at the tissue capillaries (Brauner and Wang, 1997; Harter and Brauner, 2017; Wang and Malte, 2011). To highlight this trade-off in more detail, Figure 3 shows the $M\dot{O}_{2max}$ that can theoretically be attained over the range of Hb $P_{50}$ and $G_d$ values in FW and SW teleosts. Generally, those species with the lowest $G_d$ achieved the lowest $M\dot{O}_{2max}$ and increasing $G_d$ improved $M\dot{O}_{2max}$, consistent with a diffusion limitation on $O_2$ uptake. However, $M\dot{O}_{2max}$ in species with median $G_d$ already matched those of species with the highest $G_d$, indicating a progressive perfusion limitation, which is in line with the results shown in Figure 2. Since the upper boundary for $M\dot{O}_{2max}$ was set by perfusion, both groups achieved similar $M\dot{O}_{2max}$ of ~13 μmol $g^{-1}$ $h^{-1}$ at the $Q\dot{}_{max}$ for rainbow trout (53 ml $kg^{-1}$ $min^{-1}$).

Increasing Hb-$O_2$ affinity also had beneficial effects on $M\dot{O}_{2max}$, but only until an optimum value was reached, after which further increases in Hb-$O_2$ affinity severely decreased $M\dot{O}_{2max}$. The effect of Hb-$O_2$ affinity on $M\dot{O}_{2max}$ depended on $G_d$, where a reduced $G_d$ shifted the optimal $P_{50}$ for $M\dot{O}_{2max}$ to lower values. For example, when $G_d$ was reduced from median to lowest values, the optimal $P_{50}$ shifted from 31 to 3 mmHg in FW fishes (Fig. 3A) and from 39 to 25 mmHg in SW fishes (Fig. 3B). These ranges of optimal $P_{50}$ map well onto the mined $P_{50}$ values for FW and SW fishes, respectively (Fig. 1).

Our simulations are also in line with data on rainbow trout acclimated to soft water that thickened their gills from 3 to 6 μm due to ionocyte proliferation and compensated for the impaired $G_d$ by decreasing $P_{50}$ from 18 to 12 mmHg (Greco et al., 1996; Perry, 1998; Perry et al., 1996). Gill morphology and $P_{50}$ are plastic traits, and it seems that they may respond in concert to overcome physiological challenges to $O_2$ uptake in a way that depends on environmental salinity. The situation is complicated as the diffusive pathways for $O_2$, water and ions may differ and many species exert some control over these passive fluxes, such as observed in extremely hypoxia-tolerant species (Wood et al., 2009), active species (Gonzalez and Mcdonald, 1994), and after exercise training (Gallaugher et al., 2001; Postlethwaite and McDonald, 1995). A salinity-specific effect on gill permeability has not been studied broadly, but data for a few species exist. FW-acclimated killifish have higher branchial water permeabilities than SW conspecifics, but both display similar diffusive ion fluxes (in opposite directions) during hypoxic hyperventilation

(Giacomin et al., 2019; Wood et al., 2019). However, while FW-acclimated killifish do not alter ionocyte density, they do decrease their overall GSA (Giacomin et al., 2019), which is consistent with our observation of lower $G_d$ in FW fishes that may perhaps occur via different mechanisms in different species and conditions. If this effect is substantiated more broadly, one may hypothesize that the hyper-osmoregulatory strategy of FW fishes has not only led to lower $G_d$, but also to higher Hb-$O_2$ affinities that balance the requirements for $O_2$ uptake and tissue $O_2$ extraction at the prevailing gill diffusion characteristics. Likewise, the hypo-osmoregulatory strategy of SW fishes may be permissive of high-$G_d$ gills that are best matched by higher $P_{50}$ and that enable increases in $\dot{Q}$ and $\dot{M}O_{2max}$ in species with active lifestyles where exercise performance is linked to evolutionary fitness.

To further explore the interacting effects of $G_d$ and Hb-$O_2$ affinity on $\dot{M}O_{2max}$, we plotted the upper and lower boundaries for $G_d$ as a function of Hb $P_{50}$, resulting in areas of theoretical $\dot{M}O_{2max}$ that can be achieved by the gill and blood characteristics of FW and SW fishes (Figure 4A). The lower areas were calculated at the $\dot{Q}_{max}$ for rainbow trout, and due to the perfusion limitation of $\dot{M}O_{2max}$, FW and SW fishes with the highest $G_d$ achieved similar $\dot{M}O_{2max}$. To corroborate these modelling results, we then plotted empirically measured $\dot{M}O_{2max}$ values onto the theoretical areas. Generally, the measured and calculated values agreed well. However, one cluster of FW fishes with high Hb-$O_2$ affinities fell above the predicted lines, which is likely explained by higher tissue $O_2$ conductances that maintain tissue $O_2$ extraction from high-affinity Hbs, and perhaps by lower Hill coefficients that enhance $O_2$ extraction from the water (Malte and Weber, 1987); both these values were kept constant in our model. Other FW and SW species may achieve $\dot{Q}_{max}$ values that exceed those of rainbow trout. For instance, sockeye salmon swimming in FW reached $\dot{M}O_{2max}$ of 27.2 μmol $g^{-1}$ $h^{-1}$ at a $\dot{Q}_{max}$ of 67.8 ml $kg^{-1}$ $min^{-1}$ (Steinhausen et al., 2008) and in some sockeye salmon populations with longer migration routes, $\dot{Q}_{max}$ can exceed 100 ml $kg^{-1}$ $min^{-1}$, nearly 2-fold the value of rainbow trout (Eliason et al., 2013). Whether the gill characteristics of these anadromous sockeye salmon are truly representative of a FW species is unclear, but despite their semelparous life history, they appear to exert tight control over ion homeostasis, even during their final FW stage.

We then explored whether a salinity-specific pattern in $\dot{M}O_{2max}$ exists across empirically measured values for 130 fish species (51 FW, 79 SW), and we corrected these data for allometric scaling with body mass, temperature and phylogenetic relatedness (Fig. 4B).

Across all species, there was no significant effect of salinity on $\dot{M}O_{2max}$ ($P < 0.067$; Fig. 4C), consistent with our prediction that FW and SW teleosts may achieve similar $\dot{M}O_{2max}$ through different combinations of gill and blood characteristics. However, the low $P$-value in our analysis on 130 species suggests that future studies, especially those with a larger sample sizes, may be able to resolve potential differences in $\dot{M}O_{2max}$ between FW and SW teleosts. Finally, $\dot{M}O_{2max}$ is a complex physiological trait that is strongly influenced by methodology, animal condition and behaviour (Killen et al., 2017; Norin and Clark, 2016); exploring these sources of variability and accounting for them by statistical means may be a worthwhile avenue for future analyses.

Some of the highest $\dot{M}O_{2max}$ values are found in the fast-swimming, pelagic, SW teleosts (Wegner et al., 2010), including billfishes (Istiophoridae and Xiphiidae), dolphinfishes (Coryphaenidae), jacks (Carangidae) and tunas (Scombridae). Reliable data during maximal exercise in these animals are notoriously difficult to obtain, but a few measurements are available. Based on submaximal swimming trials, the $\dot{M}O_{2max}$ in skipjack tuna has been estimated at 68.7 μmol $g^{-1}$ $h^{-1}$ (Brill, 1987; Dewar and Graham, 1994; Gooding et al., 1981) and therefore exceeds the value for rainbow trout by more than 5-fold. These high $\dot{M}O_{2max}$ are enabled by GSAs of nearly 20 cm $g^{-1}$ in yellowfin and skipjack tunas (Brill and Bushnell, 2001), and gill thicknesses as low as 0.5 μm (Hughes, 1970; Hughes, 1984), resulting in $G_d$ values that are three-fold higher than the highest values of any FW species. Clearly, a unique physiology in tunas is required to sustain such high $\dot{M}O_{2max}$ that cannot be achieved with the cardio-respiratory characteristics of rainbow trout or most other teleosts. Therefore, to determine what physiological adaptations are required in tuna to attain their high $\dot{M}O_{2max}$, we re-calibrated our model to the skipjack tuna (Fig. 4; light-shaded areas). This adjustment revealed that the high $\dot{M}O_{2max}$ in tuna are feasible only if $\dot{Q}_{max}$ is increased ~6-fold (to 318 ml $kg^{-1}$ $min^{-1}$), blood [Hb] is increased 2.3-fold, and $O_2$ conductance at the tissues is increased ~3-fold over the values in rainbow trout. Whether these values are representative of some tuna species remains to be validated experimentally; however, the available data indicate that they may not be unreasonable estimates. Even in spinally blocked skipjack tuna, $\dot{Q}_{max}$ has been measured at 132 ml $kg^{-1}$ $min^{-1}$ (Brill, 1987; Bushnell, 1988), blood [Hb] can reach 2.3 mM (Brill and Bushnell, 1991), and tissue $O_2$ conductance is undoubtedly increased by extremely high capillary densities (Hulbert et al., 1979) and myoglobin concentrations (George and Stevens, 1978; Stevens and Carey, 1981).

Theoretically, high $Q_{max}$, [Hb] and tissue $O_2$ conductances may also be attainable by FW teleosts; but matching high-$G_d$ gills may not. Our simulations show that even the highest $G_d$ values in FW fishes are insufficient to produce tuna-like $MO_{2max}$ and the upper boundary levels off at ~40 μmol $g^{-1}$ $h^{-1}$. These values are still higher than any $MO_{2max}$ recorded for a FW teleost, indicating that factors other than $G_d$ may set the actual limit. Regardless, these fundamental relationships may explain why no FW fishes match the high $MO_{2max}$ found in some SW species. Whether FW tuna-like fishes are, in fact, absent from the fossil record is a worthwhile avenue for future investigations. However, based on our findings it would seem that FW environments should remain devoid of fast-swimming pelagic analogues to the tunas.

**Prediction 3: FW and SW teleosts will achieve similar $P_{crit}$ using different combinations of GSA and Hb-$O_2$ affinity.**

Another important driver for variation in gill characteristics and Hb-$O_2$ affinity among fishes is the availability of environmental $O_2$ (Mandic et al., 2009). Under conditions of low environmental $O_2$ (hypoxia), a fish's indefinite survival depends on its ability to sustain $M\square O_{2std}$ (Hughes, 1973). The lowest water $PO_2$ at which a fish can sustain $M\square O_{2std}$ is termed the critical $O_2$ tension ($P_{crit}$), and at $PO_2$ below $P_{crit}$, a fish's survival becomes dependent on some combination of anaerobic glycolysis and metabolic depression, both of which are unsustainable in the long term (Beamish, 1964; Regan et al., 2016; Ultsch and Regan, 2019; Ultsch et al., 1978). $P_{crit}$ is therefore a common metric of hypoxia tolerance that represents the suite of aerobic contributions to hypoxia survival in a single value (Regan et al., 2019), and thus, we explored this metric in our simulations.

To estimate how differences in $G_d$ and Hb-$O_2$ affinity may influence $P_{crit}$, we adjusted our model to predict $M\square O_{2max}$ at progressively decreasing inspired water $PO_2$ (from 150 to 0 mmHg) and then fixed a horizontal line representing $MO_{2std}$ at 1.5 μmol $g^{-1}$ $h^{-1}$, which is the value reported for resting rainbow trout (Kiceniuk and Jones, 1977). The $PO_2$ at which the $M\square O_{2max}$ and $M\square O_{2std}$ lines intersect represented the theoretical $P_{crit}$ for a given set of gill and blood characteristics. Our approach assumed that $M\square O_{2max}$ and $M\square O_{2std}$ respond similarly to reductions in $PO_2$ below $P_{crit}$, and that the typical $P_{crit}$ curve is biphasic. The former assumption is consistent with previous theoretical work (Esbaugh et al., 2021), empirical measurements (Claireaux et al., 2000), and the fact that aerobic scope for activity at sub-$P_{crit}$ $PO_2$ is zero (Claireaux and Chabot, 2016; Ern et al., 2016). The latter assumption is true for some fish species, but not all (Wood, 2018; Farrell et al., 2021). However, most important for our simulations was not the shape of the $M\square O_{2std}$ curve at $PO_2$ below $P_{crit}$, but rather the presence of a benchmark $M\square O_{2std}$ value to which the sub-$P_{crit}$ $M\square O_{2max}$ line could be compared. A biphasic $P_{crit}$ curve enabled this.

The outcomes of these simulations are shown in Figure 5 for the lowest, median and highest $G_d$ that we observed in FW and SW fishes (Fig. 1). In all cases, increasing Hb-$O_2$ affinity or $G_d$ led to lower $P_{crit}$ values, representing a higher hypoxia tolerance of the fish. SW fishes achieved lower $P_{crit}$ than FW fishes when $G_d$ was at lowest or median values. However, the lowest overall $P_{crit}$ values were achieved by FW fishes with high $G_d$ and high Hb-$O_2$ affinities.

These and our previous results show that an increased Hb-$O_2$ affinity can generally compensate for impaired diffusion characteristics at the gill caused by either low $G_d$ or low environmental $O_2$. In both cases, $M\square O_2$ is limited by the reduction in $S_aO_2$ that can be improved by increasing Hb-$O_2$ affinity, and these results are in close agreement with previous work (Wang and Malte, 2011).

To validate our simulations, we plotted the $P_{crit}$ values for high and low $G_d$ as a function of Hb-$O_2$ affinity, spanning the area of theoretical $P_{crit}$ that were predicted by the model. We then overlayed empirically measured $P_{crit}$ values from the literature for FW and SW fishes (Fig. 6A). For FW fishes, we found an excellent agreement between the model predictions and measured $P_{crit}$ values. In addition, $P_{crit}$ values in FW fishes were highly variable, spanning an order of magnitude. GSA and gill thickness are plastic traits that respond to hypoxia, environmental ion concentration and temperature. The underlying mechanisms include dynamic changes in the redistribution of branchial blood flow and cellular responses that reversibly alter the morphology of the gill (Wood and Eom, 2021). For example, the crucian carp can increase GSA by ~7.5-fold after an acclimation period to hypoxia, largely through the receding of an interlamellar cell mass (Sollid et al., 2003). These acclimation responses can occur quickly, as in goldfish, where GSA increases ~2-fold in just hours (Regan et al., 2017). High GSA plasticity is often found in hypoxia-tolerant rather than -intolerant species (Dhillon et al., 2013), and in more FW than SW species (Gilmour and Perry, 2018). This may be because many FW environments are particularly hypoxia-prone (Diaz and Breitburg, 2009) or simply because relatively few SW species have been studied in this respect.

In contrast, SW fishes occupied a narrow band in $P_{crit}$ values over a broad range of Hb $P_{50}$, which is supported by the modelled and empirical data (Fig. 6A). However, the modelled data were generally offset to lower $P_{crit}$ values. The reasons for this discrepancy may relate to our model assumptions not accurately representing some of the *in vivo* characteristics of SW fishes, e.g.: i) many SW species may have higher $M\square O_{2std}$ than rainbow trout, and/or ii) the boundaries for $G_d$ in our dataset may not match those of SW species for which $P_{crit}$ and Hb $P_{50}$ data are available, both representing only a small subset of SW species. Regardless, the combined data revealed that FW and SW fishes occupy different quadrants in the $P_{crit}$ vs. Hb $P_{50}$ relationship, which indicates that they achieve their hypoxia tolerance by different mechanisms. SW fishes rely on their higher $G_d$ to maintain branchial $O_2$ uptake in hypoxia (Fig. 5), resulting in $P_{crit}$

values that are largely independent of Hb $P_{50}$ and thus fall within a narrow range (Fig. 6A). On the other hand, FW fishes rely on their high Hb-$O_2$ affinities to maintain branchial $O_2$ uptake in hypoxia and, when combined with high $G_d$ values, achieve the lowest $P_{crit}$ observed in teleosts.

To test whether there is a systematic difference in $P_{crit}$ between FW and SW teleosts, we mined literature values for 142 species (46 FW and 96 SW). After correcting for phylogenetic relatedness of species (Fig. 6B), we found no significant difference in $P_{crit}$ between FW and SW teleosts ($P = 0.891$; Fig. 6B). This stands in contrast with our data set when not corrected for phylogeny, which showed significantly lower $P_{crit}$ in FW than SW fish, and to the metanalysis of Rogers et al. (2016) and intraspecific salinity trials of Haney and Nordlie (1997), which reported lower $P_{crit}$ in FW fishes under some, but not all, conditions. Therefore, the variation in $P_{crit}$ across salinity environments appears to be driven by a few teleost clades with extreme $P_{crit}$ values, again highlighting the importance of applying appropriate corrections of the raw data. Relatedly, most SW species on which $P_{crit}$ experiments have been performed are native to environments that regularly experience hypoxia, such as coral reefs and intertidal zones. The mean SW $P_{crit}$ value may therefore be skewed towards these hypoxia-adapted SW species and away from pelagic species that are less likely to regularly encounter hypoxia. Fewer $P_{crit}$ values for pelagic species exist, perhaps because these fishes are not easily caught and experimented on. However, their addition to this comparison could further distinguish the salinity groups and is a worthwhile direction for future studies. Finally, there is substantial variability in $P_{crit}$ values in the literature and, as with other complex performance metrics, $P_{crit}$ should be interpreted with some caution regarding differences in methodology, animal condition and behaviour (Wood, 2018). Part of this variability may be due to the plasticity in gill morphology and blood characteristics in teleosts, perhaps coupled with different methodologies regarding the rates of hypoxia onset (Rogers et al., 2016). Nevertheless, the close agreement of the empirical and modelling data indicates that the general patterns in $P_{crit}$ between FW and SW fishes are largely driven by factors that are manipulated in the model. Therefore, studying the interspecific variability in $G_d$, Hb-$O_2$ affinity and their interactions may provide a roadmap for future investigations into the mechanistic basis for $P_{crit}$ in fish.

## Conclusion and Perspectives

Our analysis provides compelling evidence for systematic differences in gill and blood phenotypes between FW and SW teleosts that may be related to the osmotic characteristics of their aquatic environments. However, aerobic performance, assessed as $\dot{M}O_{2max}$ and $P_{crit}$, did not differ between FW and SW fishes; therefore, the two groups may use different combinations of gill and blood characteristics to support similar aerobic capacities. FW fishes generally had lower $G_d$, which has consequences for aerobic metabolism and may reflect a physiological constraint by their hyper-osmoregulatory strategy. The higher Hb-$O_2$ affinity of FW fishes may thus have adaptive significance, as it increases the diffusion gradient for $O_2$ across the gills. The benefits of a higher Hb-$O_2$ affinity may not fully compensate for the disadvantages of a low $G_d$, but rather, will enable a higher $\dot{M}O_{2max}$ within this constraint, by balancing the conflicting requirements for $O_2$ loading at the gills and unloading at the tissues. In contrast, the hypo-osmoregulatory strategy of SW teleosts is clearly permissive of higher $G_d$, which in turn may have lifted the brakes on the evolution of higher $\dot{Q}_{max}$ and [Hb]. In tunas, these coordinated adaptations of their cardio-respiratory systems ultimately enabled extremely high $\dot{M}O_{2max}$ that are not attainable even by FW fishes with the highest $G_d$.

Finally, gill morphology and Hb-$O_2$ binding characteristics are highly plastic traits in teleosts, and many species with euryhaline, anadromous or catadromous life cycles routinely transition between the FW and SW environments and similar transitions occurred repeatedly in the course of teleost evolution (Betancur-R et al., 2015). The evolutionary dynamics that govern these transitions are worthy of further investigation and fish clades that include species that have recently transitioned between FW and SW may be well-suited for this purpose. The osmorespiratory compromise describes the conflicting requirements for $O_2$ uptake and ion/water fluxes at the gills and has typically been studied with respect to gill surface area. Our data indicated that gill thickness and $G_d$ are important factors that determine $\dot{M}O_{2max}$ in fishes and should be considered in future studies on the osmorespiratory compromise by measuring $O_2$ uptake and unidirectional ion/water fluxes in more species in both FW and SW. Ideally, future studies should use standardized protocols for determining gill morphometrics, unidirectional ion/water fluxes, $\dot{M}O_{2max}$, $P_{crit}$, and Hb characteristics in animals acclimated to a clearly defined set of environmental conditions, allowing for more reproducible interspecies comparisons. These future studies may lay the foundation to test our hypothesis that invasions of

417 the FW environment and the divergent selective pressures of the osmorespiratory compromise
418 gave rise to adaptations that reduced $G_d$ and Hb $P_{50}$, thus profoundly shaping the aerobic
419 pathway of FW fishes.

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**Competing Interests:**

The authors declare no competing interests.

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## Data availability

All R code is publicly available on GitHub ($M\dot{O}_{2max}$ model and data analyses: github.com/tillharter/Fish_MO2max_models_in_R) and the raw data from our literature review is available in the supplement.

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## Figure Legends

**Figure 1: Systematic differences in the gill and blood characteristics of freshwater (FW, green) and seawater (SW, blue) teleost fishes.** A) The effect of salinity on mass-specific gill surface area GSA ($cm^2$ $g^{-1}$). B) There was a significant effect of body mass on GSA, as determined by a generalized linear mixed model that accounted for phylogenetic non-independence of species. Regression line: $\log_{10}$[gill surface area ($mm^2$)] = 0.63 + 0.93 $\log_{10}$[body mass (g)]. C) The residuals of this model provide a mass-independent measure of GSA and the effect of salinity was tested by a phylogenetic ANOVA simulation that revealed no differences between FW and SW species D) The blood-water diffusion distance (gill thickness, μm), E) was independent of body mass and F) the phylogenetically corrected residuals were not significantly different between FW and SW species. G) Mass-specific gill oxygen ($O_2$) conductance ($G_d$, μmol $mmHg^{-1}$ $g^{-1}$ $h^{-1}$) was calculated from GSA and thickness (and the parameters reported in Table S1). H) $G_d$ was dependent on mass (regression line: $\log_{10}$[$G_d$ (μmol $mmHg^{-1}$ $h^{-1}$)] = 0.86 + 0.92 $\log_{10}$[body mass (g)]), and I) the residuals of $\log_{10}G_d$ were lower in FW than SW species. J) Haemoglobin (Hb) $O_2$ affinity (expressed as $P_{50}$ of Hb; the $PO_2$ at which Hb is 50% saturated with $O_2$), K) was dependent on temperature (regression line: $\log_{10}$[$P_{50}$ (mmHg)] = 6.332 – 1534 temp $(K)^{-1}$) and L) the phylogenetically- and temperature-corrected residuals of log $P_{50}$ were significantly lower in FW than SW fishes. All values were mined from the literature according to the search protocols described in the Appendix. Points are values for individual fish species.

**Figure 2 Simulations modelling the outcome of counter-current oxygen ($O_2$) uptake at the fish gill.** The partial pressures of $O_2$ ($PO_2$, mmHg) in blood (red) and water are shown along the length of the gill, where blood is oxygenated in the zero to one direction and water is deoxygenated from one to zero. Panels A-C show $O_2$ uptake in freshwater fishes (FW, green) for the values of gill $O_2$ conductance ($G_d$) shown in Figure 1 (Lowest 0.02, Median 0.13 and Highest 0.36 μmol $mmHg^{-1}$ $g^{-1}$ $h^{-1}$). Panels D-F show $O_2$ uptake in seawater fishes (SW, blue) for their range of $G_d$ values (Lowest 0.10, Median 1.2 and Highest 11.4 μmol $mmHg^{-1}$ $g^{-1}$ $h^{-1}$). Haemoglobin (Hb) $O_2$ affinity (Hb $P_{50}$, the partial pressure at which Hb is 50% saturated with $O_2$) was set to cover the ranges shown in Figure 1 for both groups. The $M\dot{O}_{2max}$ and arterial Hb saturation ($S_aO_2$) values resulting from these simulations are listed in each panel.

**Figure 3 The maximal rate of oxygen consumption ($M\dot{O}_{2max}$) as a function of haemoglobin $O_2$ affinity (Hb $P_{50}$, the partial pressure at which Hb is 50% saturated with $O_2$) and gill $O_2$ conductance ($G_d$).** Panels A and B show the results for freshwater (FW; green) and seawater (SW; blue) fishes, respectively. $G_d$ and Hb $P_{50}$ values were chosen to cover the ranges reported in Figure 1 for both groups (see Fig. 2 caption for details).

**Figure 4 The maximal rates of oxygen consumption ($M\dot{O}_{2max}$) of freshwater (FW; green) and seawater (SW; blue) teleosts.** A) Haemoglobin $O_2$ affinity (Hb $P_{50}$, the partial pressure at which Hb is 50% saturated with $O_2$) was chosen to span the ranges reported in Figure 1 for both groups. $\dot{O}_{2max}$ was calculated for the upper and lower boundaries of gill diffusive conductance ($G_d$) observed in FW and SW fishes (see Fig. 2 caption for details). The dark shaded areas

represent the model results using the cardio-respiratory characteristics of rainbow trout (see Table S1). The light shaded areas represent simulations after the model was adjusted to the high Q□, [Hb] and tissue diffusive conductance of tunas (respectively, 6-fold, 2.3-fold and 3-fold higher values than in rainbow trout). Empirically measured values for Hb $P_{50}$ and $□O_{2max}$ from the literature were overlayed for FW and SW species whenever both parameters were available (1 rainbow trout, *Oncorhynchus mykiss*; 2 skipjack tuna, *Katsuwonus pelamis;* see search protocols described in the Appendix). B) There were significant effects of body mass and temperature on $□O_{2max}$, as determined by a generalized linear mixed model that accounted for phylogenetic non-independence of species. Regression line: $\log_{10}[M□O_{2max}$ (μmol $h^{-1}$)] = 0.84 + 0.019 temp (C°) + 0.88 $\log_{10}$[body mass (g)]. C) The residuals of this model provide mass- and temperature-independent measures of $□O_{2max}$ and the effect of salinity was tested by a phylogenetic ANOVA simulation that revealed no significant differences between FW and SW species.

**Figure 5 Calculated critical oxygen ($O_2$) tensions ($P_{crit}$) for freshwater (FW; green) and seawater (SW; blue) teleost fishes.** Simulations were run for the lowest, median and highest gill $O_2$ conductances ($G_d$) in FW (green; panels A-C) and SW (blue; panels D-F) fishes, respectively (see Fig. 2 for details). The rate of $O_2$ consumption ($M□O_2$) at decreasing water $PO_2$ (mmHg) was calculated with an $M□O_{2max}$ model (see Appendix), and standard $M□O_2$ ($M□O_{2std}$) was set to the value reported for rainbow trout, of 1.5 μmol $g^{-1}$ $h^{-1}$ (Kiceniuk and Jones, 1977). $P_{crit}$ was determined as the intersection between the $M□O_2$ and $M□O_{2std}$ curves over the range of haemoglobin $O_2$ affinities (Hb $P_{50}$; the $PO_2$ at which Hb is 50% saturated with $O_2$) that we observed in FW and SW fishes (Fig. 1).

**Figure 6 Modelled and empirically measured critical oxygen ($O_2$) tensions ($P_{crit}$) for freshwater (FW; green) and seawater (SW; blue) teleosts.** A) Modelling results for $P_{crit}$ of FW and SW fishes as a function of haemoglobin-oxygen affinity (Hb $P_{50}$; the $PO_2$ at which Hb is 50% saturated with $O_2$). The solid symbols and lines represent the calculated $P_{crit}$ values in FW and SW fish, as shown in Figure 5, and the shaded areas represent the combinations of $P_{crit}$ and Hb $P_{50}$ that are possible based on the gill and blood characteristics observed in both groups (Fig. 1). Empirically measured values for Hb $P_{50}$ and $P_{crit}$ from the literature were overlayed for FW and SW species whenever both parameters were available. Dashed lines indicate linear regressions through the empirical datasets for FW and SW fishes and their respective 95% confidence intervals. B) There were no significant effects of body mass on $P_{crit}$, as determined by a generalized linear mixed model that accounted for phylogenetic non-independence of species. C) The residuals of this model provide a mass-independent measure of $P_{crit}$ and the effect of salinity was tested by a phylogenetic ANOVA simulation that revealed no significant differences between FW and SW species.

## Appendix: Materials and Methods

*Literature review*

We conducted literature reviews of five cardio-respiratory variables in teleost fishes: gill surface area (GSA), blood-water diffusion distance (gill thickness), Hb-$O_2$ affinity (expressed as $P_{50}$; the $PO_2$ at which Hb is 50% saturated with $O_2$), maximal $O_2$ consumption rate ($M\square O_{2max}$), and critical $O_2$ tension ($P_{crit}$, the lowest $PO_2$ at which a fish can maintain standard $M\square O_2$). We included only teleost species in our data sets, which are by far the most diverse group of fishes and account for ~95% of all extant species (Nelson et al., 2006). The reasons for excluding other groups, such as the Chondrichthyes were based on their osmo-conforming physiology that changes the relationship between respiratory and ionoregulatory demands at the gill, and this group is almost exclusively found in the marine environment. We also excluded amphibious and obligatory air-breathing fishes (Damsgaard et al., 2020), as these traits fundamentally change the relationship between osmoregulation and respiration that we sought to examine (Graham, 1997). Additionally, the values we used were limited to those from individuals in juvenile or adult developmental stages, measured under control conditions (i.e., untreated and non-acclimated individuals).

Commenting on the methodological approaches used in literature studies was outside the scope of this review and generally, we considered data generated with different methods, while being aware that our variables of interest are sensitive to different experimental protocols. However, we did limit our search to Hb $P_{50}$ measured on whole blood, and excluded those on haemolysates, which are less representative of *in vivo* conditions even when standardising for allosteric effectors (Berenbrink, 2006). For GSA, we limited our search to measurements of total gill surface area or lamellar surface area.

All literature searches were performed on Clarivate's Web of Science®, in 2020 and 2021, using the following search terms: GSA (gill AND surface area AND fish; lamellar surface area AND fish); Hb $P_{50}$ (oxygen* binding affinity AND fish; hemoglobin* AND P50 AND fish); $P_{crit}$ (critical oxygen* AND fish; Pcrit + fish; Pcrit + hypoxia + fish); $M\square O_{2max}$ (maximum metabolic rate + fish; MO2max + fish). In instances where the mined studies cited other studies that our search did not uncover, we included these data if they were appropriate. Two recent reviews were particularly valuable in our search, providing many of the mined data points

(Killen et al., 2017; Rogers et al., 2016). Values, references and URLs for all cited studies can be found in the Excel spreadsheets in the supplementary materials.

We analysed our data in R Studio (v. 1.3.1056) using a generalized linear mixed model (GLMM) to identify correlations between continuous variables using the *MCMCglmm()*-function in the MCMCglmm-package in R (Hadfield, 2010; Hadfield and Nakagawa, 2010). This method treats individual studies on same species as a random effect (i.e., taking into account intraspecific variation) and accounted for phylogenetic non-independence of species. Here, we used a maximum clade credibility tree generated from 100 Bayesian posterior probability trees from Rabosky et al. (2018) using the *maxCladeCred()* function in the *phangorn* package in R (Schliep, 2011).

To test the effect of salinity on GSA and $G_d$, while taking into account the hypo-allometric scaling with body mass, we fitted a GLMM-model to the $\log_{10}$-transformed data and calculated the residuals. These residuals provide a mass-independent measure of GSA and $G_d$ (Garland et al., 1992). We then tested for an effect of salinity on the residuals with a phylogenetic analysis of variance simulation (Garland et al., 1993), using the *phylANOVA()*-function in the *phytools*-package in R (Revell, 2012). Similar approaches were used for the other variables:

To test for the effect of salinity on $\dot{M}O_{2max}$, while taking into account temperature and body mass, we fitted three GLMM-models to the data:

$\log_{10}(\dot{M}O_{2max}) \sim \log_{10}(\text{body mass}) + \text{temperature}$ (DIC = –124)

$\log_{10}(\dot{M}O_{2max}) \sim \text{temperature}$ (DIC = –81.6)

$\log_{10}(\dot{M}O_{2max}) \sim \log_{10}(\text{body mass})$ (DIC = 205)

We selected the full model based on the lowest deviance information criterion value (DIC), calculated the residuals, and tested for an effect of salinity on the $\dot{M}O_{2max}$ residuals, as described above. To test for the effect of salinity on Hb-$O_2$ affinity ($P_{50}$), we first fitted a GLMM-model to $\log_{10}(P_{50})$ versus absolute temperature$^{-1}$, calculated the residuals that we tested for an effect of salinity. $P_{crit}$ and gill thickness were body-mass independent thus, we used the raw values in a phylogenetic ANOVA to test for an effect of salinity.

*Mathematical model of gas exchange*

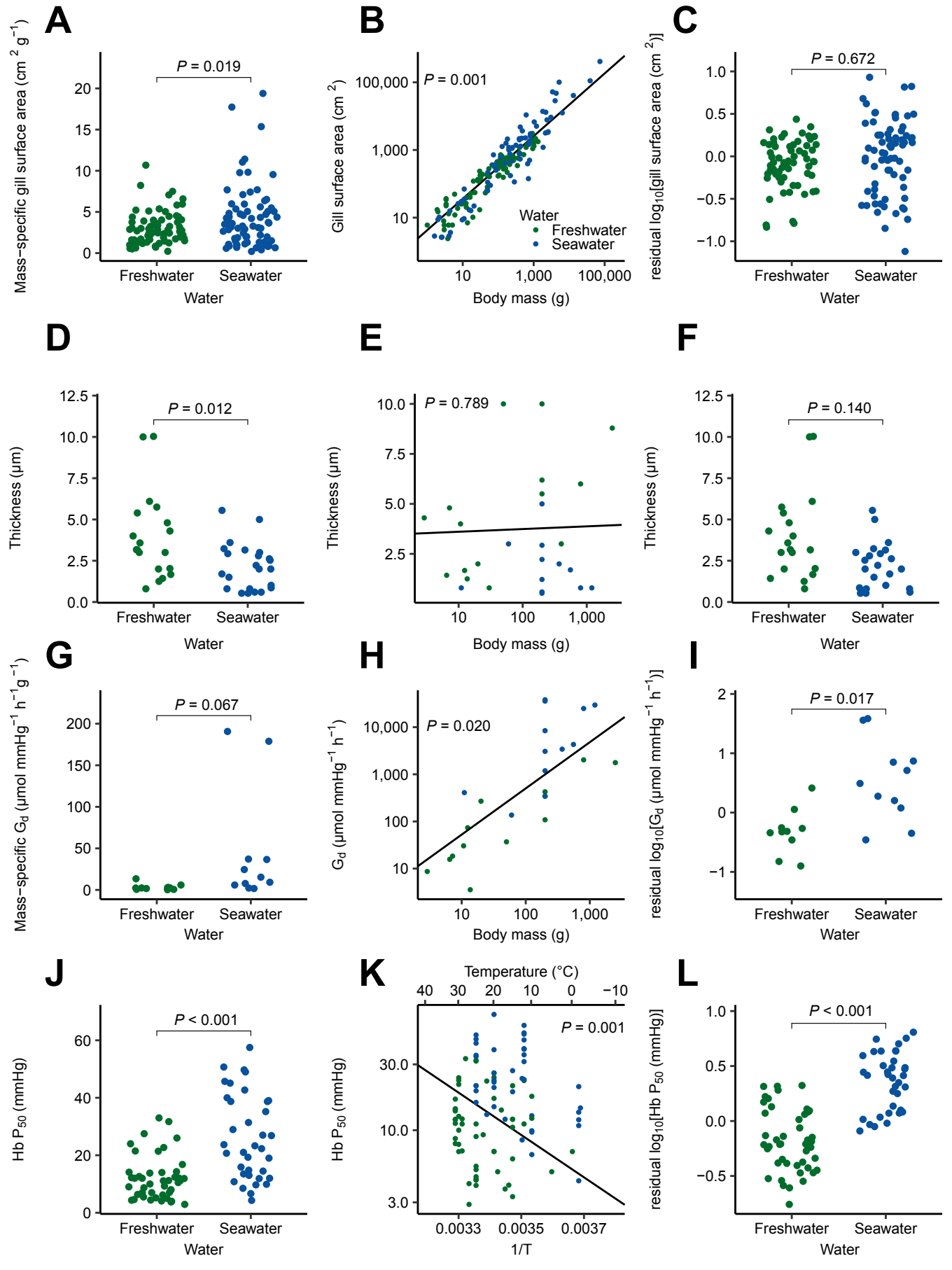
To study the effects of Hb $P_{50}$ and $G_d$ on gas exchange at the fish gills we implemented the previously published mathematical model of Malte and Weber (1985) using R v.3.6.2. in

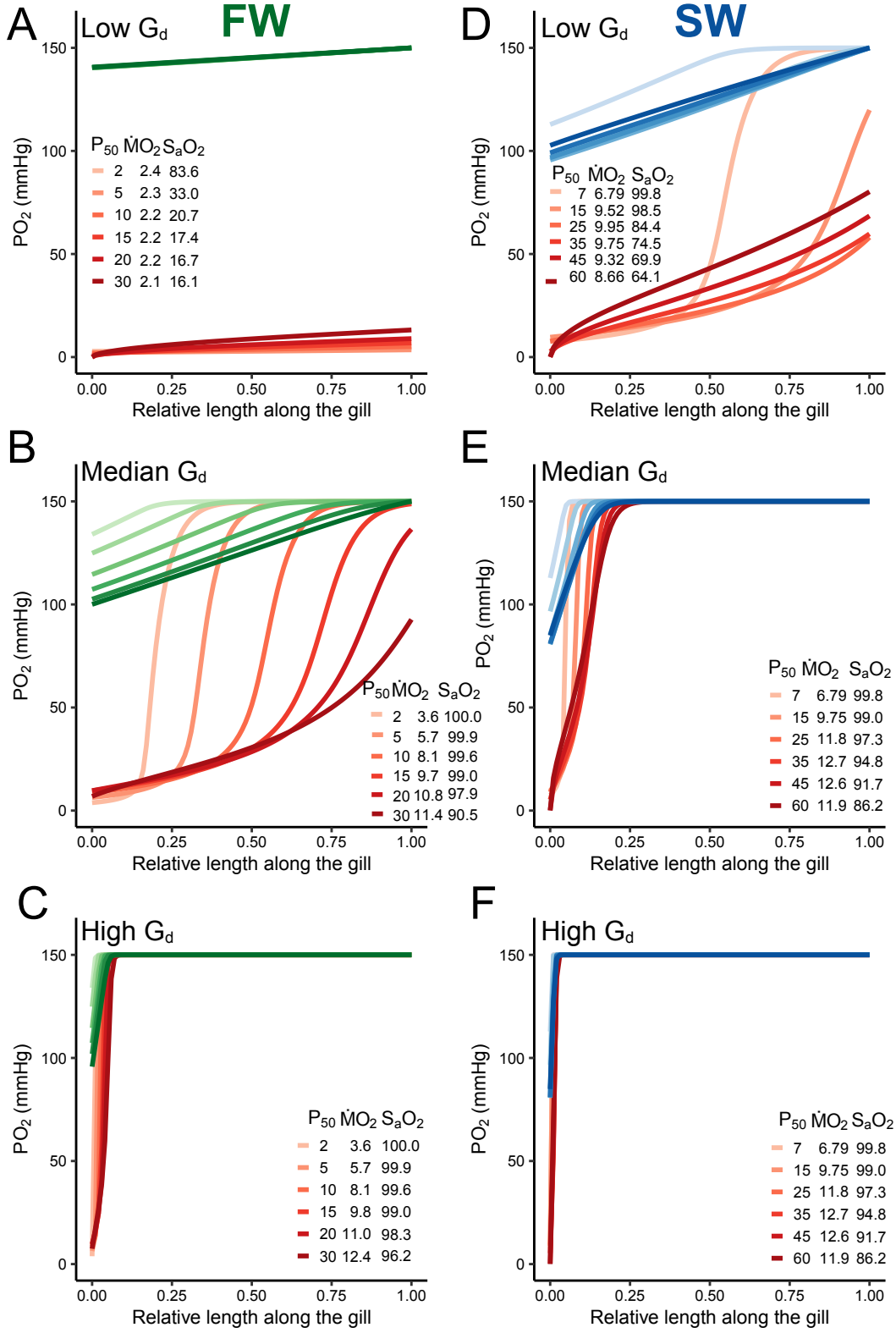
RStudio v.1.2.5033. Briefly, the model solves a system of differential equations to calculate the changes in water and blood $PO_2$ during counter-current gas exchange along the length of a model fish gill. In R, the differential equations were solved as a boundary value problem with venous $PO_2$ and inspired water $PO_2$ as the input parameters, using the bvpsolve package (Mazzia et al., 2014), with bvptwp that uses a mono-implicit Runge-Kutta (MIRK) method with deferred corrections (Cash and Mazzia, 2005; Cash and Wright, 1991) and a continuation method. The output from these gas exchange simulations are the calculated values for expired water $PO_2$ and arterial $PO_2$, which is then used to calculate other arterial blood parameters. Next the model predicts the outcome of gas exchange at the tissues to calculate venous $PO_2$ and other venous blood parameters that are then used for the next iteration of gas exchange at the gills; the cycle is repeated until the final calculated venous $PO_2$ is in equilibrium with the gas exchange systems at the gills and at the tissues (Malte and Weber, 1985; Malte and Weber, 1987; Malte and Weber, 1989; Wang and Malte, 2011). Our model focused only on the exchange of $O_2$ at the fish gill and does not take into account changes in $CO_2$ excretion, however the model does simulate a pH shift at the tissues that leads to an increase in Hb $P_{50}$ by the Bohr effect and recovery of Hb $P_{50}$ during venous transit.

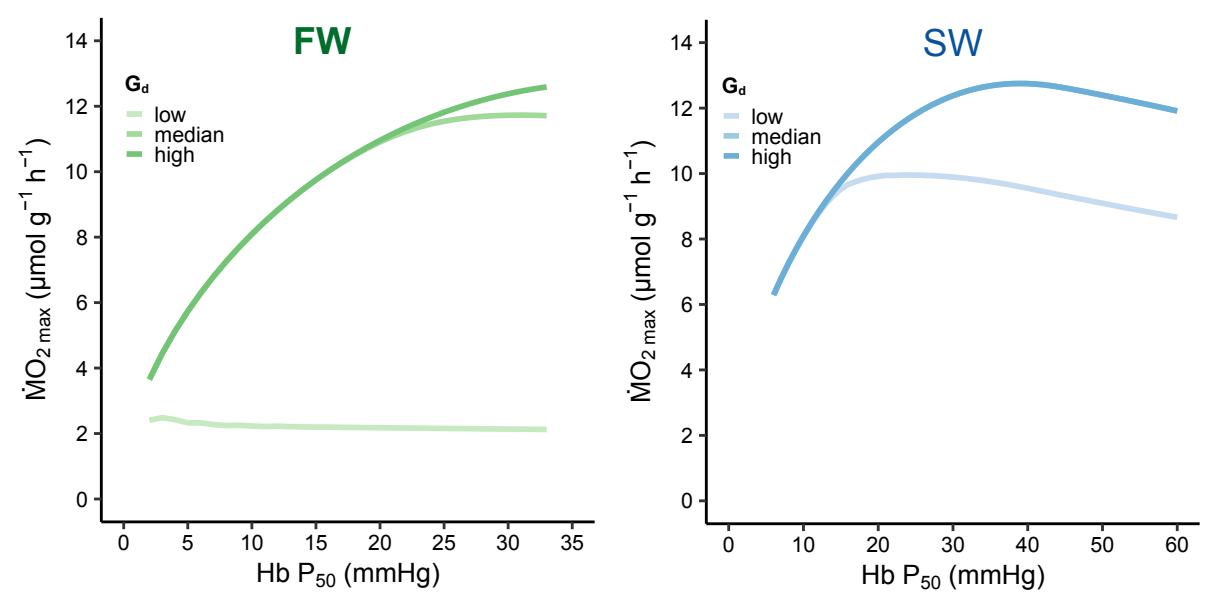
The model was "calibrated" against the cardio-respiratory characteristics measured in rainbow trout (*Oncorhynchus mykiss*) at 10°C, by Kiceniuk and Jones (1977). Specifically, the diffusion coefficient for $O_2$ at the gills ($D_{O2}$) and the tissue $O_2$ conductance, two values for which no reliable measurements exist, were adjusted so that the model output matched the empirically measured $M\dot{O}_{2max}$, arterial and venous $PO_2$, and tissue $O_2$ extraction in rainbow trout. All other input parameters for the model calibration are listed in Table S1 with their respective references.

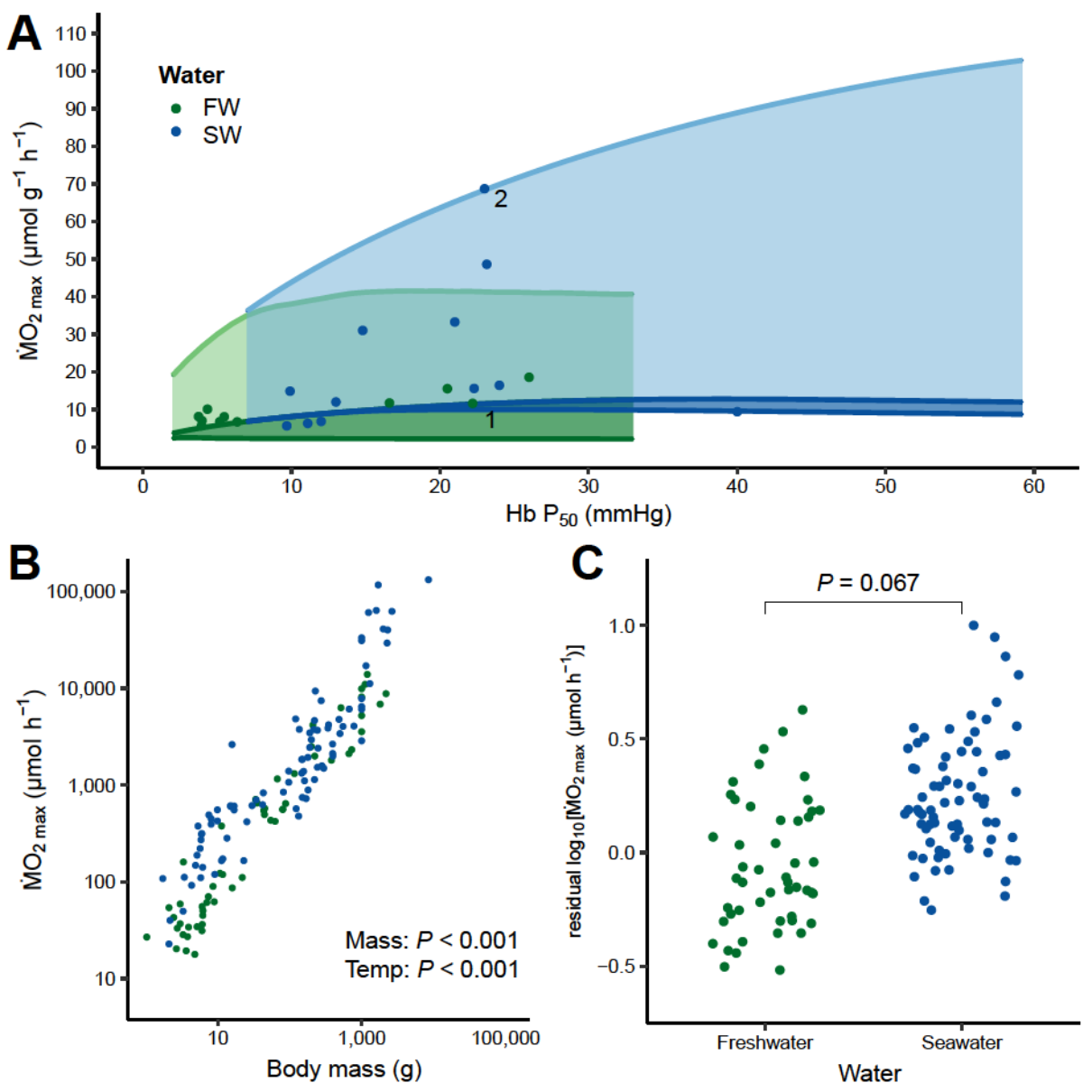
In all simulations, [Hb] was set to 1 mM based on the summary data by Gallaugher et al. (1998) and the cooperativity of Hb-$O_2$ binding (the Hill coefficient) was set to $n = 2$. While this represents a simplistic approach, the effects of cooperativity on gas exchange at the fish gill have been addressed previously (Malte and Weber, 1987) and we observed no systematic differences in cooperativity between FW and SW teleosts that would warrant a more detailed investigation within the context of our study. In addition, some groups (e.g., Cottidae) display a wide range of Hb $P_{50}$ at rather constant Hill coefficients, indicating that phylogeny may be a better predictor of Hill coefficients than e.g. Hb $P_{50}$ (Mandic et al., 2009). Finally, to test our hypothesis, we

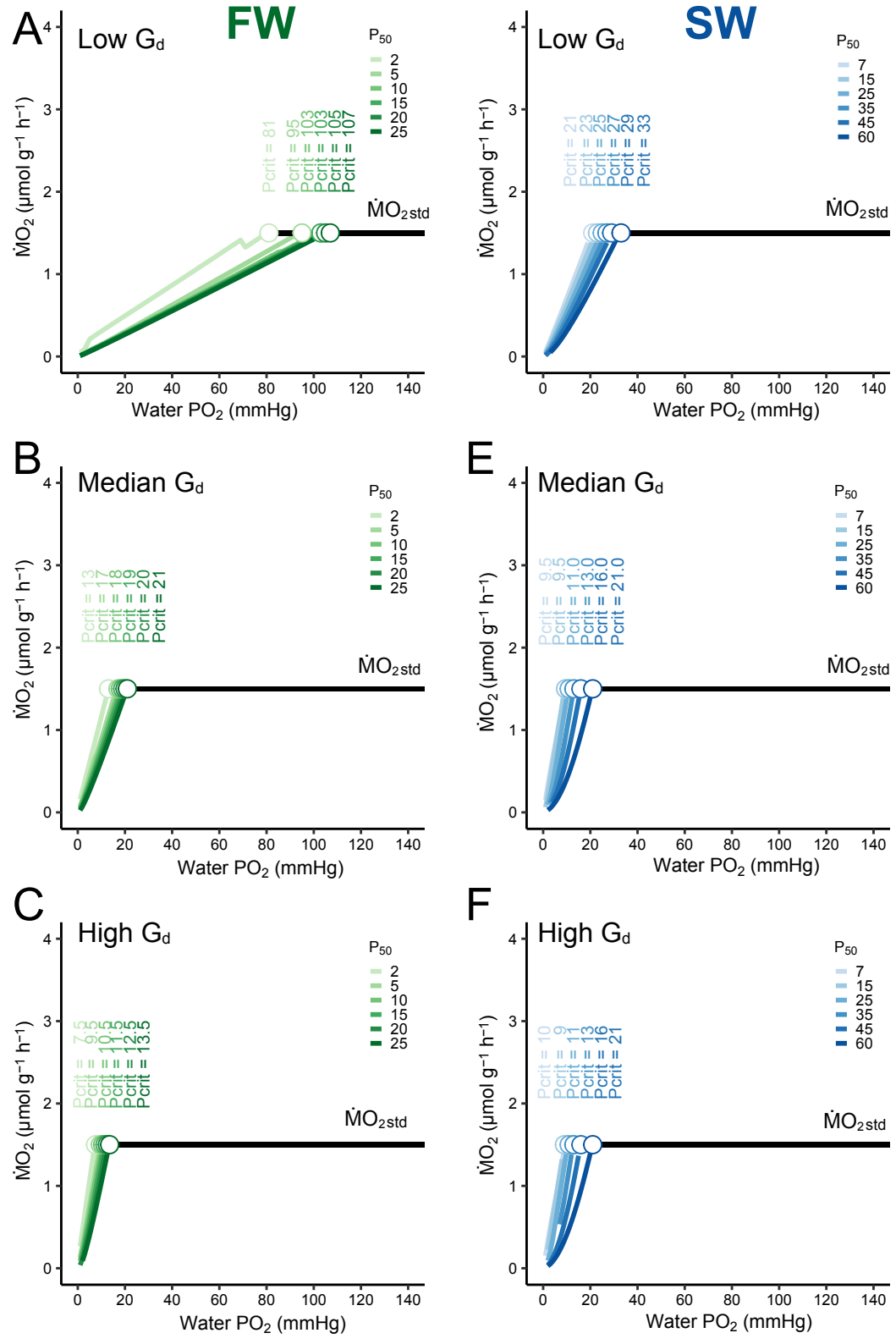
814 adjusted the parameters of $G_d$ and Hb $P_{50}$ based on the values for FW and SW fishes that resulted
815 from our review of the literature.

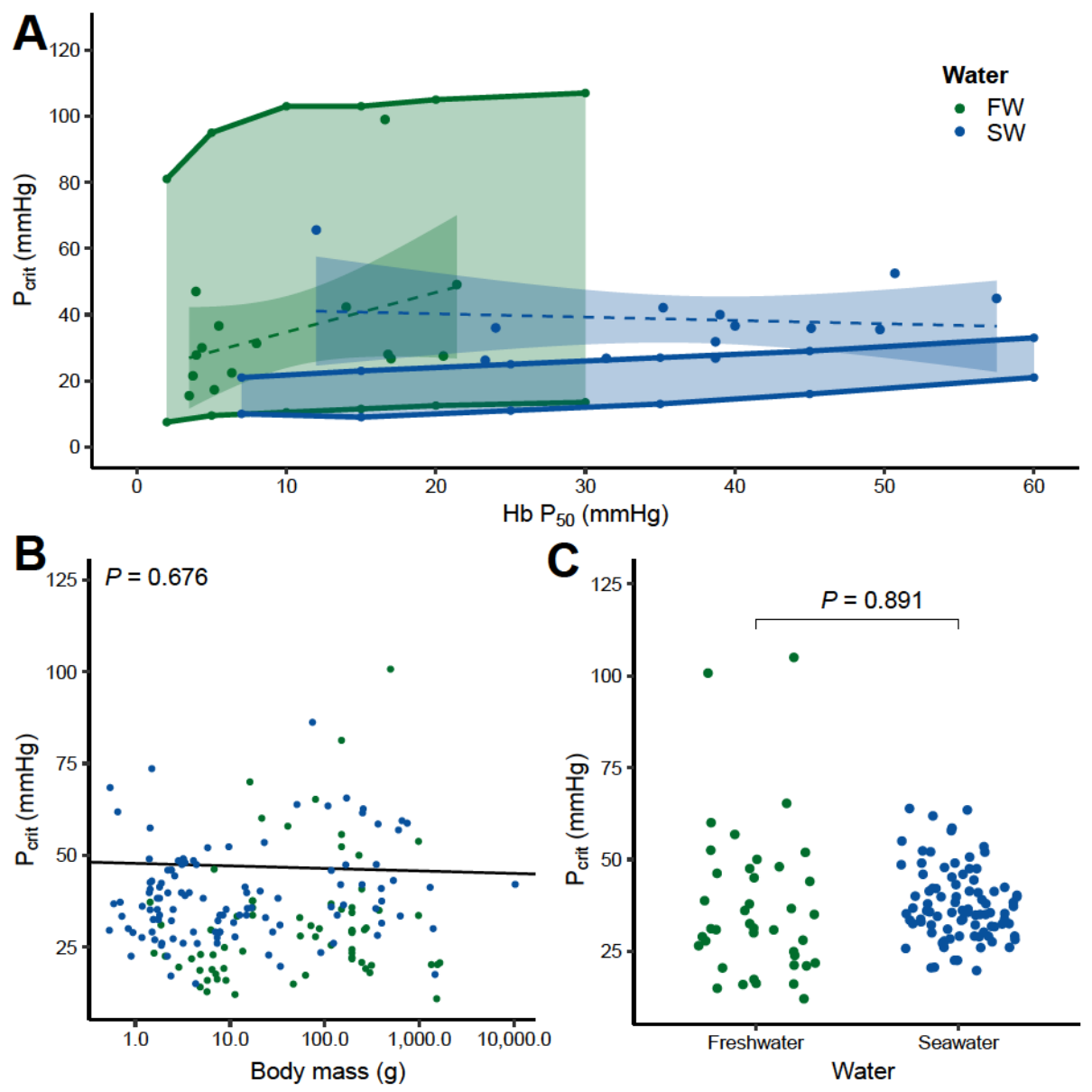

A
Mass−specific gill surface area (cm 2 g −1)
P = 0.019
Freshwater
Seawater
Water
B
Gill surface area (cm 2)
P = 0.001
Water
Freshwater
Seawater
Body mass (g)
C
residual log10[gill surface area (cm 2)]
P = 0.672
Freshwater
Seawater
Water
D
Thickness (µm)
P = 0.012
Freshwater
Seawater
Water
E
Thickness (µm)
P = 0.789
Body mass (g)
F
Thickness (µm)
P = 0.140
Freshwater
Seawater
Water
G
Mass−specific Gd (µmol mmHg−1 h−1 g−1)
P = 0.067
Freshwater
Seawater
Water
H
Gd (µmol mmHg−1 h−1)
P = 0.020
Body mass (g)
I
residual log10[Gd (µmol mmHg−1 h−1)]
P = 0.017
Freshwater
Seawater
Water
J
Hb P50 (mmHg)
P < 0.001
Freshwater
Seawater
Water
K
Temperature (°C)
P = 0.001
Hb P50 (mmHg)
1/T
L
residual log10[Hb P50 (mmHg)]
P < 0.001
Freshwater
Seawater
Water


A
Low $G_d$
FW
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
2 2.4 83.6
5 2.3 33.0
10 2.2 20.7
15 2.2 17.4
20 2.2 16.7
30 2.1 16.1
$PO_2$ (mmHg)
Relative length along the gill
D
Low $G_d$
SW
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
7 6.79 99.8
15 9.52 98.5
25 9.95 84.4
35 9.75 74.5
45 9.32 69.9
60 8.66 64.1
$PO_2$ (mmHg)
Relative length along the gill
B
Median $G_d$
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
2 3.6 100.0
5 5.7 99.9
10 8.1 99.6
15 9.7 99.0
20 10.8 97.9
30 11.4 90.5
$PO_2$ (mmHg)
Relative length along the gill
E
Median $G_d$
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
7 6.79 99.8
15 9.75 99.0
25 11.8 97.3
35 12.7 94.8
45 12.6 91.7
60 11.9 86.2
$PO_2$ (mmHg)
Relative length along the gill
C
High $G_d$
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
2 3.6 100.0
5 5.7 99.9
10 8.1 99.6
15 9.8 99.0
20 11.0 98.3
30 12.4 96.2
$PO_2$ (mmHg)
Relative length along the gill
F
High $G_d$
$P_{50}$ $\dot{M}O_2$ $S_aO_2$
7 6.79 99.8
15 9.75 99.0
25 11.8 97.3
35 12.7 94.8
45 12.6 91.7
60 11.9 86.2
$PO_2$ (mmHg)
Relative length along the gill


FW
SW
$G_d$
low
median
high
$\dot{M}O_{2\,max}$ (µmol $g^{-1}$ $h^{-1}$)
Hb $P_{50}$ (mmHg)
0
2
4
6
8
10
12
14
0
5
10
15
20
25
30
35
40
50
60


A
Water
FW
SW
2
1
$\dot{M}O_{2\,max}$ (μmol $g^{-1}$ $h^{-1}$)
Hb $P_{50}$ (mmHg)
B
$\dot{M}O_{2\,max}$ (μmol $h^{-1}$)
Mass: $P < 0.001$
Temp: $P < 0.001$
Body mass (g)
C
$P = 0.067$
residual $\log_{10}[\dot{M}O_{2\,max}$ (μmol $h^{-1}$)]
Freshwater
Seawater
Water


A
FW
Low $G_d$
$P_{50}$
2
5
10
15
20
25
Pcrit = 81
Pcrit = 95
Pcrit = 103
Pcrit = 103
Pcrit = 105
Pcrit = 107
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)
SW
Low $G_d$
$P_{50}$
7
15
25
35
45
60
Pcrit = 21
Pcrit = 23
Pcrit = 25
Pcrit = 27
Pcrit = 29
Pcrit = 33
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)
B
Median $G_d$
$P_{50}$
2
5
10
15
20
25
Pcrit = 13
Pcrit = 17
Pcrit = 18
Pcrit = 19
Pcrit = 20
Pcrit = 21
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)
E
Median $G_d$
$P_{50}$
7
15
25
35
45
60
Pcrit = 9.5
Pcrit = 9.5
Pcrit = 11.0
Pcrit = 13.0
Pcrit = 16.0
Pcrit = 21.0
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)
C
High $G_d$
$P_{50}$
2
5
10
15
20
25
Pcrit = 7.5
Pcrit = 10.5
Pcrit = 11.5
Pcrit = 12.5
Pcrit = 13.5
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)
F
High $G_d$
$P_{50}$
7
15
25
35
45
60
Pcrit = 10
Pcrit = 9
Pcrit = 11
Pcrit = 13
Pcrit = 16
Pcrit = 21
$\dot{M}O_{2std}$
$\dot{M}O_2$ (µmol $g^{-1}$ $h^{-1}$)
Water $PO_2$ (mmHg)


A
Water
FW
SW
$P_{crit}$ (mmHg)
Hb $P_{50}$ (mmHg)
B
$P$ = 0.676
$P_{crit}$ (mmHg)
Body mass (g)
C
$P$ = 0.891
$P_{crit}$ (mmHg)
Freshwater
Seawater
Water