

An image dataset of cleared, x-rayed, and fossil leaves vetted to plant family for human and machine learning

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Abstract

Leaves are the most abundant and visible plant organ, both in the modern world and the fossil record. Identifying foliage to the correct plant family based on leaf architecture is a fundamental botanical skill that is also critical for isolated fossil leaves, which often, especially in the Cenozoic, represent extinct genera and species from extant families. Resources focused on leaf identification are remarkably scarce;

however, the situation has improved due to the recent proliferation of digitized herbarium material, live-plant identification applications, and online collections of cleared and fossil leaf images. Nevertheless, the need remains for a specialized image dataset for comparative leaf architecture. We address this gap by assembling an open-access database of 30,252 images of voucherized leaf specimens vetted to family level, primarily of angiosperms, including 26,176 images of cleared and x-rayed leaves representing 354 families and 4,076 of fossil leaves from 48 families. The images maintain original resolution, have user-friendly filenames, and are vetted using APG and modern paleobotanical standards. The cleared and x-rayed leaves include the Jack A. Wolfe and Leo J. Hickey contributions to the National Cleared Leaf Collection and a collection of high-resolution scanned x-ray negatives, housed in the Division of Paleobotany, Department of Paleobiology, Smithsonian National Museum of Natural History, Washington D.C.; and the Daniel I. Axelrod Cleared Leaf Collection, housed at the University of California Museum of Paleontology, Berkeley. The fossil images include a sampling of Late Cretaceous to Eocene paleobotanical sites from the Western Hemisphere held at numerous institutions, especially from Florissant Fossil Beds National Monument (late Eocene, Colorado), as well as several other localities from the Late Cretaceous to Eocene of the Western USA and the early Paleogene of Colombia and southern Argentina. The dataset facilitates new research and education opportunities in paleobotany, comparative leaf architecture, systematics, and machine learning.

Keywords

Angiosperms, cleared leaves, data science, fossil leaves, leaf architecture, paleobotany

Introduction

General patterns of angiosperm leaf architecture, the shape and venation characters of leaves, are well known for very few of the more than 400 angiosperm families. The development of a standard descriptive terminology (von Ettingshausen 1861; Hickey 1973, 1979; Ellis et al. 2009) has catalyzed increased detail and reproducibility in species descriptions of both living and fossil leaves. However, despite the use of numerous visual examples (e.g., Ellis et al. 2009), publications to date do not inform the reader how to accomplish the fundamental task of identifying leaves that, as for the great majority of leaf fossils, are isolated from the rest of the plant and missing diagnostic information from stipules, leaf organization, and reproductive and other organs.

To build their knowledge of leaf architecture, researchers still rely primarily on “oral tradition” from a dwindling number of knowledgeable colleagues and a handful of survey papers and field guides that emphasize purportedly diagnostic leaf features (Hickey and Wolfe 1975; Gentry 1993; da Ribeiro et al. 1999; Keller 2004). There is significant literature on the leaf architecture and leaf-fossil records of various taxa (among many others, von Ettingshausen 1858; Hill 1982; Jones 1986; Manchester 1987; Todzia and Keating 1991; Gandolfo and Romero 1992; Premoli 1996; Fuller and Hickey 2005; Martínez-Millán and Cevallos-Ferriz 2005; Doyle 2007; Kellner et al. 2012). However, many of the most diverse and ecologically significant groups of angiosperms have virtually no documentation of diagnostic leaf-blade features (e.g., Asteraceae, Rubiaceae), and thus their leaf fossils remain largely unrecognized,

though probably hidden in plain sight in museum collections (see Wilf 2008; Wilf et al. 2016). More than half of fossil-leaf species in many older monographs are thought to have been misclassified (see Dilcher 1971), and most of the millions of leaf fossils in the general stratigraphic collections of the world's museums are not yet identified.

Machine-vision algorithms, as seen in popular applications such as LeafSnap (Kumar et al. 2012), Pl@ntNet (Bonnet et al. 2018), and iNaturalist (Van Horn et al. 2018), are making spectacular breakthroughs in automated species identification of live plants; however, they provide little, if any, feedback about the diagnostic features they detect. Few algorithms have attempted to generalize above the species level (Wilf et al. 2016; Carranza-Rojas et al. 2018), and so far the methods do not work on leaf fossils, which mostly represent extinct species and often extinct genera.

Increasing general knowledge of leaf architecture for both human and machine learners depends on the development of customized, accessible, vetted visual libraries that allow rapid morphological comparisons of a high phylogenetic diversity of extant and fossil leaves. The recent proliferation of digitized plant-image resources comprises an invaluable reference for plant morphology, already including tens of millions of digitized herbarium sheets on portals and aggregator sites such as JStor Global Plants (<https://plants.jstor.org>), iDigBio (<https://www.idigbio.org>), RecolNat (<https://www.recolnat.org>), and many others, as well as servers located at numerous individual herbaria worldwide (e.g., Bakker et al. 2020). However, studying leaf comparative morphology is not simple because leaves only represent part of the visual field of a herbarium sheet and appear, with overlaps, at many different angles and sizes. Computer-vision algorithms that blur text or segment leaves from background or from other plant material are likely to help solve this issue (Hussein et al. 2020, 2021; Weaver et al. 2020; de Lutio et al. 2021). However, many visual distractors remain, and critical details of higher-order venation are often not visible in digitized herbarium sheets. Assessing leaf architecture at family level from digital herbaria also requires examination of extremely large numbers of specimens for all but the most species-poor families. In this regard, JStor Global Plants stands out for prioritizing type specimens collated digitally from across the world's herbaria, thus allowing rapid surveys of the taxa in a family based on protologue voucher material. Finally, digitization efforts are far more advanced in resource-rich countries, whereas many significant collections are located in developing nations where herbarium digitization is occurring at a slower pace.

Cleared or x-rayed leaves from phylogenetically diverse taxa, selectively sampled from vouchered herbarium sheets, remain the most valuable visual reference for comparative study of leaf architecture because they have a similar visual presentation, with high capture of venation detail and comparatively few distractors. Existing collections of this type are fragile, mostly made decades ago as references for fossil leaf identification by selecting leaves from herbarium sheets, then either chemically clearing the specimens of most tissues other than veins and mounting them on glass slides or x-ray imaging them, in either case with extreme care and effort. Most cleared-leaf collections suffer from deterioration of the mounting media, which obscures large areas of the leaves; thus, photographic archiving offers a form of visual preservation before further

degradation occurs. The largest and best-known cleared-leaf collections are those of the late Drs. Jack A. Wolfe and Leo J. Hickey, together now forming the National Cleared Leaf Collection (NCLC; NCLC-W and NCLC-H, respectively), housed in the Division of Paleobotany of the Smithsonian Institution National Museum of Natural History (NMNH, repository acronym USNM, Washington, D.C.).

For the many users who may find it challenging to visit these collections in person for suitable lengths of time, many cleared and x-rayed leaf collections are already accessible from various websites or in print. These valuable resources include the NCLC-W and other collections in the Cleared Leaf Image Database (<http://clearedleavesdb.org>; Das et al. 2014); the NCLC-H served from the Yale Peabody Museum (<https://collections.peabody.yale.edu/pb/nclc>); the Daniel I. Axelrod cleared-leaf collections of the University of California Museum of Paleontology (UCMP; <https://ucmp.berkeley.edu/collections/paleobotany-collection/ucmp-cleared-leaf-collection>); the National Museum of Nature and Science (NMNS, Ibaraki, Japan) Cleared Leaf Database by Drs. Toshimasa Tanai and Kazuhiko Uemura (https://www.kahaku.go.jp/research/db/geology-paleontology/cleared_leaf/database/?lg=en); leaf x-ray images of Australian rainforest plants by the late Dr. David C. Christophel and colleagues (Christophel and Hyland 1993; Christophel and Rowett 1996), some of which are maintained in the online Australian Tropical Rainforest Plants identification system (<https://apps.lucidcentral.org/rainforest/text/intro/index.html>); and the late Dr. Edward P. Klucking's book series illustrating cleared leaves from selected families (Klucking 1986–2003). We also note an open-access image dataset of cleared leaves from Borneo, consisting of small (1 cm²) lamina samples (Blonder et al. 2019; Xu et al. 2021). In most of the online image sets, bulk downloads are not easily done, images are downsampled to low resolution, and the filenames are not standardized, requiring significant manual effort to re-organize and collate them for a particular project. Adding further complications to data modularity, taxonomic data have often become partially obsolete.

Isolated fossil leaves present an additional set of challenging problems (e.g., Wilf 2008), including incomplete preservation, morphological convergence, and the well-known legacy of innumerable taxonomic misidentifications in older publications (see Dilcher 1971, 1974; Hill 1982). Numerous high-quality systematic treatments have become available for many leaf-fossil taxa, especially over the last few decades, but the images are dispersed across publications and are usually of low resolution. An increasing number of images of vouchered fossil-leaf collections is available online from natural history museums. Examples include aggregator sites such as GBIF (gbif.org) and individual institutions such as the Yale Peabody Museum, (<https://peabody.yale.edu/collections/paleobotany>), the Burke Museum (www.burkemuseum.org/collections-and-research/geology-and-paleontology/collections-database/images.php), the University of Colorado Boulder Museum of Natural History (<https://www.colorado.edu/cumuseum/research-collections/paleontology/invertebrates-plants>), and the UCMP (<https://ucmpdb.berkeley.edu>). Nevertheless, museum servers and project sites (e.g., Traiser et al. 2018) usually retain the taxonomy as published, which is vital for the nomenclatural stability of type specimens but well known to be problematic, especially

for the many older collections that have not been revised under modern standards. All these issues make it very difficult for researchers, students, and non-specialists to form a reliable base of knowledge about fossil-leaf identification and have perhaps engendered an overreliance on methods that do not require taxonomy at all, such as leaf morphotyping (see Wilf 2008).

Here, we meet the community need for a specialized dataset of leaf images by consolidating a set of original-resolution photographs of vouchered extant and fossil specimens (Fig. 1, Table 1), primarily of angiosperms, vetted to family level and relabeled to user-friendly filenames, into an open-access archive in a single, standard file format (jpeg, at minimum possible compression). A principal goal, based on many years of practical experience using leaf-image datasets in our research, is maximum and sustained ease of use with rapid access to the entire library. Thus, instead of creating an interactive database that may become obsolete and limit resolution or user flexibility, we simply provide the image files in labeled folders that can easily be downloaded, then viewed and searched using any visual browser (e.g., Adobe Bridge, Adobe Lightroom, Windows Explorer) on any suitable device, such as a personal computer.

The full image dataset and supporting data files are available open-access for download in a single Figshare Plus data collection at <https://doi.org/10.25452/figshare.plus.14980698> (hereafter, “the Figshare archive”). The components described below

Table 1. Summary of component datasets.

Collection	Collection type	#Images	#Families	#Genera, approx.	#Species, approx.	Repository	Collection numbers†	Other data and images‡
NCLC-Wolfe	cleared leaves	16,249	267	3,893	12,439	USNM	secondary	http://clearedleavesdb.org
NCLC-Hickey	cleared leaves	6,861	313	1,678	5,723	USNM	secondary	https://collections.peabody.yale.edu/pb/nclc
Axelrod Cleared Leaves	cleared leaves	832	89	270	641	UCMP	primary	https://ucmpdb.berkeley.edu/photos/cleared_leaf.html
Wing X-Rays	x-ray negatives	2,234	26	416	890	USNM	secondary	n/a
Total extant		26,176	354	4,573	17,385			
Florissant, Meyer et al. (2008) project	fossil leaves	666	23	47	73	several	secondary	https://flfo-search.colorado.edu
Florissant, FLFO	fossil leaves	2,654	21	40	70	FLFO	primary	https://www.flickr.com/photos/155340198@N06
General fossil collection	fossil leaves	756	39	93	135	several	primary	n/a
Total fossil		4,076	48	129	222			

Abbreviations: FLFO, Florissant Fossil Beds National Monument. NCLC, National Cleared Leaf Collection. UCMP, University of California Museum of Paleontology. USNM, National Museum of Natural History, Smithsonian Institution.

†As used in records specific to these collections and our image filenames. Secondary inventory numbers are those assigned by the creators of collections that were assembled from several primary collections. Examples are the cleared and x-rayed leaf samples physically gathered from primary herbarium sheets and the Meyer et al. (2008) Florissant photographic collection of specimens housed at numerous primary repositories. The Meyer et al. secondary (photograph) numbers have an informal “CU” prefix added here to the filenames, merely to distinguish them easily from the FLFO set in searches and not to indicate primary repository. See text for more details.

‡Images, specimen inventories, and other supporting data are available in the Figshare item accompanying this article: <https://doi.org/10.25452/figshare.plus.14980698>. Fossil taxa and references are listed in Appendix 1.

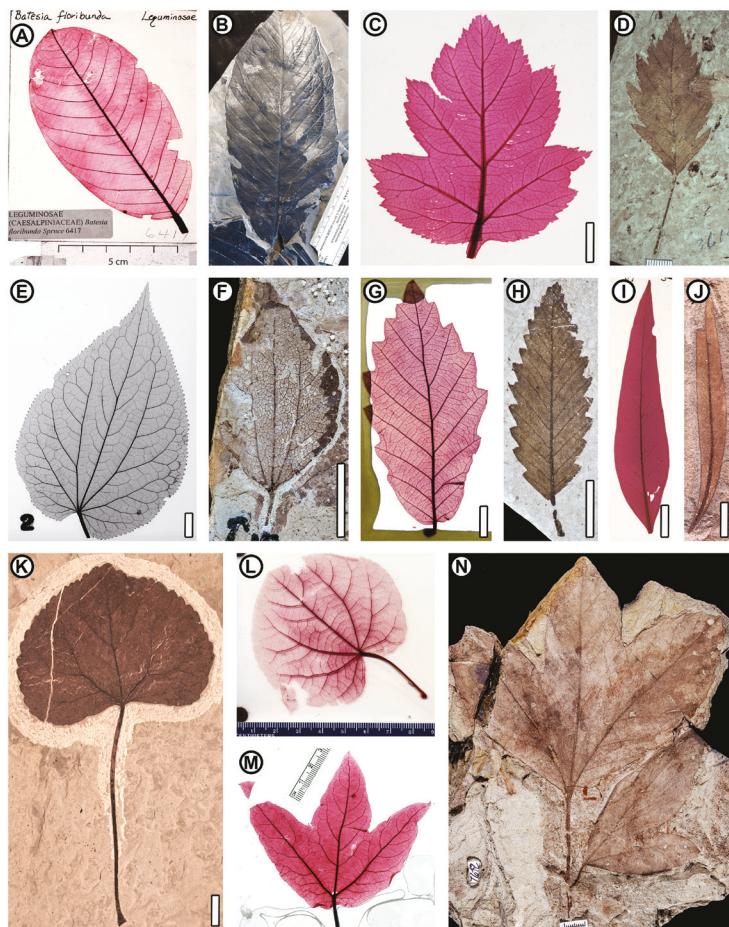


Figure 1. Selected image pairs of confamilial extant and fossil (see Appendix 1) leaves from the dataset **A** *Batesia floribunda* Spruce ex. Benth. (Fabaceae), NCLC-W 6417, showing typical layout of a cleared-leaf slide with original annotations (other examples are cropped in this figure); source voucher Froes 12074, DS 291771 (at CAS), Amazonas, Brazil **B** Fabaceae sp. CJ1, SGC-ICP-10173; Cerrejón mine, middle-late Paleocene of Guajira Peninsula, Colombia **C** *Crataegus viridis* L. (Rosaceae), NCLC-W 11951b; H. Meyer s/n (collected 1974, no other voucher), cultivated, California, USA **D** *Crataegus copeana* (Rosaceae), UCMP 3610; Florissant, late Eocene of Colorado, USA; H. Meyer photograph number 0420 **E** *Tetracentron sinense* Oliv. (Trochodendraceae), S. Wing negative 71-002; E.H. Wilson 659, US 599036, Szechuan, China **F** *Ziziphoides flabellum* (Trochodendraceae), USNM 560134; Mexican Hat, early Paleocene of Montana, USA **G** *Quercus prinus* L. (Fagaceae), NCLC-W 6137; H. Foster 8223, US 1730249, Florida, USA **H** *Fagopsis longifolia* (Fagaceae), FLFO 003432A; Florissant, late Eocene of Colorado, USA **I** *Eucalyptus astringens* (Maiden) Maiden (Myrtaceae), NCLC-W 10489; J.H. Maiden (9 November 1909), Western Australia, UC 437518 **J** *Eucalyptus frenguilliana* (Myrtaceae), MPEF-Pb 2344; Laguna del Hunco, early Eocene of Chubut, Argentina **K** *Cercidiphyllum obtitum* (Cercidiphyllaceae), DMNH 25061; Republic, early Eocene of Washington, USA **L** *Cercidiphyllum japonicum* Siebold & Zucc. ex J.J.Hoffm. & J.H.Schult.bis (Cercidiphyllaceae), Axelrod cleared leaf 166; UCMP (no other voucher) **M** *Platanus racemosa* Nutt. (Platanaceae), NCLC-H 6631; Handel s/n (collected 1985, no other voucher), California, USA **N** *Erlingdorfia montana* (compound-leaved Platanaceae), DMNH 7642; Hell Creek Formation, Late Cretaceous of North Dakota, USA. Scale bars: centimeters as labeled (**A, B, L, M**); 1 cm when not labeled (**C-K, N**).

are summarized in Table 1, along with relevant online resources where many of the specimens can already be searched, usually at lower resolution. Although individual linkage of each specimen with online resources would be desirable, it is highly impractical at present because the necessary tags and lookup tables have never been compiled and vetted for most of the collections used here. For readability, we use “leaves” to refer to all specimens discussed here, whether they are leaves, leaflets, or other plant organs that are included in small numbers.

Cleared and x-rayed leaves

The cleared and x-rayed leaf-image collections included here were chosen for availability of a large number of botanically diverse, high-quality images, accessible voucher data, and open-access re-use permissions. The collections primarily represent non-monocot (“dicot”) angiosperm leaves, with minor representation of monocots, other vascular plant groups, and non-foliar plant organs. Several other large cleared and x-rayed leaf collections exist (see Introduction) but were not used in the dataset presented here for various reasons. For example, the significant cleared-leaf atlas series by Klucking (1986–2003) was manually scanned, cropped, and made into a dataset as part of a machine-learning study (Wilf et al. 2016); however, that dataset is not retained here due to comparatively low resolution, moiré patterns, and other artifacts of printing (Wilf et al. 2016). In addition, the University of California Berkeley collection of over 800 voucherized cleared leaves (distinct from the Axelrod collection) has not been included here because it has not yet been digitized.

A master inventory of the 26,176 images of cleared and x-rayed specimens from >4,500 genera and >17,300 extant species in 354 plant families (Table 1) is provided in the accompanying Figshare archive. A small number of specimens are represented by multiple images, such as close-ups or lighting variants. Taxonomic fields include the family, genus, and species as provided in the respective collection catalog, with additional fields for updated Angiosperm Phylogeny Group (APG) family and order (APG IV 2016). Taxonomic and geographic coverage are uneven, constrained by the general availability of herbarium materials to the creators of the collections (similar issues occur even in recent, large herbarium-image datasets; e.g., de Lutio et al. 2021). Eight families are represented by more than 1,000 images each (Fabaceae, Sapindaceae, Rosaceae, Fagaceae, Annonaceae, Rubiaceae, Ulmaceae, and Malvaceae), whereas 173 families have fewer than ten images apiece. Photographs were taken by many people (see Acknowledgments) over an extended period of time and at different institutions, with a wide variety of cameras and methods that we do not attempt to detail; however, the original camera or scanner EXIF (Exchangeable Image File Format) metadata remain embedded in most of the images and are viewable in standard file browsers. We have also maintained the original pixel resolution and image dimensions in all photographs.

Catalog numbers of cleared or x-rayed leaves in the master inventory (available in the accompanying Figshare archive) refer to a unique glass slide (for the cleared leaves) or a film-negative number (for the x-rays) used to organize the respective collection,

as designated by the creator of the collection. The catalog numbers of the cleared and x-rayed leaf collections are usually secondary, i.e., specific to the collection but linked in museum records (as legacy data and thus without hyperlinks) to a primary source voucher at a herbarium (Table 1). Thus, the collection-specific secondary numbers are usually the information needed to search the specimens online (using resources listed in Table 1 and further described below) or in paper catalogs to locate the primary source-voucher data. In some cases there is no herbarium or other voucher besides the mounted slide, and then the curated cleared-leaf specimen, usually specially collected for the purpose, is a primary collection. We also provide in the accompanying Figshare archive a catalog file containing the voucher data for the Wing X-Ray Collection, for which data are not otherwise available online. In publications, specimens should be formally cited by primary voucher as well as secondary catalog number if possible (see Fig. 1).

Family and order updates were done iteratively by first doing automatic lookups to family of the catalog genera and species, using the tables provided in The Plant List (www.theplantlist.org) and its successor, World Flora Online (WFO; www.worldfloraonline.org; Borsch et al. 2020). These resources include standardized lookups to family and order for most generic and species names, modified slightly to include a few taxa not listed in WFO. Failed lookups were flagged and corrected manually. Most lookup failures resulted from typographical errors of generic and species names in the catalog data, and these were manually corrected. Others resulted from genera not being listed or having ambiguous or unverified family status in WFO; these taxa were then manually vetted using other standard resources such as Tropicos (www.tropicos.org), the International Plant Names Index (www.ipni.org), and the Angiosperm Phylogeny Website (www.mobot.org/MOBOT/research/APweb). For consistency, the WFO was designated as the priority lookup for conflicting results among taxonomic databases.

For reference, we note other online resources for batch-vetting plant names that we did not use, including Taxonomic Names Resolution Service (tnrs.iplantcollaborative.org), taxize (github.com/ropensci/taxize), and the Kew Vascular Plant Families and Genera database (data.kew.org/vpfg1992/vascplnt.html). In addition, an automated tool, the WORLDFLORA R package, is now available for batch lookups from the WFO taxonomic backbone file (Kindt 2020), although this would not have resolved the large number of taxa with uncertain status in WFO that required manual vetting.

Due to the intensive labor that would be required to update the large number of names below family level, even with the aid of batch services, and the emphasis here on family-level vetting, generic and species names were for the most part not updated except to correct misspellings that would hinder future lookups. A full vetting below family level would also require manually consulting and hyperlinking all the primary herbarium records to check for new determinations, a process of several years. However, any user can easily find taxa of interest using the specimen list provided (accompanying Figshare archive) and access updated nomenclature and voucher data using the resources listed.

The resulting master inventory of cleared and x-rayed leaves was manually inspected repeatedly to eliminate variant spellings and other inconsistencies, until no more

were found. Even after this stage, many issues remained from duplicate and corrupt files, invalid paths, labeling errors, ghost folders of problem images, and other common legacy database errors. Automated and reproducible data analysis and cleaning was done (by J. Rose and R. Saha) largely in Jupyter Notebooks and scripted in Python. In an iterative process, we used the Pandas library to load, sort, and filter the dataset in the form of a table, mapping metadata values in each column to unique specimens in each row. From there, we verified each file path's full compliance with a pair of requirements, namely that it be both (a) a unique absolute path, and (b) a valid path specifying an existing, uncorrupted image file that can be successfully opened and closed. Rows that failed this test were flagged and taken out for manual review.

Further file path cleaning included the use of a fuzzy matching algorithm, through which all possible matches between a flagged query file path q and a possible near-duplicate reference path f_R were compared by calculating the Levenshtein Distance (e.g., <https://xlinux.nist.gov/dads/HTML/Levenshtein.html>). This distance serves as a measure of the character-level similarity between two strings, from which all pairs are sorted in order of decreasing similarity to the flagged file q . Several duplicated source files that had evaded detection in previous stages were identified in this way, by manually scanning the top few most similar matches and searching for signs of typos. This procedure for automating the identification of the most likely near-duplicate strings allowed us to automatically verify that none of the tens of thousands of species in thousands of genera, hundreds of families, and dozens of orders included any artificial categories created by a misspelling. An example could be two samples from the same family, where one's family was spelled "Fabaceae" (correct), whereas the other was accidentally entered as "Fabeceae." This is an easy typo to miss, but it can skew downstream analyses.

Once all taxonomic and archival fields were validated, we assigned each sample a new filename that accomplishes both (a) directly encoding multiple levels of metadata into human-readable format within the filename, and (b) allowing easy sorting and searching of files on disk, without any additional alterations or struggling with a full relational database. The new filename format is constructed in the form: "Family_Genus_species_Collection_Catalog number". This user-friendly format facilitates, for the first time, rapid alphabetic sorting, visual inspection, and searching of all the merged images from multiple sources in standard personal-computer windows and visual browsers. In the filenames, as just described, the family is updated to APG standard according to World Flora Online and other resources, whereas the genus and species fields are usually not updated except to correct spelling errors, especially those that could cause lookup failures.

National cleared leaf collection – NCLC-W and NCLC-H

The National Cleared Leaf Collection is derived from parallel, broadly collaborative efforts supervised by the late Drs. Jack A. Wolfe (NCLC-W) and Leo J. Hickey (NCLC-H), beginning in the late 1960s. The NCLC is the world's largest and most phylogenetically comprehensive assembly of cleared, stained, and mounted leaves

sampled primarily from vouchered herbarium sheets. The collections underpinned the scientists' research on fossil leaves and leaf architecture, including their landmark evolutionary survey (Hickey and Wolfe 1975). Hickey and Wolfe (1975) reported that the clearing techniques they used were those of Foster (1952), as adapted by Hickey (1973); Dilcher (1974) further described the techniques of Hickey and Wolfe. More recent work has improved the methods for clearing and mounting leaves without deterioration and provided historical methods reviews (Vasco et al. 2014; García-Gutiérrez et al. 2020). The Wolfe and Hickey cleared-leaf collections, kept separately during the scientists' lifetimes and without any intention to merge them to our knowledge, are now curated together in the Division of Paleobotany, Department of Paleobiology, NMNH as the National Cleared Leaf Collection, constituting a monumental resource for leaf architecture that is combined here digitally for the first time. Physically, the two sub-collections are adjacent but not merged because Wolfe and Hickey used somewhat different family delimitations as they assembled their collections, and these are retained in the organization of the slides at NMNH (their systems were standardized and merged digitally for this contribution, as described earlier). The slides are organized alphabetically by family within each sub-collection.

The Wolfe contribution (NCLC-W) is the larger of the two parts, comprising over 18,000 specimens, from which 16,249 images are available here (Table 1). As described at the Cleared Leaf Image Database website (<http://clearedleavesdb.org>), the largest contributing source for NCLC-W was the University of California Herbarium (UC), Berkeley. Other significant sources were the California Academy of Sciences (CAS; including the Dudley Herbarium, DS, formerly of Stanford University), the Herbarium of the Arnold Arboretum (A) of the Harvard University Herbaria, the Missouri Botanical Garden (MO), the New York Botanical Garden (NY), the Field Museum of Natural History (F), and the National Herbarium of the Smithsonian Institution (US). Wolfe kept his collection for many years as a core reference for his voluminous body of work on fossil angiosperm leaves (see Upchurch et al. 2007), first at the United States Geological Survey (USGS) in Menlo Park, then at USGS Denver. Various photographic projects to document the collection advanced during the 1980s and 1990s, though none of these was published.

Following Dr. Wolfe's retirement in 1992, S. Wing supervised the moving and curation of the cleared-leaf collection from Denver to NMNH, as well as, after Dr. Wolfe's passing in 2005, a small portion of the collection that Wolfe had kept in his emeritus position at the University of Arizona. The collection was re-assembled, loaded into NMNH cabinetry, partially repaired, photographed, and placed under curation in the Division of Paleobotany, Department of Paleobiology, NMNH, officially as NCLC-W. A registry kept on paper by Dr. Wolfe and his team, containing the herbarium voucher data for all slides, is also kept with the collection; the registry was professionally transcribed into a digital format, then updated and corrected by E. González-Akre and several other Smithsonian staff members and volunteers. The photographs used in this contribution were made by another large group of Smithsonian staff and

volunteers (see Acknowledgments). Most slides have approximately the same physical dimensions, although some are oversize to accommodate large leaves; scale bars are included on most photographs (e.g., Fig. 1A). The photographs and collections data for NCLC-W were separately archived several years ago in the Cleared Leaves Image Database (<http://clearedleavesdb.org>; Das et al. 2014), also under open-access but at lower resolution than we provide here. We refer the reader to that useful platform to look up primary specimen metadata online using Wolfe's (secondary; Table 1) catalog numbers, including the herbarium-voucher data. Exact nomenclature may vary from what is presented here, following our separate vetting process.

The NCLC-W has been used extensively as a reference library, especially by paleobotanists; one notable example is its service as a principal reference for identifying leaf fossils from the oldest Neotropical paleorainforests, the Paleocene Cerrejón and Bogotá formation floras of Colombia (Herrera et al. 2008; Wing et al. 2009; Carvalho et al. 2021a, 2021b). Many images from NCLC-W (and NCLC-H) were used to illustrate leaf characters in the Manual of Leaf Architecture (Ellis et al. 2009), and the collection was used in a study of leaf rank and areole size (Green et al. 2014). A selection of more than 5,000 NCLC-W images was used for training and testing for family recognition as part of a machine-learning study that also included computer-marked heat maps, showing diagnostic regions for machine identification (Wilf et al. 2016).

Professor Leo J. Hickey supervised the assembly of a parallel cleared-leaf collection to Wolfe's during his time as curator of paleobotany at NMNH (Wing et al. 2014), comprising more than 7,000 slides, from which 6,861 images are included here (Table 1). Dr. Hickey made a successful effort to sample complementary taxa to Dr. Wolfe, thus increasing the combined diversity of their collections considerably (Table 1). Hickey targeted a larger number of herbaceous taxa, partly reflecting his interest in herbaceous early angiosperms (e.g., Taylor and Hickey 1992). Nearly all specimens were sampled at US, with minor contributions from MO, NY, and several other herbaria, along with a small amount of freshly sampled or fluid-preserved material. Dr. Hickey borrowed the collection that he made when he relocated to the Yale Peabody Museum of Natural History (YPM) in 1982. Web access to images of NCLC-H and additional information about the collection are still provided by the Yale Peabody Museum (<https://collections.peabody.yale.edu/pb/nclc/>), where slides were imaged and inventoried by a large team (see Acknowledgments) under the direction of Drs. Hickey and S. Hu. The same photographs are aggregated here as summarized in the master inventory (available in the accompanying Figshare archive), and full metadata and source voucher information for each slide are available at <https://collections.peabody.yale.edu/pb/nclc> and from YPM staff. In NCLC-H, primary herbarium-voucher data are usually visible in the photographs on labels that were mounted with the leaves. The physical size of the slides varies, and scale bars are included on most photographs. Following Dr. Hickey's passing in 2013, NCLC-H was returned to NMNH, where it is now curated in the Division of Paleobotany, Department of Paleobiology, adjacent to NCLC-W as just mentioned.

Axelrod cleared leaf collection

The Daniel I. Axelrod Cleared Leaf Collection at UCMP includes about 1,300 specimens that are in exceptionally good condition, compared with the NCLC, because the late Dr. Axelrod (Barbour et al. 1998) mounted them in plexiglass with a medium, possibly clear epoxy, that has remained clear for over 50 years. The slides mostly represent the California flora. They are a self-standing primary collection not linked to herbarium vouchers, and only general locality data are given on the slide labels, but nevertheless the material comprises a well-curated museum collection with good preservation and high image quality in the photographs. The UCMP has provided the Axelrod images online for many years through several portals linked from the UCMP Cleared Leaf Collection web page (<https://ucmp.berkeley.edu/collections/paleobotany-collection/ucmp-cleared-leaf-collection>). Scale bars are included in all photographs, of which 832 are used here (Table 1). A selection of images from the Axelrod collection was used for training and testing of automatic leaf recognition in the Wilf et al. (2016) machine-learning study.

Wing X-ray collection

In the early 1990s, S. Wing developed an x-ray scanning technique (Wing 1992) and used it to capture leaf and other organ images of selected families on large-format (8 by 10 inches, or 20.3 by 25.4 cm) x-ray negatives. The specimens are mostly from US, along with a variety of other herbaria and living collections; the negatives are now archived in the Division of Paleobotany, Department of Paleobiology, NMNH, and a separate data item is made available in the accompanying Figshare archive that matches the negative numbers (preserved in the current filenames) with their vouchers. The 1200-dpi cropped scans of the negatives by S. Wing are made available here digitally for the first time. Although the images lack embedded scales, the direct contact method of imaging with x-rays means that the images on physical negatives are the same size as the original specimens, and the negatives were scanned 1:1 as well. Thus, measurements can be made directly from the images or calibrated, if needed, using the post-crop image dimensions in the image metadata. Grayscale values of the scanned negatives were batch-inverted to positive here (easily reversed with a second inversion), to provide light backgrounds and improve comparability with the other image sets. The reverse grayscale tends to accentuate the visual impact of large differences in exposure caused by variation in leaf density; however, we found that standard image level and contrast adjustments are sufficient to make fine details more visible when needed. The collection includes a sizable number of x-rays of reproductive organs, especially Sapindaceae fruits, which are retained here for their general interest.

Fossil leaves

We provide 4,076 vouchered leaf-fossil images of specimens that are assigned to family level, in total covering 44 angiosperm and four non-angiosperm families from a variety of sites in the Americas that are well known to the authors (Table 1; Appendix 1).

Although far from comprehensive, this image set nevertheless covers at least a majority of angiosperm families that are reliably known in the fossil record from nearly-complete leaf remains; it provides a starter set both for comparative learning in angiosperm paleobotany and training machine-learning algorithms.

Unlike the images from the cleared and x-rayed collections, which were not adjusted except for cropping of the x-rays, the fossil-leaf images were all manually and reversibly rotated, close-cropped, and contrast- and temperature-adjusted (all whole-image adjustments, other than cropping) in Adobe Camera Raw so that they are approximately similar in relative frame alignment and overall contrast, with emphasis on making vein features visible (for some photographs taken on early-model digital cameras with barrel distortion in macro mode, the lens distortion was corrected manually using Adobe Camera Raw). This procedure minimizes strong distractors such as rock matrix for machine learning of fossil leaves, an interest of several of the authors (Wilf et al. 2016), and we found that it also enhances human learning for the same reason, by increasing visual comparability of the leaf features and eliminating distractors and variable orientation. In all cases, we have maintained the full pixel resolution and (post-crop) dimensions of the original image and resaved processed images from Camera Raw to jpeg format only once (usually with tiff format as a lossless intermediate step), using the minimum compression ratio to maintain image quality. A cost of this approach was removal of most of the scale bars. However, nearly all scaling information can be accessed if needed from online (usually much lower resolution but suitable for scaling) versions of the images or sets of uncropped originals that we have included where necessary (see General Collection, below). In addition, all physical voucher specimens can be accessed at their respective repositories. As for the cleared and x-rayed leaves, original camera or scanner EXIF data remain embedded in the image metadata.

The fossil set of 4,076 images is comprised of two parts (Table 1, Appendix 1): first, a concentrated collection of 3,320 images from a single prolific site, the late Eocene Florissant fossil beds of Colorado; and second, a smaller general collection of 756 images from a variety of latest Cretaceous and Paleogene sites in North and South America (Appendix 1; accompanying Figshare archive). Appendix 1 annotates and lists authorities and taxonomic references for the ca. 222 species used in the fossil dataset, and individual catalog numbers are embedded in the filenames of the images. Appendix 1 also lists references for site-specific collections that pertain directly to the specimens used here, if the latter are different from the taxonomic references. Some specimens are represented by multiple images, such as close-ups or lighting variations (but not by duplicate images), and many images of counterparts are included. Although the major target for the collection was “dicot” leaves, images of a few species of monocots, ferns, and conifers that were readily available were included to help seed future expansions. Several generic names that may be botanically doubtful are left in as-published or as-cataloged form (Appendix 1), but all included material is considered reliably placed at family level. Informal morphotypes are included if they have reliable features at family level. Filenames, as for the cleared and x-rayed leaves, embed taxonomy to enable rapid auto-sorting and searching in standard PC windows, followed by collection data.

Florissant collection

The late Eocene Florissant Fossil Beds Lagerstätte of Colorado is known worldwide for its long history of collection and investigation, its outstanding diversity of plant and animal fossils, and its seminal role in the conservation movement (MacGinitie 1953; Evanoff et al. 2001; Meyer 2003; Leopold et al. 2008; Veatch and Meyer 2008; Leopold and Meyer 2012). Florissant's diverse fossil flora has a long history of study, resulting in an exceptional level of taxonomic understanding (e.g., Lesquereux 1873, 1883; MacGinitie 1953; Manchester and Crane 1983; 1987; Manchester 1989a, 2001a; Jia and Manchester 2014; Herendeen and Herrera 2019). The late Harry D. MacGinitie's (1953) landmark monograph of the Florissant flora was outstanding among comparable works of the time for the high quality and botanical accuracy of his descriptions and identifications (Manchester 2001a).

Among its many distinctions, the Florissant biota was one of the first large fossil assemblages of any kind to be photographed, cataloged comprehensively, and made openly available in an internet database (Meyer et al. 2008). This massive effort by H. Meyer and associates, beginning in the 1990s, has two components as described below. The Florissant images were manually filtered and prepared by X. Zou from an initial set of 13,691 images of plant, animal, and geological specimens, of which 7,798 are of plants and 6,122 are of leaves, from which we further filtered and prepared the 3,320 images used here of leaf specimens that can be confidently placed in a plant family (Appendix 1). Vetting to plant family followed Manchester (2001a) and other publications as listed in Appendix 1.

The first of two components of the Florissant image set (Table 1) comes from the Meyer et al. (2008) project to capture high-resolution images of all type, published, and related Florissant collections, representing 5,663 specimens of ca. 1800 fossil plant and animal species as described in >300 scientific papers from a total Florissant collection of ca. 50,000 specimens. Much of this material had never been illustrated or was illustrated poorly by modern standards. The fossils are held in about 15 museums around the world as listed by Meyer et al. (2008); the largest three Florissant type and published collections are at the Smithsonian National Museum of Natural History, the Harvard University Museum of Comparative Zoology (almost entirely insects and spiders), and the University of Colorado Museum of Natural History. The original photographs on Kodachrome slides are archived at Florissant Fossil Beds National Monument (FLFO), and they were scanned twice about 12–15 years apart to take advantage of improving technology, the second time at high resolution. The resulting image database of the more recent scans (Meyer et al. 2008) was hosted on a National Park Service server initially and then moved several years ago to the University of Colorado Museum of Natural History, where it can be searched online using the Florissant Fossil Beds Collection Search at <https://flfo-search.colorado.edu>. That website provides full specimen metadata and reduced-resolution image files (with scale bars), which can be searched using the secondary inventory (photograph) numbers from the Meyer et al. (2008) project that are here embedded in the filenames (Table 1). To help distinguish images in this collection from the others rapidly in searches, we have also

attached an informal “CU” prefix (for the University of Colorado) to the secondary catalog numbers in the filenames.

The second component of the Florissant image collection provided here (Table 1) is a selection of fully digital images from the collections at Florissant Fossil Beds National Monument (primary acronym FLFO, specimen number embedded in the filename), assembled by H. Meyer and numerous interns and assistants. The images can also be searched and viewed (with scale bars but at lower resolution) by FLFO number on the park’s flickr page, located at <https://www.flickr.com/photos/155340198@N06>. The corresponding, full-resolution, uncropped images that were processed here from both Florissant image sets are archived at the University of Colorado Museum of Natural History and FLFO, respectively, and available on request to collections management.

General collection

The general collection of 756 fossil leaf images provided here (Appendix 1; specimen data in the accompanying Figshare archive) draws from a set of Late Cretaceous to Eocene fossil floras from the Americas. The general collection diversifies the phylogenetic, preservational, temporal, and geographic coverage of the overall fossil-image dataset and forms a base to encourage other teams to make similar efforts. Repositories of the material are indicated in the filenames and supplemental data files in the accompanying Figshare archive; they include the Denver Museum of Nature & Science (repository acronym DMNH); Museo Paleontológico Egidio Feruglio (MPEF-Pb, Trelew, Argentina); National Museum of Natural History, Smithsonian Institution (USNM-PAL, abbreviated here as USNM, Washington, D.C.); Colombian Geological Survey and Colombian Petroleum Institute (combined as SGC-ICP, Bogotá, Colombia); Florida Museum of Natural History (UF, Gainesville); and University of California Museum of Paleontology (UCMP, Berkeley). Filenames embed the taxonomy as well as primary repository numbers or unique field numbers, if a formal repository number is not assigned (some MPEF, USNM specimens). In a few cases, where more than one image of the same fossil is included (i.e., close-ups or unlabeled parts and counterparts), an informal tag is included in the filename in brackets to ensure uniqueness of filenames. Some material lacks formal taxonomy but is considered reliably identified at family level; there are also a few cases of historic generic identifications considered incorrect and indicated by quotations (e.g., “*Acer*,” “*Ficus*”) but still assignable to a (often different) family (Appendix 1). Because very few fossils from the general collection are otherwise viewable online to check scale bars, we provide a parallel folder of the corresponding uncropped images in the accompanying Figshare archive.

Major contributions to the general collection are briefly listed here for paleobotanists, with additional taxonomic and occurrence references listed in Appendix 1. The fossils come from (1) a suite of latest Cretaceous (late Maastrichtian, Hell Creek Formation) and early Paleocene (early Danian, Fort Union Formation) sites from western North Dakota and South Dakota USA that have been used extensively for studies of the end-Cretaceous extinction (e.g., Johnson et al. 1989; Johnson 2002; Wilf and

Johnson 2004); (2) the early Paleocene Salamanca Formation (early Danian) and Las Flores (late Danian) floras of Chubut, Argentina, known for diverse and well-preserved fossil plants and insect-feeding damage following the end-Cretaceous extinction (e.g., Iglesias et al. 2007, 2021; Clyde et al. 2014; Donovan et al. 2017; Stiles et al. 2020); (3) the early Paleocene (Danian, Fort Union Formation) Mexican Hat site in southeastern Montana, USA, known for diverse insect herbivory traces preserved in its fossil leaves (Wilf et al. 2006; Winkler et al. 2010; Donovan et al. 2014); (4) the middle-late Paleocene (Selandian-Thanetian, Cerrejón Formation) Cerrejón flora from the Guajira Peninsula, Colombia and Bogotá Formation flora of Sabana de Bogotá, central Colombia, together preserving the remains of the oldest known Neotropical rainforests (e.g., Doria et al. 2008; Herrera et al. 2008, 2019; Gómez-Navarro et al. 2009; Wing et al. 2009; Carvalho et al. 2011, 2021a, 2021b); (5) a suite of sites spanning the late Paleocene (Fort Union Formation) through early Eocene (Wasatch Formation and Little Mountain locality of the Green River Formation) of southwestern and northwestern Wyoming that have been used in many studies of floristic and plant-insect associational responses to climate change (e.g., Gemmill and Johnson 1997; Wilf et al. 1998, 2006; Wilf and Labandeira 1999; Wilf 2000; Donovan et al. 2014); (6) the early Eocene Laguna del Hunco Lagerstätte in Chubut, Argentina (Huitrera Formation), known for its outstanding diversity of fossil plants and animals, varied biogeographic connections, and large number of unique taxon occurrences for South America (e.g., Wilf et al. 2003, 2013, 2019; Gandolfo et al. 2011); (7) the late early Eocene flora of Republic, Washington (Wolfe and Wehr 1987; DeVore et al. 2005; Greenwood et al. 2016; Klondike Mountain Formation) and the middle Eocene Green River Formation flora (MacGinitie 1969; Smith et al. 2008) of Bonanza, Utah, specifically using images of field-censused collections at DMNH from both sites led by K. Johnson that were used previously for analyses of insect herbivory, fossil-leaf economics, and digital leaf physiognomy (Wilf et al. 2001, 2005b; Cariglino 2007; Royer et al. 2007; Peppe et al. 2011).

Concluding remarks

The dataset presented here consolidates thousands of hours of labor by many people (see Acknowledgments) into a single accessible platform. Due to the extraordinary effort involved, it is unlikely that many new, large-scale cleared and x-rayed leaf collections will ever be assembled and digitally processed. Thus, the future prospects for significantly increasing the overall sample size and improving the coverage of taxonomy and geography in digital leaf-reference collections most likely lie elsewhere. The greatest potential appears to come from the advancing techniques for segmenting and enhancing leaf images from the enormous, widely available resource of digitized herbarium sheets (Hussein et al. 2020; Weaver et al. 2020), which have the significant additional advantage of direct linkage to the global data infrastructure for biodiversity (e.g., Bakker et al. 2020). To reach comparability with cleared leaves, segmented leaf

images will require high pixel resolution, optimized contrast for the capture of venation details, and the careful retention of significant edge features such as the leaf margin. For leaf fossils, increasing the sample size of well-identified specimens is straightforward in principle but will require efforts far beyond the resources of a single collaboration. Thus, we plan a community initiative for this purpose.

We look forward to seeing the assembled image dataset catalyze advances in research, education, and outreach. The images and supporting data are available open-access on Figshare Plus at <https://doi.org/10.25452/figshare.plus.14980698>. Some mistakes are inevitable in a first-version database of this nature; please report any errors observed to the corresponding author. Corrections and updates may be applied to the Figshare archive under new version numbers; the version precisely corresponding to this article will remain preserved as version 1.0.

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References

APG IV (2016) An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. *Botanical Journal of the Linnean Society* 181(1): 1–20. <https://doi.org/10.1111/boj.12385>

Axelrod DI (1986) Cenozoic history of some western American pines. *Annals of the Missouri Botanical Garden* 73(3): 565–641. <https://doi.org/10.2307/2399194>

Bakker FT, Antonelli A, Clarke JA, Cook JA, Edwards SV, Ericson PGP, Faurby S, Ferrand N, Gelang M, Gillespie RG, Irestedt M, Lundin K, Larsson E, Matos-Maraví P, Müller J, von Proschwitz T, Roderick GK, Schliep A, Wahlberg N, Wiedenhoeft J, Källersjö M (2020) The Global Museum: Natural history collections and the future of evolutionary science and public education. *PeerJ* 8: e8225. <https://doi.org/10.7717/peerj.8225>

Barbour MG, Doyle J, Sanderson M (1998) Daniel Axelrod, Biological Sciences: Davis, 1910–1998. Professor of Paleoecology, Emeritus. In: Krogh D (Ed.) University of California: In Memoriam, 1998. Academic Senate, University of California, Oakland, CA, 8–11. http://content.cdlib.org/view?docId=hb1p30039g&&doc.view=entire_text

Blonder B, Both S, Jodra M, Majalap N, Burslem D, Teh YA, Malhi Y (2019) Leaf venation networks of Bornean trees: Images and hand-traced segmentations. *Ecology* 100(11): e02844. <https://doi.org/10.1002/ecy.2844>

Bonnet P, Goëau H, Hang ST, Lasseck M, Šulc M, Malécot V, Jauzein P, Melet J-C, You C, Joly A (2018) Plant identification: experts vs. machines in the era of deep learning. In: Joly A, Vrochidis S, Karatzas K, Karppinen A, Bonnet P (Eds) *Multimedia Tools and Applications for Environmental & Biodiversity Informatics*. Springer, Cham, Switzerland, 131–149. https://doi.org/10.1007/978-3-319-76445-0_8

Borsch T, Berendsohn W, Dalcin E, Delmas M, Demissew S, Elliott A, Fritsch P, Fuchs A, Geltman D, Güner A, Haevermans T, Knapp S, le Roux MM, Loizeau P-A, Miller C, Miller J, Miller JT, Palese R, Paton A, Parnell J, Pendry C, Qin H-N, Sosa V, Sosef M, von Raab-Straube E, Ranwashe F, Raz L, Salimov R, Smets E, Thiers B, Thomas W, Tulig M, Ulate W, Ung V, Watson M, Jackson PW, Zamora N (2020) *World Flora Online: Placing*

taxonomists at the heart of a definitive and comprehensive global resource on the world's plants. *Taxon* 69(6): 1311–1341. <https://doi.org/10.1002/tax.12373>

Brea M, Zamuner AB, Matheos SD, Iglesias A, Zucol AF (2008) Fossil wood of the *Mimosoideae* from the early Paleocene of Patagonia, Argentina. *Alcheringa* 32(4): 427–441. <https://doi.org/10.1080/03115510802417695>

Call VB, Dilcher DL (1994) *Parvileguminophyllum coloradensis*, a new combination for *Mimosites coloradensis* Knowlton, Green River Formation of Utah and Colorado. Review of Palaeobotany and Palynology 80(3–4): 305–310. [https://doi.org/10.1016/0034-6667\(94\)90007-8](https://doi.org/10.1016/0034-6667(94)90007-8)

Cariglino B (2007) Paleoclimatic analysis of the Eocene Laguna del Hunco, Green River, and Republic floras using digital leaf physiognomy. MS Thesis, Pennsylvania State University.

Carpenter RJ, Wilf P, Conran JG, Cúneo NR (2014) A Paleogene trans-Antarctic distribution for *Ripogonum* (Ripigonaceae: Liliales)? *Palaeontologia Electronica* 17(3): 39A. <https://doi.org/10.26879/460>

Caranza-Rojas J, Joly A, Goëau H, Mata-Montero E, Bonnet P (2018) Automated identification of herbarium specimens at different taxonomic levels. In: Joly A, Vrochidis S, Karatzas K, Karppinen A, Bonnet P (Eds) *Multimedia Tools and Applications for Environmental & Biodiversity Informatics*. Springer, Cham, Switzerland, 151–167. https://doi.org/10.1007/978-3-319-76445-0_9

Carvalho MR, Herrera FA, Jaramillo CA, Wing SL, Callejas R (2011) Paleocene Malvaceae from northern South America and their biogeographical implications. *American Journal of Botany* 98(8): 1337–1355. <https://doi.org/10.3732/ajb.1000539>

Carvalho M, Herrera F, Gómez S, Martínez C, Jaramillo C (2021a) Early records of Melastomataceae from the middle–late Paleocene rainforests of South America conflict with Laurasian origins. *International Journal of Plant Sciences* 182(5): 401–412. <https://doi.org/10.1086/714053>

Carvalho MR, Jaramillo C, de la Parra F, Caballero-Rodríguez D, Herrera F, Wing SL, Turner BL, D'Apolito C, Romero-Báez M, Narváez P, Martínez C, Gutierrez M, Labandeira CC, Bayona G, Rueda M, Paez-Reyes M, Cárdenas D, Duque Á, Crowley JL, Santos C, Silvestro D (2021b) Extinction at the end-Cretaceous and the origin of modern Neotropical rainforests. *Science* 372(6537): 63–68. <https://doi.org/10.1126/science.abf1969>

Christophel DC, Hyland BPM (1993) Leaf Atlas of Australian Tropical Rain Forest trees. CSIRO, Melbourne, 260 pp.

Christophel DC, Rowett AI (1996) Leaf and cuticle atlas of Australian leafy Lauraceae. *Flora of Australia Supplementary Series* 6: 1–217.

Clyde WC, Wilf P, Iglesias A, Slingerland RL, Barnum T, Bijl PK, Bralower TJ, Brinkhuis H, Comer EE, Huber BT, Ibañez-Mejia M, Jicha BR, Krause JM, Schueth JD, Singer BS, Raigemborn MS, Schmitz MD, Sluijs A, Zamalloa MC (2014) New age constraints for the Salamanca Formation and lower Río Chico Group in the western San Jorge Basin, Patagonia, Argentina: Implications for K/Pg extinction recovery and land mammal age correlations. *Geological Society of America Bulletin* 126(2–4): 289–306. <https://doi.org/10.1130/B30915.1>

Cockerell TDA (1908) The fossil flora of Florissant, Colorado. *Bulletin of the American Museum of Natural History* 24: 71–109. <http://hdl.handle.net/2246/914>

Crane PR, Manchester SR, Dilcher DL (1991) Reproductive and vegetative structure of *Nordenskioldia* (Trochodendraceae), a vesselless dicotyledon from the Early Tertiary of the Northern Hemisphere. *American Journal of Botany* 78(10): 1311–1334. <https://doi.org/10.1002/j.1537-2197.1991.tb12599.x>

da Ribeiro JELdS, Hopkins MJG, Vicentini A, Sothers CA, Costa MAdS, de Brito JM, de Souza MAD, Martins LHP, Lohmann LG, Assunção PACL, Pereira EdC, da Silva CF, Mesquita MR, Procópio LC (1999) Flora da Reserva Ducke. Instituto Nacional de Pesquisas da Amazônia (INPA), Manaus, 671 pp. https://ppbio.inpa.gov.br/sites/default/files/flora_da_ducke.rar

Das A, Bucksch A, Price CA, Weitz JS (2014) ClearedLeavesDB: An online database of cleared plant leaf images. *Plant Methods* 10(1): e8. <https://doi.org/10.1186/1746-4811-10-8>

de Lutio R, Little D, Ambrose B, Belongie S (2021) The Herbarium 2021 Half-Earth Challenge Dataset. arXiv preprint 2105.13808. <https://arxiv.org/abs/2105.13808>

Denk T, Dillhoff RM (2005) *Ulmus* leaves and fruits from the Early–Middle Eocene of northwestern North America: Systematics and implications for character evolution within Ulmaceae. *Canadian Journal of Botany* 83(12): 1663–1681. <https://doi.org/10.1139/b05-122>

DeVore ML, Pigg KB (2007) A brief review of the fossil history of the family Rosaceae with a focus on the Eocene Okanogan Highlands of eastern Washington State, USA, and British Columbia, Canada. *Plant Systematics and Evolution* 266(1–2): 45–57. <https://doi.org/10.1007/s00606-007-0540-3>

DeVore ML, Pigg KB, Wehr WC (2005) Systematics and phytogeography of selected Eocene Okanagan Highlands plants. *Canadian Journal of Earth Sciences* 42(2): 205–214. <https://doi.org/10.1139/e04-072>

Dilcher DL (1971) A revision of the Eocene flora of southeastern North America. *Palaeobotanist* 20(1): 7–18.

Dilcher DL (1974) Approaches to the identification of angiosperm leaf remains. *Botanical Review* 40(1): 1–157. <https://doi.org/10.1007/BF02860067>

Donovan MP, Wilf P, Labandeira CC, Johnson KR, Peppe DJ (2014) Novel insect leaf-mining after the end-Cretaceous extinction and the demise of Cretaceous leaf miners, Great Plains, USA. *PLoS ONE* 9(7): e103542. <https://doi.org/10.1371/journal.pone.0103542>

Donovan MP, Iglesias A, Wilf P, Labandeira CC, Cúneo NR (2017) Rapid recovery of Patagonian plant-insect associations after the end-Cretaceous extinction. *Nature Ecology & Evolution* 1: e0012. <https://doi.org/10.1038/s41559-016-0012>

Doria G, Jaramillo CA, Herrera F (2008) Menispermaceae from the Cerrejón Formation, middle to late Paleocene, Colombia. *American Journal of Botany* 95(8): 954–973. <https://doi.org/10.3732/ajb.2007216>

Doweld AB (2016) Nomenclatural novelties for the Palaeocene plants of North America. *Phytotaxa* 273(3): 191–199. <https://doi.org/10.11646/phytotaxa.273.3.6>

Doyle JA (2007) Systematic value and evolution of leaf architecture across the angiosperms in light of molecular phylogenetic analyses. *Courier Forschungsinstitut Senckenberg* 258: 21–37.

Ellis B, Daly DC, Hickey LJ, Johnson KR, Mitchell JD, Wilf P, Wing SL (2009) Manual of Leaf Architecture. Cornell University Press, Ithaca, New York, 200 pp. <https://repository.si.edu/handle/10088/93918>

Evanoff E, McIntosh WC, Murphey PC (2001) Stratigraphic summary and $^{40}\text{Ar}/^{39}\text{Ar}$ geochronology of the Florissant Formation, Colorado. Proceedings of the Denver Museum of Nature & Science, Series 4 1: 1–16.

Flynn S, DeVore ML, Pigg KB (2019) Morphological features of sumac leaves (*Rhus*, Anacardiaceae), from the latest early Eocene flora of Republic, Washington. International Journal of Plant Sciences 180(6): 464–478. <https://doi.org/10.1086/703526>

Foster AS (1952) Foliar venation in angiosperms from an ontogenetic standpoint. American Journal of Botany 39(10): 752–766. <https://doi.org/10.1002/j.1537-2197.1952.tb13099.x>

Fuller DQ, Hickey LJ (2005) Systematics and leaf architecture of the Gunneraceae. Botanical Review 71(3): 295–353. [https://doi.org/10.1663/0006-8101\(2005\)071\[0295:SALAOT\]2.0.CO;2](https://doi.org/10.1663/0006-8101(2005)071[0295:SALAOT]2.0.CO;2)

Gandolfo MA, Romero EJ (1992) Leaf morphology and a key to species of *Nothofagus* Bl. Bulletin of the Torrey Botanical Club 119(2): 152–166. <https://doi.org/10.2307/2997028>

Gandolfo MA, Dibbern MC, Romero EJ (1988) *Akania patagonica* n. sp. and additional material on *Akania americana* Romero & Hickey (Akanaceae), from Paleocene sediments of Patagonia. Bulletin of the Torrey Botanical Club 115(2): 83–88. <https://doi.org/10.2307/2996138>

Gandolfo MA, Hermsen EJ, Zamaloa MC, Nixon KC, González CC, Wilf P, Cúneo NR, Johnson KR (2011) Oldest known *Eucalyptus* macrofossils are from South America. PLoS ONE 6(6): e21084. <https://doi.org/10.1371/journal.pone.0021084>

García-Gutiérrez E, Ortega-Escalona F, Angeles G (2020) A novel, rapid technique for clearing leaf tissues. Applications in Plant Sciences 8(9): e11391. <https://doi.org/10.1002/aps3.11391>

Gemmill CEC, Johnson KR (1997) Paleoecology of a late Paleocene (Tiffanian) megaflora from the northern Great Divide Basin, Wyoming. Palaios 12(5): 439–448. <https://doi.org/10.2307/3515382>

Gentry AH (1993) A Field Guide to the Families and Genera of Woody Plants of Northwest South America (Colombia, Ecuador, Peru). Conservation International, Washington, D.C., 895 pp.

Gómez-Navarro C, Jaramillo C, Herrera F, Wing SL, Callejas R (2009) Palms (Arecaceae) from a Paleocene rainforest of northern Colombia. American Journal of Botany 96(7): 1300–1312. <https://doi.org/10.3732/ajb.0800378>

González CC, Gandolfo MA, Zamaloa MC, Cúneo NR, Wilf P, Johnson KR (2007) Revision of the Proteaceae macrofossil record from Patagonia, Argentina. Botanical Review 73(3): 235–266. [https://doi.org/10.1663/0006-8101\(2007\)73\[235:ROTPMR\]2.0.CO;2](https://doi.org/10.1663/0006-8101(2007)73[235:ROTPMR]2.0.CO;2)

Green WA, Little SA, Price CA, Wing SL, Smith SY, Kotrc B, Doria G (2014) Reading the leaves: A comparison of leaf rank and automated areole measurement for quantifying aspects of leaf venation. Applications in Plant Sciences 2(8): e1400006. <https://doi.org/10.3732/apps.1400006>

Greenwood DR, Pigg KB, Basinger JF, DeVore ML (2016) A review of paleobotanical studies of the Early Eocene Okanagan (Okanagan) Highlands floras of British Columbia, Canada, and Washington, USA. Canadian Journal of Earth Sciences 53(6): 548–564. <https://doi.org/10.1139/cjes-2015-0177>

Herendeen PS, Herrera F (2019) Eocene fossil legume leaves referable to the extant genus *Arcoa* (Caesalpinoideae, Leguminosae). International Journal of Plant Sciences 180(3): 220–231. <https://doi.org/10.1086/701468>

Hermsen EJ (2013) A review of the fossil record of the genus *Itea* (Iteaceae, Saxifragales) with comments on its historical biogeography. Botanical Review 79(1): 1–47. <https://doi.org/10.1007/s12229-012-9114-3>

Hermsen EJ, Gandolfo MA, Zamaloa MC (2012) The fossil record of *Eucalyptus* in Patagonia. American Journal of Botany 99(8): 1356–1374. <https://doi.org/10.3732/ajb.1200025>

Herrera F, Jaramillo CA, Dilcher DL, Wing SL, Gómez-Navarro C (2008) Fossil Araceae from a Paleocene Neotropical rainforest in Colombia. American Journal of Botany 95(12): 1569–1583. <https://doi.org/10.3732/ajb.0800172>

Herrera F, Carvalho MR, Wing SL, Jaramillo C, Herendeen PS (2019) Middle to Late Paleocene Leguminosae fruits and leaves from Colombia. Australian Systematic Botany 32(5–6): 385–408. <https://doi.org/10.1071/SB19001>

Hickey LJ (1973) Classification of the architecture of dicotyledonous leaves. American Journal of Botany 60(1): 17–33. <https://doi.org/10.1002/j.1537-2197.1973.tb10192.x>

Hickey LJ (1977) Stratigraphy and paleobotany of the Golden Valley Formation (Early Tertiary) of western North Dakota. Geological Society of America 150: 1–183.

Hickey LJ (1979) A revised classification of the architecture of dicotyledonous leaves. In: Metcalfe CR, Chalk L (Eds) Anatomy of the Dicotyledons, Vol I. Clarendon, Oxford, 25–39.

Hickey LJ, Wolfe JA (1975) The bases of angiosperm phylogeny: Vegetative morphology. Annals of the Missouri Botanical Garden 62(3): 538–589. <https://doi.org/10.2307/2395267>

Hill RS (1982) The Eocene megafossil flora of Nerriga, New South Wales, Australia. Palaeontographica. Abteilung B, Paläophytologie 181(1–3): 44–77.

Hussein BR, Malik OA, Ong W-H, Slik JWF (2020) Semantic segmentation of herbarium specimens using deep learning techniques. In: Alfred R, Lim Y, Haviluddin H, On CK (Eds) Computational Science and Technology Lecture Notes in Electrical Engineering, vol 603. Springer, Singapore, 321–330. https://doi.org/10.1007/978-981-15-0058-9_31

Hussein BR, Malik OA, Ong W-H, Slik JWF (2021) Automated extraction of phenotypic leaf traits of individual intact herbarium leaves from herbarium specimen images using deep learning based semantic segmentation. Sensors (Basel) 21(13): e4549. <https://doi.org/10.3390/s21134549>

Iglesias A, Wilf P, Johnson KR, Zamuner AB, Cúneo NR, Matheos SD, Singer BS (2007) A Paleocene lowland macroflora from Patagonia reveals significantly greater richness than North American analogs. Geology 35(10): 947–950. <https://doi.org/10.1130/G23889A.1>

Iglesias A, Wilf P, Stiles E, Wilf R (2021) Patagonia's diverse but homogeneous early Paleocene forests: Angiosperm leaves from the Danian Salamanca and Peñas Coloradas formations, San Jorge Basin, Chubut, Argentina. Palaeontologia Electronica 24(1): a02. <https://doi.org/10.26879/1124>

Jia H, Manchester SR (2014) Fossil leaves and fruits of *Cercis* L. (Leguminosae) from the Eocene of western North America. International Journal of Plant Sciences 175(5): 601–612. <https://doi.org/10.1086/675693>

Johnson KR (1996) Description of seven common plant megafossils from the Hell Creek Formation (Late Cretaceous: Late Maastrichtian), North Dakota, South Dakota, and Montana. *Proceedings of the Denver Museum of Natural History*, series 3, 3: 1–48.

Johnson KR (2002) The megaflora of the Hell Creek and lower Fort Union formations in the western Dakotas: vegetational response to climate change, the Cretaceous-Tertiary boundary event, and rapid marine transgression. *Geological Society of America Special Papers* 361: 329–391. <https://doi.org/10.1130/0-8137-2361-2.329>

Johnson KR, Plumb C (1995) Common plant fossils from the Green River Formation at Douglas Pass, Colorado, and Bonanza, Utah. In: Averett WR (Ed.) *The Green River Formation in Piceance Creek and Eastern Uinta basins*. Grand Junction Geological Society, Grand Junction, Colorado, 121–130. http://archives.datapages.com/data/grand-junction-geo-soc/data/013/013001/121_gjgs-sp0130121.htm

Johnson KR, Nichols DJ, Attrep Jr M, Orth CJ (1989) High-resolution leaf-fossil record spanning the Cretaceous-Tertiary boundary. *Nature* 340(6236): 708–711. <https://doi.org/10.1038/340708a0>

Jones JH (1986) Evolution of the Fagaceae: The implications of foliar features. *Annals of the Missouri Botanical Garden* 73(2): 228–275. <https://doi.org/10.2307/2399112>

Jud NA, Gandolfo MA, Iglesias A, Wilf P (2017) Flowering after disaster: Early Danian buckthorn (Rhamnaceae) flowers and leaves from Patagonia. *PLoS ONE* 12(5): e0176164. <https://doi.org/10.1371/journal.pone.0176164>

Jud NA, Iglesias A, Wilf P, Gandolfo MA (2018) Fossil moonseeds from the Paleogene of West Gondwana (Patagonia, Argentina). *American Journal of Botany* 105(5): 927–942. <https://doi.org/10.1002/ajb2.1092>

Jud NA, Allen SE, Nelson CW, Bastos CL, Chery JG (2021) Climbing since the early Miocene: The fossil record of Paullinieae (Sapindaceae). *PLoS ONE* 16(4): e0248369. <https://doi.org/10.1371/journal.pone.0248369>

Keller R (2004) *Identification of Tropical Woody Plants in the Absence of Flowers: a Field Guide*. Birkhäuser Verlag, Basel, Switzerland, 229 pp. <https://doi.org/10.1007/978-3-0348-7905-7>

Kellner A, Benner M, Walther H, Kunzmann L, Wissemann V, Ritz CM (2012) Leaf architecture of extant species of *Rosa* L. and the Paleogene species *Rosa lignitum* Heer (Rosaceae). *International Journal of Plant Sciences* 173(3): 239–250. <https://doi.org/10.1086/663965>

Kindt R (2020) WorldFlora: An R package for exact and fuzzy matching of plant names against the World Flora Online taxonomic backbone data. *Applications in Plant Sciences* 8(9): e11388. <https://doi.org/10.1002/aps3.11388>

Klucking EP (1986–2003) *Leaf Venation Patterns*, Vols. 1–9. J. Cramer, Berlin.

Knight CL, Wilf P (2013) Rare leaf fossils of Monimiaceae and Atherospermataceae (Laurales) from Eocene Patagonian rainforests and their biogeographic significance. *Palaeontologia Electronica* 16(3): e26A. <https://doi.org/10.26879/386>

Kumar N, Belhumeur PN, Biswas A, Jacobs DW, Kress WJ, Lopez IC, Soares JVB (2012) Leafsnap: a computer vision system for automatic plant species identification. In: Fitzgibbon A, Lazebnik S, Perona P, Sato Y, Schmid C (Eds) *Computer Vision – ECCV 2012 Lecture Notes in Computer Science*, vol. 7573. Springer, Berlin, 502–516. https://doi.org/10.1007/978-3-642-33709-3_36

Leopold EB, Meyer HW (2012) Saved in Time: the Fight to Establish Florissant Fossil Beds National Monument, Colorado. University of New Mexico Press, Albuquerque, New Mexico, 168 pp.

Leopold EB, Manchester SR, Meyer HW (2008) Phytogeography of the late Eocene Florissant flora reconsidered. Geological Society of America Special Papers 435: 53–70. [https://doi.org/10.1130/2008.2435\(04\)](https://doi.org/10.1130/2008.2435(04))

Lesquereux L (1873) Lignitic formation and fossil flora. Sixth Annual Report of the US Geological Survey of the Territories: 317–427. <https://doi.org/10.3133/70038930>

Lesquereux L (1883) Contributions to the fossil flora of the Western Territories. Part III. The Cretaceous and Tertiary floras. Government Printing Office, Washington, D.C., 283 pp. <https://play.google.com/books/reader?id=uJBPAQAAQAAJ&hl=en&pg=GBS.PP13>

MacGinitie HD (1953) Fossil plants of the Florissant beds, Colorado. Carnegie Institution of Washington Publication 599: 1–198. <https://catalog.hathitrust.org/Record/001639494>

MacGinitie HD (1969) The Eocene Green River flora of northwestern Colorado and northeastern Utah. University of California Publications in Geological Sciences 83: 1–203.

MacGinitie HD (1974) An early middle Eocene flora from the Yellowstone-Absaroka volcanic province, northwestern Wind River Basin, Wyoming. University of California Publications in Geological Sciences 108: 1–103.

Manchester SR (1986) Vegetative and reproductive morphology of an extinct plane tree (Platanaceae) from the Eocene of western North America. Botanical Gazette (Chicago, Ill.) 147(2): 200–226. <https://doi.org/10.1086/337587>

Manchester SR (1987) The fossil history of the Juglandaceae. Monographs in Systematic Botany from the Missouri Botanical Garden 21: 1–137. <https://doi.org/10.5962/bhl.title.154222>

Manchester SR (1989a) Attached reproductive and vegetative remains of the extinct American-European genus *Cedrelospermum* (Ulmaceae) from the early Tertiary of Utah and Colorado. American Journal of Botany 76(2): 256–276. <https://doi.org/10.1002/j.1537-2197.1989.tb11309.x>

Manchester SR (1989b) Systematics and fossil history of the Ulmaceae. In: Crane PR, Blackmore S (Eds) Evolution, Systematics, and Fossil History of the Hamamelidae, Vol II: “Higher” Hamamelidae. Clarendon Press, Oxford, 221–252.

Manchester SR (2001a) Update on the megafossil flora of Florissant, Colorado. Proceedings of the Denver Museum of Nature & Science, Series 4 1: 137–161.

Manchester SR (2001b) Leaves and fruits of *Aesculus* (Sapindales) from the Paleocene of North America. International Journal of Plant Sciences 162(4): 985–998. <https://doi.org/10.1086/320783>

Manchester SR (2002) Leaves and fruits of *Davidia* (Cornales) from the Paleocene of North America. Systematic Botany 27(2): 368–382. <https://www.jstor.org/stable/3093877>

Manchester SR (2014) Revisions to Roland Brown’s North American Paleocene flora. Acta Musei Nationalis Pragae 70(3–4): 153–210. <https://doi.org/10.14446/AMNP.2014.153>

Manchester SR, Chen Z (1996) *Palaeocarpinus aspinosa* sp. nov. (Betulaceae) from the Paleocene of Wyoming, U.S.A. International Journal of Plant Sciences 157(5): 644–655. <https://doi.org/10.1086/297386>

Manchester SR, Crane PR (1983) Attached leaves, inflorescences, and fruits of *Fagopsis*, an extinct genus of fagaceous affinity from the Oligocene Florissant Flora of Colorado, U.S.A. American Journal of Botany 70(8): 1147–1164. <https://doi.org/10.1002/j.1537-2197.1983.tb12464.x>

Manchester SR, Crane PR (1987) A new genus of Betulaceae from the Oligocene of western North America. Botanical Gazette (Chicago, Ill.) 148(2): 263–273. <https://doi.org/10.1086/337654>

Manchester SR, Dilcher DL (1997) Reproductive and vegetative morphology of *Polyptera* (Juglandaceae) from the Paleocene of Wyoming and Montana. American Journal of Botany 84(5): 649–663. <https://doi.org/10.2307/2445902>

Manchester SR, Dillhoff RM (2004) *Fagus* (Fagaceae) fruits, foliage, and pollen from the Middle Eocene of Pacific Northwestern North America. Canadian Journal of Botany 82(10): 1509–1517. <https://doi.org/10.1139/b04-112>

Manchester SR, Hickey LJ (2007) Reproductive and vegetative organs of *Browniea* gen. n. (Nyssaceae) from the Paleocene of North America. International Journal of Plant Sciences 168(2): 229–249. <https://doi.org/10.1086/509661>

Manchester SR, Dilcher DL, Wing SL (1998) Attached leaves and fruits of myrtaceous affinity from the middle Eocene of Colorado. Review of Palaeobotany and Palynology 102(3–4): 153–163. [https://doi.org/10.1016/S0034-6667\(98\)80002-X](https://doi.org/10.1016/S0034-6667(98)80002-X)

Manchester SR, Crane PR, Golovneva LB (1999) An extinct genus with affinities to extant *Davida* and *Camptotheca* (Cornales) from the Paleocene of North America and eastern Asia. International Journal of Plant Sciences 160(1): 188–207. <https://doi.org/10.1086/314114>

Manchester SR, Akhmetiev MA, Kodrul TM (2002) Leaves and fruits of *Celtis aspera* (Newberry) comb. nov. (Celtidaceae) from the Paleocene of North America and eastern Asia. International Journal of Plant Sciences 163(5): 725–736. <https://doi.org/10.1086/341513>

Manchester SR, Judd WS, Handley B (2006) Foliage and fruits of early poplars (Salicaceae: *Populus*) from the Eocene of Utah, Colorado, and Wyoming. International Journal of Plant Sciences 167(4): 897–908. <https://doi.org/10.1086/503918>

Manchester SR, Xiang Q-Y, Kodrul TM, Akhmetiev MA (2009) Leaves of *Cornus* (Cornaceae) from the Paleocene of North America and Asia confirmed by trichome characters. International Journal of Plant Sciences 170(1): 132–142. <https://doi.org/10.1086/593040>

Manchester SR, Pigg KB, Kvaček Z, DeVore ML, Dillhoff RM (2018) Newly recognized diversity in Trochodendraceae from the Eocene of western North America. International Journal of Plant Sciences 179(8): 663–676. <https://doi.org/10.1086/699282>

Martínez-Millán M, Cevallos-Ferriz SRS (2005) Arquitectura foliar de Anacardiaceae. Revista Mexicana de Biodiversidad 76(2): 137–190. <https://doi.org/10.22201/ib.20078706e.2005.002.308>

McClain AM, Manchester SR (2001) *Dipteronia* (Sapindaceae) from the Tertiary of North America and implications for the phytogeographic history of the Aceroideae. American Journal of Botany 88(7): 1316–1325. <https://doi.org/10.2307/3558343>

McIver EE, Basinger JF (1993) Flora of the Ravenscrag Formation (Paleocene), southwestern Saskatchewan, Canada. Palaeontographica Canadana 10: 1–167.

Merkhofer L, Wilf P, Haas MT, Kooyman RM, Sack L, Scoffoni C, Cúneo NR (2015) Resolving Australian analogs for an Eocene Patagonian paleorainforest using leaf size and floristics. *American Journal of Botany* 102(7): 1160–1173. <https://doi.org/10.3732/ajb.1500159>

Meyer HW (2003) The Fossils of Florissant. Smithsonian Books, Washington, D.C., 258 pp.

Meyer HW, Manchester SR (1997) The Oligocene Bridge Creek flora of the John Day Formation, Oregon. University of California Publications in Geological Sciences 141: 1–195.

Meyer HW, Wasson MS, Frakes BJ (2008) Development of an integrated paleontological database and Web site of Florissant collections, taxonomy, and publications. Geological Society of America Special Papers 435: 159–177. [https://doi.org/10.1130/2008.2435\(11\)](https://doi.org/10.1130/2008.2435(11))

Peppe DJ, Royer DL, Cariglino B, Oliver SY, Newman S, Leight E, Enikolopov G, Fernandez-Burgos M, Herrera F, Adams JM, Correa E, Currano ED, Erickson JM, Hinojosa LF, Hoganson JW, Iglesias A, Jaramillo CA, Johnson KR, Jordan GJ, Kraft NJB, Lovelock EC, Lusk CH, Niinemets Ü, Peñuelas J, Rapson G, Wing SL, Wright IJ (2011) Sensitivity of leaf size and shape to climate: Global patterns and paleoclimatic applications. *The New Phytologist* 190(3): 724–739. <https://doi.org/10.1111/j.1469-8137.2010.03615.x>

Pigg KB, Wehr WC, Ickert-Bond SM (2001) *Trochodendron* and *Nordenskioldia* (Trochodendraceae) from the middle Eocene of Washington State, USA. *International Journal of Plant Sciences* 162(5): 1187–1198. <https://doi.org/10.1086/321927>

Premoli AC (1996) Leaf architecture of South American *Nothofagus* (Nothofagaceae) using traditional and new methods in morphometrics. *Botanical Journal of the Linnean Society* 121(1): 25–40. <https://doi.org/10.1111/j.1095-8339.1996.tb00743.x>

Royer DL, Sack L, Wilf P, Lusk CH, Jordan GJ, Niinemets Ü, Wright IJ, Westoby M, Cariglino B, Coley PD, Cutter AD, Johnson KR, Labandeira CC, Moles AT, Palmer MB, Valladares F (2007) Fossil leaf economics quantified: Calibration, Eocene case study, and implications. *Paleobiology* 33(4): 574–589. <https://doi.org/10.1666/07001.1>

Schorn H (1998) *Holodiscus lisii* (Rosaceae): a new species of Ocean Spray from the late Eocene Florissant Formation, Colorado, USA. *PaleoBios* 18(4): 21–24. http://docubase.berkeley.edu/cgi-bin/pl_dochome?query_src=pl_search&collection=PaleoBios+Archive+Public&id=120

Smith ME, Carroll AR, Singer BS (2008) Synoptic reconstruction of a major ancient lake system: Eocene Green River Formation, western United States. *Geological Society of America Bulletin* 120(1–2): 54–84. <https://doi.org/10.1130/B26073.1>

Stiles E, Wilf P, Iglesias A, Gandolfo MA, Cúneo NR (2020) Cretaceous–Paleogene plant extinction and recovery in Patagonia. *Paleobiology* 46(4): 445–469. <https://doi.org/10.1017/pab.2020.45>

Taylor DW, Hickey LJ (1992) Phylogenetic evidence for the herbaceous origin of angiosperms. *Plant Systematics and Evolution* 180(3): 137–156. <https://doi.org/10.1007/BF00941148>

Todzia CA, Keating RC (1991) Leaf architecture of the Chloranthaceae. *Annals of the Missouri Botanical Garden* 78(2): 476–496. <https://doi.org/10.2307/2399575>

Traiser C, Roth-Nebelsick A, Grein M, Kovar-Eder J, Kunzmann L, Moraweck K, Lange J, Kvaček J, Neinhuis C, Folie A, De Franceschi D, Kroh A, Prestianni C, Poschmann M, Wuttke M (2018) MORPHYLL: A database of fossil leaves and their morphological traits. *Palaeontologia Electronica* 21(1): e1T. <https://doi.org/10.26879/773>

Upchurch Jr GR, Spicer RA, Leopold EB (2007) The life and career of Jack A. Wolfe (July 10, 1936–August 12, 2005). *Courier Forschungsinstitut Senckenberg* 258: 11–19.

Van Horn G, Aodha OM, Song Y, Cui Y, Sun C, Shepard A, Adam H, Perona P, Belongie S (2018) The iNaturalist species classification and detection dataset. 2018 IEEE/CVF Conference on Computer Vision and Pattern Recognition, Salt Lake City, 8769–8778. <https://doi.org/10.1109/CVPR.2018.00914>

Vasco A, Thadeo M, Conover M, Daly DC (2014) Preparation of samples for leaf architecture studies, a method for mounting cleared leaves. *Applications in Plant Sciences* 2(9): 1400038. <https://doi.org/10.3732/apps.1400038>

Veatch SW, Meyer HW (2008) History of paleontology at the Florissant fossil beds, Colorado. *Geological Society of America Special Papers* 453: 1–18. [https://doi.org/10.1130/2008.2435\(01\)](https://doi.org/10.1130/2008.2435(01))

von Ettingshausen CF (1858) Über die Nervation der Bombaceen mit besonderer Berücksichtigung der in der vorweltlichen Flora, repräsentirten Arten dieser Familie. Hof-und Staatsdruckerei, Vienna, 14 pp.

von Ettingshausen CF (1861) Die Blatt-Skelete der Dikotyledonen. Hof-und Staatsdruckerei, Vienna, Austria, 305 pp. <https://edoc.hu-berlin.de/handle/18452/713>

Wang Q, Manchester SR, Gregor H-J, Shen S, Li Z-Y (2013) Fruits of *Koelreuteria* (Sapindaceae) from the Cenozoic throughout the northern hemisphere: Their ecological, evolutionary, and biogeographic implications. *American Journal of Botany* 100(2): 422–449. <https://doi.org/10.3732/ajb.1200415>

Weaver WN, Ng J, Laport RG (2020) LeafMachine: Using machine learning to automate leaf trait extraction from digitized herbarium specimens. *Applications in Plant Sciences* 8(6): e11367. <https://doi.org/10.1002/aps3.11367>

Wilf P (2000) Late Paleocene-early Eocene climate changes in southwestern Wyoming: Paleobotanical analysis. *Geological Society of America Bulletin* 112(2): 292–307. [https://doi.org/10.1130/0016-7606\(2000\)112<292:LPECCI>2.0.CO;2](https://doi.org/10.1130/0016-7606(2000)112<292:LPECCI>2.0.CO;2)

Wilf P (2008) Fossil angiosperm leaves: Paleobotany's difficult children prove themselves. *Paleontological Society Papers* 14: 319–333. <https://doi.org/10.1017/S1089332600001741>

Wilf P, Johnson KR (2004) Land plant extinction at the end of the Cretaceous: A quantitative analysis of the North Dakota megafloral record. *Paleobiology* 30(3): 347–368. [https://doi.org/10.1666/0094-8373\(2004\)030<0347:LPEATE>2.0.CO;2](https://doi.org/10.1666/0094-8373(2004)030<0347:LPEATE>2.0.CO;2)

Wilf P, Labandeira CC (1999) Response of plant-insect associations to Paleocene-Eocene warming. *Science* 284(5423): 2153–2156. <https://doi.org/10.1126/science.284.5423.2153>

Wilf P, Beard KC, Davies-Vollum KS, Norejko JW (1998) Portrait of a late Paleocene (early Clarkforkian) terrestrial ecosystem: Big Multi Quarry and associated strata, Washakie Basin, southwestern Wyoming. *Palaios* 13(6): 514–532. <https://doi.org/10.2307/3515344>

Wilf P, Labandeira CC, Johnson KR, Coley PD, Cutter AD (2001) Insect herbivory, plant defense, and early Cenozoic climate change. *Proceedings of the National Academy of Sciences of the United States of America* 98(11): 6221–6226. <https://doi.org/10.1073/pnas.111069498>

Wilf P, Cúneo NR, Johnson KR, Hicks JF, Wing SL, Obradovich JD (2003) High plant diversity in Eocene South America: Evidence from Patagonia. *Science* 300(5616): 122–125. <https://doi.org/10.1126/science.1080475>

Wilf P, Johnson KR, Cúneo NR, Smith ME, Singer BS, Gandolfo MA (2005a) Eocene plant diversity at Laguna del Hunco and Río Pichileufú, Patagonia, Argentina. *American Naturalist* 165(6): 634–650. <https://doi.org/10.1086/430055>

Wilf P, Labandeira CC, Johnson KR, Cúneo NR (2005b) Richness of plant-insect associations in Eocene Patagonia: A legacy for South American biodiversity. *Proceedings of the National Academy of Sciences of the United States of America* 102(25): 8944–8948. <https://doi.org/10.1073/pnas.0500516102>

Wilf P, Labandeira CC, Johnson KR, Ellis B (2006) Decoupled plant and insect diversity after the end-Cretaceous extinction. *Science* 313(5790): 1112–1115. <https://doi.org/10.1126/science.1129569>

Wilf P, Cúneo NR, Escapa IH, Pol D, Woodburne MO (2013) Splendid and seldom isolated: The paleobiogeography of Patagonia. *Annual Review of Earth and Planetary Sciences* 41(1): 561–603. <https://doi.org/10.1146/annurev-earth-050212-124217>

Wilf P, Zhang S, Chikkerur S, Little SA, Wing SL, Serre T (2016) Computer vision cracks the leaf code. *Proceedings of the National Academy of Sciences of the United States of America* 113(12): 3305–3310. <https://doi.org/10.1073/pnas.1524473113>

Wilf P, Nixon KC, Gandolfo MA, Cúneo NR (2019) Eocene Fagaceae from Patagonia and Gondwanan legacy in Asian rainforests. *Science* 364(6444): e5139. <https://doi.org/10.1126/science.aaw5139>

Wing SL (1992) High-resolution leaf x-radiography in systematics and paleobotany. *American Journal of Botany* 79(11): 1320–1324. <https://doi.org/10.1002/j.1537-2197.1992.tb13736.x>

Wing SL (1998) Late Paleocene-early Eocene floral and climatic change in the Bighorn Basin, Wyoming. In: Aubrey M-P, Lucas S, Berggren WA (Eds) *Late Paleocene-Early Eocene Climatic and Biotic Events in the Marine and Terrestrial Records*. Columbia University Press, New York, 380–400.

Wing SL, Hickey LJ (1984) The *Platycarya* perplex and the evolution of the Juglandaceae. *American Journal of Botany* 71(3): 388–411. <https://doi.org/10.1002/j.1537-2197.1984.tb12525.x>

Wing SL, Herrera F, Jaramillo CA, Gómez-Navarro C, Wilf P, Labandeira CC (2009) Late Paleocene fossils from the Cerrejón Formation, Colombia, are the earliest record of Neotropical rainforest. *Proceedings of the National Academy of Sciences of the United States of America* 106(44): 18627–18632. <https://doi.org/10.1073/pnas.0905130106>

Wing SL, Johnson KR, Peppe DJ, Green WA, Taylor DW (2014) The multi-stranded career of Leo J. Hickey. *Bulletin - Peabody Museum of Natural History* 55(2): 69–78. <https://doi.org/10.3374/014.055.0201>

Winkler IS, Labandeira CC, Wappler T, Wilf P (2010) Distinguishing Agromyzidae (Diptera) leaf mines in the fossil record: New taxa from the Paleogene of North America and Germany and their evolutionary implications. *Journal of Paleontology* 84(5): 935–954. <https://doi.org/10.1666/09-163.1>

Wolfe JA, Schorn HE (1990) Taxonomic revision of the Spermatopsida of the Oligocene Creede flora, southern Colorado. *US Geological Survey Bulletin* 23: 1–40. <https://doi.org/10.3133/b1923>

Wolfe JA, Tanai T (1987) Systematics, phylogeny, and distribution of *Acer* (maples) in the Cenozoic of western North America. *Journal of the Faculty of Science, Hokkaido University, Series 4: Geology and Mineralogy* 22(1): 1–246. <http://hdl.handle.net/2115/36747>

Wolfe JA, Wehr WC (1987) Middle Eocene dicotyledonous plants from Republic, north-eastern Washington. US Geological Survey Bulletin 1597: 1–25. <https://pubs.usgs.gov/bul/1597/report.pdf>

Xu H, Blonder B, Jodra M, Malhi Y, Fricker M (2021) Automated and accurate segmentation of leaf venation networks via deep learning. The New Phytologist 229(1): 631–648. <https://doi.org/10.1111/nph.16923>

Appendix I

Species list for fossil-leaf images.

Species	#Images	Source, age, region†	References
Dryopteridaceae			
<i>Dryopteris guyottii</i> (Lesquereux)	64	Florissant, late Eocene, Colorado, USA	MacGinitie 1953
Cupressaceae			
<i>Chamaecyparis linguaefolia</i> (Lesquereux)	100	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
MacGinitie, <i>Chamaecyparis</i> sp.			
<i>Sequoia affinis</i> Lesquereux, <i>Sequoia</i> sp.	153	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Pinaceae			
<i>Pinus florissantii</i> Lesquereux	9	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Pinus macginitiae</i> Axelrod	1	Florissant, late Eocene, Colorado, USA	Axelrod 1986; Manchester 2001a
<i>Pinus wheeleri</i> Cockerell, <i>Pinus</i> sp.	55	Florissant, late Eocene, Colorado, USA	Cockerell 1908; MacGinitie 1953; Manchester 2001a
Taxaceae			
<i>Torreya geometrorum</i> (Cockerell)	9	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
MacGinitie, <i>Torreya</i> sp.			
Araceae			
Araceae sp. CJ80	2	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
<i>Limnobiophyllum scutatum</i> (Dawson)	2	Florissant, late Eocene, Colorado, USA	see Manchester 2001a
Krassilov			
<i>Montrichardia aquatica</i> Herrera et al.	4	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Herrera et al. 2008
<i>Petrocardium cerrejonense</i> Herrera et al.	1	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Herrera et al. 2008
<i>Petrocardium wayuuorum</i> Herrera et al.	1	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Herrera et al. 2008
Arecaceae			
Arecaceae spp. CJ67, CJ68	4	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Gómez-Navarro et al. 2009; Wing et al. 2009
Rhipogonaceae			
<i>Ripogonum americanum</i> R.J. Carpenter et al.	2	Laguna del Hunco, early Eocene, Chubut, Argentina	Carpenter et al. 2014
Zingiberaceae			
Zingiberaceae spp. CJ49, CJ65	3	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
Adoxaceae			
<i>Sambucus newtonii</i> Cockerell	33	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a

Species	#Images	Source, age, region†	References
Akaniaceae			
<i>Akania patagonica</i> Gandolfo et al.	9	Laguna del Hunco, early Eocene, Chubut, Argentina	Gandolfo et al. 1988; Wilf et al. 2005a‡
Altingiaceae			
“ <i>Acer</i> ” <i>lesquereuxi</i> Knowlton	2	Little Mountain & Bonanza, early (LM) and middle (B) Eocene, Wyoming (LM) & Utah (B), USA	MacGinitie 1969; Johnson and Plumb 1995; Wilf et al. 2005b‡
Anacardiaceae			
<i>Anardiaceae</i> sp. CJ34	4	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
<i>Anardiaceae</i> sp. TY203	1	Laguna del Hunco, early Eocene, Chubut, Argentina	P. Wilf, unpubl. obs.
<i>Rhus lesquereuxi</i> Knowlton & Cockerell	21	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Rhus malloryi</i> Wolfe & Wehr	5	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡; Flynn et al. 2019
<i>Rhus nigricans</i> (Lesquereux) Knowlton	27	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	MacGinitie 1969; Wilf 2000‡; Wilf et al. 2001‡
<i>Rhus obscura</i> (Lesquereux) MacGinitie	22	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Rhus stellariaefolia</i> (Lesquereux) MacGinitie, <i>Rhus</i> sp.	175	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Schmalzia</i> (<i>Rhus</i>) <i>vexans</i> (Lesquereux) Cockerell	5	Florissant, late Eocene, Colorado, USA	MacGinitie 1953
Apocynaceae			
<i>Apocynaceae</i> sp. RR17	1	Wasatch Fm., early Eocene, Wyoming, USA	Wing 1998; Wilf 2000‡
Atherospermataceae			
<i>Atherospermophyllum guinazui</i> (E.W. Berry) C.L. Knight	16	Laguna del Hunco, early Eocene, Chubut, Argentina	Knight and Wilf 2013
Berberidaceae			
<i>Mahonia marginata</i> (Lesquereux) Arnold	15	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Mahonia subdenticulata</i> (Lesquereux) MacGinitie, <i>Mahonia</i> sp.	11	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Betulaceae			
<i>Alnus parvifolia</i> (E.W. Berry) Wolfe and Wehr	33	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Alnus</i> sp. RR14	1	Wasatch Fm., early Eocene, Wyoming, USA	Wilf 2000
<i>Betula leopoldae</i> Wolfe and Wehr	4	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Betula</i> sp. RP21	1	Republic, early Eocene, Washington, USA	Wilf et al. 2005b
<i>Corylites</i> spp. BB05, FW01	3	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Manchester and Chen 1996; Wilf 2000‡
<i>Paracarpinus fraterna</i> (Lesquereux) Manchester & Crane, <i>Paracarpinus</i> sp.	58	Florissant, late Eocene, Colorado, USA	Manchester and Crane 1987; Manchester 2001a
Cannabaceae			
<i>Celtis aspera</i> (Newberry) Manchester et al.§	3	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000‡; Manchester et al. 2002, 2018; Wilf et al. 2006‡
Cercidiphyllaceae			
<i>Archeampelos lobatocrenata</i> (Lesquereux) Doweld	2	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	McIver and Basinger 1993; Wilf 2000‡; Wilf et al. 2006‡; Manchester 2014; Doweld 2016

Species	#Images	Source, age, region†	References
<i>Cercidiphyllum obtitum</i> (Dawson) Wolfe and Wehr	12	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Trochodendroides genetrix</i> (Newberry) Manchester	13	Fort Union Fm., several sites, early & late Paleocene, Montana, North Dakota, & Wyoming, USA	Wilf 2000‡; Johnson 2002‡; Wilf et al. 2006‡; Manchester 2014
Cornaceae			
<i>Beringiaphyllum cupanioides</i> (Newberry) Manchester et al.	3	SW Wyoming, several sites, late Paleocene, Wyoming, USA	Manchester et al. 1999; Wilf 2000‡; Wilf et al. 2006‡
<i>Brownia serrata</i> (Newberry) Manchester & Hickey	6	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000‡; Wilf et al. 2006‡; Manchester and Hickey 2007
<i>Cornus</i> sp. RP62	3	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Cornus swingii</i> Manchester et al.	2	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000‡; Wilf et al. 2006‡; Manchester et al. 2009
<i>Davida antiqua</i> (Newberry) Manchester	4	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000‡; Manchester 2002; Wilf et al. 2006‡
Cunoniaceae			
Cunoniaceae sp. SA020	13	Salamanca Fm., early Paleocene, Chubut, Argentina	Iglesias et al. 2007, 2021
Cunoniaceae sp. TY116	9	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a; Merkhofer et al. 2015
Ericaceae			
Ericaceae sp. RP41	1	Republic, early Eocene, Washington, USA	Wilf et al. 2005b‡
Rhododendron sp. RP53	1	Republic, early Eocene, Washington, USA	Wilf et al. 2005b‡
Euphorbiaceae			
Euphorbiaceae sp. CJ24	4	Cerrojón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
<i>Stillingia casca</i> Hickey	1	Wasatch Fm., early Eocene, Wyoming, USA	Hickey 1977; Wilf 2000‡
Fabaceae			
<i>Caesalpinia pecorae</i> Brown	9	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	MacGinitie 1969; Wilf 2000‡; Wilf et al. 2001‡
<i>Caesalpinites acuminatus</i> (Lesquereux) MacGinitie	13	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Caesalpinites coloradicus</i> MacGinitie	21	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Cercis parvifolia</i> Lesquereux	23	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Jia and Manchester 2014
<i>Conzzattia coriacea</i> MacGinitie, Fabaceae sp.	21	Florissant, late Eocene, Colorado, USA	MacGinitie 1953
Fabaceae sp. SA045	1	Salamanca Fm., early Paleocene, Chubut, Argentina	Iglesias et al. 2007, 2021; Brea et al. 2008
Fabaceae sp. TY117	14	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a
Fabaceae spp. CJ1, CJ19, CJ55	15	Cerrojón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009; Herrera et al. 2019
Fabaceae spp. GR554, GR567	3	Little Mountain, early Eocene, Wyoming, USA	Wilf 2000
Fabaceae spp. morphotypes 2, 3	3	Cogua & Nemocón mines, late Paleocene, Cogua, Cundinamarca, Colombia	Herrera et al. 2019
<i>Gymnocladus hesperia</i> (Brown) MacGinitie	1	Little Mountain, early Eocene, Wyoming, USA	MacGinitie 1969; Wilf 2000‡
<i>Leguminosites lespedezoides</i> MacGinitie	1	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a

Species	#Images	Source, age, region†	References
<i>Leguminosites lesquereuxiana</i> (Knowlton) Brown	6	Bonanza, middle Eocene, Utah, USA	MacGinitie 1969; Wilf et al. 2001‡
<i>Parvileguminophyllum coloradensis</i> (Knowlton) Call & Dilcher	30	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	Call and Dilcher 1994; Wilf 2000‡; Wilf et al. 2001‡
“ <i>Prosopis</i> ” <i>linearifolia</i> (Lesquereux) MacGinitie	23	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Robinia lesquereuxi</i> (Ettingshausen) MacGinitie	39	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Fagaceae			
<i>Castanea dolichophylla</i> Cockerell	15	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Castaneophyllum patagonicum</i> Wilf et al.	17	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2019
cf. <i>Quercus</i> sp. GR522	2	Little Mountain, early Eocene, Wyoming, USA	Wilf 2000
Fagaceae spp. RP060, RP154	2	Republic, early Eocene, Washington, USA	Manchester and Dillhoff 2004; Wilf et al. 2005b
<i>Fagopsis longifolia</i> (Lesquereux) Hollick, <i>Fagopsis</i> sp., cf. <i>Fagopsis</i> sp.	521	Florissant, late Eocene, Colorado, USA	Manchester and Crane 1983; Manchester 2001a
<i>Quercus dumosoides</i> MacGinitie	9	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus lyratiflormis</i> Cockerell	1	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus mohavensis</i> Axelrod	9	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus orbata</i> MacGinitie	11	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus peritula</i> Cockerell	12	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus predayana</i> MacGinitie	8	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus scottii</i> (Lesquereux) MacGinitie	37	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Quercus scudderii</i> Knowlton, <i>Quercus</i> sp.	46	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Grossulariaceae			
<i>Ribes errans</i> MacGinitie	3	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Hamamelidaceae			
<i>Langeria magnifica</i> Wolfe and Wehr	2	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
Hydrangeaceae			
<i>Philadelphus minutus</i> MacGinitie	1	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Iteaceae			
<i>Itea</i> sp. RP19	3	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡; Hermsen 2013
Juglandaceae			
<i>Carya libbeyi</i> (Lesquereux) MacGinitie, <i>Carya</i> sp.	53	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 1987; 2001a
Juglandaceae sp. BB09	1	Bison Basin, late Paleocene, Wyoming, USA	Gemmill and Johnson 1997; Wilf 2000
Juglandaceae sp. GR519	1	Little Mountain, early Eocene, Wyoming, USA	Wilf 2000
<i>Juglandiphyllites glabra</i> (R. Brown) Manchester and Dilcher	7	Fort Union Fm., several sites, early & late Paleocene, Montana & Wyoming, USA	Manchester and Dilcher 1997; Wilf 2000‡; Wilf et al. 2006‡

Species	#Images	Source, age, region†	References
<i>Platycarya americana</i> Hickey	1	Wasatch Fm., early Eocene, Wyoming, USA	Wing and Hickey 1984; Manchester 1987; Wilf 2000‡
<i>Platycarya castaneopsis</i> (Lesquereux) Wing & Hickey	1	Wasatch Fm., early Eocene, Wyoming, USA	Wing and Hickey 1984; Manchester 1987; Wilf 2000‡
Lauraceae			
“ <i>Ficus</i> ” <i>planicostata</i> Lesquereux	1	Battleship, Maastrichtian, North Dakota, USA	Johnson 2002‡
Lauraceae sp. RR19	1	Wasatch Fm. several sites, early Eocene, Wyoming, USA	Wilf 2000
Lauraceae sp. SA010	17	Salamanca Fm., early Paleocene, Chubut, Argentina	Iglesias et al. 2007, 2021
Lauraceae sp. TY084	21	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a
Lauraceae spp. CJ5, CJ22	8	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
Lauraceae spp. FW02, FW03, FW28	6	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000
<i>Laurophyllo piatnitzkyi</i> E.W. Berry	12	Salamanca & Peñas Coloradas fms., early Paleocene, Chubut, Argentina	Iglesias et al. 2007, 2021‡
<i>Lindera coloradica</i> MacGinitie	4	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Lindera varifolia</i> MacGinitie	18	Little Mountain, early Eocene, Wyoming, USA	MacGinitie 1969; Wilf et al. 2001‡
<i>Marmarthia pearsoni</i> K. Johnson	5	Hell Creek Fm., several sites, Maastrichtian, North Dakota, USA	Johnson 1996, 2002
<i>Marmarthia trivialis</i> K. Johnson	2	Madeline's Bank, Hell Creek Fm., Maastrichtian, North Dakota, USA	Johnson 1996, 2002
<i>Persites argutus</i> Hickey	11	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Hickey 1977; Wilf 2000‡; Wilf et al. 2006‡
“ <i>Sassafras</i> ” <i>hesperia</i> E.W. Berry	2	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
“ <i>Sassafras</i> ” <i>hesperia</i> E.W. Berry	6	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
Magnoliaceae			
<i>Liriodendrites bradacii</i> K. Johnson	1	Madeline's Bank, Hell Creek Fm., Maastrichtian, North Dakota, USA	Johnson 1996, 2002
Malvaceae			
“ <i>Dombeya</i> ” <i>novi-mundi</i> Hickey	1	Wasatch Fm., several sites, early Eocene, Wyoming, USA	Hickey 1977; Wilf 2000‡; Carvalho et al. 2011
Malvaceae sp. TY023	16	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a; Wilf 2008
Malvaceae spp. CJ25, CJ36	3	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009; Carvalho et al. 2011
<i>Malvaciphyllum macondicus</i> M. Carvalho	6	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009; Carvalho et al. 2011
<i>Tilia johnsoni</i> Wolfe & Wehr	1	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Triumfetta ovata</i> MacGinitie	4	Little Mountain, early Eocene, Wyoming, USA	MacGinitie 1969; Wilf 2000‡
Meliaceae			
<i>Cedrela lancifolia</i> (Lesquereux) Brown	35	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Meliaceae sp. CJ2	3	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Wing et al. 2009
Menispermaceae			
Menispermaceae sp. GR507	1	Little Mountain, early Eocene, Wyoming, USA	Wilf 2000
<i>Menispermites cerrejonensis</i> Doria et al.	6	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Doria et al. 2008

Species	#Images	Source, age, region†	References
<i>Menispermites cordatus</i> Doria et al.	1	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Doria et al. 2008
<i>Menispermites horizontalis</i> Doria et al.	1	Cerrejón mine, middle-late Paleocene, Guajira Peninsula, Colombia	Doria et al. 2008
<i>Wilkinsoniphyllum menispermoides</i> Jud et al.	1	Salamanca Fm., early Paleocene, Chubut, Argentina	Iglesias et al. 2007, 2021; Jud et al. 2018
Monimiaceae			
<i>Monimiophyllum callidentatum</i> C.L. Knight	1	Laguna del Hunco, early Eocene, Chubut, Argentina	Knight and Wilf 2013
Myricaceae			
<i>Comptonia columbiana</i> Dawson	1	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
Myrtaceae			
<i>Eucalyptus frenguelliana</i> Gandolfo & Zamaloa	13	Laguna del Hunco, early Eocene, Chubut, Argentina	Gandolfo et al. 2011; Hermsen et al. 2012
<i>Eugenia arenaceaeformis</i> (Cockerell) MacGinitie	16	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Myrtaceae sp. TY41	1	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a
<i>Syzygioides americana</i> (Lesquereux) Manchester et al.	5	Little Mountain, Bonanza, & Wasatch Fm., early and middle Eocene, Wyoming & Utah, USA	Manchester et al. 1998; Wilf 2000‡; Wilf et al. 2005b‡
Platanaceae			
<i>Erlingdorfia montana</i> (Brown) K. Johnson	15	Hell Creek Fm., several sites, Maastrichtian, North Dakota, USA	Johnson 1996, 2002
<i>Grewiopsis saportana</i> Lesquereux	8	Hell Creek Fm., several sites, Maastrichtian, North Dakota, USA	Johnson 2002‡
<i>Leepierciea preartocarpoides</i> K. Johnson	5	Hell Creek Fm., several sites, Maastrichtian, North Dakota, USA	Johnson 1996, 2002
<i>Macginitiea gracilis</i> (Lesquereux) Wolfe and Wehr	4	Fort Union Fm., several sites & Republic, late Paleocene, early Eocene, Washington & Wyoming, USA	Wolfe and Wehr 1987; Wilf 2000‡
<i>Macginitiea wyomingensis</i> (Knowlton & Cockerell) Manchester	9	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	Manchester 1986; Wilf 2000‡; Wilf et al. 2005b‡
<i>Platanites marginata</i> (Lesquereux) K. Johnson	7	Hell Creek Fm., several sites, Maastrichtian, North Dakota, USA	Johnson 1996, 2002
<i>Platanites raynoldsi</i> (Newberry) Manchester	21	Fort Union Fm., several sites, early & late Paleocene, Montana, North Dakota, & Wyoming, USA	Wilf 2000‡; Johnson 2002‡; Wilf et al. 2006‡; Manchester 2014
<i>Platanus</i> sp. GR506	1	Little Mountain, early Eocene, Wyoming, USA	Wilf 2000
Proteaceae			
<i>Lomatia occidentalis</i> (E.W. Berry) Frenguelli	32	Laguna del Hunco, early Eocene, Chubut, Argentina	González et al. 2007
<i>Lomatia preferruginea</i> E.W. Berry	9	Laguna del Hunco, early Eocene, Chubut, Argentina	González et al. 2007
Proteaceae gen. et sp. indet. (sp. TY208)	1	Laguna del Hunco, early Eocene, Chubut, Argentina	González et al. 2007
Rhamnaceae			
<i>Hovenia</i> cf. <i>H. oregonensis</i> Meyer & Manchester	1	Wasatch Fm., several sites, early Eocene, Wyoming, USA	Meyer and Manchester 1997; Wilf 2000‡
Rhamnaceae sp. TY025	15	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a
<i>Rhamnica cleburnii</i> (Lesquereux) Doweld	1	Battleship, Maastrichtian, North Dakota, USA	McIver and Basinger 1993; Johnson 2002‡; Doweld 2016
<i>Suessenia grandensis</i> Jud et al.	4	Rancho Grande, early Paleocene, Chubut, Argentina	Jud et al. 2017 (source of images used)

Species	#Images	Source, age, region†	References
<i>Zizyphus florissantii</i> (Lesquereux) MacGinitie.	11	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Rosaceae			
<i>Amelanchier scudderii</i> Cockerell, <i>Amelanchier</i> sp.	10	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Cercocarpus myricaefolius</i> (Lesquereux) MacGinitie, <i>Cercocarpus</i> sp.	146	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Wolfe and Schorn 1990; Manchester 2001a
<i>Crataegus copeana</i> (Lesquereux) MacGinitie	33	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Crataegus hendersonii</i> (Cockerell) MacGinitie	4	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Crataegus nupta</i> (Cockerell) MacGinitie, <i>Crataegus</i> sp.	10	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Crataegus</i> sp., aff. <i>Crataegus</i> spp. RP32, RP42	6	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Holodiscus lisii</i> Schorn	2	Florissant, late Eocene, Colorado, USA	Schorn 1998; Manchester 2001a
<i>Malus florissantensis</i> (Cockerell) MacGinitie	2	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Malus pseudocredneria</i> (Cockerell) MacGinitie	1	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Photinia pageae</i> Wolfe & Wehr	3	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
<i>Prunus gracilis</i> (Lesquereux) MacGinitie	4	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Prunus</i> sp. RP54	2	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡ DeVore and Pigg 2007
<i>Rosa hilliae</i> Lesquereux, <i>Rosa</i> sp.	23	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Rosaceae sp. RP61	2	Republic, early Eocene, Washington, USA	Wilf et al. 2005b
<i>Rubus coloradensis</i> (MacGinitie) Wolfe & Tanai	2	Florissant, late Eocene, Colorado, USA	Wolfe and Tanai 1987
<i>Spiraea</i> sp. RP29	8	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡; DeVore and Pigg 2007
<i>Vauquelinia coloradensis</i> (Knowlton) MacGinitie	52	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Vauquelinia lineara</i> MacGinitie, <i>Vauquelinia</i> sp.	7	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
Salicaceae			
<i>Populus cinnamomoides</i> (Lesquereux) MacGinitie	2	Little Mountain, early Eocene, Wyoming, USA	MacGinitie 1969; Wilf 2000‡; Manchester et al. 2006
<i>Populus crassia</i> (Lesquereux) Cockerell, <i>Populus</i> sp.	93	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a
<i>Populus tidwellii</i> Manchester et al.	3	Bonanza, middle Eocene, Utah, USA	MacGinitie 1969; Wilf et al. 2001‡; Manchester et al. 2006
<i>Populus wilmattae</i> Cockerell	4	Bonanza, middle Eocene, Utah, USA	MacGinitie 1969; Wilf et al. 2001‡
<i>Populus wyomingiana</i> (E.W. Berry) MacGinitie	1	Wasatch Fm., several sites, early Eocene, Wyoming, USA	MacGinitie 1974; Wilf 2000‡
<i>Salix cockerelli</i> Brown	8	Bonanza, middle Eocene, Utah, USA	MacGinitie 1969; Wilf et al. 2001‡
<i>Salix ramaleyi</i> Cockerell, <i>Salix</i> sp.	15	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; J.A. Wolfe pers. comm. in Manchester 2001a
<i>Salix taxifolioides</i> MacGinitie	2	Florissant, late Eocene, Colorado, USA	MacGinitie 1953
Sapindaceae			
<i>Acer florissantii</i> Kirchner, <i>Acer</i> sp.	36	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Wolfe and Tanai 1987
<i>Aesculus hickeyi</i> Manchester	3	Fort Union Fm., several sites, late Paleocene, Wyoming, USA	Wilf 2000‡; Manchester 2001b; Wilf et al. 2006‡

Species	#Images	Source, age, region†	References
<i>Allophylus flexifolia</i> (Lesquereux) MacGinitie	10	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	MacGinitie 1969; Wilf 2000‡; Wilf et al. 2001‡
<i>Athyana haydenii</i> (Lesquereux) MacGinitie	106	Florissant, late Eocene, Colorado, USA	MacGinitie 1953
<i>Bohleia insignis</i> (Lesquereux) Wolfe & Wehr	16	Florissant, late Eocene, Colorado, USA	Wolfe and Wehr 1987; Manchester 2001a; McClain and Manchester 2001
“ <i>Cardiospermum</i> ” <i>coloradensis</i> (Knowlton) MacGinitie	5	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	MacGinitie 1969; Wilf 2000‡; Wilf et al. 2001‡; Jud et al. 2021
“ <i>Cardiospermum</i> ” <i>terminalis</i> (Lesquereux) MacGinitie, “ <i>Cardiospermum</i> ” sp.	41	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a; Jud et al. 2021
<i>Koelreuteria allenii</i> (Lesquereux) Edwards	28	Florissant, late Eocene, Colorado, USA	MacGinitie 1953; Manchester 2001a; Wang et al. 2013
Sapindaceae sp. TY018	18	Laguna del Hunco, early Eocene, Chubut, Argentina	Wilf et al. 2005a
Schoepfiaceae			
<i>Schoepfia republicensis</i> (La Motte) Wolfe & Wehr	2	Republic, Wasatch Fm., early Eocene, Washington & Wyoming, USA	Wolfe and Wehr 1987; Wilf 2000‡; Wilf et al. 2005b‡
Theaceae			
Theaceae sp. RP49	2	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡
Trochodendraceae			
<i>Trochodendron nastae</i> Pigg et al.	1	Republic, early Eocene, Washington, USA	Pigg et al. 2001; Wilf et al. 2005b‡ Manchester et al. 2018
<i>Ziziphoides flabellum</i> (Newberry) Crane et al.	9	Fort Union Fm., several sites, early Paleocene, Montana, North Dakota, & Wyoming, USA	Crane et al. 1991; Johnson 2002‡; Wilf et al. 2006‡
<i>Ziziphoides</i> sp. RP37	1	Republic, early Eocene, Washington, USA	Pigg et al. 2001; Wilf et al. 2005b‡
Ulmaceae			
<i>Cedrelospermum lineatum</i> (Lesquereux) Manchester, <i>Cedrelospermum</i> sp.	978	Florissant, late Eocene, Colorado, USA	Manchester 1989a, 2001a
<i>Cedrelospermum nervosum</i> (Newberry) Manchester	30	Little Mountain & Bonanza, early and middle Eocene, Wyoming & Utah, USA	Manchester 1989a; Wilf 2000‡; Wilf et al. 2001‡
<i>Ulmites microphylla</i> (Newberry) Manchester	1	Wasatch Fm. several sites, early Eocene, Wyoming, USA	Wilf 2000‡; Manchester 2014
<i>Ulmus</i> sp. RP17, <i>Zelkova</i> sp. RP50	8	Republic, early Eocene, Washington, USA	Wolfe and Wehr 1987; Wilf et al. 2005b‡ Denk and Dillhoff 2005
<i>Ulmus tenuinervis</i> Lesquereux, Ulmaceae sp., <i>Ulmus</i> sp.	30	Florissant, late Eocene, Colorado, USA	Manchester 1989b, 2001a
Vitaceae			
“ <i>Vitis florissantella</i> ”	11	Florissant, late Eocene, Colorado, USA	MacGinitie 1953, 1969; Manchester 2001a

† Fossil sites listed are only those that sourced the fossils in this dataset and may not include type localities and other occurrences of the species.

‡ Occurrence reference for the dataset in this paper, when different from the taxonomic reference.

§ Doweld (2016) proposed revision of *Celtis aspera* to *Rhamnites asperus* (Newberry) Doweld, which we acknowledge but do not adopt here.