

1 **Ancient trouble in paradise: seed beetle predation on coconuts from middle–late Paleocene**
2 **rainforests of Colombia**

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21 **Abstract**

22 Modern Neotropical rainforests are characterized by the high intensity and host specificity with
23 which insects feed on plants. Previous studies have shown that, during the middle–late
24 Paleocene, the leaves of the early evolving Neotropical rainforests of tropical South America
25 were heavily herbivorized by insects. Yet, less attention has been given to insect damage found
26 on fossil fruits and seeds, despite the host specialization of many disseminule predators in
27 modern forests. Here, we present and describe borings found on a fruit compression fossil of cf.
28 *Cocos* (coconut) from the middle–late Paleocene Cerrejón Formation (58–60 Ma) of Colombia.
29 We interpret the borings as constructed by palm seed beetles (Chrysomelidae: Bruchinae:
30 Pachymerina) based on size, number, position, plant reaction tissue, and plant host selection.
31 This occurrence provides the earliest record of an ecological interaction between seed beetles
32 and palms, suggesting that this host-specific interaction has been consistently maintained for
33 several tens of millions of years.

34 **Keywords:** *Cocos*, Seed predation, Host-specialized interactions, Fossil fruits, Paleobotany,
35 Paleoecology

36 **1. Introduction**

37 A defining feature of modern Neotropical rainforests is the high intensity and host
38 specificity with which insects feed on plants (Dyer et al., 2007). Host-specific herbivory is a
39 major factor in plant population dynamics, as it creates density-dependent restrictions on
40 population growth and contributes to local and regional patterns of species diversity (Comita et
41 al., 2014; Forrister et al., 2019; Terborgh, 2012). Abundant insect-mediated leaf damage found in
42 middle–late Paleocene deposits of tropical South America indicates that herbivory of leaves was

43 intense during the early evolution of modern-like Neotropical rainforests (Carvalho et al., 2021;
44 Giraldo et al., 2021; Wing et al., 2009). Yet, much less attention has been given to insect damage
45 found on fossil fruits and seeds, even though many insect seed predators are specialized to
46 varying extent (Lewis and Gripenberg, 2008) and the host-specific nature of seed-predating
47 insects is of major importance to the evolution and maintenance of tropical forest diversity
48 (Connell, 1971; Janzen, 1970). Seed predation is a form of insect (or arthropod) attack that
49 involves the penetration of the outer seed coat and targeting of the seed's internal tissues,
50 typically resulting in the inviability of the new plant. Evidence of this form of feeding extends
51 back to the Early Pennsylvanian (Jennings, 1974; Labandeira, 2006; Scott and Taylor, 1983) and
52 is recorded as various types of scars and damaged tissues resulting from the attack. These include
53 numerous kinds of individual or grouped punctures and cratered pits into the seed main body,
54 indicating the removal and hollowing out of embryonic tissues, and circular to elliptical exit
55 holes (e.g., Barbosa dos Santos et al., 2020; Labandeira et al., 2007)

56 Currently, numerous species representing six major orders of insects are known to feed
57 on seeds, including true bugs (Hemiptera), thrips (Thysanoptera), flies (Diptera), moths
58 (Lepidoptera), wasps (Hymenoptera) (Lewis and Gripenberg, 2008; Louda, 1982; Roques et al.,
59 2016; Sweet, 1960), and, especially beetles (Coleoptera) (Janzen, 1980; Lewis and Gripenberg,
60 2008). Seed beetles (Chrysomelidae: Bruchinae), for instance, spend their larval period feeding
61 on –and living in– seeds (Johnson et al., 1995; Nilsson and Johnson, 1993), often being specific
62 to seeds of particular plant genera or species (Johnson and Slobodchikoff, 1979). Approximately
63 85% of bruchine larvae live inside legume seeds (Fabaceae), 4% within palms (Arecaceae),
64 another 4% in the morning glory family (Convolvulaceae), and 2% in the mallow family
65 (Malvaceae), whereas the remaining 4% are distributed among 29 other angiosperm plant

66 families (Borowiec, 1987; Nilsson and Johnson, 1993). Female bruchids oviposit on a fruit or
67 seed, where the first instar larva bores through the fruit and/or seed coat and enters the seed
68 cavity (Johnson et al., 1995). While inside, the larva molts into a legless grub and feeds on the
69 endosperm and embryonic tissues throughout another three molts before the larva pupates within
70 the seed (Johnson et al., 1995). Prior to pupation, the larva excavates a round exit hole –either
71 within the seed or from the seed and the fruit– which the emerging adult finishes (Johnson et al.,
72 1995). Adult bruchines feed on nectar and pollen and are not known to feed on seeds or fruits
73 (Johnson et al., 1995; Nilsson and Johnson, 1993).

74 Phylogenetically nested within the Bruchinae are the palm bruchines (subtribe
75 Pachymerina), which feed almost exclusively on palm seeds (Delobel et al., 1995; Johnson et al.,
76 1995; Nilsson and Johnson, 1993). Palm bruchines are restricted to the New World, and although
77 most species are tropical, some occur as far north as Texas and as far south as Argentina
78 (Johnson et al., 1995; Nilsson and Johnson, 1993). Currently, four palm bruchine genera are
79 recognized: *Caryoborus*, *Caryobruchus*, *Pachymerus* and *Speciomerus* (Nilsson and Johnson,
80 1993), with varying degrees of palm host specificity. Species of *Caryobruchus* show a clear
81 preference for palms in the tribes Corypheae (gebang, talipot, and buri palms), Phoeniceae (date
82 palms) and Chamaedoreeae (bamboo and parlor palms), whereas the seeds of the Phytelepheae
83 (ivory and tagua palms) and Cocoseae (coconut, oil, and jelly palms) are consumed by species of
84 *Caryoborus*, *Speciomerus* and most extensively by species of *Pachymerus* (Johnson et al., 1995).

85 Here, we present and describe an example of host-specific seed predation found on a
86 fruit compression of cf. *Cocos* sp. (Arecaceae) (Gomez- Navarro et al., 2009), a coconut from a
87 middle–late Paleocene rainforest from northern South America. Borings and exit holes observed
88 are consistent with those made by palm bruchines (Pachymerina), based on size, number,

89 position, plant reaction tissue, and plant host selection. This occurrence provides the earliest
90 record of an ecological interaction between seed beetles and palms. It also indicates that the
91 specific behavior of palm-feeding bruchines dates back to at least 60 Ma and suggests the
92 antiquity of this type of host-specific interaction through geologic time.

93 **2. Methods**

94 **2.1 Geological and environmental setting**

95 The Cerrejón Formation is a ~700 m thick sequence of sandstones, mudstones, claystones
96 and coals (Jaramillo et al., 2007) that are exposed along the Cerrejón open-pit coal mine, located
97 in the Ranchería Basin of northeastern Colombia (Figure 1). These deposits accumulated in
98 coastal plains that transitioned from estuarine to fluvial and lacustrine-influenced depositional
99 systems in an environment of warm temperatures and high precipitation (Head et al., 2012;
100 Jaramillo et al., 2011, 2007). Pollen zonation, correlations with stable carbon isotopic data and
101 marine microfossils from the Cerrejón Formation indicate a middle–late Paleocene age for the
102 entire sequence (ca. 58–60 Ma) (Jaramillo et al., 2011, 2007).

103 Along with the middle–late Paleocene Bogotá flora (Giraldo et al., 2021), the Cerrejón
104 flora localities are the earliest known examples of Neotropical rainforests (Carvalho et al., 2021;
105 Jaramillo et al., 2007; Wing et al., 2009). Extinction and turnover related to the end-Cretaceous
106 ecological crisis led to the assembly of modern-like Neotropical rainforests (Carvalho et al.,
107 2021), characterized by flowering plant dominance (Carvalho et al., 2021; Jaramillo et al., 2007;
108 Wing et al., 2009), closed, multistratal canopy structure (Graham et al., 2019), abundance of
109 legumes (Herrera et al., 2019), and hot, humid climate (Carvalho et al., 2021; Wing et al., 2009).
110 The coastal, peat-accumulating rainforests of Cerrejón included tropical tree lineages such as
111 mallows and kapok trees (Malvaceae: Malvoideae and Bombacoideae; Carvalho et al., 2011);

112 epiphytic and semiaquatic aroids (Araceae; Herrera et al., 2008); abundant vines such as
113 moonseeds (Menispermaceae; Doria et al., 2008; Herrera et al., 2011) and species of Icacinaceae
114 (Stull et al., 2012); and diverse palms (Arecaceae; Gomez- Navarro et al., 2009). Non-monocot
115 flowering plants were heavily herbivorized, as nearly half of their leaves show signs of leaf
116 chewing, galling, mining and piercing marks (Wing et al., 2009). The overall richness of insect
117 damage on leaves is nonetheless low for the Cerrejón flora, when compared to coeval floras
118 around the world (Giraldo et al., 2021).

119 **2.2 Studied material**

120 This study is based on palm fruit compressions collected from the STRI localities 0317
121 (11°14'N; 72°57'W), 0319 (11°66'N; 73°31'W), 0324 (11°62'N; 73°32'W), and La Puente-pit
122 (11°90'N; 72°30'W) (Gomez- Navarro et al., 2009; Wing et al., 2009), Cerrejón coal mines of
123 northern Colombia (Figure 1). Although the fossil fruits do not preserve internal structure,
124 multiple morphological characters (e.g., shape, longitudinally oriented fibers, inconspicuous
125 longitudinal ridges, and very large size) and the regular co-occurrence with palm leaves are
126 indicative of an Arecaceae affinity (Gomez- Navarro et al., 2009). All specimens are deposited
127 at the Paleontological Museum Royo y Gómez, Servicio Geológico Colombiano, Bogotá,
128 Colombia.

129 Examples of extant borings on palm fruits and seeds were surveyed from the available
130 entomological and forestry literature. We focused on examples that described the feeding
131 behaviour of insect larvae and/or adults on palm fruits and reported plant host and insect species,
132 plant tissue consumed, features of the borings such as size, shape and circularity of the exit
133 holes, and reaction rims (see electronic supplementary material). Additionally, the herbarium
134 collections of the Jardín Botánico Joaquín Antonio Uribe (JAUM), in Medellín, Colombia, were

135 surveyed for modern examples of borings in palm fruits. Although borings in fossil palm fruits
136 have been previously described (El Hedeny et al., 2021; Moreno-Dominguez et al., 2016), these
137 are bivalve-mediated and morphologically very distinct from the damage here described, and
138 were not considered in further comparisons.

139 Specimens were examined with a Nikon SMZ 1500 stereoscope. Fossils and botanical
140 samples were photographed using Canon EOS 5DS R and Canon EOS 5D Mark III cameras,
141 respectively. Reversible image adjustments such as white balance, temperature and contrast were
142 made using Adobe Photoshop 2021. The diameter of the borings and associated reaction tissues
143 were measured using ImageJ v1.53e (Schneider et al., 2012). The displacement of each boring,
144 or change in position in the three-dimensional space, was tracked based on the relative change in
145 position of the centroid of each boring from the outer to the innermost layers of the mesocarp.

146 **2.3 Bruchine systematics**

147 We treat Bruchinae (seed beetles) as a subfamily of Chrysomelidae (leaf beetles) (Farrell
148 and Sequeira, 2004; Gómez-Zurita et al., 2008; Morse, 2014) and have adjusted previously
149 published ranks (*sensu* Nilsson and Johnson, 1993) accordingly. Thus, palm bruchines
150 (considered as tribe Pachymerini by Nilsson and Johnson, 1993) are here adjusted to subtribe
151 Pachymerina (see Archibald et al., 2014 for a similar approach). Pachymerines have a fossil
152 record that extends to the Late Cretaceous (Poinar, 2005), ca. 20 million years earlier than the
153 fossil described here.

154 **3. Results**

155 **3.1 Borings description**

156 Of 17 palm fruits recovered from the Cerrejón locality, one specimen displayed evidence
157 of borings. Six borings were recorded in the fibrous mesocarp of specimen STRI-9938 (Plate I;
158 numbered arrows), described as cf. *Cocos* (Gomez- Navarro et al., 2009). The borings are
159 circular (3.8–7.5 mm in maximum diameter) to broadly elliptical (7.5–8.3 mm long by 3.6–3.8
160 mm wide) in shape, discernable by an outer ring of indurated reaction tissue 0.3–0.9 mm in
161 width. This fruit is preserved as an outer impression (Plate I, 1, 3) and a modular, inner
162 compression (Plate I, 2, 4) that we interpret as the innermost mesocarp layers. The preservation
163 of the mesocarp in distinct layers allows tracking of the borings in a three-dimensional manner.

164 Four borings (Plate I; arrows 1, 2, 4, 6) are easily tracked from the outer surface and into
165 the inner layers of the mesocarp. The borings show an overall expansion in diameter from the
166 outer (Plate I, 1, 3) to the innermost layers (Plate I, 2, 4), as well as some displacement in the
167 three-dimensional space. Boring 1 is 3.8 mm in diameter at the outer layer and has a 0.4 mm
168 thick reaction tissue; in the inner section, the centroid of the boring shifts 8.6 mm exmedially and
169 5 mm basally, and its diameter and reaction tissue expand to 6.3 mm and 0.7 mm, respectively.
170 Boring 2 is 5.3 mm in diameter and bears a reaction rim 0.4 mm thick at the outer section; its
171 diameter expands to 7.5 mm and reaction tissue increases to a thickness of 0.5 mm in the inner
172 section. The centroid of boring 2 shifts 2.9 mm admedially and 4 mm basally in the inner section
173 of the fruit. Boring 4 does not change in diameter (7.5 mm long by 3.6 mm wide) or reaction rim
174 thickness (0.3 mm thick) across the outer and inner mesocarp layers but it shifts 8.3 mm
175 exmedially and 14.9 mm basally in the inner mesocarp layer. Boring 6 is 4.6 mm across and has
176 a 0.3 mm thick reaction rim; this boring expands to 6.8 mm in diameter on the inner mesocarp
177 layer and shifts 1.5 mm admedially and 12.5 mm basally. The reaction rim of boring 6 is 0.9
178 mm, on the inner layer.

179 Borings 3 and 5 are only visible in the outer but not the inner sections. Boring 3 is 4.8
180 mm in diameter, associated with a 0.4 mm thick reaction rim. Boring 5 is 8.3 mm long by 3.8
181 mm wide, with a reaction rim 0.3 mm in width. Since these two borings are not discernible in the
182 inner sections, the displacement in the three-dimensional space (if any) cannot be ascertained.

183 Palm bruchine borings on modern palm fruits (Plate II) are similar in size, position, and
184 reaction tissue thickness. Although smaller borings are seen in modern specimens, their position
185 at the medial portion of the fruits is consistent with those seen in the fossil coconut. Furthermore,
186 the thickness of the reaction tissue is similar in both the fossil specimen (0.3–0.9 mm) and
187 modern palm fruits (0.1–0.6 mm). Even though not all borings on palm fruits induce such a thick
188 reaction rim (Plate II, 1–4), most of them do (Plate II, 5–10). Importantly, in all instances where
189 there is a thick reaction tissue (Plate II, 5–6, 8–9), these are outwardly directed, as seen in the
190 fossil coconut borings (Plate I).

191 **3.2 Modern palm-boring insects**

192 We compiled 244 cases describing the feeding by beetles on the fruits and seeds of extant
193 palms (electronic supplementary material). Each entry is a unique beetle-to-palm association,
194 with curated taxonomy for both groups, geographic location, and –when available– size of the
195 insect exit hole (or entrance hole for Curculionidae; see Discussion section 4.1 below). These
196 cases included 21 species of Chrysomelids (leaf beetles), 15 species of Curculionidae (weevils)
197 and one species of Cerambycidae (longhorned beetles).

198 **4. Discussion**

199 **4.1 The culprit**

200 The borings preserved in the compression fossil of a coconut from Cerrejón are
201 consistent with those made by living palm bruchines of the subtribe Pachymerina. This
202 interpretation is based on the unique combination of size, number, location, presence of a thick
203 and outwardly positioned reaction rim, and plant host selection. The thickened, outwardly flared
204 reaction tissue along the periphery of the borings define an exit hole, typical for bruchines, rather
205 than an entrance hole, as is the case with the feeding behavior of bark beetles of the tribe
206 Scolytini (see below). The size of Pachymerina exit holes depends on the size of the insects
207 fabricating the borings (Nilsson and Johnson, 1993). Exit holes as small as 3 mm and as large as
208 8 mm in diameter have been reported across species of Pachymerina (see electronic
209 supplementary material for a complete list of palm bruchine species, their palm hosts and –when
210 available– exit hole size), whereas other potential culprits, such as weevils, produce exit holes of
211 much smaller diameter. The location of these borings in the central portion of the fruit also
212 allows the elimination of other potential culprits such as the longhorn beetle *Pterolophia*
213 *apiceplagiata* Breuning (see below). Furthermore, it is common for multiple palm bruchines to
214 develop within the same seed and produce multiple exit holes, as in the fossil coconut, especially
215 among larger seeds (Draxler et al., 2011; Lau et al., 2014; Pedersen, 1995) such as those of
216 *Cocos*. The occurrence of multiple exit holes in close proximity to each other suggests that the
217 developing larvae were not cannibalistic, a behavior observed among some species of bruchids,
218 such as *Megaceros discoidus* (Say), when resources are limited (Wang and Kok, 1986).

219 Alternative insect culprits could be affiliated to other coleopteran lineages such as
220 Curculionidae or Cerambycidae, or even piercing-and-sucking hemipterans such as leaf-footed
221 bugs (Coreidae) or seed bugs (Lygaeidae). However, the damage made by these insects is
222 inconsistent with the overall morphology, size, number of borings, reaction tissue or position –or

223 a combination of these—seen in the fossil coconut. Bark beetles in the subtribe Scolytini such as
224 *Dactylotrypes*, *Hypothenemus* and especially *Coccotrypes* are particularly prolific consumers of
225 palm seeds (Atkinson and Peck, 1994; Beaver, 1987; Draxler et al., 2011; Jansen et al., 2010;
226 LaBonte and Takahashi, 2012; Siviero and Montesdeoca, 1990; Spennemann, 2019; Villalobos
227 and Blanco-Metzler, 2006; Wood, 1986). These female beetles bore into and oviposit within the
228 seed (Jansen et al., 2010; Wood, 1986), wherein multiple generations and up to 100 individuals
229 can coexist, depending on the size of the seed (Spennemann, 2019). When seed resources are
230 exhausted, adults emerge through the same entrance hole that the first female beetle created
231 (Spennemann, 2019). These entrance holes are usually less than 1 mm in diameter (Draxler et
232 al., 2011; Jansen et al., 2010) and rarely approach 1.5 mm (Siviero and Montesdeoca, 1990) (see
233 electronic supplementary material for a complete list of palm seed predating curculionids, their
234 palm hosts and, when available, entrance hole size). Although it is common for multiple females
235 to attack the same seed (Jansen et al., 2010) and abandon multiple holes as in the fossil coconut,
236 the hole diameters are 2.5–5 times smaller than the borings described herein. Importantly, as the
237 boring is excavated by the mouthparts of the female while entering the seed (Anderson, 1995),
238 the reaction tissue is not outwardly flared but rather inwardly directed. We consider that it is
239 consistent with what is known about the life habits of weevils, and this group is discarded as the
240 culprit for the damage seen in the Paleocene coconut from Cerrejón.

241 Although less common, longhorned beetles also feed on palm seeds. A report of one such
242 cerambycid, *Pterolophia apiceplagiata* Breuning, feeding on coconut fruits in Sumatra,
243 Indonesia (de Chenon et al., 1991), shows that females are likely to oviposit at the remnant tissue
244 surfaces of the calyx, given that the entrance hole made by the larva is always at the attachment
245 of the fruit to the rachillae (de Chenon et al., 1991). The larva bores through the fibrous

246 mesocarp until it reaches the germination pores, where it preferentially feeds. After obtaining
247 sufficient nutrition, the larva bores a 6 cm long gallery from the germination pores to the
248 periphery, creating a pupal chamber 18–28 mm long by 14.4–21 mm wide, followed by
249 pupation, and ending in the adult exiting the fruit through an elliptical hole 5 mm long by 3.9
250 mm wide (de Chenon et al., 1991). The size of this cerambycid exit hole is slightly less in
251 maximum dimension than the elliptical borings seen in the fossil coconut (7.5–8.3 mm long by
252 3.6–3.8 mm wide); however, only a single larva develops per fruit (de Chenon et al., 1991). It is
253 likely that the number of cerambycid larvae that develop on each fruit is restricted due to
254 nutritional constraints imposed by the coconut mesocarp tissues available, as the larvae cannot
255 penetrate the endocarp (de Chenon et al., 1991). Notably, in the fossil coconut, there is no
256 evidence of a boring near the point of attachment to the rachillae, and species of *Pterolophia* are
257 not found in the New World. Taking these observations into account, we consider that the
258 damage found in the fossil is not attributable to a cerambycid culprit.

259 Sufficiently similar lesions are caused by the coreid bug *Pseudotheraptus wayi* Brown on
260 coconuts of East Africa (Brown, 1955; Way, 1953). Females oviposit on the surfaces of flowers
261 or young fruits, and both nymphal and adult stages feed on young stems, leaves, inflorescences
262 and, especially, fruits (Egonyu et al., 2013). The damage on the fruits consists of piercing-and-
263 sucking punctures which, due to the toxic saliva of the coreid, induce sunken lesions and
264 surrounding necrotic tissue in an unorganized manner (Way, 1953). The lesions widen as the
265 fruit expands, and occasionally develop into deep slits into the exocarp (Way, 1953). Although
266 no measurements are reported for these lesions, given the necrotic nature of the damage, the lack
267 of a reaction rim along the lesion periphery, and its generalized slit-like shape, we consider that
268 the damage found on the fossil coconut is not that of a coreid.

269 Most seed bugs (Lygaeidae) are lacerate-flush feeders (Burdfield-Steel and Shuker,
270 2014), feeding mainly on mature seeds and occasionally on developing fruits (Sweet, 1960;
271 Sweet et al., 2000). Their feeding behavior creates lesions in parenchymatous tissues, emptying
272 cells of their content as insects pierce-and-suck through the surface (Panizzi et al., 2021), and
273 often these insects aggregate in large groups that can cause wilting and deformation of
274 developing fruits (Burdfield-Steel and Shuker, 2014; Sweet et al., 2000). There is no evidence of
275 wilting or deformation on the fossil coconut that could be attributed to group feeding of seed
276 bugs; moreover, the size, morphology, and reaction rim of the exit holes does not match that of
277 piercing-and-sucking marks.

278 **4.2 Plant host specificity**

279 Overall, bruchines are typically monophagous or oligophagous (Borowiec, 1987; Kergoat
280 et al., 2004), and exhibit strong conservatism in host-plant use (Kergoat et al., 2007). Studies on
281 host preference have shown that oviposition substrate affects the evolution of host-plant
282 affiliation and dietary specialization (Morse and Farrell, 2005), and presently, palm bruchines
283 feed almost exclusively on palm seeds (Nilsson and Johnson, 1993). The only possible exception
284 is a dubious record (Johnson et al., 1995) of *Pachymerus abruptestriatus* (Gyllenhal) in the seeds
285 of *Diospyros* sp. (Ebenaceae) (Bondar, 1941). The borings on the cf. *Cocos* fruit from the
286 Paleocene of Colombia are the first known case of palm bruchine feeding behaviour in the fossil
287 record.

288 Inferences on the origin and history of coevolutionary relationships between herbivorous
289 insects and their hosts are typically based on phylogenetics and divergence-age estimations of
290 living plant hosts and their pests (e.g., Kergoat et al., 2015), but rarely have there been fossil
291 occurrences for support. The specific feeding behaviour of palm bruchines observed in the

292 borings of the cf. *Cocos* fruit shows that palm bruchines have used (and likely lived in) palm
293 fruits and seeds since minimally 60 Ma and documents a coevolutionary relationship in deep
294 time. The association between palm bruchines and palms could have arisen as early as the Late
295 Cretaceous, as the fossil record of both palms (Matsunaga and Smith, 2021) and palm bruchines
296 (Poinar, 2005; see 4.3 below) extend back to this time. Early records of crown-group Cocoseae
297 include permineralized fruits of subtribe Attaleinae recovered from Danian deposits in Patagonia,
298 Argentina (Futey et al., 2012), indicating that the Cocoseae most likely diverged during the Late
299 Cretaceous, as is also suggested by divergence-age estimations (Meerow et al., 2015).

300 While it would be impossible to pinpoint a particular genus within Pachymerina as the
301 culprit of the damage found in the cf. *Cocos* fruit, most of the seeds of the tribe Cocoseae
302 (coconuts, oil and jelly palms) are predated by *Pachymerus* (Johnson et al., 1995; and see
303 electronic supplementary material). *Pachymerus bactris* L. and *P. nucleorum* Fabricius are the
304 only palm bruchines reported in *Cocos nucifera* L. seeds (see electronic supplementary material).
305 Our finding highlights the evolutionary connection between Pachymerina and palms in deep-
306 time, which extends to the early evolution of modern-like Neotropical rainforests (Carvalho et
307 al., 2021). The borings found on the cf. *Cocos* fruit also add yet another form of ecological
308 interaction between primary producers (vascular plants) and consumers (herbivorous insects) in
309 the Paleocene rainforests of northern South America (Carvalho et al., 2021). Moreover, the
310 pachymerine seed beetle–coconut palm association emphasizes that host-specific interactions are
311 a defining feature of lowland tropical rainforests (Dyer et al., 2007).

312 **4.3 Biogeography and fossil record of palm bruchines**

313 The occurrence of palm bruchines in the Cerrejón flora is consistent with their current
314 Neotropical distribution and previous findings in the Americas. Fossils of Pachymerina are

315 known from the Late Cretaceous of southern Canada (Poinar, 2005), and are a common element
316 in the early Eocene Okanagan Highland fossil sites of the Pacific Northwest of North America
317 (Archibald et al., 2014; Archibald and Mathewes, 2000). Pachymerina are also found in the late
318 Eocene Florissant Formation, Colorado (Archibald et al., 2014; Kingsolver, 1965) and the early
319 Neogene of the Dominican Republic (Poinar, 1999). A single species preserved in Late Eocene
320 Baltic amber remains the only known record of palm bruchines outside the Americas (Legalov,
321 2016). The presence of an extralimital occurrence of a bruchine outside of the Americas is a
322 common biogeographic pattern in Cenozoic insect lineages, whereby a formerly cosmopolitan
323 taxon is currently restricted to a region of the Southern Hemisphere, a prominent example of
324 which is the tsetse fly *Glossina* (Lambrecht, 1980). The Cerrejón borings on the arecaceous fruit
325 indicate that palm bruchines lived in northern South America by the middle–late Paleocene,
326 where palms were already one of the most abundant and diverse elements of the flora (Gomez-
327 Navarro et al., 2009; Wing et al., 2009). The ages and localities of known fossil occurrences of
328 Pachymerina suggest that their current Neotropical distribution is a relict from a much wider
329 distribution observed during warmer Cretaceous and Paleogene ages. The fossil record may also
330 suggest a north-to-south migration pattern, perhaps through the Late Cretaceous/Paleocene
331 Proto-Greater Antilles Island bridge (Poinar, 2005). Nonetheless, the fossil record of palm
332 bruchines is scant, and therefore a detailed interpretation of their biogeographic history remains
333 tentative.

334 **5. Conclusion**

335 Borings on an early cf. *Cocos* fruit from the middle–late Paleocene Cerrejón Formation
336 provide the earliest record of the ecological interaction between palm bruchine seed beetles
337 (Pachymerina) and coconut palms. This suggests that palm bruchines have consistently been

338 seed predators of palm fruits for minimally 60 Ma, and highlights host-specific interactions –that
339 likely act as plant diversity-promoting agents– as a defining feature of modern Neotropical
340 rainforests.

341 **Data availability.** The compilation of modern instances of beetles feeding on palm fruits and
342 seeds is provided in the electronic supplementary material. Further details on samples and
343 localities can be accessed through the Geologic sample Database of the Smithsonian Tropical
344 Research Institute at <https://biogeodb.stri.si.edu/jaramillosdb/web/fossils/>

345 **Declaration of competing interests.** The authors declare that they have no known competing
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356

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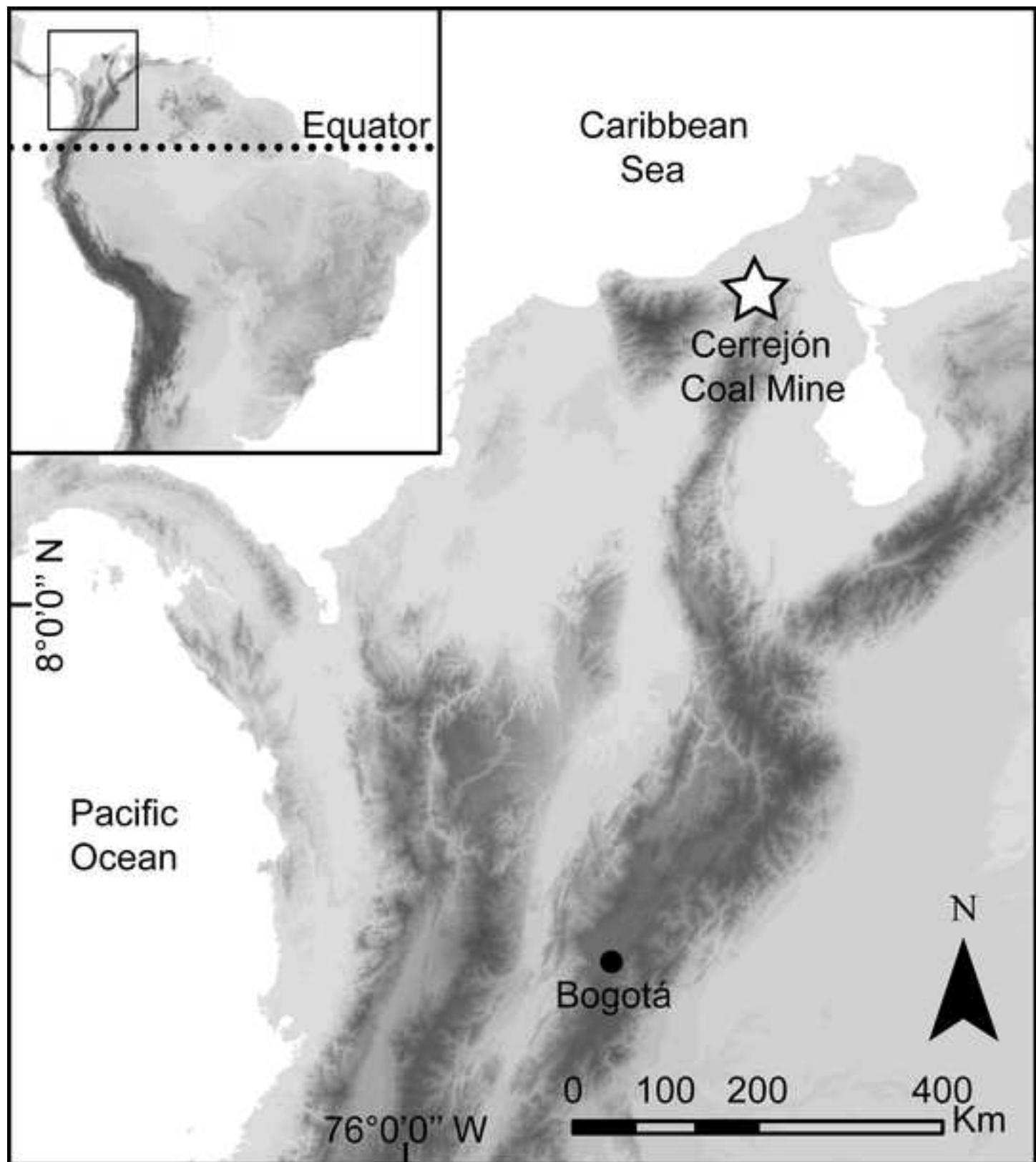
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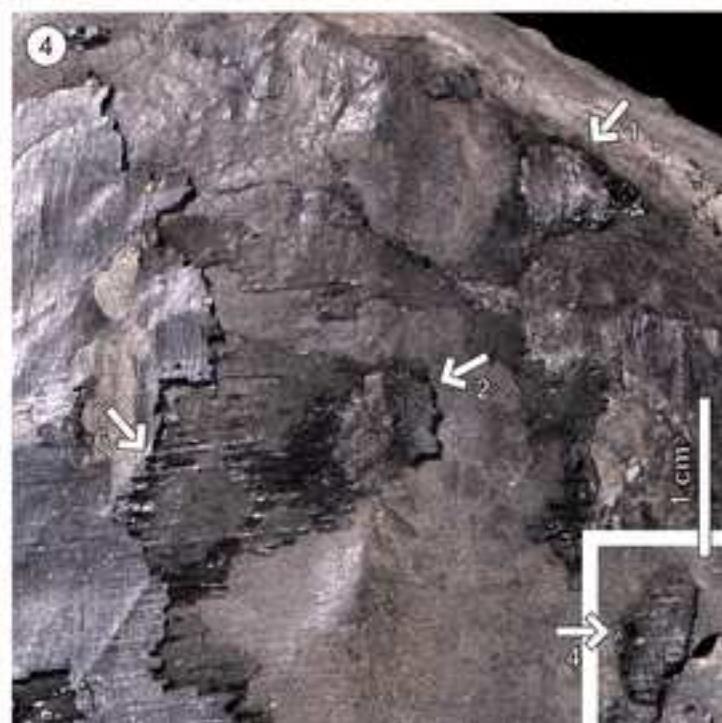
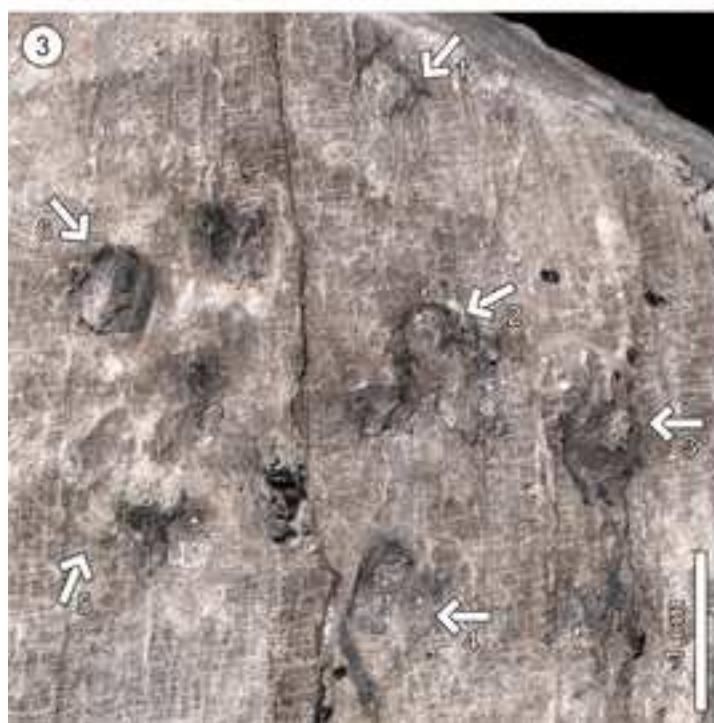
583 **Figure 1.** Collection site of cf. *Cocos* fruits of the Cerrejón Formation, Colombia, indicated by a
584 star.

585 **Plate I.** Palm bruchine borings (numbered arrows) on cf. *Cocos* fruit (STRI-9938) of the
586 Cerrejón Formation. Given that the fossil specimen is roughly fractured transversely, outer (1
587 and 3) and inner (2 and 4) mesocarp sections are discernible. Note that all the borings are circular
588 or broadly elliptical, with a thick reaction tissue flanking the periphery of the damage. Borings 1,
589 2, 4 and 6 are observable at the outer and inner mesocarp sections, allowing for a tracking of the
590 damage in a three-dimensional manner. By contrast, borings 3 and 5 are visible along the outer
591 but not the inner section. Borings are numbered clockwise.

592 **Plate II.** Palm bruchine borings on modern palm fruits. **1.** Tunnel on the mesocarp of
593 *Astrocaryum malybo* H. Karst. (JAUM 073119), enlarged at **2** (see arrow). **3.** Hole on the
594 exocarp and mesocarp of *Bactris brongniartii* Mart. (JAUM 066644), enlarged at **4**. **5.** Circular
595 hole on the exocarp and mesocarp of *Bactris chocoensis* R. Bernal, Galeano, Copete & Cámara-
596 Leret (JAUM 079698), enlarged at **6** (note the thick and outward flaring reaction tissue). **7.**
597 Circular hole on the calyx remnants, exocarp and mesocarp of *Bactris macana* (Mart.) Pittier
598 (JAUM 050273). **8.** Two circular holes of different sizes on the exocarp and mesocarp of
599 *Chamaedorea pinnatifrons* (Jacq.) Oerst. (JAUM 007592); we interpret the small hole (marked
600 with an arrow) as the entrance boring made by a larval stage bruchine, while the large one
601 represents the exit hole made by the adult. **9.** Circular holes on the exocarp and mesocarp of two
602 fruits of *Prestoea ensiformis* (Ruiz & Pav.) H.E. Moore (JAUM 058644) (note the thick reaction
603 tissue surrounding the hole of the fruit on the right). **10.** Circular and broadly elliptical holes on
604 the exocarp, mesocarp and endocarp of four fruits of the palmetto *Sabal mauritiiformis* (H.
605 Karst.) Griseb. & H. Wendl. (JAUM 072846), together with the palm bruchines that made the

606 damage (possibly *Caryobruchus gleditsiae* L.; see electronic supplementary material and Nilsson
607 and Johnson, 1993). Note that there is one palm bruchine still inside a fruit (marked arrow).







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Supplementary Material

**Supplementary material - palm fruit-seed beetle
borers.xlsx**