

Preservation and diagenesis of soft-bodied fossils and the occurrence of phosphate-associated rare earth elements in the Cambrian (Wulian) Spence Shale Lagerstätte

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ABSTRACT

The Spence Shale Lagerstätte is a middle Cambrian (Miaolingian Series; Wulian Stage) fossil deposit in northern Utah and southern Idaho, USA. At present, it is known to preserve 89 species, from at least 10 phyla, of biomineralizing and soft-bodied taxa, and represents the only major Lagerstätte of Wulian age in Laurentia outside the Canadian Rockies. In addition to taxonomic diversity, the Spence Shale exposures represent a variety of depositional environments along a continental shelf, which have been affected by several diagenetic events, making it a compelling setting to improve our understanding of taphonomic and diagenetic pathways as they affect the broader Burgess Shale-type (BST) preservation. We present analyses of 28 specimens from six localities using scanning electron microscopy (SEM) and integrated energy dispersive X-ray spectrometry (EDS), together with comparisons of published data. Taxa are predominantly preserved as a combination of carbonaceous compressions, iron oxide and pyrite replacement, but eldoniids of the genus *Eldonia* consistently show enrichment in rare earth elements (REE). The REE, predominantly lanthanum and cerium, are enriched in the phosphate mineral monazite, which occurs in the inner coil of *Eldonia ludwigi*. This pattern, seen in all sampled *Eldonia*, differs from previous reports of REE association in BST preservation in being limited to one tissue type within a specific identifiable taxon. REE enrichment of tissues can occur during the lifetime of an individual or during fossilization, and we present both possible pathways along with an outline of taphonomic processes leading to the preservation of soft-bodied fossils from the Spence Shale.

1. Introduction

The Spence Shale Lagerstätte (referred to as Spence Shale from here on) is a middle Cambrian (Miaolingian Series, Wulian Stage) Burgess Shale-type (BST) Lagerstätte that has recently been the center of renewed attention for its taxonomic diversity and complex preservation. Located in northern Utah and southern Idaho, U.S.A., it is the oldest of the five Utah Cambrian BST Lagerstätten (Kimmig et al., 2019) and the only one located outside the House Range and Drum Mountains (Robison et al., 2015; Lerosey-Aubril et al., 2018; Kimmig et al., 2019). The

fauna of the Spence Shale comprises a diverse community of, in order of generic abundance, trilobites, soft-bodied arthropods, brachiopods, echinoderms, molluscs, sponges, lophophorates, scalidophorans, hemichordates, lobopodians, algae, potential cyanobacteria, and multiple problematic taxa (e.g., *Banffia*, *Eldonia*, and *Siphonactum*) (Kimmig et al., 2019). In recent years, numerous new taxa from the Spence Shale have been described and several additional taxa have been revised, bringing the faunal diversity to at least 89 species in 74 genera (e.g., Legg and Pates, 2017; Kimmig et al., 2017; Pates et al., 2018; Wen et al., 2019; Sundberg, 2020; Whitaker et al., 2020; Kimmig and Selden,

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2021). This makes the Spence Shale one of the most diverse BST deposits (e.g., Kimmig et al., 2019; Nanglu et al., 2020; Yang et al., 2021a). The depositional conditions of the Spence Shale have been investigated, revealing rapid fluctuations from oxic to dysoxic benthic environments and moderate bioturbation (Garson et al., 2012; Kloss et al., 2015). However, investigations of the taphonomy of the member have been limited to broad overviews or specific taxonomic groups, further restricted by sampling from just a few localities. Gaines et al. (2008)

reported the prevalence of carbonaceous films as evidence of a common taphonomic pathway among BST deposits, and Broce and Schiffbauer (2017) documented variation in the preservation pathways of vermiciform fossils, including kerogenization, pyritization, aluminosilicification, and phosphatization. Broce and Schiffbauer's (2017) samples of Spence Shale fossils, however, only included examples from the Wellsville Mountains (Fig. 1). The Wellsville localities have yielded the majority of soft-bodied fossils from the Spence Shale and the greatest

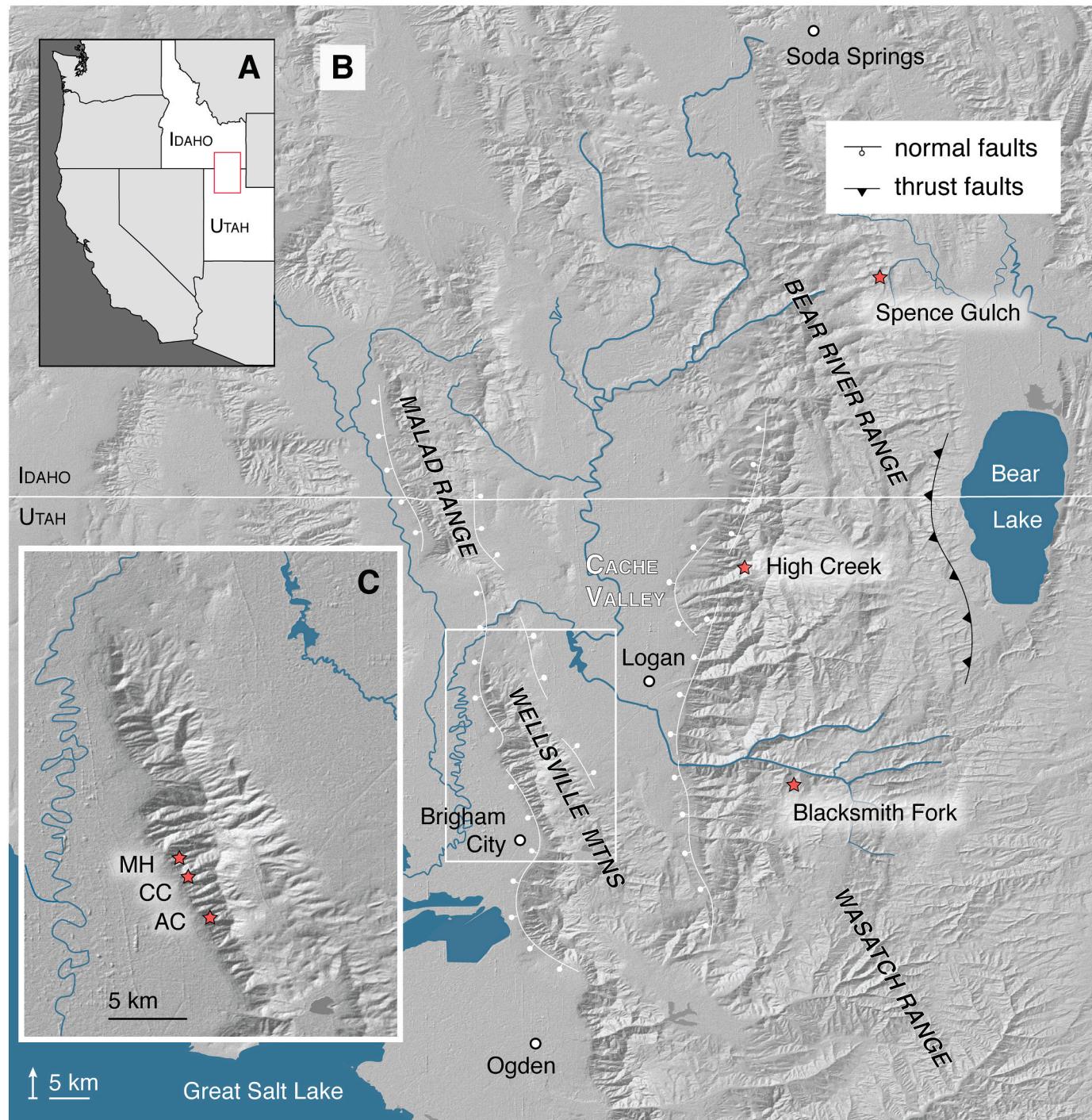


Fig. 1. Location and localities of the Spence Shale Lagerstätte. A) Map of the western United States of America showing location of the Spence Shale. B) Relief map of northern Utah and southern Idaho showing prominent localities and geologic features relating to the Spence Shale. C) Close-up of inset from (B), showing position of localities within the Wellsville Mountains. MH, Miners Hollow; CC, Cataract Canyon; and AC, Antimony Canyon. Study sites are protected, but exact location information will be shared with qualified researchers upon request. We acknowledge the use of imagery from the NASA Worldview application (<https://worldview.eosdis.nasa.gov>), part of the NASA Earth Observing System Data and Information System (EOSDIS).

diversity (Kimmig et al., 2019; Whitaker and Kimmig, 2020), but the taxonomic distributions and taphonomic conditions at other Spence Shale localities can offer new insights.

The Spence Shale differs from many other BST Lagerstätten in the preservation of depositional environments varying from shallow shelf carbonates to deeper water shales (Kimmig et al., 2019). This environmental range, as well as the occurrence of soft-tissue preservation at multiple stratigraphic horizons within exposures, makes the Spence Shale ideal for the investigation of taphonomic response to different settings. Our analyses using scanning electron microscopy (SEM) and integrated energy dispersive X-ray spectrometry (EDS) offer insights into the initial stages of fossil preservation and subsequent diagenetic history at different Spence Shale localities, their impact on the taphonomy of soft-tissues, and the importance of understanding these processes across BST Lagerstätten.

2. Geologic Context

The Spence Shale crops out in northern Utah and southern Idaho at multiple exposures in the Wellsville Mountains, the Bear River Range, the Wasatch Range, and the Malad Range (Fig. 1). The Spence Shale is the middle member of the Langston Formation, conformably overlying the Naomi Peak Limestone Member and underlying the High Creek Limestone Member at the type locality in Spence Gulch. The Langston Formation (type locality: Blacksmith Fork) conformably overlies the Geersten Canyon Quartzite and is in turn conformably overlain by the Ute Formation (Walcott, 1908; Kimmig et al., 2019). The Spence Shale straddles the *Albertella* to *Glossopleura* trilobite biozones, and is therefore slightly older than the Burgess Shale in Canada (Kimmig et al., 2019). It has been interpreted as representing deposition in the middle to outer detrital belt of a (presently) westward-oriented carbonate platform (Robison, 1991; Liddell et al., 1997). There is an overall proximal to distal trend towards the NE, with extensive soft-tissue preservation and dolomitization in the Wellsville Mountain localities, which decrease in the Bear River and Malad Range localities (Kimmig et al., 2019).

The Langston Formation has undergone extensive dolomitization, which extends into the Spence Shale (Deiss, 1938; Whitaker et al., 2020). The timing of this dolomitization has been interpreted as eogenetic to mesogenetic, and controlled largely by facies, correlating strongly with the upper peritidal deposits under a seepage reflux model (Butterbaugh, 1982). However, this model does not account for the significant dolomitization of the Spence Shale Member.

The Wasatch Range is approximately centered in the Sevier Wyoming-Idaho-Utah thrust belt, formed by the Sevier and Laramide orogenies. The Paris thrust fault, which runs roughly north-south on the eastern side of the Bear River Range (Valenti, 1982), creates a 3600 ft stratigraphic displacement, placing the Langston Formation in contact with the Furongian St. Charles Formation in the Laketown Quadrangle (Valenti, 1982).

The Utah Basin and Range extension, beginning 17 mya, formed the modern topography of the Spence Shale outcrops. These features are high-angle normal faults, creating the horst block mountain ranges and fault scarps enclosing the Cache Valley, a half-graben basin (Williams, 1948; Evans and Oaks Jr., 1996; Black et al., 2000). The relevant faults are the East Cache Fault zone, the West Cache Fault zone, and the Wasatch Fault (McCalpin, 1994; Black et al., 2000). These faults remain active to the present, with multiple seismic events in the sub-recent record (McCalpin, 1994).

The Bonneville lake cycle began approximately 30 kya, and periodically rose and fell, filling the Cache Valley. The Bonneville Shoreline level (18,000 B.P.) represents the highest paleo-shoreline (Hylland, 2007) but, as reconstructed by Chen and Maloof (2017), it does not affect known localities of the Spence Shale.

3. Material and methods

Twenty-eight specimens representing 6 localities were analyzed using scanning electron microscopy (SEM) and integrated energy dispersive X-ray spectrometry (EDS): from Antimony Canyon ($n = 8$), Blacksmith Fork ($n = 1$), Cataract Canyon ($n = 3$), High Creek ($n = 2$), Miners Hollow ($n = 9$), and Spence Gulch ($n = 5$) (Table 1). Samples were selected from the University of Kansas Invertebrate Paleontology collections based on generic occurrence at multiple localities, and the availability of stratigraphic data. Specimens represent at least 6 phyla (Lobopoda, Arthropoda, Lophotrochozoa, Mollusca, Priapulida, Hemichordata) as well as problematic taxa. An additional 18 specimens from previous studies (Marshall et al., 2012; LoDuca et al., 2015; Broce and Schiffbauer, 2017; Whitaker et al., 2020; Kimmig and Selden, 2021; Moon et al., 2021) were added to the dataset, giving a total of 46 specimens (Table 1). Genera represented are *Acinocricus*, *Anomalocaris*, *Armillimax*, *Dioxycaris*, *Eldonia*, *Hurdia*, *Isoxys*, *Mollisonia*, *Selkirkia*, *Sphenoecium*, *Utahscolex*, *Waptia*, *Wiwaxia*, along with unidentified hirdiids, vermiform taxa, and bradoriids (Table 1). The specimens represent a range of tissue types, including cellular, cuticularized and sclerotized, and multiple tissue types are often present within a specimen. Selection within a genus was determined by which individuals preserve the most tissue types. Specimens were photographed immersed in alcohol (except where noted) using a Leica DMS 300 Digital Microscope and an Olympus SZX16 microscope. Whole specimen images were created by stitching together mosaics of smaller images using Adobe Photoshop. Specimens too large for microscope photography were photographed using a Pentax K-5 SLR digital camera, and image settings were adjusted in Adobe Photoshop.

SEM imaging of KUMIP 204390 and KUMIP 314041 was conducted using an FEI Versa 3D Dual Beam FIB-SEM (focused ion beam not used, electron imaging only) at the University of Kansas Microscopy and Analytical Imaging Laboratory. EDS analysis of KUMIP 204390 was conducted using an Oxford Instruments 80 mm² x-Max silicon drift detector. Analyses conducted on this instrument used an accelerating voltage of 10 keV and a spot size of 4.5, under low vacuum conditions.

SEM and EDS analyses of KUMIP 312404 and KUMIP 314037 were conducted using a Philips XL 30 environmental scanning electron microscope (ESEM) with a Princeton Gamma-Tech EDS at Yale University; images were generated at 10 keV with a spot size of 3 (roughly 2 nm in diameter) at a working distance ~10 mm.

SEM-EDS imaging for all the other samples in the present study was conducted at the X-ray Microanalysis Core Facility at the University of Missouri, using a Zeiss Sigma 500 VP variable pressure SEM with dual, co-planar Bruker XFlash 6|30 EDS detectors (30 mm²). Analyses conducted on this instrument used the following operating conditions, with minor variation: beam accelerating voltage of 20 keV and current of 40 nA, a 60 µm aperture for imaging and a 120 µm aperture for EDS analyses, a working distance of 16 mm ± 2 mm, and a chamber vacuum of 25 ± 5 Pa.

Conditions used in previous studies are reported in the respective publications. All elements detected were mapped. Maps of prominent elements are included in figures, and all maps of all specimens are included in Appendix 1.

Saleh et al. (2020a) compared the taphonomic signals of three exceptionally preserved biotas (the Burgess Shale, the Chengjiang Biota, and the Ordovician Fezouata Shale) using the presence and co-occurrences of five tissue types. Using their approach and dataset, we added data compiled herein from the Spence Shale samples for comparison (Appendix B), evaluating each eumetazoan genus for the presence of a biomineralized skeleton, sclerotized tissue (i.e. possessing an organically strengthened part or organ), unsclerotized cuticle (i.e. a non-cellular outer body surface that is either collagenous or formed by polymerized polysaccharides), soft cellular outer layer defining at least a part of the body (e.g. hyolith tentacles), and soft internal cellular organ/tissue (e.g. digestive or nervous systems) (Saleh et al., 2020a).

Table 1

Details of specimens analyzed in this study and previously published information.

Specimen #	Taxonomic Designation	Description	Study reference	Locality	Stratigraphic Info	Preservation
KUMIP 563151	<i>Selkirkia</i>	Tube	this study	Cataract Canyon	Float	Aluminosilification
KUMIP 563145	<i>Selkirkia</i>	Tube	this study	High Creek	n/a	Iron-association/pyrite
KUMIP 563150	<i>Selkirkia</i>	Tube	this study	Spence Gulch	n/a	Iron-association/pyrite
KUMIP 314158	<i>Selkirkia spencei</i>	Tube	this study	Miners Hollow	n/a	Iron-association/pyrite
KUMIP 144845	<i>Selkirkia spencei</i>	Tube	this study	Antimony Canyon (S side)	Gunther Quarry 3 to 5 m below top (in situ)	Kerogenization; Iron-association/pyrite
KUMIP 558518	<i>Dioxykaris</i>	Isolated Carapace	this study	Spence Gulch	n/a	Kerogenization; Iron-association/pyrite
KUMIP 135145	<i>Dioxykaris argenta</i>	Isolated Carapace	this study	Antimony Canyon (S side)	About 3 m below top of Spence Tongue	Kerogenization; Iron-association/pyrite
KUMIP 579390	Bradoriida	Shell	this study	Spence Gulch	5 m in situ	Calcium association, Kerogenization; Iron-association/pyrite
KUMIP 514259	<i>Sphenoecium wheelerensis</i>	Branching tubes	this study	Antimony Canyon	n/a	Kerogenization; Iron-association/pyrite
KUMIP 495314	<i>Sphenoecium wheelerensis</i>	Branching tubes	this study	Miners Hollow	Upper Cycle 3 - Koot. Quarry (in situ)	Kerogenization; Iron-association/pyrite
KUMIP 514260	<i>Sphenoecium wheelerensis</i>	Branching tubes concentrated debris	this study	High Creek	n/a	Iron-association/pyrite
KUMIP 490937	<i>Eldonia</i>	around degraded body	this study	Cataract Canyon	Float	REE-enriched phosphatization, Kerogenization
KUMIP 204370	<i>Eldonia ludwigi</i>	Soft tissue body with gut	this study	Antimony Canyon (S side)	3 to 5 m below top (excavated, likely)	REE-enriched phosphatization, Kerogenization
KUMIP 510274-510275	<i>Eldonia ludwigi</i>	Soft tissue body with gut	this study	Miners Hollow	Upper Cycle 3 - float	REE-enriched phosphatization, Kerogenization
KUMIP 204366	<i>Wiwaxia</i>	Isolated spine	this study	Antimony Canyon (S side)	3 to 5 m below top	Kerogenization; Iron-association/pyrite
KUMIP 490944	<i>Wiwaxia Acinocricus stichus</i>	Isolated spine	this study	Miners Hollow	Upper Cycle 3 - Koot. Quarry (in situ)	Kerogenization; Iron-association/pyrite
KUMIP 490967	<i>Acinocricus stichus</i>	Isolated spines	this study	Antimony Canyon	Float	Aluminosilification
KUMIP 491082	<i>Acinocricus stichus</i>	Isolated spines	this study	Cataract Canyon	Float	Iron-association/pyrite
KUMIP 314131	<i>Acinocricus stichus</i>	Isolated spines	this study	Miners Hollow	n/a	Calcium association; Iron-association/pyrite
KUMIP 491056	<i>Hurdiidae</i>	Feeding appendage	this study	Miners Hollow	Lower Cycle 6, 155' (in situ)	Kerogenization; Iron-association/pyrite; Calcium association
KUMIP 491057	<i>Hurdiidae</i>	Isolated Carapace	this study	Antimony Canyon	Float	Kerogenization; Iron-association/pyrite; Calcium association
KUMIP 314056	<i>Hurdia victoria</i>	Isolated Carapace	this study	Miners Hollow	Cycle 3; 130 ft. above base	Aluminosilification; Iron-association/pyrite
KUMIP 204779	<i>Sidneyia?</i>	Feeding appendage	this study	Miners Hollow	Upper	Kerogenization; Iron-association/pyrite
KUMIP 204777	<i>Sidneyia?</i>	Feeding appendage	this study	Antimony Canyon	n/a	Kerogenization; Iron-association/pyrite
KUMIP 579391	Bradoriida	Shell	this study	Spence Gulch	1 m (in situ)	Kerogenization; Iron-association/pyrite
KUMIP 557975	Vermiform	Possible body fossil with gut(?)	this study	Spence Gulch	Blacksmith Fork**Possibly Ute (in situ)	Aluminosilification; Iron-association/pyrite
USNM 768408	<i>Selkirkia Mollisonia symmetrica</i>	Tube	this study	Fm. (Deiss, 1938)	n/a	Kerogenization; Iron-association/pyrite
KUMIP 314041		Articulated body fossil	this study	Miners Hollow	Cycle 3; ~130' above base of Spence Shale	Kerogenization
KUMIP 312404	<i>Isoxys</i>	Carapace with eye and traces of appendages	this study	Miners Hollow	mid?	Iron-association/pyrite
KUMIP 314037	<i>Anomalocaris</i>	Articulated body fossil, with mouthparts and appendages, nearly complete	(previously unpublished data)	Miners Hollow	n/a	Pyrite, phosphatization of muscle near gut
KUMIP 490943	<i>Armillimax pauljamisoni</i>	Articulated body fossil, soft tissue with shell and gut tract,	Kimmig and Selden, 2021	Miners Hollow	Cycle 3; ~33–36 m above base (as measured by Kimmig)	Phosphatization in shell, minor phosphatization in soft tissue, Kerogenization
KUMIP 204390 / UMNH IP 4696	<i>Utahscolex ratcliffei</i>	Articulated body fossil with gut tract, missing anterior/posterior termination	Whitaker et al., 2020	Miners Hollow	Cycle 3, float	Iron-association/pyrite
KUMIP 490902	<i>Utahscolex ratcliffei</i>		Whitaker et al., 2020	Miners Hollow	Cycle 3, float	Calcium phosphate (plates); Aluminosilification

(continued on next page)

Table 1 (continued)

Specimen #	Taxonomic Designation	Description	Study reference	Locality	Stratigraphic Info	Preservation
		Articulated body fossil with gut tract, missing posterior termination				
KUMIP 314107	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	n/a	Phosphatized Kerogenization, pyrite associated aluminosilification, bladed barite
KUMIP 314111	Annelida? Ichnofossil, <i>Planolites</i> ?	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	Cycle 3; 130 ft. above base	
KUMIP 314112	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	n/a	Pyrite
KUMIP 314114	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Antimony Canyon	“middle” spence shale	Pyrite and monazite associations
KUMIP 314159	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	n/a	Pyrite
KUMIP 314186	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	Cycle 6; 160 ft. level	Kerogenization, pyrite-association, aluminosilification, monazite, calcite
KUMIP 314191	Animalia, unknown	Body fossil?	Broce and Schiffbauer, 2017	Miners Hollow	2/3 down from top	Pyrite and barite association
KUMIP 314202	Ichnofossil		Broce and Schiffbauer, 2017	Miners Hollow	n/a	Pyrite
KUMIP 314215	Annelida?	Body fossil?	Broce and Schiffbauer, 2017	Wellsville Mtn.	n/a	Pyrite and barite
KUMIP 377069	Ichnofossil <i>Sphenoecium wheelerensis</i>	Specimen with multiple tubes	LoDuca et al., 2015	Miners Hollow	n/a	Iron-association/pyrite
KUMIP 314270	<i>Leanchoilia superlata</i>		Marshall et al., 2012	Wellsville Mtn.	~10 ft. below top	Iron-association/pyrite
KUMIP 314028	<i>Waptia</i> cf. <i>fieldensis</i>	Body fossil		Miners Hollow	n/a	Iron-association/pyrite
KUMIP 314032		Body fossil with gut tract and eggs	Moon et al., 2021	Miners Hollow	n/a	Phosphatized (eggs)

Institutional abbreviations: KUMIP, Division of Invertebrate Paleontology, Biodiversity Institute, University of Kansas, Lawrence, KS, USA; USNM PAL, Paleobiology Department, Smithsonian National Museum of Natural History, Washington, D.C., USA; UMNH IP, Natural History Museum of Utah, Salt Lake City, UT, USA.

4. Results

The specimens from the Spence Shale show a variety of preservation modes but are predominantly preserved as kerogenized/carbonaceous compressions, pyrite framboids, and weathered iron oxides. A full summary of specimen preservation is provided in Table 1; all EDS elemental maps are included in Appendix A. Soft-tissue preservation occurs in at least three sequences at Miners Hollow, within the Cycle 3 and Cycle 6 intervals (MH3L, MH3U of Cycle 3, and MH6 of Cycle 6, of Garson et al., 2012). Samples from these intervals show that MH3L is mainly non-bioturbated, MH3U is weakly bioturbated, and MH6 shows variations in bioturbation. As documented previously, bioturbation increases up-section within each cycle (Garson et al., 2012). Cycle 3 contains fossils preserved via carbonaceous compressions ($n = 7$), iron mineral associations ($n = 5$), phosphatization ($n = 2$), aluminosilification ($n = 2$), and phosphatization enriched with REEs (hereafter, REE-associated phosphatization; $n = 1$) (Table 1). Cycle 6 contains iron associations ($n = 2$), carbonaceous compressions ($n = 2$), calcium association ($n = 2$), and REE-associated phosphatization ($n = 1$) (Table 1). At Miners Hollow, all modes of preservation observed in this study were represented. Fossils with sclerotized cuticle, e.g., non-biomineralized arthropod carapaces (Appendix A) and feeding appendages (Appendix A, Fig. 3C), *Wiwaxia* spines (KUMIP 490944, Appendix A), *Selkirkia* tubes (Fig. 4D), *Sphenoecium* tubes (Fig. 3A), and *Acinocricus* spines (Fig. 2B), are predominantly preserved as carbonaceous compressions with associated iron minerals. Cellular tissue, e.g., *Eldonia* (Fig. 5 C,D), the everted pharynx of *Utahscolex* (KUMIP 490902; Whitaker et al., 2020) and the eggs of *Waptia* cf. *W. fieldensis* (KUMIP

314032; Moon et al., 2021), exhibit additional modes of preservation, including phosphatization and aluminosilification (Fig. 5C, D). Preservation quality and diversity is similar in the Cycle 3 and Cycle 6 intervals.

All modes of preservation are also represented at Antimony Canyon. Sclerotized cuticle is preserved through carbonaceous compressions and pyritization, e.g., non-biomineralized arthropod carapaces (KUMIP 491057, Appendix A; KUMIP 135145, Appendix A) and feeding appendages (Fig. 2A), *Wiwaxia* spines (Fig. 2D), *Selkirkia* tubes (Fig. 4E), and *Sphenoecium* tubes (KUMIP 514259, Appendix A). An isolated *Acinocricus* spine, KUMIP 490967 (Appendix A) is preserved by aluminosilification. The cellular tissue of *Eldonia* (Fig. 5A) is preserved by REE-associated phosphatization. An isolated *Hurdiid* carapace (KUMIP 491057) has a patchy association with Ca (Appendix A).

Cataract Canyon shows more limited preservation pathways, though this may reflect unequal sampling, and thus fewer available samples, within the Wellsville Mountains compared to other localities (Table 1; Whitaker and Kimmig, 2020). Sclerotized *Selkirkia* tubes (Fig. 4A) and *Acinocricus* spines (KUMIP 491082; Appendix A) are preserved mainly through pyritization with restricted areas as carbonaceous compressions. The *Eldonia* specimen (Fig. 5B) from Cataract Canyon shows the REE-associated phosphatization that also occurs in examples from elsewhere.

The sample of soft-bodied preservation from High Creek is limited to two taxa, *Selkirkia* (Fig. 4B) and *Sphenoecium* (Appendix A). Both are preserved through pyritization weathered to iron oxide. The preservation quality is poor, representing only the outline of the specimens.

Selkirkia (USNM 768408; Fig. 4F) is the only known genus showing soft-bodied preservation from Blacksmith Fork (based on just six *Selkirkia* specimens at the USNM). It is preserved mainly through pyritization, with very limited carbon outlines (Fig. 4F). The entire exposure of the Langston Formation at Blacksmith Fork is just 9 m, in contrast to other Spence Shale exposures which exceed 100 m at Oneida Narrows and Miners Hollow (Liddell et al., 1997; Kimmig et al., 2019).

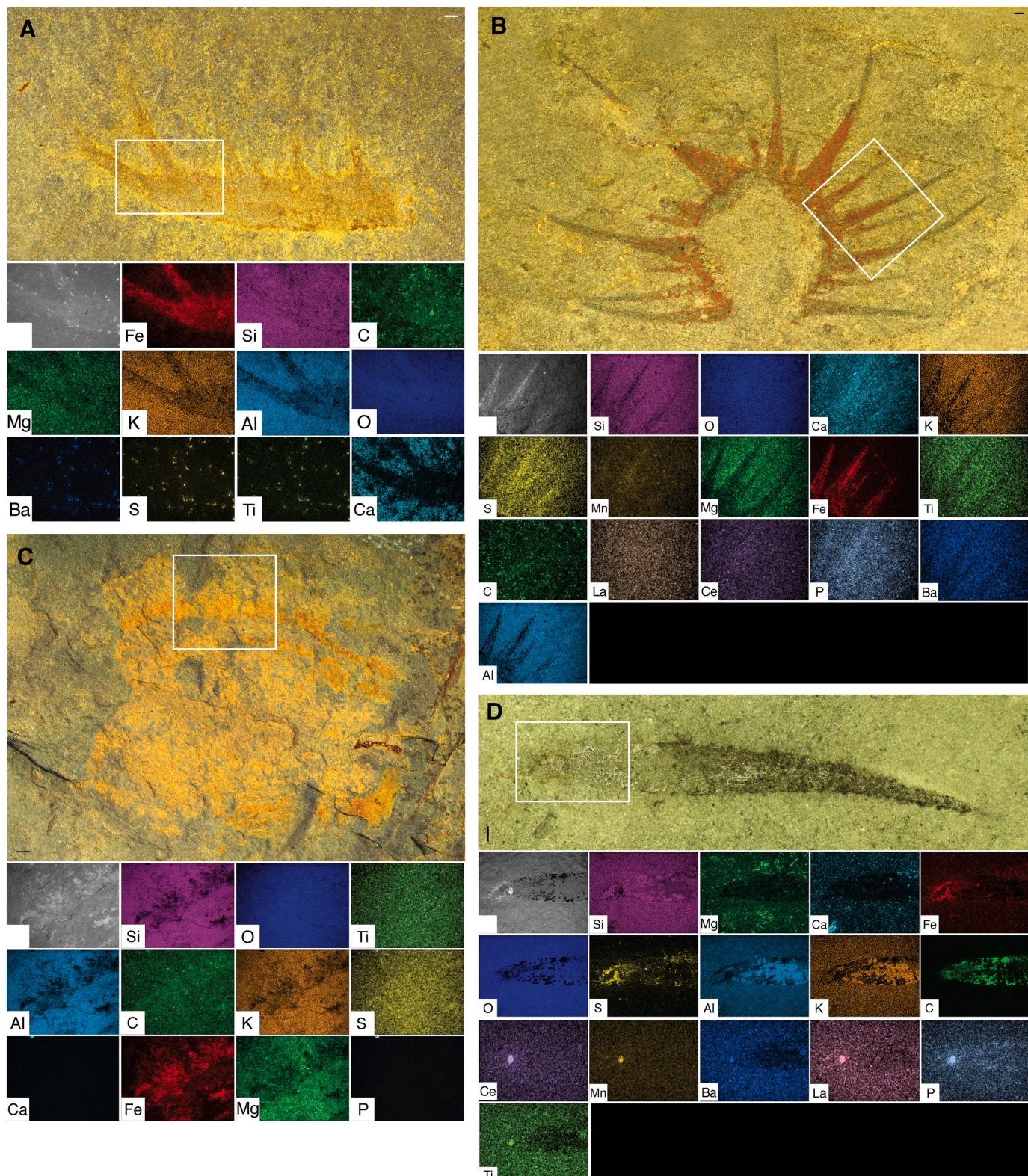


Fig. 2. Images and associated SEM-EDS elemental maps of the area highlighted by boxes in the pictures of selected fossils of diverse preservation from the Spence Shale. Elements mapped are noted in lower left corner, initial grey image is backscatter image of mapped area. A) KUMIP 204777, *Sidneyia*?, isolated feeding appendage. B) KUMIP 314131, *Acinocricus*, isolated dorsal spine set. C) KUMIP 558518, *Dioxycaris*, isolated carapace. D) KUMIP 204366, *Wiwaxia*, isolated spine. Scale bars represent A, B, D: 500 μ m; C: 1 mm.

Spence Gulch represents a middle ground between the highly diverse preservation pathways of the Wellsville Mountain localities, and the limited preservation of Blacksmith Fork and High Creek (Kimmig et al., 2019). Two bradoriid carapaces were collected in-situ from levels 4 m apart. KUMIP 579391 (Appendix A) is heavily enriched in P, in contrast to KUMIP 579390 (Fig. 3B). The carapace of *Dioxycaris* (Fig. 2C), is not enriched in Ca or P, but is preserved mainly as iron sulfides. It remains uncertain whether these differences represent taxonomic differences in original shell composition, or variation in depositional conditions within this locality.

4.1. Kerogenization

Preservation dominated by kerogenization or carbonaceous compression is present in 18 specimens from 4 localities (Miners Hollow, Antimony Canyon, Spence Gulch, and Blacksmith Fork), but is largely limited to small, visibly dark and/or reflective patches. Taxa in which carbon dominates are *Wiwaxia*, *Eldonia*, Hurdidae, and *Sphenoecium*. The fossils preserved in this manner illustrate fine morphological details. The carbon distribution in the *Wiwaxia* specimens (KUMIP 204366, 490944) clearly shows the striated 'ribbing' of the spines, whereas such features are less distinct in other taphonomic modes such as pyritization (Fig. 2D, Appendix A, respectively). Carbon in the analyzed specimens tends to be

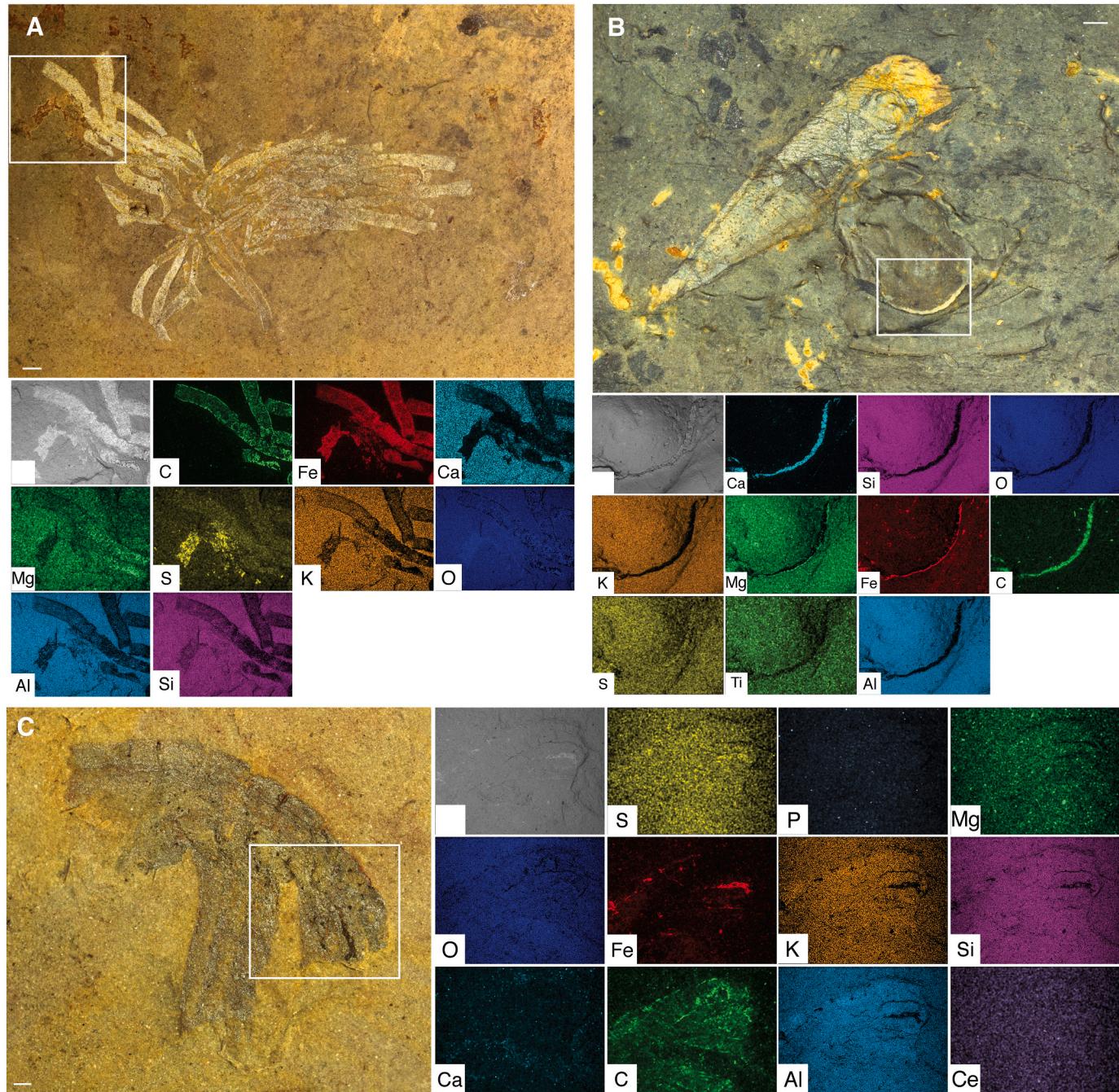


Fig. 3. Images and associated SEM-EDS elemental maps of the area highlighted by boxes in the pictures of selected fossils of diverse preservation from the Spence Shale. Elements mapped are noted in lower left corner, initial grey image is backscatter image of mapped area. A) KUMIP 495314, *Sphenoecium wheelerensis*. B) KUMIP 579390, Bradoriid, isolated carapace. C) KUMIP 491056, Hurdidiid, isolated feeding appendage. Scale bars represent A, B: 1 mm; C: 500 μ m.

concentrated along annulations, edges, and folds, where the overlapping cuticle is thicker. For example, in the huriid element (KUMIP 491056), the boundaries of the podomeres are clearly delineated by observable carbon films, which are also evident in the EDS maps (Fig. 3C). Two of the *Sphenoecium* specimens (KUMIP 495314, 514259) from the Wellsville Mountains localities are preserved as highly reflective carbon films, exhibiting parallel striations of the fusellar construction (Fig. 3A, Appendix A, respectively). However, some specimens with comparable dark coloration exhibit low carbon concentrations in elemental maps.

4.2. Pyritization

Pyrite or its iron oxide- and/or oxyhydroxide-weathering products are present in 35 analyzed samples representing all six sampled localities. While oxidized pyrite preservation is evidenced by a rusty red-orange coloration of the fossils, this mode of preservation is confirmed here by the presence of iron-bearing frambooidal textures indicating weathering-induced pseudomorphs after original pyrite. Iron-bearing mineral concentrations usually follow the margins of the specimens.

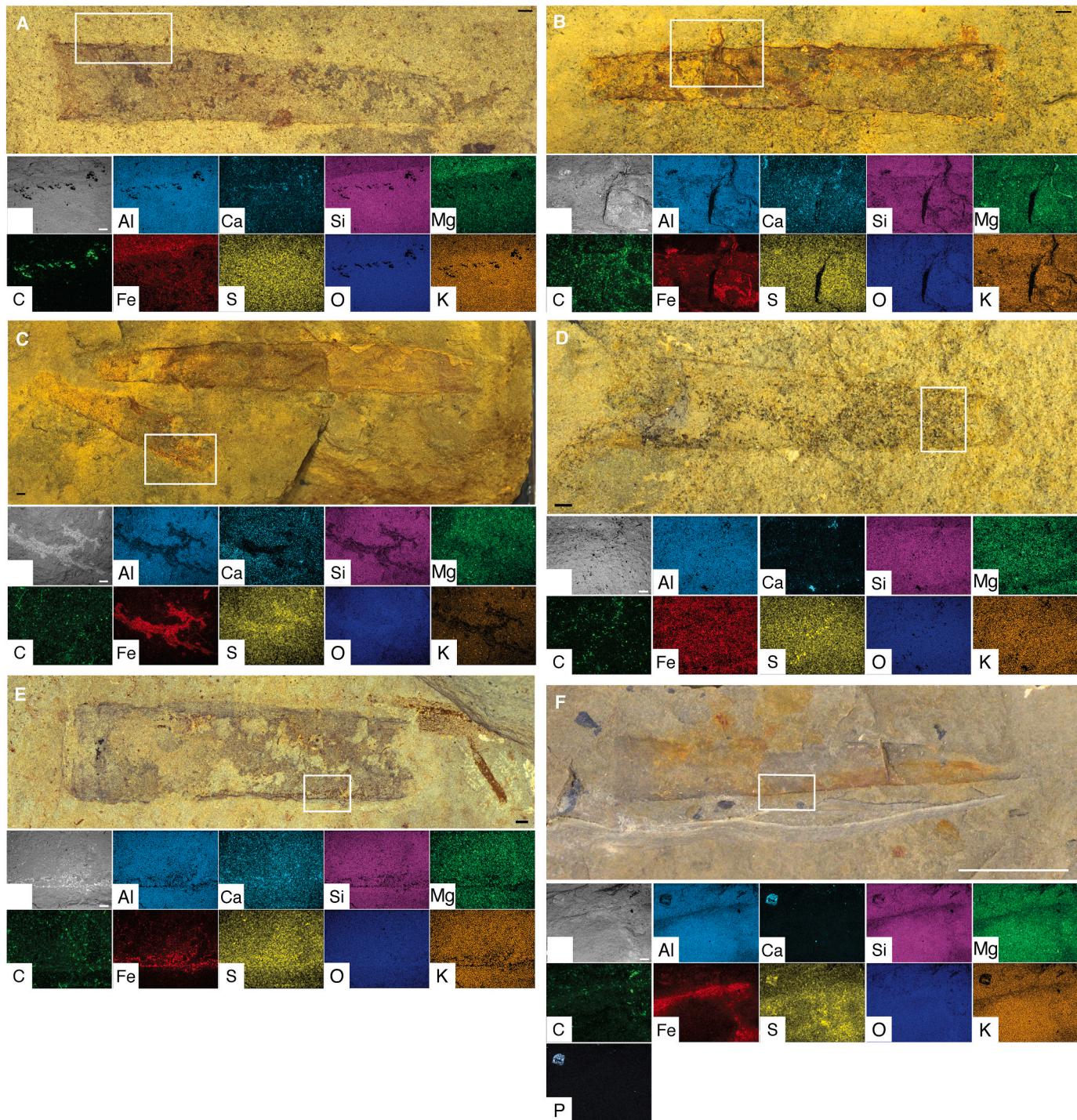


Fig. 4. *Selkirkia* specimens from the Spence Shale with associated SEM-EDS elemental maps of the area highlighted by boxes in the pictures. Elements mapped are noted in lower left corner, initial grey image is backscatter image of mapped area. A) KUMIP 563151, Cataract Canyon. B) KUMIP 563145, High Creek. C) KUMIP 563150, Spence Gulch. D) KUMIP 314158, Miners Hollow. E) KUMIP 144845, Antimony Canyon. F) USNM 768408, Blacksmith Fork. Scale bars represent A-E: 1 mm; F: 1 cm.

KUMIP 563145, *Selkirkia*, typifies this preservation style, with large patches of orange iron staining concentrated around the edges of the tube (Fig. 4B, Appendix A). The presence of unweathered pyrite is intermittent and varies within specimens. In KUMIP 135145, *Dioxycaris*, the carapace preserves scattered pyrite euhedra on the upper edge and more densely packed pyrite frambooids on the lower edge, as evidenced by coincident concentrations of Fe and S (Appendix A). Elevated concentrations of barium are found alongside iron preservation in some specimens, such as KUMIP 135145 (Appendix A) and KUMIP 144845 (Appendix A). The apparent presence of Ti is likely an instrumental artifact, owing to the overlap of the barium L α and titanium K α x-ray energies and the direct spatial overlap in their elemental maps (e.g., KUMIP 135145).

4.3. Phosphatization

The presence of calcium and phosphorus, which together reflect taphonomic phosphatization, is found sporadically in nine specimens from Antimony Canyon, Cataract Canyon, Miners Hollow, and Spence Gulch. Two bradoriid specimens from Spence Gulch, KUMIP 579390, 579391 (Fig. 3B, Appendix A respectively), show elevated calcium in their carapace, but elevated phosphorus is also evident in KUMIP 579391. Some plates of a specimen of *Utahscolex ratcliffei* from Miners Hollow, counterpart KUMIP 204390 and part UMNH IP 4696, are enriched in Ca, although they are mainly preserved in iron oxides (Fig. 2d, Whitaker et al., 2020). Plates in a second Miners Hollow specimen of *U. ratcliffei*, KUMIP 490902, show elevated Ca and P and in some cases Mg (Fig. 3g, Whitaker et al., 2020). In four eldoniid specimens from Miners Hollow (KUMIP 510274 and KUMIP 510275), Cataract Canyon (KUMIP 490937), and Antimony Canyon (KUMIP 204370), specific anatomical tissues are intermittently preserved with localized Ca. A huriid feeding appendage from Miners Hollow, KUMIP 491056, has a low elevation of Ca (Fig. 3C). Eggs of a specimen of *Waptia* cf. *W. fieldensis*, KUMIP 314032, are phosphatized, shown in this case to be controlled by whether or not the egg envelope ruptured resulting in an open or closed environment (Moon et al., 2021).

4.4. Aluminosilicification

Aluminosilicification, or preservation via clay mineral templating, is observed in seven specimens from three localities (Miners Hollow, Cataract Canyon, and Spence Gulch). These seven specimens represent the genera *Sidneyia*? (KUMIP 204779, Table 1), *Acinocrinus* (KUMIP 490967, Table 1), *Selkirkia* (KUMIP 563151, Table 1), *Utahscolex* (KUMIP 490902, Whitaker et al., 2020), unidentified vermiform organisms (KUMIP 314111, Broce and Schiffbauer, 2017; KUMIP 557975, Table 1), and an indeterminate organism (KUMIP 314186, Broce and Schiffbauer, 2017). Compositionally, these specimens show slightly elevated concentrations of Si and Al compared to the matrix.

4.5. Rare earth concentrations associated with *Eldonia*

All four analyzed specimens of *Eldonia ludwigi*, representing three separate localities (Antimony Canyon, Cataract Canyon, and Miners Hollow), revealed a unique preservation mode (Fig. 5A–D). In specimen KUMIP 204370, the coiled sac is preserved by both reflective patches and darker patches (Fig. 5A, Appendix A). The reflective patches contain high concentrations of C associated with Ba, Mn, P, and rare earth elements (REE) Ce and La, whereas the darker patches show only elevated Ca. The patchy distribution of P enrichment in KUMIP 204370 (Fig. 5A) (i.e., it only occurs in the ‘reflective’ areas) could be due to the differentiation of outer and inner tissue layers in the gut tract of *Eldonia* as observed by Butterfield (1996). Thus, the P enrichment in this specimen may indicate that the inner layer of the gut tract presents a micro environment favoring phosphate precipitation. The bifurcating internal lobes in KUMIP 204370 are preserved as darker, non-reflective areas but

are not clearly differentiated from the matrix. In KUMIP 510274 and 510275 (Fig. 5C–D), the coiled sac is preserved as a darkened area with elevated C, and a more reflective area with concentrations of Fe and Mg. Scattered throughout are bladed phosphatic crystals containing Ce and La. The fourth specimen, KUMIP 490937 (Fig. 5B), preserves concentrated detritus, collected along the margins of the poorly preserved *Eldonia*. Scattered crystals within the specimen are enriched with La, Ce, Ba, Mn, P, and irregularly, S. The margin shows elevated C, Fe, and Mg.

4.6. Histology summary

In terms of fossilized tissues, a majority of the eumetazoan genera 73% ($n = 50$) preserve one tissue type, 20% ($n = 14$) preserve two tissue types, and 7% ($n = 5$) preserve three types (Appendix B). The most common tissue type is a biomineralized skeleton (68% of genera), followed by sclerotized tissue (26%), cuticular tissue (19%), soft internal cellular tissue (13%), and soft external cellular tissue (7%). In genera with two tissues present, the two most common tissue types to occur together are sclerotized and cuticular tissue ($n = 7$), followed by sclerotized tissue and soft internal cellular tissue ($n = 2$), and biomineralized skeleton and soft internal cellular tissue ($n = 2$). Of genera preserving three tissue types, the most common co-occurrence is a biomineralized skeleton, cuticular tissue, and soft internal cellular tissue ($n = 3$).

5. Discussion

5.1. Variation in preservation by locality

The sampled genera show intra- and inter-locality variation in preservation. Each locality displayed multiple modes of preservation, including variation between and within stratigraphic horizons.

The influence of locality on preservation is largely limited to the impact of weathering. This is typified by the Spence Gulch locality, which is heavily wooded and located close to Mill Creek. Specimens recovered from float at this locality are predominantly preserved as iron oxides. Specimens recovered in situ (e.g., Fig. 2C; KUMIP 558518, Appendix A) show localized iron sulfide preservation particularly at the base of the exposure where the rocks appear less weathered. The majority of Spence Shale exposures are located on the slopes of steep gullies formed by extensive erosion, and others (i.e., Oneida Narrows) are located on the banks of small rivers. Such settings favor weathering of pyrite to iron oxides.

Soft-bodied preservation declines in abundance from localities in the Wellsville Mountains to those in the Wasatch Range and Spence Gulch in the Bear River Range. Miners Hollow is the most distal of the Wellsville Mountain localities (Kimmig et al., 2019). Geochemical analyses of sediment samples from Spence Gulch, Oneida Narrows, Miners Hollow, and High Creek indicate a predominantly oxic depositional environment, with low bioturbation (Kloss et al., 2015). Therefore, the variability in modes of preservation is likely linked to other second-order environmental factors (“distal factors” of Cai et al., 2012), such as sealing of the sediment-water interface, diagenetic fluid flow, and modern weathering.

Spence Shale specimens were buried rapidly, showing a high fidelity of preservation of labile tissues, although some show initial signs of degradation (e.g., uneven *Utahscolex* tissue margins; Fig. 3a, Whitaker et al., 2020). Collapse preceded preservation, with few exceptions (e.g., *Waptia* cf. *W. fieldensis* eggs), suggesting that this was likely due to decay. This is also supported by the incomplete nature of a large proportion of the Spence Shale soft-bodied fossils (Whitaker and Kimmig, 2020). Early diagenetic processes include phosphatization and pyritization of soft tissues (e.g., bradoriid carapaces, *Waptia* cf. *W. fieldensis* eggs). During multiple orogenic events, the Spence Shale underwent metamorphic alteration, kerogenizing organic carbon, and altering minerals (e.g., apatite to monazite; Broce and Schiffbauer, 2017). Magnesium-rich fluid movement through the Spence Shale dolomitized

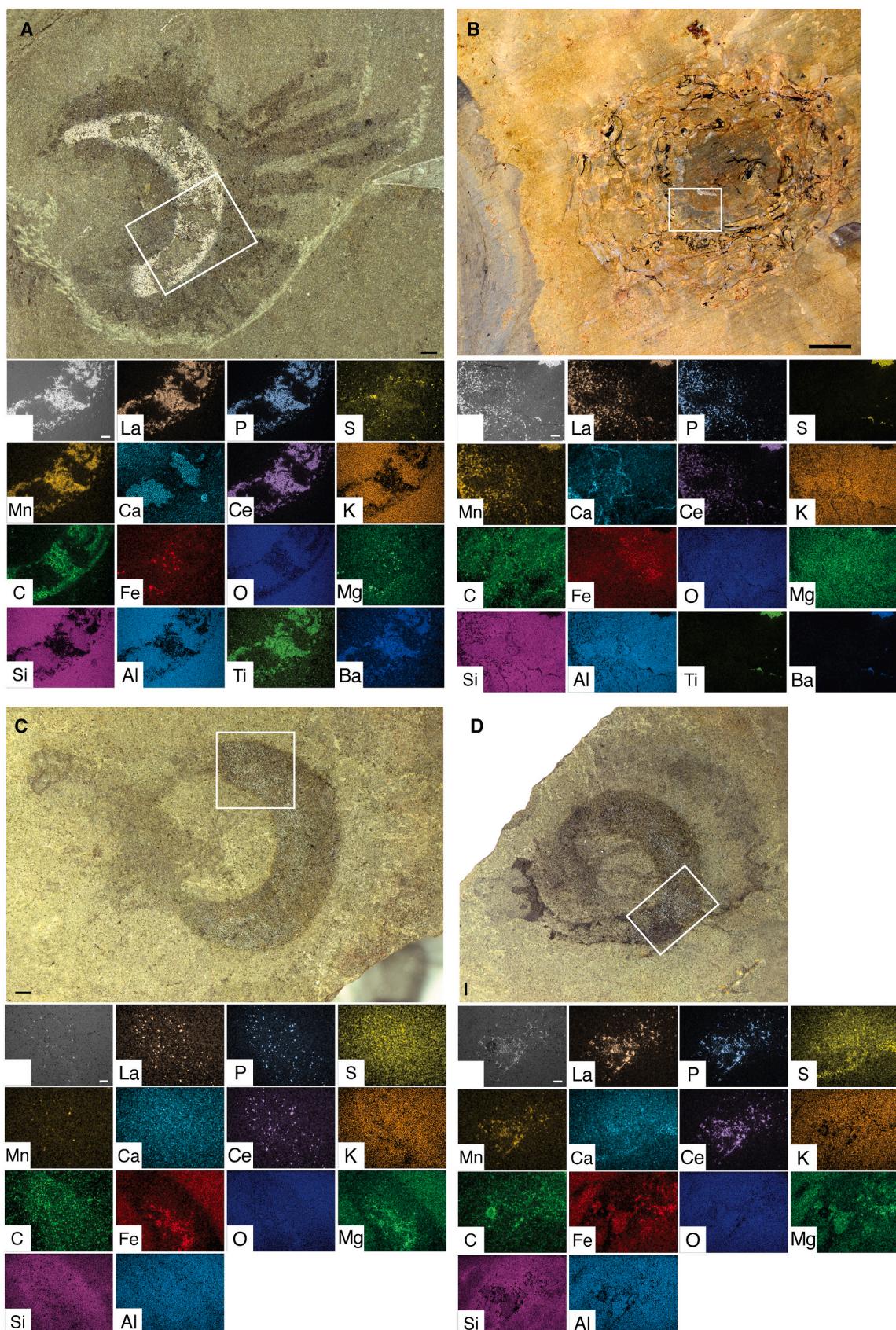


Fig. 5. *Eldonia ludwigi* specimens from the Spence Shale with associated SEM-EDS elemental maps of the area highlighted by boxes in the pictures. Elements mapped are noted in lower left corner, initial grey image is backscatter image of mapped area. A) KUMIP 204370, Antimony Canyon. B) KUMIP 490937, Cataract Canyon. C) KUMIP 510275, Miners Hollow. D) KUMIP 510274, Miners Hollow. Scale bars represent A, C, D: 1 mm; B: 1 cm.

calcium-rich features (e.g., the plates of *Utahscolex*). Pyrite was altered during diagenesis and weathering, forming iron oxides and barite (e.g., KUMIP 135145, Appendix 1; KUMIP 314191, KUMIP 314215; Broce and Schiffbauer, 2017).

5.2. Variation in preservation by taxa

The priapulid *Selkirkia* is the only genus found at all six sampled localities (Fig. 4A–F). The isolated tubes are reddish-orange, and predominantly preserved as pyrite and iron oxides, with patchy carbonaceous areas. The margins of the tubes show the strongest signal, and the annulations on the tubes are rarely evident (Fig. 4E). None of our analyzed specimens preserve the body, and possible soft parts are known in only one specimen, KUMIP 314115 (Kimmig et al., 2019, fig. 5d). The poor preservation of *Selkirkia* in the Spence Shale contrasts with that in other Cambrian BST Lagerstätten, such as the Xiaoshiba and Chengjiang, where the pharynx, digestive tract, and gut contents are preserved (Tian et al., 2015; Wang et al., 2021; Yang et al., 2021b). A higher proportion of specimens of *Selkirkia* from the Burgess Shale and Wheeler Lagerstätten also preserve soft parts, approximately 20% and 33%, respectively (Conway Morris and Robison, 1986). The tube of *Selkirkia* tube is more decay resistant than its trunk, but other stem-priapulid body fossils from Miners Hollow preserve soft tissues such as the everted proboscis and gut tract (Whitaker et al., 2020). In the Chengjiang Lagerstätte, *Selkirkia* tubes, body fossils, and eggs in the tubes are preserved in carbon and iron oxides, formed by weathering of pyrite microcrystals (Wang et al., 2021; Yang et al., 2021b). The similarities in the mode of *Selkirkia* preservation between the Spence Shale and the Chengjiang Lagerstätte emphasizes differences in the quality of preservation – it is likely some of the “empty” tubes represent exuviae of the molting *Selkirkia*, but does not fully explain the lack of soft parts.

Acinocricus was sampled from three localities in the Wellsville Mountains (Miners Hollow, Cataract Canyon and Antimony Canyon), where it is represented by the cuticle of isolated body segments (Fig. 2B, Appendix A). They are preserved in multiple ways: pyritization (Fig. 2B; KUMIP 4911082, Appendix A), aluminosilification (KUMIP 490967, Appendix A), and phosphatization (Fig. 2B). Disarticulated spines are most common, but articulated fossils of *Acinocricus* are known from the Wellsville Mountains (Caron and Aria, 2020). In KUMIP 314131 (Fig. 2B), the enrichment of Fe and S echoes the “cone-in-cone” spine structure reported by Caron and Aria (2020).

The two isolated spines of *Wiwaxia*, KUMIP 204366 (Fig. 2D) and KUMIP 490944 (Appendix A), are predominantly preserved as carbon films, as well as Fe enrichment. Acicular crystals of iron sulfide were reported within the striations on *Wiwaxia* spines from the Burgess Shale (Smith, 2014), which are predominantly carbonaceous (Butterfield et al., 2007). In the Spence Shale specimens, the striation boundaries are preserved as cracks or terminations of the carbon film (Fig. 2D) or as concentrations of carbon (Appendix A).

Two bradoriid carapaces (KUMIP 579390, Fig. 3B; and KUMIP 579391, Appendix A) are among the few Spence Shale specimens primarily preserved as calcium carbonate or calcium phosphate. This is interpreted as diagenetic and not useful as a taxonomic indicator (Vannier et al., 2005; Streng et al., 2008). The preservation of the Spence Shale bradoriids is similar to those found elsewhere (e.g., Streng et al., 2008; Peel et al., 2021), but differs from other Spence Shale soft-bodied arthropods which do not show the same diagenetic calcium enrichment (e.g., *Dioxycaris*), although the sample size is limited.

Phosphatic material, presumably apatite, is present in the eggs of a specimen of *Waptia* cf. *W. fieldensis* (KUMIP 314032) (Moon et al., 2021). The preservation of these eggs in the Spence Shale (preservation through phosphatization and carbonaceous compressions and some differentiation of the inner and outer layers of the eggs) is similar, but less detailed, to that of *W. fieldensis* eggs from the Burgess Shale, where an inner portion is preserved in carbon, calcium, phosphorus, iron, and sulfur, and an outer portion in aluminum and potassium (Caron and

Vannier, 2016). This differs in detail to the eggs of *Chuandianella ovata*, a bivalved waptiid arthropod from the Chengjiang Lagerstätte. Its eggs are primarily preserved in iron oxides, the weathering product of authigenic pyrite, with additional slight enrichment in phosphorus, sulfur, carbon, cobalt, platinum, and copper (Ou et al., 2020).

Preservation differences within taxa are largely limited to differences in specific tissue type, i.e., gut tract phosphatization or pyritization. Gut tract phosphatization is found in the Wellsville Mountain eldoniid specimens (discussed below), and an *Anomalocaris* specimen from Miners Hollow, KUMIP 314037 (Appendix A). The phosphatized patches of *Anomalocaris* are located medially on the specimen, where the digestive system would have been located (Briggs et al., 2008). Densely packed parallel fibers are arranged across the patches, potentially representing adjacent muscle fibers or mid-gut glands (Vannier et al., 2014).

5.2.1. REE enrichment

The *Eldonia* specimens are all enriched in the light Rare Earth Elements (LREE) La and Ce, in contrast to other currently sampled taxa in the Spence Shale. The LREE are associated with P, Ba, Ti, and Mn, suggesting that apatite has been altered to monazite, a LREE-bearing phosphatic mineral. Monazite has been identified associated with the gut tracts of two vermiform specimens from the Spence Shale (Broce and Schiffbauer, 2017), in isolated areas on the carapace of *Canadaspis* and surrounding sediment from the Pioche Shale Lagerstätte (Moore and Lieberman, 2009), and in association with numerous fossils from the Sirius Passet Lagerstätte (Nielsen et al., 2021). In the Pioche Shale, the monazite is found in both the fossil and sediment. Moore and Lieberman (2009) interpreted it as diagenetic rather than detrital in the sediment. REE (including La and Ce) in phosphatized palaeoscolecid casts from the Ordovician Tafilelt Konservat-Lagerstätte, were likewise interpreted as diagenetic (Gutiérrez-Marco and García-Bellido, 2015). In the Sirius Passet Lagerstätte, monazite was most commonly found in the adjacent sediment, and at the tissue-sediment interface, rather than directly associated with phosphatized or previously phosphatized tissues (Nielsen et al., 2021). The alteration of phosphate to monazite is interpreted as the result of metamorphic alteration of original phosphatic preservation (Nielsen et al., 2021).

Monazite is formed from the dissolution of LREE-bearing apatite, potentially identifying apatite, or an apatite precursor, as the authigenic mineral preserving the gut tracts of the eldoniids, as suggested for the vermiform organisms in the Spence Shale (Broce and Schiffbauer, 2017). The presence of monazite in *Eldonia* specimens is independent of locality, and appears to be taxon-specific, as co-occurring species (Table 1) did not show similar concentrations of REE with the exception of a well-defined patch of REE-enriched phosphate at the base of a *Wiwaxia* spine (KUMIP 204366; Fig. 2D). KUMIP 490937 (Fig. 5B) from Cataract Canyon preserves aggregated detritus around a poorly-preserved eldoniid disk suggesting a high-energy environment where biomineralized fragments were swept by currents and collected around the eldoniid on the sediment surface, as was observed in examples from the Miaotianshan Shale (Vannier and Chen, 2005). Despite the degradation of this eldoniid, it shows REE enrichment. Previously analyzed Spence Shale specimens where monazite is associated with a gut tract (Table 1; Broce and Schiffbauer, 2017) cannot be identified to taxon, although a plausible interpretation could be the degraded gut of an eldoniid.

Phosphatization associated with gut tract preservation has been reported in numerous taxa from other BST Lagerstätten, including trilobites (Weeks Formation), radiodonts (Spence Shale and Burgess Shale), *Leanchoilia*, *Odaraia*, *Opabinia*, *Alalcomenaeus*, *Perspicaris*, *Naraoia*, *Burgessia*, *Plenocaris*, *Molaria*, *Sidneyia*, *Isoxys* (Burgess Shale), *Pambdelurion* and *Campananuta* (Sirius Passet Lagerstätte) and an indet. *Fuxianhuia*-like arthropod (Kaili Formation) (Briggs and Whittington, 1985; Briggs and Collins, 1999; Butterfield, 2002; Zhu et al., 2004; Lerosey-Aubril et al., 2012; Schiffbauer et al., 2014; Vannier et al., 2014; Strang et al., 2016; Zacaï et al., 2016). The preservation of digestive

systems is likely facilitated by an internal source of phosphate in combination with a closed-system post-mortem (Butterfield, 2002; Leroosey-Aubril et al., 2012; Strang et al., 2016). Possible ingested sources of P have been suggested to include microbial mats (Leroosey-Aubril et al., 2012) and other arthropods (Strang et al., 2016); although “spherites” rich in Ca and P, a common mineral storage feature in the midgut tissue in modern chelicerates, crustaceans, and insects, also provide a source of P (Butterfield, 2002; Leroosey-Aubril et al., 2012; Zacaï et al., 2016).

Modern non-predatory benthic vertebrates and invertebrates have higher concentrations of REE than their predatory counterparts (Mayfield and Fairbrother, 2015; Amyot et al., 2017). Macro-invertebrates that feed by ingesting organic particulates from the sediment are positively correlated with REE concentrations, more so than other feeding styles, indicating that sediment ingestion is an effective bio-accumulation pathway (Pastorino et al., 2019). Marine invertebrates (e.g., scallops, amphipods) accumulate REE despite their relatively low concentrations in seawater, also indicating that the source is ingested via sediment (Bustamante and Miramand, 2005; Palmer et al., 2006). *Eldonia* has been interpreted as a nektobenthic filter feeder (Caron et al., 2010; MacGabhann, 2012), and the concentration of REE in its gut, as preserved, may be consistent with an in-vivo accumulation, that might have led to elevated levels of REE in the carcass, which were then preferentially incorporated in the monazite.

An alternative explanation, and more consistent with previous interpretations of REE accumulation, is that phosphate in the coiled sac of *Eldonia* favored the accumulation of LREE during diagenesis. Phosphate and calcium are stored in the gut of many arthropods (e.g., Zacaï et al., 2016). REE (Y, Nd) and Sr are enriched in the phosphatized soft tissue of decapod shrimp from the Upper Cretaceous Djebel Oum Tkout Lagerstätte (Gueriau et al., 2014). The REE element Y is concentrated in the phosphatized gut tract of penaeoid shrimp from the Triassic Paris Biota (Brayard et al., 2019). A comparison of modern and Miocene gastropods, echinoderms, and bivalves concluded that significant concentrations of REE elements are a diagenetic signal (MacFadden et al., 2015).

The “open system” behavior of apatite is well known, providing readily available sites for elemental replacement within the lattice (Kocsis et al., 2010; Herwartz et al., 2011; Herwartz et al., 2013; Keenan, 2016; Zhang et al., 2016; Decrée et al., 2018; Kocsis et al., 2020). Trace element uptake and exchange fluctuate throughout early and late diagenesis, reflecting changes in the composition of the fluid source (Herwartz et al., 2011; Herwartz et al., 2013; Decrée et al., 2018). Monazite has been experimentally formed from metasomatic dissolution of LREE-apatite under a range of simulated metamorphic temperatures and pressures, including greenschist conditions (Harlov and Förster, 2003). In this scenario, the coiled sac was diagenetically enriched in REE following initial phosphatization. The REE-enriched apatite was then altered to monazite during later diagenesis.

Several other wholly or partially phosphatized specimens do not show the same REE enrichment, but this may reflect limited sampling. Two bradoriid carapaces (KUMIP 579390 and KUMIP 579391, Fig. 3B, Appendix A, respectively) show diagenetic phosphate and calcium enrichment of the original unmineralized shell, but lack REE enrichment. Diagenetic phosphatization has been observed in other bradoriids (e.g., Peel et al., 2021) and might suggest oscillating pore-water redox during periods of prolonged oxygen depletion in the Spence Shale, which would favor phosphate nucleation during early diagenesis (Pruss et al., 2018). The eggs of *Waptia* cf. *W. fieldensis* (KUMIP 314032; Moon et al., 2021) and phosphatic patches, which might represent digestive glands (Vannier et al., 2014), in *Anomalocaris* KUMIP 314037 (Appendix A), have not been analyzed for REE. The problematic shell-bearing *Armillimax* (KUMIP 490943) and the palaeoscolecid *Utahscolex* (KUMIP 204390/UMNH IP 4696 and KUMIP 490902), which preserve gut tracts, show diagenetic alteration, i.e., the dolomitization of the original calcium carbonate shell and phosphatic plates, respectively, but lack REE enrichment within the gut tract (Whitaker et al., 2020; Kimmig and Selden, 2021).

Based on the current data from this and previous studies, it is impossible to determine whether the REE enrichment in the *Eldonia* specimens represents a biological or strictly diagenetic signal. Further analyses of *Eldonia* and additional phosphatized specimens from the Spence Shale and other deposits will elucidate whether this pattern is constrained to this genus, and determine whether the REE enrichment is limited to phosphatized gut tracts or additional tissues.

5.3. Comparison to other BST deposits

The Spence Shale is one of five Cambrian Lagerstätten in Utah, the others located in the House Range (the ‘deep’ Wheeler, Marjum and Weeks formations), and the Drum Mountains (the ‘shallow’ Wheeler Formation). In the Wheeler Formation, the extensive preservation of carbonaceous compressions has been interpreted as the result of carbonate-precipitated occluded porosity in anoxic bottom waters owing to the precipitation of carbonate bed seals (Gaines et al., 2005). Some Wheeler Formation soft-bodied arthropods are preserved through pyritization, a product of anaerobic sulfate reduction (e.g., Leroosey-Aubril et al., 2020). Articulated palaeoscolecids from the Marjum Formation are preserved through Fe, Mg, Ca, and P, but carbonaceous remains of tissues are absent (Leibach et al., 2021). This differs from the preservation of palaeoscolecids from the Spence Shale, which preserve labile tissues of the proboscis and gut tract in addition to the plates (Whitaker et al., 2020).

The representation of multiple modes of preservation, even within specimens, is shared with other BST deposits. The Chengjiang Biota features phosphatization, pyritization, Fe-rich aluminosilicates, and organic films (Zhu et al., 2005). Non-mineralizing organisms of the Chengjiang biota are predominantly preserved through Fe-oxides, whereas phosphatization is rare (Gabbott et al., 2004; Zhu et al., 2005). The Spence Shale, like the Chengjiang Biota, has undergone weathering, altering pyrite to Fe-oxides (Zhu et al., 2005). Compared to the Burgess Shale, the Spence Shale represents a different depositional environment: more rapid fluctuations between oxic, dysoxic, and anoxic conditions, limited to no transport, and less potential for immediate burial (Garson et al., 2012; Kloss et al., 2015). The Burgess Shale also features phosphatization and pyritization in addition to the predominant preservation through carbonaceous compressions (García-Bellido and Collins, 2006; Gaines et al., 2008; Gaines et al., 2012; Gaines, 2014; Vannier et al., 2014; Zacaï et al., 2016). Phosphatization is overwhelmingly found associated with gut tract preservation. Pyritization is rare in the Burgess Shale: a sample of more than 1000 analyzed *Marrella* specimens included only two with areas of pyritization (García-Bellido and Collins, 2006). Pyrite is also rarely associated with gut tract preservation (Butterfield, 2002; Lin and Briggs, 2010). Both the Spence Shale and the Burgess Shale have undergone metamorphism: the Burgess Shale has undergone multiple stages of greenschist and sub-greenschist metamorphism, with variable impact on preservation based on position relative to the escarpment (Powell, 2003). The metamorphism of the Burgess Shale primarily affects kerogenization and late-stage aluminosilification, whereas metamorphism of the Spence Shale also affected modes of mineralization (e.g., phosphatization and dolomitization).

In addition to the familiar preservation modes of BST deposits, association of unusual minerals with soft-tissues occurs in the Burgess Shale (Pushie et al., 2014), where possible biogenic copper, in the form of chalcopyrite, was reported in the arthropod *Marrella*, although the evidence for a biogenic origin of chalcopyrite has since been disputed (Gaines et al., 2019). Microbially mediated silver has been reported from the Ravens Throat River Lagerstätte (Kimmig and Pratt, 2022), indicating that biogenic minerals might be more common in BST deposits than previously thought.

Using the taphonomic comparison metrics of Saleh et al. (2020a), the Spence Shale shares most characteristics with the Fezouata Biota out of the three Lagerstätten considered (Fezouata, Burgess Shale and

Chengjiang). A majority of specimens preserve only one of the tissue types in contrast to the Burgess Shale or the Chengjiang, where multiple tissue types are more common in a fossil. The average number of tissue types per genus is comparable to that in the Fezouata Biota (1.333 in the Spence Shale, 1.316 in the Fezouata) (Appendix B). Soft cellular tissue, either external or internal, is rarely preserved in the absence of more decay-resistant biomineralized, sclerotized, or cuticular tissue, where they co-occur. The Spence Shale, like the Fezouata, has undergone modern weathering, affecting the original carbonaceous compression and pyrite preservation (Saleh et al., 2020b, 2021). The carbonate cement ‘caps’, interpreted as sealing the sediment surface and promoting preservation in other Lagerstätten (Gaines et al., 2012; Saleh et al., 2020b), are absent in the Spence Shale, as at Fezouata, creating conditions favoring pyritization.

6. Conclusion

Based on this study and previous work, the primary processes influencing preservation in the Spence Shale are, sequentially, survival of organic remains, early taphonomic precipitation of phosphate and pyrite, maturation of carbonaceous remains to kerogen, later metamorphic alteration of phosphate to monazite, and other effects of modern weathering (e.g., oxidation of pyrite). These preservation modes are comparable to preservation types from other BST deposits, against the backdrop of an atypical dynamic redox environment across the Spence depositional settings. Diagenesis has resulted in alteration of the REE-enriched phosphate in the gut tracts of *Eldonia*. The apparently unique association of REE elements with *Eldonia* in the Spence Shale warrants further investigation, as REE enrichment in BST preservation is rare. A likely contributing factor is the exclusion of REE and trace elements in previous analyses. Further SEM-EDS analyses of eldoniids from the Spence Shale and other BST Lagerstätten, as well as detailed studies of their anatomy and gut contents, will elucidate whether this pattern reflects their mode of life, or a taphonomic pathway shared by specific tissues in the Spence Shale.

Despite taphonomic conditions that limited preservation in the Spence Shale (e.g., initial decomposition, bioturbation, diagenetic alteration, modern weathering, etc.), exceptional preservation still occurred in a variety of shelf environments. The presence of preserved muscle tissue, eyes, eggs, and the discovery of new taxa emphasize the importance of this Lagerstätte. With continued study, the Spence Shale is “catching up” in importance with better known BST-deposits.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.palaeo.2022.110909>.

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